

Concentrations of Selected Essential and Non-essential Elements in Blood of Harbor Seal (*Phoca vitulina*) Pups of the German North Sea

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Abstract This study on harbor seal (*Phoca vitulina*) pups of the North Sea evaluated concentrations of 14 essential and non-essential elements (Al, As, Be, Ca, Cd, Cr, Fe, Mn, Mo, Ni, Pb, Se, Sn, and Zn) in whole blood samples. The essential elements are analyzed to give references for health status determinations of pups. The measurement of classic toxic metals, like Pb or Cd, and other elements that may be in toxic concentrations in blood, is important due to their influence on health, particularly on the immune system. Blood samples of six seal pups found on the German Wadden Sea coast of Schleswig-Holstein in 2004 and transported to the Seal Centre Friedrichskoog, Germany were collected. The blood sampling was performed three times, immediately after collection of the newborns, after 1.5 months, and after 2 months before their release back into the wild. Inductively coupled plasma mass spectrometry and total reflection X-ray fluorescence spectrometry were used to determine the element concentrations. We found higher concentrations of Al, As, Fe, Mo, and Zn in blood samples of newborns compared to samples collected later, probably due to transplacental and lactational transfer from mother to fetus. Furthermore, there is a high need for, in particular, Fe and Zn in the developing organism, which may cause reduced values after some month. In contrast, the concentrations of Be, Cd, Ca, Cr, Pb, Mn, Ni, Se, and Sn, which were low in newborns and increased during the study, may be due to the fish fed to infant pups. Compared to free-ranging adults, in pups, the concentrations of Al, Ni, and Pb were higher in contrast to lower concentrations of As, Mn, and Mo. This case study is the first report on element levels in harbor seal pups of the North Sea.

Keywords Harbor seal · Pups · Elements · Metals · Blood · Pollution · North Sea

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Introduction

The harbor seal, *Phoca vitulina*, is one important domestic marine mammal in the Wadden Sea and is considered as an indicator for changes in the ecosystem. Therefore, the “Seal Agreement” has been adopted, which include the monitoring of pollution and investigations on the effects of pollutants, as well as the assessment of the health status. Nevertheless, there is a lack of studies on seal pups. A useful way is the investigation of the newborn and infant pups living in the Seal Centre Friedrichskoog, Germany to get baseline information to health parameters of pups [1, 2].

As a monitoring tool, the multi-element measurement of elements in blood of animals provides both information to essential elements and to the levels of toxic elements. However, only few studies have performed a multi-element analysis in blood of marine mammals [3–6], in particular of the pinnipeds of the North Sea [7–10].

Since elements like Fe, Zn, or Ca have important functions in the body, i.e., Fe as component of cytochromes and oxygen-binding molecules, Zn as components of more than 70 metalloenzymes, or Ca as important element for building of bones and teeth, for blood clotting, muscle contraction, or maintenance of cell membranes, these essential elements were determined in blood serum during medical investigations, such as on marine mammals [11–17]. Some studies focused on Se due to the strong relationship to mercury and the importance of the element in the detoxification process. Most of them measured Se in tissues such as liver or kidneys [18, 19] but only few in blood [20]. Toxic elements like Cd were investigated in blood in several studies on whales and pinnipeds [20–24]. Nevertheless, all metals are toxic if they are present in living organisms in sufficiently high quantity, and there is a lack of multi-element investigations of blood in marine mammals.

Metal exposure for seal pups may result from the transplacental and lactational transfer from mother to fetus and later through contaminated prey [25–27]. Metal contamination is believed to adversely affect marine mammal health [28–30]. A relationship between stillbirths and nickel concentration in the hair of ringed seal (*Pus hispida saimensis*) pups from Finland was found [31]. Some marine mammal studies surmise that greater metal concentrations result in lesser resistance to diseases [32–34]. These relationships are substantiated by metal contamination affecting the immunocompetence and disrupt the immune homeostasis. Immunomodulation by metals in marine mammals are reported in several studies [9, 10, 35–39]. An *in vitro* study on lymphocytes of seal pups suggests that there immune cells are particularly susceptible to the toxic effects of metals during the newborn period and that this susceptibility decreases as the animals age [1]. Metals found in blood of the seal pups may have a potential influence on the immune status of the animals.

