Marijuana: Respiratory Tract Effects

Kelly P. Owen • Mark E. Sutter • Timothy E. Albertson

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Abstract Marijuana is the most commonly used drug of abuse in the USA. It is commonly abused through inhalation and therefore has effects on the lung that are similar to tobacco smoke, including increased cough, sputum production, hyperinflation, and upper lobe emphysematous changes. However, at this time, it does not appear that marijuana smoke contributes to the development of chronic obstructive pulmonary disease. Marijuana can have multiple physiologic effects such as tachycardia, peripheral vasodilatation, behavioral and emotional changes, and possible prolonged cognitive impairment. The carcinogenic effects of marijuana are unclear at this time. Studies are mixed on the ability of marijuana smoke to increase the risk for head and neck squamous cell carcinoma, lung cancer, prostate cancer, and cervical cancer. Some studies show that marijuana is protective for development of malignancy. Marijuana smoke has been shown to have an inhibitory effect on the immune system. Components of cannabis are under investigation as treatment for autoimmune diseases and malignancy. As marijuana becomes legalized in many states for medical and recreational use, other forms of tetrahydrocannabinol (THC) have been developed, such as food products and beverages. As most research on marijuana at this time has been on whole marijuana smoke, rather than

K. P. Owen (⊠) • M. E. Sutter
Department of Emergency Medicine, University of California
Davis Medical Center, 4150 V St., Suite 2100,
Sacramento, CA 95817, USA
e-mail: kelly.owen@ucdmc.ucdavis.edu

T. E. Albertson

Department of Internal Medicine, University of California Davis Medical Center, 4150 V St., Suite 3100, Sacramento, CA 95817, USA

T. E. Albertson

Department of Medicine, Veterans Administration Northern California Health Care System, 10535 Hospital Way, Mather, CA 95655, USA THC, it is difficult to determine if the currently available data is applicable to these newer products.

Keywords Marijuana · Chronic obstructive pulmonary disease · Malignancy · Chronic bronchitis · Immune system

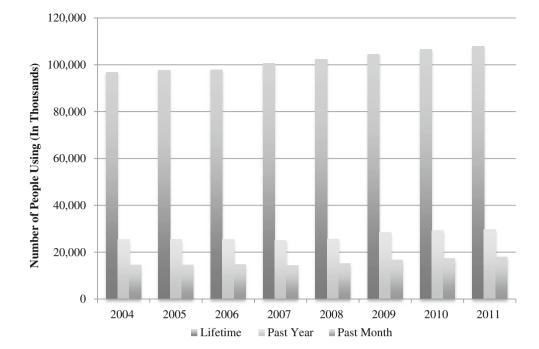
Introduction

In the USA, marijuana was used medicinally until the passage of the Marijuana Tax Act in 1937, which made use illegal. Under the controlled substance act of 1970, marijuana was classified as a Schedule I drug. Schedule I drugs are defined as drugs with no currently accepted medical use and a high potential for abuse [1]. Since then, 18 states have legalized marijuana for different degrees of medical use, and two states have legalized marijuana for personal consumption. To date, federal law has not changed its stance on marijuana, but federal prosecution of users has been limited.

Data from the 2011 National Survey on Drug Use and Health show that marijuana is the most commonly used drug of abuse [2]. Annual use of marijuana has increased over the past decade to 7.0 % of the population or 18.1 million people age 12 or older. In 2011, 2.6 million people used marijuana for the first time, which averages to 7,200 new users per day. On average, about five million people are using marijuana on a daily or nearly daily basis. Despite the recent increase in use, the number of current or new users is still lower than it was at the peak of use in the 1970s (Fig. 1).

Marijuana is derived from the *Cannabis sativa* plant. The stalks, leaves, flowers, and seeds of the plant contain cannabinoids, which are the active ingredients. The drug is usually prepared by drying the flowering top and leaves of the plant. Over 400 compounds have been found in marijuana including 60 different cannabinoids [3]. The cannabinoid 1-delta-9-tetrahydrocannabinol (THC) was identified

Fig. 1 Number of people (in thousands) using marijuana in their lifetime, past year, and past month for the years 2004–2011. Graph created with data from the National Survey on Drug Use and Health. Marijuana use declined from 1979 to 1990, before leveling off. However, current use is increasing over recent years (see [1])



in 1964 [4], and is thought to be responsible for most of the psychoactive effects of the drug. Other cannabinoids have been identified such as cannabinol (CBN), cannabidiol (CBD), and delta-8-THC. CBD does not have psychoactive effects, though has been shown to have anti-epileptic, anxiolytic, and antiemetic actions as well as causing drug-induced prolongation of sleep [5]. The average content of dried marijuana leaves confiscated from the USA between 1993 and 2008 was 4.5 % THC, 0.4 % CBD, and 0.3 % CBN [6]. The cannabinoid content in hashish (a more concentrated form) was 14.1 % THC, 2.4 % CBD, and 1.9 % CBN [6] (Fig. 2).

The composition of marijuana smoke and tobacco smoke are complex, but thought to be similar, with the exception of the active ingredients (cannabinoids and nicotine, respectively). Therefore, the respiratory effects of marijuana smoke are, in general, similar to tobacco, including chronic bronchitis, chronic obstructive pulmonary disease (COPD), and emphysema, as well as development of pulmonary, laryngeal, and head and neck cancer [3, 7]. However, studies proving these associations are mixed and have significant limitations.

Research on the potential health effects of marijuana is ongoing. Despite the research, their remains debate about potential health consequences. This is due to a multitude of factors. First, though research can be performed using purified THC, marijuana smoke contains multiple different compounds. Second, studies of marijuana users often are confounded by concurrent use of tobacco or other drugs. Finally, given that marijuana is an illicit substance, use may be underreported. The health effects of marijuana are summarized in Table 1.

Pharmacology

In the past, the main exposure to cannabinoids was through smoking of marijuana joints. As medicinal use of marijuana has increased, multiple different products containing THC have been developed, including oils, baked products, and beverages. Ways of smoking marijuana have also changed with the advent of vaporizers, which allow the combustion and release of THC with reduced combustion-related byproducts of traditional marijuana smoke. The average THC content of a joint in the 1960–1970s contained about 10 mg of THC. Current marijuana rolled cigarettes or joints may contain 150 mg of THC, or up to 300 mg if hashish oil is added [3]. The THC content in edibles and oils is not standardized and has not been well studied.

Marijuana Smoke Composition

Over 6,000 different chemical components have been found in tobacco smoke, with several thousand-fold more trace components [8]. Though marijuana smoke has not been as well characterized, studies have found that the pyrolization of both marijuana and tobacco results in smoke with similar components [9–24]. Both tobacco and marijuana smoke contain vinyl chlorides, phenol, nitrosamines, reactive oxygen species, and various polycyclic aromatic hydrocarbons (PAH), which are known carcinogens and co-carcinogens [25]. Furthermore, marijuana tar has been found to have higher concentrations of benzo(a)pyrene, a procarcinogenic PAH, than tobacco tar [25].

When evaluating the concentrations of individual chemicals in marijuana smoke and tobacco smoke, the data is conflicting.

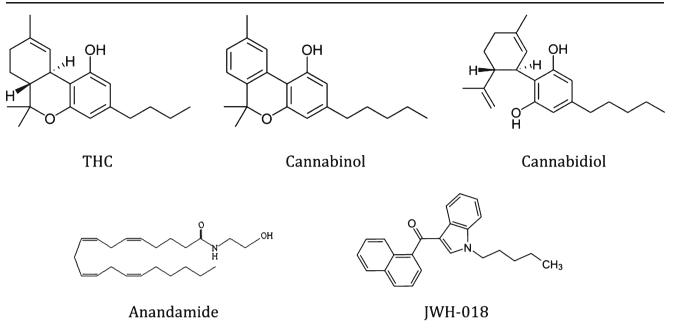


Fig. 2 Molecular structure of natural cannabinoids delta-9-tetrahydracannabinol (THC), cannabinol, cannabidiol, endogenous cannabinoid anandamide, and synthetic cannabinoid JWH-018

Huber et al. [8] compiled data from multiple studies and determined that, in general, marijuana smoke tended to contain higher amounts of ammonia, hydrogen cyanide, and some PAH. In this compilation, tobacco smoke was found to contain higher amounts of carbon monoxide. A recent study by Moir et al. [26] found that marijuana smoke contained three to five times higher levels of nitrogen, both as nitrate and ammoniacal nitrogen, hydrogen cyanide, and aromatic amines. Ammonia was found in marijuana smoke at levels 20 times higher than tobacco smoke. PAH, formaldehyde, carbon monoxide, and acetaldehyde were found at higher levels in tobacco smoke than marijuana amoke. A summary of the components of marijuana and tobacco smoke are found in Table 2.

The way a marijuana cigarette is smoked can also change what is delivered to the lungs. Smokers of marijuana have been found to inhale deeply and hold the smoke in their lungs [27, 28]. The holding of smoke in the lungs has been shown to result in higher delivery of tar, carbon monoxide, and THC to the lungs [29]. Relative to tobacco smokers, smokers of marijuana may inhale more than three times as much tar, and more than one third more tar may be deposited in the respiratory tract [28]. These findings, along with histopathologic changes, have led to the hypothesis that marijuana smoke is carcinogenic [30].

Given the potential carcinogenicity of marijuana smoke from combustion, alternate mechanisms of delivery have been developed. Vaporizers are machines that heat cannabis to a temperature where active cannabis vapors form but below the temperature at which smoke and other carcinogens form [31]. This delivery system has been found to have similar delivery of THC as smoking, with reduced levels of carbon monoxide [32]. Analysis of vapor smoke demonstrated higher levels of THC than smoking, with lower levels of by-products than joint smoke. However, in this

Table 1 Potential health consequences of marijuana smoking

Organ system	Manifestation	
Cell metabolism	Decreased cell proliferation, protein synthesis	
Genetic	Abnormal mitoses, DNA synthesis alterations	
Immunologic	Impairs the following: cell-mediated immunity, macrophage activation, accessory T cell, cytotoxic T cell, and NK cell function. Modifies cytokine release	
Neurologic	Acute: behavioral and emotional changes, impaired psychomotor performance, psychotic reaction	
	Chronic: possible prolonged cognitive impairment	
Cardiovascular	Peripheral vasodilatation, tachycardia, increased oxygen consumption	
Reproductive	Postural hypotension, syncope	
	Gynecomastia, decreased sperm count, impaired ovulation	

