Selenium and Mercury in Native and Introduced Fish Species of Patagonian Lakes, Argentina

M. A. Arribére • S. Ribeiro Guevara • D. F. Bubach • M. Arcagni • P. H. Vigliano

Received: 24 October 2007 / Revised: 20 November 2007 / Accepted: 1 December 2007 / Published online: 23 January 2008 © Humana Press Inc. 2007

Abstract A survey of mercury (Hg) and selenium (Se) contents was performed in fish collected from lakes located in two National Parks of the northern patagonian Andean range. Two native species, catfish (*Diplomystes viedmensis*) and creole perch (*Percichthys trucha*), and three introduced species, brown trout (*Salmo trutta*), rainbow trout (*Oncorhynchus mykiss*), and brook trout (*Salvelinus fontinalis*), were caught from lakes Nahuel Huapi, Moreno, Traful, Espejo Chico, and Guillelmo belonging to Nahuel Huapi National Park and from lakes Futalaufquen and Rivadavia, Los Alerces National Park. In lake Moreno, fish diet items were analyzed and rainbow trout grown in a farm. Hg and Se were measured in muscle and liver tissues by instrumental neutron activation analysis. The average concentrations in muscle of Hg for all species, ages, and lakes are between 0.4 to $1.0 \ \mu g g^{-1}$ dry weight (DW) with a few fish, mainly native, exceeding the United States

M. A. Arribére · S. Ribeiro Guevara · D. F. Bubach · M. Arcagni

Laboratorio de Análisis por Activación Neutrónica (LAAN), Unidad de Energía Nuclear, Comisión Nacional de Energía Atómica, Bustillo 9500, 8400 Bariloche, Argentina

S. Ribeiro Guevara e-mail: ribeiro@cab.cnea.gov.ar

D. F. Bubach e-mail: bubachd@cab.cnea.gov.ar

M. Arcagni e-mail: marina.arcagni@cab.cnea.gov.ar

M. A. Arribére Instituto Balseiro, Universidad Nacional de Cuyo, Bustillo 9500, 8400 Bariloche, Argentina

P. H. Vigliano Grupo de Evaluación y Manejo de Recursos Icticos, Universidad Nacional del Comahue, Centro Regional Universitario Bariloche, Quintral 1250, 8400 Bariloche, Argentina

M. A. Arribére (⊠) Laboratorio de Análisis por Activación Neutrónica, Centro Atómico Bariloche, Bustillo 9500, 8400 Bariloche, Argentina e-mail: arribere@cab.cnea.gov.ar Environmental Protection Agency health advisory for freshwater fish limited consumption, and from 0.8 to 1.5 μ g g⁻¹ DW for Se. Average concentrations in liver of Hg in all species range from 0.4 to 0.9 μ g g⁻¹ DW. Brown trout, the top predator in these lakes, showed the lowest average Hg burden in both tissues. Se concentrations in the liver of brown and rainbow trout, up to 279 μ g g⁻¹ DW, are higher than those expected for nearly pristine lakes, exceeding 20 μ g g⁻¹ DW, the threshold concentration associated with Se toxicity. These species show lower Hg contents in muscle, suggesting a possible detoxification of Hg by a Se-rich diet. Creole perch and velvet catfish livers have lower Se concentrations, with a narrower span of values (2.3 to 8.5 μ g g⁻¹ and 3.3 to 5.5 μ g g⁻¹ DW respectively).

Keywords Selenium · Mercury · Freshwater fish · Northern Patagonian lakes · Nahuel Huapi National Park · INAA

Introduction

Selenium (Se) is an element widely distributed in nature; it is nutritionally important as an essential trace element for some plants and animals but is harmful at slightly higher concentrations. The major natural source of environmental Se is the weathering of rocks. For example, in North America, Se tends to be present in large amounts in areas where the soils have been derived from Cretaceous rocks [1–3].

