

# Current Neurogenic and Neuroprotective Strategies to Prevent and Treat Neurodegenerative and Neuropsychiatric Disorders

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**Abstract** The adult central nervous system is commonly known to have a very limited regenerative capacity. The presence of functional stem cells in the brain can therefore be seen as a paradox, since in other organs these are known to counterbalance cell loss derived from pathological conditions. This fact has therefore raised the possibility to stimulate neural stem cell differentiation and proliferation or survival by either stem cell replacement therapy or direct administration of neurotrophic factors or other proneurogenic molecules, which in turn has also originated regenerative medicine for the treatment of otherwise incurable neurodegenerative and neuropsychiatric disorders that take a huge toll on society. This may be facilitated by the fact that many of these disorders converge on similar pathophysiological pathways: excitotoxicity, oxidative stress, neuroinflammation, mitochondrial failure, excessive intracellular calcium and apoptosis. This review will therefore focus on the most promising achievements in promoting neuroprotection and neuroregeneration reported to date.

**Keywords** Neurogenesis · Neurotrophic factors · Neural stem cells · Neurodegenerative disorders · Retinoic acid · Curcuminoids

## Introduction

Although the subject of the brain as a “renewable” organ was controversial for many years, it has been discovered that it is in fact capable of significant, yet limited, plasticity and neurogenesis. This could explain the occurrence of spontaneous rehabilitation after brain damage (Fuchs and Flügge 2014). The neurogenic potential can be defined as the ability of neural stem cells (NSCs) or neural precursor cells (NPCs), under specific conditions, to differentiate into mature central nervous system (CNS) cells such as neurons, astrocytes and oligodendrocytes (Gage 2000), and functionally integrate into the surrounding neuronal network. This allows the establishment and maintenance of polarized excitatory synaptic contacts and the restoration of the structure and function of the impaired nervous system (Fuchs and Flügge 2014; Lepski 2012; Liebau et al. 2007).

The existence of NPCs was first reported in the rodent brain (Altman and Das 1965; Altman 1962) and later isolated and successfully differentiated *in vitro* (Reynolds and Weiss 1992). Studies in the human brain also found the presence of these cells (Eriksson et al. 1998). While some authors defend that NPCs are present almost ubiquitously throughout the adult brain (Arsenijevic et al. 2001; Nunes et al. 2003; Richardson et al. 2006; Sanai et al. 2004), most studies report their presence in specific neurogenic sites (Kukekov et al. 1999; Leonard et al. 2009; Sanai et al. 2004), the subventricular zone (SVZ) and the subgranular zone (SGZ) in the hippocampus (Eriksson et al. 1998; Zhao et al. 2008). It is possible that, since fibroblast-like cells (FbC) are phenotypically very similar to NPCs, these (FbC) are the actual mitotic active cells found in non-neurogenic sites (Park et al. 2012). Under physiological conditions the NPCs from the SVZ and the SGZ proliferate and migrate, respectively, to the olfactory bulb and the dentate gyrus

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and generate new neurons and astrocytes (Curtis et al. 2007; Doetsch and Alvarez-Buylla 1996; Lois et al. 1996). Contrary to what was initially thought, neurogenesis is a constant occurrence in these brain regions, and a large amount of experimental studies have shown that it may be altered in many neurodegenerative diseases (Lamm et al. 2014), as a possible consequence of brain aging (Bolognin et al. 2014), in stress-induced situations, or in psychiatric disorders such as depression (Conti et al. 2003; DeCarolis and Eisch 2010; Dwivedi 2009; Eisch et al. 2008; Kempermann et al. 2008; Kuhn et al. 2001; Steiner et al. 2006). While it was initially suggested that neurodegenerative diseases are confluent with a reduced precursor cell viability (Lovell et al. 2006), it was demonstrated that by using short postmortem, freshly harvested and immediately used SVZ samples, there was no significant NPC functionality reduction in elderly humans with Parkinson's disease (PD), Alzheimer's disease, progressive supranuclear palsy disease, dementia with Lewy bodies and in normal elderly control cases (Leonard et al. 2009). In fact, there are evidences that suggest that some neurodegenerative disorders such as traumatic brain injury (TBI) may even slightly stimulate neurogenesis (Curtis et al. 2003; Levin 2003; Macas et al. 2006; Richardson et al. 2007). In other words this means that even with the occurrence of such disorders, the brain still retains the capacity to generate fully functional NSCs/NPCs, which can be pharmacologically enhanced.

This knowledge has unveiled a new world of possibilities for the treatment of otherwise incurable neurodegenerative disorders. The current available therapies slow down the degenerative process but still leave the brain with irreversible damage and result in insufficient or no functional recovery. By stimulating neuron development we might be able not only to stop the progression of a disease but also to reverse its deleterious effects and therefore restore a normal cerebral function.

## Neural Stem Cell Transplantation

Ever since the discovery of human embryonic stem cells (hESC) in 1998, a heavy burden of expectation has been placed upon the use of undifferentiated cells for regenerative medicine. These cells, which can be obtained from fetal precursor cells, embryonic tissues or through the reprogramming of an individual's own somatic cells, possess an unlimited self-renewal capacity which allows them to differentiate into any cell type suitable for the treatment of an unlimited variety of diseases (Ormerod et al. 2008). Cell replacement therapy by delivering specific cells to the brain can be an effective approach to treat several brain pathologies that are affected by loss of neural cells, or

neural cell function as is the case of most neurodegenerative and neuropsychiatric disorders.

## Fetal Neuroprecursors

Stem cell transplantation can be made primarily from fetal neuroprecursors, which have been shown to survive many years after transplantation (Mendez et al. 2005), differentiate into mature neurons (Uchida et al. 2005) and reverse some neurological deficits accompanying PD (Freed et al. 2001; Mendez et al. 2005), Huntington's disease (HD) (Capetian et al. 2009), spinal cord injury (Bohl et al. 2008), stroke (Bliss et al. 2006) and brain injury (Gaillard et al. 2007). These neuroprecursors of fetal origin can be obtained from the bone marrow, umbilical cord blood or fetal brain. However, the difficulties in obtaining these cells for scientific research, the ethical issues associated with it and most importantly the histocompatibility concerns have limited their broader use in the clinical field. As a safer alternative, the reprogramming of an individual's own somatic cells back to the pluripotent state by using oocytes has also been suggested (Tachibana et al. 2013). However, the use of oocytes still raises ethical issues and this technology is rather inefficient (Noisa and Parnpai 2011).

## Embryonic Cells

Embryonic neuroprecursors have also been widely tested, and while their extremely high proliferative capacity can lead to a more efficient neurogenesis, it also hampers its use in humans due to the reported formation of malignant teratoma in 25 % of the implanted animal subjects and the inability to control their differentiation and proliferation (Bido et al. 2009; Hicks et al. 2009; Kim et al. 2002; Kim et al. 2008a, b; Maucksch et al. 2013). As an alternative, several researchers have attempted to commit the stem cells to a specific lineage prior to their transplantation, successfully converting them into specific neural cells: neural progenitor cells (Carpenter et al. 2001; Chaddah et al. 2012; Gerrard et al. 2005; Itsykson et al. 2005; Narkilahti et al. 2007; Piña-Crespo et al. 2012; Reubinoff et al. 2001; Trounson 2006; Zhang et al. 2001), neural crest cells (Lee et al. 2007a), dopaminergic neurons (Cho et al. 2008; Perrier et al. 2004), motor neurons (Erceg et al. 2010; Hester et al. 2011; Lee et al. 2007b; Zhang et al. 2011b), sensory neurons (Chen et al. 2012) and specific glial subtypes (Krencik et al. 2011). However, achieving an uniform differentiation of these cells *in vitro* still remains a challenge because of the many specific requirements in terms of growth factors or other small molecules for each cell subtype (Chen et al. 2012; Gerrard et al. 2005; Itsykson et al. 2005; Koch et al. 2009) and due to cluster formation

(Wichterle et al. 2002) that complicates the uniform diffusion of the substrate.

### Induced Pluripotent Stem Cells

The issues that arise from the use of the previous cells can be largely resolved by using induced pluripotent stem cells (iPSC). These are obtained by transfecting mature somatic cells, from an adult source, with retroviral vectors (Takahashi et al. 2007; Wernig et al. 2008). The somatic cells are obtained from the human donor who will also later be the recipient for the differentiated cells. This method largely mitigates the occurrence of immune reactions during transplantation (Kimbrel and Lanza 2015).

Current cell-based regenerative strategies and on-going clinical trials by NSC transplantation have so far generated extremely positive results, even considering the strict regulations imposed for their manipulation. The many promising applications of the use of iPSC and hESC include ocular disorders such as macular degeneration (Schwartz et al. 2012, 2014), diabetes (D'Amour et al. 2006; Kelly et al. 2011; Pagliuca et al. 2014; Schulz et al. 2012) and heart failure (Bellamy et al. 2014; Chong et al. 2014; Menasché et al. 2015). In the neurodegeneration field, hESC have been studied for the treatment of PD and have been shown to produce fully functional and transplantable dopaminergic neurons with a long-term survival and high capacity of integrating into the surrounding neuronal network (Doi et al. 2014; Grealish et al. 2014). New studies and ongoing clinical trials are, however, focused on surpassing some challenges still encountered in the cell transplantation field. Specifically, some “rate-limiting” steps, namely long-term survival of transplanted cells, cell targeting to injured tissues and cells’ successful integration in the host’s organism, may delay the progress in this field. For brain delivery specifically, the successful incorporation of the transplanted stem cells in the local stem cell niche plays a decisive role in cell viability and functionality (Lepski 2012), and the interaction between the new transfected cells and the local microenvironment is still not fully comprehended. Also, like in any surgical transplantation, local inflammation and immune reaction may occur, which might compromise survival and differentiation of the engrafted stem cells (Lepski 2012). Further research in this area will not only contribute to a greater availability of therapeutic strategies in regenerative medicine but it will also be crucial to gain a better understanding of the development of the brain and the intrinsic molecular interactions that lead to the generation of specific cell types. This could enable the discovery of new compounds with neurogenic properties.

### Neural Growth Factors

Given the limitations of cell replacement therapy which slow down its progress, different approaches have been explored. Several studies and human clinical trials have shown that the constituents of the microenvironments surrounding the stem cell niche and the intervening molecules in the neurogenesis process are just as fundamental as the presence of the NSCs. Probably the most important are the neurotrophic factors (NF) which are small proteins secreted by target tissues in the brain after being stimulated by the surrounding neural cells. They are capable of signaling the surrounding cells to differentiate and grow, but also of preventing them from initiating programmed cell death (Zhang et al. 2014). A large number of growth factors have been reported to possess neurotrophic properties. These include ciliary neurotrophic factor (CNTF), glial cell-line-derived neurotrophic factor (GDNF), insulin-like growth factor, fibroblast-like growth factor (FGF) and the major players in neurogenesis, the neurotrophins: nerve growth factor (NGF), brain-derived neurotrophic factor (BDNF), neurotrophin-3 (NT-3) and neurotrophin-4 (NT-4) (Jagasia et al. 2009; Weissmiller and Wu 2012; Yoo et al. 2007).

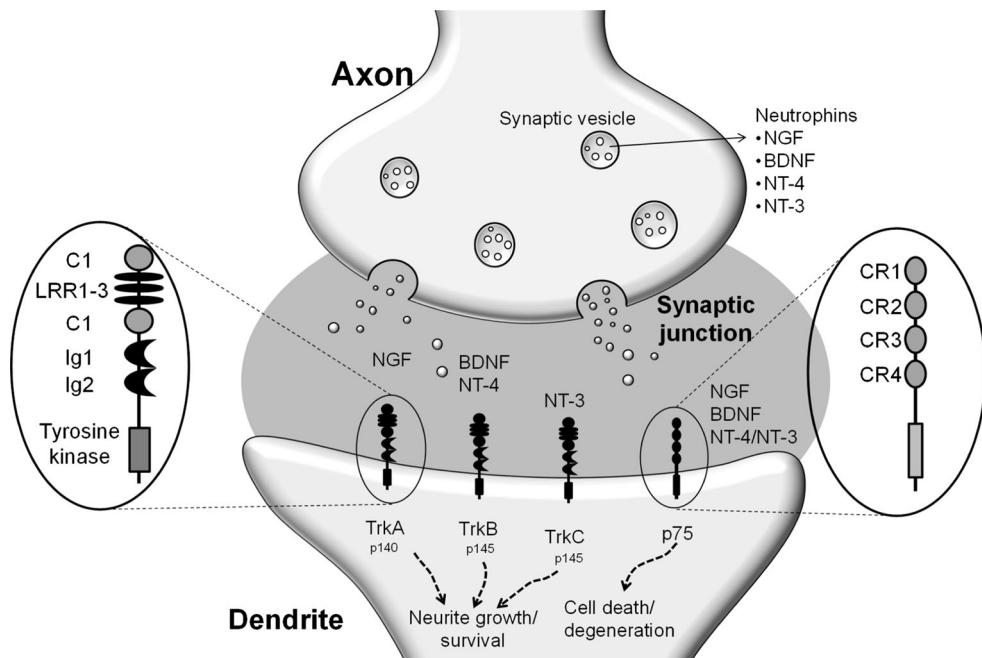
The neurotrophin family of the neurotrophic factors is primarily responsible for stimulating neural cell differentiation, growth and survival, as well as controlling the brain’s synaptic function and plasticity (Skaper 2012). They are produced under physiological conditions by originating neurons after receiving trophic input from their target neurotrophin-sensitive neurons with which they are connected (Thoenen 1995). Alternatively, the exposure of an autocrine or a non-target-derived paracrine neurotrophin has also been suggested. This goes for all neurotrophic factors, and while most are target- or paracrine-delivered, recent evidences have demonstrated that progranulin functions as an autocrine growth factor (Ahmed et al. 2007; Inestrosa and Arenas 2010). After release, neurotrophins are recognized by two specific receptor classes, Trks and p75<sup>NTR</sup>, which are expressed on the surface of the neurotrophin-sensitive neurons. The neurotrophins exhibit specific interactions with the Trk receptors. TrkA binds NGF (Kaplan et al. 1991; Klein et al. 1991a), TrkB binds mBDNF and NT-4 (Klein, et al. 1991b; Squinto et al. 1991) and TrkC binds NT-3 (Lamballe et al. 1991). Under specific conditions, NT-3 can also interact with TrkA and TrkB with less affinity. All neurotrophins, however, interact with and activate the p75 receptors (Bothwell 1995; Skaper 2012), which initiate an apoptotic pathway (Chao 2003). Specifically for BDNF, its precursor (proBDNF) binds primarily to p75 receptors, therefore initiating programmed cell death. When proteolytically processed due to

neuronal activity, proBDNF is converted to mature BDNF (mBDNF) which specifically binds to the TrkB receptors (Je et al. 2013) (Fig. 1). Neural survival will therefore depend on the level of neurotrophins secreted and the type of receptors expressed (Weissmiller and Wu 2012). In the CNS, due to the abundant expression of TrkB and TrkA (Weissmiller and Wu 2012) the most active neurotrophins are BDNF followed by NGF because of their high specificity for these receptors.

