

Aluminum Chloride Causes the Dysfunction of Testes Through Inhibiting the ATPase Enzyme Activities and Gonadotropin Receptor Expression in Rats

Xudong Sun^{1,2} · Hao Sun³ · Kaiyuan Yu¹ · Zhongying Wang⁴ · Yanfen Liu¹ · Kexiang Liu¹ · Yanzhu Zhu⁵ · Yanfei Li¹

Received: 24 June 2017 / Accepted: 4 August 2017 / Published online: 30 August 2017
© Springer Science+Business Media, LLC 2017

Abstract The aim of this experiment is to explore the effects of aluminum chloride (AlCl_3) on the ATPase enzymes and gonadotropin receptors in the testes. Eighty male Wistar rats were orally exposed to 0 mg/kg body weight (BW) (control group, CG), 64 mg/kg BW (low-dose group, LG), 128 mg/kg BW (mid-dose group, MG), or 256 mg/kg BW (high-dose group, HG) for 120 days. The microstructure and ultrastructure of testes; the activities of $\text{Na}^+\text{-K}^+\text{-ATPase}$, $\text{Mg}^{2+}\text{-ATPase}$, and $\text{Ca}^{2+}\text{-ATPase}$; and the mRNA and protein expressions of follicle-stimulating hormone receptor (FSHR) and luteinizing hormone receptors (LHR) in the testes were examined. The results showed that the testes histological structure were damaged; the activities of $\text{Na}^+\text{-K}^+\text{-ATPase}$, $\text{Mg}^{2+}\text{-ATPase}$, and $\text{Ca}^{2+}\text{-ATPase}$, the mRNA and protein expressions of FSHR and LHR in the testes were all decreased in the rats with AlCl_3 exposure. It indicates that AlCl_3 causes the dysfunction of testes in rats.

Keywords Aluminum chloride-rat · Testes · ATPase enzymes · Gonadotropin receptor

Introduction

Human male infertility is a major health problem that affects approximately 10 to 15% of couples worldwide [1]. Male infertility, as the most difficult form of infertility, is governed by a variety of causative factors, including environmental disruptors, genetic defects, physiological and endocrine failure, and testis pathologies [2]. Despite 60–75% of male infertility are idiopathic, the disease is usually accompanied by qualitative (asthenospermia, teratozoospermia, and necrospermia) and quantitative (azoospermia, cryptozoospermia, and oligoasthenozoospermia) abnormalities [3, 4]. Aluminum (Al) exposure is one of the important pathogenesis for male infertility [5]. Al, as the third most abundant element on Earth's crust, is widely used in Al containers and utensils, medicines, water purifiers, and food additives, which induces a global public health problem [6, 7]. Aluminum trichloride (AlCl_3) (34 mg/kg body weight (BW); 25 mg/kg BW) impaired the structure of testes and epididimides in the rat and mice, respectively [8, 9]. Results obtained from Yousef et al. [10] revealed that rabbits which were orally administered AlCl_3 at 34 mg/kg BW every other day for 16 weeks exerted significant decrease in sperm activities and numbers [10]. Ige and Akhigbe [6] suggested that AlCl_3 exposure caused male rats infertility via oxidative damage. These evidences demonstrate that Al is a potential risk for male infertility, but the investigation on the ionic transport or gonadotropin receptors in male rat testes exposure to AlCl_3 are lacking.

One of the major mechanisms behind toxic effects of Al on reproductive system has been attributed to ionic disorders.

Xudong Sun and Hao Sun contributed equally to this study.

✉ Yanfei Li
liyanfei@neau.edu.cn

¹ Key Laboratory of the Provincial Education Department of Heilongjiang for Common Animal Disease Prevention and Treatment, Northeast Agricultural University, Harbin 150030, China

² Key Laboratory of Zoonosis, Ministry of Education, College of Veterinary Medicine, Jilin University, 5333 Xi'an Road, Changchun, Jilin 130062, China

³ Laishan District Bureau of Commerce of Yantai city, Yantan, Shandong Province 264003, China

⁴ Vascular Surgery Department, The First hospital of Jilin University, Changchun 130021, China

⁵ Institute of Special Animal and Plant Sciences of Chinese Academy of Agricultural Sciences, Changchun 130112, China

ATPases, which constitute a major category of ion transporters in the human body, have a variety of significant biological and pathological roles. ATPases can maintain proper function of the vasculature through regulating the transmembrane ionic balance [10], and many studies have shown that ionic disorders in the vasculature give rise to testes dysfunction. With sublethal dose (3.5 mg/kg BW) of Al acetate, total ATPase activity decreased in testes of albino mice [11]. Our previous research observed that chronic AlCl_3 administration decreased ovary $\text{Na}^+\text{-K}^+\text{-ATPase}$, $\text{Mg}^{2+}\text{-ATPase}$, and $\text{Ca}^{2+}\text{-ATPase}$ activity; intracellular energy production; and incomplete cell membrane [12]. However, whether there is a similar effect in rat testes needs to be explored.

The follicle-stimulating hormone receptor (FSHR) and luteinizing hormone receptor (LHR) were expressed on sertoli cells and leydig cells, and their activation promoted the support and nourishment of germ cells during spermatogenesis. The FSHR and LHR are the basis for the physiological functions of follicular stimulating hormone (FSH) and luteinizing hormone (LH). One study showed that male rats orally administered AlCl_3 at 100 mg/kg BW for 8 weeks exerted significant decrease of FSH and LH [6]. Also, male albino rat exposure to aluminum sulfate (50 mg/kg BW) for 45 consecutive days showed significant decrease in serum FSH and LH concentration [13]. But, limited data showed the effects of AlCl_3 on the protein and mRNA expressions of FSHR and LHR in testes.

