

## Novel antioxidants in anthracycline cardiotoxicity

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Published online: 3 May 2007  
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**Abstract** It has been suggested nitroxides and their amine precursors prevent incidence of damage caused by superoxide and hydroxyl radicals formed during the oxidative metabolism of doxorubicin (DOX) and daunorubicin (DAU). Both doxorubicin and daunorubicin have been associated with cardiac toxicity in both adults and children. The authors herein suggest that cardioprotective molecules modified by nitroxides and their secondary amine precursors can prevent or diminish the anthracycline-induced cardiomyopathy by accumulating in cardiomyocytes.

**Keywords** Anthracyclines · Antioxidants · Cardiomyopathy · EPR · Free radicals · Nitroxides

### Introduction

Anthracyclines are widely used for treating a range of malignancies such as sarcomas, lymphomas, breast cancer, myeloma, small-cell lung cancer, bladder cancer and paediatric solid tumors. However, drugs such as DOX and DAU also cause oxygen-dependent DNA damage. Anthracyclines may also cause protein-associated breaks. These breaks are induced by the reaction of anthracyclines with topoisomerase II, which promotes DNA strand cleavage and reannealing [1].

The DNA damage could also be attributed to electron transfer processes. A one-electron – reduction of anthracyclines, catalyzed by flavoenzymes produces the anthracycline semiquinone radical, which can be oxidized to

anthracycline with a reduction of oxygen to superoxide anion radical ( $O_2^-$ ). Both superoxide and semiquinone can generate hydroxyl radicals involved in a variety of deleterious biological processes, including DNA breaks (Scheme 1, Eqs. 1–3) [2]. The third possibility for DNA damage is the formation of a doxorubicin-ferric complex, which binds to DNA by a different mechanism than the intercalation of anthracyclines. This ferric complex is reduced by superoxide to yield a ferrous complex. The latter catalyses hydroxyl radical formation in a Fenton reaction near the DNA strands, cleaving them (Scheme 1, Eqs. 4&5) [3]. This latter process contribution has been confirmed by the fact that the addition of Dexrazoxane (an iron-chelator) prevents cardiac toxicity in humans [4, 5]. Beyond this, several other attempts has been made to reduce the cardiotoxicity of anthracyclines including special modes of administration, development of new formulations, liposome encapsulation [6], less cardiotoxic analogues or co-administration of antioxidant molecules such as flavones [7]. The results and ideas we present here are related to the latter concept. Nitroxides, and their amino precursors, can participate in one-electron processes and are capable of acting as multifunctional antioxidants.