Given the crucial role of NFs in modulating neurogenesis in the adult brain, it is not surprising that the occurrence of most neurodegenerative or neuropsychiatric disorders is associated with an altered NF expression or their respective receptors, and several studies have successfully identified a specific NF-related cause and/or consequence for the occurrence of neurological disorders (Conti et al. 2003). GDNF and VEGF have been shown to promote the differentiation and survival of dopaminergic neurons (Eggert et al. 1999; Grandoso et al. 2007; Lindvall and Wahlberg 2008; Theofilopoulos et al. 2001; Zeng et al. 2006) and also exhibit a close correlation with motor neuron protection and reversal of degeneration (Drinkut et al. 2012; Eslamboli et al. 2005; Kells et al. 2010). In diseases associated with progressive motor impairment, like amyotrophic lateral sclerosis (ALS) and PD, these NFs are the most studied and effective in delaying the onset of neuronal and motor degeneration in rodents (Azzouz et al. 2004; Wang et al. 2002, 2007b), along with IGF (Dodge et al. 2008; Kaspar et al. 2003). NGF and BDNF have been primarily studied and mobilized toward the prevention and reversion of atrophy and loss of basal forebrain cholinergic

neurons which are highly associated with brain aging dementia (Fischer et al. 1987), AD (Bruno and Cuello 2006; Connor et al. 1997; Cuello and Bruno 2007; Peng et al. 2004) and neuronal degeneration and dementia in Down's Syndrome (Castrén and Tanila 2006; Dorsey et al. 2006; Fukuda et al. 2010). Likewise, BDNF has been successful in reducing or delaying the onset of dementia in patients with Rett Syndrome (RS) (Katz 2014). Mature BDNF, as the best-characterized of all neurotrophins in terms of its role in brain plasticity, has shown to be essential for differentiation and survival of dopaminergic, cholinergic, GABAergic and serotonergic neurons (Pillai 2008) and is thus closely involved in cognition and mood-related behaviors (Cowansage et al. 2010). A decreased BDNF expression has been associated with the clinical manifestations of HD (Cattaneo et al. 2005; Zuccato and Cattaneo 2007), AD (Zhang et al. 2012) and stroke, whereas a BDNF hyperactivity has been reported in epilepsy, autism and manic-depressive disorders (Tsai 2006). Furthermore, a reduced BDNF expression occurs in stressful situations and animal models lacking BDNF expression do not respond to antidepressant treatment (Adachi et al. 2008; Monteggia et al. 2004, 2007; Saarelaisten et al. 2003), which suggests a close relationship between their mechanisms of action. It has also been suggested that BDNF is involved in the respiratory control and rhythmogenesis generated and modulated in the brainstem, being therefore useful to stabilize the respiratory rhythm in respiratory disorders with a neurological cause, such as RS (Caravagna et al. 2013). FGF-2 plays a key role in angiogenesis, preventing and reducing neuronal damage

**Fig. 1** Neuron growth and survival mediation through extracellular activation of Trk receptors and cell death and degeneration through extracellular activation of p75 receptors (*CR1–CR4* cysteine-rich motifs, *C1/C2* cysteine-rich clusters, *LRR1–3* leucine-rich repeats, *Ig1/Ig2* immunoglobulin-like domains, *p140* 140,000 molecular weight protein, *p145* 145,000 molecular weight protein)



after cerebral infarction (Kawamata et al. 1997), focal ischemia (Ma et al. 2008) and seizures (Liu and Holmes 1997). It stimulates a cerebral plastic response, reduces inflammation and promotes neuronal survival (Uzun et al. 2010). Another NF, P021 which is a small neurotrophic compound derived from the CNTF activates endogenous neuroprotective mechanisms that could be used to treat and prevent cognitive aging (Bolognin et al. 2014).

Despite the therapeutic potential of these NF and their success in the laboratory, several drawbacks have hindered their broader use, namely their short biological half-life, rapid degradation rate and limited blood–brain barrier (BBB) permeability by oral or intravenous drug delivery (Cai et al. 2014; Herrán et al. 2014; Stankowski and Gupta 2011). All these factors result in an extremely low NF bioavailability in the brain, which is not sufficient to produce the required neurotrophic effects. On the contrary, if the amount of NF administered is too high, besides the occurrence of peripheral side effects, the overstimulation of their receptors and neuronal synapses may result in epileptic seizures (Claire Géral et al. 2013), or alternatively, as several evidences suggest, it could downregulate the expression of Trk receptors therefore inhibiting their neuroprotective effect altogether (Frank et al. 1996; Knusel et al. 1997; Sommerfeld et al. 2000). The unpredictability of the amount of NF that reaches the CNS is therefore the biggest challenge to be overcome. In addition, some diseases such as stroke have a relatively short timeframe in which these NF can be effective. In order to surpass the limitations encountered in conventional drug delivery, several surgical approaches have been proposed, such as intracerebral administrations of the NFs (Kawamata et al. 1997; Kells et al. 2010; Liu and Holmes 1997; Shirakura et al. 2004; Van Kampen et al. 2014; Wu 2005; Zhu et al. 2014), the implant of programmable pumps (Grondin et al. 2002, 2003; Yanamoto et al. 2000) and the administration of gene-carrying viral vectors (Björklund et al. 2000; Choi-Lundberg et al. 1997) which, although effective, are more invasive and complex. In addition, gene therapy is irreversible. Nanoparticle-based strategies have also been explored to improve BBB permeation and promote a targeted and sustained drug release in the brain (Herrán et al. 2014; Migliore et al. 2014; Yurek et al. 2009) (Table 1). Among the many advantages offered by the use of nanoparticles, some of the most important include their ability to protect the NFs from enzymatic degradation and other stressors (Claire Géral et al. 2013), their ability to evade the host's immune system when coated with hydrophilic polymers (e.g., polyethylene glycol) (Angelov et al. 2011; Kurakhmaeva et al. 2009) with minimal loss of biological activity (Sakane and Pardridge 1997), and the possibility to target specific areas while minimizing systemic side effects (Géral et al. 2013). Furthermore, there is the possibility to anchor the surface of the nanoparticles with specific ligands toward receptor recognition in the CNS (Pang et al. 2008; Xie

et al. 2005). Other more advanced strategies that have been exploited combine both of the above methods: sustained release of GDNF from encapsulated, genetically engineered fibroblasts (Sajadi et al. 2006) or genetically modified NSCs for the overexpression of NT-3 when implanted in rats (Lu et al. 2003).

While it is still difficult to deliver NFs to the brain, the wide knowledge of the molecular structure of both NFs, their receptors, and their interaction may prove useful in the design of new mimetic molecules with improved pharmacokinetic and pharmacodynamic characteristics (Webster and Pirrung 2008), as is the case of P021 which is a CNTF derivate (Bolognin et al. 2014).

## Proneurogenic Molecules

Several molecules have been proposed as potential neurogenesis promoters either due to their antioxidant and anti-inflammatory properties or more specifically due to their ability to modulate neural stem cell differentiation and proliferation or apoptotic pathways. Among them, glutamate has been demonstrated to play a key role in neurogenesis in association with neurotrophic factors. It stimulates BDNF expression, which in turn modifies neuron glutamate sensitivity (Mattson 2008). However, overactivation of glutamate receptors may result in neuronal degeneration (Mattson 2008), which leads to a necessity for optimized glutamatergic signaling. Galanin, a widely expressed neuropeptide, has also been shown to exhibit an intense neuroprotective and neuroproliferative potential by modulating noradrenergic, serotonergic and cholinergic neurotransmission and by protecting neurons from excessive glutamate and amyloid  $\beta$  cytotoxicity (Agasse et al. 2013). However, retinoic acid, curcumin, allopregnanolone and antidepressants have received the widest attention due to their positive and very promising effects on several distinct neurogenesis pathways. A list of some of the most promising results achieved with the use of several classes of proneurogenic molecules can be seen on Table 2.

### Retinoic Acid (Vitamin A)

Retinoids are a class of molecules, produced in the olfactory epithelium in the adult SVZ and hippocampus (Calza et al. 2003; Haskell and LaMantia 2005; Jacobs et al. 2006) from retinaldehyde with the enzyme retinaldehyde dehydrogenase 3 (RALDH3), which are involved in several distinct stem cell differentiation pathways (Paschaki et al. 2013) (Fig. 2). They modulate glucocorticosteroids (GCs) availability in the brain, which are released to cope with stress and are directly related to brain aging, cognitive decline, loss of brain plasticity (Cameron and Gould 1994; Suri and

**Table 1** Nanosystems for neurotrophic factor delivery

Nanosystem	Disease/model	References
<b>BDNF</b>		
Nanoporous PGA particles	Deafness/in vitro (guinea pig)	Tan et al. (2012)
Layer-by-layer films on agarose hydrogel scaffolds	Spinal cord injury/in vitro	Mehrotra et al. (2010)
Neurotensin-polyplex nanocarrier	PD/in vitro (rat)	Martinez-Fong et al. (2012)
PEGylated cationic lipid nanoparticles	in vitro	Angelov et al. (2011)
Cubosome nanoparticles with omega-3	in vitro	Géral et al. (2012)
PLGA nanoparticles	Cauda equina syndrome/in vivo (dog)	Tan et al. (2013)
<b>GDNF</b>		
PLGA nanospheres	PD/in vivo (rat)	Herrán et al. (2014)
Neurotensin-polyplex nanocarrier	PD/in vitro (rat)	Martinez-Fong et al. (2012)
Trojan horse nanocarriers with brain-specific promoters	PD/in vivo (rat and rhesus monkey)	Boado and Pardridge (2011), Xia et al. (2008)
Cationic liposomes	Spinal cord injury/in vitro	Martina et al. (2008), Yang et al. (1997)
	PD/in vivo (rat)	Migliore et al. (2014)
<b>VEGF</b>		
PLGA nanospheres	AD/in vitro, in vivo (mice)	Herrán et al. (2013)
	PD/in vivo (rat)	Herrán et al. (2014)
Iron-oxide nanoparticles with monoclonal antibodies	Brain tumor/in vivo (rat)	Abakumov et al. (2012)
<b>NGF</b>		
PEGylated—PBCA nanoparticles	PD/in vivo (rat)	Kurakhmaeva et al. (2009)
Targeted liposomes	AD/in vivo	(Xie et al. 2005)
Cationic liposomes	Spinal cord injury/in vitro	Martina et al. (2008), Yang et al. (1997)
<b>NT-3</b>		
PBCA nanoparticles	Hypertensive intracerebral hemorrhage/in vivo(rat)	Chung et al. (2013)
<b>FGF-2</b>		
Organically modified silica nanoplexes with r-FGF	Neurodegenerative disorders/in vitro	Stachowiak et al. (2009)

PGA poly-L-glutamic acid, PBCA poly-butyl cyanoacrylate

Vaidya 2013) and even contribute to the development of depressive symptoms (Christoph Anacker et al. 2013). In addition, it has been suggested that GCs alter BDNF expression and signaling, therefore reducing the brain's neurogenesis potential (Suri and Vaidya 2013). Thus, the modulation of GCs availability by retinoids is an important biological mechanism that can be explored in many stress-related pathologies to prevent plasticity alterations in the hippocampus (Bonhomme et al. 2014). Retinoids are also known to alleviate microglial chronic activation which occurs after brain trauma or inflammation and is associated with a progressive neurodegeneration in AD and PD and in a number of other CNS pathologies (Brossaud et al. 2013; Rangarajan et al. 2013). In addition, retinoic acid (RA) is also involved in the Wnt pathway, being therefore essential for hippocampal neurogenesis (Elizalde et al. 2011).

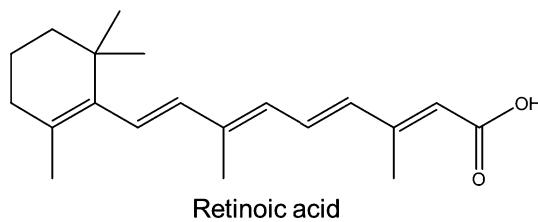
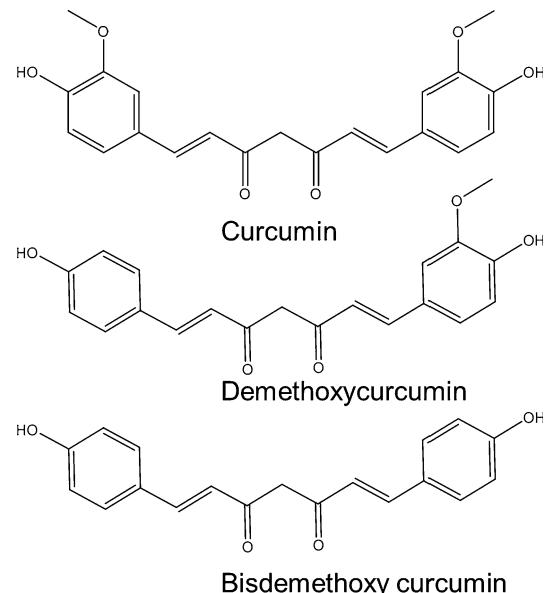
All-trans retinoic acid (ATRA), which is the active form of RA, has proven to be capable of inducing neuronal

outgrowth and differentiation from embryonic stem cells (Lu et al. 2009; Stavridis et al. 2010; Yurek et al. 2009), NSCs (Takahashi et al. 1999; Wang et al. 2005a, b), dorsal root ganglia (Corcoran and Maden 1999) or mouse teratocarcinoma (Hádinger et al. 2009). It can be especially effective for the regeneration of neuronal cells after peripheral or CNS injury due to the increased RALDH3 expression in these situations (Santos et al. 2012).

However, ATRA is rapidly metabolized by cells, has low aqueous solubility and requires a fine-tuning of the concentration window to achieve efficacy, posing difficulties in the delivery of therapeutic doses (Santos et al. 2012). In this regard, nanoparticle formulations seem to be the most effective in promoting a prolonged ATRA release in the SVZ, and these include the use of polyelectrolyte nanoparticles (Maia et al. 2011; Santos et al. 2012), polyethyleneimine complex nanoparticles (Ku et al. 2013) and polymer-oil nanostructured carriers (Narvekar et al. 2012).

**Table 2** List of some of the most relevant mechanisms of action of several proneurogenic molecules

Proneurogenic Molecule	Mechanism of action	References
All-trans retinoic acid	Reduction in oxidative stress	Nair et al. (2015)
	Promotion of NGF release	Palencia et al. (2014)
	Downregulation of NMDAR and KLF11 expression	Nair et al. (2015)
	Reduction in glucocorticosteroid expression	Christoph Anacker et al. (2013), Bonhomme et al. (2014), Suri and Vaidya (2013)
	Reduction in microglial chronic activation	(Brossaud et al. 2013; Rangarajan et al. 2013)
	Activation of the canonical Wnt/β-catenin pathway	(Elizalde et al. 2011)
	Reduction in proinflammatory and oxidative responses	Mansouri et al. (2012)
	Reduction in amyloid-B aggregates	Garcia-Alloza et al. (2007), Thapa et al. (2013)
	Reduction in kainic acid-induced seizures	Sumanont et al. (2007)
	Inhibition of homocysteine production	Mansouri et al. (2012)
Curcuminoids	Inhibition of the Cu(II)-induced oxidative damage	Huang et al. (2011)
	Activation of the canonical Wnt/β-catenin pathway	Tiwari et al. (2014)
	Promotion of BDNF release in the hippocampus	Liu et al. (2014)
	Regulation of GABAergic and glutamatergic neurons and pro-inflammatory genes	He et al. (2004), Roof et al. (1997)
	Activation of microglial cells	Pettus et al. (2005)
	Increased BDNF expression	Gonzalez et al. (2005)
	Increased activation of serotonergic receptors	Bristot et al. (2014), Marx et al. (2003)
	20–40 % increase in proliferative cells in the dentate gyrus and up-regulation of BDNF expression	Malberg et al. (2000)
	Increased neural progenitor cells proliferation in the dentate gyrus	McAvoy et al. (2015)
Neurosteroids (progesterone, alopregnanolone)		
Antidepressants		
Fluoxetine, tranylcypromine and reboxetine		
Fluoxetine		

**Fig. 2** Retinoic acid chemical structure

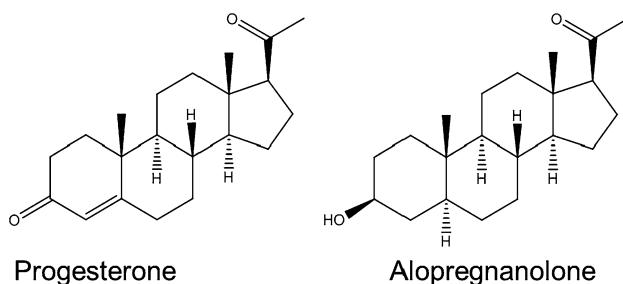
## Curcumin

Curcumin is a highly pleiotropic polyphenolic molecule with anti-inflammatory, anti-amyloid, anti-apoptotic and antioxidant properties (Mansouri et al. 2012), which can be extracted from turmeric (*Curcuma longa*) (Kim et al. 2008a, b) (Fig. 3). It is involved in hippocampal neurogenesis and can enhance neural plasticity and repair (Kim et al. 2008a, b) through activation of the canonical Wnt/β-catenin pathway (Tiwari et al. 2014). In addition, due to its

**Fig. 3** Curcumin and its main analogs' chemical structure

potent anti-inflammatory potential, curcumin can also prevent neurodegeneration by inhibiting proinflammatory responses in the injured, aged or diseased brain (Wang et al. 2005a, b). It has been shown to slow the progression of AD by reducing amyloid  $\beta$  in the brain (Garcia-Alloza et al. 2007; Thapa et al. 2013; Yang et al. 2005). Moreover, curcumin reduces the incidence of kainic acid-induced seizures (Sumanont et al. 2007) and promotes neuroprotection in PD by reducing homocysteine production which is a risk factor for vascular disease and brain atrophy (Mansouri et al. 2012), by inhibiting the Cu(II)-induced oxidative damage (Huang et al. 2011) and by inhibiting the inflammatory glial response and reducing superoxide dismutase function in the brain (Tripanichkul and Jaroensupparerch 2013). More recently, it has also been suggested that curcumin exhibits an antidepressant-like activity by promoting BDNF release in the hippocampus (Liu et al. 2014) and ameliorates mitochondrial dysfunctions in HD (Sandhir et al. 2014). Curcumin analogs such as demethoxycurcumin and bisdemethoxycurcumin, which can often be found in curcuminoid commercial powders, have also been reported to possess considerable antioxidant, anti-inflammatory and anti-proliferative activities (Kim et al. 2001; Sandur et al. 2007; Simon et al. 1998) (Fig. 4).

