Maternal Nutritional Status and Development of Atopic Dermatitis in Their Offspring



Chun-Min Kang^{1,2} · Bor-Luen Chiang³ · Li-Chieh Wang¹

Published online: 10 March 2020 © Springer Science+Business Media, LLC, part of Springer Nature 2020

Abstract

Atopic dermatitis (AD) is the leading chronic skin inflammatory disease and the initial manifestation of atopic march. Available evidence supports the notion that primary prevention early in life leads to a decreased incidence of AD, thus possibly decreasing the subsequent occurrence of atopic march. Nutritional status is essential to a proper functioning immune system and is valued for its important role in AD. Essential nutrients, which include carbohydrates, proteins, lipids, vitamins, and minerals, are transferred from the mother to the fetus through the placenta during gestation. Various nutrients, such as polyunsaturated fatty acids (PUFAs) and vitamin D, were studied in relation to maternal status and offspring allergy. However, no strong evidence indicates that a single nutrient or food in mothers' diet significantly affects the risk of childhood AD. In the light of current evidence, mothers should not either increase nor avoid consuming these nutrients to prevent or ameliorate allergic diseases in their offspring. Each essential nutrient has an important role in fetal development, and current government recommendations suggest specific intake amounts for pregnant women. This review discusses evidence on how various nutrients, including lipids (monounsaturated fatty acids, PUFAs, saturated fatty acids, and short-chain fatty acids), carbohydrates (oligosaccharides and polysaccharides), proteins, vitamins (A, B, C, D, and E), and trace minerals (magnesium, iron, zinc, copper, selenium, and strontium) in maternal status are associated with the development of AD and their possible mechanisms.

Keywords Atopic dermatitis · Carbohydrate · Lipid · Maternal status · Mineral · Protein · Vitamin

Introduction

Atopic dermatitis (AD) or atopic eczema, similar to other atopic disorders such as asthma and allergic rhinoconjunctivitis, is a growing problem worldwide, affecting approximately 10–20% of young children globally. AD is an inflammatory disease characterized by pruritic skin lesions, immunodysregulation, disrupted epidermal barrier function, and immunoglobulin E (IgE)–mediated sensitization to food and environmental allergens [1]. AD is a complex inflammatory process in which innate and adaptive immune cells contribute to the complex immune network underlying cutaneous inflammation. The imbalance between T helper (Th) 1 and Th2 cells results in an increased secretion of interleukin (IL)-4, IL-5, and IL-13 by Th2-expressing cells [2]. The damage to and dysfunction of the epidermal barrier and the lack of antimicrobial peptides on the skin surface lead to a significantly increased risk of skin infections in AD patients [3]. The chronic relapsing inflammation of skin and the disrupted skin barrier cause severe itchiness and wound infection, largely impairing the quality of life [4, 5]. As a multifactorial disease and given that approximately 60% of the onset of childhood AD occurs before the age of 1 year, a variety of factors in the early years of life are being studied [5]. AD commonly develops as the initial manifestation of atopic march, and sensitization through the skin is likely an important initial step in the development of other allergic diseases [6]. Evidence indicates that primary prevention through the application of emollients early in life leads to a decreased incidence of AD [7], thus possibly decreasing the subsequent development of atopic march. Nutrition, as a major environmental factor, is valued for its important role in AD. Maternal diet and antenatal/ perinatal nutrition have gained interest because of their

Li-Chieh Wang lcwang5@ntu.edu.tw

Department of Pediatrics, National Taiwan University Hospital, No. 7, Chung Shan South Road, Taipei 10002, Taiwan, Republic of China

² Department of Laboratory Medicine, National Taiwan University Hospital, Taipei, Taiwan

³ Department of Medical Research, National Taiwan University Hospital, Taipei, Taiwan

considerable influence on the nutrition status during the first year of life, which may in turn alter the immune response, thus manipulating the chances of developing future atopy [8-10].

Essential nutrients, which include carbohydrates, proteins, lipids, vitamins, and minerals, are transferred from the mother to fetus through the placenta during gestation. Maternal nutrition status could thus substantially affect fetal development by altering fetal genome expression and consequently give rise to diseases after birth or later in adulthood, a phenomenon termed "developmental programming" or "fetal programming" [11]. Various food categories, including vegetables, fruits, dairy products, nuts, fish, oil, and antioxidants, were studied for the associations between maternal intake and offspring atopy in the past few decades [8, 12–18]. However, clinicians only recently focused more attention to the effect of individual nutrients. Maternal diet may also affect the content of breast milk, in which certain food allergens such as peanut could be detected [19]. The intake of dietary supplements in large amounts, for example, vitamin D, may also elevate its level in breast milk [20]. However, whether the changes in breast milk content may alter the incidence of AD development among breastfed children remains controversial [21, 22]. In this article, we will review individual nutrients in maternal diet during pregnancy and/or breastfeeding and their effect on the development of AD in the offspring.

Lipids

Lipids are main constituents of human body cells and play an important part in physiological functions; thus, they are critically required for fetal development [23]. Given the natural deficiency of certain fatty acid desaturases in human beings, we cannot synthesize specific crucial fatty acids, that is, essential fatty acids, and must acquire them from dietary sources. Fatty acids directly influence the behavior of a number of proteins involved in immune cell activation, including those associated with T cell responses, antigen presentation, and fatty acid-derived inflammatory mediator production [24]. Fatty acids can have powerful antiinflammatory and immunomodulatory activities in a wide array of diseases (e.g., autoimmunity, arthritis, and infection) [25]. Studies reported the relation of fatty acids and development of allergies. Saturated, unsaturated, and short-chain fatty acids (SCFAs) will be discussed here.

Saturated Fatty Acids

Saturated fatty acids (SFAs) refer to the fatty acids without double bonds. These molecules are mostly present in animal fat products and certain plant oils. The modern Western dietary pattern is rich in foods of animal origin, such as red meat and dairy products, which contain abundant SFAs. In contrast to polyunsaturated fatty acids (PUFAs), SFAs are usually considered "bad fats" because their high intake is associated with a variety of diseases, including cardiovascular diseases, metabolic syndrome, or cancer [26]. Palmitic acid (PA) is the most common SFA in the human body [27], accounting for around 65% of SFAs and 28–32% of the total fatty acids in serum [28]. PA can posttranslationally modify proteins in a process called palmitoylation, in which PA is covalently linked to proteins through a thioester bond, thus regulating protein function and performing pathogenic roles in metabolic syndrome, cardiovascular diseases, cancer, neurodegenerative diseases, and inflammation [29]. PA also promotes cell apoptosis and autophagy through stimulation of phosphorylation of mitogenactivated protein kinases and AMP-activated protein kinase, inhibition of the phosphorylation of Akt and mammalian target of rapamycin [30], and stimulation of NO production through the production of superoxide, nuclear factor (NF)-KB activation, and increase in inducible NO synthase protein content [31]. PA can stimulate an inflammatory response through the Toll-like receptor (TLR) 4 signaling pathway [32].

The quantification of SFA intake is difficult, although several serum markers, such as myristic acid, have been determined as good reflectors of dietary intake [33]. Most studies approximate the amount of SFA intake by detailed record of daily diet with further calculation. Other research relates the consumption of SFAs to atopic diseases, but the findings are inconsistent [34, 35]. A study on adolescents showed a positive association between SFA intake and asthma [36]. Hoppu et al. in Finland used a software to quantify different nutrient proportions in breastfeeding mothers [37]. They observed that atopic mothers had higher percentage of fat and SFAs and lower percentage of carbohydrates of total energy intake than non-atopic mothers; a higher SFA intake during breastfeeding was associated with higher atopic sensitization of infants in terms of skin-prick test (odds ratio (OR) 1.16; 95% confidence interval (CI) 1.001–1.36; p = 0.048) [37]. Saito et al. conducted a questionnaire-based study on 771 Japanese motherinfant pairs and observed that maternal intake of either PUFAs, SFAs, or monounsaturated fatty acids (MUFAs) showed no association with atopic eczema in infants aged 3-4 months [38]. Barman et al. demonstrated that children with atopic eczema or other allergic diseases had significantly higher cord blood PUFAs levels, whereas those who remained non-allergic at the age of 13 had lower cord blood PUFAs levels and higher SFA and MUFA levels [39]. The authors also reported no correlation in most fatty acid levels between mothers' and children's sera except for one in the long-chain (LC) PUFA species. The result agrees with the assumption of previous studies indicating that SFAs and MUFAs mainly enter the fetal circulation by passive diffusion [40]. No conclusion could be made for the role of dietary SFAs in childhood AD based on the current evidence. Nevertheless, daily SFA

intake per person, whether pregnant or not, is designated to a limited amount by many health authorities because of its evident association with other health problems [41].

Monounsaturated Fatty Acids

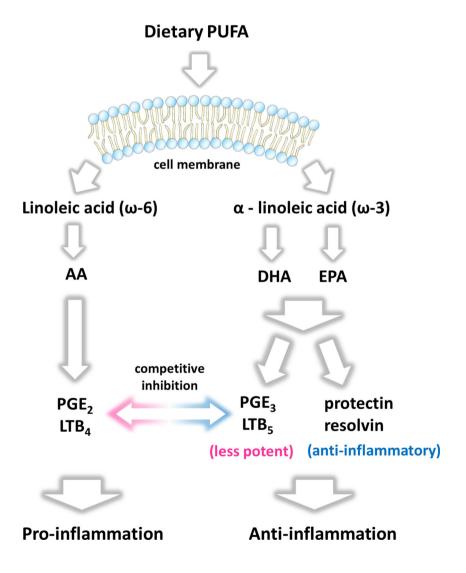
Oleic acid is the most abundant MUFA in the human diet (C18:1 n-9). In Mediterranean countries, olive oil is the main source of MUFAs. Other oil sources of MUFAs include canola, peanut, sunflower, corn, soybean, and safflower oil [42]. MUFAs are associated with atopy development. A European ecological study [43] showed that a high MUFA intake (palmitoleic and oleic acids) was positively associated with sensitization prevalence in 20–44-year-old adults (OR 1.59; p = 0.035). In a German prospective study, Nagel et al. [44] revealed a significantly positive association between the dietary intake of oleic acid (C18:1) and hay fever in adulthood (OR 2.86; 95% CI 1.22–6.70; p = 0.04). However, the mechanism of MUFAs has not been studied, and the association

Fig. 1 Mechanism of PUFAs. The ω -6 PUFAs, such as AA, are precursors for PGE₂ and LTB₄, which are proinflammatory mediators and chemoattractants. The w-3 PUFAs, such as EPA and DHA, are precursors for PGE₃ and LTB₅, which compete with AA in the synthesis of PGE2 and LTB₄, respectively. The less potent eicosanoids and antiinflammatory mediators, such as protectin and resolvin, reduce the inflammatory process and exert a protective role in various diseases

between maternal MUFA intake and offspring atopy has not been well studied.

Polyunsaturated Fatty Acids

The main categories of PUFAs are ω -3 LC PUFAs and ω -6 LC PUFAs, and both are characterized by multiple double bonds in the long hydrocarbon chain, with the first double bond located between the third and fourth carbons or between the sixth and seventh carbons, respectively. Both classes of PUFAs are important (Fig. 1). Omega-6 PUFAs are subject to consumption for energy and the major ingredients for proinflammatory eicosanoids [45]. The metabolites of ω -6 PUFAs include the eicosanoids prostaglandin (PG)E₂ and leukotriene (LT)B₄, which are proinflammatory and neutrophil chemotactic agents [46]. The modern Western diet often provides more than the necessary amount of ω -6 PUFAs, and excessive ω -6 PUFAs are associated with a variety of health problems, including obesity, cardiovascular diseases, and proinflammation status [45, 47]. Omega-3 PUFAs, on the other



hand, have gained vast interests over the years because of their multiple health benefits [48]. The incorporation of ω -3 PUFAs into the lipid bilayer increases the membrane fluidity, thereby enhancing membrane-mediated processes, such as phagocytosis and endosome/exosome formation, which regulates the function of immune cells [49]. Omega-3 PUFAs might inhibit the production of PGE₂, which is derived from arachidonic acid (AA), and Th2 responses [50], affect the TLRs, and decrease the transcription of proinflammatory genes, influencing the immunologic response [51-54]. Given that ω -3 and ω -6 fatty acids are metabolized by the same enzymatic pathway in a competitive manner, an imbalanced ω -6 PUFA-to- ω -3 PUFA ratio in the diet is implicated in various diseases [55, 56]. A high proportion of ω -3 LC PUFAs in diet lowers the incidence of cancer, cardiovascular, and neurological diseases [47, 48] and possibly alleviates the inflammatory process and allergic reaction [57-59].

The most well-known members of the w-3 LC PUFA family are the 20-carbon eicosapentaenoic acid (EPA) and the 22carbon docosahexaenoic acid (DHA). Oily fish (salmon, tuna, mackerel, sardines, herring, etc.) or commercial fish oil are abundant sources of EPA and DHA. The intake of fish oil had been proven to alter the composition of cell membrane lipids. In a study by Yaqoob et al. in 2000, regular supplementation with fish oil capsule (2.1 g EPA + 1.1 g DHA) significantly increased the percentage of EPA in the plasma membrane in 4 weeks (from 0.5 to 3.7%), replacing the relatively abundant 20-carbon AA (2.0 to 1.3%), which belongs the ω -6 PUFA family and is also the main ingredient of pro-allergic eicosanoids, namely, PGs and LTs [60]. The substitution of AA to EPA dampens the prostanoid signaling pathway, most significantly the PG endoperoxide H synthase-1, which is responsible for downstream products that finally convert into PGDs, PGEs, and PGFs [61]. EPA is also a substrate to 5lipoxygenase which competes with AA in the pathway of transforming AA into LTB_4 [62]. When supplemented to healthy pregnant women, w-3 LC PUFAs not only increase EPA and DHA levels in serum or plasma, lowering the levels of the ω -6 LC PUFA family, but also significantly decrease the PGE₂ level. Prenatal supplementation of ω -3 LC PUFAs is recommended in health programs of several countries based on proofs of their multiple health benefits. The European Food Safety Authority has recommended in their 2010 report the supplementation of 100-200 mg preformed DHA in addition to the daily adequate intake (250 mg EPA + DHA daily) for mothers during pregnancy and lactation [63]. Guidelines from the UK government (Scientific Advisory Committee on Nutrition & Committee on Toxicity) also recommend that pregnant and lactating women should consume 1-2 portions (around 140 g each) of fish per week, with at least one being an oily fish. One to two portions of oily fish per week will provide ω -3 LC PUFAs approximately equal to 450 mg EPA and DHA daily [64]. However, an amount exceeding the upper limit of two portions of oily fish per week leads to the intake of additional contaminants, particularly methylmercury [65].

Scientific evidence shows that alteration of PUFA ratio in maternal diet changes the serum fatty acid profile and cytokine/chemokine profiles of the fetus and the offspring risk of AD (Table 1). Dunstan et al. first examined in 2003 the effect of fish oil supplements in Australian pregnant mothers on their children [66]. In this double-blind, placebo-controlled, randomized clinical trial (DBPCRCT), 40 atopic mothers received fish oil capsules, and 43 in the control group received olive oil capsules from 20 weeks of gestation until delivery. As a result, the fatty acid composition of neonate red blood cell showed significantly increased ω -3 PUFAs in the fish oil group compared with the control group; the plasma IL-13 level of the fish oil group was significantly lower, whereas no differences were observed in other cytokines (IL-4, IL-5, IL-6, IL-10, IL-12, tumor necrosis factor (TNF)- α , and interferon (IFN)- γ) or IgE levels between groups [66]. On the other hand, in a later German multicenter, DBPCRCT in 2008, Krauss-Etschmann et al. studied Th1 and Th2-related molecules in the cord blood and observed the increased mRNA for transforming growth factor (TGF)- β , a cytokine associated with induction of tolerance, and decreased mRNA for Th2-related cytokines IL-4 and IL-13 in the fish oilsupplemented group (p < 0.01) [71]. Other studies showed the correlation of oily fish intake/supplementary fish oil with decreased inflammation- or allergy-associated cytokine PGE₂ and LTB₄ and increased T cell protein kinase C ζ (PKC ζ), a kinase isoform which exists in significantly low levels in neonates with allergic diseases, particularly eczema [67, 68, 72, 95]. The changes in cytokine profile by maternal supplementation of EPA or DHA are implicative of the antiallergic role of prenatally administered ω -3 LC PUFAs.

Several studies investigated the direct association of clinically diagnosed childhood AD with modification of maternal dietary fatty acid composition during prenatal stage or lactation. However, the results were inconsistent. Furuhjelm et al. conducted a DBPCRCT in Sweden involving 145 pregnant women (70 supplied with daily 1.6 g EPA + 1.1 g DHA and 75 with placebo) and followed up their 117 children up to 3 years old [74]. A lower incidence of IgE-associated eczema was detected in the ω -3 LC PUFA supplement group (p < 0.05). Interestingly, the ω -6/ ω -3 LC PUFA ratio of the mothers' serum showed no statistically significant difference in the study [74]. Other clinical trials, including a larger trial by Palmer et al. involving 706 mother-infant pairs, had failed to find clinical differences in the incidence of offspring AD between intervention and placebo groups [69, 72, 75]. The results of observational studies also varied. Many studies generated promising, statistically significant results, indicating that maternal ω -3 LC PUFA supplement reduces the risk of childhood AD [8, 14, 17, 77, 79-81, 83]. A recent

Interventional study		
Study group	Study design/intervention/population	Significant results about AD in intervention group compared to placebo/control
"The Infant Fish Oil Supplementation Study (IFOS)" in Perth, Australia [66–70]	Study design: DBPCRCT Intervention: fish oil capsule vs placebo Duration: GA 20 weeks until delivery Population and size: atopic pregnant mothers (n = 98)	 ↑ ω-3 PUFA level in erythrocyte membrane [66] ↓ Plasma IL-13 [66] ↓ IL-6, IL-10 production in neutrophils [67] ↓ LTB₄ (only trend) production in neutrophils [67] ↑ PKCζ expression in neonatal T cells [68] ↓ AD severity [69] ↔ No difference in frequency of AD [69]
In Neuherberg, Germany [71]	Study design: DBPCRCT, multicenter Intervention: fish oil (DHA + EPA) vs placebo Duration: GA 22 weeks until delivery Population and size: pregnant women ($n = 311$)	 ↑ mRNA of TGF-β in maternal and cord blood ↓ IFN-γ in mothers' blood ↓ mRNA of IL-4, IL-13, and CCR4 in cord blood ↓ Natural killer cell and CD8⁺ T cells in cord blood
"The Salmon in Pregnancy Study (SiPS)" in Southampton, UK [72, 73]	 Study design: randomized, single-blind controlled trial Intervention: 2 portions of oily fish/week vs habitual diet Duration: GA 20 weeks until delivery Population and size: pregnant women (n = 123) 	 ↑ DHA and EPA intake [73] ↑ Percentage of DHA and EPA in maternal and cord plasma phosphatidylcholine [73] ↓ IL-2, IL-4, IL-5, IL-10, TNF-α, and PGE₂ production in cord blood mononuclear cell [72] ↔ No differences in IgE and skin prick tests [72]
In Linköping, Sweden [74]	Study design: randomized placebo-controlled tri- al Intervention: DHA + EPA vs placebo Duration: GA 25 weeks to 3–4 months of breastfeeding	 ↓ Prevalence of food allergy at 1 year old (2% vs 15%) ↓ Incidence of IgE-associated eczema (8% vs 24%)
"The Docosahexaenoic Acid to Optimize Mother Infant Outcome (DOMInO) Trial" in Adelaide, Australia [75, 76]	Population and size: pregnant women $(n = 145)$ with positive family history of atopic diseases Study design: randomized controlled trial Intervention: fish oil capsule vs vegetable oil capsule Duration: GA 21 weeks until delivery Population and size: mother-infant dyads (n = 706) at high risk of atopic diseases	 ↓ Percentage of atopic eczema (7% vs 12%), not significant after adjustment ↓ Sensitization to egg (9% vs 15%) ↔ No difference in IgE associated allergies
Observational study		
Study group	Study design/methods/population	Significant results about AD
In Aberdeen, UK [17]	 Study design: population-based study, single center Methods: questionnaire, skin prick tests and blood tests since GA 12 weeks until 5 years after birth Population and size: mother-children dyads (n = 1924) 	↓ Doctor-confirmed eczema for maternal fish consumption
"The Kyushu Okinawa Maternal and Child Health Study (KOMCHS)" in Fukuoka, Japan [77, 78]	Study design: prospective prebirth cohort study Methods: self-administered questionnaire Population and size: Japanese mother–child pairs (n = 1354)	For maternal intake of EPA and EPA + DHA: ↓ Infantile wheeze ↔ No relation with infantile eczema
In Krakow, Poland [79]	(n - 1504) Study design: prospective birth cohort study Methods: detailed, standardized, face-to-face in- terview every 3 months after delivery to 1 year Population and size: mothers giving birth to term babies ($n = 469$)	↓ Risk of infantile eczema ^a for maternal fish consumption
In Sabadell, Spain [80]	Study design: population-based birth cohort study, single center Methods: blood sampling at GA 12 weeks and questionnaire when the child is 6–14 years old Population and size: non-atopic mothers ($n = 211$) and children	↓ Atopic eczema for maternal LC PUFA ↓ Atopic eczema for cord blood DHA, total ω-3 and ω-3 LC PUFAs
"The Generation R Study" in Rotterdam, the Netherlands [81, 82]	Study design: prospective population-based co- hort study	