Although the sensitivity to Se and its compounds is extremely variable, a diet containing 0.05 to 0.1 μ g g⁻¹ of Se provides adequate protection to humans and to various species of fish and livestock against Se deficiency, whereas waterfowl has reportedly been adversely affected by diets containing 12 to 280 μ g g⁻¹ of Se. Laboratory and field studies with fish, mammals, and birds have led to agreement that elevated concentrations of Se in diet or water are associated with reproductive abnormalities, congenital malformations, and growth retardation [1].

In general, concern levels for Se in fish as diet for other fish are $3-7 \ \mu g \ g^{-1}$, and the toxic threshold is $7 \ \mu g \ g^{-1}$ fresh weight (FW) as whole-body. Lemly [4] reports $20-40 \ \mu g \ g^{-1}$ dry weight (DW) in viscera as associated with deformities in fish and reports $20 \ \mu g \ g^{-1}$ DW as a toxic threshold. Green sunfish from a lake in North Carolina receiving Se as fly ash wastes from a coal-fired power station had Se levels in liver up to $21.4 \ \mu g \ g^{-1}$ FW resulting in reproduction failure and population decline [5].

There is evidence that selenides of some heavy metals, such as mercury, arsenic, cadmium, and thallium are very insoluble and contribute to keep these metals biologically unavailable [6]. Moreover, Se has been introduced in aquatic systems to diminish Hg bioaccumulation [7]. Mercury (Hg) is considered one of the most toxic elements; although it is not essential for any metabolic process, it can be readily accumulated by biota [8–10]. Experiments performed in enclosures at the Hg contaminated English-Wabigoon river system showed that there was a reduction of Hg burden in fish proportional to the Se accumulation [11]. In another work, performed in 25 Norwegian lakes, Fjield and Rognerud [12] found that Se deposited from the atmosphere seemed to lower the bioavailability of Hg for brown trout, a top predator. A steady increase of Hg contents, from 0.02 to 0.61 μ g g⁻¹ FW, in muscle of large mouth bass from Rogers Quarry was observed after the elimination of selenium-rich discharges of fly ash to the quarry between 1990 to 1998 [13].

Laboratory tests showed that selenium-rich diets tend to reduce the retention of methylmercury (MeHg) in tissues of rainbow trout [14]. In this work, the researchers

studied the effect of dietary Se on the retention of injected organic and inorganic mercury. Rainbow trout fed with a Se-supplemented diet (approximately 10 μ g g⁻¹) augmented the elimination of organic mercury from muscle, liver, kidney, bile, and erythrocytes, compared to those fed with a 1 μ g g⁻¹ Se diet. Elimination of inorganic Hg increased in muscle and kidney, with a rise of Se content in the trout livers from 1 to 26 μ g g⁻¹ during the exposure.

The cases of antagonistic character of Se and Hg mentioned above and others reported in the literature [15–17] show that it depends on the chemical form of both elements. Researchers have found that Se to Hg molar ratios in tissues of marine mammals are around 1:1, and although the correlation is not so evident in fish tissues, Se to Hg molar ratios are in general greater than 1. In addition, cases of synergistic enhancement of Se and Hg toxicity have been reported, as well as the situation that Hg and Se may be antagonistic to each other for adults and synergistic to the young in birds [15, 18–19].

Historically, the Northern Patagonian Andean range was mostly protected from anthropogenic contamination due to the low population density and the access difficulty. Nahuel Huapi National Park, located on the Argentinian side of the northern Patagonian Andes, encloses a 7,850-km² drainage basin with three major river systems and numerous lakes, including lake Nahuel Huapi (Fig. 1). The park encompasses from strictly controlled conservation areas to popular tourist sites. Three major urban settlements are located within the park; the largest of them is the city of Bariloche with circa 120,000 inhabitants.

Some areas surrounding the lakes are experiencing population growth, and the study of lacustrine sediment sequences, suspended particulate material, mussels, and lichens gave evidence of recent metal contamination [20–25]. At present, there are no relevant industrial or extensive agricultural activities in the surrounding area. Potential sources for contaminants, besides the volcanic activity of the area, could be found in human settlements around the lakes, although widespread regional and global contaminants can reach the area through atmospheric transport and subsequent deposition.