The neuroprotective efficacy of curcumin is, however, limited by its low brain bioavailability due to poor absorption, rapid metabolism and systemic elimination, and limited blood brain barrier (BBB) permeability (Yang et al. 2007). Therefore, the production of curcumin-loaded nanoparticles for brain delivery has been increasingly reported, namely the use of liposomes and phospholipid complexes (Tsai et al. 2011), biodegradable poly(lactic-co-glycolic acid) (PLGA) nanoparticles (Tiwari et al. 2014; Tsai et al. 2011) and liposomes (Chiu et al. 2013), which allow for a superior bioavailability and BBB permeability, reduced elimination and a slow and sustained curcumin release in the SVZ.



**Fig. 4** Progesterone and pregnanolone's chemical structure

## Neurosteroids

Neurosteroids, which are steroids produced in the brain, regulate physiological functions in the CNS. Among them, pregnane neurosteroids, such as progesterone (PROG) and its metabolite alopregnanolone, (AP) have been receiving increasing attention due to their neuroprotective effects in several neurodegenerative and neuropsychiatric disorders (Fig. 4). PROG and AP regulate both GABAergic and glutamatergic receptors and are also known to prevent lipid peroxidation, the generation of isoprostanes (Roof et al. 1997), the expression of pro-inflammatory genes (He et al. 2004), the activation of microglial cells (Pettus et al. 2005), and of regulating BDNF expression (Gonzalez et al. 2005). PROG has also been reported to regulate the expression of anti-apoptotic proteins, such as Bcl-2 (Bristot et al. 2014). Several in vitro and in vivo preclinical studies have demonstrated AP's high potency and efficacy in promoting neurogenesis in aged brain and in the neuronal restoration after TBI (He et al. 2004), AD (Brinton and Wang 2006a, b) and posttraumatic stress disorder (Pibiri et al. 2008). Studies in humans have primarily shown a correlation between the downregulation of AP biosynthesis and the occurrence of psychiatric symptoms ranging from anxiety and depression (Uzunova et al. 1998), aggressiveness (Bristot et al. 2014), schizophrenia (Marx et al. 2003) and bipolarity (Johansson et al. 2011) due to their effect on serotonergic receptors. Additionally, even though AP has the advantage of being a small hydrophobic BBB penetrating molecule (Brinton and Wang 2006a), it is quickly inactivated by glucuronidation, sulfate conjugation and oxidation (Bristot et al. 2014).

## Antidepressants

The potential influence of psychoactive drugs in brain neurogenesis is a very widely studied area, and while it has been demonstrated that opioids and alcohol negatively influence neurogenesis (Canales 2007; Crews and Nixon 2003; Eisch et al. 2000), antidepressants (ADPs), atypical antipsychotics and mood modulators have been shown to positively impact neurogenesis (Duman et al. 2001a, b; Harwood 2003; Malberg 2004; Wakade et al. 2002). There is evidence which suggests that stress-induced neurobiological disorders are highly associated with a decreased production of NFs or abnormalities in their receptors (Dwivedi 2009), hippocampal morphological alterations including atrophy and loss of CA3 pyramidal neurons (McEwen 1999). While these are not the sole cause for the occurrence of these disorders, they are likely major contributors. Therefore, it has been demonstrated that ADPs possess neuroprotective and neuroregenerative effects by mediating the expression of neurotrophic and protective factors such as bcl-2, BDNF and NGF (Croce et al. 2014), and by regulating the glucocorticoid receptor

function (Anacker et al. 2011). Some ADPs also increase AP expression in rodents and the levels of neurosteroids in depression and panic disorders (Bristot et al. 2014). Some authors report, however, that ADPs such as the widely studied fluoxetine (serotonin-reuptake inhibitor) have a small influence on hippocampal neurogenesis when compared to increased exercise in mice, and in some cases, such as for duloxetine (dual serotonergic-noradrenergic reuptake inhibitor) show no increased neuron regeneration or survival (Hanson et al. 2011; Marlatt et al. 2010). Although the exact mechanism of action of ADPs in neuroprotective and neuroregenerative processes is still not fully comprehended, therefore eliciting conflicting opinions on the subject, it is common knowledge that due to their ability to revert neural impairment either by reducing stress or by actively promoting neuron generation and proliferation, these molecules are very promising in the regenerative medicine field.

## Hyperbaric Oxygen Therapy

Hyperbaric oxygen therapy (HBOT) refers to the use of oxygen as a therapeutically active substance, at a superior level than the atmospheric pressure (Mu et al. 2011). The temporary elevated partial oxygen pressure in the body leads to a superior erythrocyte transporting capacity and tissue regenerating processes. Therefore, it was initially used for the treatment of diabetic foot, crush injuries, skin grafts, thermal burns and several neurological diseases (Mychaskiw 2010). However, since neurons are highly energy-demanding and require a constant oxygen supply (Mu et al. 2011), several studies have supported HBOT's further applicability for the treatment and prevention of neurodegenerative disorders (Godman et al. 2010; Günther et al. 2005; Wang et al. 2009; Zhang et al. 2010, 2011a). Although there is yet a lot to be learned about its underlying mechanisms of action, HBOT's neuroprotection and neurogenic potential have been correlated with the activation of several signaling pathways and transcription factors, such as the Wnt, hypoxia-inducible factors (HIF) and cAMP response element-binding (CREB). HBOT's neuroprotective potential by the Wnt pathway is thought to be regulated by decreasing the negative influence of  $\beta$ -catenin in cell proliferation (Wang et al. 2007a; Zhang et al. 2011a). Similarly, inhibition of HIF through HBOT also triggers  $\beta$ -catenin's reduced expression (Mazumdar et al. 2010; Milosevic et al. 2009). It has also been suggested that HBOT blocks CREB's degradation (Mu et al. 2011), which is involved in the up-regulation of BDNF, VEGF and Bcl-2 genes (Zhu et al. 2004). Therefore, HBOT, in addition to increasing oxygen supply to the brain, also promotes NSC proliferation and differentiation into neurons and oligodendrocytes, which makes it useful for the treatment of

neurodegenerative conditions such as stroke (Lee et al. 2013; Shin et al. 2008; Yin and Zhang 2005; Yin et al. 2003) and TBI (Kernie and Parent 2010; Rockswold et al. 2007). Certain physiological abnormalities including cerebral hypoperfusion, inflammation, mitochondrial dysfunction and oxidative stress that occur in autistic individuals and lead to neurodegeneration may also be ameliorated by HBOT therapy (Jepson et al. 2011; Rossignol et al. 2009; Sampanthavivat et al. 2012). HBOT's role in other conditions has also been studied, namely in AD by reducing cell toxicity and oxidative stress (Tian et al. 2012) and cerebral palsy by improving brain oxygenation and ameliorating the motor function (Mukherjee et al. 2014).

## Conclusion

Current advances in the field of neurobiology are allowing a better understanding of the role of NPCs in brain renewal after traumatic or ischemic brain injury and in several neurodegenerative and neuropsychiatric disorders, such as Alzheimer's disease, PD, Huntington's disease, autism, bipolar disease, among many others. The most promising approaches for stimulating neurogenesis include the transplantation of NSCs programmed toward a specific cellular line, the administration of neural growth factors, namely neurotrophic factors, the administration of proneurogenic molecules and the use of hyperbaric oxygen therapy. Although stem cell therapy has been on the verge of incredible breakthrough, therapy with proneurogenic molecules due to fewer limitations for its use will most likely evolve more rapidly. It is also possible to ally this strategy with hyperbaric oxygen therapy which would potentiate the results. There are still several significant setbacks to be overcome in order for a successful treatment of these debilitating disorders to be achieved, which means that a lot remains undiscovered. However, considerable progress has been made, and these advances will lead to new technologies and innovative therapies which will undoubtedly make a huge change in society.

## Compliance with Ethical Standards

**Conflict of interest** The authors report no conflicts of interest and have no proprietary or commercial interests in any concept or product discussed in this paper.

## References

- Abakumov, M. A., Shein, S. A., Vishvasrao, H., Nukolova, N. V., Sokol'ski-Papkov, M., Sandalova, T. O., et al. (2012). Visualization of experimental glioma C6 by MRI with magnetic nanoparticles conjugated with monoclonal antibodies to vascular endothelial growth factor. *Bulletin of experimental biology and*

- medicine*, 154(2), 274–277. <http://www.ncbi.nlm.nih.gov/pubmed/23330142>. Accessed September 21, 2014.
- Adachi, M., Barrot, M., Autry, A. E., Theobald, D., & Monteggia, L. M. (2008). Selective loss of brain-derived neurotrophic factor in the dentate gyrus attenuates antidepressant efficacy. *Biological Psychiatry*, 63(7), 642–649. doi:10.1016/j.biopsych.2007.09.019.
- Agasse, F., Xapelli, S., Coronas, V., Christiansen, S. H., Rosa, A. I., Sardá-Arroyo, L., et al. (2013). Galanin promotes neuronal differentiation in murine subventricular zone cell cultures. *Stem Cells and Development*, 22(11), 1693–1708. doi:10.1089/scd.2012.0161.
- Ahmed, Z., Mackenzie, I. R. A., Hutton, M. L., & Dickson, D. W. (2007). Progranulin in frontotemporal lobar degeneration and neuroinflammation. *Journal of Neuroinflammation*, 4, 7. doi:10.1186/1742-2094-4-7.
- Altman, J. (1962). Are new neurons formed in the brains of adult mammals? *Science (New York, N.Y.)*, 135(3509), 1127–1128. <http://www.ncbi.nlm.nih.gov/pubmed/13860748>. Accessed June 15, 2014.
- Altman, J., & Das, G. D. (1965). Post-natal origin of microneurones in the rat brain. *Nature*, 207(5000), 953–956. <http://www.ncbi.nlm.nih.gov/pubmed/5886931>. Accessed June 3, 2014.
- Anacker, C., Zunszain, P. A., Cattaneo, A., Carvalho, L. A., Garabedian, M. J., Thuret, S., et al. (2011). Antidepressants increase human hippocampal neurogenesis by activating the glucocorticoid receptor. *Molecular psychiatry*, 16(7), 738–750. doi:10.1038/mp.2011.26.
- Anacker, C., Cattaneo, A., Luoni, A., Musaelyan, K., Zunszain, P. A., Milanesi, E., et al. (2013). Glucocorticoid-related molecular signaling pathways regulating hippocampal neurogenesis. *Neuropsychopharmacology: Official Publication of the American College of Neuropsychopharmacology*, 38(5), 872–883. doi:10.1038/npp.2012.253.
- Angelov, B., Angelova, A., Filippov, S. K., Karlsson, G., Terrill, N., Lesieur, S., & Štěpánek, P. (2011). Topology and internal structure of PEGylated lipid nanocarriers for neuronal transfection: synchrotron radiation SAXS and cryo-TEM studies. *Soft Matter*, 7(20), 9714. doi:10.1039/c1sm06447a.
- Arsenijevic, Y., Villemure, J. G., Brunet, J. F., Bloch, J. J., Déglon, N., Kostic, C., et al. (2001). Isolation of multipotent neural precursors residing in the cortex of the adult human brain. *Experimental Neurology*, 170(1), 48–62. doi:10.1006/exnr.2001.7691.
- Azzouz, M., Ralph, G. S., Storkebaum, E., Walmsley, L. E., Mitrophanos, K. A., Kingsman, S. M., et al. (2004). VEGF delivery with retrogradely transported lentivector prolongs survival in a mouse ALS model. *Nature*, 429(6990), 413–417. doi:10.1038/nature02544.
- Bellamy, V., Vanneaux, V., Bel, A., Nemetalla, H., Emmanuelle Boitard, S., Farouz, Y., et al. (2014). Long-term functional benefits of human embryonic stem cell-derived cardiac progenitors embedded into a fibrin scaffold. *The Journal of Heart and Lung Transplantation: The Official Publication of the International Society for Heart Transplantation*, . doi:10.1016/j.healun.2014.10.008.
- Björklund, A., Kirik, D., Rosenblad, C., Georgievska, B., Lundberg, C., & Mandel, R. J. (2000). Towards a neuroprotective gene therapy for Parkinson's disease: Use of adenovirus, AAV and lentivirus vectors for gene transfer of GDNF to the nigrostriatal system in the rat Parkinson model. *Brain research*, 886(1–2), 82–98. <http://www.ncbi.nlm.nih.gov/pubmed/11119690>. Accessed June 27, 2014.
- Bliss, T. M., Kelly, S., Shah, A. K., Foo, W. C., Kohli, P., Stokes, C., et al. (2006). Transplantation of hNT neurons into the ischemic cortex: Cell survival and effect on sensorimotor behavior. *Journal of Neuroscience Research*, 83(6), 1004–1014. doi:10.1002/jnr.20800.
- Boado, R. J., & Pardridge, W. M. (2011). The Trojan horse liposome technology for nonviral gene transfer across the blood-brain barrier. *Journal of Drug Delivery*, 2011, 296151. doi:10.1155/2011/296151.
- Bohl, D., Liu, S., Blanchard, S., Hocquemiller, M., Haase, G., & Heard, J.-M. (2008). Directed evolution of motor neurons from genetically engineered neural precursors. *Stem Cells (Dayton, Ohio)*, 26(10), 2564–2575. doi:10.1634/stemcells.2008-0371.
- Boido, M., Rupa, R., Garbossa, D., Fontanella, M., Ducati, A., & Vercelli, A. (2009). Embryonic and adult stem cells promote raphespinal axon outgrowth and improve functional outcome following spinal hemisection in mice. *The European journal of neuroscience*, 30(5), 833–846. doi:10.1111/j.1460-9568.2009.06879.x.
- Bolognin, S., Buffelli, M., Puoliväli, J., & Iqbal, K. (2014). Rescue of cognitive-aging by administration of a neurogenic and/or neurotrophic compound. *Neurobiology of Aging*, . doi:10.1016/j.neurobiolaging.2014.02.017.
- Bonhomme, D., Minni, A. M., Alfos, S., Roux, P., Richard, E., Higueret, P., et al. (2014). Vitamin A status regulates glucocorticoid availability in Wistar rats: Consequences on cognitive functions and hippocampal neurogenesis? *Frontiers in Behavioral Neuroscience*, 8, 20. doi:10.3389/fnbeh.2014.00020.
- Bothwell, M. (1995). Functional interactions of neurotrophins and neurotrophin receptors. *Annual Review of Neuroscience*, 18, 223–253. doi:10.1146/annurev.ne.18.030195.001255.
- Brinton, R. D., & Wang, J. M. (2006a). Therapeutic potential of neurogenesis for prevention and recovery from Alzheimer's disease: Allopregnanolone as a proof of concept neurogenic agent. *Current Alzheimer Research*, 3(3), 185–190. <http://www.ncbi.nlm.nih.gov/pubmed/16842093>. Accessed June 5, 2014.
- Brinton, R. D., & Wang, J. M. (2006b). Preclinical analyses of the therapeutic potential of allopregnanolone to promote neurogenesis in vitro and in vivo in transgenic mouse model of Alzheimer's disease. *Current Alzheimer Research*, 3(1), 11–17. <http://www.ncbi.nlm.nih.gov/pubmed/16472197>. Accessed July 15, 2014.
- Bristot, G., Ascoli, B., Gubert, C., Panizzutti, B., Kapczinski, F., & Rosa, A. R. (2014). Progesterone and its metabolites as therapeutic targets in psychiatric disorders. *Expert Opinion on Therapeutic Targets*, 18(6), 679–690. doi:10.1517/14728222.2014.897329.
- Brossaud, J., Roumes, H., Moisan, M.-P., Pallet, V., Redonnet, A., & Corcuff, J.-B. (2013). Retinoids and glucocorticoids target common genes in hippocampal HT22 cells. *Journal of Neurochemistry*, 125(4), 518–531. doi:10.1111/jnc.12192.
- Bruno, M. A., & Cuello, A. C. (2006). Activity-dependent release of precursor nerve growth factor, conversion to mature nerve growth factor, and its degradation by a protease cascade. *Proceedings of the National Academy of Sciences of the United States of America*, 103(17), 6735–6740. doi:10.1073/pnas.0510645103.
- Cai, J., Hua, F., Yuan, L., Tang, W., Lu, J., Yu, S., et al. (2014). Potential therapeutic effects of neurotrophins for acute and chronic neurological diseases. *BioMed Research International*, 2014, 601084. doi:10.1155/2014/601084.
- Calza, L., Giuliani, A., Fernandez, M., Pirondi, S., D'Intino, G., Aloe, L., & Giardino, L. (2003). Neural stem cells and cholinergic neurons: regulation by immunolesion and treatment with mitogens, retinoic acid, and nerve growth factor. *Proceedings of the National Academy of Sciences of the United States of America*, 100(12), 7325–7330. doi:10.1073/pnas.1132092100.
- Cameron, H. A., & Gould, E. (1994). Adult neurogenesis is regulated by adrenal steroids in the dentate gyrus. *Neuroscience*, 61(2), 203–209. <http://www.ncbi.nlm.nih.gov/pubmed/7969902>. Accessed June 17, 2014.