	Methods: questionnaire, blood tests and lung function tests Population and size: mothers from early pregnancy to post-delivery, in pairs with	 ↑ Risk of childhood eczema for maternal total PUFA (OR 1.16) and total ω-6 PUFA (OR 1.21) ↔ No association with total ω-3 PUFA or ω-63
	children ($n = 4976$)	ω-3 PUFA ratio
"The Urban Child Institute CANDLE Study" in Memphis, TN, USA [83]	Study design: prospective prenatal cohort study, single center	↑ AD in children of maternal atopy for higher 2nd trimester ω -6 PUFAs
	Population and size: racially diverse mother–infant dyads (<i>n</i> = 1131)	\leftrightarrow No association with prenatal ω -3 PUFAs and ω -6: ω -3 PUFAs
"The Finnish Type 1 Diabetes Prediction and Prevention (DIPP) Nutrition Study" in Finland [84, 85]	Study design: multidisciplinary population-based prospective birth cohort study, multicenter Methods: questionnaire (181 item FFQ) Population and size: newborn infants with HLA-conferred susceptibility to type 1 diabe- tes (<i>n</i> = 2441)	fats and fatty acids after adjustment for confounders
"The Avon Longitudinal Study of Parents and Children (ALSPAC)" in Avon, UK [86, 87]	Study design: prospective population-based co- hort study, multicenter Methods: questionnaire and blood tests, including cord blood Population and size: 14,541 pregnancies resulting	 ↑ Eczema at 18–30 months old for the ratio of AA: EPA in cord blood RBC (OR 1.14) ↔ No longer significant after adjustment
"The Growing Up in Singapore Toward healthy Outcome (GUSTO) birth cohort" in Singapore [88, 89]	in 14,062 live births (born to 13,866 mothers). n = 1238 and $n = 2945$ for cord and maternal analyses, respectively Study design: prospective population-based co- hort study Methods: detailed interview and blood tests Population and size: mother–infant pairs, n = 1162; $n = 883$ analyzed for eczema out- come.	↔ No association with offspring rhinitis, eczema, wheezing in maternal total ω-3, ω-6 PUFA status and the ω-6: ω-3 PUFA ratio after adjustment
"Life-style Related Factors on the Immune System and the Development of Allergies in Childhood PLUS the influence of traffic emissions and genetics (LISAplus) study" in Munich, Germany [90, 91]	 Study design: prospective population-based birth cohort study Methods: questionnaires completed at 2, 6, and 10 years old. Cord blood and blood tests. Population and size: children (<i>n</i> = 436) from the Munich LISAplus birth cohort 	↔ No association with eczema or other allergic diseases in ω-3 LC PUFA, ω-6 LC PUFA, or the ω-6: ω-3 ratio in cord blood
"Child, Parent and health: Lifestyle and Genetic constitution (KOALA) cohort" in the Netherlands [92, 93]	 Study design: prospective population-based birth cohort study Methods: blood tests for mother at GA 34–36 weeks and for children at age 24 months. Repeated parental questionnaires. Home-visit by trained nurse for atopic dermatitis at age 24 months Population and size: mothers (<i>n</i> = 1275) and children (<i>n</i> = 807 for home visit and <i>n</i> = 951 for follow-up at 6–7 years) from the KOALA cohort 	 ↓ Risk of eczema in the child with high ratio of maternal ω-6: ω-3 LC PUFAs ↓ Risk of eczema in the first 7 months of life with AA
"Perturbateurs endocriniens: Étude Longitudinale sur les Anomalies de la Grossesse, l'Infertilité et l'Enfance (PELAGIE) cohort" in Brittany, France [94]	conort Study design: prospective population-based birth cohort study Methods: questionnaires (FFQ) at 2 years old Population and size: mothers (<i>n</i> = 1500) from the PELAGIE cohort	↔ No association with childhood eczema in maternal seafood consumption

^a The results also showed that exposure to air pollutants prenatally and postnatally positively correlates with the occurrence of infantile eczema

DBPCRCT double-blind, placebo-controlled, randomized clinical trial, *GA* gestational age, *IL* interleukin, *LTB*₄ leukotriene B₄, *PKC* ζ protein kinase C ζ , *DHA* docosahexaenoic acid, *EPA* eicosapentaenoic acid, *TGF*- β transforming growth factor- β , *IFN*- γ interferon- γ , *CCR4* C-C chemokine receptor 4, *TNF*- α tumor necrosis factor- α , *PGE*₂ prostaglandin E₂, *LC PUFA* long-chain PUFA, *FFQ* food frequency questionnaire, *AA* arachidonic acid

observational study published in 2019 by Gardner et al. based on a racially diverse cohort suggested an increased OR (OR 1.25; 95% CI 1.01–1.54) of childhood AD in mothers with increased ω -6 PUFA intake, but no association was found with ω -3 PUFA intake alone nor

with ω -6/ ω -3 PUFA ratio [83]. Other studies either reported no significant differences in the clinical outcomes [84, 86, 88, 90, 123] or inverse findings (that is, ω -3 LC PUFA supplement increased the risk of childhood eczema) [92, 124].

In general, several studies suggest the protective role of low ω -6/ ω -3 PUFA ratio (low ω -6 and/or high ω -3) in maternal diet, whereas other research showed no effect. Meanwhile, several studies obtained the opposite results. Carefully designed large-scale studies with longer follow-up periods may be required to generate more solid results. Ethnic and environmental diversity and different risk stratifications of mothers giving birth to babies with atopic diseases should also be considered in future studies.

Short-Chain Fatty Acids

SCFAs contain less than six carbon atoms, with acetate, propionate, and butyrate being the most common. SCFAs are the main energy sources for enterocyte regeneration; they modulate the enteric microbial community and contribute to increasing host health [125]. SCFAs have antiinflammatory properties, including the reduction in expression and signaling of different proinflammatory cytokines, induction of nitric oxide synthesis and metalloproteinases, reduction and activation of lymphocyte proliferation [126], promotion of regulatory T cell (Treg) generation in the colon of mice [127, 128], increase in intestinal IgA secretion, and improvement of Th1/Th2 ratio [129]. Supplementation of SCFAs at high concentration reduces inflammation through altered cytokine expression and enhances tissue repair and mucus secretion [125], affecting the abundance of bacterial populations in intestines [130].

The potential role of SCFAs in eczema has been addressed recently. One study noted the lower levels of different SCFAs compared with non-allergic controls in 1-year old allergic children [131]. Another study noted the inverse correlation of severity of eczema with the amount of butyrate-producing bacteria (r = -0.52, p = 0.005) in 6-month-old infants [132]. Kim et al. noted the lower amounts of SCFAs in fecal samples of children with later developing eczema in a DBPCRCT of probiotics supplement in pregnant mothers and infants in the first year of life [133]. The intestinal SCFAs may play an important role in the development of eczema in early life, but direct maternal supplementation has not been studied.

Carbohydrates

Carbohydrates are the main energy source of humans. Different sources of carbohydrates have variable digestion rates. Thus, their effects on blood glucose and insulin levels also vary. High glycemic index foods, including rice, white bread, and potatoes, cause a sharp rise in blood glucose levels which declines rapidly, whereas low glycemic index foods, such as fruits or dairy, have slowly digestible carbohydrates that result in a lower postprandial glucose response [134]. Immune cells have various demands for nutrients, including glucose, glutamine, and fatty acids, which are metabolized to

generate adenosine triphosphate for energy expenditure; competition for nutrients of different immune cells may regulate immune responses [135]. Dietary fiber describes a variety of plant-based carbohydrates that are resistant to digestion by human gastrointestinal enzymes. They include soluble fiber (fruits, vegetables, and legumes), insoluble fiber (nuts, wholegrain bread, or cereals), or resistant starch (cooked potato and rice). Diets high in fiber and with a low glycemic index can promote laxation, reduce blood cholesterol, and modulate blood glucose and hence may be beneficial in pregnancy [134]. Saccharides, especially oligosaccharides as prebiotics, will be discussed here.

Monosaccharides/Disaccharides

Free sugars consisting mostly of mono- or disaccharides are frequently added by manufacturers in foods or drinks to sweeten the taste. Additionally, high consumption of sugar had been linked to increased risk of atopic diseases, particularly asthma [136]. However, the correlation between sugarrich foods and the severity of AD lacks clarity [137]. Bedard et al. studied the association between maternal sugar consumption and atopic diseases, suggesting the increased risk of asthma and overall atopy with high sugar intake. However, no significant association was observed between maternal sugar intake and offspring atopic eczema [138].

Oligosaccharides

Prebiotics are a group of food ingredients that are degraded by gut microorganisms and may induce their growth and activity, thus changing the composition of gut microflora. Nondigestible oligosaccharides, most commonly fructooligosaccharides (FOS), galacto-oligosaccharides (GOS), and trans-GOS, comprise mostly of known prebiotics, whereas specific resistant starch or non-carbohydrates could also serve as prebiotics [139]. Prebiotics are non-digestible food ingredients that beneficially influence host health by indirect (acting as a fermentable substrate for specific commensal host bacteria, leading to the release of SCFAs in the gut intestinal tract and influencing various molecular and cellular processes) and direct effects (acting directly on several compartments and specifically on different arrangements of cells (epithelial and immune cells)) [140]. SCFAs, as previously stated, play a role in the immunomodulation of gut mucosa [129]. Prebiotics also directly promote barrier integrity to prevent pathogeninduced barrier disruptions [141]. Absorbed prebiotics may be in direct contact with circulating immune cells. For example, inulin and FOS induce the secretion of IL-10, IL-1β, and TNF- α by blood monocytes [142]. In murine models, prebiotics reinforce gut barrier function and reduce allergic reactions [143, 144]. Compared with the more established effects of probiotics in improving symptoms of AD, however, studies on postnatal prebiotics administration in human yielded inconclusive results [145]. Prenatal administration of prebiotics was studied in a murine model in 2009 by Fujiwara et al., who demonstrated the beneficial effect of supplementation during pregnancy on reducing severity of AD-like skin lesions in mice offspring [146]. A prior study conducted by the same group demonstrated that by supplementing FOS in female mice during pregnancy and lactation, the gut microbiota composition of suckling mice was altered. The result possibly links maternal intake of prebiotics to a preventive role against allergic disease development later in life.

In humans, several clinical trials used prebiotics as an allergy prevention strategy by intervening in infancy by supplementation of infant formula. In World Allergy Organization Guideline for Allergic Disease Prevention, Cuello-Garcia et al. conducted a systematic review including 22 randomized trials with a minimum follow-up of 4 weeks, comparing any type of prebiotics against placebo or no prebiotics [147]. They concluded the high uncertainty of reducing the risk of developing allergies using the currently available evidence on prebiotics supplementation in infancy. To date, no human studies in which prebiotics have been given to mothers during pregnancy and/or breastfeeding have been published; however, two clinical studies are currently being conducted: SYMBA is a DBPCRCT investigating the effects of maternal prebiotics supplementation (GOS:FOS at a ratio of 9:1) from 18 to 20 weeks of gestation during pregnancy until 6 months of lactation on the development of infant allergic disease [148]; PREGRALL is a multicenter DBPCRCT that aims to evaluate the effectiveness of gestational prebiotics supplementation (from 20th week of gestation until delivery, 11.8 g 9:1 GOS/ inulin powder per day) versus placebo on the occurrence of AD at 1 year of age in high-risk children (defined as having a maternal history of atopic disease) [149]. The outcomes of these trials might further verify the effect of maternal prebiotics supplementation on allergic diseases in offspring.

Polysaccharides

Dietary fibers are non-starch polysaccharides consisting of subtypes, including soluble fiber, insoluble fiber, and resistant starch [150, 151]. These polysaccharides are subject to fermentation of gut microbiome [152], leading to the generation of SCFAs which deliver antiinflammatory and immunomodulatory effects in a manner similar to prebiotics. Maternal diet rich in vegetables (especially green- and yellow-leaf vegetables) and fruits during pregnancy is beneficial to offspring with eczema [12, 16]. However, these information do not imply the effect of dietary fibers given that they are inevitably confounded by other coexisting nutrients and antioxidants such as β -carotene. A study conducted by Pretorius et al. [153] followed up over 600 mother–infant pairs and analyzed the relationship between eczema or wheezes, both physician-

diagnosed or parent-reported, and the amount of dietary fiber in different subtypes taken by mothers during gestation. Of all different subtypes, statistical significance was observed in resistant starch, which had a negative correlation with clinical wheezes but a positive correlation with atopic eczema. Thus, high maternal intake of resistant starch is associated with a high risk of parent-reported eczema (adjusted OR (aOR) 1.27; 95% CI 1.09–1.49; p < 0.01) and physician-diagnosed eczema (aOR 1.19; 95% CI 1.01–1.41; p = 0.04). The result was contrary to the hypothesis that AD may be reduced by immunomodulatory effects of SCFAs, which are generated by fermentation of dietary fibers.

Proteins

Proteins are involved in both structural (keratin and collagen) and functional (enzymes, protein transport, and hormones) biological roles [134]. Proteins, out of all kinds of nutrients, are the major allergens for food allergy [154]. Childhood AD is strongly associated with food allergy, with at least 30% of children with moderate to severe AD documented for food allergies [155, 156]. The association between protein intake and AD has long been discussed, and questions have also been raised about whether maternal intake of protein food allergen could possibly cause food allergy in children. Given their importance in fetal development, proteins digested by mothers are generally transferred through the placenta in the form of amino acids by specific amino acid transport proteins [157]. Nevertheless, certain food component proteins, namely, βlactoglobulin (BLG) and ovalbumin (OVA), are detectable in cord blood and placenta tissues of mothers ingesting milk and eggs, indicating their capability to cross the placenta and be delivered directly to the fetus, thereby mediating fetal T cell priming [158–160]. Other studies showed that component proteins, including BLG [161], OVA [162-164], and peanut antigen [19, 165], are also present in breast milk.

Despite proofs of maternal-fetal passage of food proteins, whether maternal intake of certain food proteins is associated with increased childhood allergic diseases remains controversial. Two aspects are being discussed: maternal diet modification during pregnancy and breastfeeding. An exclusion diet (excluding common food allergens, such as cow's milk, egg, nuts, or fish, in several studies) during lactation decreased the level of related component protein levels in breast milk in several studies [166] but caused no difference in antigenspecific antibody production in infants [166, 167]. No difference in antibody profile was found in the fetal cord blood in the comparison of mothers with and without diet restriction during pregnancy [167]. In their 2008 report, American Academy of Pediatrics (AAP) had concluded that data are lacking to affirm the role of maternal exclusion diet during pregnancy and lactation on childhood food allergy, and by

far, no new clinical trial is being conducted to change this conclusion [168]. A 2012 meta-analysis of Cochrane library by Kramer et al. included five articles, with two studies directly addressing the outcomes of offspring atopic eczema, had also drawn a similar conclusion, that is, the exclusion of common food allergen protein in the mother's diet causes no effect on the incidence of childhood allergic reactions, including AD [166, 169, 170]. A systemic review by Netting et al. in 2014 listed seven interventional studies (five RCTs and two nonrandomized comparisons) and sixteen observational studies examining in part the correlation between maternal diet and eczema in children [13]. In the five reviewed RCTs [171–175], including the Isle of Wight prevention study [174], two eliminated cow's milk during pregnancy and lactation in the intervention group and followed up the children's outcomes until 18 months old [172, 175]; one had the intervention group restricted from cow's milk and egg during pregnancy and lactation and followed up the offspring until 5 years of age [171]; one excluded cow's milk, egg, and peanut from diet and limited soy and wheat intake in the intervention group during pregnancy and lactation and followed up the children until 7 years of age [173]; the Isle of Wight study restricted cow's milk, egg, nut, and fish intake in the intervention group during lactation only and followed up the children until 12 months old, with controlled dust mite concentration in the environment and delayed introduction of allergenic food [174]. Overall, these RCTs found no differences in the outcomes in offspring AD between two groups, except for the Isle of Wight study, which addressed a significantly greater prevalence of eczema (OR 3.6; 95% CI 1.0-12.5) in the control group. The two non-randomized studies showed no evidence of maternal restriction diet to improve the outcome on childhood atopic eczema [176, 177]. The results of observational studies were inconsistent. One study suggested that excessive meat consumption by the mother increases the risk of eczema [38]; however, another study failed to reach the same conclusion [123]. Dairy product also causes no effect on the increased risk of eczema [8, 15]. Several studies suggested a possible protective role for maternal fish intake, which echoes the conclusive result of this systemic review, that is, maternal diets rich in fish, fruits, vegetables, vitamin D, and Mediterranean dietary patterns are associated with a low risk for allergic disease in children [8, 13]. However, fish is a source of ω -3 PUFAs and proteins [9, 48]; thus, the causality may very well be that from lipids rather than proteins themselves.

Meanwhile, evidence supporting the protective role of eliminating common protein allergens from maternal diet is lacking. Both AAP and European Academy of Allergy and Clinical Immunology advise a normal diet without restriction for allergenic foods for mothers who are pregnant or breastfeeding [168, 178]. An RCT with large sample size and long follow-up period may be needed to make further suggestions.

Vitamins

Vitamin A and β-Carotene (Provitamin A)

The vitamin A group consists of retinol (the usually called vitamin A) and more than 600 carotenoids [179]. β -Carotene is abundant in plants and fruits and can be converted into vitamin A by dioxygenase in the intestinal mucosa. This compound is a member of the carotenoid group which exhibits a strong antioxidant activity. Vitamin A can influence the immune system by converting into retinoic acid (RA). RA can inhibit Th1 cells to become pathogenic Th17 cells and block the development of innate lymphoid cell (ILC) 2s while promoting the differentiation and expansion of ILC3s and imprinting dendritic cells (DCs) with the ability to produce RA. In the mesenteric lymph node, RA-producing DCs can induce gut tropism on T and B cells and promote the differentiation of Treg. Furthermore, vitamin A can promote Tr1 and Treg differentiation and inhibit NF-KB signaling in macrophages [180].

Many studies surveyed the effect of antioxidants on AD, because oxidative stress takes part in the pathogenesis of AD [181, 182]. However, studies for vitamin A or β -carotene are limited (Table 2). Serum vitamin A levels were reported to be significantly lower in adults AD patients compared with the controls [183]. An Australian study by West et al. [100] investigated the association between maternal intake of β -carotene, vitamin C, vitamin E, zinc, and copper during pregnancy and the risk of offspring atopic diseases, including eczema, wheezing, and food allergy (sample size 300). For β -carotene, the result suggested no association between maternal intake and childhood atopy. Another study based in the UK (sample size 1942) also found no relation in the prenatal intake of β carotene and childhood eczema [101]. Our study with a small sample size also showed that retinol levels were not significantly different in breast milk for 2-4-month-old AD infants and healthy controls [122]. Therefore, further observational studies for vitamin A intake in pregnant and breastfeeding mothers and RCTs with vitamin A supplementation may be needed to clarify the role of maternal vitamin A in offspring AD.