Therefore, the effects of AlCl_3 exposure on the testes **histological** structure, the ATPase activities ($\text{Na}^+\text{-K}^+\text{-ATPase}$, $\text{Mg}^{2+}\text{-ATPase}$, and $\text{Ca}^{2+}\text{-ATPase}$), and the gonadotropin receptors (measured as protein and mRNA expressions of FSHR and LHR) in the testes of rats were examined to depict the basic toxicological mechanisms of AlCl_3 on the testes.

Material and Methods

Rats

Eighty healthy male Wistar rats (3 weeks old) that weighed 70–95 g were allocated into four groups according to their weights (each group 20 rats). All the rats were acclimatized for 1 week, and then were provided with 0 (control group, CG), 0.4 mg/mL (low-dose group, LG), 0.8 mg/mL (mid-dose group, MG), and 1.6 mg/mL (high-dose group, HG) AlCl_3 in drinking water for 120 days, respectively. During the whole exposure duration, the water consumption was increased gradually with the increased of rat BW. And the water consumption of the individual rat averaged at 16 ± 2 mL/day for 100 g BW per day, resulting in the dose of AlCl_3 at 0, 64, 128, or 256 mg/kg BW for treatment groups. The dose of this experiment was determined referencing the research of Zhu et al.

[14], which simulated the process of Al exposure in the environment, especially for workers who worked in an electroplating factory and Al mines or factories. Rats were housed in the biomedical research center, northeast agriculture university. The housing conditions were maintained according to the request of rats. The rats were kept in plastic cages (5 rats per cage). The size of all the cages is $470 \times 300 \times 150$ mm, large enough for five mature rats. Rats were given the drinking water and food ad libitum. The health status of rats was daily monitored.

Sample Collection

After 120 days, the rats were sacrificed under light ether anesthesia, and the testes were collected from each rat. The testes were used to observe the testes morphology and examine the activities of $\text{Na}^+\text{-K}^+\text{-ATPase}$, $\text{Mg}^{2+}\text{-ATPase}$, and $\text{Ca}^{2+}\text{-ATPase}$ and the protein and mRNA expressions of FSHR and LHR.

Observation of Testes Microstructure

The 10% neutral buffered formalin solution-fixed testes were embedded in paraffin; sections with a thickness of 5–6 μm were sliced from the paraffin-embedded blocks and stained with hematoxylin and eosin (H&E) stain [15]. Then, the sections were viewed and photographed using an Olympus light microscope (Olympus BX51, Tokyo, Japan) with an attached photographic machine (Olympus E-330, Olympus Optical Co. Ltd., Japan).

Observation of Testes Ultrastructure

The testes samples were fixed in 2.5% glutaraldehyde solution for a week. The samples were embedded in Spurr's resin by using Leica/LKB Embedding Capsules Easy Molds. Embedded samples were sectioned to a thickness of 600 \AA and stained with lead staining solution and analyzed with a transmission electron microscope T-400 (Philips, Eindhoven, NL).

The Detection of ATPase Activity in the Testes

The one part of each testis was quantified 0.1 g, and the sample was processed to obtain 10 mL of 10% testes tissue homogenate. The tissue homogenate had been centrifuged at 3000 rpm for 10 min. The supernatant of the tissue homogenate was collected and used to detect the activity of $\text{Na}^+\text{-K}^+\text{-ATPase}$, $\text{Mg}^{2+}\text{-ATPase}$, and $\text{Ca}^{2+}\text{-ATPase}$ using ^{125}I radioimmunoassay (RIA) kits (New Bay Biological Technology Co., Ltd., Tianjin, China) [12]. The experiment procedure was followed by kit introduction.

Table 1 The primer sequence of the FSHR, LHR, and β -actin

Gene	Number	Upstream and downstream primer sequence	Primer length (bp)	Product length (bp)
FSHR	NM_199237	F:5'-GTCCTCATCAAGCGACACCA-3'	23	103
		R:5'-GGAGGCAGAAATGGCAAAGA-3'	23	
LHR	NM_012978	F:5'-CTATCTCCCTGTCAAAGTAA-3'	22	384
		R:5'-CCATTTCCATACAGTTTTAG-3'	22	
β -actin	NM_031144	F:5'-CGGGACCTGACAGACTACCT-3'	20	328
		R:5'-AGACAGCACTGTGTTGGCATAG-3'	20	

Detection of FSHR and LHR Protein Expression

The testes sample was fixed and embedded by paraffin, then stained with FSHR and LHR immunohistochemistry kits (Beijing Biosynthesis Biotechnology Co. LTD, Beijing, China/Wuhan Biosynthesis Biotechnology Co. LTD., Wuhan, China). Sections were observed under a microscope (model: BA400, Motic, German). The positive gray value was measured by Motic image 3.2 micrograph analysis software (Motic, German), which was applied to quantify the nuclear FSHR and LHR contents. The positive gray value was negatively correlated with the protein expression of FSHR and LHR.

Determination of FSHR and LHR mRNA Expression

The expressions of the FSHR and LHR mRNA were detected in the testes by quantitative real-time PCR (QRT-PCR) with SYBR Premix Ex Taq kit (Takara, Shiga, Japan). The primers of FSHR and LHR were designed by Premier Premier 5.0 and OLIGO 6.0 Software and were synthesized by Sangon biotech (Shanghai, China) (Table 1). The relative expression of target genes was determined by the $2^{-\Delta\Delta CT}$ method

[16]. The detail process was conducted according to Sun et al. [17].

Statistical Analysis

Statistical analyses were done using SPSS 22.0 package programmer (SPSS Inc., Chicago, IL, USA) with one-way analysis of variance followed by LSD test. Data are shown as least square means and standard deviation (SD, bar on the top of each column). The p values of less than 0.05 were considered significant, and p values of less than 0.01 were considered markedly significant.