This pilot study is the first investigation that measured element levels in harbor seal pups from the German North Sea coast.

Materials and Methods

Animals

We analyzed blood samples of six harbor seal pups found on the German Wadden Sea coast of Schleswig-Holstein in 2004 (Table 1). The pups were transported to the Seal Centre Friedrichskoog, Germany and were examined by a veterinarian. This investigation included

Table 1 Harbour Seal (*Phoca vitulina*) Pups Analyzed in this Study

Seal	Arrival in the Seal Centre	Weight (kg)	Length (cm)	Sex	WBC (*10 ³ mm ⁻³)	Blood sampling	Age (days)
01	20.06.04	8.9	58	f	10	21.06.04	8
						03.08.04	51
						26.08.04	74
02	20.06.04	8.6	57	f	9	21.06.04	8
						03.08.04	51
						26.08.04	74
03	20.06.04	10.5	59	f	9	21.06.04	8
						03.08.04	51
						26.08.04	74
04	21.06.04	8.9	57	f	7	21.06.04	7
						03.08.04	50
						26.08.04	73
05	21.06.04	8.7	54	m	10	21.06.04	7
						03.08.04	50
						26.08.04	73
06	21.06.04	10.9	65	m	8	21.06.04	10
						03.08.04	53
						26.08.04	76

f Female, *m* male, *WBC* white blood cells

a full external examination, auscultation of heart and lung, and measurements of body temperature, mass, and length. Information on the intake of breast milk was not available. Blood samples were obtained from the epidural vertebral vein. The numbers of white blood cells were investigated (Dr. Driver, Kleintierpraxis, Reinsbuettel, Germany) and were in the normal range for harbor seals with $6\text{--}12 \times 10^3 \text{ mm}^{-3}$ [40]. The first blood sampling was performed in June immediately after their arrival at the Seal Centre (age of pups, 7–10 days). The second sampling took place in the beginning of August when these animals were between 50 and 53 days old. The seals were sampled a third time just before their release back into the sea after gaining a weight of 25 kg (age of pups, 73–76 days).

Multi-element Analysis

Concentrations of 14 elements (Al, As, Be, Ca, Cd, Cr, Fe, Mn, Mo, Ni, Pb, Se, Sn, and Zn) were determined in whole blood samples. Whole blood samples were collected in special lithium heparin monovettes for metal analysis (Sarstedt, Nümbrecht, Germany) and stored at -20°C . For multi-element determination, a microwave digestion system (MarsXpress, CEM GmbH, Kamp-Lintfort, Germany) was used. Five hundred microliters of whole blood was pipetted into perfluoroalkoxy vessels. Two milliliters sub-boiled nitric acid, 1 ml of hydrogen peroxide, and 50 μl internal standard (1 mg l^{-1} of yttrium, Merck, Darmstadt, Germany) were added and the vessels heated in a three-step program up to 180°C .

Concentrations of 14 trace elements were determined with two different methods. Al, Be, Cd, Cr, Mn, Mo, Ni, Pb, and Sn were analyzed using an inductively coupled plasma–mass spectrometer (ICP-MS) with a collision cell (Agilent 7500c ICP-MS, Agilent Technologies, Tokyo, Japan). Matrix effects and instrumental drift of the ICP-MS were

corrected by using yttrium as an internal standard. For calculation, external calibration was carried out with diluted standard solutions (Merck). The standard mode without collision gas was used for Al, Be, Pb, and Sn. For the other elements, better results were obtained with helium as the collision gas (flow rate, 3.0 ml min⁻¹).

Measurements of As, Ca, Fe, Se, and Zn were performed by total X-ray fluorescence spectrometry (TXRF; Atomika TXRF 8030 C, FEI Company, Oberschleissheim, Germany). Digested samples (20 µl) were pipetted onto the sample carrier and evaporated to dryness. The Mo K α excitation was selected for the detection of the elements. Yttrium as internal standard was used to calculate the concentrations.