Vitamin B9 (Folate)

Folate, or vitamin B9, is essential for cell metabolism by acting as methyl donor for DNA methylation, an important process carried out by DNA methyl transferases [184]. Folate is abundant in a variety of foods, including dark green vegetables, fruits, nuts, beans, grains, eggs, dairy products, and meat. Folic acid, on the other hand, is an oxidized synthetic form of folate and is usually used as supplement given that it is more stable and better absorbed than folate [9]. The requirement for folate or folic acid increases in pregnant women given the

Table 2 Studies about maternal vitamins and offspring AD

Interventional study			
Study group	Study design/intervention/population		Significant results about AD in intervention group compared to placebo/control
"The Copenhagen Prospective Studies	Study design: DBPCRCT		\downarrow Risk of persistent wheeze through
on Asthma in Childhood 2010 cohort" in Denmark [96]	Intervention: vitamin D ₃ (2400 IU/day) vs		age 3 years
	placebo; all with a prenatal 400 IU vitamin D_3		\leftrightarrow No difference for eczema
	Duration: GA 24 weeks until 1 week after		
	delivery		
	Population and size: mother–child pairs $(n = 623)$		
"The Vitamin D Antenatal Asthma	Study design: DBPCRCT, multicenter		\leftrightarrow Lower the incidence of asthma and
Reduction Trial" in USA [97]	Intervention: daily 4000 IU vitamin D vs		recurrent wheezing at age 3 years,
	placebo; all with a prenatal vitamin containing 400 IU vitamin D		but not significant ↔ No difference for eczema
	Duration: GA 10-18 weeks until delivery		
	Population and size: high-risk mothers for offspring asthma $(n = 881)$		
In London, UK [98]	Study design: randomized controlled trial		\leftrightarrow No difference for atopy and
	Intervention: no vitamin D, 800 IU		eczema risk at age 3
	ergocalciferol daily until delivery or single oral bolus of 200,000 IU cholecalciferol		
	Duration: GA 27 weeks until delivery		
	Population and size: mother–child pairs $(n = 180)$		
In Japan [99]	Study design: DBPCRCT, multicenter		\leftrightarrow No difference in SCORAD at age
	Intervention: vitamin D ₃ supplements (800 IU/day) vs placebo		3 month ↑ Doctor-diagnosed food allergy at
	Duration: 6 months		age 2 years
	Population and size: mothers with exclusively		
	breastfed infants of facial eczema at 1 month ($n = 164$)		
Observational study			
Study group	Study design/methods/population	Study targets	Significant results about AD
"The Infant Fish Oil Supplementation	Study design: prospective observational study	β-Carotene	\downarrow Risk of any diagnosed infant
Study (IFOS)" in Perth, Australia	based on cohort from an associated clinical	Vitamin C	allergic disease and wheeze for
[70, 100]	trial (IFOS)	Vitamin E	higher maternal dietary vitamin C
	Methods: semi-quantitative FFQ and inter- view since GA 28 weeks	(Copper) (Zinc)	intake. ↔ No relationships between vitamin
	Population and size: atopic mothers paired		C and eczema.
	with infants $(n = 300)$		\leftrightarrow No relationships between
			β -carotene, vitamin E or zinc and any allergic outcomes.
			(↓ Risk of eczema for higher dietary copper)
In Aberdeen, UK [101]	Study design: prospective population-based	β-Carotene	\leftrightarrow No association of eczema and
	cohort study, single center	Vitamin C	wheezing in the 1st year
	Methods: semi-quantitative FFQ at GA	Vitamin E	↓ Childhood eczema born to atopic
	34 weeks, 6, 12, 24 months after birth and cord blood sample		mothers during the 2nd year for maternal vitamin E intake
	Population and size: mother children pairs		

Population and size: mother-children pairs

(n = 1924)

Table 2 (continued)

			↑ "Ever wheeze" and eczema during the 2nd year for maternal vitamin C intake
In Turku, Finland [102]	 Study design: prospective observational study, single center Methods: questionnaire and personal interview at GA 35–36 weeks; skin prick test; breast milk collection at 1 month of age; clinical examination for infants at 1, 3, 6, 12 months of age Population and size: breast milk from atopic 	β-Carotene Vitamin C Vitamin E (α-tocopherol)	 make ↓ Risk of atopy in the infant (OR 0.30) for higher concentration of vitamin C in breast milk ↔ No consistent relationship of α-tocopherol and atopy.
"The Generation R Study" in Rotterdam, the Netherlands [103]	 mothers (n = 34) Study design: prospective population-based cohort study Methods: questionnaire and blood tests from early pregnancy to child age of 48 months Population and size: mother-infant pairs (n = 8742) 	Vitamin B9 (folate) Vitamin B12	 ↑ The development of AD (aOR 1.18) for high maternal folate ↑ The development of AD (aOR 1.30) for high maternal vitamin B12
In Perth, Australia [104]	 Study design: prospective population-based cohort study Methods: FFQ and blood tests; pregnant mother recruited in third trimester; Children followed-up to 1 year old Population and size: pregnant women (<i>n</i> = 628) and their infants (<i>n</i> = 484) (partially overlap with IFOS cohort [70]) 	Vitamin B9 (folate)	 ↑ Subsequent eczema for folate supplements (especially > 500 µg folic acid/day) ↑ Sensitization risk for cord blood folate levels < 50 nmol/l and > 75 nmol/l
"Child, Parent and health: Lifestyle and Genetic constitution (KOALA) cohort" in the Netherlands [93, 105]	 Study design: prospective population-based birth cohort study Methods: questionnaires and blood tests Population and size: mother-child pairs (n = 2834) 	Folic acid	↔ No association with any of the atopic outcomes
"Prevention and Incidence of Asthma and Mite Allergy (PIAMA) birth cohort study" in the Netherlands [106]	 Study design: prospective population-based birth cohort study Methods: questionnaires and blood tests; mothers recruited during pregnancy; children followed up to 8 years old Population and size: mother-child pairs (n = 3786) 	Folic acid	↔ No association with eczema or other respiratory allergic outcomes
"The Osaka Matemal and Child Health Study (OMCHS)" in Neyagawa City, Osaka, Japan [107, 108]	 (n = 5780) Study design: prospective population-based birth cohort study Methods: diet history questionnaire; Mothers recruited during pregnancy; Children followed up to 8 years old Population and size: mother-child pairs (n = 763) 	Vitamin B2 Vitamin B6 Vitamin B9 (folate) Vitamin B12	↔ No association with the risk of wheeze or eczema in the offspring for maternal consumption of folate, vitamins B12, B6, and B2 during pregnancy after adjustment
"The Mothers and Children's Environ-mental Health (MOCEH) study" in Seoul, South Korea [109, 110]	 Study design: prospective population-based birth cohort study Methods: questionnaires and blood tests; mothers recruited at mid- or late- pregnancy; children followed up to 24 months old Population and size: mother-child pairs (<i>n</i> = 917) 	Vitamin B9 (folate)	 ↓ Cord blood eosinophil count ↑ Cord blood IL-10 ↓ Risk for offspring lower respiratory tract infections at 6 months of age (aOR 0.50) and AD at 24 months (aOR 0.52)
		Vitamin B9 (folate)	\leftrightarrow No association with childhood AD

"The Conditions Affecting Neurocognitive Development	Study design: prospective population-based birth cohort study		\downarrow Odds of current wheeze at age 3
and Learning in Early Childhood (CANDLE) Study" in Memphis, TN, USA [111, 112]	Methods: blood tests and clinical diagnosis of atopic diseases; Serum folate measured at 2nd and 3rd trimester; Diagnosis of atopic diseases made at 3 years of age.		
	Population and size: mother-child dyads $(n = 858)$		
"The Generation R Study" in Rotterdam, the Netherlands [113]	Study design: prospective population-based cohort studyMethods: questionnaire and blood tests; data collection at birth (cord blood), 4 years old (eczema), and 6 years old (FeNO and Rint)	Vitamin B9 (folate) Vitamin B12 (homocysteine) (<i>MTHFR</i> gene SNP	↔ No association with any of the outcome (eczema, wheezing) at any age for cord blood folate, B12 and homocysteine ↑ Risk of eczema for folate in children
	Population and size: Caucasian children $(n = 2001)$	C0//1 and A1298C)	carrying C677T mutations in MTHFR
"The Southampton Women's Survey (SWS)" in Southampton, UK [114]	Study design: prospective population-based cohort study Methods: blood tests, questionnaire and examination by trained nurse	Nicotinamide (metabolites: kynurenine, kynurenic acid, anthranilic acid,	↔ No association with offspring atopic eczema at age 6 months for maternal nicotinamide and related metabolite
	Population and size: mother-infant pairs $(n = 497)$	tryptophan, N1-methylnicotinamid- e)	↓ Risk of eczema at age 12 months for nicotinamide and anthranilic acid
"The Finnish type 1 Diabetes Prediction and Prevention Nutrition Study" in Finland [115]	Study design: prospective population-based cohort study, multicenterMethods: maternal pregnancy FFQ; allergen specific IgE at 5 years of agePopulation and size: newborn infants with	Citrus fruits Vitamin D	 ↑ Sensitization to inhalant allergen for citrus fruits intake (OR 1.14) ↓ Sensitization to food allergen for vitamin D intake (OR 0.56)
	HLA-conferred susceptibility to type 1 di- abetes $(n = 931)$		
"The Etude des Déterminants pré et post natals du développement et de la santé de l'Enfant (EDEN) birth cohort" in France [116]	Study design: prospective birth cohort Methods: cord blood 25(OH)D; International Study of Asthma and allergies in childhood-based symptom questionnaires at 1, 2, 3, and 5 years	25(OH)D	 ↓ Risk of transient early wheezing and early- and late-onset AD, as well as AD, by the ages of 1, 2, 3 and 5 years ↔ No association with asthma and
	Population and size: newborn $(n = 239)$		allergic rhinitis at age 5 years
In Perth, Australia [117]	 Study design: prospective birth cohort Methods: cord blood 25-hydroxyvitamin D₃ [25(OH)D₃] concentration; allergic outcomes in the first year of life Population and size: high-risk mother–infant pairs (n = 669) 	25(OH)D ₃	 ↓ Risk of eczema for high cord blood 25(OH)D₃ ↔ No association with allergen sensitization, IgE-mediated food allergy, and eczema severity
In Southampton, UK [118]	Study design: prospective cohort study	25(OH)D	↑ Risk of eczema at age 9 months
	Methods: maternal blood tests at late pregnancy; atopic eczema assessed at 9 months old; asthma at age 9 years Population and size: mother-infant pairs		(OR 3.26) ↑ Risk of asthma at age 9 years (OR 5.40)
"The Avon Longitudinal Study of Parents and Children (ALSPAC) study" in UK [119]	(n = 466)Study design: prospective population-based cohort study, multicenterMethods: maternal blood 25(OH)D; allergic	25(OH)D	↔ No association with wheeze, asthma, eczema, atopy and hay fever at age 7.5 years.
	outcome at age 7.5 years Population and size: mother–infant pairs ($n = 5513$)		

"The KOMCHS prebirth cohort" in Japan [120]	Study design: prospective prebirth cohort Methods: maternal diet history questionnaire; offspring allergic outcome by questionnaires at age 23– 29 months. Population and size: mother–infant pairs (<i>n</i> = 1354)	Vitamin D	↑ Risk of infantile eczema (aOR 1.63)
"The Prediction of Allergies in Taiwanese Children (PATCH) study" in Taiwan [121]	Study design: prospective birth cohort Methods: maternal and cord blood 25(OH)D; allergic outcome at age 4 Population and size: mother–infant pairs (n = 164)	25(OH)D	↓ Risk of eczema (OR 0.12) and asthma (OR 0.22) at age 4 for high maternal 25(OH)D
In Taiwan [122]	 Study design: prospective observational cohort Methods: breast milk retinol and 25(OH)D₃; objective SCORAD in infants Population and size: breast milk for exclusively breastfed AD and healthy infants (n = 90) 	Retinol 25(OH)D ₃	 ↔ No association with AD for retinol ↑ objective SCORAD for lower 25(OH)D₃ ↔ No association with persistent AD till age 3–4 years for 25(OH)D₃

DBPCRCT double-blind, placebo-controlled, randomized clinical trial, SCORAD SCORing Atopic Dermatitis index, GA gestational age, FFQ food frequency questionnaire, FeNO fractional exhaled nitric oxide, Rint interrupter resistance, SNP single-nucleotide polymorphism, MTHFR methylene-tetrahydrofolate reductase, OR odds ratio, HLA human leukocyte antigen

needs of the developing fetus for processing DNA synthesis and rapid cell division. An inadequate periconceptional maternal folate status may lead to defects in fetal development, most notably neural tube defects, such as spinal bifida, meningocele, or an encephaly [184]. Hence, the importance of folic acid status and supplementation during pregnancy has been addressed in prenatal care programs of various countries [185]. In January 1992, the UK government first suggested that pregnant women should consume 400 mg folic acid per day for the first 12 weeks of pregnancy and preferably before conception. Later the same year in September, the US government made similar recommendations with same dosage (400 mg daily) for all women of childbearing age [186]. In Asia, the Ministry of Health and Welfare in Taiwan recommends a 400-mg folic acid intake before conception and an increased amount to 600 mg daily during pregnancy.

As a key participant in DNA methylation, folate/folic acid can therefore regulate transcriptional activity by facilitating in the methylation of specific regulatory regions of genes, thereby silencing their expression [9]. Given that mammals are highly dependent on dietary methyl donors and cofactors for the methyl groups to convey all biological methylation reactions, early administration of methyl donors, such as folic acid, in diet could cause significant epigenetic alterations [187]. This epigenetic function may influence the Th1/Th2 polarization of the immune system. Animal studies showed that methylation of Th1-related genes caused by in utero exposure drives the Th1/Th2 balance toward Th2 reactions, thus making the host more prone to developing atopic diseases [188]. The reduced expression of typical Th1 cytokine IFN- γ by methylation of the promoter region, for example, increases the risk of AD [189, 190]. However, whether early folic acid supplementation specifically facilitates hypermethylation of cytokine expressions in a certain T helper pathway is unclear. Further studies may be required to clarify the causality.

Although the mechanism remains unclear, clinical studies are being conducted to examine whether folic acid supplementation during pregnancy or lactation influences the risk of developing allergic diseases, whereas most focus on airway allergies rather than eczema or AD. The 2012 Generation R Study in the Netherlands conducted by Kiefte-de Jong et al. is by far the largest in scale, with a population-based birth cohort of 8742 children followed up from fetal life to 48 months old. This study revealed that maternal folate > 16.2 nmol/L was positively associated with the development of AD (aOR 1.18; 95% CI 1.05–1.33) [103]. A high maternal vitamin B12 level > 178 pmol/L demonstrated a similar association with AD (aOR 1.30; 95% CI 1.06–1.60) in the same study [103]. Another study by Dunstan et al. in Australia in 2012 showed that relatively high intake of folate supplement by the mother $(> 500 \mu g/day)$ increased the probability of infants to develop eczema at the age of one than those whose mother had low levels of folate supplement (< 200 µg/day; OR 1.85; 95% CI 1.14-3.02; p = 0.013) [104]. Interestingly, the amount of maternal folate intake from ordinary food causes no difference in the risk of offspring eczema. The folate level in cord blood shows no difference in children with or without eczema, but it is possibly associated with sensitization, indicating a greater risk when the level is either low or high (< 50 or > 75 nmol/L, respectively) [104]. Other studies, however, showed different trends. Two separate studies in the Netherlands [105, 106],

including one based on the KOALA cohort [105] and a Japanese study [107] investigating outcomes on both airway allergy and eczema, found no association between maternal intake of folate and eczema. Two studies reported that a high folate level in the middle of gestation possibly demonstrates a protective rather than harmful effect on the risk of offspring atopy. A South Korean study with a prospective cohort of 917 mother-child pairs showed that a high maternal serum folate level (\geq 9.5 ng/mL) during gestational age (GA) of 12-28 weeks was associated with the reduced risk of maternalreported AD at 24 months; however, no relationship was observed for the high folate level in late pregnancy (29-42 weeks) or AD in other ages [109]. Another recent study based in the USA found decreased odds of wheezing in children whose mothers had 2nd trimester folate \geq 20 ng/mL, but no association was noted for AD [111].

Overall, current studies focusing on the relationship between maternal folic acid/folate supplement and childhood AD had shown inconsistent results. All research about maternal folic acid status and childhood atopic diseases are observational cohort studies, partly due to the difficulties and infeasibility of conducting clinical trials and given that deficiency or overdosing of folic acid during the gestation stage could be harmful [191]. A meta-analysis investigated the effect of prenatal folic acid supplement on childhood risk of asthma and discovered no association [192]. However, in the case of AD, thus far, neither meta-analysis nor systemic review is available to generate a conclusive result. The role of maternal folic acid supplement on offspring AD remains unclear. A larger cohort study with detailed documentation of maternal folic acid status and prolonged follow-up period of the children might be needed to gain deeper insights into this topic.

Vitamins B2, B6, and B12

Vitamins B2, B6, and B12 participate in the DNA methylation pathway, which is carried out mainly by folate. Deficiency in vitamin B2, B6, or B12 may impair folate metabolism [193]. For pregnant women, these group B vitamins, including folate (B9), are often supplemented in combination. Little is known regarding the roles of vitamin B2 and B6 in association with atopic diseases. A Japanese study conducted by Miyake et al. analyzed the maternal consumption of vitamin B2, B6, folate, and B12 and its effects on childhood atopic diseases including AD [107]. None of the vitamins analyzed in the study affected the risk of AD.

Vitamin B12 (cobalamin) is a group of cobalt-containing vitamins. Although this vitamin can be synthesized by microorganisms in human gut, most of the vitamin B12 in our body comes from food sources, especially those of animal origins, such as milk, cheese, and eggs [194]. Vitamin B12 facilitates normal physiological function in humans by multiple metabolic functions. One member of B12, methylcobalamin, acts as a co-enzyme that methylates homocysteine into methionine, which is an important step to convert folate (B9) into metabolically active form to perform its function as methyl group donor [194]. Methylcobalamin can also suppress cytokine production by T cells in vitro and modulate lymphocyte function through augmenting Treg activities, although the mechanism is not fully understood. Topical vitamin B12 is effective in treating AD in both adults and children [195, 196].

Vitamin B12 is often supplemented together with folate for pregnant women. Despite the claimed effect of improving AD, current studies, including a Japanese study on vitamin B group and the Generation R Study mentioned above, found no beneficial role for vitamin B12 in childhood AD when supplemented to mothers during pregnancy [103, 107]. The Generation R Study identified an increased risk with maternal supplementation of vitamin B12. One study investigated the relation between cord blood folate, homocysteine, and vitamin B12 levels and childhood asthma and eczema and found no association [113].

A polymorphism in the gene encoding methylenetetrahydrofolate reductase, the MTHFR gene, is also being studied for its role in manipulating folate and vitamin B12 status, which could render susceptibility to atopic diseases. With a prevalence of 5-15% in the general population, the singlenucleotide polymorphism C677T in MTHFR gene reduces the activity of the enzyme, causes decreased re-methylation of homocysteine to methionine by vitamin B12, and subsequently alters folate distribution [197, 198]. The presence of homozygous MTHFR C677T may further augment the effect of low folate or vitamin B12 status on DNA methylation in lymphocytes and affect the risk of developing allergic diseases. Several studies investigated maternal and/or fetus MTHFR C677T polymorphism, along with maternal folate and vitamin B12 status, and their association with offspring AD [199]. However, of all the genetic combinations analyzed, no difference was found in the outcomes of the studies [103,200].

Vitamin B3 (Niacin)

Niacin (vitamin B3) is found in a variety of food sources, including fish, poultry, meat, mushroom, and nuts. Nicotinamide is the amide form of niacin, which could be supplemented by niacin intake and shares similar physiological functions with niacin. Niacin can be converted from tryptophan, an essential amino acid, by the kynurenine pathway. An increased intake of tryptophan-containing foods or supplemental niacin can increase the serum level of nicotinamide; topical and oral forms of nicotinamide are effective in the treatment of dermatitis by reducing transdermal water loss [201, 202].