In the present work, we studied the contents of Se and total Hg in muscle and liver of five fish species of seven lakes within Nahuel Huapi and Los Alerces National Parks. The existence of wild and farmed rainbow trout, fed only with commercial pelletized food, at lake Moreno, arose the possibility of investigating the impact of different diets on the same species elemental contents. Therefore, farmed rainbow trout, commercial fish food, and the more significant food items of wild fish from lake Moreno were analyzed.

Study Area

The lakes chosen for this study belong to Nahuel Huapi and Los Alerces National Parks. Nahuel Huapi National Park (see Fig. 1) extends between the parallel 40°8′S on the northern border to the 41°36′S in the South, and from 71°2′W to 71°6′W, in the northern Patagonian Andean range. There is a strong vegetation gradient from dense temperate rain forest in the West to dry grasslands in the East. The West to East precipitation gradient is steep, from 3,000 mm y⁻¹ in the West to 500 mm y⁻¹ at the East. All the watersheds within the park are of glacial origin. Most of the lakes are ultraoligotrophic, monomictic, with thermal stratification in summer and complete mixing from autumn to spring [26]. The main human settlements in this Park are Bariloche, Dina Huapi, and Villa La Angostura on the South, East, and North margins of lake Nahuel Huapi, respectively, and Villa Traful in the South margin of lake Traful (Fig. 1). These towns are supported by tourism and some small scale cattle rising. On the Southern shore of lake Moreno East branch, there is a small



Fig. 1 Map of the area and sampling sites

200 inhabitant settlement and a fish farm facility that raises 20 tons of rainbow trout per year in fish pens. The only potential source of Se anthropogenic contamination identified in the area is the use of Se compounds (sodium selenite) as a feeding supplement for the fish at the trout farm.

Los Alerces National Park (between 42°56'S and 43°12'S and 71°34'W and 72°07'W) located 150 km to the South has similar geological, climatological, and biological characteristics as Nahuel Huapi National Park. There are no permanent human settlements

Lake	Lake Code	Area (km ²)	рН	$\begin{array}{c} Conductivity \\ (\mu S \ cm^{-1}) \end{array}$	Maximum Depth (m)	Average Depth (m)	Surface Tempera	ature (°C)	Maximum Length (km)	Maximum Width (km)
							Winter	Summer		
Nahuel Huapi	Ν	557.0	6.8	30.9	464	175	7	14.8	40	8.3
West Moreno	М	4.8	6.8	37.1	110	60	8	16.5	4.7	2.0
East Moreno		5.4	6.8	37.1	128	69	8	17.5	4.4	1.8
Traful	Т	171.5	6.91	41.8	339	173	7	17.0	21.9	4.8
Espejo Chico	Е	2.5	6.5	29.1	60	30	7	14.8	3.1	0.8
Guillelmo	G	5.4	-	-	-	-	7	14.5	7.2	0.9
Futalaufquen	F	44.6	7.8	66	168	101	7	14.5	8.3	1.5
Rivadavia	R	21.7	-	56	147	104	8	14.8	12.9	2.9

 Table 1 General Data of Lakes Belonging to Nahuel Huapi and Los Alerces National Parks^a

^a From [26]

in this park. The lakes chosen are Futalaufquen and Rivadavia. Relevant data about these lakes are shown in Table 1.

Patagonian lakes are poor in fish species. The most widely distributed native fish species on Andean lakes are creole perch (*Percichthys trucha*), the small puyen (*Galaxias maculatus*), and the creole silverside (*Odontesthes hatcherii*) [27, 28]. The native catfishes (*Diplomystes viedmensis* and *Diplomystes mesembranquinus*) are present in lakes and rivers of the Atlantic drainage, including lakes and rivers of Nahuel Huapi National Park. Pacific and Atlantic salmonid species were introduced by the National Park Administration in early 1900 to enhance sport-fishing in the area [29]. Eggs of trout were imported from the Northern Hemisphere to supply fish farms in the area. Both native and introduced species that inhabit the water bodies of Patagonia are considered opportunistic predators of a wide trophic spectrum, being capable to adaptation to different diet types according to the availability of resources [30].