Results

BW of Rats

As shown in Fig. 1, the BW of rats was decreased gradually with the increase of $AlCl_3$ dose, and the BW of rats in LG, MG, and HE was significantly lower than that in CG on day 120 ($p < 0.05$; $p < 0.01$).

Clinical Symptoms

The control rats had nice sprite, normal eating and drinking skills, light clothing hair, and normal dung. With the increase of the $AlCl_3$ dose, the rats exerted bad, anorexia, loose clothing hair. The rats walk stumble and the tail of the rats got pale. The response to the outside stimulation turned weakness and the condition would get worse with the increase of $AlCl_3$ dose (Fig. 2a). In MG and HG, the rats exerted polypnea and anorexia. The rats in HG were dissected and found that the testes represented congestion, bleed, edema, and wither compared with CG (Fig. 2b).

Microstructure of Testes

The testes microstructure image were examined using paraffin section that was stained routinely with H&E (Fig. 3). The testes microstructural features in the CG revealed intact

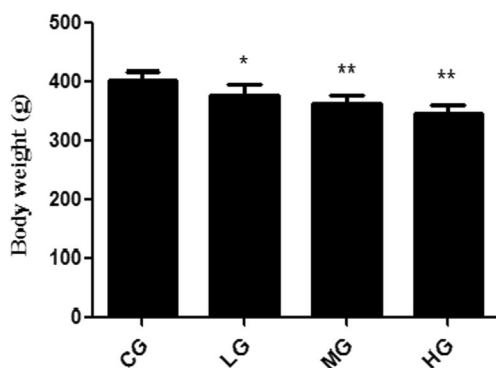
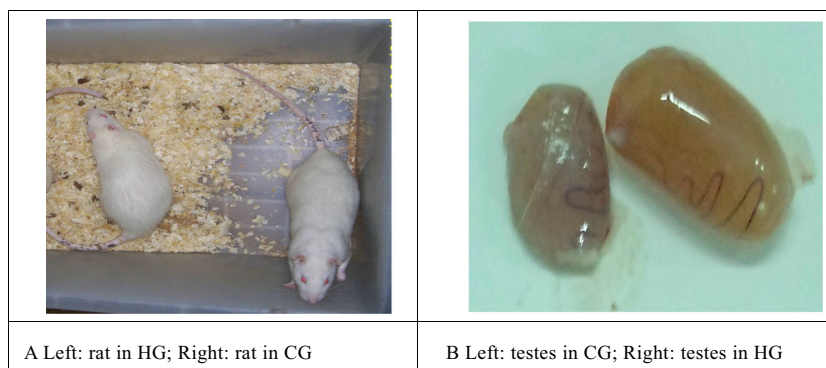


Fig. 1 Effects of $AlCl_3$ on the BW of rats. The rats were orally exposed to 0 (control group, CG), 0.4 mg/mL (low-dose group, LG), 0.8 mg/mL (mid-dose group, MG), and 1.6 mg/mL (high-dose group, HG) $AlCl_3$ in drinking water for 120 days. Data are means \pm SD ($n = 20$ per group). * $p < 0.05$, ** $p < 0.01$ versus CG

Fig. 2 Effects of AlCl_3 on the clinical symptoms of rats. CG control group, HG high-dose group

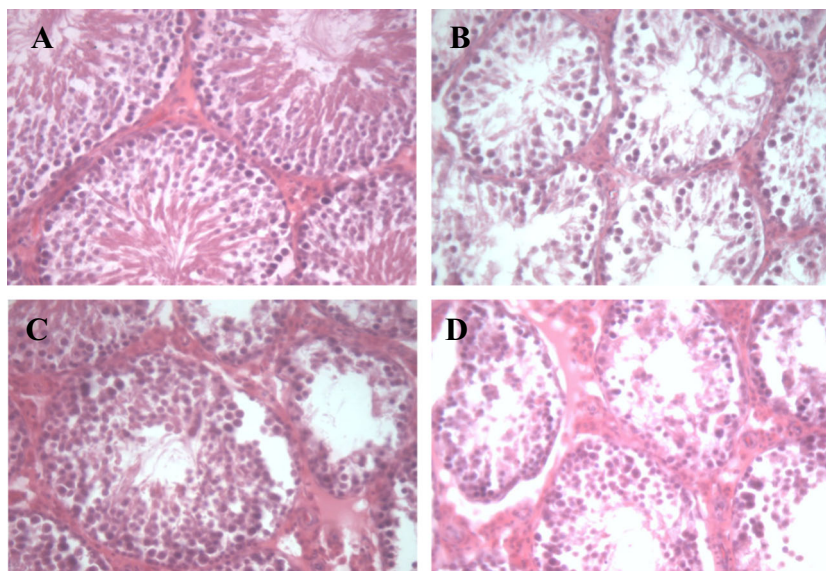


morphology and structure of spermatogenic epithelial cells, without shedding of cells in lumen, and a large number of sperms (Fig. 3a). We found that testicular stroma slightly expanded, the number of spermatogenic cells decreased, the number of sperm decreased in the LG (Fig. 3b); testicular interstitial eosinophilic enhanced, spermatogenic cells fall into the lumen, the number of sperm significantly decreased in the MG (Fig. 3c); testicular stroma significantly expanded, the seminiferous tubule lumen was narrow, fewer sperm in the HG (Fig. 3d).

Ultrastructure of Testes

The testes ultrastructural features was observed using transmission electron microscope (Fig. 4). The testes ultrastructural features in the CG revealed intact cell membrane, dispersed chromatin, and normal morphology of mitochondria (Fig. 4a). In the treatment groups, we found that cell disintegration and incomplete cell membrane, lots of foam-like structure, mitochondrial swelling, irregular nuclear envelope (Fig. 4b–d).

