

Aqueous Synthesis and Concentration-Dependent Dermal Toxicity of TiO₂ Nanoparticles in Wistar Rats

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Abstract A number of dermal toxicological studies using TiO₂ nanoparticles exist which are based on the study of various animal models like mice, rabbits etc. However, a well-defined study is lacking on the dermal toxic effects of TiO₂ nanoparticles on rats, which are the appropriate model for systemic absorption study of nanoparticles. Furthermore, toxicity of TiO₂ nanoparticles varies widely depending upon the size, concentration, crystallinity, synthesis method etc. This study was conducted to synthesize TiO₂ nanoparticles of different sizes (~15 to ~30 nm) by aqueous method, thereby evaluating the concentration-dependent toxicological effects of the ~20-nm sized nanoparticles on Wistar rats. Characterization of the particles was done by transmission electron microscope, dynamic light scattering instrument, X-ray diffractometer, and ultraviolet spectrophotometer. The toxicity study was conducted for 14 days (acute), and it is observed that TiO₂ nanoparticles (~20 nm) at a concentration of 42 mg/kg, when applied topically showed toxicity on rat skin at the biochemical level. However, the histopathological studies did not show any observable effects at tissue level. Our data suggest that well-crystallized spherical-shaped ~20 nm anatase TiO₂ nanoparticles synthesized in aqueous medium can induce concentration-dependent biochemical alteration in rat skin during short-term exposure.

Keywords TiO₂ nanoparticles · Acute toxicity · Topical application · Wistar rat · Characterization

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Introduction

Titanium dioxide (TiO₂) is one of the nanomaterials that have attracted a great attention due to its unique properties. Titanium dioxide is classified as being physiologically inert in both humans and animals [1], hence it has been widely used in many ways; as an additive, including as a white pigment in paint, as food colorant, in sunscreens and in cosmetic creams. It is also a well-known photocatalyst [2, 3], due to which it is applied in environment application and in waste water as a disinfectant [4]. Recently, TiO₂ was used as a photosensitizer for photodynamic therapy of endobronchial and esophageal cancers [5]. Despite these bright outlooks, there is an increasing concern that nanosized TiO₂ particles may adversely affect the health of humans and his environment. It is expected that the minute size of nanomaterials which gives them unusual properties of strength and reactivity, the same property would give them the unpredicted properties of toxicity also [6]. The toxicological concern is due to the distinct properties of nanoparticles, such as small size, high number per given mass and large specific surface area. It has been reported that the biological responses to nanoparticles may exceed those elicited by micron-sized particles [7, 8]. The physical properties of these nanomaterials would allow them to catalyze a number of bio-molecular interactions, which potentially could produce adverse toxicological effects.

Many *in vivo* studies on generation of inflammatory response due to accumulation of nanosized TiO₂ particles in the liver, kidney, spleen, lung, heart, and brain has been reported [9–16]. For instance, intraperitoneal administration of TiO₂ nanoparticles of smaller size (~5 nm) can easily enter mouse liver cells and bond to liver DNA, at higher doses [17]. Intra-gastric administration of TiO₂ nanoparticles for 30 consecutive days could lead to liver function damage in mice [18] and induces liver hepatitis when administered at a dose of 10 and 50 mg/kg body weight (BW) [19]. Studies had shown that by intra-tracheal instillation of TiO₂ nanoparticles in rats, pulmonary inflammation and cytotoxicity is observed [20–22]. TiO₂ nanoparticles with oral gavages increased the activity of lactate dehydrogenase causing hepatocyte necrosis in mice [23]. Intraperitoneal injection of various doses of TiO₂ nanoparticles can lead to its accumulation in the mouse spleen [9]. Higher doses of anatase TiO₂ by intra-gastric administration exert toxicity through oxidative stress [24].