Two native and three introduced species were considered for the study: velvet catfish (*D. viedmensis*), creole perch (*P. trucha*), brook trout (*Salvelinus fontinalis*), rainbow trout (*Oncorhynchus mykiss*), and brown trout (*Salmo trutta*). Preliminary studies indicate that salmonids and creole perch have high mobility in the lakes, whereas velvet catfish, although it moves always along the bottom meanwhile it feeds, is not considered a great swimmer as compared to salmonids.

D. viedmensis feeds exclusively of benthic organisms in all studied environments. Creole perch feeds mostly on macrozoobenthic organisms and has fish as secondary prey. Rainbow trout is the species that shows a greater diet variation, with fish being the most important item in lakes Moreno and Nahuel Huapi, macrozoobenthic organisms and fish in lakes Espejo Chico and Traful, and macrozoobenthic and terrestrial organisms in lakes Rivadavia and Futalaufquen. Finally, brown trout behaves as an almost strict piscivorous fish on all lakes studied. Adult catfish is not a prey for any fish species. [31]

Fish were sampled from five sites of lake Nahuel Huapi: (Rincón Branch, BR; Huemul Branch, BH; López Bay, BL; Puerto Cisnes, PC; and Dina Huapi, DH), lakes Moreno East and West branches, Traful, Espejo Chico, and Guillelmo. Relevant data about these lakes are presented in Table 1.

Materials and Methods

Fish sampling was conducted following the methodology described elsewhere [29, 32]. Fish from lakes Nahuel Huapi, Traful, and Espejo Chico were sampled during the period of thermal stratification (that will be designated as "summer") and during the period of thorough mixing ("winter"). Lake Guillelmo and the lakes of Los Alerces National Park were sampled only in summer. Rainbow trout, brook trout, and creole perch were sampled every season (summer, fall, winter, and spring) during 2 years from the West and East branches of lake Moreno. Rainbow trout were bought at the fish farm, as well as commercial pelletized fish food.

Fish were frozen after being caught. At the laboratory, before analysis, the individuals were thawed, and the whole liver, about 10 g of muscle tissue, stomach, and head were removed using titanium knives. Scales were extracted for age determination.

For each season and site, muscle tissue and the whole liver of the individuals of the each species differing in less than 10 mm length were used to create pooled samples. The individuals of the largest set of fish of each species caught at a same time in one site were analyzed separately to obtain information on intraspecific variability. Details of sample handling and conditioning are described in detail elsewhere [33]. Sample conditioning included freeze-drying and homogenization; handling was performed with Teflon[®] and titanium tools. Aliquots of about 120 to 150 mg of the dried material were sealed in Suprasil AN[®] quartz ampoules for analysis.

Se and Hg were determined by Instrumental Neutron Activation Analysis (INAA) in the RA-6 reactor at Bariloche. Corrections for interference due to impurities in the ampoules were performed, all being negligible. Se was determined by the ⁷⁵Se(n,γ)⁷⁵Se reaction. The mercury concentration measurements were done using the 279.2 keV gamma ray resulting from de decay of the ²⁰²Hg(n,γ)²⁰³Hg reaction product and the low energy lines (67.0, 68.8, and 77.9 keV) of ^{197g}Hg. The high contents of Se in several liver samples induced a significant interference in the 279.2 keV gamma ray peak, which prevented the use of this line for analytical purposes. In those cases, only the ¹⁹⁶Hg(n,γ)^{197g}Hg reaction was used. Very good agreement between both reactions was observed when it was possible to obtain