Fig. 3 Effects of AlCl_3 on the testes microstructure of rats. The microstructure of testes was measured by H&E stain. **a** CG (control group). **b** LG (low-dose group). **c** MG (mid-dose group). **d** HG (high-dose group). ($\times 400$)



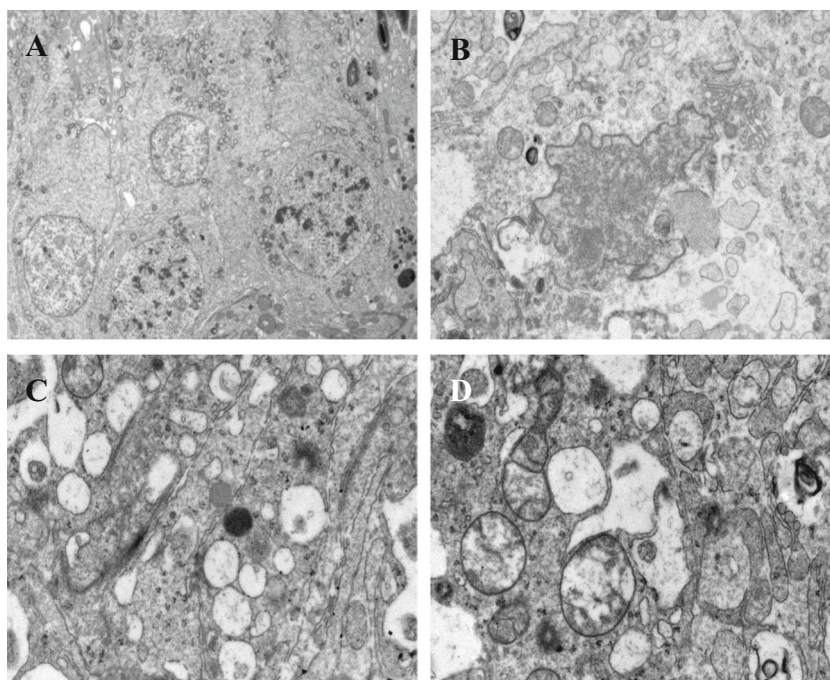
The Activities of $\text{Na}^+\text{-K}^+\text{-ATPase}$, $\text{Mg}^{2+}\text{-ATPase}$, and $\text{Ca}^{2+}\text{-ATPase}$ in Testes

The activities of $\text{Na}^+\text{-K}^+\text{-ATPase}$, $\text{Mg}^{2+}\text{-ATPase}$, and $\text{Ca}^{2+}\text{-ATPase}$ in testes were gradually decreased with the increase of AlCl_3 (Fig. 5). The activity of $\text{Mg}^{2+}\text{-ATPase}$ in MG ($p < 0.05$) and HG ($p < 0.01$) was significantly lower than those in CG. The activities of $\text{Na}^+\text{-K}^+\text{-ATPase}$ and $\text{Ca}^{2+}\text{-ATPase}$ in HG ($p < 0.05$) were significantly lower than those in CG.

The Protein Expressions of FSHR and LHR in Testes

The protein expressions of FSHR and LHR were showed in the four groups (Figs. 6 and 7). The positive gray value of FSHR and LHR was increased in AlCl_3 -treated rat testes. The positive gray values of FSHR and LHR in MG ($p < 0.05$) and HG ($p < 0.01$) were significantly higher than those in CG (Fig.~8). The positive gray values of FSHR and LHR in testes were negatively correlated with the protein

Fig. 4 Effects of AlCl_3 on the testes ultrastructure of rats. The ultrastructure of testes was observed by transmission electron microscope. **a** Testes tissue of rats in CG (control group) ($\times 2550$). Testes tissue of rats in HG (high-dose group), respectively, at $\times 9000$ (**b**), $\times 16,500$ (**c**), and $\times 16,500$ (**d**)



expressions of FSHR and LHR. Therefore, the protein expressions of FSHR and LHR were decreased gradually with the increase dose of AlCl_3 . The protein expressions of FSHR in MG and HG were significantly lower than in CG ($p < 0.01$).

The mRNA Expressions of FSHR and LHR in Testes

The FSHR and LHR mRNA expressions were gradually decreased in AlCl_3 -treated rats compared with CG

(Fig.~9). The FSHR and LHR mRNA expressions in MG and HG were markedly lower than those in CG ($p < 0.01$). The FSHR and LHR mRNA expressions in testes were positively correlated with the protein expressions of FSHR and LHR. Therefore, FSHR and LHR mRNA expressions were gradually decreased in AlCl_3 -treated rats compared with CG. The FSHR and LHR mRNA expressions in MG and HG were markedly lower than in CG ($p < 0.01$).

Fig. 5 Effects of AlCl_3 on the activities of Na^+ - K^+ -ATPase, Mg^{2+} -ATPase, and Ca^{2+} -ATPase in testes. The activities of Na^+ - K^+ -ATPase, Mg^{2+} -ATPase, and Ca^{2+} -ATPase were detected by ^{125}I radioimmunoassay (RIA) kits. CG control group, LG low-dose group, MG mid-dose group, HG high-dose group. Data are means \pm SD ($n = 20$ per group). * $p < 0.05$, ** $p < 0.01$ versus CG

