



Evaluation of the Methylation of *MIR129-2* Gene in Gastric Cancer

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Abstract

Background Genetic and epigenetic changes have strong role in the development of gastric cancer. The mutation of the *MIR129-2* gene is one of the major causes in many cancers, especially gastric cancer. The aim of this study was to investigate the methylation changes of the *MIR129-2* gene in tumor and normal tissue of patients with gastric cancer.

Method In this study, 50 gastric cancer patients with Iranian Azari ethnic origin without any familial relations were included. Genomic DNAs was extracted from the tumoral and normal tissues. Then the promotor regions of the *MIR129-2* gene were analyzed by methylation-specific PCR (MSP) to evaluate the presence or absence of methylated CpG sites.

Results There was a statistically significant difference in methylation level of *MIR129-2* gene between tumoral and normal tissues. It was observed that 84 out of 100 CpG sites were methylated in tumoral tissues in comparison to 13 out of 100 CpG sites in normal tissues.

Conclusion *MIR129-2* gene was hypermethylated in tumoral tissues, suggesting that methylation is involved in the development of gastric cancer.

Keywords Gastric cancer · *MIR129-2* gene · CpG site

Introduction

Cancer is an asymmetric division of the cells of the body, in which the cells have no longer normal mechanisms of cell growth and division. The exact reason of this phenomenon is uncertain, but genetic factors or cases that interfere with the activity of the cells are likely to play a role in this occurrence [1, 2]. Gastric cancer, is a malignancy caused by the proliferation and spread of gastrointestinal cells [3]. Gastric

cancer is responsible for 7% of the total cancers and is ranked fifth among the most common cancers whereas it is responsible for 9% of cancer deaths worldwide [4]. Gastrointestinal cancer predisposing factors are categorized into two genetic and environmental factors; environmental factors include infection with *Helicobacter pylori* or Epstein–Barr virus, smoking, and diet with greasy foods and high amount of salt [2, 5]. After environmental factors, the second important factor involved in the development of cancer is genetics. About 10% of all people with gastric cancer present the disease as heredity, which suggests that, like other cancers, genetics has an outstanding role in the creation and development of this malignancy [6, 7]. The most important and famous gene involved in gastric cancer is the *Cadherin 1 (CDH1)* gene, also known as Hereditary Diffuse Gastric Cancer (HDGC), which encodes the cadherin E protein and is located on chromosome 6. Although mutation in *CDH1* has not been the only genetic cause of gastric cancer, yet many mutations in several genes, such as *P53*, *BRCA2*, and epigenetic processes, have been responsible for causing this disease [8, 9].

MicroRNAs (miRNAs) are regulatory, small, and noncoding RNAs and are known as gene expression modulators that act at the posttranscriptional level [10], controlling the expression of most of the human protein-coding genes. Recently, it has been observed that different miRNAs are controlling cell

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signaling pathways in normal and tumor tissues [11, 12]. Some of the roles of miRNAs include degradation or translational inhibition of the target mRNAs by base-pairing with their 3' untranslated region (3'UTR) [13]. Another role include regulation of thousands of genes, playing an essential role in cell development, proliferation, differentiation, chromatin structure, apoptosis, metabolism, and morphogenesis [14]. The role of epigenetics with processes such as methylation and alkylation and acetylation has been shown in the regulation of expression profiles of miRNAs in gastric cancer [15–17]. DNA methylation is a process by which methyl groups are added to the cytosine nucleotides of DNA molecules, resulting in suppression of gene expression. *MIR129-2* is an important miRNA involved in different cancers [18]. Generally, *MIR129* has been reported as a tumor suppressor gene in most cancers, and overall downregulated expression of this molecule has been reported in cancers and also, it should be considered that there are different mechanisms for reducing the expression of a miRNA in cancers, and the most important of which are deletion, point mutations, regulatory effects of other genes, and ultimately epigenetic changes, such as methylation, which seems to be the last option in the regulation of *MIR129-2* gene and is playing the most important role [19–21]. The purpose of the present study was to investigate the methylation changes of the *MIR129-2* gene in tumoral and normal tissue of patients with gastric cancer.