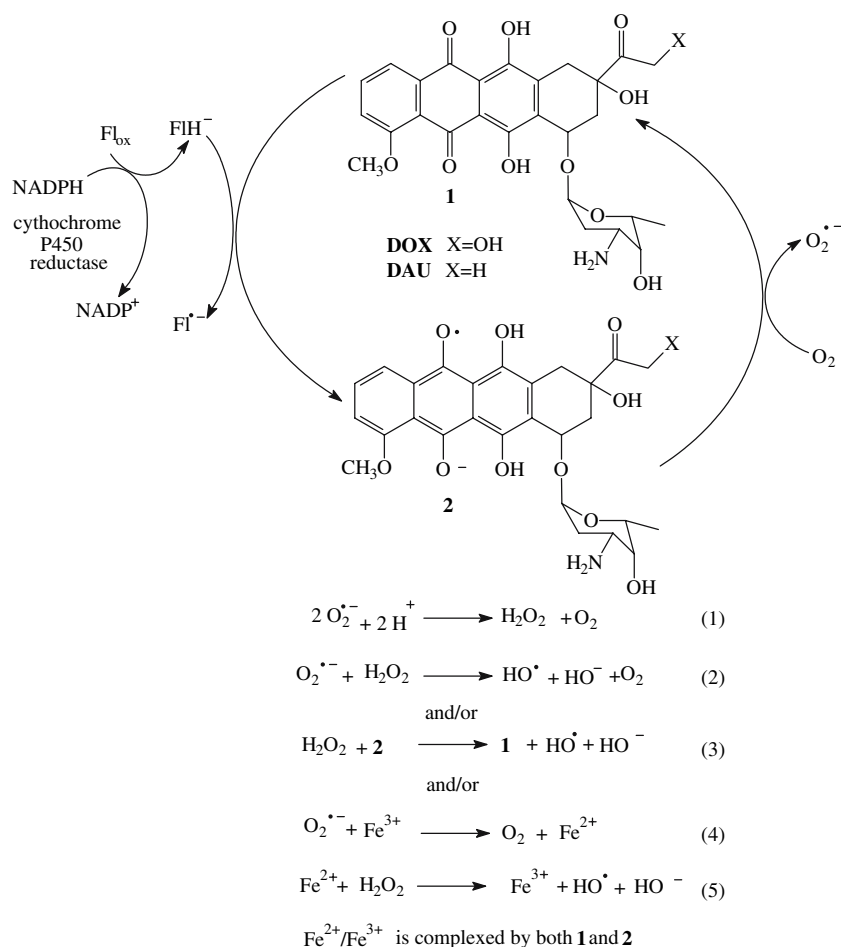
### Results and discussion

It is well documented that TEMPOL (4-hydroxy-2,2,6,6-tetramethyl-piperidine), at 2.5 mM concentration, inhibits the DOX-induced membrane lipid peroxidation by free radical trapping in an isolated rat heart model perfused with 100 µg/ml DOX and by stabilizing the oxidized form of iron blocking the Fenton-reaction [8]. It has also been reported that TEMPOL has no effect on DOX induced cytotoxicity on Chinese hamster V79 cells [9]. These

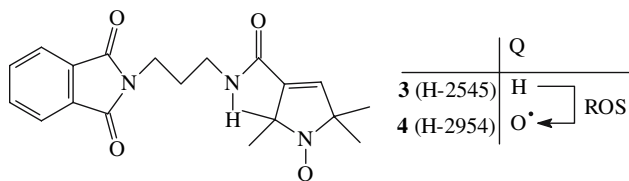
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**Scheme 1** Possible mechanism of ROS production during oxygen dependent metabolism of anthracycline antibiotics

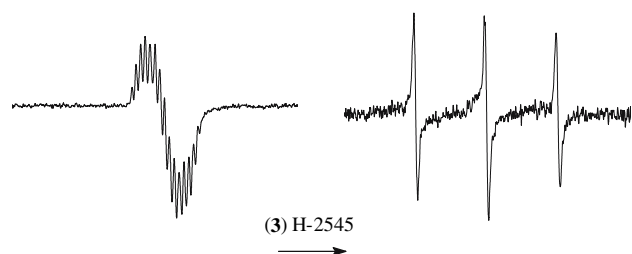


observations prompted us to test our experimental drug H-2545 and its metabolite H-2954 (Scheme 2: compounds 3 and 4) in order to determine whether they can prevent the acute deterioration of cardiac function following DOX administration [10]. It was reported that compound 3 (Fig. 1) could scavenge free radicals with the formation of compound 4 [11–13]. The hypothesis of preventing the DOX-induced cardiotoxicity with antioxidants 3 and 4 was proved with Langendorff-perfused rat heart experiments studying high-energy phosphate levels, contractile function, lipid peroxidation, protein oxidation and Akt phosphorylation [14].



**Scheme 2** H-2545 and its oxidative metabolite H-2954

From these experiments we determined that compounds H-2545 and H-2954 in 10 μM and 20 μM concentrations, had greater recovery of adenosine triphosphate (ATP) and creatine phosphate (PCr) levels when compared to DOX-only treated hearts, (Table 1) following ischemia/reperfusion (1 h). Protein oxidation and lipid peroxidation induced by doxorubicin administration, was inhibited by the experimental antioxidant drugs (3, 4). For instance, compound 3 in 20 μM concentration reduced the lipid peroxi-



**Fig. 1** EPR spectra of DOX-generated ROS trapping by compound 3. The resultant EPR signal is characteristic of compound 4