One study carried out in the UK by El-Heis et al. examined the relation of maternal serum concentrations of nicotinamide and tryptophan metabolites in the kynurenine pathway to the risk of atopic eczema in offspring [114]. The study included 497 mother–infant pairs, and the outcomes were analyzed at 6 and 12 months of age. The results showed that high concentrations of nicotinamide and anthranilic acid were associated with a low risk of eczema at age 12 months but not at 6 months [114]. The other metabolites showed no association with AD. However, given the limited number of studies and their relatively small sample size, the relationship between maternal niacin intake and childhood AD remains to be concluded.

Vitamin C

Vitamin C, or ascorbic acid, is a water-soluble vitamin with well-known antioxidant properties and is abundant in fruits and vegetables [203]. The rationale of the effect of vitamins C on AD originates from two aspects: vitamin C plays a role in maintaining the integrity of skin by aiding in the synthesis of ceramides in the epidermis [204], and as an antioxidant, vitamin C reduces the oxidative stress that plays a part in the pathogenesis of AD [205]. Adult studies with small sample sizes had demonstrated that serum vitamin C level is low in patients with AD and is inversely correlated with severity in terms of SCORing Atopic Dermatitis index (SCORAD) [183, 204].

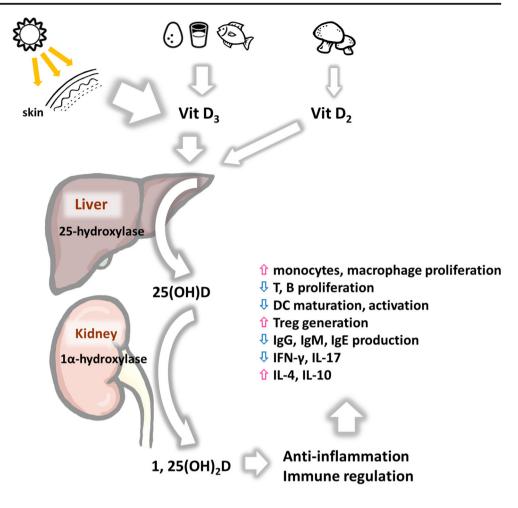
Despite several proven rationales, however, the results of mother-infant studies showed inconsistencies. The Australian study by West et al. investigated the maternal intake of multiple antioxidants during pregnancy; a high dietary vitamin C was correlated with reduced offspring wheezes but not eczema [100]. A breast milk analysis by Hoppu et al. measured the antioxidant composition based on vitamin C and E levels of 34 mothers and revealed that high concentrations of vitamin C in breast milk reduced the risk of atopy in infants (p = 0.038), which is defined by the presence of AD during the first year of life and a positive skin-prick test reaction at 12 months [37]. By contrast, a prospective questionnaire-based UK study of 1924 mother-infant pairs revealed that a high maternal intake of vitamin C was associated with weak but positive risk of AD in infants in their second year of life [101]. Results of other studies based specifically on children population are also inconsistent [205]. Evidence is still inadequate to make suggestions for vitamin C intake of pregnant or breastfeeding mothers.

Vitamin D

Vitamin D is a fat-soluble vitamin that occurs in two main forms: ergocalciferol (vitamin D_2 , produced by plants) and cholecalciferol (vitamin D_3 , derived from animals) (Fig. 2) [206]. Humans predominately derive vitamin D by cutaneous synthesis under the influence of sunlight, with limited vitamin D sourced from dietary intake [207]. Therefore, the UK Department of Health recommends that women consume a daily vitamin D supplement of 400 IU throughout pregnancy and lactation, whereas the AAP suggests that all infants and children should have a minimum intake of 400 IU vitamin D per day beginning immediately after birth. Vitamin D potentially modulates allergy outcomes via its multifaceted effects on altered epidermal barrier function, immune dysregulation, and inadequate bacterial defense [208]. Systemically, vitamin D is an immunomodulator that targets innate and adaptive immune cells, including monocytes, macrophages, DCs, T cells, and B cells. Vitamin D decreases excessive inflammation by suppressing TLR production by monocytes, enhancing the mast cell production of IL-10, inhibiting DC activation by lipopolysaccharides, decreasing cytokine secretion from Th1 cells, inducing Treg activities, and inhibiting B lymphocyte function and IgE secretion [209, 210]. Deficiencies in vitamin D levels and/or signaling would favor a predominant Th2 response and IgE elevation. In the skin, vitamin D exhibits pleiotropic effects ranging from keratinocyte proliferation, differentiation, and apoptosis to barrier maintenance and immunoregulatory processes [206]. Vitamin D plays an important role in epidermal differentiation and barrier function through the regulation of calcium, antimicrobial peptides (i.e., cathelicidin), and TLRs [211-213]. Vitamin D prevents skin T cell infiltration by downregulating the expression of cutaneous lymphocyte-associated antigen [214]. Therefore, vitamin D deficiency might lead to the dysfunction of the skin barrier, infiltration of T cells, and increased predisposition of patients with AD to skin superinfection by Staphylococcus aureus or its superantigens [213].

Although several studies reported no significant relationship [215], most of the research found a negative correlation between AD severity and vitamin D levels with dosedependent effects [216–218]. Low serum vitamin D levels were associated with elevated serum IgE levels [218] and increased house dust mite sensitization in AD patients [219]. A specific vitamin D receptor gene polymorphism occurs frequently in patients with severe AD, thus suggesting the important role of vitamin D in the pathogenesis of the disease [217]. Maternal vitamin D intake in pregnancy is associated with the reduced risk of detection of IgE, which is specific to food allergens, in offspring at 5 years of age [115].

Controversies surround the results regarding the association of vitamin D status in utero and AD development. The EDEN prospective birth cohort study in France found a significant inverse association between the cord serum 25hydroxyvitamin D [25(OH)D] levels and risk of AD by the ages of 1, 2, 3, and 5 years [116]. The prospective study with 669 mother–infant pairs in Australia showed a low cord serum vitamin D in infants that developed eczema (p = 0.018); eczema was significantly more likely to occur in those with Fig. 2 Vitamin D metabolism. Vitamin D is mainly derived from cutaneous synthesis under the sunlight, with limited vitamin D sourced from dietary intake. The main dietary sources are egg, fish, milk (vitamin D₃ from animals), and mushroom (vitamin D₂ from plants). After metabolized by the liver and kidney, 1,25 (OH)2D regulates the immune system by various effects, such as inhibition of T and B cell proliferation, decrease in IgE production, inhibition of DC activation and maturation, and increase in Treg generation



vitamin D concentrations < 50 nmol/L in comparison with those with concentrations \geq 75 nmol/L (OR 2.66; 95% CI 1.24-5.72; p = 0.012 [117]. A UK study showed that children born to mothers in the highest quartile of the cohort for maternal serum 25(OH)D concentration in late pregnancy had an increased risk of eczema at 9 months of age and asthma at 9 years of age [118]. Altogether, observational studies support a protective relationship between vitamin D status in utero and the risk of eczema development, whereas others suggest that high levels may be a risk factor. In an Avon Longitudinal Study of Parents and Children (ALSPAC) study, maternal serum 25(OH)D concentration was measured at each stage of pregnancy and showed no association with parentally reported allergic diseases (wheeze, asthma, eczema, or hay fever) or sensitization to three common aeroallergens [119]. These studies measured maternal 25(OH)D levels during pregnancy [116, 119, 220] and/or in cord blood [117, 118, 220] at birth and reported the eczema outcomes in infants. A major limitation of these studies is that 25(OH)D levels were only measured once, thus failing to capture the effects of likely changes in 25(OH)D status and exposure to the fetus over the course of pregnancy [208]. However, limited information is available regarding the primary prevention of allergic diseases after vitamin D supplementation. The two combined independent RCTs of vitamin D supplementation during pregnancy resulted in a significantly reduced risk of asthma/recurrent wheezing, but not eczema, in the offspring [96, 97, 221]. In a UK randomized study, 180 women received no supplement, 800 IU per day ergocalciferol (vitamin D2), or a single bolus dose of 200,000 IU cholecalciferol (vitamin D3) at 27 weeks of gestation [98]. No significant differences were observed between the control group versus the intervention groups for atopic sensitization or risk of eczema. In exclusively breastfed infant, our study showed that vitamin D levels in breast milk were negatively associated with objective SCORAD at age of 2–4 months (p = 0.003) [122].

In summary, conflicting evidence is available about the association between maternal vitamin D status and risk for AD development in offspring. However, from the serum status and AD severity in children and adults, vitamin D plays a protective role in AD. Although serum 25(OH)D concentration reflects dietary intake and UVB exposure, a number of other factors influence circulating 25(OH)D. Genetic variation in a number of genes, including the vitamin D binding protein, the carrier molecule that delivers all vitamin D and vitamin D metabolites to tissues, influences serum 25(OH)D [222]. More

double-blind placebo-controlled vitamin D intervention studies are required. These studies need to consider the supplement type (vitamin D3 vs. D2), timing (a highly developed immune system is already present by 20 weeks of pregnancy), and dosage [9].

Vitamin E

Vitamin E is a family of fat-soluble compounds that deliver antiinflammatory properties and act as powerful antioxidants [205]. Two principal classes of vitamin E, tocopherols and tocotrienols, exist, and they can be further divided into four forms: α , β , δ , and γ [223]. These eight forms of vitamin E are naturally present in food sources, most abundantly in plant oils, cereals, and nuts. The strong antioxidative property of vitamin E has inspired studies inspecting its nutrient–disease association with AD. In adult studies including DBPCRCT, vitamin E supplementation has significantly improved AD in terms of symptoms, SCORAD, and serum IgE level [224, 225]. In children, a questionnaire-based study has shown that AD patients have significantly lower dietary vitamin E intake than those without the disease.

In mother-infant studies, however, not all results are in agreement. A UK prospective study investigating 1924 mother-infant pairs in multiple antioxidants showed that maternal intake of vitamine E was inversely associated with the risk of childhood eczema in atopic mothers (p = 0.024), but no statistical significance was observed for the whole population [101]. In the Australian study by West et al., no association was found between maternal intake of vitamin E and eczema in offspring [100]. In a Finnish study in which 37 atopic mothers were enrolled, the ratios of infant serum γ - and α tocopherol to fat were significantly correlated with maternal serum ratios (infant α -tocopherol/fat was higher, and γ -tocopherol/fat ratio was lower than those of mothers), indicating that maternal vitamin E status may affect that of the infants. Nevertheless, the serum tocopherol levels showed no correlation with the clinical presentation of AD or skin-prick test [226]. In general, vitamin E may have a protective role against AD, but its association with offspring AD when supplemented in maternal diet during pregnancy remains unclear.

Minerals and Trace Elements

Magnesium

Magnesium assists in the activation of vitamin D, which in turn regulates serum calcium and phosphate levels and facilitates immune function; this element has thus been widely studied for its relationship with AD [227]. Magnesium itself also exhibits antiinflammatory activity on the skin and improves skin barrier function by participating in cell proliferation and differentiation [228, 229]. In a small-scale children's study, AD patients had significantly lower serum magnesium level compared with the controls (p = 0.007). Erythrocyte zinc levels were significantly lower in AD patients in the same study [230]. However, the mechanism is still unclear. Studies on magnesium intake in the mother–fetal or mother–childhood atopy relationship are lacking.

Iron

Iron supplementation is essential for pregnant women, and the World Health Organization has made recommendations of 400 µg daily iron supplement along with folic acid to prevent maternal anemia, puerperal sepsis, low birth weight, and preterm birth [231]. Iron is often supplemented to infants or toddlers because of possible iron deficiency or anemia associated with exclusive breast milk or formula feeding, as suggested by AAP [232]. The relationship between iron status and the immune system is complicated. Free ferrous ion in the blood can interact with H₂O₂ through a mechanism called Fenton reaction, leading to the production of free radicals and increased oxidative stress within cells [233]. On the other hand, a deficient iron status may also be harmful. Poor iron status at birth may compromise Th1 lymphocytes and bias the immune response toward the Th2 pathway, increasing the risk of development of allergic diseases [234].

An exploratory study by Nwaru et al. investigated the associations between maternal iron status in pregnancy and childhood wheezing and atopy in their first 10 years of life. A reduced maternal serum iron level is significantly associated with childhood wheezes. The risk of "doctor-diagnosed eczema" is high with low maternal iron status, but the association is of borderline significance [235]. In part of the ALSPAC study by Shaheen et al., high iron levels in cord blood are negatively associated with later onset of eczema (OR 0.90), but the cord blood level shows no relation with the maternal intake [236]. Fortes et al. noted that prenatal cosupplementation of iron and folic acid leads to a fourfold decreased risk of AD (OR 0.22; 95% CI 0.06-0.79; p = 0.02) after adjusting for possible confounding factors [237]. Although studies with large sample size are lacking, current available evidence indicates a possible protective role of adequate iron supplement in perinatal or infancy against AD.

Zinc

Zinc is an essential micro-nutrient [205] that is related to the integrity and immune status of the skin barrier in multiple functional pathways [238]. Zinc acts as the structural cofactor of zinc-finger motifs, which are present in the proteins involved in the expression of filaggrin, a key component involved in the pathogenesis of AD [239]. Topical zinc oxide application can alter the dermal cytokine profile into an

antiinflammatory pattern (increased IL-10 and decreased IL-1 β , IL-6, and TNF- α levels) [239]. An in vitro study has shown a possible antimicrobial activity of zinc oxide against methicillin-resistant *Staphylococcus aureus* [240]. Although the mechanism between zinc and AD is still not fully known, several pilot studies have suggested the protective role of high serum, hair, and erythrocyte zinc level against the development of AD [238].

Dietary zinc is most abundant in meat, shellfish, nuts, and certain vegetables. A Mediterranean diet pattern may generally satisfy the zinc requirement [241, 242]; maternal Mediterranean diet pattern may be beneficial for childhood atopy [12]. Nevertheless, a limited number of studies focused specifically on zinc in maternal diet and its effect in offspring eczema or AD. Currently existing large-scale studies analyzing antioxidants in maternal diet and offspring atopy discovered no associations between zinc intake and childhood eczema [10, 16, 100]. Whether zinc in maternal diet would influence childhood atopic disease remains unclear.

Copper

The rationale of how copper influences the development of allergic disease is not fully discovered. Copper is suggested to have antioxidant activities and act as a cofactor that participates in cell growth and function [243]. A 1987 study by Di Toro et al. investigated both zinc and copper status in allergic children. However, in contrast to zinc, whose level in hair is negatively correlated with development of AD, a significantly higher hair copper level was found in children with the disease [244]. Similarly, in 1990, el-Kholy et al. observed a significantly high serum ceruloplasmin in children with AD (p < 0.001), whereas high serum and hair copper levels were associated with asthma [243].

In spite of these early studies suggesting that high copper levels may induce atopy, recent maternal–infant studies valued copper for its antioxidant role, whereas others reported different results. The Australian study by West et al. showed that a high dietary copper intake is associated with reduced risk of offspring eczema and other allergic diseases [100]. Another cohort study by Martindale et al. found no relationship [101]. Interestingly, maternal intake of copper showed an opposite influence compared with the copper status of the children themselves. Given that the role of copper in allergic diseases is not fully elucidated, additional studies have to be carried out to clarify the related mechanism.

Selenium

Selenium is known for its role as an antioxidant in human health. Environmentally, selenium is ubiquitous and can be found in rock, water, and soil. This element enters the food chain by being taken up by plants from the soil and finally reaches animals through bioaccumulation. The most abundant food sources of selenium include fish, egg, meat, and Brazilian nuts. Biologically, selenium is a key component in several enzymes, such as glutathione peroxidase, thioredoxin reductase, and iodothyronine deiodinases, which carry out antioxidative functions [245]. Sufficient amount of selenium is therefore necessary for maintaining optimal antioxidative capacity in the body [205]. In studies, selenium status is being linked to diseases, including AD, because of the possible role of oxidative stress in their pathogenesis [181, 182, 246]. Limited clinical evidence supports the beneficial role of selenium in improving AD. A DBPCRCT conducted in 1989 by Fairris et al. enrolled 60 adults with AD, who were supplemented with either selenium-enriched yeast plus vitamin E, selenium-enriched yeast only, or placebo. No difference was observed in the severity of eczema or the concentration of cutaneous selenium among groups [247].

For pregnant women, serum selenium concentration decreases significantly during gestation [248]; a deficient selenium status is associated with multiple health problems, such as recurrent miscarriages, preterm delivery, gestational diabetes mellitus, thyroid peroxidase antibody-positive autoimmune thyroiditis, and neural tube defect of the fetus [249]. Adequate selenium intake or supplement is thus suggested by authorities, especially during pregnancy [250, 251]. Regarding the antioxidative property, several studies investigated the association of maternal selenium status or selenium supplementation and the risk of atopic diseases, including eczema, in children. In the ALSPAC study that analyzed trace elements in the umbilical cord, high cord blood selenium level was negatively associated with childhood wheezing, but no association was found with eczema [236]. Another study on UK cohort by Martindale et al. revealed no consistent statistically significant associations between eczema in the first 2 years of life and total maternal intake of selenium [101]. A Japanese cohort study of 1036 mother-infant pairs by Yamada et al. analyzed 32 measurable minerals in the hair of infants and mothers using proton-induced X-ray emission (PIXE); only selenium and strontium demonstrated significant association with childhood dermatitis [252]. Selenium deficiency in either infant or mother increased the risk of AD at the age of 10 months, but for mothers, the result was borderline significant (p = 0.048 for infants and p = 0.062 for mothers). Compared with childhood asthma or wheezing, of which more evidence is needed for a relatively concrete conclusion, little is known for selenium's role in AD and how mother's selenium status affects children.

Strontium

Strontium is present in seawater and soil. The strontium we obtain mostly originates from seafoods, cereals, and grains. Strontium is scarcely studied for its biological role compared

with other trace elements. Most of the physiological functions currently known for strontium are related to the metabolism of bones [253]. Less is known for its association with atopic diseases. A study by Barneo-Caragol et al. revealed that strontium level rose significantly during the third trimester of pregnancy, possibly to cope up with the oxidative damage that developed physiologically during this period [254]. Yamada et al. discovered that among all 32 minerals tested in infants' and mothers' hair by PIXE in their study, only selenium and strontium had significant association with the risk of childhood AD. In contrast to selenium, whose levels in both infants' and mothers' hair are inversely related to AD risk, the hair strontium level in mother is positively associated with AD risk in children [252]. With the tool, the authors of this study developed a logistic prediction model for childhood AD using the levels of these elements, with a sensitivity of 65.9% and specificity of 70.5% [252].

Discussion

In consideration of children, maternal nutrition status may have substantial effects as one of the most modifiable environmental factors and may play a prominent role in the development of chronic and multifactorial diseases, such as allergy. However, studying the causality of mothers' food intake to offspring disease could be difficult. The nutrients from mothers are delivered through the placenta to the fetus or secreted in breast milk. Thus, the levels of most nutrients in cord blood or breast milk and in mothers' and children's sera must be examined to establish a robust correlation. Several nutrients may be quantified indirectly using other specimens, for example, minerals which can be measured in urine or hair [252]. However, certain obstacles could prevent the acquisition of samples in real clinical settings, especially with vulnerable groups as subjects. Not all nutrients have highly correlated, measurable markers. Most of the available motherfetal or mother-infant studies utilized well-designed, validated questionnaires to assess the nutrient composition of maternal diet; however, although they are easy to administer, such questionnaires could be affected by recall bias [255]. The often observed inconsistencies in the amounts of specific nutrients from mother's intake, maternal serum concentration to cord blood, and fetal serum concentration also suggest a complicated physiological mechanism involved in the process of nutrition transportation [104, 236].

Compared with childhood asthma and wheezes, studies rarely evaluated the influence of maternal nutrition status to examine the outcome of childhood AD or eczema. The mechanism behind the causality is also less explored in AD, either in animal models or in nutrient–genome interaction. The paucity of studies causes difficulty in developing a solid conclusion. Of all the nutrients that were reviewed, ω -3 PUFA, folic acid, and vitamin D are the most studied. Small-scale RCTs are conducted for ω -3 PUFA and vitamin D, of which commercialized supplementary products are available because of a relative common deficient status in modern Western lifestyle. Other nutrients are mostly investigated in observational studies, of which several use a large sample size and long period follow-up periods, for example, the Generation R Study or the ALSPAC [103, 236]. However, neither solid proof nor concordant results of being beneficial or risky to offspring with AD has been found for any of the nutrients. Systemic reviews or meta-analyses are also lacking due to the highly heterogenous study designs and methodology and inadequate number of studies.