Table 2 (Components	of marijuana	and tobacc	o smoke
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Component	Huber et al.		Moir et al.			
			ISO		Extreme	
	Marijuana	Tobacco	Marijuana	Tobacco	Marijuana	Tobacco
Acetaldehyde	1,200	980	448±44*	872±101	1,021±99*	1,555±222
Acetone	443	578	237±23*	454±44	514±32*	826±93
Acrolein	92	85	54.3±4.5*	125±13	148±13*	251±32
Ammonia	228	198	720±84*	35.5 ± 2.4	$1,315\pm106$	$67 {\pm} 9.9$
Benz(a)anthracene	75	43	26.2±3.4*	$30.5 {\pm} 2.5$	43.1±7.9*	52 ± 5.8
Benz(a)pyrene	31	22	8.67±1.12*	14.3 ± 1.2	15.5±2.9*	25.1±2.5
Benzene	76	67	58.3 ± 5.9	62.2±3.5	84.4±8.9*	94.6±2.6
Carbon Monoxide (ppm/cig)	2,600	4,100				
Carbon monoxide (mg/cig)			13.4±1.6*	20.8 ± 1.9	35.3±2.9*	41.5±4.0
Hydrogen cyanide	532	498	525±46*	208 ± 24	1,668±159*	320±29
Isoprene	83	310	74±6.5*	286±15	132±19*	540 ± 18
<i>m</i> - and <i>p</i> -Cresol	54.4	65	$57.8 {\pm} 6.7$	55.4 ± 3.8	157±12*	114 ± 7
Naphthalene	3,000	1,200	2,070±290*	$2,907 \pm 159$	$4,459 \pm 646$	4,908±456
o-Cresol	17.9	24	17.6 ± 1.5	25.5±1.9	46.8±3.9*	51.5±3.3
Phenol	76.8	39	91.5±10.5*	137±11	265±20	283 ± 20

Tobacco and marijuana smoke have many similar components. The ISO smoking group represents a puff volume of 35 mL, a puff duration of 2 s, and a puff interval of 60 s. The extreme conditions are thought to more accurately represent the way that marijuana is smoked and used a puff volume of 70 mL, a puff duration of 2 s, and a puff interval of 30 s. All concentrations are in micrograms unless otherwise noted. Table comprised from data from [8] and [26]

*Denotes statistically significant difference between marijuana and tobacco smoke for Moir et al. [26]

study, the by-products were not analyzed for content [33]. A small non-randomized trial of 12 frequent marijuana smokers who switched to vaporizers showed a significant reduction in respiratory symptoms and improvement in forced vital capacity (FVC) [34].

Absorption and Metabolism

The rate of absorption of cannabis is dependent on both the potency of the specific cannabis product and the mechanism of consumption (IV, inhalational, or oral). About 10–30 % of available THC is absorbed after smoking, as 50 % of THC is lost from smoke escaping into the air or respiratory dead space, and more is lost with combustion [7]. After smoking, THC is rapidly transferred from the lungs to the blood, with peak concentrations occurring between 3 and 10 min [35, 36]. When injected in the IV form, similar times to peak were observed, with higher concentrations than with inhalation [35]. Onset of psychotropic effects is noted within seconds to minutes with a duration of 2–3 h [7]. This is in contrast with oral ingestion, where onset is variable, usually 0.5–2 h, and effects can last 5–6 h [7]. In both cases, THC levels do not correlate with degree of psychotropic effects [35].

Once absorbed, THC follows a biphasic distribution pattern and eventually accumulates in lipophilic tissue. Initially, THC distributes to highly vascularized organs such as the liver, kidneys, heart, and muscle tissue. The distribution half-life is less than 10 min following smoking or IV injection [36, 37]. Following initial distribution, THC accumulates more slowly in body fat and less vascularized tissue.

THC is metabolized by hepatic microsomal hydroxylation and oxidation by CYP 2C9 and 3A4 in the liver [38]. The cytochrome 2C9 is responsible for the oxidation of THC at the C11 position, to its major active metabolite 11-hydroxy- Δ^9 -THC (11-OH-THC) [39, 40]. CYP 3A4 is responsible for oxidation at the 8β -position [39]. Subsequent oxidation of 11-OH-THC leads to the major inactive metabolite 11-nordelta9-carboxy-tetrahydrocannabino-9-carboxylic acid (THC-COOH), which is then eliminated in the urine. The UDPglucuronosyltransferase enzymes also play a role in the metabolism of THC and its metabolites [41]. Fifteen to twenty percent of a dose of THC is eliminated as urinary metabolites, and 65 % is eliminated by fecal excretion as 11-OH-THC and THC-COOH [42]. Data from in vitro studies suggests that THC can inhibit 2C9, though to a lesser degree than CBD [43]. It is unclear if this effect is clinically relevant.

CBD is metabolized through the CYP 2C19 pathway in the liver. It has been found to be a potent inhibitor of CYP1A1, CYP2B6, CYP2C9, CYP2C19, CYP2D6, CYP3A4, and CYP3A5 [44]. The degree of in vivo inhibition is unclear at

this time. As these enzymes are important in the metabolism of many common drugs, the potential for drug/drug interactions is a concern, and further research is needed.

Cannabinoid Receptor Physiology

Cannabinoid receptors have been isolated in central nervous system tissue of multiple species including rat [45], mouse [46, 47], bovine, feline [48], leech [49], puffer fish [50], and newt [51]. The discovery of endogenous cannabinoids (endocannabinoids) has accelerated research on the cannabinoid receptor system. The first endocannabinoid was identified in 1992 and was named "anandamide" after the Sanskrit word for "bliss" [52]. Next, 2-arachidonylglyerol was discovered in 1995 [53, 54]. Finally, in 2001, 2-arachidonyl glycerol ether (noladin ether) was identified in porcine brain [55].

There are two main cannabinoid receptors, CB1 and CB2. Overall, more is known about the CB1 receptor. THC is thought to mediate its psychotropic effects through the CB1 receptor. CB1 was initially cloned from human cells in 1991 [56] and is found in CNS, as well as some peripheral cells. A second receptor was cloned in 1993 from macrophages and spleen, and was termed CB2 [57]. When compared to the CB1 receptor, the CB2 receptor is 68 % homologous in membrane spanning regions and 44 % homologous overall [57]. Both CB1 and CB2 are G-protein coupled, with seven transmembrane spanning regions [58] (Table 3).

The highest density of CB1 receptors in the rodent brain is in the basal ganglia, substantia nigra, globus pallidus, cerebellum, and hippocampus. They are absent in the brainstem [59]. The CB1 receptor inhibits adenylate cyclase activity via Gi/o [60]. Inhibition causes downregulation of cyclic adenosine monophosphate (cAMP), resulting in less accumulation and therefore inhibition of cAMP-dependent protein kinase (PKA). Activation of the CB1 receptor increases mitogen-activated protein (MAP) kinase activity. This mechanism is thought to affect synaptic plasticity, cell migration, and possibly neuronal growth [61]. This receptor modulates calcium conductance through N-and P/Q-type calcium channels and may also increase potassium conductance through an inwardly rectifying potassium channel [62–66]. CB1 expression is particularly dense in presynaptic terminals, and endocannabinoids have been shown to act as retrograde messengers at CNS synapses [60]. CB1 receptors are found on both GABAergic and glutamatergic neurons [61] and are thought to be involved in neurotransmission.

The CB2 receptor was initially thought to only be a peripheral receptor. However, it has now been found throughout the CNS [60], particularly in microglial cells [59]. There is theory that the CB2 receptor is part of a general protective system as part of the immune system [67]. CB2 receptors are found on multiple immune system cells with the amount of mRNA greatest in B cells>natural

killer cells>monocytes>polymorphonuclear leukocytes>T cells [68]. Unlike the CB1 receptor, CB2 receptors do not seem to alter the on inward potassium rectifier channels or N or P/Q type calcium channels [69].

Subsequently, multiple different splice variants of the CB1 and CB2 receptors have been found. Receptor variants have been linked to vulnerability to substance abuse in certain populations [70, 71], though the strength of this association is not definitive [72]. It is hypothesized that the association is due to the role of the cannabinoid receptor on the dopamine and cannabinoid reward systems [71]. Research is ongoing to determine the effects of this genetic variation on response to marijuana. For example, Hopfer et al. found that a common CB1 haplotype was associated with fewer cannabis dependence symptoms in adolescents [73].

Synthetic Cannabinoid Agonists

Multiple compounds that bind to cannabinoid receptors have been developed, such as HU-210, nabilone, and dronabinol [74]. Pfizer developed the cyclohexylphenol (CP) series in the 1970s [75]. In 1994, Huffman et al. created multiple different synthetic cannabinoids known as aminoalkylindoles or JWH compounds [74]. These compounds have been identified in the epidemic of novel illicit drug use involving "spice" compounds.

The FDA has approved three synthetic compounds for medicinal use. Nabilone (Cesamet®) was licensed in 1981 and is a schedule II drug (high potential for abuse). It is a synthetic derivative of THC, which is approved to reduce nausea and vomiting associated with chemotherapy [76]. It has been approved in Canada since 1981, but just began being marketed in the USA in 2006. Though it has better efficacy than older antiemetics, no trials have compared it to new antiemetics such as ondansetron. In 1985, dronabinol (Marinol[®]) was approved as an antiemetic for nausea and vomiting associated with cancer chemotherapy [77]. In 1992, it gained additional marketing approval for appetite stimulation in conditions associated with excessive weight loss such as that occurring in AIDS [78]. The newest drug Sativex[®] is a combination product of THC and CBD. It was approved in 2005 for management of pain in end-stage cancer patients as well as for pain associated with multiple sclerosis patients [79].

Psychotropic/Neurologic Effects of Cannabis

Common psychotropic effects of marijuana intoxication include induction of a relaxed or dreamy state, loss of time perception, anxiety, and psychosis. Acute, residual, and chronic cognitive effects have been reported.

There is some data that shows that alterations in memory and attention can persist even after acute intoxication has

	CB1	CB2
Receptor type	G-protein coupled	G-protein coupled
Receptor action	Gi/o	Gi/o
Location	Central nervous system (basal ganglia, substantia nigra, globus pallidus, cerebellum, and hippocampus)	Multiple immune cells (B cells, NK cells, monocytes, PMN, T cells)
		Spleen and bone marrow
	Expression particularly dense in presynaptic terminals	Microglial cells of CNS
Effect	Modulation of calcium conductance	Modulation of immune function
	Increased potassium conductance through inwardly rectifying potassium channel	
	Psychotropic effects of cannabinoids	

Table 3 Characteristics of cannabinoid receptors

To date, two cannabinoid receptors have been described with multiple splice variants. The CB1 receptor is thought to be the central receptor responsible for the psychotropic effects of cannabinoids, whereas the CB2 receptor is thought to be a more peripheral receptor with immune modulation effects. Research is ongoing to determine downstream cellular effects of both receptors

resolved. Pope et al. studied heavy cannabis users (>5,000) uses during lifetime and found that they had deficits in memory of word use after a 0-, 1-, and 7-day abstinence period, but by day 28, no significant difference could be found when compared to controls [80]. A separate study examined PET scans during the Iowa Gambling task (a decision making task) and found a difference in brain activity only in heavy users (smoking 53–84 joints/week) when compared to moderate users or controls [81].

A meta-analysis by Crean et al. [82] examined the acute (0–6 h), residual (7 h to 20 days), and persistent (>3 weeks) effects of marijuana use. Overall attention and cognition was impaired acutely in light users, but normal in heavy users. Residual effects were mixed, and long-term attention and cognition was largely normal. Decision making and risk taking were impaired in the residual and persistent time period, but acutely, there were mixed findings. Working memory was impaired acutely, but normalized at the residual and long-term time period.