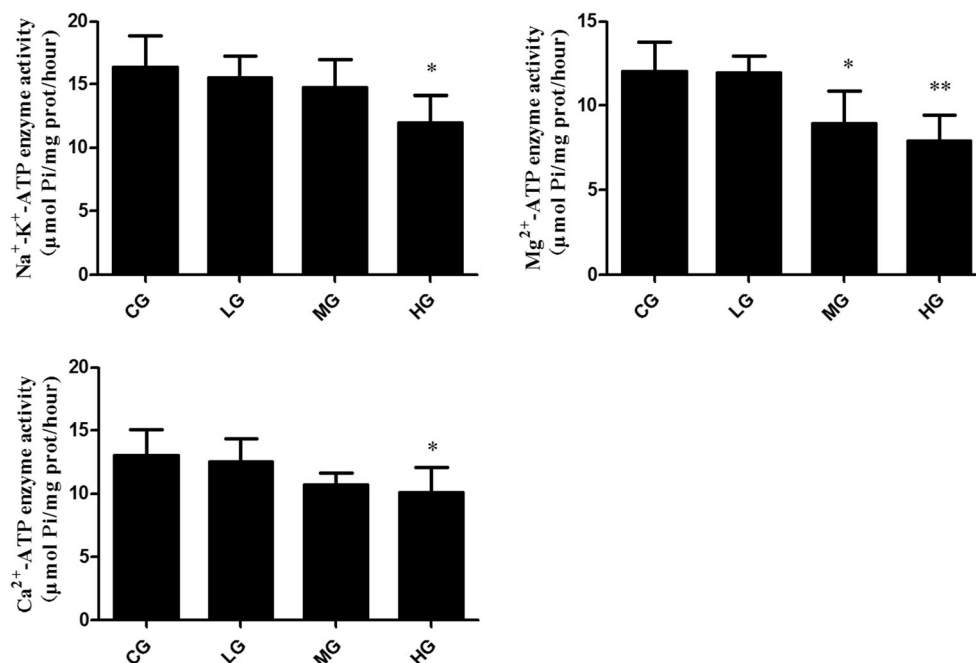
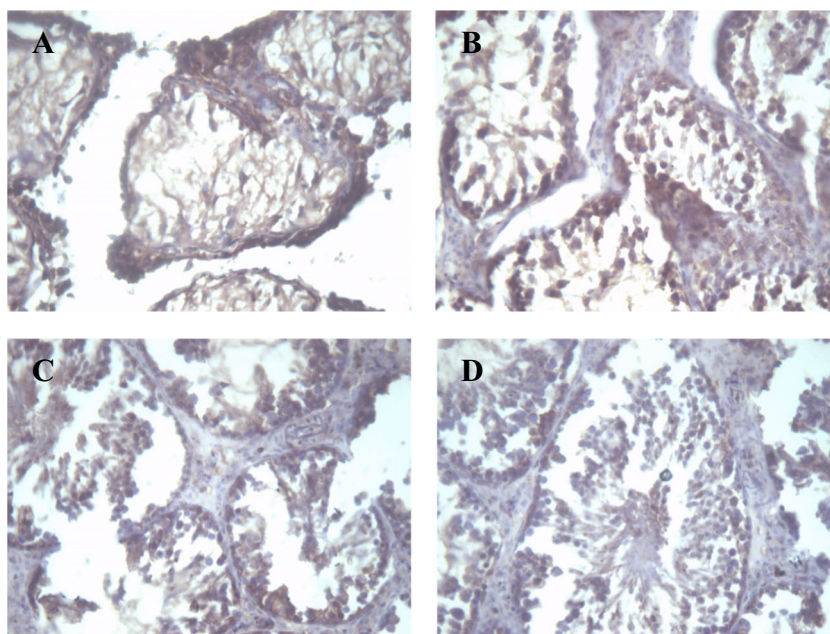


Fig. 6 Effects of AlCl_3 on the protein expression of FSHR in testes. The protein expression of FSHR was detected by immunohistochemistry kits. *CG* control group (a), *LG* low-dose group (b), *MG* mid-dose group (c), *HG* high-dose group (d) $\times 400$



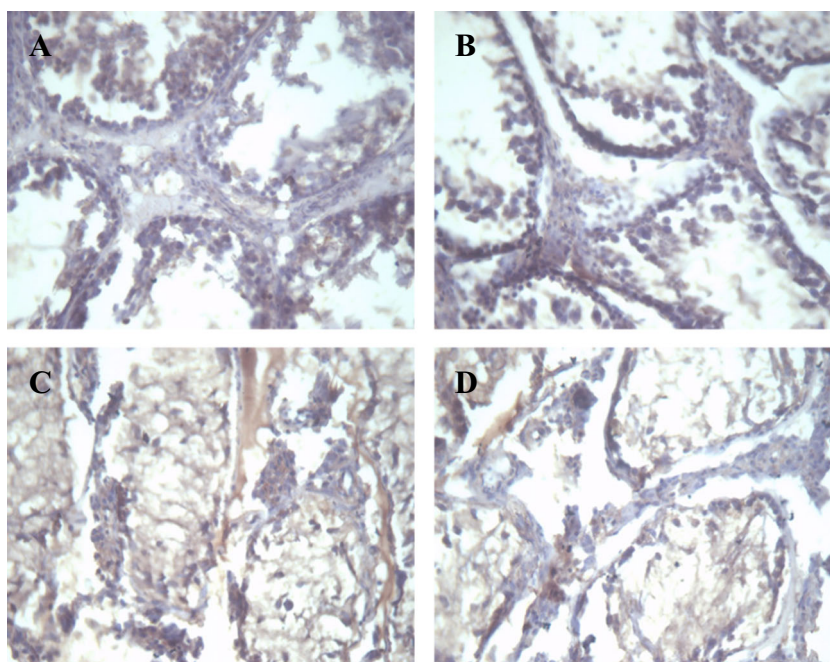
Discussion

Al is well-known water contaminant. Atmospheric acidification and bauxite mine over-exploitation are causing a progressively massive export of Al from mountains to surface waters, putting animals and humans in contact with absorbable cationic Al [18]. In this experiment, Wistar rats were orally administered of AlCl_3 in drinking water, which simulated the process of Al exposure in the environment. Furthermore, orally administered AlCl_3 in drinking water can reduce the stress response of rats caused by intragastric administration or intraperitoneal injection of AlCl_3 . Thus, oral delivery of aluminum to rats was chosen in this study.