Several studies investigated the influence of maternal diet in terms of specific food category or dietary pattern, instead of focusing on single nutrients. A few research suggested a potentially protective role of maternal Mediterranean diet pattern, which consists of high proportions of vegetables, fruits, nuts, seafoods, and grains, against AD or other atopic diseases [12, 13, 241, 242]. Additional intake of fish [8, 17], vegetables, and fruits [8, 16] is also beneficial [256]. Nevertheless, whether the effect is delivered mainly by a sole key nutrient in certain foods (e.g., w-3 PUFA in fish or vitamin C in citrus fruits) is unclear. One RCT specifically examined the level of single nutrient (ω -3 PUFA) after increased intake of salmon and detected an increase in EPA in maternal serum and cord blood [73]. A change in the cytokine and Ig profile was also observed in the fetus, but no difference was noted in the outcomes [72]. This condition demonstrates that certain food source modifications in maternal diet can alter the nutrition status of offspring using a measurable nutrient component, although no correlation was observed between the laboratory test results and clinical presentations. Given the complicated mechanisms involved in nutrition physiology, concluding the net effect of individual nutrients would be extremely difficult, especially when certain food or diet is composed of various portions of different nutrients. The interplay of other environmental factors, genetic factors, and microbiota also increases the complexity. More strategically designed, carefully conducted, and less biased studies are required to shed a light on this topic.

To conclude, no strong evidence indicates that a single nutrient or food in maternal diet significantly affects the risk of childhood AD. Nevertheless, a balanced diet is not only always helpful for humans in pregnancy or during lactation but also a key to healthy immune function against allergies and other diseases. A healthy lifestyle with healthy, balanced dietary intake is thus encouraged.

Compliance with Ethical Standards

Conflict of Interest The authors declare that they have no conflict of interest.

Ethical Approval and Informed Consent Not applicable.

References

- Misery L (2011) Atopic dermatitis: new trends and perspectives. Clin Rev Allergy Immunol 41(3):296–297. https://doi.org/10. 1007/s12016-010-8247-6
- Ong PY, Leung DY (2006) Immune dysregulation in atopic dermatitis. Curr Allergy Asthma Rep 6(5):384–389. https://doi.org/ 10.1007/s11882-996-0008-5
- Ong PY, Leung DY (2016) Bacterial and viral infections in atopic dermatitis: a comprehensive review. Clin Rev Allergy Immunol 51(3):329–337. https://doi.org/10.1007/s12016-016-8548-5
- Nutten S (2015) Atopic dermatitis: global epidemiology and risk factors. Ann Nutr Metab 66(Suppl 1):8–16. https://doi.org/10. 1159/000370220
- Roduit C, Frei R, Loss G, Buchele G, Weber J, Depner M, Loeliger S, Dalphin ML, Roponen M, Hyvarinen A, Riedler J, Dalphin JC, Pekkanen J, von Mutius E, Braun-Fahrlander C, Lauener R, Protection Against Allergy-Study in Rural Environments study g (2012) Development of atopic dermatitis according to age of onset and association with early-life exposures. J Allergy Clin Immunol 130(1):130–136 e135. https://doi. org/10.1016/j.jaci.2012.02.043
- Han H, Roan F, Ziegler SF (2017) The atopic march: current insights into skin barrier dysfunction and epithelial cell-derived cytokines. Immunol Rev 278(1):116–130. https://doi.org/10. 1111/imr.12546
- Simpson EL, Berry TM, Brown PA, Hanifin JM (2010) A pilot study of emollient therapy for the primary prevention of atopic dermatitis. J Am Acad Dermatol 63(4):587–593. https://doi.org/ 10.1016/j.jaad.2009.11.011
- Sausenthaler S, Koletzko S, Schaaf B, Lehmann I, Borte M, Herbarth O, von Berg A, Wichmann HE, Heinrich J, Group LS (2007) Maternal diet during pregnancy in relation to eczema and allergic sensitization in the offspring at 2 y of age. Am J Clin Nutr 85(2):530–537. https://doi.org/10.1093/ajcn/85.2.530
- Miles EA, Calder PC (2015) Maternal diet and its influence on the development of allergic disease. Clin Exp Allergy 45(1):63–74. https://doi.org/10.1111/cea.12453
- Bedard A, Northstone K, Holloway JW, Henderson AJ, Shaheen SO (2018) Maternal dietary antioxidant intake in pregnancy and childhood respiratory and atopic outcomes: birth cohort study. Eur Respir J 52(2). https://doi.org/10.1183/13993003.00507-2018
- Wu G, Bazer FW, Cudd TA, Meininger CJ, Spencer TE (2004) Maternal nutrition and fetal development. J Nutr 134(9):2169– 2172. https://doi.org/10.1093/jn/134.9.2169
- Chatzi L, Torrent M, Romieu I, Garcia-Esteban R, Ferrer C, Vioque J, Kogevinas M, Sunyer J (2008) Mediterranean diet in pregnancy is protective for wheeze and atopy in childhood. Thorax 63(6):507–513. https://doi.org/10.1136/thx.2007.081745
- Netting MJ, Middleton PF, Makrides M (2014) Does maternal diet during pregnancy and lactation affect outcomes in offspring? A systematic review of food-based approaches. Nutrition (Burbank, Los Angeles County, Calif) 30(11–12):1225–1241. https://doi. org/10.1016/j.nut.2014.02.015
- Romieu I, Torrent M, Garcia-Esteban R, Ferrer C, Ribas-Fito N, Anto JM, Sunyer J (2007) Maternal fish intake during pregnancy and atopy and asthma in infancy. Clin Exp Allergy 37(4):518– 525. https://doi.org/10.1111/j.1365-2222.2007.02685.x
- Miyake Y, Sasaki S, Tanaka K, Hirota Y (2010) Dairy food, calcium and vitamin D intake in pregnancy, and wheeze and eczema in infants. Eur Respir J 35(6):1228–1234. https://doi.org/10.1183/ 09031936.00100609

- Miyake Y, Sasaki S, Tanaka K, Hirota Y (2010) Consumption of vegetables, fruit, and antioxidants during pregnancy and wheeze and eczema in infants. Allergy 65(6):758–765. https://doi.org/10. 1111/j.1398-9995.2009.02267.x
- Willers SM, Devereux G, Craig LCA, McNeill G, Wijga AH, Abou El-Magd W, Turner SW, Helms PJ, Seaton A (2007) Maternal food consumption during pregnancy and asthma, respiratory and atopic symptoms in 5-year-old children. Thorax 62(9): 773–779. https://doi.org/10.1136/thx.2006.074187
- Miyake Y, Okubo H, Sasaki S, Tanaka K, Hirota Y (2011) Maternal dietary patterns during pregnancy and risk of wheeze and eczema in Japanese infants aged 16-24 months: the Osaka Maternal and Child Health Study. Pediatr Allergy Immunol 22(7):734–741. https://doi.org/10.1111/j.1399-3038.2011.01176.
- Miyake Y, Yura A, Iki M (2003) Breastfeeding and the prevalence of symptoms of allergic disorders in Japanese adolescents. Clin Exp Allergy 33(3):312–316. https://doi.org/10.1046/j.1365-2222. 2003.t01-1-01607.x
- Hollis BW, Wagner CL, Howard CR, Ebeling M, Shary JR, Smith PG, Taylor SN, Morella K, Lawrence RA, Hulsey TC (2015) Maternal versus infant vitamin D supplementation during lactation: a randomized controlled trial. Pediatrics 136(4):625–634. https://doi.org/10.1542/peds.2015-1669
- Kim JH (2017) Role of breast-feeding in the development of atopic dermatitis in early childhood. Allergy, Asthma Immunol Res 9(4):285–287. https://doi.org/10.4168/aair.2017.9.4.285
- Lien TY, Goldman RD (2011) Breastfeeding and maternal diet in atopic dermatitis. Can Fam Physician 57(12):1403–1405
- Guardamagna O, Cagliero P (2016) Lipid metabolism in the human fetus development. In: Bhattacharya N, Stubblefield PG (eds) Human fetal growth and development: first and second trimesters. Springer International Publishing, Cham, pp 183–195. https://doi. org/10.1007/978-3-319-14874-8 12
- Yaqoob P, Calder PC (2007) Fatty acids and immune function: new insights into mechanisms. Br J Nutr 98(Suppl 1):S41–S45. https://doi.org/10.1017/S0007114507832995
- Fritsche K (2006) Fatty acids as modulators of the immune response. Annu Rev Nutr 26:45–73. https://doi.org/10.1146/ annurev.nutr.25.050304.092610
- German JB, Dillard CJ (2004) Saturated fats: what dietary intake? Am J Clin Nutr 80(3):550–559. https://doi.org/10.1093/ajcn/80.3. 550
- Carta G, Murru E, Banni S, Manca C (2017) Palmitic acid: physiological role, metabolism and nutritional implications. Front Physiol 8:902. https://doi.org/10.3389/fphys.2017.00902
- Yu Y, Cai Z, Zheng J, Chen J, Zhang X, Huang XF, Li D (2012) Serum levels of polyunsaturated fatty acids are low in Chinese men with metabolic syndrome, whereas serum levels of saturated fatty acids, zinc, and magnesium are high. Nutr Res (New York, NY) 32(2):71–77. https://doi.org/10.1016/j.nutres.2011.12.004
- Fatima S, Hu X, Gong RH, Huang C, Chen M, Wong HLX, Bian Z, Kwan HY (2019) Palmitic acid is an intracellular signaling molecule involved in disease development. Cell Mol Life Sci. https://doi.org/10.1007/s00018-019-03092-7
- 30. Liang Z, Yuan Z, Guo J, Wu J, Yi J, Deng J, Shan Y (2019) Ganoderma lucidum polysaccharides prevent palmitic acidevoked apoptosis and autophagy in intestinal porcine epithelial cell line via restoration of mitochondrial function and regulation of MAPK and AMPK/Akt/mTOR signaling pathway. Int J Mol Sci 20(3). https://doi.org/10.3390/ijms20030478
- 31. Lambertucci RH, Leandro CG, Vinolo MA, Nachbar RT, Dos Reis Silveira L, Hirabara SM, Curi R, Pithon-Curi TC (2012) The effects of palmitic acid on nitric oxide production by rat skeletal muscle: mechanism via superoxide and iNOS activation. Cell

Physiol Biochem 30(5):1169–1180. https://doi.org/10.1159/000343307

- 32. Kim F, Pham M, Luttrell I, Bannerman DD, Tupper J, Thaler J, Hawn TR, Raines EW, Schwartz MW (2007) Toll-like receptor-4 mediates vascular inflammation and insulin resistance in dietinduced obesity. Circ Res 100(11):1589–1596. https://doi.org/10. 1161/circresaha.106.142851
- Moilanen T, Rasanen L, Viikari J, Akerblom HK, Nikkari T (1992) Correlation of serum fatty acid composition with dietary intake data in children and young adults. Ann Med 24(1):67–70. https://doi.org/10.3109/07853899209164147
- Dunder T, Kuikka L, Turtinen J, Rasanen L, Uhari M (2001) Diet, serum fatty acids, and atopic diseases in childhood. Allergy 56(5): 425–428. https://doi.org/10.1034/j.1398-9995.2001.056005425.x
- 35. Venter C, Meyer RW, Nwaru BI, Roduit C, Untersmayr E, Adel-Patient K, Agache I, Agostoni C, Akdis CA, Bischoff SC, du Toit G, Feeney M, Frei R, Garn H, Greenhawt M, Hoffmann-Sommergruber K, Lunjani N, Maslin K, Mills C, Muraro A, Pali-Scholl I, Poulson LK, Reese I, Renz H, Roberts GC, Smith P, Smolinska S, Sokolowska M, Stanton C, Vlieg-Boerstra B, O'Mahony L (2019) EAACI position paper: influence of dietary fatty acids on asthma, food allergy, and atopic dermatitis. Allergy 74(8):1429–1444. https://doi.org/10.1111/all.13764
- Woods RK, Raven JM, Walters EH, Abramson MJ, Thien FC (2004) Fatty acid levels and risk of asthma in young adults. Thorax 59(2):105–110. https://doi.org/10.1136/thorax.2003. 009498
- Hoppu U, Kalliomaki M, Isolauri E (2000) Maternal diet rich in saturated fat during breastfeeding is associated with atopic sensitization of the infant. Eur J Clin Nutr 54(9):702–705. https://doi. org/10.1038/sj.ejcn.1601079
- Saito K, Yokoyama T, Miyake Y, Sasaki S, Tanaka K, Ohya Y, Hirota Y (2010) Maternal meat and fat consumption during pregnancy and suspected atopic eczema in Japanese infants aged 3-4 months: the Osaka Maternal and Child Health Study. Pediatr Allergy Immunol 21(1 Pt 1):38–46. https://doi.org/10.1111/j. 1399-3038.2009.00897.x
- Barman M, Johansson S, Hesselmar B, Wold AE, Sandberg AS, Sandin A (2013) High levels of both n-3 and n-6 long-chain polyunsaturated fatty acids in cord serum phospholipids predict allergy development. PLoS One 8(7):e67920. https://doi.org/10.1371/ journal.pone.0067920
- Kamp F, Guo W, Souto R, Pilch PF, Corkey BE, Hamilton JA (2003) Rapid flip-flop of oleic acid across the plasma membrane of adipocytes. J Biol Chem 278(10):7988–7995. https://doi.org/ 10.1074/jbc.M206648200
- Ministry of Health (2006) (revised 2008) Food and nutrition guidelines for healthy pregnant and breastfeeding women: a background paper. Ministry of Health, Wellington. https://www.health. govt.nz/system/files/documents/publications/food-and-nutritionguidelines-preg-and-bfeed.pdf. Accessed 13 Dec 2019
- 42. Nicklas TA, Hampl JS, Taylor CA, Thompson VJ, Heird WC (2004) Monounsaturated fatty acid intake by children and adults: temporal trends and demographic differences. Nutr Rev 62(4): 132–141. https://doi.org/10.1111/j.1753-4887.2004.tb00035.x
- Heinrich J, Hölscher B, Bolte G, Winkler G (2001) Allergic sensitization and diet: ecological analysis in selected European cities. Eur Respir J 17(3):395–402. https://doi.org/10.1183/09031936. 01.17303950
- Nagel G, Nieters A, Becker N, Linseisen J (2003) The influence of the dietary intake of fatty acids and antioxidants on hay fever in adults. Allergy 58(12):1277–1284. https://doi.org/10.1046/j.1398-9995.2003.00296.x
- 45. Khanapure SP, Garvey DS, Janero DR, Letts LG (2007) Eicosanoids in inflammation: biosynthesis, pharmacology, and

therapeutic frontiers. Curr Top Med Chem 7(3):311–340. https:// doi.org/10.2174/156802607779941314

- Miles EA, Calder PC (2017) Can early omega-3 fatty acid exposure reduce risk of childhood allergic disease? Nutrients 9(7). https://doi.org/10.3390/nu9070784
- Simopoulos AP (2008) The importance of the omega-6/omega-3 fatty acid ratio in cardiovascular disease and other chronic diseases. Exp Biol Med (Maywood) 233(6):674–688. https://doi. org/10.3181/0711-MR-311
- Shahidi F, Ambigaipalan P (2018) Omega-3 polyunsaturated fatty acids and their health benefits. Annu Rev Food Sci Technol 9: 345–381. https://doi.org/10.1146/annurev-food-111317-095850
- Calder PC (2013) n-3 fatty acids, inflammation and immunity: new mechanisms to explain old actions. Proc Nutr Soc 72(3): 326–336. https://doi.org/10.1017/S0029665113001031
- Dooper MMBW, Wassink L, M'Rabet L, Graus YMF (2002) The modulatory effects of prostaglandin-E on cytokine production by human peripheral blood mononuclear cells are independent of the prostaglandin subtype. Immunology 107(1):152–159. https://doi. org/10.1046/j.1365-2567.2002.01474.x
- Prescott SL, Calder PC (2004) N-3 polyunsaturated fatty acids and allergic disease. Curr Opin Clin Nutr Metab Care 7(2):123–129. https://doi.org/10.1097/00075197-200403000-00004
- 52. Stephensen CB (2004) Fish oil and inflammatory disease: is asthma the next target for n-3 fatty acid supplements? Nutr Rev 62(12):486–489. https://doi.org/10.1111/j.1753-4887.2004. tb00021.x
- Wong KW (2005) Clinical efficacy of n-3 fatty acid supplementation in patients with asthma. J Am Diet Assoc 105(1):98–105. https://doi.org/10.1016/j.jada.2004.10.009
- Plat J, Mensink RP (2005) Food components and immune function. Curr Opin Lipidol 16(1):31–37. https://doi.org/10.1097/ 00041433-200502000-00007
- Simopoulos AP (2006) Evolutionary aspects of diet, the omega-6/ omega-3 ratio and genetic variation: nutritional implications for chronic diseases. Biomed Pharmacother 60(9):502–507. https:// doi.org/10.1016/j.biopha.2006.07.080
- Simopoulos AP (2002) The importance of the ratio of omega-6/ omega-3 essential fatty acids. Biomed Pharmacother 56(8):365– 379. https://doi.org/10.1016/s0753-3322(02)00253-6
- Oddy WH, de Klerk NH, Kendall GE, Mihrshahi S, Peat JK (2004) Ratio of omega-6 to omega-3 fatty acids and childhood asthma. J Asthma 41(3):319–326. https://doi.org/10.1081/jas-120026089
- Simopoulos AP (2002) Omega-3 fatty acids in inflammation and autoimmune diseases. J Am Coll Nutr 21(6):495–505. https://doi. org/10.1080/07315724.2002.10719248
- Calder PC (2006) n-3 polyunsaturated fatty acids, inflammation, and inflammatory diseases. Am J Clin Nutr 83(6 Suppl):1505S– 1519S. https://doi.org/10.1093/ajcn/83.6.1505S
- Yaqoob P, Pala HS, Cortina-Borja M, Newsholme EA, Calder PC (2000) Encapsulated fish oil enriched in alpha-tocopherol alters plasma phospholipid and mononuclear cell fatty acid compositions but not mononuclear cell functions. Eur J Clin Investig 30(3):260–274. https://doi.org/10.1046/j.1365-2362.2000.00623. x
- 61. Wada M, DeLong CJ, Hong YH, Rieke CJ, Song I, Sidhu RS, Yuan C, Warnock M, Schmaier AH, Yokoyama C, Smyth EM, Wilson SJ, FitzGerald GA, Garavito RM, Sui de X, Regan JW, Smith WL (2007) Enzymes and receptors of prostaglandin pathways with arachidonic acid-derived versus eicosapentaenoic acidderived substrates and products. J Biol Chem 282(31):22254– 22266. https://doi.org/10.1074/jbc.M703169200
- James MJ, Gibson RA, Cleland LG (2000) Dietary polyunsaturated fatty acids and inflammatory mediator production. Am J Clin