A few studies have investigated dermal toxicity and systemic exposure of nanoscale TiO₂ particles by topical application on rabbits and mice [25–30]. However, there are no reported toxicity studies on TiO₂ nanoparticles when rat skin is used as a portal entry for whole body exposure. The ability of TiO₂ nanomaterials to traverse the skin is a primary determinant of its dermato-toxic potential. That is, TiO₂ nanoparticles must penetrate the stratum corneum in order to exert toxicity in lower cell layers. The quantitative prediction of toxicity is carried out in the current research and is studied by biochemical alteration and histological changes in the rat skin. This research aims to synthesize TiO₂ nanoparticles of different sizes (~15 to ~30 nm) by aqueous method, characterize its property and assessing its concentration based toxicological properties by topical application on Wistar rats.

Materials and Methods

Chemicals

Titanium tetrachloride (TiCl₄; 1 M in Toluene) was purchased from Spectrochem (India). Tri-sodium citrate 2-hydrate GR (>99% purity) was purchased from Merck (India). Acetic acid glacial extra-pure was purchased from SD-Fine Chem Ltd. (India). Reduced

glutathione, 1-chloro-2,4-dinitrobenzene (CDNB), thiobarbituric acid, and bovine serum albumin (BSA) were obtained from Sigma-Aldrich, USA. Trichloroacetic acid was purchased from Central Drug House (CDH), India. Biological Assay kit for the estimation of lactate dehydrogenase (LDH) was purchased from Reckon (India). All other chemicals were of analytical grade. Milli-Q water was used for the whole experiment.

Synthesis of TiO₂ Nanoparticles

Titanium dioxide nanoparticles of different sizes were prepared by aqueous method in the following manner. Tri-sodium citrate at varying concentration (0.01, 0.05, 0.08, and 0.1 M, respectively) was dissolved in 100 ml of Milli-Q water. The solution was vortexed thoroughly and then sonicated for 5 min. The solution was then kept under constant stirring under magnetic stirrer. To this solution, 500 μ L of TiCl₄ was added dropwise under vigorous stirring. The reaction was performed at thermostatically controlled temperature and stirring was continued for 24 h. Since the particle size depends on the concentration of capping agent and the metal ion, we prepared TiO₂ nanoparticles of a broad range of size (~15, ~20, ~25, and ~30 nm) following the above protocol. The prepared particles were studied further for characterization.

Characterization of TiO₂ Nanoparticles

Transmission Electron Microscopy

The size of the nanoparticles was measured by Transmission Electron Microscope (TEM; Philips Morgagni). Measurements were made using computerized image analyzer and the average size of nanoparticles was noted.

Dynamic Light Scattering

Dynamic light scattering (photon correlation spectroscopy) is a technique used to determine the size distribution profile of small particles in suspension. The technique was performed on "Malvern ZS" instrument.

X-ray Diffractometry

For determining the crystallographic nature of the sample, X-ray diffraction (XRD) measurements were performed on PANalytical instrument, using powdered samples of the TiO₂ nanoparticles.

Ultraviolet Spectroscopy

All the spectrophotometer studies were done using a Spectroscan 80 DV spectrophotometer.

Animal Model

Both male and female rats of Wistar strain, approximately 8 weeks old (weights in the range of 150–200 g) were used in this study. The rats were obtained from the Central Animal House facility of Jamia Hamdard, New Delhi and were housed in a well-ventilated room at $\pm 22^{\circ}\text{C}$, under a 12-h light/dark cycle. Research on the experimental animals was conducted

in accordance with the internationally accepted principles for laboratory animal's use and care as found in the guidelines laid down by the Indian Ethical Committee, Committee for the Purpose of Control and Supervision of Experiments on Animals. The rats, in groups of five, were kept hygienically in separate polypropylene cages. They were acclimatized for 1 week before the start of the study and were allowed free access to standard laboratory feed (Hindustan Lever Ltd, India) and water ad libitum. The dorsal portions of the rat's skin were shaven with an electric clipper (Oster A2) followed by the application of hair removing cream (Anne French, Geoffrey Manners, India) 2 days before treatment. Excess cream was washed off with cotton sorbs dipped in lukewarm water. Only rats that showed no signs of hair regrowth were included in the experiment.

Treatment Regimen

Effects of TiO₂ nanoparticles (~20 nm) on rat's skin was studied by randomly allocating 25 Wistar rats (both male and female) into five groups, each having five rats. All animals of the experimental study received topical application of nanoparticles for a period of 14 consecutive days (acute toxicity study).