In this study, no deaths were observed in AlCl_3 -treated rats. But the rats exerted bad, anorexia, loose clothing hair. Compared with CG, congestion, bleed, edema, and wither were observed in the testes of HG. Furthermore, AlCl_3 exposure damaged the cell nucleus, mitochondria, and cell membrane, disordered testicular stroma, and decreased the number of sperm, which agreed with previous study [10]. These results implied that AlCl_3 exposure disrupted the structure and inhibited the development of the testes, which preliminarily confirmed that the rat model with AlCl_3 exposure was built.

Na^+/K^+ -ATPase, Mg^{2+} -ATPase, and Ca^{2+} -ATPase were the markers of the cell impairment during toxic exposure

Fig. 7 Effects of AlCl_3 on the protein expression of LHR in testes. The protein expression of LHR was detected by immunohistochemistry kits. *CG* control group (a), *LG* low-dose group (b), *MG* mid-dose group (c), *HG* high-dose group (d) $\times 400$



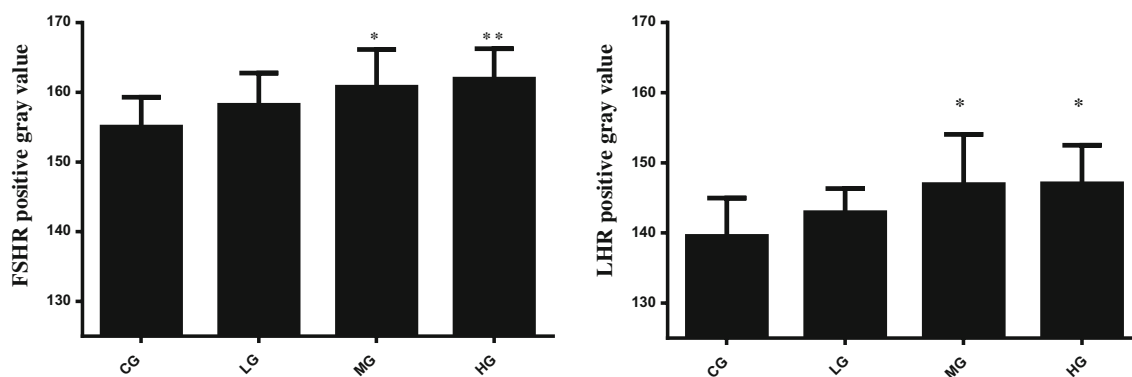


Fig. 8 The positive gray value of FSHR and LHR in testes. The positive gray value was negatively correlated with the protein expressions of FSHR and LHR. CG control group, LG low-dose group, MG mid-dose group, HG high-dose group. * $p < 0.05$, ** $p < 0.01$ versus CG

[19]. In this experiment, the activities of the $\text{Na}^+\text{-K}^+\text{-ATPase}$, $\text{Mg}^{2+}\text{-ATPase}$, and $\text{Ca}^{2+}\text{-ATPase}$ were gradually decreased with AlCl_3 exposure, indicating that AlCl_3 exposure induced the intracellular accumulation of Na^+ and Ca^{2+} , leading to the cytotoxic effect and impairment of the cells in the testes. Al can directly interfere with -SH groups of enzyme at the active site, thus prevents the -SH groups from functioning in certain chemical reactions [20]. The -SH groups are involved in the maintenance of the membrane-bound $\text{Na}^+\text{-K}^+\text{-ATPase}$ oligomeric structure [21]. It indicates that Al lowers the activities of $\text{Na}^+\text{-K}^+\text{-ATPase}$, $\text{Ca}^{2+}\text{-ATPases}$, and $\text{Mg}^{2+}\text{-ATPases}$ by the reduction of -SH groups. In addition, Al perturbed the structure and functions of cell membranes through regulating apical Cl^- secretion and ATPase inactivation in class of lipids [22, 23]. $\text{Na}^+\text{-K}^+\text{-ATPase}$ enzyme presents the cell membrane of all the animals. Thus, AlCl_3 can decrease the activity of $\text{Na}^+\text{-K}^+\text{-ATPase}$ by disruption of the membrane. Calcium (Ca) is actively transported into intracellular organelles and out of the cytoplasm by $\text{Ca}^{2+}/\text{Mg}^{2+}\text{-ATPases}$ located in the endoplasmic reticulum and plasma membranes [24]. A recent investigation showed that disintegration of endoplasmic reticulum was observed when young gerbils were injected intraperitoneally with AlCl_3 for 5 weeks

[25]. Thus, the decrease of $\text{Ca}^{2+}/\text{Mg}^{2+}\text{-ATPases}$ attributed to the disintegration of endoplasmic reticulum induced by Al. Al binds to the plasma membrane phospholipids, alters the lipid protein interaction, and modifies the activities of the transporters [26]. Al^{3+} accelerates Ca^{2+} release from mitochondria and strongly inhibits $\text{Ca}^{2+}\text{-ATPase}$ activity [27]. These results indicate that AlCl_3 can induce ionic disorders by altering ATPases activity and contribute to testes dysfunction during intoxication.