	Hg Contents (µg	$g g^{-1}$)	Se contents ($\lg g^{-1}$)
	Analysis Results	Certified Value	Analysis Results	Certified Value
NRCC-DORM2 ^a (Dogfish muscle)	4.33±0.53		1.46±0.13	
	4.93 ± 0.66		1.53 ± 0.16	
	5.24 ± 0.71	4.64 ± 0.26	1.51 ± 0.16	$1.40 {\pm} 0.09$
	4.81 ± 0.65		1.44 ± 0.14	
	$4.80 {\pm} 0.60$		1.66 ± 0.21	
	4.61 ± 0.63		1.40 ± 0.14	
NRCC-DOLT2 ^a (Dogfish liver)	2.16 ± 0.38	$2.14{\pm}0.28$	6.22 ± 0.54	6.06 ± 0.49
	2.28 ± 0.24		5.79 ± 0.47	
NRCC-TORT2 ^a (Lobster hepatopancreas)	$0.271 {\pm} 0.047$	$0.27 {\pm} 0.06$	5.67±0.46	5.63±0.67

Table 2	Analy	sis of	Certified	Reference	Material
	/				

Concentrations in dry weight basis

^a Supplied by National Research Council of Canada

significant Hg concentrations from ²⁰³Hg measurements after the correction for the Se interference [33].

Reference materials NRCC-DORM-2 Dogfish Muscle, NRCC-DOLT-2 Dogfish Liver and NRCC-TORT-2 Lobster Hepatopancreas were analyzed together with the samples for analytical quality control, showing good agreement with the certified concentrations (Table 2).

Results and Discussion

Fish muscle and liver contents were measured for dry tissue. Dry to wet weight mass ratios for muscle ranged from 0.21 to 0.27 for brown trout, rainbow, brook trout, and creole perch and from 0.17 to 0.20 for velvet catfish. For liver, dry to wet weight ratios ranged from 0.21 to 0.54 for brown trout, 0.22 to 0.30 for rainbow and brook trout, 0.23 to 0.43 for creole perch, and 0.18 to 0.57 for velvet catfish.

The number of individual specimens caught in the nets varied with the season and the site.

Tables 3, 4, 5, 6, and 7 show the Hg and Se contents in muscle and liver of brown trout, rainbow trout, brook trout, creole perch, and velvet catfish. The number of specimens analyzed per site and their average lengths and ages are included in the tables. These tables also show the range of concentrations measured when individuals, instead of pooled samples, were analyzed for one site and the overall average for the species. Analytical uncertainties for Se determinations were about 11% in muscle samples and 9% in liver

	Brown Trout	Season ^a	Number of	Length ^b	Age ^c	Musc	le	Liver	
			Individuals (n)	(mm)	(years)	Se	Hg	Se	Hg
Nahuel Huapi	Lake Espejo Chico	Summer	1	402	5	1.2	0.74	140	1.3
National Park		Winter	1	384	5	1.6	0.48	89	0.49
	Lake Traful	Summer	6	537	>6	1.5	0.53	90	0.75
		Winter	11	505	>6	1.6	0.39	58	0.29
	Lake Moreno-West	Summer	1	575	5	0.76	0.12	10.1	0.15
		Winter	-	-	-	-	-	-	-
	Lake Nahuel Huapi-	Summer	8	560	5	0.76	0.55	24	1.0
	Rincón Branch	Winter	1	635	8	0.85	0.36	27	0.30
	Lake Nahuel	Summer	3	665	6	0.73	1.36	70	0.43
	Huapi–López Bay	Winter	5	486	4	0.66	0.06	12	0.06
	Lake Nahuel Huapi-	Summer	1	600	6	0.67	0.27	61	0.28
	Huemul Branch	Winter	2	488	4	0.64	0.17	7.2	0.06
	Lake Nahuel	Summer	5	532	5	0.56	0.095	15.7	0.056
	Huapi-Puerto Cisne	Winter	5	447	4	0.68	< 0.05	7.5	< 0.01
	Lake Nahuel	Summer	3	623	6	0.71	0.35	37.6	0.36
	Huapi–Dina Huapi	Winter	1	455	4	0.72	0.20	7.3	0.45
Los Alerces	Lake Futalaufquen	Summer	10	466	4	1.5	1.06	90	1.02
National Park	Range of 10					1.2-	0.27-	21-	0.28-
	individuals					1.7	2.4	279	2.7
	Lake Rivadavia	Summer	1	458	4	1.2	0.20	41	0.28
	Overall average					0.96	0.41	46.3	0.40