**Table 1** Effect of H-2545 (**3**) and H-2954 (**4**) on creatine phosphate and ATP level

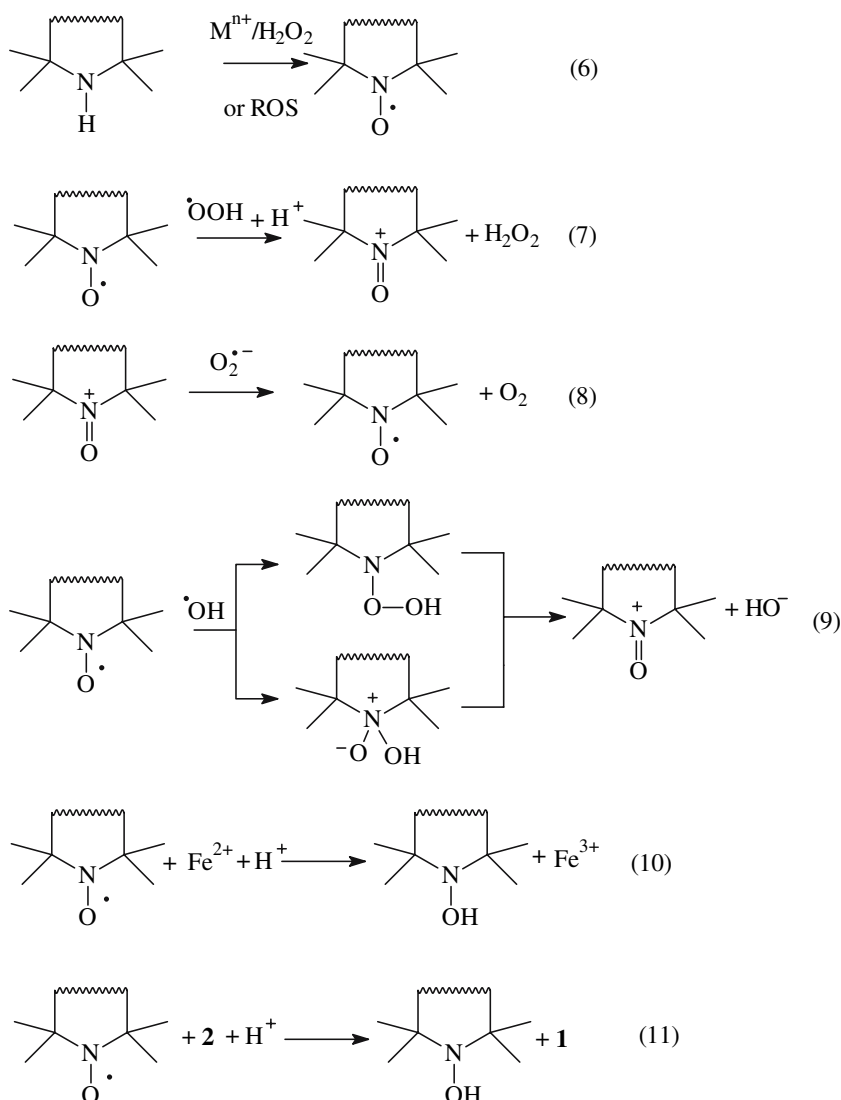
	Control	Dox	<b>3</b> (10 $\mu$ M)	<b>3</b> (20 $\mu$ M)	<b>4</b> (10 $\mu$ M)	<b>4</b> (20 $\mu$ M)
PCr (% recovery)	81 $\pm$ 9	20 $\pm$ 4	48 $\pm$ 7	62 $\pm$ 7	40 $\pm$ 6	61 $\pm$ 10
ATP(% recovery)	85 $\pm$ 4	15 $\pm$ 10	47 $\pm$ 8	61 $\pm$ 7	41 $\pm$ 9	60 $\pm$ 8

**Table 2** Effect of H-2545 (**3**) and H-2954 (**4**) on lipid peroxidation and protein oxidation

	Control	Dox	<b>3</b> (10 $\mu$ M)	<b>3</b> (20 $\mu$ M)	<b>4</b> (10 $\mu$ M)	<b>4</b> (20 $\mu$ M)
Lipid peroxidation (nM/gwt)	40 $\pm$ 1	110 $\pm$ 5	80 $\pm$ 5	50 $\pm$ 5	81 $\pm$ 5	58 $\pm$ 9
Protein oxidation (nM/gwt)	1350 $\pm$ 100	2100 $\pm$ 150	1650 $\pm$ 70	1500 $\pm$ 100	1620 $\pm$ 100	1550 $\pm$ 120

dation value almost to the control level (Table 2). The concentration dependence of antioxidant activity suggests that compounds **3** and **4** participate not only in catalytic, but in stoichiometric radical scavenging processes as well. These compounds reduced the Akt phosphorylation by

preventing activation of the Akt kinase cascade, and improved the contractile function when compared to DOX-only perfused hearts. The co-administration of antioxidants **3** and **4** with DOX did not alter the anticancer properties verified on malignant cell cultures (HeLa, PANC-1, HEPG-

**Scheme 3** Possible mechanism of detoxification of ROS formed during DOX metabolism

2). Although free radicals participate in the DOX anticancer effect, DOX can intercalate between DNA strands and inhibit topoisomerase II, blocking DNA and RNA synthesis. Presumably, this mechanism is not involved in cardiotoxicity because in some cases heart failure developed after doxorubicin administration [15].