At the end of the experiment, animals of all the groups were sacrificed under mild anesthesia. Blood was taken for various serological parameters. Skin from dorsal area was removed and cleaned off extraneous tissue. A piece of skin was preserved in 10% neutral buffered formalin for histopathological investigation. Skin homogenates were prepared in chilled phosphate buffer (0.1 M, pH 7.4) using polytron homogenizer and then filtered through muslin cloth. The homogenized tissue was centrifuged at 10,500 rpm for 30 min at 4°C to obtain post-mitochondrial supernatant (PMS). PMS was used in various biochemical measurements as detailed in [Biochemical assays](#).

Biochemical Assays

Glutathione-S-transferase (GST) activity was measured by the method of Habig et al [31], and expressed as nanomoles of CDNB conjugates formed per minute per milligram of protein. Catalase (CAT) activity was assayed by the method of Claiborne et al [32] and expressed as nanomoles of H₂O₂ consumed per minute per milligrams of protein. Superoxide dismutase (SOD) was examined by a modified method of Misra and Fridovich [33], and the activity was expressed in micromolars of epinephrine oxidized per minute per milligram of protein. Estimation of lipid peroxidation was done according to the method of Wright et al. [34]. The results were expressed as nanomoles of Malon dialdehyde (MDA) formed per hour per gram of tissue at 37°C using a molar extinction coefficient of $1.56 \times 10^5 \text{M}^{-1} \text{cm}^{-1}$. LDH activity was estimated in serum by the standard protocol method mentioned in the biological assay kit. Protein estimation in all samples was done using the method of Lowry et al. [35] using BSA as standard.

Histopathological Investigation

The skin samples were processed with haematoxylin and eosin stain for gross tissue histo-architecture evaluation. The formalin fixed skin samples were dehydrated with graded ethanol (Merck) and embedded in paraffin (Hi-Media Labs, India) after rinsing with distilled water. The samples were cut by microtome at 5-mm thick and mounted on glass slides. The slides were studied using "Olympus DP71 Biological" microscope.

Statistical Analysis

The level of significance between different groups is based on analysis of variance test followed by Dunnett's *t* test.

Results and Discussions

Characterization of Nano-TiO₂ Samples

Transmission Electron Microscope

TEM images of TiO₂ nanoparticles synthesized at varying concentrations (0.01, 0.05, 0.08, and 0.1 M, respectively) are shown in Fig. 1a–d. Well-defined nanoscale size distribution of highly monodisperse TiO₂ nanoparticles with spherical shape was obtained.