Table 3 Concentrations of Se and Hg in Muscle and Liver of Brown Trout ($\mu g g^{-1} DW$)

^a Summer-stratified; winter-non-stratified

^b In mm. Length of the individual (if n=1) or average of the *n* individuals

^c Age of the individual (if n=1) or average of the n individuals

Induviduals (r) Se Hg		Rainbow trout	Season ^a	Number of	Length ^b (mm)	Age ^c (years)	Muscle		Liver	
Namel Huapi National Park Lake Espejo Chico Summer 4 402 50 16 0.36 26 Namel Huapi National Park Lake Tarti Summer 4 40 16 0.39 26 Lake Tarti Summer 10 510 4,0 1,4 0.39 26 Lake Guillelno Summer 10 334 - 16 0.37 23 Lake Moreno-East Summer 6 330 3,4 0,38 0,37 24 Lake Moreno-East Summer 6 390 3,4 0,38 0,37 25 Lake Moreno-East Summer 6 390 3,4 0,38 0,37 26 Take Moreno-East Summer 6 390 3,4 4,3 4,4 0,37 26 26 Take Moreno-East Summer 5 3,9 3,4 4,2 0,39 0,40 4,6 4,6 4,6 4,6 4,6 4,6 4,6				Individuals (n)			Se	Hg	Se	Hg
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Nahuel Huapi National Park	Lake Espejo Chico	Summer	4	402	5.0	1.6	0.36	26	0.44
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	1		Winter	4	384	5.0	1.6	0.50	26	0.94
Winter 10 510 4.0 1.4 0.29 55 Lake Guillelno Summer 10 334 - 1.6 0.17 30 Lake Moreno-East Winter 16 350 3.0 0.78 0.37 22 Lake Moreno-East Winter 16 370 3.0 0.78 0.37 22 Lake Moreno-East Winter 16 370 3.4 4.3 0.78 0.37 22 Lake Moreno-East Summer 1 3.4 4.3 4.2 0.78 0.37 23 Lake Moreno-East Summer 1 3.4 4.3 4.2 0.78 0.37 26 Minter 5 300 3.4 4.2 0.79 0.40 51 Lake Nahuel Huapi-Rucko Branch Summer 3 4.87 4.0 0.75 1.67 26 University fam at Guiterrez stream Summer 5 4.87 4.0 0.75 1.67		Lake Traful	Summer	8	430	4.0	1.6	0.54	LL	0.76
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$			Winter	10	510	4.0	1.4	0.29	55	0.28
$ \begin{array}{rrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrr$		Lake Guillelmo	Summer	10	334	I	1.6	0.17	30	0.22
$ \begin{array}{rcccccccccccccccccccccccccccccccccccc$		Lake Moreno-West	Summer	2	450	4.5	0.98	0.84	55	0.49
$ \begin{array}{llllllllllllllllllllllllllllllllllll$			Winter	16	350	3.0	0.78	0.33	22	0.52
		Lake Moreno-East	Summer	9	390	3.4	0.88	0.57	26	0.69
			Winter	43	443	4.2	0.72	0.60	23	0.43