For internal quality control, the reliability of the analytical procedures was checked with the human reference material Clin Check[®] Whole Blood Control Level II lot no. 932 (Recipe, Chemicals + Instruments, Munich, Germany) and animal reference material IAEA-A-13 (International Atomic Energy Agency, Vienna, Austria; Table 2).

Statistical Analysis

For the results of the six pups investigated three times, we used the Friedman tests in comparing the element concentrations in the whole blood (Table 3). All indicated *P* values are two-tailed. Statistical significance was designated as *P* ≤ 0.05.

Results

Two trends were found for elements in whole blood of harbor seal pups during the time of investigation. Lower concentrations of Be, Ca, Cd, Cr, Mn, Ni, Se, and Sn were measured directly after collection of the newborns compared to measurements in the following time when the animals were older (Fig. 1). All results are significant (Table 3). Although the results for Pb were not significant, for four of six pups, similar trends were found, and Pb therefore was included in Fig. 1.

Table 2 Concentrations of Elements (Mean Value, *n*=10) of the Reference Materials Clin Check (Human Whole Blood, Level II) Measured with ICP-MS and IAEA-A-13 (Animal Whole Blood) Measured with TXRF

Element	Certified mean value	Control range	Measured mean value (<i>n</i> =10)	±SD
ICP-MS measurement: ClinCheck, human whole blood, Level II, lyophilized (µg l ⁻¹)				
Cr	7.1	6.0–8.2	7.26	0.872
Mn	27	22–32	31.3	1.29
Co	4.7	3.7–5.7	5.06	0.251
Ni	8.3	6.4–10.2	8.24	1.06
Cd	4.3	3.5–5.1	3.42	0.210
Pb	304	258–350	272	11.3
TXRF measurement: IAEA-A-13, animal whole blood, freeze dried (mg kg ⁻¹)				
K	2,500	2,100–2,700	2,615	60
Ca	286	226–332	268	42
Fe	2,400	2,200–2,500	2,363	115
Cu	4.3	3.7–4.8	4.25	0.51
Zn	13	12–14	12.6	0.52
Se	0.24	0.15–0.31	0.25	0.02

Table 3 Statistical Analysis of Time-Dependent Differences Between Element Concentrations ($N=14$) in Whole Blood Samples Collected Three Times During 2 Months (Friedman Test, $k=3$, $n=6$)

Metal	F_r	P Value
Al	9.33	0.006*
As	6.33	0.052
Be	11.47	0.004*
Ca	9.33	0.006*
Cd	7.14	0.020*
Cr	12.00	<0.001*
Fe	8.33	0.012*
Mn	8.33	0.012*
Mo	2.33	0.430
Ni	9.33	0.006*
Pb	1.33	0.570
Se	9.33	0.006*
Sn	9.65	0.003*
Zn	8.33	0.012

* $P \leq 0.05$, significant differences

In contrast, higher concentrations of Al, Fe, and Zn were measured in the samples of the newborns compared to the samples collected when these animals were older (Fig. 2). All results are significant (Table 3). The results for As and Mo were not significant; nevertheless, similar decreasing trends were found, and the results of both elements are inserted in Fig. 2.

Compared to free-ranging adults of the North Sea, higher concentrations of Al, Ni, and Pb and lower concentrations of As, Mn, and Mo were found in the blood of the pups (Figs. 1 and 2).

Discussion

Blood is the medium for the transport of elements, including essential elements such as Ca, Fe, Mn, Se, or Zn, as well as several important contaminants such as As, Be, Cd, or Pb. In this study, the concentrations of 14 elements in blood of harbor seal pups were investigated during their first 2 months of life. Metals could be grouped into two classes: increasing and decreasing concentrations with age during juvenile stages.