Material and Methods

Population and Sampling

In this study, 50 patients were diagnosed with gastric cancer. The disease was identified by a gastroenterologist and patients were referred to the surgery. Participants in this disease were all from the Azari population living in the northwest of Iran. Tumoral and healthy marginal tissues around the tumor were obtained from each study subject. Inclusion criteria of this study were gastric cancer disease in stage 2, having Azerbaijani ethnicity and habitation in the northwest of Iran. Exclusion criteria included existence of family relationship with other patients in the study, metastasis, unwillingness to

continue cooperation in research, simultaneous diagnosis of another malignant disease other than gastric cancer, history of chemotherapy, and radiation therapy. Consent forms were obtained from each study subject, and the local ethical committee of Tabriz University of Medical Sciences approved the protocol of the study.

DNA Extraction and Bisulfite Treatment

DNA extraction was performed according to the protocol of phenol-chloroform approach. The quality and quantity of the DNAs were evaluated by nanodrop spectrophotometer. Before performing methylation-specific PCR (MSP), extracted DNA was treated with sodium bisulfite according to the protocol of ZYMO RESEARCH kit. During this process, all non-methyl cytosine is converted to uracil, but methylcytosine remains intact.

Methylation-Specific PCR

This technique requires use of two primers for the methylated DNA (M primer) and unmethylated DNA (U primer). In fact, for each sample, two PCR reactions were performed separately with each of the primers. The amplification with M primer represents methylation in CpG islands and amplification with U primer represents the lack of methylation in the examined region. Amplification with both primers represents partial methylation in the target area. The product of the M primer was 189 bp length and the length of the U primer was 188 bp. After performing the PCR reaction, the product of all samples (300 reactions) was electrophoresed on the agarose gel to ensure the reaction of the products. Examples of agarose gel electrophoresis related to the met primer with sequences of 189 nucleotides and unmet primer with sequences of 189 nucleotides with 100 bp ladder are shown in Fig. 1.

The Primer 3 software was used for designing the U and M primers (Table 1). The primers were then examined by NCBI Blast for specificity and proper melting temperature was determined for each primer with a temperature gradient.

PCR conditions were determined by using various variables including DNA and MgCl₂ concentrations and temperature profiles for PCR reaction conditions is presented below. The

Fig. 1 For each sample, the PCR reaction was performed for *MIR129-9* and for both met and unmet primers. Samples with methylation in both alleles responded only with 189 and non-methylation samples with only 188 nucleotide long primers. Samples with partial methylation reacted with both primers

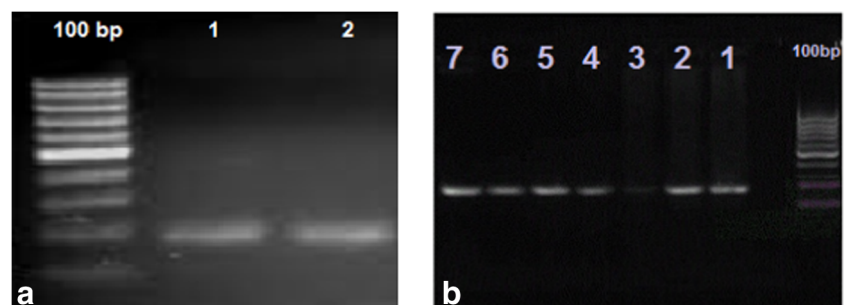


Table 1 Characteristics of the primers used in the study

Target	Forward primer (5'–3')	Reverse primer (5'–3')	Amplicon size (bp)	T _m (°C)
Unmethylated	GAGTTGGGGGATTGTGGAT	AATATACCAACTTCTTCAATTCACCA	188	59
Methylated	GAGTTGGGGGATCGCGGAC	ATATACCGACTTCTTCGATTGCGCCG	189	55

amount of consumables in the PCR reaction for U primer is Master Mix Red 10 μ l, DNA Template 2 μ l, Primer 1 μ l, Depc Water 7 μ l, and for M primer is Master Mix Red 10 μ l, DNA template 2 μ l, Primer 1 μ l, Depc Water 7 μ l, and Mgcl₂ (25 Mm).

Statistical Analysis

Statistical package for the Social Sciences (SPSS) software v.22 used to analyze data and plotting was performed using GraphPad Prism software v.6. The Pearson's chi-square and Fisher's exact test were used to check if there is a significant difference between the two groups in terms of methylation. The significant level was considered as $P < 0.05/100$ (Fig. 2).