Despite these adverse effects on memory, attention, and cognition, newer data supports a possible neuroprotective mechanism of cannabinoids in brain injury. This effect seems to be dose related, with lower doses causing injury and higher doses being neuroprotective [83]. One example is the finding that THC inhibits formation of amyloid beta fibrils important in the pathogenesis of Alzheimer's disease [84]. Research is ongoing to determine the effects of THC and synthetic cannabinoids on many neurodegenerative diseases such as Alzheimer's disease, Parkinson's disease, and Huntington's disease [85].

Currently, there is conflicting data on the effect of cannabis on mental illness. There is some evidence that the age of initiation of marijuana predicts the time to onset of mental illness [86–88]. Interestingly, CBD, which does not have any of the psychotropic effects of THC, has been proposed as a potential treatment for schizophrenia and psychosis [89, 90].

Physiologic Effects

Acutely, the physiologic effects of marijuana include tachycardia, bronchodilation, conjunctival irritation, reduced lacrimation, and decreased intraocular pressure [91]. Multiple studies have evaluated the short-term effect of marijuana on bronchodilation. Studies have measured specific airway conductance (sGaw) or forced expiratory volume in 1 s (FEV1) in both asthmatics and non-asthmatics. The majority of studies show increased sGaw or FEV1 in both asthmatics and controls. In general, these effects were significant within 15 min and lasted at least 1 h [92–103]. Marijuana was able to reverse methacholine or exercise-induced bronchospasm [94]. Marijuana has been reported to stimulate respiratory drive in animal models; however, in a study of chronic users, no increases in hypoxic or hypercapnic-induced respiratory drive were noted [97].

The cardiovascular effects of marijuana are likely due to CB2 receptors. Postural hypotension with a reflex tachycardia has been well documented [7]. There has been increased interest in the role of cannabinoids as modulators of cardiac risk after a study showed that TCH inhibited atherosclerotic plaque formation via a CB2 receptor-dependent anti-inflammatory mechanism [104]. Research is continuing to elucidate the role of cannabinoids in ischemia and reperfusion injury.

Respiratory Tract Effects

It is generally accepted that the development of obstructive lung disease and respiratory carcinoma is due to the compounds in tobacco smoke. Obstructive lung disease is thought to be due to protein and lipid modifications from compounds in the smoke. Bronchogenic carcinoma is thought to be from the mutagenic/carcinogenic effects from compounds such as nitrosamines and benzy(a)pyrene [91]. Given the similarity of marijuana smoke to tobacco smoke, it would logically seem that the effects on the respiratory system would be similar (Table 2).

Much of the early research on marijuana smoke has focused on early pathologic changes, or effects on cell lines. Attempts to extrapolate this data and form definitive conclusions regarding the development of malignancy or obstructive lung disease have produced conflicting data. Research is complicated as marijuana is an illicit substance in most countries and has only been legal for medical use over the past decade; therefore, obtaining accurate longitudinal data is difficult. It seems reasonable to first discuss the data on inflammatory changes on the respiratory tract, as these are believed to precede development of disease processes and then explore the data on specific diseases.

It is important to note that the majority of studies look at either marijuana smoke or users who smoke marijuana in the form of cigarettes. It is possible that different effects could occur from the use of "water pipes" in which the smoke passes through a water chamber prior to being inhaled, or vaporizers in which the plant material is heated to a temperature where THC is released, but smoke is not generated.

Cellular Changes in the Respiratory Tract

Marijuana and tobacco smoking can lead to cytologic changes in the respiratory tract. Evaluation with bronchoscopy in marijuana smokers, compared to smokers of tobacco, versus tobacco and marijuana, and non-smoking controls showed that 91 % of subjects in the smoking groups (32 people) had visible airway hyperemia [105]. Basal cell and goblet cell hyperplasia was more significant in the marijuana group versus the non-smokers. Cellular disorganization was more prevalent in marijuana versus tobacco smokers. Interestingly, squamous metaplasia was found with significantly higher prevalence in the group with concomitant use of marijuana and tobacco than all other groups, leading to a hypothesis that there are additive effects of marijuana and tobacco smoke.

A second study using bronchoscopy found significantly higher bronchitis index scores in marijuana smokers, tobacco smokers, and smokers of both marijuana and tobacco [106]. When biopsies were evaluated, there was a significant increase in the presence of vascular hyperplasia, submucosal edema, inflammatory cell infiltrates, and goblet cell hyperplasia in smokers versus non-smokers.

It is difficult and impossible to study the entire respiratory tract in humans; therefore, a study was performed in rhesus monkeys [107]. Monkeys were exposed to either high-dose (1 joint/day), low-dose (2 joints/week), placebo (one extracted marijuana cigarette 7 days/week), or sham (sham smoke 7 days/week) for 12 months. A 7-month washout period was used, in which animals were not exposed to any smoke, before evaluation of lung tissue. A higher frequency of alveolitis, alveolar cell hyperplasia, and granulomatous inflammation were found in all smoking groups. The marijuana group had a higher incidence of bronchiolitis, bronchiolar squamous metaplasia, and interstitial fibrosis compared to the other smoking groups. Finally, alveolar cell hyperplasia with atypia was only found in the marijuana group. A higher degree of changes was found in the high-dose marijuana group when compared to the low-dose group, which implies dose-related changes.

Bronchoalveolar lavage (BAL) has also been use to characterize cellular changes in smokers. Total number of cells, as grams per milliliter of BAL fluid, was significantly higher in smokers of tobacco, marijuana, and tobacco plus marijuana when compared to non-smokers [108]. In all groups, macrophages were the predominant cell type found. Elevated neutrophil counts were found in all smoking groups, with the highest number in smokers of tobacco plus marijuana. Macrophages, in tobacco or marijuana smokers, were found to have a higher rate of proliferation when compared to non-smokers, which could lead to the higher cell numbers found in BAL fluid of smokers [109]. At this time, the stimulus for cellular proliferation is unknown. Tobacco smokers, unlike marijuana smokers, have significantly lower percentages of CD4 cells, higher levels of CD8 cells, and lower ratios of CD4:CD8 cells in BAL fluid, when compared to non-smoking controls [110].

Marijuana smoke has been found to have multiple effects in cultured cell lines, including increased oxidative stress, suppression of receptor-mediated apoptosis, increase in necrotic cell death, and disruption of mitochondrial function and cellular energetics [111–113]. In an in vitro model using human small airway epithelial cells, THC was found to have a time- and concentration-dependent decrease in mitochondrial membrane potential, ATP level, and cell viability [114]. How these cellular changes translate into disease states remains to be elucidated.

Lung Disease

Given the similarity between tobacco and marijuana smoke, it would stand to reason that smokers of marijuana would be at risk for the same pulmonary complications as tobacco smokers, such as chronic obstructive pulmonary disease (COPD), chronic bronchitis, emphysema, and asthmatic bronchitis. However, the evidence to show this association is difficult to obtain, as the studies are complicated by a multitude of factors. First, the concomitant use of tobacco is not controlled for in all studies. Second, the dose of marijuana is difficult to define. Some studies use marijuana cigarette years as duration of exposure, with four to five marijuana cigarettes per day defined as being equivalent to 20 cigarettes per day, whereas other studies use number of times exposed to marijuana. Next, studies have generally been performed on younger patients, which may not allow enough time for pulmonary disease to manifest. In many studies, an attempt is made to control for this limitation by comparing results with tobacco smokers within the same cohort. Finally, outcome measures are not always standardized. With these limitations in mind, the data is reviewed below and summarized in Table 4.

Pulmonary Function

There are studies that have found impairment in respiratory function from marijuana use. An epidemiologic study found small but significant decreases in expiratory flow rate and FEV1/forced vial capacity (FVC) ratio in non-tobacco cigarette smokers when compared to tobacco smokers [115]. When the same group extended research longitudinally, slight decreases were seen in FEV1 and FEV1/forced expiratory volume (FEV) ratio in past smokers of marijuana but not current smokers [116]. One of the limitations of these studies is that they did not quantify marijuana use, and instead used non-tobacco cigarette smokers, with the assumption that this represented marijuana use. An additional cross-sectional study found 36 % of marijuana smokers had a FEV1/FVC ratio <80 %, compared to 20 % of non-smokers [117].

When computerized tomography (CT) was used to evaluate pulmonary parenchymal changes, marijuana use was associated with evidence of hyperinflation, with little evidence of emphysema. In this same group, pulmonary function testing showed evidence of airway obstruction as measured by the FEV1/FVC ratio, though less so than in tobacco smokers [118].

The data supporting a lack of effect of marijuana smoking on pulmonary function is more robust. An observational cohort study showed a decline in FEV1 in male tobacco smokers over an 8-year period, but did not show similar decline in marijuana smokers [119]. A dose-related response was not seen, and there was not an additive effect between marijuana and tobacco smoke. A second cohort study found persistent decrease in FEV1/FVC values over time in marijuana smokers when compared to non-smokers; however, when after adjusting for age, tobacco smoking, and weight, there was no statistically significant difference found [120]. Another study using data from the National Health and Nutrition Examination Survey (NHANES III) found a decrease in FEV1/FVC in marijuana smokers (defined as lifetime use more than 100 times and use at least one in the past month), but when controlling for tobacco use, no difference was found [121].

A recent meta-analysis of 34 studies assessed the impact of short-term and long-term marijuana use. They concluded that there is a short-term bronchodilating effect that is evident soon after smoking. They could not find a consistent association between long-term marijuana smoking and FEV1/FVC ratio, DLCO, or airway hyper-reactivity [122]. Since the publication of the meta-analysis, more studies have been performed.

In a cohort study of 878 people over the age of 40, smokers of tobacco or tobacco plus marijuana were more likely to have respiratory symptoms, and evidence of COPD, as defined by spirometric evidence of a ratio of FEV1 to FVC of less than 0.70 after bronchodilation, than non-smokers [123]. Marijuana smokers were defined as having lifetime use of 50 or more marijuana cigarettes. Marijuana smoking alone did not cause an increase in respiratory symptoms and was not an independent risk factor for development of COPD. Use of marijuana and tobacco together created increased risk of respiratory symptoms and COPD than tobacco alone, suggesting an additive effect. In commentary on this study, Tashkin performed a literature review and concluded that smoking marijuana alone can lead to respiratory symptoms, however, that we can be close to concluding that smoking marijuana alone does not lead to COPD [124]. Another large cohort study of 1.037 patients found that marijuana smokers did have a tendency toward hyperinflation and air trapping, though they did not have evidence of airflow obstruction. Marijuana users were found to have higher FVC levels [125].