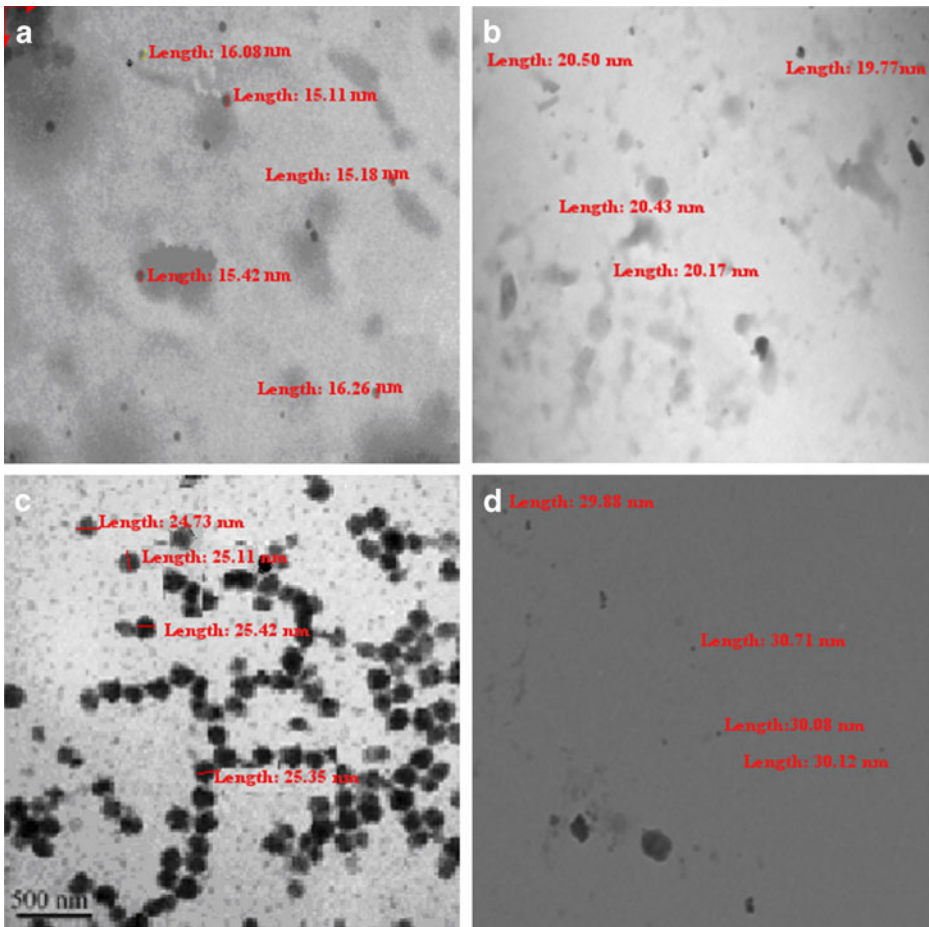


Fig. 1 a–d TEM images of different sizes of TiO₂ nanoparticles with spherical shape. **a** ~15, **b** ~20, **c** ~25, and **d** ~30 nm

Dynamic Light Scattering

The size distribution of TiO₂ particles (0.05 M) suspended in water was taken by dynamic light scattering (DLS) and shown in Fig. 2. TiO₂ nanoparticles have a negative surface charge, thus stabilizing the suspensions via repulsive forces. The figure reveals that the size of synthesized TiO₂ is less than 50 nm.

X-Ray Diffraction

XRD addresses the structural information of a large portion of nanosized sample. Sharp peaks at 101 of the nanosized TiO₂ particles (0.05 M) were observed, which indicate the crystalline nature of the sample. All prominent peaks show the tetragonal crystal structure of anatase-TiO₂ (Fig. 3). Calculation by the Scherrer's equation showed that the average crystal size of synthesized TiO₂ was ~20 nm.

UV Spectroscopy

TiO₂ nanoparticles exhibit broad absorption bands in the ultraviolet visible range. These are due to the excitation of plasma resonance or interband transition and are a characteristic property of the metallic nature of the nanosized TiO₂ particle. Nanosized spherical-shaped TiO₂ particles exhibit a surface plasmon peak at around 230 nm, characterizing the synthesized samples to be anatase. In Fig. 4a–d represent the respective peaks for ~15, ~20, ~25, and ~30 nm, respectively.