Nutr 71(1 Suppl):343S-348S. https://doi.org/10.1093/ajcn/71.1. 343s

- Scientific Opinion on Dietary Reference Values for fats, including saturated fatty acids, polyunsaturated fatty acids, monounsaturated fatty acids, trans fatty acids, and cholesterol (2010). EFSA Journal 8 (3). doi:https://doi.org/10.2903/j.efsa.2010.1461
- 64. The Scientific Advisory Committee on Nutrition & Committee on Toxicity (2004) Advice on fish consumption: benefits and risks. The Stationary Office, London. https://assets.publishing.service. gov.uk/government/uploads/system/uploads/attachment_data/file/ 338801/SACN_Advice_on_Fish_Consumption.pdf. Accessed 13 Dec 2019
- 65. Brown TJ, Brainard J, Song F, Wang X, Abdelhamid A, Hooper L, Group P (2019) Omega-3, omega-6, and total dietary polyunsaturated fat for prevention and treatment of type 2 diabetes mellitus: systematic review and meta-analysis of randomised controlled trials. BMJ 366:14697. https://doi.org/10.1136/bmj.14697
- Dunstan JA, Mori TA, Barden A, Beilin LJ, Taylor AL, Holt PG, Prescott SL (2003) Maternal fish oil supplementation in pregnancy reduces interleukin-13 levels in cord blood of infants at high risk of atopy. Clin Exp Allergy 33(4):442–448. https://doi.org/10. 1046/j.1365-2222.2003.01590.x
- Prescott SL, Barden AE, Mori TA, Dunstan JA (2007) Maternal fish oil supplementation in pregnancy modifies neonatal leukotriene production by cord-blood-derived neutrophils. Clin Sci (Lond) 113(10):409–416. https://doi.org/10.1042/CS20070111
- Prescott SL, Irvine J, Dunstan JA, Hii C, Ferrante A (2007) Protein kinase Czeta: a novel protective neonatal T-cell marker that can be upregulated by allergy prevention strategies. J Allergy Clin Immunol 120(1):200–206. https://doi.org/10.1016/ j.jaci.2007.03.045
- Dunstan JA, Mori TA, Barden A, Beilin LJ, Taylor AL, Holt PG, Prescott SL (2003) Fish oil supplementation in pregnancy modifies neonatal allergen-specific immune responses and clinical outcomes in infants at high risk of atopy: a randomized, controlled trial. J Allergy Clin Immunol 112(6):1178–1184. https://doi.org/ 10.1016/j.jaci.2003.09.009
- Meldrum SJ, D'Vaz N, Dunstan J, Mori TA, Prescott SL (2011) The Infant Fish Oil Supplementation Study (IFOS): design and research protocol of a double-blind, randomised controlled n–3 LCPUFA intervention trial in term infants. Contemp Clin Trials 32(5):771–778. https://doi.org/10.1016/j.cct.2011.05.019
- Krauss-Etschmann S, Hartl D, Rzehak P, Heinrich J, Shadid R, Del Carmen Ramirez-Tortosa M, Campoy C, Pardillo S, Schendel DJ, Decsi T, Demmelmair H, Koletzko BV, Nutraceuticals for Healthier Life Study G (2008) Decreased cord blood IL-4, IL-13, and CCR4 and increased TGF-beta levels after fish oil supplementation of pregnant women. J Allergy Clin Immunol 121(2): 464–470 e466. https://doi.org/10.1016/j.jaci.2007.09.018
- Noakes PS, Vlachava M, Kremmyda LS, Diaper ND, Miles EA, Erlewyn-Lajeunesse M, Williams AP, Godfrey KM, Calder PC (2012) Increased intake of oily fish in pregnancy: effects on neonatal immune responses and on clinical outcomes in infants at 6 mo. Am J Clin Nutr 95(2):395–404. https://doi.org/10.3945/ajcn. 111.022954
- 73. Miles EA, Noakes PS, Kremmyda LS, Vlachava M, Diaper ND, Rosenlund G, Urwin H, Yaqoob P, Rossary A, Farges MC, Vasson MP, Liaset B, Froyland L, Helmersson J, Basu S, Garcia E, Olza J, Mesa MD, Aguilera CM, Gil A, Robinson SM, Inskip HM, Godfrey KM, Calder PC (2011) The Salmon in Pregnancy Study: study design, subject characteristics, maternal fish and marine n-3 fatty acid intake, and marine n-3 fatty acid status in maternal and umbilical cord blood. Am J Clin Nutr 94(6 Suppl): 1986S–1992S. https://doi.org/10.3945/ajcn.110.001636
- Furuhjelm C, Warstedt K, Larsson J, Fredriksson M, Bottcher MF, Falth-Magnusson K, Duchen K (2009) Fish oil supplementation in

pregnancy and lactation may decrease the risk of infant allergy. Acta Paediatr 98(9):1461–1467. https://doi.org/10.1111/j.1651-2227.2009.01355.x

- Palmer DJ, Sullivan T, Gold MS, Prescott SL, Heddle R, Gibson RA, Makrides M (2012) Effect of n-3 long chain polyunsaturated fatty acid supplementation in pregnancy on infants' allergies in first year of life: randomised controlled trial. BMJ (Clin Res ed) 344:e184. https://doi.org/10.1136/bmj.e184
- Makrides M, Gibson RA, McPhee AJ, Yelland L, Quinlivan J, Ryan P, Team DOI (2010) Effect of DHA supplementation during pregnancy on maternal depression and neurodevelopment of young children: a randomized controlled trial. JAMA 304(15): 1675–1683. https://doi.org/10.1001/jama.2010.1507
- 77. Miyake Y, Tanaka K, Okubo H, Sasaki S, Arakawa M (2013) Maternal fat intake during pregnancy and wheeze and eczema in Japanese infants: the Kyushu Okinawa Maternal and Child Health Study. Ann Epidemiol 23(11):674–680. https://doi.org/10.1016/j. annepidem.2013.08.004
- Miyake Y, Tanaka K, Okubo H, Sasaki S, Arakawa M (2013) Fish and fat intake and prevalence of depressive symptoms during pregnancy in Japan: baseline data from the Kyushu Okinawa Maternal and Child Health Study. J Psychiatr Res 47(5):572– 578. https://doi.org/10.1016/j.jpsychires.2013.01.012
- Jedrychowski W, Perera F, Maugeri U, Mrozek-Budzyn D, Miller RL, Flak E, Mroz E, Jacek R, Spengler JD (2011) Effects of prenatal and perinatal exposure to fine air pollutants and maternal fish consumption on the occurrence of infantile eczema. Int Arch Allergy Immunol 155(3):275–281. https://doi.org/10.1159/ 000320376
- Montes R, Chisaguano AM, Castellote AI, Morales E, Sunyer J, Lopez-Sabater MC (2013) Fatty-acid composition of maternal and umbilical cord plasma and early childhood atopic eczema in a Spanish cohort. Eur J Clin Nutr 67(6):658–663. https://doi.org/ 10.1038/ejcn.2013.68
- Rucci E, den Dekker HT, de Jongste JC, Steenweg-de-Graaff J, Gaillard R, Pasmans SG, Hofman A, Tiemeier H, Jaddoe VW, Duijts L (2016) Maternal fatty acid levels during pregnancy, childhood lung function and atopic diseases. The Generation R Study. Clin Exp Allergy 46(3):461–471. https://doi.org/10.1111/cea. 12613
- 82. Jaddoe VW, van Duijn CM, Franco OH, van der Heijden AJ, van lizendoorn MH, de Jongste JC, van der Lugt A, Mackenbach JP, Moll HA, Raat H, Rivadeneira F, Steegers EA, Tiemeier H, Uitterlinden AG, Verhulst FC, Hofman A (2012) The Generation R Study: design and cohort update 2012. Eur J Epidemiol 27(9): 739–756. https://doi.org/10.1007/s10654-012-9735-1
- Gardner KG, Gebretsadik T, Hartman TJ, Rosa MJ, Tylavsky FA, Adgent MA, Moore PE, Kocak M, Bush NR, Davis RL, Lewinn KZ, Wright RJ, Carroll KN (2019) Prenatal omega-3 and omega-6 polyunsaturated fatty acids and childhood atopic dermatitis. J Allergy Clin Immunol Pract. https://doi.org/10.1016/j.jaip.2019. 09.031
- Nwaru BI, Erkkola M, Lumia M, Kronberg-Kippila C, Ahonen S, Kaila M, Ilonen J, Simell O, Knip M, Veijola R, Virtanen SM (2012) Maternal intake of fatty acids during pregnancy and allergies in the offspring. Br J Nutr 108(4):720–732. https://doi.org/10. 1017/S0007114511005940
- 85. Kupila A, Muona P, Simell T, Arvilommi P, Savolainen H, Hamalainen AM, Korhonen S, Kimpimaki T, Sjoroos M, Ilonen J, Knip M, Simell O, Juvenile Diabetes Research Foundation Centre for the Prevention of Type IDiF (2001) Feasibility of genetic and immunological prediction of type I diabetes in a population-based birth cohort. Diabetologia 44(3):290–297. https://doi.org/10.1007/s001250051616
- Newson RB, Shaheen SO, Henderson AJ, Emmett PM, Sherriff A, Calder PC (2004) Umbilical cord and maternal blood red cell fatty

acids and early childhood wheezing and eczema. J Allergy Clin Immunol 114(3):531–537. https://doi.org/10.1016/j.jaci.2004.05. 010

- Golding J, Pembrey M, Jones R, Team AS (2001) ALSPAC–the Avon Longitudinal Study of Parents and Children. I Study Method Paediatr Perinat Epidemiol 15(1):74–87. https://doi.org/10.1046/j. 1365-3016.2001.00325.x
- Yu YM, Chan YH, Calder PC, Hardjojo A, Soh SE, Lim AL, Fisk HL, Teoh OH, Goh A, Saw SM, Kwek K, Gluckman PD, Godfrey KM, Chong YS, Shek LP, Pan A, Chong MF, van Bever HP, group Gs (2015) Maternal PUFA status and offspring allergic diseases up to the age of 18 months. Br J Nutr 113(6):975–983. https://doi.org/ 10.1017/S000711451500001X
- Soh SE, Lee SS, Hoon SW, Tan MY, Goh A, Lee BW, Shek LP, Teoh OH, Kwek K, Saw SM, Godfrey K, Chong YS, Gluckman P, van Bever HP (2012) The methodology of the GUSTO cohort study: a novel approach in studying pediatric allergy. Asia Pac Allergy 2(2):144–148. https://doi.org/10.5415/apallergy.2012.2. 2.144
- Standl M, Demmelmair H, Koletzko B, Heinrich J (2014) Cord blood LC-PUFA composition and allergic diseases during the first 10 yr. results from the LISAplus study. Pediatr Allergy Immunol 25(4):344–350. https://doi.org/10.1111/pai.12212
- 91. Heinrich J, Bolte G, Holscher B, Douwes J, Lehmann I, Fahlbusch B, Bischof W, Weiss M, Borte M, Wichmann HE, Group LS (2002) Allergens and endotoxin on mothers' mattresses and total immunoglobulin E in cord blood of neonates. Eur Respir J 20(3): 617–623. https://doi.org/10.1183/09031936.02.02322001
- 92. Notenboom ML, Mommers M, Jansen EH, Penders J, Thijs C (2011) Maternal fatty acid status in pregnancy and childhood atopic manifestations: KOALA Birth Cohort Study. Clin Exp Allergy 41(3):407–416. https://doi.org/10.1111/j.1365-2222.2010.03672. x
- 93. Kummeling I, Thijs C, Penders J, Snijders BE, Stelma F, Reimerink J, Koopmans M, Dagnelie PC, Huber M, Jansen MC, de Bie R, van den Brandt PA (2005) Etiology of atopy in infancy: the KOALA Birth Cohort Study. Pediatr Allergy Immunol 16(8):679–684. https://doi.org/10.1111/j.1399-3038. 2005.00333.x
- Garlantezec R, Monfort C, Rouget F, Cordier S (2009) Maternal occupational exposure to solvents and congenital malformations: a prospective study in the general population. Occup Environ Med 66(7):456–463. https://doi.org/10.1136/oem.2008.041772
- 95. D'Vaz N, Ma Y, Dunstan JA, Lee-Pullen TF, Hii C, Meldrum S, Zhang G, Metcalfe J, Ferrante A, Prescott SL (2012) Neonatal protein kinase C zeta expression determines the neonatal T-cell cytokine phenotype and predicts the development and severity of infant allergic disease. Allergy 67(12):1511–1518. https://doi. org/10.1111/all.12027
- 96. Chawes BL, Bonnelykke K, Stokholm J, Vissing NH, Bjarnadottir E, Schoos AM, Wolsk HM, Pedersen TM, Vinding RK, Thorsteinsdottir S, Arianto L, Hallas HW, Heickendorff L, Brix S, Rasmussen MA, Bisgaard H (2016) Effect of vitamin D3 supplementation during pregnancy on risk of persistent wheeze in the offspring: a randomized clinical trial. JAMA 315(4):353–361. https://doi.org/10.1001/jama.2015.18318
- 97. Litonjua AA, Carey VJ, Laranjo N, Harshfield BJ, McElrath TF, O'Connor GT, Sandel M, Iverson RE Jr, Lee-Paritz A, Strunk RC, Bacharier LB, Macones GA, Zeiger RS, Schatz M, Hollis BW, Hornsby E, Hawrylowicz C, Wu AC, Weiss ST (2016) Effect of prenatal supplementation with vitamin D on asthma or recurrent wheezing in offspring by age 3 years: the VDAART randomized clinical trial. JAMA 315(4):362–370. https://doi.org/10.1001/ jama.2015.18589
- Goldring ST, Griffiths CJ, Martineau AR, Robinson S, Yu C, Poulton S, Kirkby JC, Stocks J, Hooper R, Shaheen SO, Warner

JO, Boyle RJ (2013) Prenatal vitamin d supplementation and child respiratory health: a randomised controlled trial. PLoS One 8(6): e66627. https://doi.org/10.1371/journal.pone.0066627

- 99. Norizoe C, Akiyama N, Segawa T, Tachimoto H, Mezawa H, Ida H, Urashima M (2014) Increased food allergy and vitamin D: randomized, double-blind, placebo-controlled trial. Pediatr Int 56(1):6–12. https://doi.org/10.1111/ped.12207
- West CE, Dunstan J, McCarthy S, Metcalfe J, D'Vaz N, Meldrum S, Oddy WH, Tulic MK, Prescott SL (2012) Associations between maternal antioxidant intakes in pregnancy and infant allergic outcomes. Nutrients 4(11):1747–1758. https://doi.org/10.3390/ nu4111747
- 101. Martindale S, McNeill G, Devereux G, Campbell D, Russell G, Seaton A (2005) Antioxidant intake in pregnancy in relation to wheeze and eczema in the first two years of life. Am J Respir Crit Care Med 171(2):121–128. https://doi.org/10.1164/rccm.200402-220OC
- Hoppu U, Rinne M, Salo-Vaananen P, Lampi AM, Piironen V, Isolauri E (2005) Vitamin C in breast milk may reduce the risk of atopy in the infant. Eur J Clin Nutr 59(1):123–128. https://doi. org/10.1038/sj.ejcn.1602048
- 103. Kiefte-de Jong JC, Timmermans S, Jaddoe VW, Hofman A, Tiemeier H, Steegers EA, de Jongste JC, Moll HA (2012) High circulating folate and vitamin B-12 concentrations in women during pregnancy are associated with increased prevalence of atopic dermatitis in their offspring. J Nutr 142(4):731–738. https://doi. org/10.3945/jn.111.154948
- 104. Dunstan JA, West C, McCarthy S, Metcalfe J, Meldrum S, Oddy WH, Tulic MK, D'Vaz N, Prescott SL (2012) The relationship between maternal folate status in pregnancy, cord blood folate levels, and allergic outcomes in early childhood. Allergy 67(1): 50–57. https://doi.org/10.1111/j.1398-9995.2011.02714.x
- 105. Magdelijns FJH, Mommers M, Penders J, Smits L, Thijs C (2011) Folic acid use in pregnancy and the development of atopy, asthma, and lung function in childhood. Pediatrics 128(1):e135–e144. https://doi.org/10.1542/peds.2010-1690
- 106. Bekkers MBM, Elstgeest LEM, Scholtens S, Haveman-Nies A, de Jongste JC, Kerkhof M, Koppelman GH, Gehring U, Smit HA, Wijga AH (2012) Maternal use of folic acid supplements during pregnancy, and childhood respiratory health and atopy. Eur Respir J 39(6):1468–1474. https://doi.org/10.1183/09031936.00094511
- 107. Miyake Y, Sasaki S, Tanaka K, Hirota Y (2011) Maternal B vitamin intake during pregnancy and wheeze and eczema in Japanese infants aged 16-24 months: the Osaka Maternal and Child Health Study. Pediatr Allergy Immunol 22(1 Pt 1):69–74. https://doi.org/ 10.1111/j.1399-3038.2010.01081.x
- 108. Miyake Y, Tanaka K, Sasaki S, Kiyohara C, Ohya Y, Fukushima W, Yokoyama T, Hirota Y, Osaka M, Child Health Study G (2009) Breastfeeding and atopic eczema in Japanese infants: the Osaka Maternal and Child Health Study. Pediatr Allergy Immunol 20(3): 234–241. https://doi.org/10.1111/j.1399-3038.2008.00778.x
- 109. Kim JH, Jeong KS, Ha EH, Park H, Ha M, Hong YC, Bhang SY, Lee SJ, Lee KY, Lee SH, Kim Y, Kim MH, Chang N (2015) Relationship between prenatal and postnatal exposures to folate and risks of allergic and respiratory diseases in early childhood. Pediatr Pulmonol 50(2):155–163. https://doi.org/10.1002/ppul. 23025
- 110. Kim BM, Ha M, Park HS, Lee BE, Kim YJ, Hong YC, Kim Y, Chang N, Roh YM, Kim BN, Oh SY, Ha EH, Group MS (2009) The Mothers and Children's Environmental Health (MOCEH) study. Eur J Epidemiol 24(9):573–583. https://doi.org/10.1007/ s10654-009-9370-7
- 111. Volgyi E, Carroll KN, Hare ME, Ringwald-Smith K, Piyathilake C, Yoo W, Tylavsky FA (2013) Dietary patterns in pregnancy and effects on nutrient intake in the mid-south: the Conditions Affecting Neurocognitive Development and Learning in Early

Childhood (CANDLE) study. Nutrients 5(5):1511–1530. https:// doi.org/10.3390/nu5051511

- 112. Roy A, Kocak M, Hartman TJ, Vereen S, Adgent M, Piyathilake C, Tylavsky FA, Carroll KN (2018) Association of prenatal folate status with early childhood wheeze and atopic dermatitis. Pediatr Allergy Immunol 29(2):144–150. https://doi.org/10.1111/pai. 12834
- 113. van der Valk RJ, Kiefte-de Jong JC, Sonnenschein-van der Voort AM, Duijts L, Hafkamp-de Groen E, Moll HA, Tiemeier H, Steegers EA, Hofman A, Jaddoe VW, de Jongste JC (2013) Neonatal folate, homocysteine, vitamin B12 levels and methylenetetrahydrofolate reductase variants in childhood asthma and eczema. Allergy 68(6):788–795. https://doi.org/10.1111/all.12146
- 114. El-Heis S, Crozier SR, Robinson SM, Harvey NC, Cooper C, Inskip HM, Southampton Women's Survey Study G, Godfrey KM (2016) Higher maternal serum concentrations of nicotinamide and related metabolites in late pregnancy are associated with a lower risk of offspring atopic eczema at age 12 months. Clin Exp Allergy 46(10):1337–1343. https://doi.org/10.1111/cea. 12782
- 115. Nwaru BI, Ahonen S, Kaila M, Erkkola M, Haapala AM, Kronberg-Kippila C, Veijola R, Ilonen J, Simell O, Knip M, Virtanen SM (2010) Maternal diet during pregnancy and allergic sensitization in the offspring by 5 yrs of age: a prospective cohort study. Pediatr Allergy Immunol 21(1 Pt 1):29–37. https://doi.org/ 10.1111/j.1399-3038.2009.00949.x
- 116. Baiz N, Dargent-Molina P, Wark JD, Souberbielle JC, Annesi-Maesano I, Group EM-CCS (2014) Cord serum 25hydroxyvitamin D and risk of early childhood transient wheezing and atopic dermatitis. J Allergy Clin Immunol 133(1):147–153. https://doi.org/10.1016/j.jaci.2013.05.017
- 117. Jones AP, Palmer D, Zhang G, Prescott SL (2012) Cord blood 25hydroxyvitamin D3 and allergic disease during infancy. Pediatrics 130(5):e1128–e1135. https://doi.org/10.1542/peds.2012-1172
- 118. Gale CR, Robinson SM, Harvey NC, Javaid MK, Jiang B, Martyn CN, Godfrey KM, Cooper C, Princess Anne Hospital Study G (2008) Maternal vitamin D status during pregnancy and child outcomes. Eur J Clin Nutr 62(1):68–77. https://doi.org/10.1038/sj. ejcn.1602680
- 119. Wills AK, Shaheen SO, Granell R, Henderson AJ, Fraser WD, Lawlor DA (2013) Maternal 25-hydroxyvitamin D and its association with childhood atopic outcomes and lung function. Clin Exp Allergy 43(10):1180–1188. https://doi.org/10.1111/cea.12172
- 120. Miyake Y, Tanaka K, Okubo H, Sasaki S, Arakawa M (2014) Maternal consumption of dairy products, calcium, and vitamin D during pregnancy and infantile allergic disorders. Ann Allergy Asthma Immunol 113(1):82–87. https://doi.org/10.1016/j.anai. 2014.04.023
- 121. Chiu CY, Huang SY, Peng YC, Tsai MH, Hua MC, Yao TC, Yeh KW, Huang JL (2015) Maternal vitamin D levels are inversely related to allergic sensitization and atopic diseases in early childhood. Pediatr Allergy Immunol 26(4):337–343. https://doi.org/10. 1111/pai.12384
- Wang LC, Chiang BL, Huang YM, Shen PT, Huang HY, Lin BF (2019) Lower vitamin D levels in the breast milk is associated with atopic dermatitis in early infancy. Pediatr Allergy Immunol. https://doi.org/10.1111/pai.13179
- 123. Miyake Y, Sasaki S, Tanaka K, Ohfuji S, Hirota Y (2009) Maternal fat consumption during pregnancy and risk of wheeze and eczema in Japanese infants aged 16-24 months: the Osaka Maternal and Child Health Study. Thorax 64(9):815–821. https://doi.org/10.1136/thx.2009.115931
- 124. Pele F, Bajeux E, Gendron H, Monfort C, Rouget F, Multigner L, Viel JF, Cordier S (2013) Maternal fish and shellfish consumption and wheeze, eczema and food allergy at age two: a prospective

cohort study in Brittany, France. Environ Health 12:102. https:// doi.org/10.1186/1476-069X-12-102