A cohort study over a 20-year time period using data from the Coronary Artery Risk Development in Young Adults (CARDIA) data found the expected decrease in FEV1 and FVC in tobacco smokers. However, with low levels of marijuana use, defined as 7 joint-years of life (with 1 joint-year equivalent to 365 joints or filled pipe bowels),

 Table 4 Respiratory effects of marijuana smoking

Cellular changes	Airway hyperermia, basal and goblet cell hyperplasia, cellular disorganization, squamous metaplasia
Respiratory symptoms	Cough, wheeze, shortness of breath, sputum production, chest tightness, pharyngitis, hoarse voice, worsening asthma symptoms
Lung function	Hyperinflation, increase/decrease or no change on FEV1, FVC, and FEV. COPD (possible)

there was no adverse effect on pulmonary function. In fact, FEV1 and FVC were actually increased with low levels of exposure. With increasing levels of exposure, FEV1 was not significantly different from baseline, and FVC remained significantly elevated [126]. There are several reports of emphysematous bullous disease in heavy marijuana smokers. The majority of the disease is seen on CT in the upper lobes. Despite CT findings, pulmonary function testing and chest radiographs in these patients tend to be normal [127]. Despite the limitations of the data, a meta-analysis, and the large cohort studies that were published after the meta-analysis, there is evidence for hyperinflation without significant airway obstruction. The clinical relevance of the hyperinflation remains to be determined.

Respiratory Symptoms

Though the evidence for obstructive lung disease is sometimes conflicting, data shows that respiratory symptoms are uniformly increased in marijuana smokers. In the NHANES III study, both marijuana and tobacco smokers had increased likelihood of cough, sputum production, wheezing, chest sounds, and chronic bronchitis [121]. Shortness of breath was increased with tobacco use, but not with marijuana use. Increases in pneumonia were not found with either marijuana or tobacco use. In a longitudinal study, which followed 299 participants, a mean of 9.8 years, current smokers of either tobacco or marijuana were more likely to have symptoms of chronic bronchitis. However, former smokers of either substance were no more likely to have symptoms, showing that cessation of smoking can have clinical benefit [128].

Overall, increases in cough [115, 116, 118, 129], wheeze [115, 116, 118, 129], shortness of breath [115], sputum production [115, 116, 118, 129], chest tightness [117, 118], pharyngitis [130, 131], hoarse voice [131, 132], and worsening asthma symptoms [130, 131] are found in marijuana smokers when compared to non-smokers. Interestingly, one study demonstrated an increase in cough with marijuana smokers (OR, 1.3; CI, 1.0–1.6) and tobacco smokers (OR, 1.4; CI, 1.1–1.9). However, in combined smokers, there was not an association with cough (OR, 1.0; CI, 0.7–1.4) [118].

Neoplastic Effects

Cigarette smoke is a known carcinogen, and given the similarity of marijuana and tobacco smoke, especially with known carcinogens, it would be reasonable to hypothesize that marijuana smoke should be carcinogenic. However, the literature is mixed with regards to the ability of marijuana to cause cancer, and there is some data that synthetic cannabinoids as well as THC have the ability to kill cancer cells. When approaching this data, it is important to evaluate both the effects of marijuana smoke and the effects of cannabinoids alone. A summary of the potential malignant effects of marijuana is summarized in Table 5.

Evidence that marijuana smoke can cause cancer started with findings from the Ames Salmonella mutagenesis assay, which is used to determine the ability of compounds to cause cellular mutations. Two studies have demonstrated the ability of marijuana smoke to induce mutations in the Ames assay [9, 24]. When the components of the smoke were further analyzed to determine the causative factors of marijuana smoke, the highest mutagenic potential was found in the basic fraction [133]. The basic fraction of a high-dose marijuana cigarette was found to be sevenfold higher than the base fraction of tobacco smoke. Painting marijuana tar onto mouse skin causes cancerous effects [134]. Though it is unknown how cancerous effects from skin paining on mice correlates to human toxicity, this model is widely accepted as a screening process for the carcinogenicity of chemical compounds including tobacco smoke [135]. In human lung explants, marijuana smoke induces chromosomal and DNA alterations [136]. Furthermore, DNA damage in macrophages has been demonstrated in human marijuana smokers [137]. In contrast to marijuana smoke, THC alone is not carcinogenic in rodent skin tests [138], and is not mutagenic using the Ames test [139].

Carcinogen-DNA adducts are thought to be precursors to the formation of neoplastic cells. In human tissue samples of tobacco smokers, carcinogen-DNA adducts have been found in samples of peripheral lung, urinary bladder, oral mucosa, and placenta [140]. In rhesus monkeys exposed to marijuana smoke for 1 year at either doses of 7 days per week or 2 days per week, there was no increase in carcinogen-DNA adducts found when compared to sham controls [140]. Levels of adducts were higher in animals exposed to ethanol extracted marijuana smoke.

Human epidemiological studies have been mixed with the development of carcinoma. Initial evidence that linked marijuana use with head and neck squamous cell carcinoma (HNSCC) was based on case reports. In a hospital-based case-control study, ever user of marijuana had a 2.6 (CI, 1.1-6.6)-fold increase in HNSCC when compared to bloodbank controls. This association was stronger in people age 55 and younger (OR, 3.1; CI, 1.0-9.7). There was a trend toward increase risk with increasing frequency of use per day and increased years of use [141]. In a North African population, cannabis use had an odds ratio of 2.62 (CI, 1-6.86) for development of nasopharyngeal carcinoma, after attempting to control for tobacco use [142]. However, a potential confounder is that it is common for tobacco to be mixed with marijuana in North Africa. A third study found that human papillomavirus (HPV)-16-positive HNSCC was associated with use of marijuana; however, HPV-16 negative cancer was not [143].

Table 5 Malignancy from marijuana	Ames test: marijuana smoke	Mutagenic
, jan a	Ames test: THC	Non-mutagenic
	Probable increased risk	Non-seminoma germ cell tumors
		Adult-onset glioma
	Possible increased risk	Head and neck squamous cell carcinoma
		Lung cancer
		Prostate cancer
Care tract Care data la		Cervical cancer
See text for details		

In contrast, seven studies have been unable to find an association between marijuana use and HNSCC. Two small population-based studies from England, performed by the same group, failed to find an association between cannabis smoking and oral cancer [144, 145]. Another small study, with only 75 cases, also did not find increased risk, but may have been limited by the sample size [146]. Two larger population-based studies did not find increase risk in patients with oral squamous cell carcinoma [147, 148] or esophageal cancer [148]. Using data from the INHANCE consortium, a pooled analysis of 4,085 cases and 5,125 controls was performed. The data from the other two large studies noted above was included in their analysis. An association between marijuana use and head and neck cancer was not found (OR, 0.85; CI, 0.53-1.35) [149]. Finally, a case-control study from Massachusetts found that 10 to 20 years of marijuana use was associated with a decreased risk of HNSCC (OR, 0.38; CI, 0.22-0.67). They concluded that moderate marijuana use is associated with a decreased risk of HNSCC [150].

The evidence for marijuana use contributing to the development of lung cancer is equally divided. A Tunisian case-control study showed a markedly elevated increase risk for lung cancer in marijuana smokers (OR, 8.2; CI, 1.3-15.5). However, the study was small, 110 cases, the duration and frequency of marijuana and tobacco use were not reported, and as noted above, in this region, tobacco and marijuana are often smoked together. A systematic review in 2006 concluded that there was no data showing an association of marijuana smoking with lung cancer [151]. They did note several limitations to the studies such as lack of adjustment for tobacco smoking, small sample size, selection bias, limited generalizability, and young age, which might give insignificant lag time for the development of malignancy. Since the systematic review, further studies have been performed.

A case–control study of lung cancer patients under 55 showed an increase risk of lung cancer of 8 % (CI, 2–15) for every year of cannabis use [152]. When adjusted for tobacco smoking, this risk only persisted for patients with >10.5 joint/years exposure (RR, 5.7; CI, 1.5–21.6). A second case–control study with 1,212 cases found no increase

risk from marijuana smoking, even in patients with >60 joint-years smoking (OR, 0.62; CI, 0.32–1.2) [148]. A review of all prior studies with repeat statistical analysis performed in 2008 found that marijuana was protective for development of lung carcinoma, tobacco smokers were at increased risk, and that smokers of both marijuana and tobacco were at higher risk of developing cancer than marijuana smokers, but lower risk than tobacco smokers [153].

Marijuana smoking has also been associated with the development of other malignancies. In a large cohort study, 64,855 patients were assessed using self-assessment questioners, and followed a mean of 8.6 years. In non-smokers, ever users of marijuana did not have an increased risk of lung, melanoma, prostate, breast, or cervical cancer. In tobacco smokers, ever using marijuana increased the risk of prostate cancer (RR, 2.1; CI, 1.0–9.5) and trended toward risk of cervical cancer (RR, 1.4; CI, 1.0–2.1) [154]. Three small studies have correlated marijuana use with the development of nonseminoma testicular germ cell tumors [155–157]. A large cohort study of 133,811 patients demonstrated an increased risk for malignant primary adultonset glioma in individuals who smoked marijuana at least once a month (RR, 2.8; CI, 1.3–6.2) [158].

Despite the link between marijuana and development of malignancy, cannabinoids have also been found to have an anti-tumor effect. In a mouse model of lung adenocarcinoma, developed in 1975, oral administration of THC, delta-8-THC, and CBN decreased tumor size and increased survival [159]. Despite this finding many years ago, research on cannabinoids as cancer treatment has only recently been aggressively pursued. Though much of this research has been performed on in vitro models, there is evidence that cannabinoids, including synthetic cannabinoids, could be beneficial in treatment of breast, prostate, and bone cancer [160], as well as glioblastoma multiforme [161].

In summary, marijuana smoke has been shown to cause cellular changes, some of which are concerning as possible precursors to malignant changes. The development of carcinoma has been difficult to definitively prove in human epidemiological studies. There are limitations to these studies, which include underreporting due to marijuana's illegal status, confounders such as concurrent tobacco, ethanol, or other drug use, as well as time, as cancer takes years to develop, and many participants in these studies were young. Therefore, more research is needed on this complex subject before a final determination can be made.

Immune System Effects

The effects of cannabinoids on the immune system are dependent on the specific cannabinoid studied, cell lines used, and dose given. Though CB1 and CB2 receptors are both expressed on immune system, CB2 receptors are expressed in a much higher level. When looking at the distribution of CB2 mRNA, the levels are greatest in B cells>NK»monocytes>PMNs>T8 cells>T4 cells [162]. The exact mechanism through which exogenous and endogenous cannabinoids upregulate and downregulate immune function is a subject of continued investigation. The effect of cannabinoids on immune function has been extensively reviewed in two articles [163, 164]; therefore, an overview of effects on immune cells will be provided below.

Lymphocyte Effects

In humans, the effect of marijuana on T cell proliferation has been shown to have no effect [165, 166] or has demonstrated decrease in sensitivity [167]. A study of patients who ingested marijuana over 6 to 36 months showed a decrease in the number of T and B cell lymphocytes when compared to controls [168]. However, other studies have shown no difference in the number of T and B cell lymphocytes in chronic marijuana smokers [169]. Overall, there is evidence of a biphasic response of both T and B cell lymphocytes to cannabinoids, where low doses seem to simulate cells and high doses tend to decrease numbers of lymphocytes [163]. In a recent randomized, placebo-controlled trial of HIVpositive patients randomly assigned to marijuana cigarette, dronabinol capsule, or placebo, viral load decreased, and there was not an effect on CD4+ or CD8+ cells for both marijuana cigarette and dronabinol capsule groups when compared to placebo [170]. In a rat model, injected CBD decreased total leukocyte numbers [171].