- Canales, J. J. (2007). Adult neurogenesis and the memories of drug addiction. *European Archives of Psychiatry and Clinical Neuroscience*, 257(5), 261–270. doi:10.1007/s00406-007-0730-6.
- Capetian, P., Knoth, R., Maciaczyk, J., Pantazis, G., Ditter, M., Bokla, L., et al. (2009). Histological findings on fetal striatal grafts in a Huntington's disease patient early after transplantation. *Neuroscience*, 160(3), 661–675. doi:10.1016/j.neuroscience.2009.02.035.
- Caravagna, C., Soliz, J., & Seaborn, T. (2013). Brain-derived neurotrophic factor interacts with astrocytes and neurons to control respiration. *The European Journal of Neuroscience*, 38(9), 3261–3269. doi:10.1111/ejn.12320.
- Carpenter, M. K., Inokuma, M. S., Denham, J., Mujtaba, T., Chiu, C. P., & Rao, M. S. (2001). Enrichment of neurons and neural precursors from human embryonic stem cells. *Experimental Neurology*, 172(2), 383–397. doi:10.1006/exnr.2001.7832.
- Castrén, E., & Tanila, H. (2006). Neurotrophins and dementia—keeping in touch. *Neuron*, 51(1), 1–3. doi:10.1016/j.neuron.2006.06.019.
- Cattaneo, E., Zuccato, C., & Tartari, M. (2005). Normal huntingtin function: An alternative approach to Huntington's disease. *Nature Reviews Neuroscience*, 6(12), 919–930. doi:10.1038/nrn1806.
- Chaddah, R., Arntfield, M., Runciman, S., Clarke, L., & van der Kooy, D. (2012). Clonal neural stem cells from human embryonic stem cell colonies. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, 32(23), 7771–7781. doi:10.1523/JNEUROSCI.3286-11.2012.
- Chao, M. V. (2003). Neurotrophins and their receptors: a convergence point for many signalling pathways. *Nature Reviews Neuroscience*, 4(4), 299–309. doi:10.1038/nrn1078.
- Chen, W., Jongkamoniwat, N., Abbas, L., Eshtan, S. J., Johnson, S. L., Kuhn, S., et al. (2012). Restoration of auditory evoked responses by human ES-cell-derived otic progenitors. *Nature*, 490(7419), 278–282. doi:10.1038/nature11415.
- Chiu, S., Terpstra, K. J., Bureau, Y., Hou, J., Raheb, H., Cernovsky, Z., et al. (2013). Liposomal-formulated curcumin [Lipocurc™] targeting HDAC (histone deacetylase) prevents apoptosis and improves motor deficits in Park 7 (DJ-1)-knockout rat model of Parkinson's disease: implications for epigenetics-based nanotechnology-driven drug platform. *Journal of Complementary & Integrative Medicine*,. doi:10.1515/jcim-2013-0020.
- Cho, M. S., Lee, Y.-E., Kim, J. Y., Chung, S., Cho, Y. H., Kim, D.-S., et al. (2008). Highly efficient and large-scale generation of functional dopamine neurons from human embryonic stem cells. *Proceedings of the National Academy of Sciences of the United States of America*, 105(9), 3392–3397. doi:10.1073/pnas.0712359105.
- Choi-Lundberg, D. L., Lin, Q., Chang, Y. N., Chiang, Y. L., Hay, C. M., Mohajeri, H., et al. (1997). Dopaminergic neurons protected from degeneration by GDNF gene therapy. *Science (New York, N.Y.)*, 275(5301), 838–841. <http://www.ncbi.nlm.nih.gov/pubmed/9012352>. Accessed June 5, 2014.
- Chong, J. J. H., Yang, X., Don, C. W., Minami, E., Liu, Y.-W., Weyers, J. J., et al. (2014). Human embryonic-stem-cell-derived cardiomyocytes regenerate non-human primate hearts. *Nature*, 510(7504), 273–277. doi:10.1038/nature13233.
- Chung, C.-Y., Yang, J.-T., & Kuo, Y.-C. (2013). Polybutylcyanoacrylate nanoparticles for delivering hormone response element-conjugated neurotrophin-3 to the brain of intracerebral hemorrhagic rats. *Biomaterials*, 34(37), 9717–9727. doi:10.1016/j.biomaterials.2013.08.083.
- Connor, B., Young, D., Yan, Q., Faull, R. L., Synek, B., & Dragunow, M. (1997). Brain-derived neurotrophic factor is reduced in Alzheimer's disease. *Brain Research Molecular Brain Research*, 49(1–2), 71–81. <http://www.ncbi.nlm.nih.gov/pubmed/9387865>. Accessed July 6, 2014.
- Conti, L., Cataudella, T., & Cattaneo, E. (2003). Neural stem cells: a pharmacological tool for brain diseases? *Pharmacological Research: The Official Journal of the Italian Pharmacological Society*, 47(4), 289–297. <http://www.ncbi.nlm.nih.gov/pubmed/12644385>. Accessed June 28, 2014.
- Corcoran, J., & Maden, M. (1999). Nerve growth factor acts via retinoic acid synthesis to stimulate neurite outgrowth. *Nature Neuroscience*, 2(4), 307–308. doi:10.1038/7214.
- Cowansage, K. K., LeDoux, J. E., & Monfils, M.-H. (2010). Brain-derived neurotrophic factor: a dynamic gatekeeper of neural plasticity. *Current Molecular Pharmacology*, 3(1), 12–29. <http://www.ncbi.nlm.nih.gov/pubmed/20030625>. Accessed July 7, 2014.
- Crews, F. T., & Nixon, K. (2003). Alcohol, neural stem cells, and adult neurogenesis. *Alcohol Research & Health: The Journal of the National Institute on Alcohol Abuse and Alcoholism*, 27(2), 197–204. <http://www.ncbi.nlm.nih.gov/pubmed/15303631>. Accessed June 28, 2014.
- Croce, N., Mathé, A. A., Gelfo, F., Caltagirone, C., Bernardini, S., & Angelucci, F. (2014). Effects of lithium and valproic acid on BDNF protein and gene expression in an in vitro human neuron-like model of degeneration. *Journal of Psychopharmacology (Oxford, England)*,. doi:10.1177/0269881114529379.
- Cuello, A. C., & Bruno, M. A. (2007). The failure in NGF maturation and its increased degradation as the probable cause for the vulnerability of cholinergic neurons in Alzheimer's disease. *Neurochemical Research*, 32(6), 1041–1045. doi:10.1007/s11064-006-9270-0.
- Curtis, M. A., Penney, E. B., Pearson, A. G., van Roon-Mom, W. M. C., Butterworth, N. J., Dragunow, M., et al. (2003). Increased cell proliferation and neurogenesis in the adult human Huntington's disease brain. *Proceedings of the National Academy of Sciences of the United States of America*, 100(15), 9023–9027. doi:10.1073/pnas.1532244100.
- Curtis, M. A., Kam, M., Nannmark, U., Anderson, M. F., Axell, M. Z., Wikkelso, C., et al. (2007). Human neuroblasts migrate to the olfactory bulb via a lateral ventricular extension. *Science (New York, N.Y.)*, 315(5816), 1243–1249. doi:10.1126/science.1136281.
- D'Amour, K. A., Bang, A. G., Eliazer, S., Kelly, O. G., Agulnick, A. D., Smart, N. G., et al. (2006). Production of pancreatic hormone-expressing endocrine cells from human embryonic stem cells. *Nature Biotechnology*, 24(11), 1392–1401. doi:10.1038/nbt1259.
- DeCarolis, N. A., & Eisch, A. J. (2010). Hippocampal neurogenesis as a target for the treatment of mental illness: A critical evaluation. *Neuropharmacology*, 58(6), 884–893. doi:10.1016/j.neuropharm.2009.12.013.
- Dodge, J. C., Haidet, A. M., Yang, W., Passini, M. A., Hester, M., Clarke, J., et al. (2008). Delivery of AAV-IGF-1 to the CNS extends survival in ALS mice through modification of aberrant glial cell activity. *Molecular Therapy: The Journal of the American Society of Gene Therapy*, 16(6), 1056–1064. doi:10.1038/mt.2008.60.
- Doetsch, F., & Alvarez-Buylla, A. (1996). Network of tangential pathways for neuronal migration in adult mammalian brain. *Proceedings of the National Academy of Sciences of the United States of America*, 93(25), 14895–14900. <http://www.ncbi.nlm.nih.gov/articlerender.fcgi?artid=26233&tool=pmcentrez&rendertype=abstract>. Accessed June 15, 2014.
- Doi, D., Samata, B., Katsukawa, M., Kikuchi, T., Morizane, A., Ono, Y., et al. (2014). Isolation of human induced pluripotent stem cell-derived dopaminergic progenitors by cell sorting for successful transplantation. *Stem Cell Reports*, 2(3), 337–350. doi:10.1016/j.stemcr.2014.01.013.
- Dorsey, S. G., Renn, C. L., Carim-Todd, L., Barrick, C. A., Bambrick, L., Krueger, B. K., et al. (2006). In vivo restoration of

- physiological levels of truncated TrkB<sup>T1</sup> receptor rescues neuronal cell death in a trisomic mouse model. *Neuron*, 51(1), 21–28. doi:10.1016/j.neuron.2006.06.009.
- Drinkut, A., Tereshchenko, Y., Schulz, J. B., Bähr, M., & Kügler, S. (2012). Efficient gene therapy for Parkinson's disease using astrocytes as hosts for localized neurotrophic factor delivery. *Molecular Therapy: The Journal of the American Society of Gene Therapy*, 20(3), 534–543. doi:10.1038/mt.2011.249.
- Duman, R. S., Malberg, J., & Nakagawa, S. (2001). Regulation of adult neurogenesis by psychotropic drugs and stress. *The Journal of Pharmacology and Experimental Therapeutics*, 299(2), 401–407. <http://www.ncbi.nlm.nih.gov/pubmed/11602648>. Accessed June 28, 2014.
- Duman, R. S., Nakagawa, S., & Malberg, J. (2001b). Regulation of adult neurogenesis by antidepressant treatment. *Neuropsychopharmacology: Official Publication of the American College of Neuropsychopharmacology*, 25(6), 836–844. doi:10.1016/S0893-133X(01)00358-X.
- Dwivedi, Y. (2009). Brain-derived neurotrophic factor: Role in depression and suicide. *Neuropsychiatric Disease and Treatment*, 5, 433–449. <http://www.ncbi.nlm.nih.gov/pmc/articles/der.fcgi?artid=2732010&tool=pmcentrez&rendertype=abstract>. Accessed June 5, 2014.
- Eggert, K., Schlegel, J., Oertel, W., Würz, C., Krieg, J. C., & Vedder, H. (1999). Glial cell line-derived neurotrophic factor protects dopaminergic neurons from 6-hydroxydopamine toxicity in vitro. *Neuroscience Letters*, 269(3), 178–182. <http://www.ncbi.nlm.nih.gov/pubmed/10454161>. Accessed July 4, 2014.
- Eisch, A. J., Barrot, M., Schad, C. A., Self, D. W., & Nestler, E. J. (2000). Opiates inhibit neurogenesis in the adult rat hippocampus. *Proceedings of the National Academy of Sciences of the United States of America*, 97(13), 7579–7584. doi:10.1073/pnas.120552597.
- Eisch, A. J., Cameron, H. A., Encinas, J. M., Meltzer, L. A., Ming, G.-L., & Overstreet-Wadiche, L. S. (2008). Adult neurogenesis, mental health, and mental illness: Hope or hype? *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, 28(46), 11785–11791. doi:10.1523/JNEUROSCI.3798-08.2008.
- Elizalde, C., Campa, V. M., Caro, M., Schlangen, K., Aransay, A. M., Vivanco, M. dM, & Kypta, R. M. (2011). Distinct roles for Wnt-4 and Wnt-11 during retinoic acid-induced neuronal differentiation. *Stem cells (Dayton, Ohio)*, 29(1), 141–153. doi:10.1002/stem.562.
- Erceg, S., Ronaghi, M., Oria, M., Roselló, M. G., Aragó, M. A. P., Lopez, M. G., et al. (2010). Transplanted oligodendrocytes and motoneuron progenitors generated from human embryonic stem cells promote locomotor recovery after spinal cord transection. *Stem cells (Dayton, Ohio)*, 28(9), 1541–1549. doi:10.1002/stem.489.
- Eriksson, P. S., Perfilieva, E., Björk-Eriksson, T., Alborn, A. M., Nordborg, C., Peterson, D. A., & Gage, F. H. (1998). Neurogenesis in the adult human hippocampus. *Nature Medicine*, 4(11), 1313–1317. doi:10.1038/3305.
- Eslamboli, A., Georgievska, B., Ridley, R. M., Baker, H. F., Muzyczka, N., Burger, C., et al. (2005). Continuous low-level glial cell line-derived neurotrophic factor delivery using recombinant adeno-associated viral vectors provides neuroprotection and induces behavioral recovery in a primate model of Parkinson's disease. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, 25(4), 769–777. doi:10.1523/JNEUROSCI.4421-04.2005.
- Fischer, W., Wictorin, K., Björklund, A., Williams, L. R., Varon, S., & Gage, F. H. (1987). Amelioration of cholinergic neuron atrophy and spatial memory impairment in aged rats by nerve growth factor. *Nature*, 329(6134), 65–68. doi:10.1038/329065a0.
- Frank, L., Ventimiglia, R., Anderson, K., Lindsay, R. M., & Rudge, J. S. (1996). BDNF down-regulates neurotrophin responsiveness, TrkB protein and TrkB mRNA levels in cultured rat hippocampal neurons. *The European Journal of Neuroscience*, 8(6), 1220–1230. <http://www.ncbi.nlm.nih.gov/pubmed/8752592>. Accessed September 14, 2014.
- Freed, C. R., Greene, P. E., Breeze, R. E., Tsai, W. Y., DuMouchel, W., Kao, R., et al. (2001). Transplantation of embryonic dopamine neurons for severe Parkinson's disease. *The New England Journal of Medicine*, 344(10), 710–719. doi:10.1056/NEJM200103083441002.
- Fuchs, E., & Flügge, G. (2014). Adult neuroplasticity: More than 40 years of research. *Neural Plasticity*. doi:10.1155/2014/541870.
- Fukuda, Y., Berry, T. L., Nelson, M., Hunter, C. L., Fukuhara, K., Imai, H., et al. (2010). Stimulated neuronal expression of brain-derived neurotrophic factor by Neurotropin. *Molecular and Cellular Neurosciences*, 45(3), 226–233. doi:10.1016/j.mcn.2010.06.013.
- Gage, F. H. (2000). Mammalian neural stem cells. *Science (New York, N.Y.)*, 287(5457), 1433–1438. <http://www.ncbi.nlm.nih.gov/pubmed/10688783>. Accessed June 2, 2014.
- Gaillard, A., Prestoz, L., Dumartin, B., Cantereau, A., Morel, F., Roger, M., & Jaber, M. (2007). Reestablishment of damaged adult motor pathways by grafted embryonic cortical neurons. *Nature Neuroscience*, 10(10), 1294–1299. doi:10.1038/nn1970.
- Garcia-Alloza, M., Borrelli, L. A., Rozkalne, A., Hyman, B. T., & Bacskai, B. J. (2007). Curcumin labels amyloid pathology in vivo, disrupts existing plaques, and partially restores distorted neurites in an Alzheimer mouse model. *Journal of Neurochemistry*, 102(4), 1095–1104. doi:10.1111/j.1471-4159.2007.04613.x.
- Géral, C., Angelova, A., Angelov, B., Nicolas, V., & Lesieur, S. (2012). Multicompartment lipid nanocarriers for targeting of cells expressing brain receptors. In N. Garti, R. Mezzenga, & P. Somasundaran (Eds.), *Self-assembled supramolecular architectures: Lyotropic liquid crystals* (pp. 319–355). New York, USA: Wiley.
- Géral, C., Angelova, A., & Lesieur, S. (2013). From molecular to nanotechnology strategies for delivery of neurotrophins: emphasis on brain-derived neurotrophic factor (BDNF). *Pharmaceutics*, 5(1), 127–167. doi:10.3390/pharmaceutics5010127.
- Gerrard, L., Rodgers, L., & Cui, W. (2005). Differentiation of human embryonic stem cells to neural lineages in adherent culture by blocking bone morphogenetic protein signaling. *Stem Cells (Dayton, Ohio)*, 23(9), 1234–1241. doi:10.1634/stemcells.2005-0110.
- Godman, C. A., Joshi, R., Giardina, C., Perdrizet, G., & Hightower, L. E. (2010). Hyperbaric oxygen treatment induces antioxidant gene expression. *Annals of the New York Academy of Sciences*, 1197, 178–183. doi:10.1111/j.1749-6632.2009.05393.x.
- Gonzalez, S. L., Labombarda, F., Deniselle, M. C. G., Mougel, A., Guennoun, R., Schumacher, M., & De Nicola, A. F. (2005). Progesterone neuroprotection in spinal cord trauma involves up-regulation of brain-derived neurotrophic factor in motoneurons. *The Journal of Steroid Biochemistry and Molecular Biology*, 94(1–3), 143–149. doi:10.1016/j.jsbmb.2005.01.016.
- Grandoso, L., Ponce, S., Manuel, I., Arrué, A., Ruiz-Ortega, J. A., Ulibarri, I., et al. (2007). Long-term survival of encapsulated GDNF secreting cells implanted within the striatum of parkinsonized rats. *International Journal of Pharmaceutics*, 343(1–2), 69–78. doi:10.1016/j.ijpharm.2007.05.027.
- Grealish, S., Diguet, E., Kirkeby, A., Mattsson, B., Heuer, A., Bramouille, Y., et al. (2014). Human ESC-derived dopamine neurons show similar preclinical efficacy and potency to fetal neurons when grafted in a rat model of Parkinson's Disease. *Cell Stem Cell*, 15(5), 653–665. doi:10.1016/j.stem.2014.09.017.