Fig. 2 Size distribution of ~20-nm TiO₂ particle

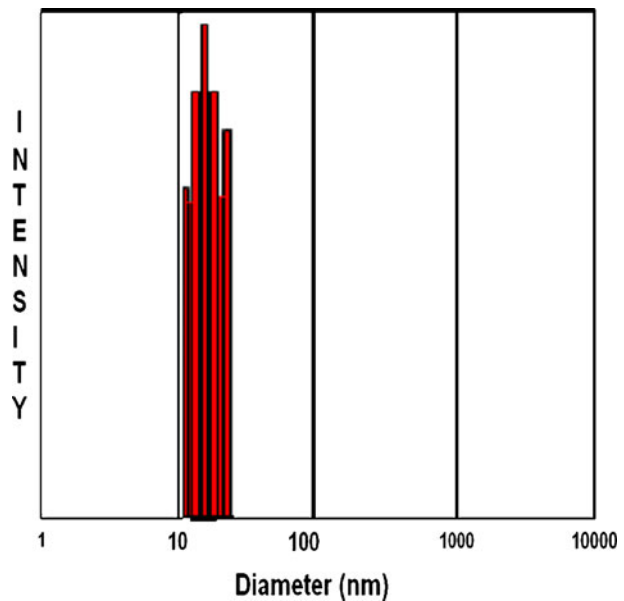
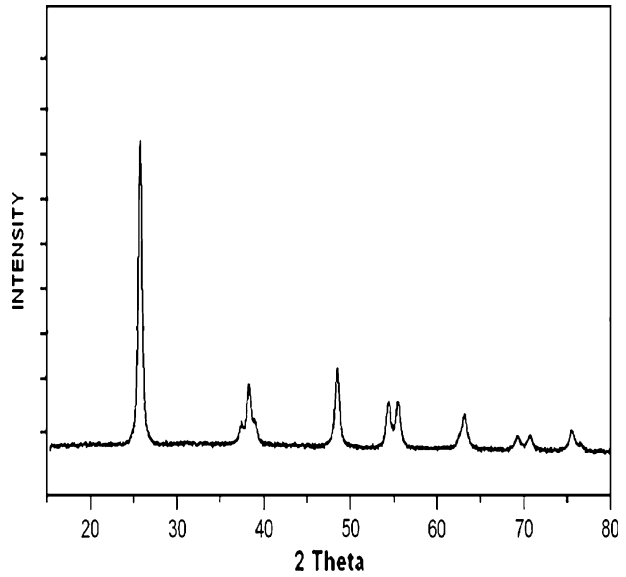


Fig. 3 Characteristic XRD pattern of ~20-nm powdered TiO₂ nanoparticles



Toxicological Study

Biochemical Estimation

The effects of topical treatment of ~20 nm TiO₂ nanoparticles on the skin of rats of different groups at different concentration are studied and shown in the figures below. Group I is the control group while the remaining groups II, III, IV, and V are the nanoparticles-treated group with a concentration level of 14, 28, 42, and 56 mg/kg BW, respectively.

Fig. 4 UV-visible spectra of TiO₂ nanoparticles (~15 to ~30 nm)

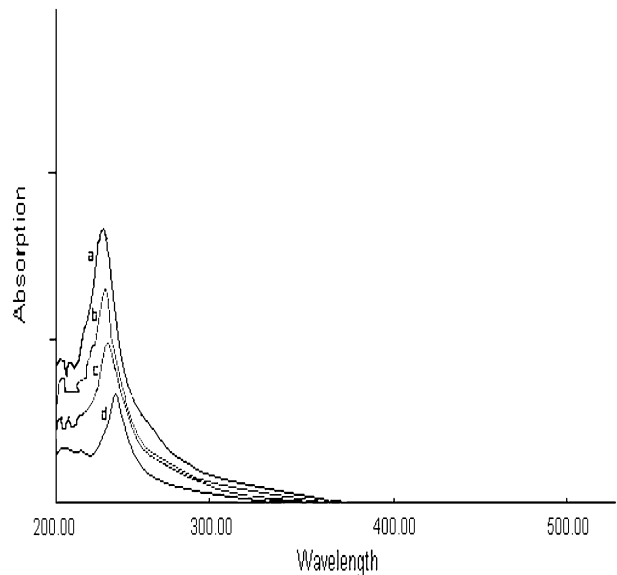
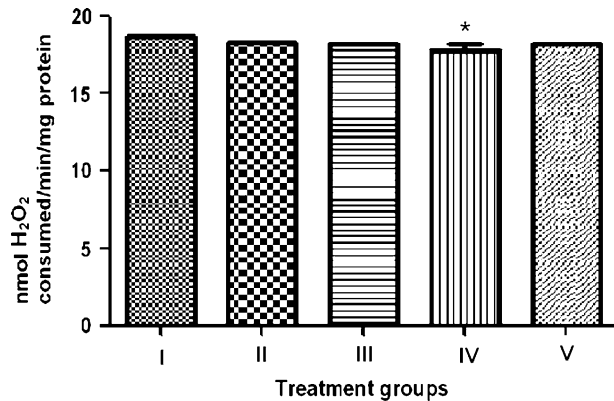


Fig. 5 Study of the effect of TiO₂-induced cutaneous catalase activity. Results represent mean \pm SE ($n=5$) measured as nanomoles H₂O₂ consumed per minute per milligram of protein. * $P<0.05$, significant when compared with acetone-treated control group



The result depicts the induced oxidative stress and alteration in the rat skin on exposure to concentration of 42 mg/kg BW of TiO₂ nanoparticles through topical application. There is a significant depletion in the activity of the antioxidant enzyme, catalase (Fig. 5), and SOD (Fig. 6) in group IV ($p<0.05$) when compared with acetone-treated control (group I) animals. No significant change was observed in the other groups.