The concentrations of the major element Ca and of the trace elements Be, Cd, Cr, Mn, Ni, Pb, Se, and Sn increased during the time of investigation. Young animals have an elevated Ca metabolism due to bone development and alteration processes, which cause elevated levels in blood. Similarly, increases of Ca concentrations in blood of harbor and gray seal pups during the first weeks of life were measured [13, 41]. Greater concentrations of Ca concentrations in serum were determined in harp seal pups compared to adult harp seals [42]. Furthermore, increasing concentrations of elements, e.g., of toxic metals like Be or Cd during the time of investigation, are probably caused by contaminated prey. In general, for marine mammals, the main intake of pollutants is due to contaminated prey [43]. The species fed to the harbor seal pups at the Seal Centre was herring (*Clupea harengus*) from the Baltic Sea (personal communication: Seal Centre Friedrichskoog), which was probably the source of the metals [44–46].

Higher concentrations of Al, As, Fe, Mo, and Zn were determined in the blood samples of the newborns compared to samples taken 2 months later. Such elevated concentrations of elements in blood of newborns were probably caused by transplacental and/or lactational

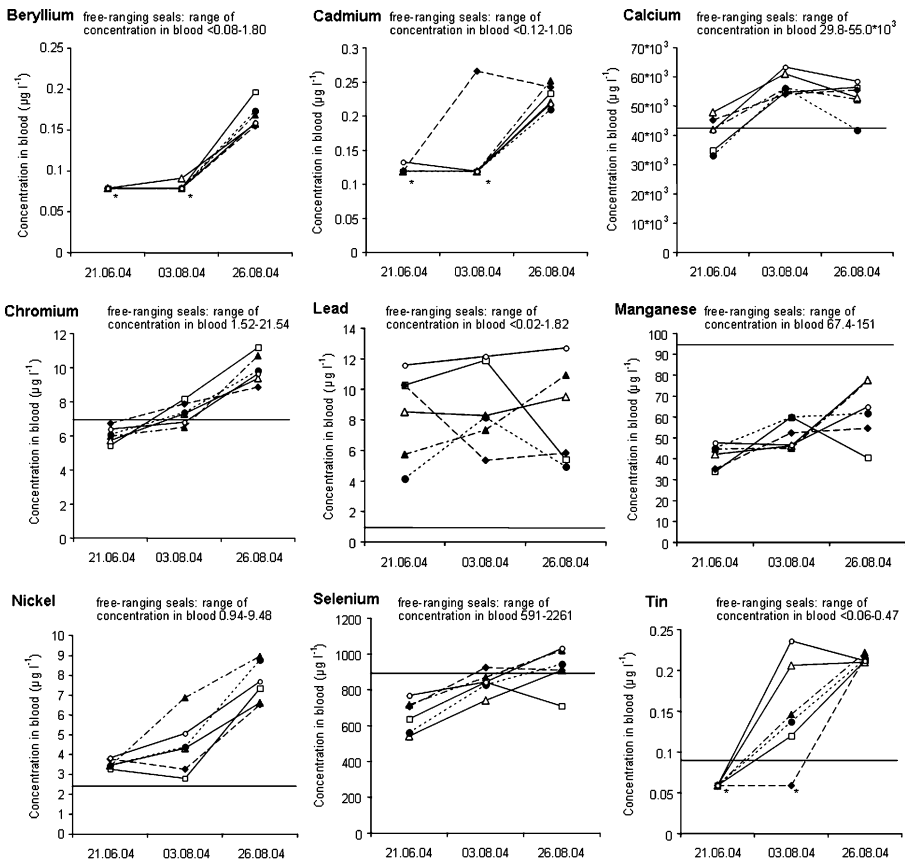


Fig. 1 Concentrations ($\mu\text{g l}^{-1}$) of Be, Cd, Ca, Cr, Pb, Mn, Ni, Se, and Sn in whole blood samples of six harbor seal pups (* values <math><\text{LOD}</math>) measured three times. Mean values (lines; Be and Cd, no mean values published) and ranges of concentrations in blood of free-ranging seals of the North Sea, Lorenzenplate [8]

transfers from mother to pup. A study on a nursing *Tursiops truncatus* female revealed that metal pollutants pass from the tissue of the female into the milk [47]. A transplacental transfer from mother to pup was described for different marine mammals and several pollutants including metals [25–27, 31]. Furthermore, a decrease can be caused by a high consumption of several elements in the first time of life. Concentrations of Fe and Zn in newborn pups were comparable to free-ranging, older seals and decreased in the following 2 months, which may be caused by the utilization of these elements through the high metabolism and blood formation of the newborn organism. Additionally, higher Fe concentrations in newborns may have been caused by elevated hemoglobin concentrations during this period [41]. Human studies showed higher concentrations of Fe in newborns compared to nursing infants and children [48].