- Grondin, R., Zhang, Z., Yi, A., Cass, W. A., Maswood, N., Andersen, A. H., et al. (2002). Chronic, controlled GDNF infusion promotes structural and functional recovery in advanced parkinsonian monkeys. *Brain: Journal of Neurology*, 125(Pt 10), 2191–2201. <http://www.ncbi.nlm.nih.gov/pubmed/12244077>. Accessed June 5, 2014.
- Grondin, R., Zhang, Z., Ai, Y., Gash, D. M., & Gerhardt, G. A. (2003). Intracranial delivery of proteins and peptides as a therapy for neurodegenerative diseases. *Progress in drug research. Fortschritte der Arzneimittelforschung. Progrès des recherches pharmaceutiques*, 61, 101–123. <http://www.ncbi.nlm.nih.gov/pubmed/14674610>. Accessed July 8, 2014.
- Günther, A., Küppers-Tiedt, L., Schneider, P.-M., Kunert, I., Berrouschot, J., Schneider, D., & Rossner, S. (2005). Reduced infarct volume and differential effects on glial cell activation after hyperbaric oxygen treatment in rat permanent focal cerebral ischaemia. *The European Journal of Neuroscience*, 21(11), 3189–3194. doi:10.1111/j.1460-9568.2005.04151.x.
- Hádinger, N., Varga, B. V., Berzsényi, S., Környei, Z., Madarász, E., & Herberth, B. (2009). Astroglia genesis in vitro: Distinct effects of retinoic acid in different phases of neural stem cell differentiation. *International Journal of Developmental Neuroscience: The Official Journal of the International Society for Developmental Neuroscience*, 27(4), 365–375. doi:10.1016/j.ijdevneu.2009.02.004.
- Hanson, N. D., Nemeroff, C. B., & Owens, M. J. (2011). Lithium, but not fluoxetine or the corticotropin-releasing factor receptor 1 receptor antagonist R121919, increases cell proliferation in the adult dentate gyrus. *The Journal of Pharmacology and Experimental Therapeutics*, 337(1), 180–186. doi:10.1124/jpet.110.175372.
- Harwood, A. J. (2003). Neurodevelopment and mood stabilizers. *Current Molecular Medicine*, 3(5), 472–482. <http://www.ncbi.nlm.nih.gov/pubmed/12943000>. Accessed June 28, 2014.
- Haskell, G. T., & LaMantia, A.-S. (2005). Retinoic acid signaling identifies a distinct precursor population in the developing and adult forebrain. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, 25(33), 7636–7647. doi:10.1523/JNEUROSCI.0485-05.2005.
- He, J., Evans, C.-O., Hoffman, S. W., Oyesiku, N. M., & Stein, D. G. (2004). Progesterone and allopregnanolone reduce inflammatory cytokines after traumatic brain injury. *Experimental Neurology*, 189(2), 404–412. doi:10.1016/j.expneurol.2004.06.008.
- Herrán, E., Pérez-González, R., Igartua, M., Pedraz, J. L., Carro, E., & Hernández, R. M. (2013). VEGF-releasing biodegradable nanospheres administered by craniotomy: A novel therapeutic approach in the APP/Ps1 mouse model of Alzheimer's disease. *Journal of Controlled Release: Official Journal of the Controlled Release Society*, 170(1), 111–119. doi:10.1016/j.jconrel.2013.04.028.
- Herrán, E., Requejo, C., Ruiz-Ortega, J. A., Aristieta, A., Igartua, M., Bengoetxea, H., et al. (2014). Increased antiparkinson efficacy of the combined administration of VEGF- and GDNF-loaded nanospheres in a partial lesion model of Parkinson's disease. *International Journal of Nanomedicine*, 9(Suppl 1), 2677–2687. doi:10.2147/IJN.S61940.
- Hester, M. E., Murtha, M. J., Song, S., Rao, M., Miranda, C. J., Meyer, K., et al. (2011). Rapid and efficient generation of functional motor neurons from human pluripotent stem cells using gene delivered transcription factor codes. *Molecular Therapy: The Journal of the American Society of Gene Therapy*, 19(10), 1905–1912. doi:10.1038/mt.2011.135.
- Hicks, A. U., Lappalainen, R. S., Narkilahti, S., Suuronen, R., Corbett, D., Sivenius, J., et al. (2009). Transplantation of human embryonic stem cell-derived neural precursor cells and enriched environment after cortical stroke in rats: Cell survival and functional recovery. *The European Journal of Neuroscience*, 29(3), 562–574. doi:10.1111/j.1460-9568.2008.06599.x.
- Huang, H.-C., Lin, C.-J., Liu, W.-J., Jiang, R.-R., & Jiang, Z.-F. (2011). Dual effects of curcumin on neuronal oxidative stress in the presence of Cu(II). *Food and Chemical Toxicology: An International Journal Published for the British Industrial Biological Research Association*, 49(7), 1578–1583. doi:10.1016/j.fct.2011.04.004.
- Inestrosa, N. C., & Arenas, E. (2010). Emerging roles of Wnts in the adult nervous system. *Nature Reviews Neuroscience*, 11(2), 77–86. doi:10.1038/nrn2755.
- Itsykson, P., Ilouz, N., Turetsky, T., Goldstein, R. S., Pera, M. F., Fishbein, I., et al. (2005). Derivation of neural precursors from human embryonic stem cells in the presence of noggin. *Molecular and Cellular Neurosciences*, 30(1), 24–36. doi:10.1016/j.mcn.2005.05.004.
- Jacobs, S., Lie, D. C., DeCicco, K. L., Shi, Y., DeLuca, L. M., Gage, F. H., & Evans, R. M. (2006). Retinoic acid is required early during adult neurogenesis in the dentate gyrus. *Proceedings of the National Academy of Sciences of the United States of America*, 103(10), 3902–3907. doi:10.1073/pnas.0511294103.
- Jagasia, R., Steib, K., Englberger, E., Herold, S., Faus-Kessler, T., Saxe, M., et al. (2009). GABA-cAMP response element-binding protein signaling regulates maturation and survival of newly generated neurons in the adult hippocampus. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, 29(25), 7966–7977. doi:10.1523/JNEUROSCI.1054-09.2009.
- Je, H. S., Yang, F., Ji, Y., Potluri, S., Fu, X.-Q., Luo, Z.-G., et al. (2013). ProBDNF and mature BDNF as punishment and reward signals for synapse elimination at mouse neuromuscular junctions. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, 33(24), 9957–9962. doi:10.1523/JNEUROSCI.0163-13.2013.
- Jepson, B., Granpeesheh, D., Tarbox, J., Olive, M. L., Stott, C., Braud, S., et al. (2011). Controlled evaluation of the effects of hyperbaric oxygen therapy on the behavior of 16 children with autism spectrum disorders. *Journal of Autism and Developmental Disorders*, 41(5), 575–588. doi:10.1007/s10803-010-1075-y.
- Johansson, A. G. M., Nikamo, P., Schalling, M., & Landén, M. (2011). AKR1C4 gene variant associated with low euthymic serum progesterone and a history of mood irritability in males with bipolar disorder. *Journal of Affective Disorders*, 133(1–2), 346–351. doi:10.1016/j.jad.2011.04.009.
- Kaplan, D. R., Hempstead, B. L., Martin-Zanca, D., Chao, M. V., & Parada, L. F. (1991). The trk proto-oncogene product: a signal transducing receptor for nerve growth factor. *Science (New York, N.Y.)*, 252(5005), 554–558. <http://www.ncbi.nlm.nih.gov/pubmed/1850549>. Accessed June 29, 2014.
- Kaspars, B. K., Lladó, J., Sherkat, N., Rothstein, J. D., & Gage, F. H. (2003). Retrograde viral delivery of IGF-1 prolongs survival in a mouse ALS model. *Science (New York, N.Y.)*, 301(5634), 839–842. doi:10.1126/science.1086137.
- Katz, D. M. (2014). Brain-derived neurotrophic factor and Rett syndrome. *Handbook of Experimental Pharmacology*, 220, 481–495. doi:10.1007/978-3-642-45106-5\_18.
- Kawamata, T., Dietrich, W. D., Schallert, T., Gotts, J. E., Cocke, R. R., Benowitz, L. I., & Finklestein, S. P. (1997). Intracisternal basic fibroblast growth factor enhances functional recovery and up-regulates the expression of a molecular marker of neuronal sprouting following focal cerebral infarction. *Proceedings of the National Academy of Sciences of the United States of America*, 94(15), 8179–8184. <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC199336/>. Accessed June 30, 2014.