The nitroxides and their sterically hindered secondary amine precursors can be regarded as multifunctional non-enzymatic antioxidants. The sterically hindered amine reacts with hydrogen peroxide ( $H_2O_2$ ) in the presence of transition metal ions to yield non-toxic nitroxides. However, other reactive oxygen species (ROS) (for example  $^1O_2$ ) also oxidize them to nitroxide (Eq.6) (Scheme 3) [16]. This process was proven in an in vitro EPR experiment by adding compound 3 to a solution of compound 1 (DOX) in a buffer (pH = 8), resulting in a loss of DOX signal (one band with a fine structure) and the appearance of an isotropic triplet signal, characteristic of nitroxide free radicals such as compound 4. This triplet appears within 5 min, and it remains consistent over a period of 24 h. (Scheme 1)(Belagyi, and Hideg, unpublished results).

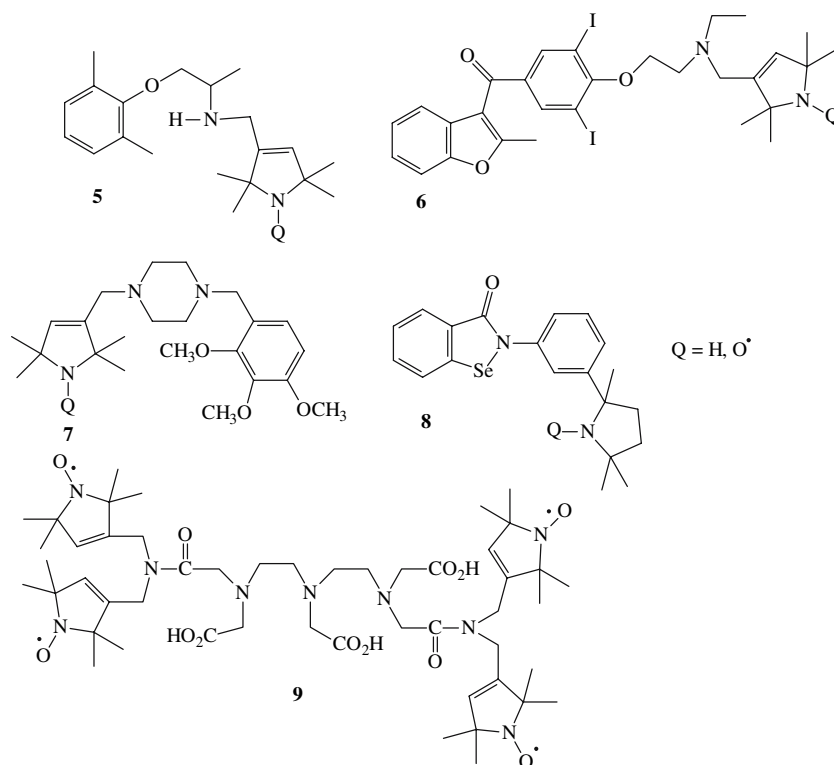
The superoxide dismutase activity of nitroxides is well known (Eqs. 7 and 8) [17]. The process is acid catalyzed, but the reaction is rather slow at physiological pH values. Thus the superoxide dismutating activity, presumably, is not the only cause of antioxidant activity of compound 4 (H-2954) [18]. The nitroxides scavenge hydroxyl radicals (Eq. 9), oxidise transition metals such as  $Fe^{2+}$  (Eq. 10),

pre-empting their participation in Fenton reaction (Eq. 5) [19], and oxidize semiquinones to quinones (Eq. 11) without oxygen consumption or superoxide radical formation in a manner analogous to the juglone semiquinone radical oxidation (Scheme 3) [20]. It is interesting to note that nitroxides are also efficient scavengers of  $NO_2$  and  $CO_3^{\cdot-}$ . Although these species do not form during the oxidative metabolism of DOX, hydroxyl radicals formed during the metabolism may react further with  $NO_2$  to yield  $NO_2^{\cdot}$  or with  $CO_3^{\cdot-}$  to give  $CO_3^{\cdot-}$ . Considering that both anions are possible putative parts of biological systems, this scavenging capability may be important [18].