Macrophage Effects

THC has been shown to have multiple effects on macrophage function including phagocytic ingestion, spreading, oxidative burst, protein expression, DNA synthesis, cytolysis, arachidonic acid release, and antigen presentation [164]. In alveolar macrophages isolated from rhesus monkeys chronically exposed to marijuana smoke, morphologic changes and alterations in protein expression were seen, after a 7-month period without exposure [172]. Alveolar macrophages from human smokers of marijuana showed decreased antimicrobial activity, which was thought to be due to decreased production of nitric oxide [173].

Natural Killer Cell Effects

In the presence of THC, natural killer (NK) cells show diminished production of tumor necrosis factor alpha (TNF- α) when challenged with *Candida albicans* [174]. In vitro, natural killer cells have decreased antitumor activity when exposed to THC [174–176]. In a murine model, THC significantly inhibited NK cell activity [177]. CBD did not have effects on NK cells in a rat model [171].

Neutrophil Effects

There is not data on the effect of THC on the polymorphonuclear leukocyte line. Cannabinoid receptors have not been found on the neutrophil surface, and therefore, it is unlikely that THC exhibits direct effect on neutrophils.

Effects on Cytokine Production

Cytokines are important mediators in host response to infection. In macrophages, THC has been shown to cause impaired tumor necrosis factor alpha (TNF- α) release and enhance interleukin 1 (IL-1) secretion [178–181]. In mouse splenocyte cultures exposed to THC, or mice injected with THC, interferon (INF) production is reduced [182]. INF- α/β production was reduced in a mouse model where mice were injected with THC and subsequently infected with the human simplex virus [183]. Finally, IL-2 production is suppressed after exposure to THC. The mechanism of IL-2 reduction appears to be decreased protein synthesis of IL-2 receptor proteins with subsequent downregulation of IL-2 receptors [184].

Other Effects

Barotrauma

Case reports and case series describe young patients with predominately upper lobe bullous disease leading to complications such as pneumothorax [185, 186]. This has been termed marijuana lung [187] or "bong lung" [188]. Smokers of marijuana tend to differ from those of other substances, in that they inhale more deeply and hold the smoke in their lungs, performing a Valsalva maneuver. These techniques may put marijuana smokers at increased risk for bleb rupture, leading to pneumothorax or pneumomediastinum. Pneumorachis (air in the spinal canal) and pneumomediastinum have also been reported after marijuana smoking from a homemade apparatus

[189], and pneumomediastinum has been reported after alternate use of cocaine and marijuana [190].

Aspergillus Infection

Aspergillus fumigatus spores can contaminate marijuana. In one study, precipitins to Aspergillus were detected in 11 of 21 marijuana smokers tested [191]. The development of chronic pulmonary aspergillosis (CPA) has been associated with marijuana use in patients on chemotherapy, with leukemia, renal transplant, and acquired immunodeficiency syndrome (AIDS) [192]. The development of CPA in marijuana smokers is uncommon, and most case reports are in patients with underlying immunodeficiency. One immunocompetent habitual marijuana smoker developed invasive aspergillosis involving the sphenoid sinus [193]. It is unclear if the Aspergillus infection is due to the handling of the marijuana or from actually smoking.

Summary and Conclusions

Cannabinoids have multiple effects on the pulmonary system, immune system, and on cancer cells. Daily marijuana smoking has been shown to increase the risk for pulmonary symptoms such as wheeze, cough, and sputum production, though the risk for development of COPD is questionable. It is difficult to differentiate the effects of individual cannabinoids with the effect of marijuana smoke.

Though laboratory data has demonstrated the mutagenic and carcinogenic properties of marijuana smoke, human epidemiologic studies have been equivocal in finding a link between marijuana and cancer. However, it also took time to determine the causality of tobacco in the development of malignancy. New evidence shows that cannabinoids may have some cancer fighting properties. Further longitudinal studies are needed to continue to examine the effects of cannabinoids on malignancy.

The effect of cannabinoids on the immune system is unclear. Overall, it appears that THC has a depressant effect, which could lead to increased susceptibility to infection. Research is demonstrating that the immunomodulatory properties of cannabinoids may have clinical use in the treatment in autoimmune diseases.

References

- US Department of Justice (2013) http://www.justice.gov/dea/ druginfo/ds.shtml. Accessed 10 Feb 2013
- SAMHSA (2012) http://www.samhsa.gov/data/NSDUH/2k11Results/ NSDUHresults2011.htm. Accessed 10 Feb 2013
- Ashton CH (2001) Pharmacology and effects of cannabis: a brief review. Br J Psychiatry 178:101–106

- Mechoulam R, Gaoni Y (1965) A total synthesis of delta-1tetrahydrocannabinol, the active constituent of hashish. J Am Chem Soc 87:3273–3275
- Mechoulam R, Parker LA, Gallily R (2002) Cannabidiol: an overview of some pharmacological aspects. J Clin Pharmacol 42:11S–19S
- Mehmedic Z, Chandra S, Slade D, Denham H, Foster S, Patel AS et al (2010) Potency trends of Delta9-THC and other cannabinoids in confiscated cannabis preparations from 1993 to 2008. J Forensic Sci 55:1209–1217
- Maykut MO (1985) Health consequences of acute and chronic marihuana use. Prog Neuropsychopharmacol Biol Psychiatry 9:209–238
- Huber GL, First MW, Grubner O (1991) Marijuana and tobacco smoke gas-phase cytotoxins. Pharmacol Biochem Behav 40:629–636
- Busch FW, Seid DA, Wei ET (1979) Mutagenic activity of marihuana smoke condensates. Cancer Lett 6:319–324
- Fentiman AF Jr, Foltz RL, Kinzer GW (1973) Identification of noncannabinoid phenols in marihuana smoke condensate using chemical ionization mass spectrometry. Anal Chem 45:580–583
- Haq MZ, Rose SJ, Deiderich LR, Patel AR (1974) Identification and quantitative measurement of some N-heterocyclics in marijuana smoke condensates. Anal Chem 46:1781–1784
- Kettenes-van den Bosch JJ, Salemink CA (1977) Cannabis. XVI. Constituents of marihuana smoke condensate. J Chromatogr 131:422–424
- Kier LD, Yamasaki E, Ames BN (1974) Detection of mutagenic activity in cigarette smoke condensates. Proc Natl Acad Sci U S A 71:4159–4163
- Lee ML, Novotny M, Bartle KD (1976) Gas chromatography/mass spectrometric and nuclear magnetic resonance spectrometric studies of carcinogenic polynuclear aromatic hydrocarbons in tobacco and marijuana smoke condensates. Anal Chem 48:405–416
- Mann PE, Cohen AB, Finley TN, Ladman AJ (1971) Alveolar macrophages. Structural and functional differences between nonsmokers and smokers of marijuana and tobacco. Lab Invest 25:111–120
- Novotny M, Lee ML, Bartle KD (1976) A possible chemical basis for the higher mutagenicity of marijuana smoke as compared to tobacco smoke. Experientia 32:280–282
- Novotny M, Lee ML, Low CE, Raymond A (1976) Analysis of marijuana samples from different origins by high-resolution gas– liquid chromatography for forensic application. Anal Chem 48:24–29
- Okamoto T, Chan PC, So BT (1972) Effect of tobacco, marijuana and benzo(a)pyrene on aryl hydrocarbon hydroxylase in hamster lung. Life Sci II 11:733–741
- Patel AR, Gori GB (1975) Preparation and monitoring of marijuana smoke condensate samples. Bull Narc 27:47–54
- Rickert WS, Robinson JC, Rogers B (1982) A comparison of tar, carbon monoxide and pH levels in smoke from marihuana and tobacco cigarettes. Can J Public Health 73:386–391
- Stedman RL (1968) The chemical composition of tobacco and tobacco smoke. Chem Rev 68:153–207
- Swain AP, Cooper JE, Stedman RL (1969) Large-scale fractionation of cigarette smoke condensate for chemical and biologic investigations. Cancer Res 29:579–583
- 23. Tomkins BA, Jenkins RA, Griest WH, Reagan RR, Holladay SK (1985) Liquid chromatographic determination of benzo[a]pyrene in total particulate matter of cigarette smoke. J Assoc Off Anal Chem 68:935–940
- Wehner FC, van Rensburg SJ, Thiel PG (1980) Mutagenicity of marijuana and Transkei tobacco smoke condensates in the *Salmonella*/microsome assay. Mutat Res 77:135–142
- Hoffmann DBK, Gori GB, Wynder EL (1975) On the carcinogenicity of marijuana smoke. Recent Adv Phytochem 9:63–81

- 26. Moir D, Rickert WS, Levasseur G, Larose Y, Maertens R, White P et al (2008) A comparison of mainstream and sidestream marijuana and tobacco cigarette smoke produced under two machine smoking conditions. Chem Res Toxicol 21:494–502
- Simmons MS, Tashkin DP (1995) The relationship of tobacco and marijuana smoking characteristics. Life Sci 56:2185–2191
- Wu TC, Tashkin DP, Djahed B, Rose JE (1988) Pulmonary hazards of smoking marijuana as compared with tobacco. N Engl J Med 318:347–351
- Tashkin DP, Gliederer F, Rose J, Chang P, Hui KK, Yu JL et al (1991) Tar, CO and delta 9THC delivery from the 1st and 2nd halves of a marijuana cigarette. Pharmacol Biochem Behav 40:657–661
- Hashibe M, Straif K, Tashkin DP, Morgenstern H, Greenland S, Zhang ZF (2005) Epidemiologic review of marijuana use and cancer risk. Alcohol 35:265–275
- Hazekamp A, Ruhaak R, Zuurman L, van Gerven J, Verpoorte R (2006) Evaluation of a vaporizing device (Volcano) for the pulmonary administration of tetrahydrocannabinol. J Pharm Sci 95:1308– 1317
- Abrams DI, Vizoso HP, Shade SB, Jay C, Kelly ME, Benowitz NL (2007) Vaporization as a smokeless cannabis delivery system: a pilot study. Clin Pharmacol Ther 82:572–578
- Pomahacova B, Van der Kooy F, Verpoorte R (2009) Cannabis smoke condensate III: the cannabinoid content of vaporised *Cannabis sativa*. Inhal Toxicol 21:1108–1112
- Van Dam NT, Earleywine M (2010) Pulmonary function in cannabis users: support for a clinical trial of the vaporizer. Int J Drug Policy 21:511–513
- 35. Hollister LE, Gillespie HK, Ohlsson A, Lindgren JE, Wahlen A, Agurell S (1981) Do plasma concentrations of delta 9tetrahydrocannabinol reflect the degree of intoxication? J Clin Pharmacol 21:171S–177S
- Huestis MA, Henningfield JE, Cone EJ (1992) Blood cannabinoids. I. Absorption of THC and formation of 11-OH-THC and THCCOOH during and after smoking marijuana. J Anal Toxicol 16:276–282
- Ohlsson A, Lindgren JE, Wahlen A, Agurell S, Hollister LE, Gillespie HK (1980) Plasma delta-9 tetrahydrocannabinol concentrations and clinical effects after oral and intravenous administration and smoking. Clin Pharmacol Ther 28:409– 416
- Watanabe K, Yamaori S, Funahashi T, Kimura T, Yamamoto I (2007) Cytochrome P450 enzymes involved in the metabolism of tetrahydrocannabinols and cannabinol by human hepatic microsomes. Life Sci 80:1415–1419
- Bornheim LM, Lasker JM, Raucy JL (1992) Human hepatic microsomal metabolism of delta 1-tetrahydrocannabinol. Drug Metab Dispos 20:241–246
- 40. Watanabe K, Matsunaga T, Yamamoto I, Funae Y, Yoshimura H (1995) Involvement of CYP2C in the metabolism of cannabinoids by human hepatic microsomes from an old woman. Biol Pharm Bull 18:1138–1141
- 41. Mazur A, Lichti CF, Prather PL, Zielinska AK, Bratton SM, Gallus-Zawada A et al (2009) Characterization of human hepatic and extrahepatic UDP-glucuronosyltransferase enzymes involved in the metabolism of classic cannabinoids. Drug Metab Dispos 37:1496–1504
- Hunt CA, Jones RT (1980) Tolerance and disposition of tetrahydrocannabinol in man. J Pharmacol Exp Ther 215:35–44
- Yamaori S, Okamoto Y, Yamamoto I, Watanabe K (2011) Cannabidiol, a major phytocannabinoid, as a potent atypical inhibitor for CYP2D6. Drug Metab Dispos 39:2049–2056
- 44. Jiang R, Yamaori S, Okamoto Y, Yamamoto I, Watanabe K (2013) Cannabidiol is a potent inhibitor of the catalytic activity of cytochrome P450 2C19. Drug Metab Pharmacokinet (in press)