The concentrations of the majority of trace elements in blood of harbor seal pups are comparable to the values of free-ranging animals of the North Sea [8]. Nevertheless, metal-specific differences were found, which can be caused by ingestion of different metal-

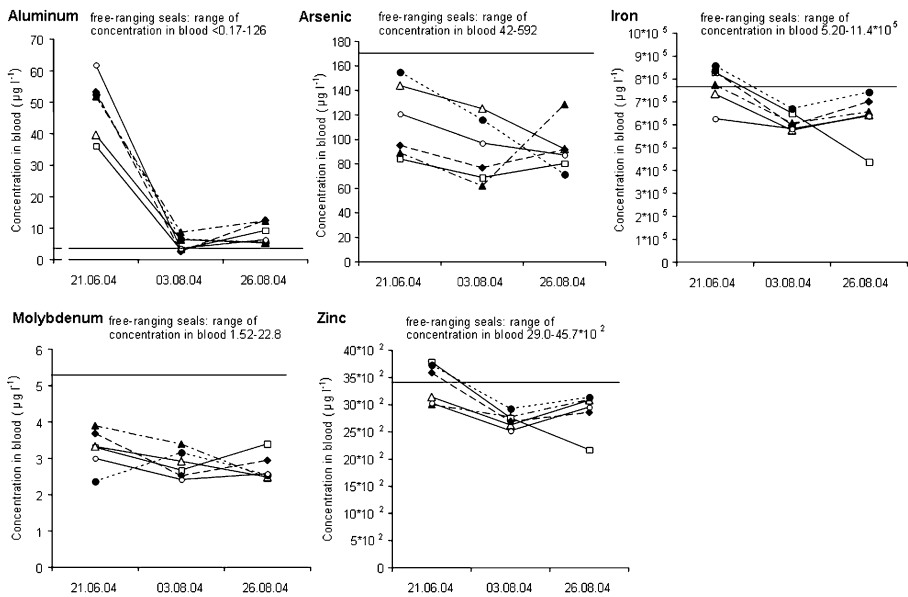


Fig. 2 Concentrations ($\mu\text{g l}^{-1}$) of Al, As, Fe, Mo, and Zn in whole blood samples of six harbor seal pups (* values <math>< \text{LOD}</math>) measured three times. Mean values (lines) and ranges of concentrations in blood of free-ranging seals of the North Sea, Lorenzenplate [8]

contaminated prey. While seals from the North Sea are mostly benthic feeders and the fish species vary with offer and feeding location [49, 50], the species fed to the seals in the Seal Centre was only one, the pelagic fish species, herring *C. harengus*.

Our results show that newborn seals in the North Sea have high concentrations in blood of selected metals, which may have a potential immunological impact on the pups. The immunosuppressive influence of metals on parameters of cellular immunity was investigated for several marine mammal species [35–39]. A study on harbor seal pups, in particular, suggests a higher susceptibility to toxic effects of metals on precursor stages of immune cells isolated from blood of newborns and a decreased susceptibility when the pups were older [1]. Metal pollutants may also induce an activation (e.g., inflammatory process) or dysregulations, e.g., hypersensitivity. Metals found in blood of the pups, e.g., Pb or Ni are known sensitizers for harbor seals [9]. Although a direct relationship to metal body burden cannot be shown, investigations of cytokines of harbor seal pups from the German North Sea coast suggest an activated immune system of the newborns directly after collection of the newborns [1, 2].