- 125. Jiminez JA, Uwiera TC, Abbott DW, Uwiera RRE, Inglis GD (2017) Butyrate supplementation at high concentrations alters enteric bacterial communities and reduces intestinal inflammation in mice infected with Citrobacter rodentium. mSphere 2(4):e00243– e00217. https://doi.org/10.1128/mSphere.00243-17
- Leonel AJ, Alvarez-Leite JI (2012) Butyrate: implications for intestinal function. Curr Opin Clin Nutr Metab Care 15(5):474–479. https://doi.org/10.1097/MCO.0b013e32835665fa
- 127. Furusawa Y, Obata Y, Fukuda S, Endo TA, Nakato G, Takahashi D, Nakanishi Y, Uetake C, Kato K, Kato T, Takahashi M, Fukuda NN, Murakami S, Miyauchi E, Hino S, Atarashi K, Onawa S, Fujimura Y, Lockett T, Clarke JM, Topping DL, Tomita M, Hori S, Ohara O, Morita T, Koseki H, Kikuchi J, Honda K, Hase K, Ohno H (2013) Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells. Nature 504(7480): 446–450. https://doi.org/10.1038/nature12721
- 128. Arpaia N, Campbell C, Fan X, Dikiy S, van der Veeken J, deRoos P, Liu H, Cross JR, Pfeffer K, Coffer PJ, Rudensky AY (2013) Metabolites produced by commensal bacteria promote peripheral regulatory T-cell generation. Nature 504(7480):451–455. https:// doi.org/10.1038/nature12726
- 129. Fernández J, Redondo-Blanco S, Gutiérrez-del-Río I, Miguélez EM, Villar CJ, Lombó F (2016) Colon microbiota fermentation of dietary prebiotics towards short-chain fatty acids and their roles as anti-inflammatory and antitumour agents: a review. J Funct Foods 25:511–522. https://doi.org/10.1016/j.jff.2016.06.032
- Vinolo MA, Rodrigues HG, Nachbar RT, Curi R (2011) Regulation of inflammation by short chain fatty acids. Nutrients 3(10):858–876. https://doi.org/10.3390/nu3100858
- Böttcher MF, Nordin EK, Sandin A, Midtvedt T, Björkstén B (2000) Microflora-associated characteristics in faeces from allergic and nonallergic infants. Clin Exp Allergy 30(11):1591–1596. https://doi.org/10.1046/j.1365-2222.2000.00982.x
- 132. Nylund L, Nermes M, Isolauri E, Salminen S, de Vos WM, Satokari R (2015) Severity of atopic disease inversely correlates with intestinal microbiota diversity and butyrate-producing bacteria. Allergy 70(2):241–244. https://doi.org/10.1111/all.12549
- 133. Kim HK, Rutten NB, Besseling-van der Vaart I, Niers LE, Choi YH, Rijkers GT, van Hemert S (2015) Probiotic supplementation influences faecal short chain fatty acids in infants at high risk for eczema. Benefic Microbes 6(6):783–790. https://doi.org/10.3920/ BM2015.0056
- Mousa A, Naqash A, Lim S (2019) Macronutrient and micronutrient intake during pregnancy: an overview of recent evidence. Nutrients 11(2). https://doi.org/10.3390/nu11020443
- Kedia-Mehta N, Finlay DK (2019) Competition for nutrients and its role in controlling immune responses. Nat Commun 10(1): 2123. https://doi.org/10.1038/s41467-019-10015-4
- 136. Berentzen NE, van Stokkom VL, Gehring U, Koppelman GH, Schaap LA, Smit HA, Wijga AH (2015) Associations of sugarcontaining beverages with asthma prevalence in 11-year-old children: the PIAMA birth cohort. Eur J Clin Nutr 69(3):303–308. https://doi.org/10.1038/ejcn.2014.153
- Ehlers I, Worm M, Sterry W, Zuberbier T (2001) Sugar is not an aggravating factor in atopic dermatitis. Acta Derm Venereol 81(4): 282–284. https://doi.org/10.1080/00015550152572930
- Bedard A, Northstone K, Henderson AJ, Shaheen SO (2017) Maternal intake of sugar during pregnancy and childhood respiratory and atopic outcomes. Eur Respir J 50(1). https://doi.org/10. 1183/13993003.00073-2017
- Davani-Davari D, Negahdaripour M, Karimzadeh I, Seifan M, Mohkam M, Masoumi SJ, Berenjian A, Ghasemi Y (2019) Prebiotics: definition, types, sources, mechanisms, and clinical applications. Foods 8(3). https://doi.org/10.3390/foods8030092

- Brosseau C, Selle A, Palmer DJ, Prescott SL, Barbarot S, Bodinier M (2019) Prebiotics: mechanisms and preventive effects in allergy. Nutrients 11(8). https://doi.org/10.3390/nu11081841
- 141. Wu RY, Abdullah M, Määttänen P, Pilar AVC, Scruten E, Johnson-Henry KC, Napper S, O'Brien C, Jones NL, Sherman PM (2017) Protein kinase C δ signaling is required for dietary prebiotic-induced strengthening of intestinal epithelial barrier function. Sci Rep 7(1). https://doi.org/10.1038/srep40820
- 142. Capitan-Canadas F, Ortega-Gonzalez M, Guadix E, Zarzuelo A, Suarez MD, de Medina FS, Martinez-Augustin O (2014) Prebiotic oligosaccharides directly modulate proinflammatory cytokine production in monocytes via activation of TLR4. Mol Nutr Food Res 58(5):1098–1110. https://doi.org/10.1002/mnfr.201300497
- Cummings JH, Pomare EW, Branch WJ, Naylor CP, Macfarlane GT (1987) Short chain fatty acids in human large intestine, portal, hepatic and venous blood. Gut 28(10):1221–1227. https://doi.org/ 10.1136/gut.28.10.1221
- 144. Fukuda S, Toh H, Hase K, Oshima K, Nakanishi Y, Yoshimura K, Tobe T, Clarke JM, Topping DL, Suzuki T, Taylor TD, Itoh K, Kikuchi J, Morita H, Hattori M, Ohno H (2011) Bifidobacteria can protect from enteropathogenic infection through production of acetate. Nature 469(7331):543–547. https://doi.org/10.1038/ nature09646
- 145. Rusu E, Enache G, Cursaru R, Alexescu A, Radu R, Onila O, Cavallioti T, Rusu F, Posea M, Jinga M, Radulian G (2019) Prebiotics and probiotics in atopic dermatitis. Exp Ther Med 18(2):926–931. https://doi.org/10.3892/etm.2019.7678
- 146. Fujiwara R, Takemura N, Watanabe J, Sonoyama K (2010) Maternal consumption of fructo-oligosaccharide diminishes the severity of skin inflammation in offspring of NC/Nga mice. Br J Nutr 103(4):530-538. https://doi.org/10.1017/ S000711450999198X
- 147. Cuello-Garcia CA, Fiocchi A, Pawankar R, Yepes-Nunez JJ, Morgano GP, Zhang Y, Ahn K, Al-Hammadi S, Agarwal A, Gandhi S, Beyer K, Burks W, Canonica GW, Ebisawa M, Kamenwa R, Lee BW, Li H, Prescott S, Riva JJ, Rosenwasser L, Sampson H, Spigler M, Terracciano L, Vereda A, Waserman S, Schunemann HJ, Brozek JL (2016) World allergy organization-McMaster University Guidelines for Allergic Disease Prevention (GLAD-P): prebiotics. World Allergy Organ J 9:10. https://doi. org/10.1186/s40413-016-0102-7
- 148. Pretorius R, Prescott SL, Palmer DJ (2018) Taking a prebiotic approach to early immunomodulation for allergy prevention. Expert Rev Clin Immunol 14(1):43–51. https://doi.org/10.1080/ 1744666X.2018.1411191
- 149. Cabridain C, Aubert H, Kaeffer B, Badon V, Boivin M, Dochez V, Winer N, Faurel-Paul E, Planche L, Riochet D, Maruani A, Perrotin F, Droitcourt C, Lassel L, Tching-Sin M, Rogers NK, Bodinier M, Barbarot S (2019) Effectiveness of an antenatal maternal supplementation with prebiotics for preventing atopic dermatitis in high-risk children (the PREGRALL study): protocol for a randomised controlled trial. BMJ Open 9(4):e024974. https:// doi.org/10.1136/bmjopen-2018-024974
- 150. Roberfroid M (1993) Dietary fiber, inulin, and oligofructose: a review comparing their physiological effects. Crit Rev Food Sci Nutr 33(2):103-148. https://doi.org/10.1080/ 10408399309527616
- 151. Gibson GR, Hutkins R, Sanders ME, Prescott SL, Reimer RA, Salminen SJ, Scott K, Stanton C, Swanson KS, Cani PD, Verbeke K, Reid G (2017) Expert consensus document: The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of prebiotics. Nat Rev Gastroenterol Hepatol 14(8):491–502. https://doi. org/10.1038/nrgastro.2017.75
- 152. The Scientific Advisory Committee on Nutrition (2006) Folate and disease prevention. The Stationary Office, London. https://

assets.publishing.service.gov.uk/government/uploads/system/ uploads/attachment_data/file/338892/SACN_Folate_and_ Disease_Prevention_Report.pdf. Accessed 13 Dec 2019

- 153. Centers for Disease Control and Prevention (1992) Recommendations for the use of folic acid to reduce the number of cases of spina bifida and other neural tube defects. MMWR Recomm Rep 41(Rr–14):1–7
- 154. Nowak-Wegrzyn A (2007) Food allergy to proteins. Nestle Nutr Workshop Ser Pediatr Program 59:17–31; discussion 31-16. https://doi.org/10.1159/000098510
- 155. Eigenmann PA, Sicherer SH, Borkowski TA, Cohen BA, Sampson HA (1998) Prevalence of IgE-mediated food allergy among children with atopic dermatitis. Pediatrics 101(3):E8. https://doi.org/10.1542/peds.101.3.e8
- Bergmann MM, Caubet JC, Boguniewicz M, Eigenmann PA (2013) Evaluation of food allergy in patients with atopic dermatitis. J Allergy Clin Immunol Pract 1(1):22–28. https://doi.org/10. 1016/j.jaip.2012.11.005
- Hay WW Jr (1994) Placental transport of nutrients to the fetus. Horm Res 42(4–5):215–222. https://doi.org/10.1159/000184196
- Edelbauer M, Loibichler C, Nentwich I, Gerstmayr M, Urbanek R, Szepfalusi Z (2004) Maternally delivered nutritive allergens in cord blood and in placental tissue of term and preterm neonates. Clin Exp Allergy 34(2):189–193. https://doi.org/10.1111/j.1365-2222.2004.01848.x
- Jones CA, Vance GH, Power LL, Pender SL, Macdonald TT, Warner JO (2001) Costimulatory molecules in the developing human gastrointestinal tract: a pathway for fetal allergen priming. J Allergy Clin Immunol 108(2):235–241. https://doi.org/10.1067/ mai.2001.117178
- Loibichler C, Pichler J, Gerstmayr M, Bohle B, Kisst H, Urbanek R, Szepfalusi Z (2002) Materno-fetal passage of nutritive and inhalant allergens across placentas of term and pre-term deliveries perfused in vitro. Clin Exp Allergy 32(11):1546–1551. https://doi. org/10.1046/j.1365-2222.2002.01479.x
- 161. Sorva R, Makinenkiljunen S, Juntunenbackman K (1994) β-Lactoglobulin secretion in human milk varies widely after cow's milk ingestion in mothers of infants with cow's milk allergy. J Allergy Clin Immunol 93(4):787–792. https://doi.org/10.1016/ 0091-6749(94)90259-3
- 162. Cant A, Marsden RA, Kilshaw PJ (1985) Egg and cows' milk hypersensitivity in exclusively breast fed infants with eczema, and detection of egg protein in breast milk. Br Med J (Clin Res Ed) 291(6500):932–935. https://doi.org/10.1136/bmj.291.6500. 932
- Palmer DJ, Gold MS, Makrides M (2008) Effect of maternal egg consumption on breast milk ovalbumin concentration. Clin Exp Allergy 38(7):1186–1191. https://doi.org/10.1111/j.1365-2222. 2008.03014.x
- 164. Palmer DJ, Gold MS, Makrides M (2005) Effect of cooked and raw egg consumption on ovalbumin content of human milk: a randomized, double-blind, cross-over trial. Clin Exp Allergy 35(2):173–178. https://doi.org/10.1111/j.1365-2222.2005.02170.
- 165. Vadas P, Wai Y, Burks W, Perelman B (2001) Detection of peanut allergens in breast milk of lactating women. JAMA 285(13): 1746–1748. https://doi.org/10.1001/jama.285.13.1746
- 166. Lovegrove JA, Morgan JB, Hamptom SM (1996) Dietary factors influencing levels of food antibodies and antigens in breast milk. Acta Paediatr 85(7):778–784. https://doi.org/10.1111/j.1651-2227.1996.tb14151.x
- 167. Lilja G, Dannaeus A, Foucard T, Graff-Lonnevig V, Johansson SG, Oman H (1991) Effects of maternal diet during late pregnancy and lactation on the development of IgE and egg- and milk-specific IgE and IgG antibodies in infants. Clin Exp Allergy

21(2):195–202. https://doi.org/10.1111/j.1365-2222.1991. tb00830.x

- 168. Greer FR, Sicherer SH, Burks AW, Committee On N, Section On A, Immunology (2019) The effects of early nutritional interventions on the development of atopic disease in infants and children. Pediatrics 143(4). https://doi.org/10.1542/peds.2019-0281
- Cant AJ, Bailes JA, Marsden RA, Hewitt D (1986) Effect of maternal dietary exclusion on breast fed infants with eczema: two controlled studies. Br Med J (Clin Res Ed) 293(6541):231–233. https://doi.org/10.1136/bmj.293.6541.231
- Kramer MS, Kakuma R (2012) Maternal dietary antigen avoidance during pregnancy or lactation, or both, for preventing or treating atopic disease in the child. Cochrane Database Syst Rev 9:CD000133. https://doi.org/10.1002/14651858.CD000133.pub3
- 171. Falth-Magnusson K, Oman H, Kjellman NI (1987) Maternal abstention from cow milk and egg in allergy risk pregnancies. Effect on antibody production in the mother and the newborn. Allergy 42(1):64–73. https://doi.org/10.1111/j.1398-9995.1987.tb02189.x
- 172. Lilja G, Dannaeus A, Foucard T, Graff-Lonnevig V, Johansson SG, Oman H (1989) Effects of maternal diet during late pregnancy and lactation on the development of atopic diseases in infants up to 18 months of age–in-vivo results. Clin Exp Allergy 19(4):473–479. https://doi.org/10.1111/j.1365-2222.1989.tb02416.x
- 173. Zeiger R, Heller S, Mellon M, Forsythe A, Oconnor R, Hamburger R, Schatz M (1989) Effect of combined maternal and infant food-allergen avoidance on development of atopy in early infancy: a randomized study. J Allergy Clin Immunol 84(1): 72–89. https://doi.org/10.1016/0091-6749(89)90181-4
- 174. Arshad SH, Matthews S, Gant C, Hide DW (1992) Effect of allergen avoidance on development of allergic disorders in infancy. Lancet 339(8808):1493–1497. https://doi.org/10.1016/0140-6736(92)91260-f
- 175. Lovegrove JA, Hampton SM, Morgan JB (1994) The immunological and long-term atopic outcome of infants born to women following a milk-free diet during late pregnancy and lactation: a pilot study. Br J Nutr 71(2):223–238. https://doi.org/10.1079/ bjn19940129
- 176. Webster D, Dhondt JL, Hannon WH, Loeber G, Torresani T (1999) Quality assurance and standardization: summary of the satellite meeting, Turku, Finland, 11-12 June 1999. Acta Paediatr 88:7–12. https://doi.org/10.1080/08035259950170510
- 177. Herrmann ME, Dannemann A, Gruters A, Radisch B, Dudenhausen JW, Bergmann R, Coumbos A, Weitzel HK, Wahn U (1996) Prospective study of the atopy preventive effect of maternal avoidance of milk and eggs during pregnancy and lactation. Eur J Pediatr 155(9):770–774. https://doi.org/10.1007/ bf02002904
- 178. de Silva D, Geromi M, Halken S, Host A, Panesar SS, Muraro A, Werfel T, Hoffmann-Sommergruber K, Roberts G, Cardona V, Dubois AE, Poulsen LK, Van Ree R, Vlieg-Boerstra B, Agache I, Grimshaw K, O'Mahony L, Venter C, Arshad SH, Sheikh A (2014) Primary prevention of food allergy in children and adults: systematic review. Allergy 69(5):581–589. https://doi.org/10. 1111/all.12334
- Devereux G, Seaton A (2005) Diet as a risk factor for atopy and asthma. J Allergy Clin Immunol 115(6):1109–1117; quiz 1118. https://doi.org/10.1016/j.jaci.2004.12.1139
- Parigi SM, Eldh M, Larssen P, Gabrielsson S, Villablanca EJ (2015) Breast milk and solid food shaping intestinal immunity. Front Immunol 6:415. https://doi.org/10.3389/fimmu.2015.00415
- 181. Omata N, Tsukahara H, Ito S, Ohshima Y, Yasutomi M, Yamada A, Jiang M, Hiraoka M, Nambu M, Deguchi Y, Mayumi M (2001) Increased oxidative stress in childhood atopic dermatitis. Life Sci 69(2):223–228. https://doi.org/10.1016/s0024-3205(01)01124-9