- 45. Matsuda LA, Lolait SJ, Brownstein MJ, Young AC, Bonner TI (1990) Structure of a cannabinoid receptor and functional expression of the cloned cDNA. Nature 346:561–564
- Chakrabarti A, Onaivi ES, Chaudhuri G (1995) Cloning and sequencing of a cDNA encoding the mouse brain-type cannabinoid receptor protein. DNA Seq 5:385–388
- 47. Abood ME, Ditto KE, Noel MA, Showalter VM, Tao Q (1997) Isolation and expression of a mouse CB1 cannabinoid receptor gene. Comparison of binding properties with those of native CB1 receptors in mouse brain and N18TG2 neuroblastoma cells. Biochem Pharmacol 53:207–214
- Gebremedhin D, Lange AR, Campbell WB, Hillard CJ, Harder DR (1999) Cannabinoid CB1 receptor of cat cerebral arterial muscle functions to inhibit L-type Ca2+ channel current. Am J Physiol 276:H2085–2093
- Stefano GB, Salzet B, Salzet M (1997) Identification and characterization of the leech CNS cannabinoid receptor: coupling to nitric oxide release. Brain Res 753:219–224
- Yamaguchi F, Macrae AD, Brenner S (1996) Molecular cloning of two cannabinoid type 1-like receptor genes from the puffer fish *Fugu rubripes*. Genomics 35:603–605
- Soderstrom K, Leid M, Moore FL, Murray TF (2000) Behaviroal, pharmacological, and molecular characterization of an amphibian cannabinoid receptor. J Neurochem 75:413–423
- 52. Devane WA, Hanus L, Breuer A, Pertwee RG, Stevenson LA, Griffin G et al (1992) Isolation and structure of a brain constituent that binds to the cannabinoid receptor. Science 258:1946–1949
- 53. Mechoulam R, Ben-Shabat S, Hanus L, Ligumsky M, Kaminski NE, Schatz AR et al (1995) Identification of an endogenous 2-monoglyceride, present in canine gut, that binds to cannabinoid receptors. Biochem Pharmacol 50:83–90
- 54. Sugiura T, Kondo S, Sukagawa A, Nakane S, Shinoda A, Itoh K et al (1995) 2-Arachidonoylglycerol: a possible endogenous cannabinoid receptor ligand in brain. Biochem Biophys Res Commun 215:89–97
- 55. Hanus L, Abu-Lafi S, Fride E, Breuer A, Vogel Z, Shalev DE et al (2001) 2-Arachidonyl glyceryl ether, an endogenous agonist of the cannabinoid CB1 receptor. Proc Natl Acad Sci U S A 98:3662–3665
- Gerard CM, Mollereau C, Vassart G, Parmentier M (1991) Molecular cloning of a human cannabinoid receptor which is also expressed in testis. Biochem J 279(Pt 1):129–134
- Munro S, Thomas KL, Abu-Shaar M (1993) Molecular characterization of a peripheral receptor for cannabinoids. Nature 365:61–65
- Hirst RA, Lambert DG, Notcutt WG (1998) Pharmacology and potential therapeutic uses of cannabis. Br J Anaesth 81:77–84
- 59. Mechoulam R, Parker LA (2013) The endocannabinoid system and the brain. Annu Rev Psychol 64:21–47
- Onaivi ES (2009) Cannabinoid receptors in brain: pharmacogenetics, neuropharmacology, neurotoxicology, and potential therapeutic applications. Int Rev Neurobiol 88:335–369
- Howlett AC, Barth F, Bonner TI, Cabral G, Casellas P, Devane WA et al (2002) International Union of Pharmacology. XXVII. Classification of cannabinoid receptors. Pharmacol Rev 54:161– 202
- Mackie K, Devane WA, Hille B (1993) Anandamide, an endogenous cannabinoid, inhibits calcium currents as a partial agonist in N18 neuroblastoma cells. Mol Pharmacol 44:498–503
- Mackie K, Hille B (1992) Cannabinoids inhibit N-type calcium channels in neuroblastoma-glioma cells. Proc Natl Acad Sci U S A 89:3825–3829
- 64. Mackie K, Lai Y, Westenbroek R, Mitchell R (1995) Cannabinoids activate an inwardly rectifying potassium conductance and inhibit Q-type calcium currents in AtT20 cells transfected with rat brain cannabinoid receptor. J Neurosci 15:6552–6561

- 65. Caulfield MP, Brown DA (1992) Cannabinoid receptor agonists inhibit Ca current in NG108-15 neuroblastoma cells via a pertussis toxin-sensitive mechanism. Br J Pharmacol 106:231–232
- Childers SR, Deadwyler SA (1996) Role of cyclic AMP in the actions of cannabinoid receptors. Biochem Pharmacol 52:819–827
- Pacher P, Mechoulam R (2011) Is lipid signaling through cannabinoid 2 receptors part of a protective system? Prog Lipid Res 50:193–211
- Klein TW, Newton C, Larsen K, Lu L, Perkins I et al (2003) The cannabinoid system and immune modulation. J Leukoc Biol 74:486–496
- 69. Felder CC, Joyce KE, Briley EM, Mansouri J, Mackie K, Blond O et al (1995) Comparison of the pharmacology and signal transduction of the human cannabinoid CB1 and CB2 receptors. Mol Pharmacol 48:443–450
- Zhang PW, Ishiguro H, Ohtsuki T, Hess J, Carillo F, Walther D et al (2004) Human cannabinoid receptor 1: 5' exons, candidate regulatory regions, polymorphisms, haplotypes and association with polysubstance abuse. Mol Psychiatry 9:916–931
- Comings DE, Muhleman D, Gade R, Johnson P, Verde R, Saucier G et al (1997) Cannabinoid receptor gene (CNR1): association with i.v. drug use. Mol Psychiatry 2:161–168
- Benyamina A, Kebir O, Blecha L, Reynaud M, Krebs MO (2011) CNR1 gene polymorphisms in addictive disorders: a systematic review and a meta-analysis. Addict Biol 16:1–6
- 73. Hopfer CJ, Young SE, Purcell S, Crowley TJ, Stallings MC, Corley RP et al (2006) Cannabis receptor haplotype associated with fewer cannabis dependence symptoms in adolescents. Am J Med Genet B Neuropsychiatr Genet 141B:895–901
- Vardakou I, Pistos C, Spiliopoulou C (2010) Spice drugs as a new trend: mode of action, identification and legislation. Toxicol Lett 197:157–162
- Pertwee RG (2006) Cannabinoid pharmacology: the first 66 years. Br J Pharmacol 147(Suppl 1):S163–171
- Einhorn LH, Nagy C, Furnas B, Williams SD (1981) Nabilone: an effective antiemetic in patients receiving cancer chemotherapy. J Clin Pharmacol 21:648–69S
- Ungerleider JT, Sarna G, Fairbanks LA, Goodnight J, Andrysiak T, Jamison K (1985) THC or compazine for the cancer chemotherapy patient—the UCLA study. Part II: patient drug preference. Am J Clin Oncol 8:142–147
- Struwe M, Kaempfer SH, Geiger CJ, Pavia AT, Plasse TF, Shepard KV et al (1993) Effect of dronabinol on nutritional status in HIV infection. Ann Pharmacother 27:827–831
- 79. Smith PF (2004) GW-1000. GW pharmaceuticals. Curr Opin Investig Drugs 5:748–754
- Pope HG Jr, Gruber AJ, Hudson JI, Huestis MA, Yurgelun-Todd D (2002) Cognitive measures in long-term cannabis users. J Clin Pharmacol 42:41S–47S
- Bolla KI, Eldreth DA, Matochik JA, Cadet JL (2005) Neural substrates of faulty decision-making in abstinent marijuana users. NeuroImage 26:480–492
- 82. Crean RD, Crane NA, Mason BJ (2011) An evidence based review of acute and long-term effects of cannabis use on executive cognitive functions. J Addict Med 5:1–8
- Sarne Y, Asaf F, Fishbein M, Gafni M, Keren O (2011) The dual neuroprotective-neurotoxic profile of cannabinoid drugs. Br J Pharmacol 163:1391–1401
- 84. Eubanks LM, Rogers CJ, Beuscher AE IV, Koob GF, Olson AJ, Dickerson TJ et al (2006) A molecular link between the active component of marijuana and Alzheimer's disease pathology. Mol Pharm 3:773–777
- Saito VM, Rezende RM, Teixeira AL (2012) Cannabinoid modulation of neuroinflammatory disorders. Curr Neuropharmacol 10:159–166