There is also depletion in the level of an important phase II enzyme, glutathione-*S*-transferase ($p<0.05$) as compared with the control-treated group (Fig. 7). Groups II, III, and V do not show any significant alterations and the results are close to normal.

Group IV animals showed slight significant enhancement in levels of LDH (Fig. 8) and MDA (Fig. 9) formation ($p<0.05$), when compared with group I (control group). No significant difference was observed in other treated groups (groups II, III, and V).

Histopathological Findings

When treated with ~20 nm TiO₂ nanoparticles, the rat skin did not show any marked alterations. The histo-architecture of rat skin of group I rats showed normal histology of well differentiated dermal layer and thin wavy epidermis with basal levels of neutrophils. Among the treatment groups, group IV showed very slight changes in cutaneous architectures as compared with control group which are not significantly visible. All the other treatment groups (groups II, III, and V) do not show any significant differences when compared with the control-treated group (group I)

Fig. 6 Study of the effect of TiO₂-induced cutaneous SOD activity. Results represent mean \pm SE ($n=5$) measured as micromolars of epinephrine oxidized per minute per milligram of protein. * $P<0.05$, significant when compared with acetone-treated control group

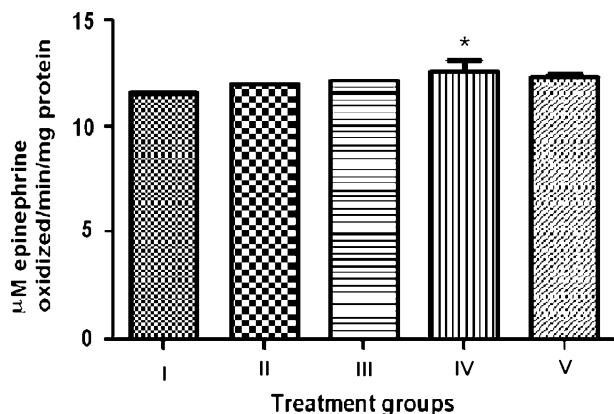
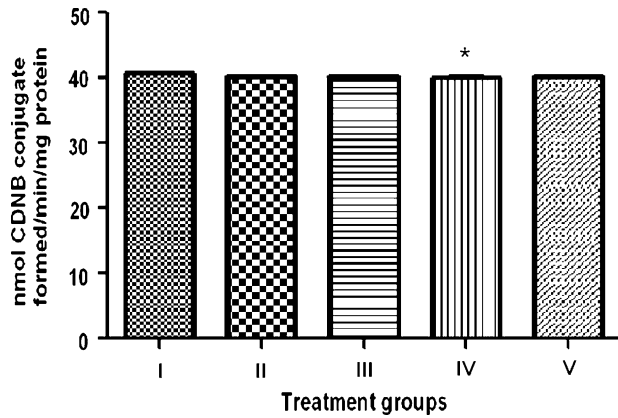


Fig. 7 Study of the effect of TiO₂-induced cutaneous GST activity. Results represent mean \pm SE ($n=5$) measured as nanomoles of 1-chloro-2,4-dinitrobenzene (CDNB) conjugates formed per minute per milligram of protein. * $P<0.05$, significant when compared with acetone-treated control group



Discussion

There are many advantages in the dedicated synthesis of nanoparticles for toxicity studies. A good control of particle-size range within each batch is also important if reliable links between toxicity and size are to be made. Our synthesis focused primarily on the methodology of preparing TiO₂ nanoparticles by aqueous method under controlled condition. Since particle size depends on the concentration, we synthesized nanosized TiO₂ of different sizes ranging from approximately ~15 to ~30 nm using the aqueous method. The synthesized particles were further characterized by TEM, XRD, DLS, and UV spectrophotometer. We then followed this synthesis work with in vivo toxicological studies. Because of their diminutive size, nanoparticles carry several inherent properties. Firstly, ultrafine particles have larger surface areas per unit mass. Secondly, particle toxicity is determined by surface reactivity. Thus, given their structure, nanoparticles exhibit greater harm compared with larger particles because of their proportionally increased surface area. The large surface area also provides a distinctive interface for catalytic reactions of surface-located mediators with biological targets such as proteins.