### Conclusions and outlook

Our results indicate that nitroxides or their amine precursors play multiple roles in elimination of ROS formed during DOX-oxidative metabolism without reducing their anticancer effect. The target-oriented nitroxide 4 or nitroxide precursor 3 are proven to be effective in the prevention of the DOX-induced cardiotoxicity. Modified cardioactive compounds such as mexiletine derivative 5, [21] amiodarone analog 6, [22] trimetazidine derivative 7, [23] ebselen analog 8 [24] and complex forming compound 9 [25], which presumably also have a protective effects (Scheme 4). Compounds 5–7 are advantageous in that they accumulate in membranes of cardiomyocytes and can

**Scheme 4** Potentially active nitroxides and amine precursors in prevention or decrease of DOX-induced cardiotoxicity



detoxify ROS in statu nascendi by both stoichiometric and catalytic pathways. Compounds **8**, as an ebselen analog, and compound **9**, as a paramagnetic MRI agent, may not be target specific, however beyond the catalytic and stoichiometric ROS detoxifying capability of nitroxides they have glutathion-peroxidase-like activity and metal complex forming capability, respectively. Further investigations of compounds **5–9** are in progress.

**Acknowledgment** This work was supported Hungarian National Research Fund OTKAT048334 and OTKA T042951.

## References

- Ross, W. E., Glaubiger, D. L., & Kohn, K. W. (1978). Protein associated DNA breaks in cells treated with adriamycin or ellipticine. *Biochimica et Biophysica Acta*, *519*, 23–30.
- Hertzberg, R. P., & Dervan, P. B. (1984). Cleavage of DNA with methidiumpropyl-EDTA-Iron(II)-reaction conditions and product analyses. *Biochemistry*, *23*, 3934–3945.
- Walling, C. (1975). Fenton's reagent revisited. *Accounts of Chemical Research*, *8*, 125–131.
- Hasinoff, B. B., Kuschak, T. I., Jalowich, J. C., & Creighton, A. M. (1995). A QSAR study comparing the cytotoxicity and DNA topoisomerase II inhibitory effects of bisdioxopiperazine analogs of ICRF-187 (Dexrazoxane). *Biochemical Pharmacology*, *50*, 953–958.
- Wiseman, L. R., & Spencer, C. M. (1998). Dexrazoxane. A review of its use as a cardioprotective agent in patients receiving anthracycline-based chemotherapy. *Drugs*, *56*, 385–403.
- Swenson, C. E., Bolcsak, L. E., Batist, G., Guthrie, T. H., Tkaczuk, K. H., Boxenbaum, H., Welles, L., Show, S. C., Bhamra, R., & Chakin, P. (2003). Pharmacokinetics of doxorubicin administered i.v. as myocet (TLC D-99; liposome-encapsulated doxorubicin citrate) compared with conventional doxorubicin when given in combination with cyclophosphamide in patients with metastatic breast cancer. *Anti-Cancer Drugs*, *14*, 239–246.
- Abou El Hassan, M. A. I., Kedde, M. A., Zwiers, U. T. H., Tourn, E., Haennen, G. R. M., Bast, A., & van der Vijgh, W. J. F. (2003). Bioavailability and pharmacokinetics of the cardioprotecting flavonoid 7-mono-hydroxyethylrutin in mice. *Cancer Chemotherapy and Pharmacology*, *52*, 371–376.
- Monti, E., Cova, D., Guido, E., Morelli, R., & Oliva, C. (1996). Protective effect of the nitroxide tempol against the cardiotoxicity of adriamycin. *Free Radical Biology and Medicine*, *21*, 463–470.
- DeGraff, W., Hahn, S. M., Mitchell, J. B., & Krishna, M. C. (1994). Free radical modes of cytotoxicity of adriamycin<sup>®</sup> and streptonigrin. *Biochemical Pharmacology*, *48*, 1427–1435.
- Hankovszky, H. O., Hideg, K., Bódi, I., & Frank, L. (1986). New Antiarrhythmic Agents. 2,2,5,5-Tetramethyl-3-pyrroline-3-carboxamides and 2,2,5,5-tetramethyl-pyrrolidine-3-carboxamides. *Journal of Medicinal Chemistry*, *29*, 1138–1152.
- Twomey, P., Taira, J., DeGraff, W., Mitchell, J. B., Russo, A., Krishna, M. C., Hankovszky, H. O., Frank, L., & Hideg, K. (1997). Direct evidence for *In Vivo* nitroxide free radical production from a new antiarrhythmic drug by EPR spectroscopy. *Free Radical Biology and Medicine*, *22*, 909–916.