FSH and LH, synthesized and secreted by the pituitary, regulate spermatogenesis through sperms and testosterone during adulthood. Sun et al. [17] found that the concentration of LH decreased in all AlCl_3 -treated groups. In addition, AlCl_3 decreased the zinc (Zn) concentration in rats [5]. And Zn-deficient animals, chiefly males, have been shown to have lower concentrations of FSH and LH [28]. Thus, AlCl_3 might inhibit the secretion of FSH and LH through reducing Zn concentration. The physiological functions of FSH and LH are mediated by specific receptors—FSHR and LHR. In this experiment, the mRNA and protein expressions of FSHR and LHR in the testes decreased in the rats with AlCl_3 exposure. Rawi and Seif Al Nassr [13] found that the concentration of LH was decreased, but there were no significant changes in

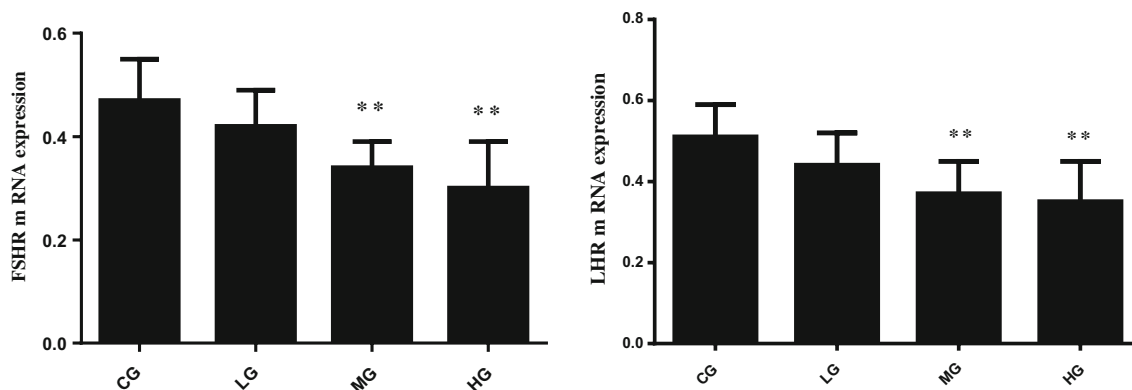


Fig. 9 Effects of AlCl_3 on FSHR and LHR mRNA expressions in testes. The FSHR and LHR mRNA expressions in testes were detected by QRT-PCR. CG control group, LG low-dose group, MG mid-dose group, HG high-dose group. Data are means \pm SD ($n = 20$ per group). ** $p < 0.01$ versus CG

FSH concentration in AlCl_3 -treated male rats. Thus, we deduced that the lower level of LH was a reason for the degeneration of LHR with AlCl_3 exposure. So, AlCl_3 might inhibit not only the secretion of LH, but also the expressions of FSHR and LHR. However, the FSH concentration is not related to the protein and mRNA expressions of FSHR and LHR, which needs to be explored.

Al toxicity elicits a dysfunctional tricarboxylic acid cycle and impedes ATP production [29]. Study showed that mitochondrial ATP production was suppressed by generating ROS via transduction [30]. Studies showed that Al accumulation induced testicular oxidative stress [31, 32]. Thus, we deduced that Al may also suppress mitochondrial ATP production by transduction of generating ROS. Then, the lower energy will block the mRNA and protein expressions of FSHR and LHR in the testes. Furthermore, the lower activities of $\text{Na}^+\text{-K}^+$ ATPase, Ca^{2+} -ATPases, and Mg^{2+} -ATPases induced by AlCl_3 lead to the intracellular accumulation of Na^+ and Ca^{2+} , inducing cytotoxic effect and impairment of the cells in the testes (sertoli cells and leydig cells). So, the lower mRNA and protein expressions of FSHR and LHR may attribute to the lower activity of $\text{Na}^+\text{-K}^+$ ATPase, Ca^{2+} -ATPases, and Mg^{2+} -ATPases. The FSHR and LHR expressed on sertoli cells and leydig cells in the testes. Leydig cell hyperplasia and a destruction of inter-sertoli cell tight junctions were observed [5]. These results indicate that AlCl_3 exposure to the decrease of protein expression of FSHR and LHR may cause the dysfunction of testes.

In conclusion, AlCl_3 exposure damaged the structure of the test, inhibited the ATPase enzymes activities and protein and mRNA expressions of gonadotropin receptors in the testes of rats, which indicated that AlCl_3 disrupted the reproductive regulation of testes.

Acknowledgements This study was supported by a grant from the Natural Science Foundation of Heilongjiang Province (C201425).

Compliance with Ethical Standards The experimental protocol was approved by the Ethics Committee on the Use and Care of Animals, Northeast Agricultural University, China.

Conflict of Interest The authors declare that they have no conflict of interest.