- Kells, A. P., Eberling, J., Su, X., Pivirotto, P., Bringas, J., Hadaczek, P., et al. (2010). Regeneration of the MPTP-lesioned dopaminergic system after convection-enhanced delivery of AAV2-GDNF. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, 30(28), 9567–9577. doi:10.1523/JNEUROSCI.0942-10.2010.
- Kelly, O. G., Chan, M. Y., Martinson, L. A., Kadoya, K., Ostertag, T. M., Ross, K. G., et al. (2011). Cell-surface markers for the isolation of pancreatic cell types derived from human embryonic stem cells. *Nature Biotechnology*, 29(8), 750–756. doi:10.1038/nbt.1931.
- Kempermann, G., Krebs, J., & Fabel, K. (2008). The contribution of failing adult hippocampal neurogenesis to psychiatric disorders. *Current Opinion in Psychiatry*, 21(3), 290–295. doi:10.1097/YCO.0b013e3282fad375.
- Kernie, S. G., & Parent, J. M. (2010). Forebrain neurogenesis after focal Ischemic and traumatic brain injury. *Neurobiology of Disease*, 37(2), 267–274. doi:10.1016/j.nbd.2009.11.002.
- Kim, D. S., Park, S. Y., & Kim, J. K. (2001). Curcuminoids from *Curcuma longa* L. (Zingiberaceae) that protect PC12 rat pheochromocytoma and normal human umbilical vein endothelial cells from betaA(1-42) insult. *Neuroscience Letters*, 303(1), 57–61. <http://www.ncbi.nlm.nih.gov/pubmed/11297823>. Accessed July 29, 2014.
- Kim, J.-H., Auerbach, J. M., Rodríguez-Gómez, J. A., Velasco, I., Gavin, D., Lumelsky, N., et al. (2002). Dopamine neurons derived from embryonic stem cells function in an animal model of Parkinson's disease. *Nature*, 418(6893), 50–56. doi:10.1038/nature00900.
- Kim, M., Lee, S.-T., Chu, K., & Kim, S. U. (2008a). Stem cell-based cell therapy for Huntington disease: A review. *Neuropathology: Official Journal of the Japanese Society of Neuropathology*, 28(1), 1–9. doi:10.1111/j.1440-1789.2007.00858.x.
- Kim, S. J., Son, T. G., Park, H. R., Park, M., Kim, M.-S., Kim, H. S., et al. (2008b). Curcumin stimulates proliferation of embryonic neural progenitor cells and neurogenesis in the adult hippocampus. *The Journal of biological chemistry*, 283(21), 14497–14505. doi:10.1074/jbc.M708373200.
- Kimbrel, E. A., & Lanza, R. (2015). Hope for regenerative treatments: Toward safe transplantation of human pluripotent stem-cell-based therapies. *Regenerative Medicine*, 10(2), 99–102. doi:10.2217/rme.14.89.
- Klein, R., Jing, S. Q., Nanduri, V., O'Rourke, E., & Barbacid, M. (1991a). The trk proto-oncogene encodes a receptor for nerve growth factor. *Cell*, 65(1), 189–197. <http://www.ncbi.nlm.nih.gov/pubmed/1849459>. Accessed June 29, 2014.
- Klein, R., Nanduri, V., Jing, S. A., Lamballe, F., Tapley, P., Bryant, S., et al. (1991b). The trkB tyrosine protein kinase is a receptor for brain-derived neurotrophic factor and neurotrophin-3. *Cell*, 66(2), 395–403. <http://www.ncbi.nlm.nih.gov/article/der.fcgi?artid=2710095&tool=pmcentrez&rendertype=abstract>. Accessed June 29, 2014.
- Knusel, B., Gao, H., Okazaki, T., Yoshida, T., Mori, N., Hefti, F., & Kaplan, D. R. (1997). Ligand-induced down-regulation of Trk messenger RNA, protein and tyrosine phosphorylation in rat cortical neurons. *Neuroscience*, 78(3), 851–862. <http://www.ncbi.nlm.nih.gov/pubmed/9153663>. Accessed September 14, 2014.
- Koch, P., Opitz, T., Steinbeck, J. A., Ladewig, J., & Brüstle, O. (2009). A rosette-type, self-renewing human ES cell-derived neural stem cell with potential for in vitro instruction and synaptic integration. *Proceedings of the National Academy of Sciences of the United States of America*, 106(9), 3225–3230. doi:10.1073/pnas.0808387106.
- Krencik, R., Weick, J. P., Liu, Y., Zhang, Z.-J., & Zhang, S.-C. (2011). Specification of transplantable astroglial subtypes from human pluripotent stem cells. *Nature Biotechnology*, 29(6), 528–534. doi:10.1038/nbt.1877.
- Ku, B., Kim, J., Chung, B. H., & Chung, B. G. (2013). Retinoic acid-polyethyleneimine complex nanoparticles for embryonic stem cell-derived neuronal differentiation. *Langmuir: The ACS Journal of Surfaces and Colloids*, 29(31), 9857–9862. doi:10.1021/la4015543.
- Kuhn, H. G., Palmer, T. D., & Fuchs, E. (2001). Adult neurogenesis: compensatory mechanism for neuronal damage. *European Archives of Psychiatry and Clinical Neuroscience*, 251(4), 152–158. <http://www.ncbi.nlm.nih.gov/pubmed/11697579>. Accessed June 28, 2014.
- Kukekov, V. G., Laywell, E. D., Suslov, O., Davies, K., Scheffler, B., Thomas, L. B., et al. (1999). Multipotent stem/progenitor cells with similar properties arise from two neurogenic regions of adult human brain. *Experimental Neurology*, 156(2), 333–344. doi:10.1006/exnr.1999.7028.
- Kurakhmaeva, K. B., Djindjikhashvili, I. A., Petrov, V. E., Balabanyan, V. U., Voronina, T. A., Trofimov, S. S., et al. (2009). Brain targeting of nerve growth factor using polybutyl cyanoacrylate) nanoparticles. *Journal of Drug Targeting*, 17(8), 564–574. doi:10.1080/10611860903112842.
- Lamballe, F., Klein, R., & Barbacid, M. (1991). trkC, a new member of the trk family of tyrosine protein kinases, is a receptor for neurotrophin-3. *Cell*, 66(5), 967–979. doi:10.1016/0092-8674(91)90442-2.
- Lamm, O., Ganz, J., Melamed, E., & Offen, D. (2014). Harnessing neurogenesis for the possible treatment of Parkinson's disease. *The Journal of Comparative Neurology*. doi:10.1002/cne.23607.
- Lee, G., Kim, H., Elkabetz, Y., Al Shamy, G., Panagiotakos, G., Barberi, T., et al. (2007a). Isolation and directed differentiation of neural crest stem cells derived from human embryonic stem cells. *Nature Biotechnology*, 25(12), 1468–1475. doi:10.1038/nbt1365.
- Lee, H., Shamy, G. Al, Elkabetz, Y., Schofield, C. M., Harrsion, N. L., Panagiotakos, G., et al. (2007b). Directed differentiation and transplantation of human embryonic stem cell-derived motoneurons. *Stem Cells (Dayton, Ohio)*, 25(8), 1931–1939. doi:10.1634/stemcells.2007-0097.
- Lee, Y.-C., Chio, C.-C., Chang, C.-P., Wang, L.-C., Chiang, P.-M., Niu, K.-C., & Tsai, K.-J. (2013). Long course hyperbaric oxygen stimulates neurogenesis and attenuates inflammation after ischemic stroke. *Inflammatory Mediators*, 2013, 13.
- Leonard, B. W., Mastroeni, D., Grover, A., Liu, Q., Yang, K., Gao, M., et al. (2009). Subventricular zone neural progenitors from rapid brain autopsies of elderly subjects with and without neurodegenerative disease. *The Journal of Comparative Neurology*, 515(3), 269–294. doi:10.1002/cne.22040.
- Lepski, G. (2012). What do we know about the neurogenic potential of different stem cell types? *Arquivos de Neuro-Psiquiatria*, 70(7), 540–546. doi:10.1590/S0004-282X2012000700013.
- Levin, H. S. (2003). Neuroplasticity following non-penetrating traumatic brain injury. *Brain Injury : [BI]*, 17(8), 665–674. doi:10.1080/0269905031000107151.
- Liebau, S., Vaida, B., Storch, A., & Boeckers, T. M. (2007). Maturation of synaptic contacts in differentiating neural stem cells. *Stem Cells (Dayton, Ohio)*, 25(7), 1720–1729. doi:10.1634/stemcells.2006-0823.
- Lindvall, O., & Wahlberg, L. U. (2008). Encapsulated cell biodelivery of GDNF: a novel clinical strategy for neuroprotection and neuroregeneration in Parkinson's disease? *Experimental Neurology*, 209(1), 82–88. doi:10.1016/j.expneuro.2007.08.019.
- Liu, Z., & Holmes, G. L. (1997). Basic fibroblast growth factor is highly neuroprotective against seizure-induced long-term behavioural deficits. *Neuroscience*, 76(4), 1129–1138. <http://www.ncbi.nlm.nih.gov/pubmed/9027873>. Accessed June 30, 2014.

- Liu, D., Wang, Z., Gao, Z., Xie, K., Zhang, Q., Jiang, H., & Pang, Q. (2014). Effects of curcumin on learning and memory deficits, BDNF, and ERK protein expression in rats exposed to chronic unpredictable stress. *Behavioural Brain Research*, 271C, 116–121. doi:10.1016/j.bbr.2014.05.068.
- Lois, C., García-Verdugo, J. M., & Alvarez-Buylla, A. (1996). Chain migration of neuronal precursors. *Science (New York, N.Y.)*, 271(5251), 978–981. <http://www.ncbi.nlm.nih.gov/pubmed/8584933>. Accessed June 15, 2014.
- Lovell, M. A., Geiger, H., Van Zant, G. E., Lynn, B. C., & Markesberry, W. R. (2006). Isolation of neural precursor cells from Alzheimer's disease and aged control postmortem brain. *Neurobiology of Aging*, 27(7), 909–917. doi:10.1016/j.neurobiaging.2005.05.004.
- Lu, P., Jones, L. L., Snyder, E. Y., & Tuszynski, M. H. (2003). Neural stem cells constitutively secrete neurotrophic factors and promote extensive host axonal growth after spinal cord injury. *Experimental Neurology*, 181(2), 115–129. <http://www.ncbi.nlm.nih.gov/pubmed/12781986>. Accessed May 31, 2014.
- Lu, J., Tan, L., Li, P., Gao, H., Fang, B., Ye, S., et al. (2009). All-trans retinoic acid promotes neural lineage entry by pluripotent embryonic stem cells via multiple pathways. *BMC Cell Biology*, 10, 57. doi:10.1186/1471-2121-10-57.
- Ma, Y.-P., Ma, M.-M., Cheng, S.-M., Ma, H.-H., Yi, X.-M., Xu, G.-L., & Liu, X.-F. (2008). Intranasal bFGF-induced progenitor cell proliferation and neuroprotection after transient focal cerebral ischemia. *Neuroscience Letters*, 437(2), 93–97. doi:10.1016/j.neulet.2008.04.003.
- Macas, J., Nern, C., Plate, K. H., & Momma, S. (2006). Increased generation of neuronal progenitors after ischemic injury in the aged adult human forebrain. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, 26(50), 13114–13119. doi:10.1523/JNEUROSCI.4667-06.2006.
- Maia, J., Santos, T., Aday, S., Agasse, F., Cortes, L., Malva, J. O., et al. (2011). Controlling the neuronal differentiation of stem cells by the intracellular delivery of retinoic acid-loaded nanoparticles. *ACS Nano*, 5(1), 97–106. doi:10.1021/nn101724r.
- Malberg, J. E. (2004). Implications of adult hippocampal neurogenesis in antidepressant action. *Journal of Psychiatry & Neuroscience: PN*, 29(3), 196–205. <http://www.ncbi.nlm.nih.gov/pubmed/15048977>. Accessed June 28, 2014.
- Malberg, J. E., Eisisch, A. J., Nestler, E. J., & Duman, R. S. (2000). Chronic antidepressant treatment increases neurogenesis in adult rat hippocampus. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, 20(24), 9104–9110. <http://www.ncbi.nlm.nih.gov/pubmed/11124987>. Accessed June 13, 2014.
- Mansouri, Z., Sabetkasaei, M., Moradi, F., Masoudnia, F., & Ataie, A. (2012). Curcumin has neuroprotection effect on homocysteine rat model of Parkinson. *Journal of Molecular Neuroscience: MN*, 47(2), 234–242. doi:10.1007/s12031-012-9727-3.
- Marlatt, M. W., Lucassen, P. J., & van Praag, H. (2010). Comparison of neurogenic effects of fluoxetine, duloxetine and running in mice. *Brain Research*, 1341, 93–99. doi:10.1016/j.brainres.2010.03.086.
- Martina, M.-S., Wilhelm, C., & Lesieur, S. (2008). The effect of magnetic targeting on the uptake of magnetic-fluid-loaded liposomes by human prostatic adenocarcinoma cells. *Biomaterials*, 29(30), 4137–4145. doi:10.1016/j.biomaterials.2008.07.011.
- Martinez-Fong, D., Bannon, M. J., Trudeau, L.-E., Gonzalez-Barrios, J. A., Arango-Rodriguez, M. L., Hernandez-Chan, N. G., et al. (2012). NTS-Polyplex: A potential nanocarrier for neurotrophic therapy of Parkinson's disease. *Nanomedicine: Nanotechnology, Biology and Medicine*, 8(7), 1052–1069. doi:10.1016/j.nano.2012.02.009.
- Marx, C. E., VanDoren, M. J., Duncan, G. E., Lieberman, J. A., & Morrow, A. L. (2003). Olanzapine and clozapine increase the GABAergic neuroactive steroid allopregnanolone in rodents. *Neuropsychopharmacology: Official Publication of the American College of Neuropsychopharmacology*, 28(1), 1–13. doi:10.1038/sj.npp.1300015.
- Mattson, M. P. (2008). Glutamate and neurotrophic factors in neuronal plasticity and disease. *Annals of the New York Academy of Sciences*, 1144, 97–112. doi:10.1196/annals.1418.005.
- Maucksch, C., Vazey, E. M., Gordon, R. J., & Connor, B. (2013). Stem cell-based therapy for Huntington's disease. *Journal of Cellular Biochemistry*, 114(4), 754–763. doi:10.1002/jcb.24432.
- Mazumdar, J., O'Brien, W. T., Johnson, R. S., LaManna, J. C., Chavez, J. C., Klein, P. S., & Simon, M. C. (2010). O2 regulates stem cells through Wnt/β-catenin signalling. *Nature Cell Biology*, 12(10), 1007–1013. doi:10.1038/ncb2102.
- McAvoy, K., Russo, C., Kim, S., Rankin, G., & Sahay, A. (2015). Fluoxetine induces input-specific hippocampal dendritic spine remodeling along the septotemporal axis in adulthood and middle age. *Hippocampus*. doi:10.1002/hipo.22464.
- McEwen, B. S. (1999). Stress and hippocampal plasticity. *Annual Review of Neuroscience*, 22, 105–122. doi:10.1146/annurev.neuro.22.1.105.
- Mehrotra, S., Lynam, D., Maloney, R., Pawelec, K. M., Tuszynski, M. H., Lee, I., et al. (2010). Time controlled protein release from layer-by-layer assembled multilayer functionalized agarose hydrogels. *Advanced Functional Materials*, 20(2), 247–258. doi:10.1002/adfm.200901172.
- Menasché, P., Vanneaux, V., Fabreguettes, J.-R., Bel, A., Tosca, L., Garcia, S., et al. (2015). Towards a clinical use of human embryonic stem cell-derived cardiac progenitors: A translational experience. *European Heart Journal*, 36(12), 743–750. doi:10.1093/eurheartj/ehu192.
- Mendez, I., Sanchez-Pernaute, R., Cooper, O., Viñuela, A., Ferrari, D., Björklund, L., et al. (2005). Cell type analysis of functional fetal dopamine cell suspension transplants in the striatum and substantia nigra of patients with Parkinson's disease. *Brain: A Journal of Neurology*, 128(Pt 7), 1498–1510. doi:10.1093/brain/awh510.
- Migliore, M. M., Ortiz, R., Dye, S., Campbell, R. B., Amiji, M. M., & Waszcak, B. L. (2014). Neurotrophic and neuroprotective efficacy of intranasal GDNF in a rat model of Parkinson's disease. *Neuroscience*, 274, 11–23. doi:10.1016/j.neuroscience.2014.05.019.
- Milosevic, J., Adler, I., Manaenko, A., Schwarz, S. C., Walkinshaw, G., Arend, M., et al. (2009). Non-hypoxic stabilization of hypoxia-inducible factor alpha (HIF-alpha): Relevance in neural progenitor/stem cells. *Neurotoxicity Research*, 15(4), 367–380. doi:10.1007/s12640-009-9043-z.
- Monteggia, L. M., Barrot, M., Powell, C. M., Berton, O., Galanis, V., Gemelli, T., et al. (2004). Essential role of brain-derived neurotrophic factor in adult hippocampal function. *Proceedings of the National Academy of Sciences of the United States of America*, 101(29), 10827–10832. doi:10.1073/pnas.0402141101.
- Monteggia, L. M., Luikart, B., Barrot, M., Theobold, D., Malkovska, I., Nef, S., et al. (2007). Brain-derived neurotrophic factor conditional knockouts show gender differences in depression-related behaviors. *Biological Psychiatry*, 61(2), 187–197. doi:10.1016/j.biopsych.2006.03.021.
- Mu, J., Krafft, P. R., & Zhang, J. H. (2011). Hyperbaric oxygen therapy promotes neurogenesis: Where do we stand? *Medical Gas Research*, 1(1), 14. doi:10.1186/2045-9912-1-14.
- Mukherjee, A., Raison, M., Sahni, T., Arya, A., Lambert, J., Marois, P., et al. (2014). Intensive rehabilitation combined with HBO2 therapy in children with cerebral palsy: a controlled longitudinal study. *Undersea & Hyperbaric Medicine: Journal of the*