- Stefanis NC, Dragovic M, Power BD, Jablensky A, Castle D, Morgan VA (2013) Age at initiation of cannabis use predicts age at onset of psychosis: the 7- to 8-year trend. Schizophr Bull 39:251– 254
- Galvez-Buccollini JA, Proal AC, Tomaselli V, Trachtenberg M, Coconcea C, Chun J et al (2012) Association between age at onset of psychosis and age at onset of cannabis use in nonaffective psychosis. Schizophr Res 139:157–160
- Dragt S, Nieman DH, Schultze-Lutter F, van der Meer F, Becker H, de Haan L et al (2012) Cannabis use and age at onset of symptoms in subjects at clinical high risk for psychosis. Acta Psychiatr Scand 125:45–53
- Zuardi AW, Crippa JA, Hallak JE, Moreira FA, Guimaraes FS (2006) Cannabidiol, a *Cannabis sativa* constituent, as an antipsychotic drug. Braz J Med Biol Res 39:421–429
- 90. Deiana S (2013) Medical use of cannabis. Cannabidiol: a new light for schizophrenia? Drug Test Anal 5:46–51
- Van Hoozen BE, Cross CE (1997) Marijuana. Respiratory tract effects. Clin Rev Allergy Immunol 15:243–269
- Tashkin DP, Reiss S, Shapiro BJ, Calvarese B, Olsen JL, Lodge JW (1977) Bronchial effects of aerosolized delta 9tetrahydrocannabinol in healthy and asthmatic subjects. Am Rev Respir Dis 115:57–65
- Tashkin DP, Shapiro BJ, Frank IM (1973) Acute pulmonary physiologic effects of smoked marijuana and oral 9tetrahydrocannabinol in healthy young men. N Engl J Med 289:336–341
- Tashkin DP, Shapiro BJ, Lee YE, Harper CE (1975) Effects of smoked marijuana in experimentally induced asthma. Am Rev Respir Dis 112:377–386
- Tashkin DP, Shapiro BJ, Lee YE, Harper CE (1976) Subacute effects of heavy marihuana smoking on pulmonary function in healthy men. N Engl J Med 294:125–129
- Vachon L, FitzGerald MX, Solliday NH, Gould IA, Gaensler EA (1973) Single-dose effects of marihuana smoke. Bronchial dynamics and respiratory-center sensitivity in normal subjects. N Engl J Med 288:985–989
- Wu HD, Wright RS, Sassoon CS, Tashkin DP (1992) Effects of smoked marijuana of varying potency on ventilatory drive and metabolic rate. Am Rev Respir Dis 146:716–721
- Vachon L, Sulkowski A, Rich E (1974) Marihuana effects on learning, attention and time estimation. Psychopharmacologia 39:1–11
- Tashkin DP, Shapiro BJ, Frank IM (1974) Acute effects of smoked marijuana and oral delta9-tetrahydrocannabinol on specific airway conductance in asthmatic subjects. Am Rev Respir Dis 109:420–428
- Bernstein JG, Kuehnle JC, Mendelson JH (1976) Medical implications of marijuana use. Am J Drug Alcohol Abuse 3:347–361
- 101. Laviolette M, Belanger J (1986) Role of prostaglandins in marihuana-induced bronchodilation. Respiration 49:10–15
- 102. Renaud AM, Cormier Y (1986) Acute effects of marihuana smoking on maximal exercise performance. Med Sci Sports Exerc 18:685–689
- 103. Steadward RD, Singh M (1975) The effects of smoking marihuana on physical performance. Med Sci Sports 7:309–311
- 104. Steffens S, Pacher P (2012) Targeting cannabinoid receptor CB(2) in cardiovascular disorders: promises and controversies. Br J Pharmacol 167:313–323
- 105. Gong H Jr, Fligiel S, Tashkin DP, Barbers RG (1987) Tracheobronchial changes in habitual, heavy smokers of marijuana with and without tobacco. Am Rev Respir Dis 136:142–149
- 106. Roth MD, Arora A, Barsky SH, Kleerup EC, Simmons M, Tashkin DP (1998) Airway inflammation in young marijuana and tobacco smokers. Am J Respir Crit Care Med 157:928–937

- 107. Fligiel SE, Beals TF, Tashkin DP, Paule MG, Scallet AC, Ali SF et al (1991) Marijuana exposure and pulmonary alterations in primates. Pharmacol Biochem Behav 40:637–642
- 108. Barbers RG, Gong H Jr, Tashkin DP, Oishi J, Wallace JM (1987) Differential examination of bronchoalveolar lavage cells in tobacco cigarette and marijuana smokers. Am Rev Respir Dis 135:1271–1275
- 109. Barbers RG, Evans MJ, Gong H Jr, Tashkin DP (1991) Enhanced alveolar monocytic phagocyte (macrophage) proliferation in tobacco and marijuana smokers. Am Rev Respir Dis 143:1092– 1095
- 110. Wallace JM, Oishi JS, Barbers RG, Simmons MS, Tashkin DP (1994) Lymphocytic subpopulation profiles in bronchoalveolar lavage fluid and peripheral blood from tobacco and marijuana smokers. Chest 105:847–852
- 111. Sarafian TA, Kouyoumjian S, Khoshaghideh F, Tashkin DP, Roth MD (2003) Delta 9-tetrahydrocannabinol disrupts mitochondrial function and cell energetics. Am J Physiol Lung Cell Mol Physiol 284:L298–306
- 112. Sarafian TA, Magallanes JA, Shau H, Tashkin D, Roth MD (1999) Oxidative stress produced by marijuana smoke. An adverse effect enhanced by cannabinoids. Am J Respir Cell Mol Biol 20:1286–1293
- 113. Sarafian TA, Tashkin DP, Roth MD (2001) Marijuana smoke and delta(9)-tetrahydrocannabinol promote necrotic cell death but inhibit Fas-mediated apoptosis. Toxicol Appl Pharmacol 174:264–272
- 114. Sarafian T, Habib N, Mao JT, Tsu IH, Yamamoto ML, Hsu E et al (2005) Gene expression changes in human small airway epithelial cells exposed to delta9-tetrahydrocannabinol. Toxicol Lett 158:95–107
- 115. Bloom JW, Kaltenborn WT, Paoletti P, Camilli A, Lebowitz MD (1987) Respiratory effects of non-tobacco cigarettes. Br Med J (Clin Res Ed) 295:1516–1518
- 116. Sherrill DL, Krzyzanowski M, Bloom JW, Lebowitz MD (1991) Respiratory effects of non-tobacco cigarettes: a longitudinal study in general population. Int J Epidemiol 20:132–137
- 117. Taylor DR, Poulton R, Moffitt TE, Ramankutty P, Sears MR (2000) The respiratory effects of cannabis dependence in young adults. Addiction 95:1669–1677
- 118. Aldington S, Williams M, Nowitz M, Weatherall M, Pritchard A, McNaughton A et al (2007) Effects of cannabis on pulmonary structure, function and symptoms. Thorax 62:1058–1063
- 119. Tashkin DP, Simmons MS, Sherrill DL, Coulson AH (1997) Heavy habitual marijuana smoking does not cause an accelerated decline in FEV1 with age. Am J Respir Crit Care Med 155:141– 148
- 120. Taylor DR, Fergusson DM, Milne BJ, Horwood LJ, Moffitt TE, Sears MR et al (2002) A longitudinal study of the effects of tobacco and cannabis exposure on lung function in young adults. Addiction 97:1055–1061
- 121. Moore BA, Augustson EM, Moser RP, Budney AJ (2005) Respiratory effects of marijuana and tobacco use in a U.S. sample. J Gen Intern Med 20:33–37
- 122. Tetrault JM, Crothers K, Moore BA, Mehra R, Concato J, Fiellin DA (2007) Effects of marijuana smoking on pulmonary function and respiratory complications: a systematic review. Arch Intern Med 167:221–228
- 123. Tan WC, Lo C, Jong A, Xing L, Fitzgerald MJ, Vollmer MJ et al (2009) Marijuana and chronic obstructive lung disease: a population-based study. CMAJ 180:814–820
- 124. Tashkin DP (2009) Does smoking marijuana increase the risk of chronic obstructive pulmonary disease? CMAJ 180:797–798
- 125. Hancox RJ, Poulton R, Ely M, Welch D, Taylor DR, McLachlan CR et al (2010) Effects of cannabis on lung function: a population-based cohort study. Eur Respir J 35:42–47

- 126. Pletcher MJ, Vittinghoff E, Kalhan R, Richman J, Safford M, Sidney S et al (2012) Association between marijuana exposure and pulmonary function over 20 years. JAMA 307:173–181
- Lee MH, Hancox RJ (2011) Effects of smoking cannabis on lung function. Expert Rev Respir Med 5:537–546 (quiz 547)
- Tashkin DP, Simmons MS, Tseng CH (2012) Impact of changes in regular use of marijuana and/or tobacco on chronic bronchitis. COPD 9:367–374
- 129. Tashkin DP, Coulson AH, Clark VA, Simmons M, Bourque LB, Duann S et al (1987) Respiratory symptoms and lung function in habitual heavy smokers of marijuana alone, smokers of marijuana and tobacco, smokers of tobacco alone, and nonsmokers. Am Rev Respir Dis 135:209–216
- 130. Henderson RL, Tennant FS, Guerry R (1972) Respiratory manifestations of hashish smoking. Arch Otolaryngol 95:248–251
- Chopra GS (1973) Studies on psycho-clinical aspects of longterm marihuana use in 124 cases. Int J Addict 8:1015–1026
- 132. Boulougouris JC, Panayiotopoulos CP, Antypas E, Liakos A, Stefanis C (1976) Effects of chronic hashish use on medical status in 44 users compared with 38 controls. Ann N Y Acad Sci 282:168–172
- Sparacino CM, Hyldburg PA, Hughes TJ (1990) Chemical and biological analysis of marijuana smoke condensate. NIDA Res Monogr 99:121–140
- 134. Cottrell JC, Sohn SS, Vogel WH (1973) Toxic effects of marihuana tar on mouse skin. Arch Environ Health 26:277–278
- 135. Walaszek Z, Hanausek M, Slaga TJ (2007) The role of skin painting in predicting lung cancer. Int J Toxicol 26:345–351
- 136. Leuchtenberger C, Leuchtenberger R, Ritter U, Inui N (1973) Effects of marijuana and tobacco smoke on DNA and chromosomal complement in human lung explants. Nature 242:403–404
- 137. Sherman MP, Aeberhard EE, Wong VZ, Simmons MS, Roth MD, Tashkin DP (1995) Effects of smoking marijuana, tobacco or cocaine alone or in combination on DNA damage in human alveolar macrophages. Life Sci 56:2201–2207
- 138. Chan PC, Sills RC, Braun AG, Haseman JK, Bucher JR (1996) Toxicity and carcinogenicity of delta 9-tetrahydrocannabinol in Fischer rats and B6C3F1 mice. Fundam Appl Toxicol 30:109– 117
- 139. Hall W, MacPhee D (2002) Cannabis use and cancer. Addiction 97:243–247
- 140. Talaska G, Schamer M, Bailey JR, Ali SF, Scallet AC, Slikker W et al (1992) No increase in carcinogen-DNA adducts in the lungs of monkeys exposed chronically to marijuana smoke. Toxicol Lett 63:321–332
- 141. Zhang ZF, Morgenstern H, Spitz MR, Tashkin DP, Yu GP, Marshall JR et al (1999) Marijuana use and increased risk of squamous cell carcinoma of the head and neck. Cancer Epidemiol Biomarkers Prev 8:1071–1078
- 142. Feng BJ, Khyatti M, Ben-Ayoub W, Dahmoul S, Ayad M, Maachi F et al (2009) Cannabis, tobacco and domestic fumes intake are associated with nasopharyngeal carcinoma in North Africa. Br J Cancer 101:1207–1212
- 143. Gillison ML, D'Souza G, Westra W, Sugar E, Xiao W, Begum S et al (2008) Distinct risk factor profiles for human papillomavirus type 16-positive and human papillomavirus type 16-negative head and neck cancers. J Natl Cancer Inst 100:407–420
- 144. Llewellyn CD, Linklater K, Bell J, Johnson NW, Warnakulasuriya S (2004) An analysis of risk factors for oral cancer in young people: a case–control study. Oral Oncol 40:304–313
- 145. Llewellyn CD, Johnson NW, Warnakulasuriya KA (2004) Risk factors for oral cancer in newly diagnosed patients aged 45 years and younger: a case–control study in Southern England. J Oral Pathol Med 33:525–532