Summarizing this pilot study indicates that the multi-element measurement is a highly effective method to get baseline information to essential elements and to the levels of toxic elements in the blood of animals investigated and is the first study on harbor seal pups of the German Wadden Sea coast.

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References

1. Kakuschke A, Valentine-Thon E, Fonfara S, Griesel S, Rosenberger T, Siebert U, Prange A (2008) Metal-induced impairment of the cellular immunity of newborn harbour seals (*Phoca vitulina*). Arch Environ Contam Toxicol 55:129–136
2. Fonfara S, Kakuschke A, Rosenberger T, Siebert U, Prange A (2008) Cytokine and acute phase protein expression in blood samples of harbor seal pups. Mar Biol 155:337–345
3. Stavros HCW, Bossart GD, Hulsey TC, Fair PA (2008) Trace element concentrations in blood of free-ranging bottlenose dolphins (*Tursiops truncatus*): influence of age, sex and location. Mar Pollut Bull 56:371–379
4. Stavros HCW, Bonde RK, Fair PA (2008) Concentrations of trace elements in blood and skin of Florida manatees (*Trichechus manatus latirostris*). Mar Pollut Bull 56:1215–1233
5. Gray R, Canfield P, Rogers T (2008) Trace element analysis in the serum and hair of Antarctic leopard seal, *Hydrurga leptonyx*, and Weddell seal, *Leptonychotes weddellii*. Sci Total Environ 399:202–215
6. Agusa T, Nomura K, Kunito T, Anan Y, Iwata H, Miyazaki N, Tatsukawa R, Tanabe S (2008) Interelement relationships and age-related variation of trace element concentrations in liver of striped dolphins (*Stenella coeruleoalba*) from Japanese coastal waters. Mar Pollut Bull 57:807–815
7. Griesel S, Mundry R, Kakuschke A, Fonfara S, Siebert U, Prange A (2006) Mineral elements and essential trace elements in blood of seals of the North Sea measured by total-reflection X-ray fluorescence analysis. Spectrochim Acta Part B Atom Spectrosc 61:1158–1165
8. Griesel S, Kakuschke A, Siebert U, Prange A (2008) Trace element concentrations in blood of harbor seals (*Phoca vitulina*) from the Wadden Sea. Sci Total Environ 392:313–323
9. Kakuschke A, Valentine-Thon E, Griesel S, Fonfara S, Siebert U, Prange A (2005) Immunological impact of metals in harbor seals (*Phoca vitulina*) of the North Sea. Environ Sci Technol 39:7568–7575
10. Kakuschke A, Valentine-Thon E, Griesel S, Rosenberger T, Mundry R, Siebert U, Prange A (2008) Blood metal levels and metal-influenced immune functions of harbour seals in captivity. Mar Pollut Bull 56:764–769
11. Bossart GD, Reidarson TH, Dierauf LA, Duffield DA (2001) Clinical pathology. In: Dierauf LA, Gulland FMD (eds) Marine mammal medicine. CRC, Boca Raton, FL