Results

Totally, in tumor samples, the prevalence of methylation of the gene was 84 out of 100 CpG sites and in healthy samples, this number was 12. This indicates an increase in a significant difference between the two groups with respect to methylation of *MIR129-2* gene. Regarding the pattern of the methylation of tumors and marginal samples, there were no significant differences in the prevalence of methylation with respect to sex, age, history of cancer, smoking, and alcohol consumption.

Discussion

In this study, 50 samples were examined for the methylation level of the CpG sites in the promoter of *MIR129-2* gene. There was a significant difference in the distribution of

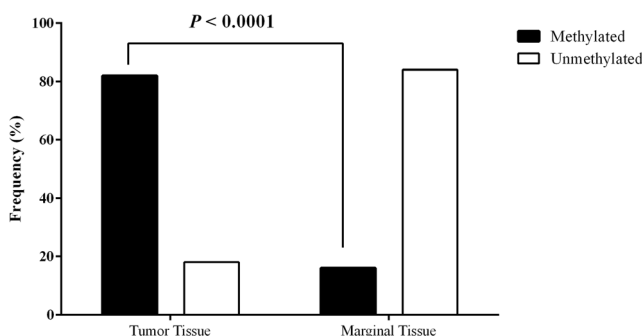


Fig. 2 Comparison of methylation percentile of tumor and margin tissues from patients with gastric cancer

methylated sites between tumoral and marginal tissues from gastric cancer subjects.

For the first time, Katada and his colleagues in a project published in 2009 revealed miR-129 as a distinct risk factor for gastric cancer [22]. Another study by Bandres et al. evaluated the methylation of *MIR129-2* gene in colorectal cancer and it was introduced as a potential biomarker for colorectal cancer. One of the differences between this study and our research was the type of involved tissue and another was the method of investigation. A similar study evaluated the expression profile of miR-129 in colorectal cancer and introduced it as a biomarker for this disease [23].

A study by Liu and colleagues in hepatocellular cancer also confirmed the role of miR-129 in this cancer [24]. Although the study by Chen et al. on esophagus cancer with respect to the DNA methylation of *MIR129-2* gene did not report a relationship between methylation level and development of this cancer [25]. A study by Kang and colleagues on esophageal cancer [26], as confirmed by the results of the Chen study, is in contradiction with our studies, which contradicts the role of different gene and molecular pathways in various cancers. The study by Chen is one of the few studies performed on methylation of gastric cancer [25], the results of which generally confirm the results of our review.

Although the P value and odds ratio varies with the results of our study, this difference can be due to a variety of reasons, including the difference in the population studied and the method used in the two studies. In all of the above studies, the sample population is different from our society. Generally, these studies are a statistical community with a higher frequency of our study and at least 100 patients. However, in this study, only 50 patients were studied and the difference in the volume of the studied population can be considered as one of the most important factors justifying the differences. The second leading cause of this difference can be the difference in the type of study, as in this study, we examined the level of methylation of the desired gene, while some previous studies only evaluated the expression. The third major reason for using methyl specific (MS)-PCR in this study is a qualitative method, while some previous studies have used quantitative methods to investigate the amount of methylation. The fourth factor that can somehow explain the differences is race. This raises the issue of racial differentiation as one of the factors contributing to the development of the disease. The fifth factor that can be the most important factor in making the difference

in the data of this gene in our study compared with other similar studies is the limitation in collecting clinical data from patients. Generally, due to limitations in collecting samples and access to patient's information, we were unfortunately unable to perform appropriate statistical surveys.

In conclusion, the methylation of *MIR129-2* gene in patients with gastric cancer in the northwestern community of Iran has a significant difference in tumor samples with healthy samples and these changes can be considered as a risk factor for gastric cancer. The drugs that reverse the methylation of CpG sites can be further investigated for possible treatment of patients with gastric cancer.

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Compliance with Ethical Standards

Ethics Approval and Consent to Participate Informed consent was obtained from each patient and the ethical approaches were approved by the ethic committee of Tabriz University of Medical Sciences.

Conflict of Interest The authors declare that they have no conflict of interests.

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