- 146. Aldington S, Harwood M, Cox B, Weatherall M, Beckert L, Hansell A et al (2008) Cannabis use and cancer of the head and neck: case–control study. Otolaryngol Head Neck Surg 138:374–380
- 147. Rosenblatt KA, Daling JR, Chen C, Sherman KJ, Schwartz SM (2004) Marijuana use and risk of oral squamous cell carcinoma. Cancer Res 64:4049–4054
- 148. Hashibe M, Morgenstern H, Cui Y, Tashkin DP, Zhang ZF, Cozen W et al (2006) Marijuana use and the risk of lung and upper aerodigestive tract cancers: results of a populationbased case–control study. Cancer Epidemiol Biomarkers Prev 15:1829–1834
- 149. Berthiller J, Lee YC, Boffetta P, Wei Q, Sturgis EM, Greenland S et al (2009) Marijuana smoking and the risk of head and neck cancer: pooled analysis in the INHANCE consortium. Cancer Epidemiol Biomarkers Prev 18:1544–1551
- 150. Liang C, McClean MD, Marsit C, Christensen B, Peters E, Nelson HH et al (2009) A population-based case–control study of marijuana use and head and neck squamous cell carcinoma. Cancer Prev Res (Phila) 2:759–768
- 151. Mehra R, Moore BA, Crothers K, Tetrault J, Fiellin DA (2006) The association between marijuana smoking and lung cancer: a systematic review. Arch Intern Med 166:1359–1367
- 152. Aldington S, Harwood M, Cox B, Weatherall M, Beckert L, Hansell A et al (2008) Cannabis use and risk of lung cancer: a case–control study. Eur Respir J 31:280–286
- 153. Chen AL, Chen TJ, Braverman ER, Acuri V, Kemer M, Varshavskiy M et al (2008) Hypothesizing that marijuana smokers are at a significantly lower risk of carcinogenicity relative to tobacco-non-marijuana smokers: evidenced based on statistical reevaluation of current literature. J Psychoactive Drugs 40:263–272
- 154. Sidney S, Quesenberry CP Jr, Friedman GD, Tekawa IS (1997) Marijuana use and cancer incidence (California, United States). Cancer Causes Control 8:722–728
- 155. Daling JR, Doody DR, Sun X, Trabert BL, Weiss NS, Chen C et al (2009) Association of marijuana use and the incidence of testicular germ cell tumors. Cancer 115:1215–1223
- Trabert B, Sigurdson AJ, SweeneyAM SSS, McGlynn KA (2011) Marijuana use and testicular germ cell tumors. Cancer 117:848–853
- 157. Lacson JC, Carroll JD, Tuazon E, Castelao EJ, Bernstein L, Cortessis VK (2012) Population-based case–control study of recreational drug use and testis cancer risk confirms an association between marijuana use and nonseminoma risk. Cancer 118:5374–5383
- 158. Efird JT, Friedman GD, Sidney S, Klatsky A, Habel LA, Udaltsova NV et al (2004) The risk for malignant primary adult-onset glioma in a large, multiethnic, managed-care cohort: cigarette smoking and other lifestyle behaviors. J Neurooncol 68:57–69
- Munson AE, Harris LS, Friedman MA, Dewey WL, Carchman RA (1975) Antineoplastic activity of cannabinoids. J Natl Cancer Inst 55:597–602
- 160. Guindon J, Hohmann AG (2011) The endocannabinoid system and cancer: therapeutic implication. Br J Pharmacol 163:1447–1463
- 161. Torres S, Lorente M, Rodriguez-Fornes F, Hernandez-Tiedra S, Salazar M, Garcia-Taboada E et al (2011) A combined preclinical therapy of cannabinoids and temozolomide against glioma. Mol Cancer Ther 10:90–103
- 162. Galiegue S, Mary S, Marchand J, Dussossoy D, Carriere D, Carayon P et al (1995) Expression of central and peripheral cannabinoid receptors in human immune tissues and leukocyte subpopulations. Eur J Biochem 232:54–61
- 163. Croxford JL, Yamamura T (2005) Cannabinoids and the immune system: potential for the treatment of inflammatory diseases? J Neuroimmunol 166:3–18
- Klein TW, Newton C, Friedman H (1998) Cannabinoid receptors and immunity. Immunol Today 19:373–381

- Lau RJ, Tubergen DG, Barr M Jr, Domino EF, Benowitz N, Jones RT (1976) Phytohemagglutinin-induced lymphocyte transformation in humans receiving delta9-tetrahydrocannabinol. Science 192:805–807
- 166. White SC, Brin SC, Janicki BW (1975) Mitogen-induced blastogenic responses of lymphocytes from marihuana smokers. Science 188:71–72
- 167. Nahas GG, Suciu-Foca N, Armand JP, Morishima A (1974) Inhibition of cellular mediated immunity in marihuana smokers. Science 183:419–420
- 168. El-Gohary M, Eid MA (2004) Effect of cannabinoid ingestion (in the form of bhang) on the immune system of high school and university students. Hum Exp Toxicol 23:149–156
- Rachelefsky GS, Opelz G, Mickey MR, Lessin P, Kiuchi M, Silverstein MJ et al (1976) Intact humoral and cell-mediated immunity in chronic marijuana smoking. J Allergy Clin Immunol 58:483–490
- 170. Abrams DI, Hilton JF, Leiser RJ, Shade SB, Elbeik TA, Aweeka FT et al (2003) Short-term effects of cannabinoids in patients with HIV-1 infection: a randomized, placebo-controlled clinical trial. Ann Intern Med 139:258–266
- 171. Ignatowska-Jankowska B, Jankowski M, Glac W, Swiergel AH (2009) Cannabidiol-induced lymphopenia does not involve NKT and NK cells. J Physiol Pharmacol 60(Suppl 3):99–103
- 172. Cabral GA, Stinnett AL, Bailey J, Ali SF, Paule MG, Scallet AC et al (1991) Chronic marijuana smoke alters alveolar macrophage morphology and protein expression. Pharmacol Biochem Behav 40:643– 649
- 173. Roth MD, Whittaker K, Salehi K, Tashkin DP, Baldwin GC (2004) Mechanisms for impaired effector function in alveolar macrophages from marijuana and cocaine smokers. J Neuroimmunol 147:82–86
- 174. Kusher DI, Dawson LO, Taylor AC, Djeu JY (1994) Effect of the psychoactive metabolite of marijuana, delta 9-tetrahydrocannabinol (THC), on the synthesis of tumor necrosis factor by human large granular lymphocytes. Cell Immunol 154:99–108
- 175. Kawakami Y, Klein TW, Newton C, Djeu JY, Dennert G, Specter S et al (1988) Suppression by cannabinoids of a cloned cell line with natural killer cell activity. Proc Soc Exp Biol Med 187:355– 359
- 176. Specter SC, Klein TW, Newton C, Mondragon M, Widen R, Friedman H (1986) Marijuana effects on immunity: suppression of human natural killer cell activity of delta-9-tetrahydrocannabinol. Int J Immunopharmacol 8:741–745
- 177. Wang M, Richards AL, Friedman H, Djeu JY (1991) Selective inhibition of natural killer but not natural cytotoxic activity in a cloned cell line by delta-9-tetrahydrocannabinol. J Leukoc Biol 50:192–197
- Shivers SC, Newton C, Friedman H, Klein TW (1994) Delta 9-Tetrahydrocannabinol (THC) modulates IL-1 bioactivity in human monocyte/macrophage cell lines. Life Sci 54:1281–1289
- 179. Zhu W, Newton C, Daaka Y, Friedman H, Klein TW (1994) Delta 9-tetrahydrocannabinol enhances the secretion of interleukin 1 from endotoxin-stimulated macrophages. J Pharmacol Exp Ther 270:1334–1339
- Zheng ZM, Specter S, Friedman H (1992) Inhibition by delta-9tetrahydrocannabinol of tumor necrosis factor alpha production by mouse and human macrophages. Int J Immunopharmacol 14:1445–1452
- 181. Fischer-Stenger K, Dove Pettit DA, Cabral GA (1993) Delta 9tetrahydrocannabinol inhibition of tumor necrosis factor-alpha: suppression of post-translational events. J Pharmacol Exp Ther 267:1558–1565
- 182. Blanchard DK, Newton C, Klein TW, Stewart WE 2nd, Friedman H (1986) In vitro and in vivo suppressive effects of delta-9tetrahydrocannabinol on interferon production by murine spleen cells. Int J Immunopharmacol 8:819–824
- 183. Cabral GA, Lockmuller JC, Mishkin EM (1986) Delta 9tetrahydrocannabinol decreases alpha/beta interferon response to

herpes simplex virus type 2 in the B6C3F1 mouse. Proc Soc Exp Biol Med 181:305–311

- 184. Zhu W, Igarashi T, Friedman H, Klein TW (1995) Delta 9tetrahydrocannabinol (THC) causes the variable expression of IL2 receptor subunits. J Pharmacol Exp Ther 274:1001–1007
- 185. Beshay M, Kaiser H, Niedhart D, Reymond MA, Schmid RA (2007) Emphysema and secondary pneumothorax in young adults smoking cannabis. Eur J Cardiothorac Surg 32:834–838
- Hii SW, Tam JD, Thompson BR, Naughton MT (2008) Bullous lung disease due to marijuana. Respirology 13:122–127
- 187. Hii S, Naughton MT, Young A (2006) Marijuana lung. Intern Med J 36:270–271
- 188. Gao Z, Wood-Baker R, Harle R, Muller K, Hauser J, Reid DW (2010) "Bong lung" in cystic fibrosis: a case report. J Med Case Rep 4:371

- 189. Hazouard E, Koninck JC, Attucci S, Fauchier-Rolland F, Brunereau L, Diot P (2001) Pneumorachis and pneumomediastinum caused by repeated Muller's maneuvers: complications of marijuana smoking. Ann Emerg Med 38:694–697
- 190. Luque MA 3rd, Cavallaro DL, Torres M, Emmanual P, Hillman JV (1987) Pneumomediastinum, pneumothorax, and subcutaneous emphysema after alternate cocaine inhalation and marijuana smoking. Pediatr Emerg Care 3:107–109
- 191. Kagen SL (1981) Aspergillus: an inhalable contaminant of marihuana. N Engl J Med 304:483–484
- 192. Gargani Y, Bishop P, Denning DW (2011) Too many mouldy joints—marijuana and chronic pulmonary aspergillosis. Mediterr J Hematol Infect Dis 3:e2011005
- 193. Schwartz IS (1985) Marijuana and fungal infection. Am J Clin Pathol 84:256