Since the skin is an important interface between man and his environment, it is a significant portal of entry of hazardous agents and a vulnerable target organ system. The skin is endowed with a versatile group of adaptive and defensive mechanism. The

Fig. 8 Study of the effect of TiO₂-induced cutaneous LDH activity. Results represent mean \pm SE ($n=5$) measured as nanomoles NADH oxidized per minute per milligram of protein. * $P<0.05$, significant when compared with acetone-treated control group

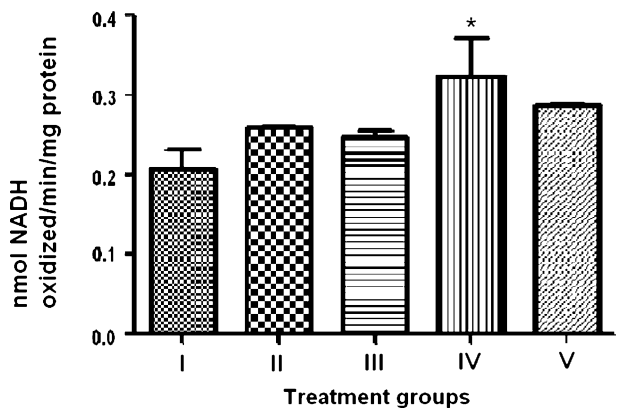
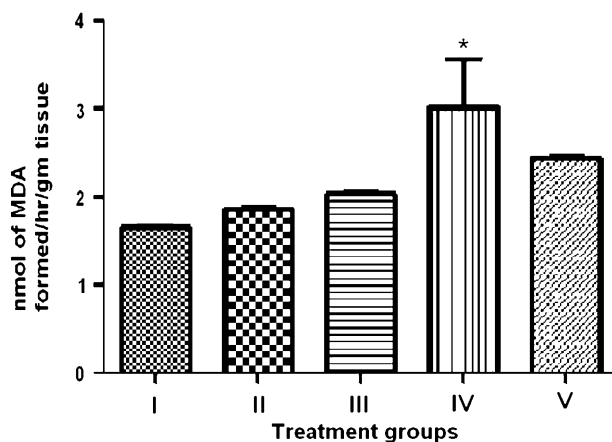


Fig. 9 Study of the effect of TiO₂-induced cutaneous LPO. Results represent mean \pm SE ($n=5$) measured as nanomoles Malon dialdehyde (MDA) formed per hour per gram of tissue. * $P<0.05$, significant when compared with acetone-treated control group



penetration of materials into the stratum corneum is limited by molecular size. The intercellular space between the cells composing the stratum corneum measures approximately 100 nm³ and may be widened with topical application of various products [36]. This raises the question of whether the particles used in TiO₂-based sunscreens have the potential to penetrate the stratum corneum. Studies reporting the dermal toxicity concluded that TiO₂ do not reach the viable cells [6]. These findings are based on the study on various animal models like mice, rabbits etc.; and that too for a period of 3 days maximum. However, a well-defined documentary report is lacking in the dermal toxicity of TiO₂ nanoparticles on rats, which are the appropriate model for system absorption study of nanoparticles [37].