- Ji H, Li XK (2016) Oxidative stress in atopic dermatitis. Oxidative Med Cell Longev 2016:2721469. https://doi.org/10.1155/2016/ 2721469
- Sivaranjani N, Rao SV, Rajeev G (2013) Role of reactive oxygen species and antioxidants in atopic dermatitis. J Clin Diagn Res 7(12):2683–2685. https://doi.org/10.7860/jcdr/2013/6635.3732
- Crider KS, Yang TP, Berry RJ, Bailey LB (2012) Folate and DNA methylation: a review of molecular mechanisms and the evidence for folate's role. Adv Nutr 3(1):21–38. https://doi.org/10.3945/an. 111.000992
- 185. Folate and disease prevention. (2006). Scientific Advisory Committee on Nutrition
- 186. (1992) Recommendations for the use of folic acid to reduce the number of cases of spina bifida and other neural tube defects. MMWR vol 41(RR-14):1–7
- Waterland RA, Jirtle RL (2004) Early nutrition, epigenetic changes at transposons and imprinted genes, and enhanced susceptibility to adult chronic diseases. Nutrition 20(1):63–68. https://doi.org/ 10.1016/j.nut.2003.09.011
- Prescott SL, Clifton V (2009) Asthma and pregnancy: emerging evidence of epigenetic interactions in utero. Curr Opin Allergy Clin Immunol 9(5):417–426. https://doi.org/10.1097/ACI. 0b013e328330634f
- Warner JA, Miles EA, Jones AC, Quint DJ, Colwell BM, Warner JO (1994) Is deficiency of interferon gamma production by allergen triggered cord blood cells a predictor of atopic eczema? Clin Exp Allergy 24(5):423–430. https://doi.org/10.1111/j.1365-2222. 1994.tb00930.x
- 190. Herberth G, Heinrich J, Roder S, Figl A, Weiss M, Diez U, Borte M, Herbarth O, Lehmann I, group Ls (2010) Reduced IFN-gamma- and enhanced IL-4-producing CD4+ cord blood T cells are associated with a higher risk for atopic dermatitis during the first 2 yr of life. Pediatr Allergy Immunol 21(1 Pt 1):5–13. https://doi.org/10.1111/j.1399-3038.2009.00890.x
- 191. Keating E, Correia-Branco A, Araújo JR, Meireles M, Fernandes R, Guardão L, Guimarães JT, Martel F, Calhau C (2015) Excess perigestational folic acid exposure induces metabolic dysfunction in post-natal life. J Endocrinol 224(3):245–259. https://doi.org/10. 1530/joe-14-0448
- 192. Crider KS, Cordero AM, Qi YP, Mulinare J, Dowling NF, Berry RJ (2013) Prenatal folic acid and risk of asthma in children: a systematic review and meta-analysis. Am J Clin Nutr 98(5): 1272–1281. https://doi.org/10.3945/ajcn.113.065623
- 193. Husemoen LL, Toft U, Fenger M, Jorgensen T, Johansen N, Linneberg A (2006) The association between atopy and factors influencing folate metabolism: is low folate status causally related to the development of atopy? Int J Epidemiol 35(4):954–961. https://doi.org/10.1093/ije/dy1094
- Moll R, Davis B (2017) Iron, vitamin B 12 and folate. Medicine 45(4):198–203. https://doi.org/10.1016/j.mpmed.2017.01.007
- 195. Stucker M, Pieck C, Stoerb C, Niedner R, Hartung J, Altmeyer P (2004) Topical vitamin B12–a new therapeutic approach in atopic dermatitis-evaluation of efficacy and tolerability in a randomized placebo-controlled multicentre clinical trial. Br J Dermatol 150(5):977–983. https://doi.org/ 10.1111/j.1365-2133.2004.05866.x
- Januchowski R (2009) Evaluation of topical vitamin B(12) for the treatment of childhood eczema. J Altern Complement Med 15(4): 387–389. https://doi.org/10.1089/acm.2008.0497
- 197. Molloy AM, Daly S, Mills JL, Kirke PN, Whitehead AS, Ramsbottom D, Conley MR, Weir DG, Scott JM (1997) Thermolabile variant of 5, 10-methylenetetrahydrofolate reductaseassociated with low red-cell folates: implications for folate intake recommendations. Lancet 349(9065):1591–1593. https://doi.org/10.1016/s0140-6736(96)12049-3

- 198. Puri M, Kaur L, Walia GK, Mukhopadhhyay R, Sachdeva MP, Trivedi SS, Ghosh PK, Saraswathy KN (2013) MTHFR C677T polymorphism, folate, vitamin B12 and homocysteine in recurrent pregnancy losses: a case control study among north Indian women. J Perinat Med 41(5):549–554. https://doi.org/10.1515/jpm-2012-0252
- 199. Liew SC, Gupta ED (2015) Methylenetetrahydrofolate reductase (MTHFR) C677T polymorphism: epidemiology, metabolism and the associated diseases. Eur J Med Genet 58(1):1–10. https://doi. org/10.1016/j.ejmg.2014.10.004
- 200. Granell R, Heron J, Lewis S, Davey Smith G, Sterne JA, Henderson J (2008) The association between mother and child MTHFR C677T polymorphisms, dietary folate intake and childhood atopy in a population-based, longitudinal birth cohort. Clin Exp Allergy 38(2):320–328. https://doi.org/10.1111/j.1365-2222. 2007.02902.x
- Soma Y, Kashima M, Imaizumi A, Takahama H, Kawakami T, Mizoguchi M (2005) Moisturizing effects of topical nicotinamide on atopic dry skin. Int J Dermatol 44(3):197–202. https://doi.org/ 10.1111/j.1365-4632.2004.02375.x
- 202. Chen AC, Martin AJ, Dalziell RA, Halliday GM, Damian DL (2016) Oral nicotinamide reduces transepidermal water loss: a randomized controlled trial. Br J Dermatol 175(6):1363–1365. https://doi.org/10.1111/bjd.14648
- 203. Kouda K, Tanaka T, Kouda M, Takeuchi H, Takeuchi A, Nakamura H, Takigawa M (2000) Low-energy diet in atopic dermatitis patients: clinical findings and DNA damage. J Physiol Anthropol Appl Hum Sci 19(5):225–228. https://doi.org/10. 2114/jpa.19.225
- Shin J, Kim YJ, Kwon O, Kim NI, Cho Y (2016) Associations among plasma vitamin C, epidermal ceramide and clinical severity of atopic dermatitis. Nutr Res Pract 10(4):398–403. https://doi. org/10.4162/nrp.2016.10.4.398
- 205. Vaughn AR, Foolad N, Maarouf M, Tran KA, Shi VY (2019) Micronutrients in atopic dermatitis: a systematic review. J Altern Complement Med 25(6):567–577. https://doi.org/10.1089/acm. 2018.0363
- Umar M, Sastry KS, Al Ali F, Al-Khulaifi M, Wang E, Chouchane AI (2018) Vitamin D and the pathophysiology of inflammatory skin diseases. Skin Pharmacol Physiol 31(2):74–86. https://doi. org/10.1159/000485132
- Holick MF (2007) Vitamin D deficiency. N Engl J Med 357(3): 266–281. https://doi.org/10.1056/NEJMra070553
- Palmer DJ (2015) Vitamin D and the development of atopic eczema. J Clin Med 4(5):1036–1050. https://doi.org/10.3390/ jcm4051036
- Muehleisen B, Gallo RL (2013) Vitamin D in allergic disease: shedding light on a complex problem. J Allergy Clin Immunol 131(2):324–329. https://doi.org/10.1016/j.jaci.2012.12.1562
- Di Filippo P, Scaparrotta A, Rapino D, Cingolani A, Attanasi M, Petrosino MI, Chuang K, Di Pillo S, Chiarelli F (2015) Vitamin D supplementation modulates the immune system and improves atopic dermatitis in children. Int Arch Allergy Immunol 166(2): 91–96. https://doi.org/10.1159/000371350
- 211. Bikle DD, Chang S, Crumrine D, Elalieh H, Man MQ, Choi EH, Dardenne O, Xie Z, Arnaud RS, Feingold K, Elias PM (2004) 25 Hydroxyvitamin D 1 alpha-hydroxylase is required for optimal epidermal differentiation and permeability barrier homeostasis. J Investig Dermatol 122(4):984–992. https://doi.org/10.1111/j. 0022-202X.2004.22424.x
- 212. Liu PT, Stenger S, Li H, Wenzel L, Tan BH, Krutzik SR, Ochoa MT, Schauber J, Wu K, Meinken C, Kamen DL, Wagner M, Bals R, Steinmeyer A, Zugel U, Gallo RL, Eisenberg D, Hewison M, Hollis BW, Adams JS, Bloom BR, Modlin RL (2006) Toll-like receptor triggering of a vitamin D-mediated human antimicrobial

response. Science 311(5768):1770–1773. https://doi.org/10.1126/ science.1123933

- 213. Schauber J, Dorschner RA, Coda AB, Buchau AS, Liu PT, Kiken D, Helfrich YR, Kang S, Elalieh HZ, Steinmeyer A, Zugel U, Bikle DD, Modlin RL, Gallo RL (2007) Injury enhances TLR2 function and antimicrobial peptide expression through a vitamin D-dependent mechanism. J Clin Invest 117(3):803–811. https://doi.org/10.1172/jci30142
- 214. Yamanaka K, Dimitroff CJ, Fuhlbrigge RC, Kakeda M, Kurokawa I, Mizutani H, Kupper TS (2008) Vitamins A and D are potent inhibitors of cutaneous lymphocyte-associated antigen expression. J Allergy Clin Immunol 121(1):148–157 e143. https://doi.org/10. 1016/j.jaci.2007.08.014
- 215. Thuesen BH, Heede NG, Tang L, Skaaby T, Thyssen JP, Friedrich N, Linneberg A (2015) No association between vitamin D and atopy, asthma, lung function or atopic dermatitis: a prospective study in adults. Allergy 70(11):1501–1504. https://doi.org/10. 1111/all.12704
- 216. WHO (2012) Guideline: daily iron and folic acid supplementation in pregnant women. World Health Organization, Geneva [archived]. https://www.who.int/nutrition/publications/ micronutrients/guidelines/daily_ifa_supp_pregnant_women/en/. Accessed 13 Dec 2019
- 217. Heine G, Hoefer N, Franke A, Nothling U, Schumann RR, Hamann L, Worm M (2013) Association of vitamin D receptor gene polymorphisms with severe atopic dermatitis in adults. Br J Dermatol 168(4):855–858. https://doi.org/10.1111/bjd.12077
- Wang SS, Hon KL, Kong AP, Pong HN, Wong GW, Leung TF (2014) Vitamin D deficiency is associated with diagnosis and severity of childhood atopic dermatitis. Pediatr Allergy Immunol 25(1):30–35. https://doi.org/10.1111/pai.12167
- 219. Jang YH, Sim HB, Moon SY, Lee WJ, Lee SJ, Jin M, Kim SH, Kim DW (2017) House dust mite sensitization is inversely associated with plasma 25-hydroxyvitamin D3 levels in patients with severe atopic dermatitis. Ann Dermatol 29(4):400–406. https:// doi.org/10.5021/ad.2017.29.4.400
- 220. Weisse K, Winkler S, Hirche F, Herberth G, Hinz D, Bauer M, Roder S, Rolle-Kampczyk U, von Bergen M, Olek S, Sack U, Richter T, Diez U, Borte M, Stangl GI, Lehmann I (2013) Maternal and newborn vitamin D status and its impact on food allergy development in the German LINA cohort study. Allergy 68(2):220–228. https://doi.org/10.1111/all.12081
- 221. Wolsk HM, Chawes BL, Litonjua AA, Hollis BW, Waage J, Stokholm J, Bonnelykke K, Bisgaard H, Weiss ST (2017) Prenatal vitamin D supplementation reduces risk of asthma/ recurrent wheeze in early childhood: a combined analysis of two randomized controlled trials. PLoS One 12(10):e0186657. https:// doi.org/10.1371/journal.pone.0186657
- 222. Carpenter TO, Zhang JH, Parra E, Ellis BK, Simpson C, Lee WM, Balko J, Fu L, Wong BY, Cole DE (2013) Vitamin D binding protein is a key determinant of 25-hydroxyvitamin D levels in infants and toddlers. J Bone Miner Res 28(1):213–221. https:// doi.org/10.1002/jbmr.1735
- 223. Tucker LA (2017) Alpha- and gamma-tocopherol and telomere length in 5768 US men and women: a NHANES study. Nutrients 9(6). https://doi.org/10.3390/nu9060601
- 224. Jaffary F, Faghihi G, Mokhtarian A, Hosseini SM (2015) Effects of oral vitamin E on treatment of atopic dermatitis: a randomized controlled trial. J Res Med Sci 20(11):1053–1057. https://doi.org/ 10.4103/1735-1995.172815
- 225. Tsoureli-Nikita E, Hercogova J, Lotti T, Menchini G (2002) Evaluation of dietary intake of vitamin E in the treatment of atopic dermatitis: a study of the clinical course and evaluation of the immunoglobulin E serum levels. Int J Dermatol 41(3):146–150. https://doi.org/10.1046/j.1365-4362.2002.01423.x

- Hoppu U, Salo-Vaananen P, Lampi AM, Isolauri E (2005) Serum alpha- and gamma-tocopherol levels in atopic mothers and their infants are correlated. Biol Neonate 88(1):24–26. https://doi.org/ 10.1159/000084068
- Uwitonze AM, Razzaque MS (2018) Role of magnesium in vitamin D activation and function. J Am Osteopath Assoc 118(3): 181–189. https://doi.org/10.7556/jaoa.2018.037
- 228. Proksch E, Nissen HP, Bremgartner M, Urquhart C (2005) Bathing in a magnesium-rich Dead Sea salt solution improves skin barrier function, enhances skin hydration, and reduces inflammation in atopic dry skin. Int J Dermatol 44(2):151–157. https://doi.org/10.1111/j.1365-4632.2005.02079.x
- 229. Mazur A, Maier JA, Rock E, Gueux E, Nowacki W, Rayssiguier Y (2007) Magnesium and the inflammatory response: potential physiopathological implications. Arch Biochem Biophys 458(1): 48–56. https://doi.org/10.1016/j.abb.2006.03.031
- Toyran M, Kaymak M, Vezir E, Harmanci K, Kaya A, Ginis T, Kose G, Kocabas CN (2012) Trace element levels in children with atopic dermatitis. J Investig Allergol Clin Immunol 22(5):341– 344
- 231. Daily iron and folic acid supplementation in pregnant women World Health Organization
- Baker RD, Greer FR, Committee on Nutrition American Academy of P (2010) Diagnosis and prevention of iron deficiency and irondeficiency anemia in infants and young children (0-3 years of age). Pediatrics 126(5):1040–1050. https://doi.org/10.1542/peds.2010-2576
- MacKenzie EL, Iwasaki K, Tsuji Y (2008) Intracellular iron transport and storage: from molecular mechanisms to health implications. Antioxid Redox Signal 10(6):997–1030. https://doi.org/10. 1089/ars.2007.1893
- Roth-Walter F, Pacios LF, Bianchini R, Jensen-Jarolim E (2017) Linking iron-deficiency with allergy: role of molecular allergens and the microbiome. Metallomics 9(12):1676–1692. https://doi. org/10.1039/c7mt00241f
- 235. Nwaru BI, Hayes H, Gambling L, Craig LC, Allan K, Prabhu N, Turner SW, McNeill G, Erkkola M, Seaton A, McArdle HJ, Devereux G (2014) An exploratory study of the associations between maternal iron status in pregnancy and childhood wheeze and atopy. Br J Nutr 112(12):2018–2027. https://doi.org/10. 1017/S0007114514003122
- 236. Shaheen SO, Newson RB, Henderson AJ, Emmett PM, Sherriff A, Cooke M, Team AS (2004) Umbilical cord trace elements and minerals and risk of early childhood wheezing and eczema. Eur Respir J 24(2):292–297. https://doi.org/10.1183/09031936.04. 00117803
- 237. Fortes C, Mastroeni S, Mannooranparampil TJ, Di Lallo D (2019) Pre-natal folic acid and iron supplementation and atopic dermatitis in the first 6 years of life. Arch Dermatol Res 311(5):361–367. https://doi.org/10.1007/s00403-019-01911-2
- Gray NA, Dhana A, Stein DJ, Khumalo NP (2019) Zinc and atopic dermatitis: a systematic review and meta-analysis. J Eur Acad Dermatol Venereol 33(6):1042–1050. https://doi.org/10. 1111/jdv.15524
- Maarouf M, Vaughn AR, Shi VY (2018) Topical micronutrients in atopic dermatitis-an evidence-based review. Dermatol Ther 31(5): e12659. https://doi.org/10.1111/dth.12659
- Kadiyala U, Turali-Emre ES, Bahng JH, Kotov NA, VanEpps JS (2018) Unexpected insights into antibacterial activity of zinc oxide nanoparticles against methicillin resistant Staphylococcus aureus (MRSA). Nanoscale 10(10):4927–4939. https://doi.org/10.1039/ c7nr08499d

- 241. Mesias M, Seiquer I, Navarro MP (2012) Is the Mediterranean diet adequate to satisfy zinc requirements during adolescence? Public Health Nutr 15(8):1429–1436. https://doi.org/10.1017/ S1368980011003429
- Castro-Quezada I, Roman-Vinas B, Serra-Majem L (2014) The Mediterranean diet and nutritional adequacy: a review. Nutrients 6(1):231–248. https://doi.org/10.3390/nu6010231
- el-Kholy MS, Gas Allah MA, el-Shimi S, el-Baz F, el-Tayeb H, Abdel-Hamid MS (1990) Zinc and copper status in children with bronchial asthma and atopic dermatitis. J Egypt Public Health Assoc 65(5–6):657–668
- 244. Di Toro R, Galdo Capotorti G, Gialanella G, Miraglia del Giudice M, Moro R, Perrone L (1987) Zinc and copper status of allergic children. Acta Paediatr Scand 76(4):612–617. https://doi.org/10. 1111/j.1651-2227.1987.tb10530.x
- 245. Tinggi U (2008) Selenium: its role as antioxidant in human health. Environ Health Prev Med 13(2):102–108. https://doi.org/10.1007/ s12199-007-0019-4
- Pastore S, Korkina L (2010) Redox imbalance in T cell-mediated skin diseases. Mediat Inflamm 2010:861949. https://doi.org/10. 1155/2010/861949
- 247. Fairris GM, Perkins PJ, Lloyd B, Hinks L, Clayton BE (1989) The effect on atopic dermatitis of supplementation with selenium and vitamin E. Acta Derm Venereol 69(4):359–362
- Ba Z (2016) Selenium in pregnant women: mini review. J Nutr Food Sci 06(03). https://doi.org/10.4172/2155-9600.1000492
- Pieczynska J, Grajeta H (2015) The role of selenium in human conception and pregnancy. J Trace Elem Med Biol 29:31–38. https://doi.org/10.1016/j.jtemb.2014.07.003
- Scientific Opinion on Dietary Reference Values for selenium (2014). EFSA Journal 12 (10). doi:https://doi.org/10.2903/j.efsa. 2014.3846
- 251. Hubalewska-Dydejczyk A, Duntas L, Gilis-Januszewska A (2019) Pregnancy, thyroid, and the potential use of selenium. Hormones (Athens). https://doi.org/10.1007/s42000-019-00144-2
- 252. Yamada T, Saunders T, Kuroda S, Sera K, Nakamura T, Takatsuji T, Fukuoka College of O, Gynecologists PAoFD, Hara T, Nose Y (2013) Cohort study for prevention of atopic dermatitis using hair mineral contents. J Trace Elem Med Biol 27(2):126–131. https://doi.org/10.1016/j.jtemb.2012.08.003
- 253. Pilmane M, Salma-Ancane K, Loca D, Locs J, Berzina-Cimdina L (2017) Strontium and strontium ranelate: historical review of some of their functions. Mater Sci Eng C Mater Biol Appl 78:1222– 1230. https://doi.org/10.1016/j.msec.2017.05.042
- 254. Barneo-Caragol C, Martinez-Morillo E, Rodriguez-Gonzalez S, Lequerica-Fernandez P, Vega-Naredo I, Alvarez Menendez FV (2018) Strontium and oxidative stress in normal pregnancy. J Trace Elem Med Biol 45:57–63. https://doi.org/10.1016/j.jtemb. 2017.09.021
- 255. Venter C, Higgins B, Grundy J, Clayton CB, Gant C, Dean T (2006) Reliability and validity of a maternal food frequency questionnaire designed to estimate consumption of common food allergens. J Hum Nutr Diet 19(2):129–138. https://doi.org/10.1111/ j.1365-277X.2006.00677.x
- Venter C, Brown KR, Maslin K, Palmer DJ (2017) Maternal dietary intake in pregnancy and lactation and allergic disease outcomes in offspring. Pediatr Allergy Immunol 28(2):135–143. https://doi.org/10.1111/pai.12682

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.