- Shankar, R.A., Hideg, K., Zweier, J. L., & Kuppusamy, P. (2000). Targeted antioxidant properties of *N*-[(2,2,5,5-tetramethyl-3-pyrroline-3-carboxamido) propylphthalimide], a new antiarrhythmic drug and its nitroxide-metabolite in preventing postischemic myocardial injury. *Journal of Pharmacology and Experimental Therapeutics*, *292*, 838–845.
- Marton, Zs., Halmosi, R., Horváth, B., Alexy, T., Késmárky, G., Vékási, J., Battyány, I., Hideg, K., & Tóth, K. (2001). Scavenger effect of experimental and clinically used cardiovascular drugs. *Journal of Cardiovascular Pharmacology*, *38*, 745–753.
- Deres, P., Halmosi, R., Tóth, A., Kovács, K., Pálfi, A., Habon, T., Czopf, L., Kálai, T., Hideg, K., Sümegi, B., & Tóth, K. (2005). Prevention of doxorubicin-induced acute cardiotoxicity by an experimental antioxidant compound. *Journal of Cardiovascular Pharmacology*, *45*, 36–43.
- Mordente, A., Meucci, E., Martorana, G. E., Giardina, B., & Minotti, G. (2001). Human heart cytosolic reductases and anthracycline cardiotoxicity. *IUBMB Life*, *52*, 83–88.
- Kálai, T., Hideg, É., Vass, I. & Hideg, K. (1998) Double (Fluorescent and Spin) Sensors for detection of reactive oxygen species in the thylakoid membrane. *Free Radical Biology and Medicine*, *24*, 649–652.
- Krishna, M. C., Russo, A., Mitchell, J. B., Goldstein, S., Hafini, H., & Samuni, A. (1996). Do nitroxide antioxidants act as scavengers of O<sub>2</sub><sup>-</sup> or as SOD mimics? *Journal of Biological Chemistry*, *271*, 26026–26031.
- Goldstein, S., Samuni, A., Hideg, K., & Merényi, G. (2006). Structure-activity relationship of cyclic nitroxides as SOD mimics and scavengers of nitrogen dioxide and carbonate radicals. *Journal of Physical Chemistry A*, *110*, 3679–3685.
- Glebska, J., Pulaski, L., Gwozdinski, K., & Kolimowski, J. (2001). Structure-activity relationship studies of protective function of nitroxides in Fenton system. *BioMetals*, *14*, 159–170.
- Zhang, R., Hirsch, O., Mohsen, M., & Samuni, A. (1994). Effects of nitroxide stable radicals on juglone cytotoxicity. *Archives of Biochemistry and Biophysics*, *312*, 385–391.
- Li, H., Xu, K. Y., Zhou, L., Kálai, T., Zweier, J. L., Hideg, K., & Kuppusamy, P. (2000). A pyrroline derivative of mexiletine offers marked protection against ischemia/reperfusion-induced myocardial contractile dysfunction. *Journal of Pharmacology and Experimental Therapeutics*, *295*, 563–571.
- Bognár, Z., Kálai, T., Pálfi, A., Hantó, K., Bognár, B., Márk, L., Szabó, Z., Tapodi, A., Radnai, B., Sárszegi, Zs., Szántó, Á. Jr., Gallyas, F., Hideg, K., Sümegi, B., & Várbró, G. (2006). A novel SOD-mimetic permeability transition inhibitor agent protects ischemic heart by inhibiting both apoptotic and necrotic cell death. *Free Radical Biology and Medicine*, *41*, 835–848.
- Kutala, V. K., Khan, M., Mandal, R., Ganesan, L. P., Tridandapani, S., Kálai, T., Hideg, K., & Kuppusamy, P. (2006). Attenuation of myocardial ischemia-reperfusion injury by trimetazidine derivatives functionalized with antioxidant properties. *Journal of Pharmacology and Experimental Therapeutics*, *317*, 921–928.
- Kálai, T., Muges, G., Roy, G., Sies, H., Berente, Z., & Hideg, K. (2005). Combining benzol[*d*]isosenazol-3-ones with sterically hindered alicyclic amines and nitroxides: enhanced activity as glutathione peroxidase mimics. *Organic Biomolecular Chemistry*, *3*, 3564–3569.
- Jászberényi, Z., Brücher, E., Jekó, J., Hideg, K., Kálai, T. & Király, R. (2003). Synthesis, equilibrium and kinetic properties of the Gd<sup>3+</sup> complexes of three spin labeled DTPA-bis(amide) derivative ligands. *European Journal of Inorganic Chemistry*, *2003*, 3601–3608.