Size, crystal structure, and surface chemistry (such as coating) are among the factors that influence the effects of nano-TiO₂ particles. Other physicochemical properties, such as shape [38], manufacturing process, doping, and purity (or impurities) could also play a role in the toxicity of nano-TiO₂, but such information is usually not reported in toxicological studies. Physicochemical properties, experimental conditions, and the immediate environment can all influence the ecological and health effects of nano-TiO₂ particles. Another important parameter of nanoparticles' toxicity study is the purity of water used for synthesis. It affects the degree of aggregation, which in turn may affect exposure-dose and toxicity. The degree of aggregation generally increases with the presence of salt, minerals, and organic matter in water [39, 40]. Although the influences of media and vehicle and dispersion methods on particle aggregation and distribution have been reported, information on these influences on health effects is very scarce [41].

In our present study, we formulated the TiO₂ nanoparticles via aqueous route and further studied to reveal its characteristic properties for in vivo toxicological studies. We investigated the effect of TiO₂ on a number of biochemical parameters mainly SOD, CAT, GST, LPO, and LDH. Enzymes SOD and CAT remove reactive oxygen species (ROS) generated by the free radicals. SOD converts O₂⁻ into H₂O₂ and O₂ while CAT reduces H₂O₂ into H₂O and O₂. Thus SOD and CAT prevent further ROS generation in cells. GST family of phase II detoxification enzymes catalyzes the conjugation of glutathione to a wide variety of endogenous and exogenous electrophilic compounds, such as therapeutic drugs, environmental toxins, and products of oxidative stress. Lipid peroxidation is the oxidative degradation of lipids in which free radicals steal electrons from the lipids in cell membranes, resulting in cell damage. LDH is an enzyme present in

body tissues that get released during tissue damage. With the experimental setting of the current study, we aim to address the current concern that once exposed to TiO₂ nanoparticles for consecutive days; it may become systemically available and cause toxic effects. This can be seen from the results of the biochemical assay carried out on rat skin exposed to nanosized TiO₂ particles. There was a significant depletion in the activities of catalase, SOD, and GST with a concomitant increase in the LPO and LDH activity in group IV, at a concentration of 42 mg/kg BW. The toxicity level at a concentration of 42 mg/kg BW vis-à-vis control group is observed to be significant as compared with other concentrations. The decrease in level of toxicity at 56 mg/kg BW is attributed to the coagulation of the nanosized TiO₂ particles as the concentration increased.

The most discussed aspect at present is the induction of reactive oxygen species due to the chemical properties of TiO₂. Overall results of this study indicate that rats exposed to TiO₂ nanoparticles by topical route showed significant health effects at the dose level of 42 mg/kg BW, even though no marked histological alteration was seen. It is evident from our study that at lower concentration of TiO₂ nanoparticles, there was no noticeable level of toxicity. This study will provide valuable information regarding the safety evaluation of TiO₂ nanoparticles as the synthesis protocol and the species studied (rat model) to test the toxicological properties of TiO₂ nanoparticles is different from the previously reported studies [37].

The science of toxicology has always provided the foundation for understanding the interactions between chemistry and biology. Consequently, the unique physical-chemical characteristics of engineered nanomaterials that lead to their distinctive properties will likely contribute to the hazards associated with these materials. Therefore, the approach to addressing the safety of these materials will best be conducted via multidisciplinary teams.

Conclusions

The inferences drawn from this study highlight the toxicity parameters in experimental rat model after topical application of nanosized TiO₂ nanoparticles of ~20 nm size at a defined concentration of 42 mg/kg, for 14 consecutive days. The study also confirms that nanoparticles' toxicity is due to the oxidant generation and the resultant oxidant stress to cells. Oxidative stress has been clearly shown to occur during TiO₂ dermal application on rats. Superoxide anions are generated which will lead to the formation of highly reactive species, hydroxyl radicals that attack proteins and lipid membranes causing cell damage or genetic alterations. LPO and SOD are therefore, the markers for assessing the extent of damage in cells. With the application of TiO₂ nanoparticles, the level of the cellular antioxidant enzyme and glutathione is depleted. Additionally, the levels of MDA increased which indicated increased peroxidation of lipids. The histopathology study showed only slight changes in cutaneous architectures. The data suggest that well-crystallized spherical-shaped anatase TiO₂ synthesized in aqueous medium induced concentration-dependent biochemical alteration in rat model with no marked alteration on the histology of skin tissue.

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Conflicts of Interest Statement None.

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