- Undersea and Hyperbaric Medical Society, Inc.*, 41(2), 77–85. <http://www.ncbi.nlm.nih.gov/pubmed/24851544>. Accessed July 29, 2014.
- Mychaskiw, G. (2010). Hyperbaric oxygen therapy and neurologic disease: the time has come. *Undersea & Hyperbaric Medicine: Journal of the Undersea and Hyperbaric Medical Society, Inc.*, 37(2), xi–xiii. <http://www.ncbi.nlm.nih.gov/pubmed/20462138>. Accessed July 29, 2014.
- Nair, S. S., Prathibha, P., Syam Das, S., Kavitha, S., & Indira, M. (2015). All trans retinoic acid (ATRA) mediated modulation of N-methyl D-aspartate receptor (NMDAR) and Kruppel like factor 11 (KLF11) expressions in the mitigation of ethanol induced alterations in the brain. *Neurochemistry International*, 83–84, 41–47. doi:[10.1016/j.neuint.2015.02.007](https://doi.org/10.1016/j.neuint.2015.02.007).
- Narkilahti, S., Rajala, K., Pihlajamäki, H., Suuronen, R., Hovatta, O., & Skottman, H. (2007). Monitoring and analysis of dynamic growth of human embryonic stem cells: Comparison of automated instrumentation and conventional culturing methods. *Biomedical Engineering Online*, 6, 11. doi:[10.1186/1475-925X-6-11](https://doi.org/10.1186/1475-925X-6-11).
- Narvekar, M., Xue, H. Y., & Wong, H. L. (2012). A novel hybrid delivery system: polymer-oil nanostructured carrier for controlled delivery of highly lipophilic drug all-trans-retinoic acid (ATRA). *International Journal of Pharmaceutics*, 436(1–2), 721–731. doi:[10.1016/j.ijpharm.2012.07.042](https://doi.org/10.1016/j.ijpharm.2012.07.042).
- Noisa, P., & Parnpai, R. (2011). Technical challenges in the derivation of human pluripotent cells. *Stem cells international*, 2011, 907961. doi:[10.4061/2011/907961](https://doi.org/10.4061/2011/907961).
- Nunes, M. C., Roy, N. S., Keyoung, H. M., Goodman, R. R., McKhann, G., Jiang, L., et al. (2003). Identification and isolation of multipotential neural progenitor cells from the subcortical white matter of the adult human brain. *Nature Medicine*, 9(4), 439–447. doi:[10.1038/nm837](https://doi.org/10.1038/nm837).
- Ormerod, B. K., Palmer, T. D., & Caldwell, M. A. (2008). Neurodegeneration and cell replacement. *Philosophical Transactions of the Royal Society of London. Series B, Biological sciences*, 363(1489), 153–170. doi:[10.1098/rstb.2006.2018](https://doi.org/10.1098/rstb.2006.2018).
- Pagliuca, F. W., Millman, J. R., Gürler, M., Segel, M., Van Dervort, A., Ryu, J. H., et al. (2014). Generation of functional human pancreatic β cells in vitro. *Cell*, 159(2), 428–439. doi:[10.1016/j.cell.2014.09.040](https://doi.org/10.1016/j.cell.2014.09.040).
- Palencia, G., Hernández-Pedro, N., Saavedra-Perez, D., Peña-Curiel, O., Ortiz-Plata, A., Ordoñez, G., et al. (2014). Retinoic acid reduces solvent-induced neuropathy and promotes neural regeneration in mice. *Journal of Neuroscience Research*, 92(8), 1062–1070. doi:[10.1002/jnr.23376](https://doi.org/10.1002/jnr.23376).
- Pang, Z., Lu, W., Gao, H., Hu, K., Chen, J., Zhang, C., et al. (2008). Preparation and brain delivery property of biodegradable polymersomes conjugated with OX26. *Journal of Controlled Release: Official Journal of the Controlled Release Society*, 128(2), 120–127. doi:[10.1016/j.jconrel.2008.03.007](https://doi.org/10.1016/j.jconrel.2008.03.007).
- Park, T. I.-H., Monzo, H., Mee, E. W., Bergin, P. S., Teoh, H. H., Montgomery, J. M., et al. (2012). Adult human brain neural progenitor cells (NPCs) and fibroblast-like cells have similar properties in vitro but only NPCs differentiate into neurons. *PLoS ONE*, 7(6), e37742. doi:[10.1371/journal.pone.0037742](https://doi.org/10.1371/journal.pone.0037742).
- Paschaki, M., Cammas, L., Muta, Y., Matsuoka, Y., Mak, S.-S., Rataj-Baniowska, M., et al. (2013). Retinoic acid regulates olfactory progenitor cell fate and differentiation. *Neural Development*, 8, 13. doi:[10.1186/1749-8104-8-13](https://doi.org/10.1186/1749-8104-8-13).
- Peng, S., Wuu, J., Mufson, E. J., & Fahnestock, M. (2004). Increased proNGF levels in subjects with mild cognitive impairment and mild Alzheimer disease. *Journal of Neuropathology and Experimental Neurology*, 63(6), 641–649. <http://www.ncbi.nlm.nih.gov/pubmed/15217092>. Accessed June 15, 2014.
- Perrier, A. L., Tabar, V., Barberi, T., Rubio, M. E., Bruses, J., Topf, N., et al. (2004). Derivation of midbrain dopamine neurons from human embryonic stem cells. *Proceedings of the National Academy of Sciences of the United States of America*, 101(34), 12543–12548. doi:[10.1073/pnas.0404700101](https://doi.org/10.1073/pnas.0404700101).
- Pettus, E. H., Wright, D. W., Stein, D. G., & Hoffman, S. W. (2005). Progesterone treatment inhibits the inflammatory agents that accompany traumatic brain injury. *Brain Research*, 1049(1), 112–119. doi:[10.1016/j.brainres.2005.05.004](https://doi.org/10.1016/j.brainres.2005.05.004).
- Pibiri, F., Nelson, M., Guidotti, A., Costa, E., & Pinna, G. (2008). Decreased corticolimbic allopregnanolone expression during social isolation enhances contextual fear: A model relevant for posttraumatic stress disorder. *Proceedings of the National Academy of Sciences of the United States of America*, 105(14), 5567–5572. doi:[10.1073/pnas.0801853105](https://doi.org/10.1073/pnas.0801853105).
- Pillai, A. (2008). Brain-derived neurotropic factor/TrkB signaling in the pathogenesis and novel pharmacotherapy of schizophrenia. *Neuro-Signals*, 16(2–3), 183–193. doi:[10.1159/000111562](https://doi.org/10.1159/000111562).
- Piña-Crespo, J. C., Talantova, M., Cho, E.-G., Soussou, W., Dolatabadi, N., Ryan, S. D., et al. (2012). High-frequency hippocampal oscillations activated by optogenetic stimulation of transplanted human ESC-derived neurons. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, 32(45), 15837–15842. doi:[10.1523/JNEUROSCI.3735-12.2012](https://doi.org/10.1523/JNEUROSCI.3735-12.2012).
- Rangarajan, P., Eng-Ang, L., & Dheen, S. T. (2013). Potential drugs targeting microglia: current knowledge and future prospects. *CNS & Neurological Disorders Drug Targets*, 12(6), 799–806. <http://www.ncbi.nlm.nih.gov/pubmed/24047522>. Accessed June 5, 2014.
- Reubinoff, B. E., Itsykson, P., Turetsky, T., Pera, M. F., Reinhartz, E., Itzik, A., & Ben-Hur, T. (2001). Neural progenitors from human embryonic stem cells. *Nature Biotechnology*, 19(12), 1134–1140. doi:[10.1038/nbt1201-1134](https://doi.org/10.1038/nbt1201-1134).
- Reynolds, B., & Weiss, S. (1992). Generation of neurons and astrocytes from isolated cells of the adult mammalian central nervous system. *Science*, 255(5052), 1707–1710. doi:[10.1126/science.1553558](https://doi.org/10.1126/science.1553558).
- Richardson, R. M., Holloway, K. L., Bullock, M. R., Broaddus, W. C., & Fillmore, H. L. (2006). Isolation of neuronal progenitor cells from the adult human neocortex. *Acta Neurochirurgica*, 148(7), 773–777. doi:[10.1007/s00701-006-0778-5](https://doi.org/10.1007/s00701-006-0778-5).
- Richardson, R. M., Sun, D., & Bullock, M. R. (2007). Neurogenesis after traumatic brain injury. *Neurosurgery Clinics of North America*, 18(1), 169–181, xi. doi:[10.1016/j.nec.2006.10.007](https://doi.org/10.1016/j.nec.2006.10.007).
- Rockswold, S. B., Rockswold, G. L., & Defillo, A. (2007). Hyperbaric oxygen in traumatic brain injury. *Neurological Research*, 29(2), 162–172. doi:[10.1179/016164107X181798](https://doi.org/10.1179/016164107X181798).
- Roof, R. L., Hoffman, S. W., & Stein, D. G. (1997). Progesterone protects against lipid peroxidation following traumatic brain injury in rats. *Molecular and Chemical Neuropathology/Sponsored by the International Society for Neurochemistry and the World Federation of Neurology and Research Groups on Neurochemistry and Cerebrospinal Fluid*, 31(1), 1–11. <http://www.ncbi.nlm.nih.gov/pubmed/9271001>. Accessed July 15, 2014.
- Rossignol, D. A., Rossignol, L. W., Smith, S., Schneider, C., Logerquist, S., Usman, A., et al. (2009). Hyperbaric treatment for children with autism: A multicenter, randomized, double-blind, controlled trial. *BMC Pediatrics*, 9, 21. doi:[10.1186/1471-2431-9-21](https://doi.org/10.1186/1471-2431-9-21).
- Saarelainen, T., Hendolin, P., Lucas, G., Koponen, E., Sairanen, M., MacDonald, E., et al. (2003). Activation of the TrkB neurotrophin receptor is induced by antidepressant drugs and is required for antidepressant-induced behavioral effects. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, 23(1), 349–357. <http://www.ncbi.nlm.nih.gov/pubmed/12514234>. Accessed July 7, 2014.

- Sajadi, A., Bensadoun, J.-C., Schneider, B. L., Lo Bianco, C., & Aebscher, P. (2006). Transient striatal delivery of GDNF via encapsulated cells leads to sustained behavioral improvement in a bilateral model of Parkinson disease. *Neurobiology of Disease*, 22(1), 119–129. doi:10.1016/j.nbd.2005.10.006.
- Sakane, T., & Pardridge, W. M. (1997). Carboxyl-directed pegylation of brain-derived neurotrophic factor markedly reduces systemic clearance with minimal loss of biologic activity. *Pharmaceutical Research*, 14(8), 1085–1091. <http://www.ncbi.nlm.nih.gov/pubmed/9279893>. Accessed September 22, 2014.
- Sampanthavivat, M., Singkhwa, W., Chaiyakul, T., Karoonyawanich, S., & Ajpru, H. (2012). Hyperbaric oxygen in the treatment of childhood autism: a randomised controlled trial. *Diving and Hyperbaric Medicine: the Journal of the South Pacific Underwater Medicine Society*, 42(3), 128–133. <http://www.ncbi.nlm.nih.gov/pubmed/22987458>. Accessed July 24, 2014.
- Sanai, N., Tramontin, A. D., Quiñones-Hinojosa, A., Barbaro, N. M., Gupta, N., Kunwar, S., et al. (2004). Unique astrocyte ribbon in adult human brain contains neural stem cells but lacks chain migration. *Nature*, 427(6976), 740–744. doi:10.1038/nature02301.
- Sandhir, R., Yadav, A., Mehrotra, A., Sunkaria, A., Singh, A., & Sharma, S. (2014). Curcumin nanoparticles attenuate neurochemical and neurobehavioral deficits in experimental model of Huntington's disease. *NeuroMolecular Medicine*, 16(1), 106–118. doi:10.1007/s12017-013-8261-y.
- Sandur, S. K., Pandey, M. K., Sung, B., Ahn, K. S., Murakami, A., Sethi, G., et al. (2007). Curcumin, demethoxycurcumin, bis-demethoxycurcumin, tetrahydrocurcumin and turmerones differentially regulate anti-inflammatory and anti-proliferative responses through a ROS-independent mechanism. *Carcinogenesis*, 28(8), 1765–1773. doi:10.1093/carcin/bgm123.
- Santos, T., Ferreira, R., Maia, J., Agasse, F., Xapelli, S., Cortes, L., et al. (2012). Polymeric nanoparticles to control the differentiation of neural stem cells in the subventricular zone of the brain. *ACS Nano*, 6(12), 10463–10474. doi:10.1021/nn304541h.
- Schulz, T. C., Young, H. Y., Agulnick, A. D., Babin, M. J., Baetge, E. E., Bang, A. G., et al. (2012). A scalable system for production of functional pancreatic progenitors from human embryonic stem cells. *PLoS ONE*, 7(5), e37004. doi:10.1371/journal.pone.0037004.
- Schwartz, S. D., Hubschman, J.-P., Heilwell, G., Franco-Cardenas, V., Pan, C. K., Ostrick, R. M., et al. (2012). Embryonic stem cell trials for macular degeneration: A preliminary report. *Lancet*, 379(9817), 713–720. doi:10.1016/S0140-6736(12)60028-2.
- Schwartz, S. D., Regillo, C. D., Lam, B. L., Elliott, D., Rosenfeld, P. J., Gregori, N. Z., et al. (2014). Human embryonic stem cell-derived retinal pigment epithelium in patients with age-related macular degeneration and Stargardt's macular dystrophy: Follow-up of two open-label phase 1/2 studies. *The Lancet*, 385(9967), 509–516. doi:10.1016/S0140-6736(14)61376-3.
- Shin, H. Y., Kim, J. H., Phi, J. H., Park, C.-K., Kim, J. E., Kim, J.-H., et al. (2008). Endogenous neurogenesis and neovascularization in the neocortex of the rat after focal cerebral ischemia. *Journal of Neuroscience Research*, 86(2), 356–367. doi:10.1002/jnr.21494.
- Shirakura, M., Inoue, M., Fujikawa, S., Washizawa, K., Komaba, S., Maeda, M., et al. (2004). Postischemic administration of Sendai virus vector carrying neurotrophic factor genes prevents delayed neuronal death in gerbils. *Gene Therapy*, 11(9), 784–790. doi:10.1038/sj.gt.3302224.
- Simon, A., Allais, D. P., Duroux, J. L., Basly, J. P., Durand-Fontanier, S., & Delage, C. (1998). Inhibitory effect of curcuminoids on MCF-7 cell proliferation and structure-activity relationships. *Cancer Letters*, 129(1), 111–116. <http://www.ncbi.nlm.nih.gov/pubmed/9714342>. Accessed July 29, 2014.
- Skaper, S. D. (2012). The neurotrophin family of neurotrophic factors: an overview. *Methods in Molecular Biology (Clifton, N.J.)*, 846, 1–12. doi:10.1007/978-1-61779-536-7\_1.
- Sommerfeld, M. T., Schweigreiter, R., Barde, Y. A., & Hoppe, E. (2000). Down-regulation of the neurotrophin receptor TrkB following ligand binding. Evidence for an involvement of the proteasome and differential regulation of TrkA and TrkB. *The Journal of Biological Chemistry*, 275(12), 8982–8990. <http://www.ncbi.nlm.nih.gov/pubmed/10722747>. Accessed September 14, 2014.
- Squinto, S. P., Stitt, T. N., Aldrich, T. H., Davis, S., Bianco, S. M., Radziejewski, C., et al. (1991). trkB encodes a functional receptor for brain-derived neurotrophic factor and neurotrophin-3 but not nerve growth factor. *Cell*, 65(5), 885–893. <http://www.ncbi.nlm.nih.gov/pubmed/1710174>. Accessed June 29, 2014.
- Stachowiak, E. K., Roy, I., Lee, Y.-W., Capacchietti, M., Aletta, J. M., Prasad, P. N., & Stachowiak, M. K. (2009). Targeting novel integrative nuclear FGFR1 signaling by nanoparticle-mediated gene transfer stimulates neurogenesis in the adult brain. *Integrative Biology: Quantitative Biosciences from Nano to Macro*, 1(5–6), 394–403. doi:10.1039/b902617g.
- Stankowski, J. N., & Gupta, R. (2011). Therapeutic targets for neuroprotection in acute ischemic stroke: Lost in translation? *Antioxidants & Redox Signaling*, 14(10), 1841–1851. doi:10.1089/ars.2010.3292.
- Stavridis, M. P., Collins, B. J., & Storey, K. G. (2010). Retinoic acid orchestrates fibroblast growth factor signalling to drive embryonic stem cell differentiation. *Development (Cambridge, England)*, 137(6), 881–890. doi:10.1242/dev.043117.
- Steiner, B., Wolf, S., & Kempermann, G. (2006). Adult neurogenesis and neurodegenerative disease. *Regenerative Medicine*, 1(1), 15–28. doi:10.2217/17460751.1.1.15.
- Sumanont, Y., Murakami, Y., Tohda, M., Vajragupta, O., Watanabe, H., & Matsumoto, K. (2007). Effects of manganese complexes of curcumin and diacetylcurcumin on kainic acid-induced neurotoxic responses in the rat hippocampus. *Biological & Pharmaceutical Bulletin*, 30(9), 1732–1739. <http://www.ncbi.nlm.nih.gov/pubmed/17827730>. Accessed July 10, 2014.
- Suri, D., & Vaidya, V. A. (2013). Glucocorticoid regulation of brain-derived neurotrophic factor: Relevance to hippocampal structural and functional plasticity. *Neuroscience*, 239, 196–213. doi:10.1016/j.neuroscience.2012.08.065.
- Tachibana, M., Amato, P., Sparman, M., Gutierrez, N. M., Tippner-Hedges, R., Ma, H., et al. (2013). Human embryonic stem cells derived by somatic cell nuclear transfer. *Cell*, 153(6), 1228–1238. doi:10.1016/j.cell.2013.05.006.
- Takahashi, J., Palmer, T. D., & Gage, F. H. (1999). Retinoic acid and neurotrophins collaborate to regulate neurogenesis in adult-derived neural stem cell cultures. *Journal of Neurobiology*, 38(1), 65–81. <http://www.ncbi.nlm.nih.gov/pubmed/10027563>. Accessed July 8, 2014.
- Takahashi, K., Tanabe, K., Ohnuki, M., Narita, M., Ichisaka, T., Tomoda, K., & Yamanaka, S. (2007). Induction of pluripotent stem cells from adult human fibroblasts by defined factors. *Cell*, 131(5), 861–872. doi:10.1016/j.cell.2007.11.019.
- Tan, J., Wang, Y., Yip, X., Glynn, F., Shepherd, R. K., & Caruso, F. (2012). Nanoporous peptide particles for encapsulating and releasing neurotrophic factors in an animal model of neurodegeneration. *Advanced Materials (Deerfield Beach, Fla.)*, 24(25), 3362–3366. doi:10.1002/adma.201200634.
- Tan, J., Shi, J., Shi, G., Liu, Y., Liu, X., Wang, C., et al. (2013). Changes in compressed neurons from dogs with acute and severe cauda equina constrictions following intrathecal injection of brain-derived neurotrophic factor-conjugated polymer nanoparticles. *Neural Regeneration Research*, 8(3), 233–243. doi:10.3969/j.issn.1673-5374.2013.03.005.