12. De Swart RL, Ross PS, Vedder LJ, Boink PBTJ, Reijnders PJH, Mulder PGH, Osterhaus ADME (1995) Haematology and clinical chemistry values for harbour seals (*Phoca vitulina*) fed environmentally contaminated herring remain within normal ranges. Can J Zool Rev Can Zool 73:2035–2043
13. Barnett JEF, Turner L, Booth PA, Hunt AE (2007) Haematological and biochemical values for grey seal pups (*Halichoerus grypus*) during early rehabilitation. Vet Rec 161:447–451
14. Boily F, Beaudoin S, Measures LN (2006) Hematology and serum chemistry of harp (*Phoca groenlandica*) and hooded seals (*Cystophora cristata*) during the breeding season, in the Gulf of St. Lawrence, Canada. J Wildl Dis 42:115–132
15. Sarran D, Greig DJ, Rios CA, Zabka TS, Gulland FMD (2006) Evaluation of aqueous humor as a surrogate for serum biochemistry in California sea lions (*Zalophus californianus*). Aquat Mamm 34:157–165
16. Roletto J (1993) Hematology and serum chemistry values for clinically healthy and sick pinnipeds. J Zoo Wildl Med 24:145–157
17. Tryland M, Krafft BA, Lydersen C, Kovacs KM, Thoresen SI (2006) Serum chemistry values for free-ranging ringed seals (*Pusa hispida*) in Svalbard. Vet Clin Pathol 35:405–412
18. Augier H, Benkoel L, Chamlian A, Park WK, Ronneau C (1993) Mercury, zinc and selenium bioaccumulation in tissues and organs of Mediterranean striped dolphins *Stenella coeruleoalba* Meyer toxicological result of their interaction. Cell Mol Biol 39:621–634
19. Das K, Debacker V, Bouquegneau JM (2000) Metallothioneins in marine mammals. Cell Mol Biol 46:283–294
20. Nielsen JB, Nielsen F, Jorgensen PJ, Grandjean P (2000) Toxic metals and selenium in blood from pilot whales (*Globicephala melas*) and sperm whales (*Physeter catodon*). Mar Pollut Bull 40:348–351
21. Baraj B, Bianchini A, Niencheski LFH, Campos CCR, Martinez PE, Robaldo RB, Muelbert MMC, Colares EP, Zarzur S (2001) The performance of ZEISS GFAAS-5 instrument on the determination of trace metals in whole blood samples of southern elephant seals (*Mirounga leonina*) from Antarctica. Fresenius Environ Bull 10:859–862
22. Fujise Y, Honda K, Tatsukawa R, Mishima S (1988) Tissue distribution of heavy metals in Dalls porpoise in the Northwestern Pacific. Mar Pollut Bull 19:226–230
23. Caurant F, Amiardtriquet C (1995) Cadmium contamination in pilot whales *Globicephala melas*—source and potential hazard to the species. Mar Pollut Bull 30:207–210
24. Yamamoto Y, Honda K, Hidaka H, Tatsukawa R (1987) Tissue distribution of heavy metals in Weddell seals (*Leptonychotes weddellii*). Mar Pollut Bull 18:164–169