References

- Tahmasbpour E, Balasubramanian D, Agarwal A (2014) A multifaceted approach to understanding male infertility: gene mutations, molecular defects and assisted reproductive techniques (ART). *J Assist Reprod Genet* 31(9):1115–1137
- Agarwal A, Tvrdá E, Sharma R (2014) Relationship amongst teratozoospermia, seminal oxidative stress and male infertility. *Reprod Biol Endocrinol* 12:45
- Oberholster PJ, Myburgh JG, Ashton PJ, Coetzee JJ, Botha AM (2012) Bioaccumulation of aluminium and iron in the food chain of Lake Loskop, South Africa. *Ecotoxicol Environ Saf* 75(1):134–141
- Rengel Z (2004) Aluminum cycling in the soil-plant-animal-human continuum. *Biometals* 17:669–689
- Djouina M, Esquerre N, Desreumaux P, Vignal C, Body-Malapel M (2016) Toxicological consequences of experimental exposure to aluminum in human intestinal epithelial cells. *Food Chem Toxicol* 91:108–116
- Ige SF, Akhigbe RE (2012) The role of *Allium Cepa* on aluminum-induced reproductive dysfunction in experimental male rat models. *J Hum Reprod Sci* 5(2):200–205
- Zhang F, Sun X, Yu H, Yang X, Song M, Han Y et al (2017) Effects of aluminum trichloride on the cartilage stimulatory growth factors in rats. *Biometals* 30(1):143–150
- Moselhy WA, Helmy NA, Abdel-Halim BR, Nabil TM, Abdel-Hamid MI (2012) Role of ginger against the reproductive toxicity of aluminium chloride in albino male rats. *Reprod Domest Anim* 47(2):335–343
- Abdel-Moneim AM (2013) Effects of taurine against histomorphological and ultrastructural changes in the testes of mice exposed to aluminium chloride. *Arh Hig Rada Toksikol* 64(3):405–414
- Yousef MI, El-Morsy AM, Hassan MS (2005) Aluminium-induced deterioration in reproductive performance and seminal plasma biochemistry of male rabbits: protective role of ascorbic acid. *Toxicology* 215(1–2):97–107
- Sushma NJ, Rao KJ (2007) Total ATPases activity in different tissues of albino mice exposed to aluminium acetate. *J Environ Biol* 28(2 Suppl):483–484
- Fu Y, Jia FB, Wang J, Song M, Liu SM, Li YF et al (2014) Effects of sub-chronic aluminum chloride exposure on rat ovaries. *Life Sci* 100(1):61–66
- Rawi SM, Seif Al Nassr FM (2015) Zinc sulphate and vitamin E alleviate reproductive toxicity caused by aluminium sulphate in male albino rats. *Toxicol Ind Health* 31(3):221–234
- Zhu YZ, Sun H, Fu Y, Wang J, Song M, Li M et al (2014) Effects of sub-chronic aluminum chloride on spermatogenesis and testicular enzymatic activity in male rats. *Life Sci* 102:36–40
- Li X, Huang W, Gu J, Du X, Lei L, Yuan X et al (2015) SREBP-1c overactivates ROS-mediated hepatic NF- κ B inflammatory pathway in dairy cows with fatty liver. *Cell Signal* 27(10):2099–2109
- Sun X, Yuan X, Chen L, Wang T, Wang Z, Sun G et al (2017) Histamine induces bovine rumen epithelial cell inflammatory response via NF- κ B pathway. *Cell Physiol Biochem* 42(3):1109–1119
- Sun H, Hu C, Jia L, Zhu Y, Zhao H, Shao B et al (2011) Effects of aluminum exposure on serum sex hormones and androgen receptor expression in male rats. *Biol Trace Elem Res* 144(1–3):1050–1058
- Kopáček J, Hejzlar J, Kana J, Norton SA, Porcal P, Turek J (2009) Trends in aluminium export from a mountainous area to surface waters, from deglaciation to the recent: effects of vegetation and soil development, atmospheric acidification, and nitrogen-saturation. *J Inorg Biochem* 103:1439–1448
- Speit G, Merckens O (2002) Evaluation of mutagenic effects of formaldehyde in vitro: detection of cross-links and mutat in mouse lymphoma cells. *Mutagenesis* 17:183–187
- Zaman K, Zaman W, Siddique H (1993) Hematological and enzymatic results of aluminum intoxication in rats. *Comp Biochem Physiol C* 105(1):73–76
- Silva VS, Gonçalves PP (2003) The inhibitory effect of aluminium on the ($\text{Na}^+\text{/K}^+$)ATPase activity of rat brain cortex synaptosomes. *J Inorg Biochem* 97(1):143–150

22. Ohyashiki T, Karino T, Suzuki S, Matsui K (1996) Effect of aluminum ion on Fe(2+)-induced lipid peroxidation in phospholipid liposomes under acidic conditions. *J Biochem* 120(5):895–900
23. Suwalsky M, Norris B, Villena F, Cuevas F, Sotomayor P, Zatta P (2004) Aluminum fluoride affects the structure and functions of cell membranes. *Food Chem Toxicol* 42(6):925–933
24. Mundy WR, Kodavanti PR, Dulchinos VF, Tilson HA (1994) Aluminum alters calcium transport in plasma membrane and endoplasmic reticulum from rat brain. *J Biochem Toxicol* 9(1):17–23
25. Garrosa M, Llanes F, Gayoso MJ (2011) Histopathological changes in gerbil liver and kidney after aluminum subchronic intoxication. *Histol Histopathol* 26(7):883–892
26. Nichol BE, Oliveira LA, Glass A, Siddiqi MY (1993) The effects of aluminum on the influx of calcium, potassium, ammonium, nitrate, and phosphate in an aluminum-sensitive cultivar of barley (*Hordeum Vulgare* L.) *Plant Physiol* 101(4):1263–1266
27. Gandolfi L, Stella MP, Zambenedetti P, Zatta P (1998) Aluminum alters intracellular calcium homeostasis in vitro. *Biochim Biophys Acta* 1406(3):315–320
28. Chaitanya TV, Mallipedi K, Bondili JS, Nayak P (2012) Effect of aluminum exposure on superoxide and peroxide handling capacities by liver, kidney, testis and temporal cortex in rat. *Indian J Biochem Biophys* 49(5):395–398
29. Mailloux RJ, Hamel R, Appanna VD (2006) Aluminum toxicity elicits a dysfunctional TCA cycle and succinate accumulation in hepatocytes. *J Biochem Mol Toxicol* 20(4):198–208
30. Kajikawa M, Fujimoto S, Tsuura Y, Mukai E, Takeda T, Hamamoto Y et al (2002) Ouabain suppresses glucose-induced mitochondrial ATP production and insulin release by generating reactive oxygen species in pancreatic islets. *Diabetes* 51(8):2522–2529
31. Guo CH, Hsu GS, Chuang CJ, Chen PC (2009) Aluminum accumulation induced testicular oxidative stress and altered selenium metabolism in mice. *Environ Toxicol Pharmacol* 27(2):176–181
32. Maghraoui S, Clichici S, Ayadi A, Login C, Moldovan R, Daicoviciu D (2014) Oxidative stress in blood and testicle of rat following intraperitoneal administration of aluminum and indium. *Acta Physiol Hung* 101(1):47–58