- Thapa, A., Vernon, B. C., De la Peña, K., Soliz, G., Moreno, H. A., López, G. P., & Chi, E. Y. (2013). Membrane-mediated neuroprotection by curcumin from amyloid- $\beta$ -peptide-induced toxicity. *Langmuir: The ACS Journal of Surfaces and Colloids*, 29(37), 11713–11723. doi:10.1021/la4020459.
- Theofilopoulos, S., Goggi, J., Riaz, S. S., Jauniaux, E., Stern, G. M., & Bradford, H. F. (2001). Parallel induction of the formation of dopamine and its metabolites with induction of tyrosine hydroxylase expression in foetal rat and human cerebral cortical cells by brain-derived neurotrophic factor and glial-cell derived neurotrophic factor. *Brain Research. Developmental Brain Research*, 127(2), 111–122. doi:10.1016/S0165-3806(01)00125-0.
- Thoenen, H. (1995). Neurotrophins and neuronal plasticity. *Science (New York, N.Y.)*, 270(5236), 593–598. <http://www.ncbi.nlm.nih.gov/pubmed/7570017>. Accessed June 29, 2014.
- Tian, X., Wang, J., Dai, J., Yang, L., Zhang, L., Shen, S., & Huang, P. (2012). Hyperbaric oxygen and Ginkgo Biloba extract inhibit A $\beta$ 25-35-induced toxicity and oxidative stress in vivo: A potential role in Alzheimer's disease. *The International Journal of Neuroscience*, 122(10), 563–569. doi:10.3109/00207454.2012.690797.
- Tiwari, S. S. K., Agarwal, S., Seth, B., Yadav, A., Nair, S., Bhatnagar, P., et al. (2014). Curcumin-loaded nanoparticles potently induce adult neurogenesis and reverse cognitive deficits in Alzheimer's disease model via canonical Wnt/ $\beta$ -catenin pathway. *ACS Nano*, 8(1), 76–103. doi:10.1021/nn405077y.
- Tripanichkul, W., & Jaroensuppaperch, E.-O. (2013). Ameliorating effects of curcumin on 6-OHDA-induced dopaminergic degeneration, glial response, and SOD1 reduction in the striatum of hemiparkinsonian mice. *European Review for Medical and Pharmacological Sciences*, 17(10), 1360–1368. <http://www.ncbi.nlm.nih.gov/pubmed/23740450>. Accessed July 10, 2014.
- Trounson, A. (2006). The production and directed differentiation of human embryonic stem cells. *Endocrine Reviews*, 27(2), 208–219. doi:10.1210/er.2005-0016.
- Tsai, S.-J. (2006). TrkB partial agonists: potential treatment strategy for epilepsy, mania, and autism. *Medical Hypotheses*, 66(1), 173–175. doi:10.1016/j.mehy.2005.05.033.
- Tsai, Y.-M., Chien, C.-F., Lin, L.-C., & Tsai, T.-H. (2011). Curcumin and its nano-formulation: The kinetics of tissue distribution and blood-brain barrier penetration. *International Journal of Pharmaceutics*, 416(1), 331–338. doi:10.1016/j.ijpharm.2011.06.030.
- Uchida, K., Momiyama, T., Okano, H., Yuzaki, M., Koizumi, A., Mine, Y., & Kawase, T. (2005). Potential functional neural repair with grafted neural stem cells of early embryonic neuroepithelial origin. *Neuroscience Research*, 52(3), 276–286. doi:10.1016/j.neures.2005.03.015.
- Uzun, G., Subhani, D., & Amor, S. (2010). Trophic factors and stem cells for promoting recovery in stroke. *Journal of Vascular and Interventional Neurology*, 3(1), 3–12. <http://www.ncbi.nlm.nih.gov/articlerender.fcgi?aid=3317290&tool=pmcentrez&rendertype=abstract>. Accessed July 7, 2014.
- Uzunova, V., Sheline, Y., Davis, J. M., Rasmusson, A., Uzunov, D. P., Costa, E., et al. (1998). Increase in the cerebrospinal fluid content of neurosteroids in patients with unipolar major depression who are receiving fluoxetine or fluvoxamine. *Proceedings of the National Academy of Sciences of the United States of America*, 95(6), 3239–3244. <http://www.ncbi.nlm.nih.gov/articlerender.fcgi?aid=19726&tool=pmcentrez&rendertype=abstract>. Accessed July 15, 2014.
- Van Kampen, J. M., Baranowski, D., & Kay, D. G. (2014). Progranulin gene delivery protects dopaminergic neurons in a mouse model of Parkinson's disease. *PLoS ONE*, 9(5), e97032. doi:10.1371/journal.pone.0097032.
- Wakade, C. G., Mahadik, S. P., Waller, J. L., & Chiu, F.-C. (2002). Atypical neuroleptics stimulate neurogenesis in adult rat brain. *Journal of Neuroscience Research*, 69(1), 72–79. doi:10.1002/jnr.10281.
- Wang, L.-J., Lu, Y.-Y., Muramatsu, S., Ikeguchi, K., Fujimoto, K., Okada, T., et al. (2002). Neuroprotective effects of glial cell line-derived neurotrophic factor mediated by an adeno-associated virus vector in a transgenic animal model of amyotrophic lateral sclerosis. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, 22(16), 6920–6928.
- Wang, Q., Sun, A. Y., Simonyi, A., Jensen, M. D., Shelat, P. B., Rottinghaus, G. E., et al. (2005a). Neuroprotective mechanisms of curcumin against cerebral ischemia-induced neuronal apoptosis and behavioral deficits. *Journal of Neuroscience Research*, 82(1), 138–148. doi:10.1002/jnr.20610.
- Wang, T.-W., Zhang, H., & Parent, J. M. (2005b). Retinoic acid regulates postnatal neurogenesis in the murine subventricular zone-olfactory bulb pathway. *Development (Cambridge, England)*, 132(12), 2721–2732. doi:10.1242/dev.01867.
- Wang, X.-L., Yang, Y.-J., Xie, M., Yu, X.-H., Liu, C.-T., & Wang, X. (2007a). Proliferation of neural stem cells correlates with Wnt-3 protein in hypoxic-ischemic neonate rats after hyperbaric oxygen therapy. *NeuroReport*, 18(16), 1753–1756. doi:10.1097/WNR.0b013e3282f0ec09.
- Wang, Y., Mao, X. O., Xie, L., Banwait, S., Marti, H. H., Greenberg, D. A., & Jin, K. (2007b). Vascular endothelial growth factor overexpression delays neurodegeneration and prolongs survival in amyotrophic lateral sclerosis mice. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, 27(2), 304–307. doi:10.1523/JNEUROSCI.4433-06.2007.
- Wang, X.-L., Yang, Y.-J., Xie, M., Yu, X.-H., & Wang, Q.-H. (2009). [Hyperbaric oxygen promotes the migration and differentiation of endogenous neural stem cells in neonatal rats with hypoxic-ischemic brain damage]. *Zhongguo dang dai er ke za zhi = Chinese Journal of Contemporary Pediatrics*, 11(9), 749–52. <http://www.ncbi.nlm.nih.gov/pubmed/19755026>. Accessed July 24, 2014.
- Webster, N. J. G., & Pirrung, M. C. (2008). Small molecule activators of the Trk receptors for neuroprotection. *BMC Neuroscience*, 9 Suppl 2(Suppl 2), S1. doi:10.1186/1471-2202-9-S2-S1.
- Weissmiller, A. M., & Wu, C. (2012). Current advances in using neurotrophic factors to treat neurodegenerative disorders. *Translational Neurodegeneration*, 1(1), 14. doi:10.1186/2047-9158-1-14.
- Wernig, M., Zhao, J.-P., Pruszak, J., Hedlund, E., Fu, D., Soldner, F., et al. (2008). Neurons derived from reprogrammed fibroblasts functionally integrate into the fetal brain and improve symptoms of rats with Parkinson's disease. *Proceedings of the National Academy of Sciences of the United States of America*, 105(15), 5856–5861. doi:10.1073/pnas.0801677105.
- Wichterle, H., Lieberam, I., Porter, J. A., & Jessell, T. M. (2002). Directed differentiation of embryonic stem cells into motor neurons. *Cell*, 110(3), 385–97. <http://www.ncbi.nlm.nih.gov/pubmed/12176325>. Accessed May 27, 2014.
- Wu, D. (2005). Neuroprotection in experimental stroke with targeted neurotrophins. *NeuroRx: the journal of the American Society for Experimental NeuroTherapeutics*, 2(1), 120–128. doi:10.1602/neurrx.2.1.120.
- Xia, C.-F., Boado, R. J., Zhang, Y., Chu, C., & Pardridge, W. M. (2008). Intravenous glial-derived neurotrophic factor gene therapy of experimental Parkinson's disease with Trojan horse liposomes and a tyrosine hydroxylase promoter. *The Journal of Gene Medicine*, 10(3), 306–315. doi:10.1002/jgm.1152.
- Xie, Y., Ye, L., Zhang, X., Cui, W., Lou, J., Nagai, T., & Hou, X. (2005). Transport of nerve growth factor encapsulated into liposomes across the blood-brain barrier: In vitro and in vivo studies. *Journal of Controlled Release: Official Journal of the Controlled Release Society*, 105(1–2), 106–119. doi:10.1016/j.jconrel.2005.03.005.

- Yanamoto, H., Nagata, I., Sakata, M., Zhang, Z., Tohnai, N., Sakai, H., & Kikuchi, H. (2000). Infarct tolerance induced by intracerebral infusion of recombinant brain-derived neurotrophic factor. *Brain Research*, 859(2), 240–248. <http://www.ncbi.nlm.nih.gov/pubmed/10719070>. Accessed July 1, 2014.
- Yang, K., Clifton, G. L., & Hayes, R. L. (1997). Gene therapy for central nervous system injury: the use of cationic liposomes: an invited review. *Journal of Neurotrauma*, 14(5), 281–297. <http://www.ncbi.nlm.nih.gov/pubmed/9199395>. Accessed September 21, 2014.
- Yang, F., Lim, G. P., Begum, A. N., Ubeda, O. J., Simmons, M. R., Ambegaokar, S. S., et al. (2005). Curcumin inhibits formation of amyloid beta oligomers and fibrils, binds plaques, and reduces amyloid in vivo. *The Journal of biological chemistry*, 280(7), 5892–5901. doi:10.1074/jbc.M404751200.
- Yang, K.-Y., Lin, L.-C., Tseng, T.-Y., Wang, S.-C., & Tsai, T.-H. (2007). Oral bioavailability of curcumin in rat and the herbal analysis from *Curcuma longa* by LC-MS/MS. *Journal of Chromatography, B: Analytical Technologies in the Biomedical and Life Sciences*, 853(1–2), 183–189. doi:10.1016/j.jchromb.2007.03.010.
- Yin, D., & Zhang, J. H. (2005). Delayed and multiple hyperbaric oxygen treatments expand therapeutic window in rat focal cerebral ischemic model. *Neurocritical Care*, 2(2), 206–211. doi:10.1385/NCC:2:2:206.
- Yin, D., Zhou, C., Kusaka, I., Calvert, J. W., Parent, A. D., Nanda, A., & Zhang, J. H. (2003). Inhibition of apoptosis by hyperbaric oxygen in a rat focal cerebral ischemic model. *Journal of Cerebral Blood Flow and Metabolism: Official Journal of the International Society of Cerebral Blood Flow and Metabolism*, 23(7), 855–864. doi:10.1097/01.WCB.0000073946.29308.55.
- Yoo, M., Joung, I., Han, A. M., Yoon, H. H., & Kwon, Y. K. (2007). Distinct effect of neurotrophins delivered simultaneously by an adenoviral vector on neurite outgrowth of neural precursor cells from different regions of the brain. *Journal of Microbiology and Biotechnology*, 17(12), 2033–2041. <http://www.ncbi.nlm.nih.gov/pubmed/18167452>. Accessed June 17, 2014.
- Yurek, D. M., Fletcher, A. M., Smith, G. M., Seroogy, K. B., Ziady, A. G., Molter, J., et al. (2009). Long-term transgene expression in the central nervous system using DNA nanoparticles. *Molecular Therapy: The Journal of the American Society of Gene Therapy*, 17(4), 641–650. doi:10.1038/mt.2009.2.
- Zeng, X., Chen, J., Deng, X., Liu, Y., Rao, M. S., Cadet, J.-L., & Freed, W. J. (2006). An in vitro model of human dopaminergic neurons derived from embryonic stem cells: MPP + toxicity and GDNF neuroprotection. *Neuropsychopharmacology: Official Publication of the American College of Neuropsychopharmacology*, 31(12), 2708–2715. doi:10.1038/sj.npp.1301125.
- Zhang, S. C., Wernig, M., Duncan, I. D., Brüstle, O., & Thomson, J. A. (2001). In vitro differentiation of transplantable neural precursors from human embryonic stem cells. *Nature Biotechnology*, 19(12), 1129–1133. doi:10.1038/nbt1201-1129.
- Zhang, T., Yang, Q.-W., Wang, S.-N., Wang, J.-Z., Wang, Q., Wang, Y., & Luo, Y.-J. (2010). Hyperbaric oxygen therapy improves neurogenesis and brain blood supply in piriform cortex in rats with vascular dementia. *Brain Injury : [BI]*, 24(11), 1350–1357. doi:10.3109/02699052.2010.504525.
- Zhang, X.-Y., Yang, Y.-J., Xu, P.-R., Zheng, X.-R., Wang, Q.-H., Chen, C.-F., & Yao, Y. (2011a). The role of β-catenin signaling pathway on proliferation of rats neural stem cells after hyperbaric oxygen therapy in vitro. *Cellular and Molecular Neurobiology*, 31(1), 101–109. doi:10.1007/s10571-010-9559-z.
- Zhang, Y., Wang, J., Chen, G., Fan, D., & Deng, M. (2011b). Inhibition of Sirt1 promotes neural progenitors toward motoneuron differentiation from human embryonic stem cells. *Biochemical and Biophysical Research Communications*, 404(2), 610–614. doi:10.1016/j.bbrc.2010.12.014.
- Zhang, F., Kang, Z., Li, W., Xiao, Z., & Zhou, X. (2012). Roles of brain-derived neurotrophic factor/tropomyosin-related kinase B (BDNF/TrkB) signalling in Alzheimer's disease. *Journal of Clinical Neuroscience: Official Journal of the Neurosurgical Society of Australasia*, 19(7), 946–949. doi:10.1016/j.jocn.2011.12.022.
- Zhang, E., Shen, J., & So, K. F. (2014). Chinese Traditional Medicine and Adult Neurogenesis in the Hippocampus. *Journal of Traditional and Complementary Medicine*, 4(2), 77–81. doi:10.4103/2225-4110.130372.
- Zhao, C., Deng, W., & Gage, F. H. (2008). Mechanisms and functional implications of adult neurogenesis. *Cell*, 132(4), 645–660. doi:10.1016/j.cell.2008.01.033.
- Zhu, D. Y., Lau, L., Liu, S. H., Wei, J. S., & Lu, Y. M. (2004). Activation of cAMP-response-element-binding protein (CREB) after focal cerebral ischemia stimulates neurogenesis in the adult dentate gyrus. *Proceedings of the National Academy of Sciences of the United States of America*, 101(25), 9453–9457. doi:10.1073/pnas.0401063101.
- Zhu, G., Chen, G., Shi, L., Feng, J., Wang, Y., Ye, C., et al. (2014). PEGylated rhFGF-2 conveys long-term neuroprotection and improves neuronal function in a rat model of Parkinson's disease. *Molecular Neurobiology*, . doi:10.1007/s12035-014-8750-5.
- Zuccato, C., & Cattaneo, E. (2007). Role of brain-derived neurotrophic factor in Huntington's disease. *Progress in Neurobiology*, 81(5–6), 294–330. doi:10.1016/j.pneurobio.2007.01.003.