25. Itano K, Kawai S, Miyazaki N, Tatsukawa R, Fujiyama T (1984) Mercury and selenium levels at the fetal and suckling stages of striped dolphin, *Stenella coeruleoalba*. *Agric Biol Chem* 48:1691–1698
26. Miyazaki N (1994) Contaminant monitoring studies using marine mammals and the need for establishment of an international environmental specimen bank. *Sci Total Environ* 154:249–256
27. Storelli MM, Marcotrigiano GO (2000) Environmental contamination in bottlenose dolphin (*Tursiops truncatus*): Relationship between levels of metals, methylmercury, and organochlorine compounds in an adult female, her neonate, and a calf. *Bull Environ Contam Toxicol* 64:333–340
28. O'Shea TJ (1999) Environmental contaminants and marine mammals. In: Reynolds JE, Rommel SA (eds) *Biology of marine mammals*. Smithsonian Institution Press, Washington
29. Reijnders PJH, Aguilar A, Donovan GP (1999) Chemical pollutants and cetaceans. *J Cetacean Res Manag Spec Issue* 1:273
30. Das K, Debacker V, Pillet S, Bouquegneau JM (2003) Heavy metals in marine mammals. In: Vos JG, Bossart GD, Fournier M, O'Shea TJ (eds) *Toxicology of marine mammals*. Taylor & Francis, London, pp 135–167
31. Hyvärinen H, Sipilä T (1984) Heavy metals and high pup mortality in the Saimaa ringed seal population in Eastern Finland. *Mar Pollut Bull* 15:335–337
32. Siebert U, Joiris C, Holsbeek L, Benke H, Failing K, Frese K, Petzinger E (1999) Potential relation between mercury concentrations and necropsy findings in cetaceans from German waters of the North and Baltic Seas. *Mar Pollut Bull* 38:285–295
33. Kannan K, Agusa T, Perrotta E, Thomas NJ, Tanabe S (2006) Comparison of trace element concentrations in livers of diseased, emaciated and non-diseased southern sea otters from the California coast. *Chemosphere* 65:2160–2167
34. Bennett PM, Jepson PD, Law RJ, Jones BR, Kuiken T, Baker JR, Rogan E, Kirkwood JK (2001) Exposure to heavy metals and infectious disease mortality in harbour porpoises from England and Wales. *Environ Pollut* 112:33–40
35. De Guise S, Bernier J, Martineau D, Beland P, Fournier M (1996) Effects of *in vitro* exposure of beluga whale splenocytes and thymocytes to heavy metals. *Environ Toxicol Chem* 15:1357–1364
36. Pillet S, Lesage V, Hammill M, Cyr DG, Bouquegneau JM, Fournier M (2000) *In vitro* exposure of seal peripheral blood leukocytes to different metals reveal a sex-dependent effect of zinc on phagocytic activity. *Mar Pollut Bull* 40:921–927
37. Nakata H, Sakakibara A, Kanoh M, Kudo S, Watanabe H, Nagai N, Miyazaki N, Asano Y, Tanabe S (2002) Evaluation of mitogen-induced responses in marine mammal and human lymphocytes by *in vitro* exposure of butyltins and non-ortho coplanar PCBs. *Environ Pollut* 120:245–253
38. Lalancette A, Morin Y, Measures L, Fournier M (2003) Contrasting changes of sensitivity by lymphocytes and neutrophils to mercury in developing grey seals. *Dev Comp Immunol* 27:735–747
39. Kakuschke A, Valentine-Thon E, Fonfara S, Griesel S, Siebert U, Prange A (2006) Metal sensitivity of marine mammals: a case study of a gray seal (*Halichoerus grypus*). *Mar Mamm Sci* 22:985–996
40. Siebert U, Fonfara S, Mundry R, Lehnert K, Seibel H, Rademaker M (2006) Untersuchungen zum Gesundheitszustand von Robben in Schleswig-Holstein im Jahr 2005, Bericht an das Ministerium für Landwirtschaft, Umwelt und ländliche Räume des Landes Schleswig-Holstein und das Landesamt für den Nationalpark Schleswig-Holsteinisches Wattenmeer, Research and Technology Center, University of Kiel, Büsum, Germany
41. Lander ME, Harvey JT, Gulland FMD (2003) Hematology and serum chemistry comparisons between free-ranging and rehabilitated harbor seal (*Phoca vitulina richardsi*) pups. *J Wildl Dis* 39:600–609
42. Nordoy ES, Thoresen SI (2002) Reference values for serum biochemical parameters in free-ranging harp seals. *Vet Clin Pathol* 31:98–105
43. Law RJ (1996) Metals in marine mammals. In: Beyer WN, Heinz GH, Redmon-Norwood AW (eds) *Environmental contaminants in wildlife: interpreting tissue concentrations*. CRC, Boca Raton, FL, pp 357–376
44. Perttälä M, Tervo V, Parmanne R (1982) Age dependence of the concentrations of harmful substances in Baltic herring (*Clupea harengus*). *Chemosphere* 11:1019–1026
45. Perttälä M, Tervo V, Parmanne R (1982) Heavy metals in Baltic herring and cod. *Mar Pollut Bull* 13:391–393
46. Skwarzec B, Holm E, Roos P, Pempkowiak J (1994) Ni-63 in Baltic fish and sediments. *Appl Radiat Isot* 45:609–611
47. Frodello JP, Viale D, Marchand B (2002) Metal concentrations in the milk and tissues of a nursing *Tursiops truncatus* female. *Mar Pollut Bull* 44:551–554
48. Koletzko B (2000) *Kinderheilkunde*. Springer, Berlin
49. Behrends G (1985) Food selection of common seals (*Phoca vitulina* L) in the coastal shallows of Schleswig-Holstein (W Germany). *Z Jagdwiss* 31:3–14
50. Sievers U (1989) Stomach content-analysis in the harbor seal (*Phoca vitulina*) from the Schleswig-Holstein Wadden Sea. *Zool Anz* 222:249–260