fish farm (pooled, n=12) 0.80-1.0 0.11-0.60 4.0-1.0 Range of 4 individuals University farm at Gutiérrez stream Spring 12 0.777 0.37 4.6 University farm at Gutiérrez stream Spring 12 2 0.777 0.37 4.6 University farm at Gutiérrez stream Spring 12 2 0.777 0.37 4.6 Lake Nahuel Huapi-Rincón Branch Summer 3 487 4.0 0.73 0.21 14 Lake Nahuel Huapi-López Bay Summer 5 577 5.0 0.77 0.33 0.21 14 Lake Nahuel Huapi-Lúpez Bay Summer 5 577 5.0 0.77 0.33 0.21 14 Lake Nahuel Huapi-Lúpez Bay Summer 5 577 5.0 0.77 0.33 23 Lake Nahuel Huapi-Lúpez Bay Summer 5 570 4.0 0.77 0.33 27 Lake Nahuel Huapi-Dina Huapi Summer 8 570 5.0 0.77 0.33 0.76 27 Lake Futalaufquen Summer		Lake Moreno-East. Commercial	Spring	12		2	0.89	0.40	5.1	0.72
Range of 4 individuals 0.80-1.0 0.11-0.60 4.0 University farm at Guičerrez stream Spring 12 2 0.77 0.37 4.6 University farm at Guičerrez stream Spring 12 2 0.77 0.37 4.6 University farm at Guičerrez stream Spring 12 2 0.77 0.37 4.6 University farm at Guičerrez stream Spring 12 2 0.77 0.37 1.67 92 Lake Nahuel Huapi-López Bay Summer 5 571 5.0 0.73 0.21 14 Lake Nahuel Huapi-Huemul Branch Summer 2 571 5.0 0.73 0.21 14 Lake Nahuel Huapi-Huemul Branch Summer 2 571 5.0 0.76 0.13 23 Lake Nahuel Huapi-Puerto Cisne Summer 7 530 0.74 0.11 27 Lake Nahuel Huapi-Dina Huapi Winter 8 3.0 0.74 0.11 27 Lake Nahuel Huapi-Dina Huapi Summer 7 4.38 3.0 0.74 0.11 25 <td></td> <td>fish farm (pooled, $n=12$)</td> <td>)</td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td>		fish farm (pooled, $n=12$))							
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$		Range of 4 individuals					0.80 - 1.0	0.11 - 0.60	4.0 - 5.8	0.35 - 0.50
Lake Nahuel Huapi-Rincón Branch Summer 3 487 4.0 0.75 1.67 92 Lake Nahuel Huapi-López Bay Winter 5 577 5.0 0.73 0.78 21 Lake Nahuel Huapi-López Bay Winter 5 571 5.0 0.73 0.21 14 Lake Nahuel Huapi-Huemul Branch Winter 8 510 4.0 0.70 0.18 23 Lake Nahuel Huapi-Huemul Branch Winter 2 571 5.0 0.70 0.18 23 Lake Nahuel Huapi-Puerto Cisne Summer 2 571 5.0 0.74 0.11 27 Lake Nahuel Huapi-Puerto Cisne Summer 7 4.38 0.67 0.35 38 Lake Nahuel Huapi-Dina Huapi Winter 10 530 4.0 0.71 0.35 38 Lake Nahuel Huapi-Dina Huapi Summer 7 4.38 3.0 0.74 0.11 27 Lake Nahuel Huapi-Dina Huapi Summer 7 4.38 3.0 0.67 0.51 25 Lake Putalaufquen Summe		University farm at Gutiérrez stream	Spring	12		2	0.77	0.37	4.6	0.42
		Lake Nahuel Huapi-Rincón Branch	Summer	.0	487	4.0	0.75	1.67	92	2.1
Lake Nahuel Huapi-López Bay Summer 6 380 3.0 0.73 0.21 14 Lake Nahuel Huapi-López Bay Winter 8 510 4.0 0.70 0.18 23 Lake Nahuel Huapi-Huemul Branch Winter 5 571 5.0 0.70 0.18 23 Lake Nahuel Huapi-Puerto Cisne Winter 5 550 4.8 0.68 0.13 23 Lake Nahuel Huapi-Puerto Cisne Summer 8 420 3.0 0.74 0.11 27 Lake Nahuel Huapi-Dina Huapi Summer 8 420 3.0 0.74 0.11 27 Lake Nahuel Huapi-Dina Huapi Summer 8 307 2.0 0.67 0.51 25 Lake Futalaufquen Summer 10 4.38 3.0 0.67 0.51 25 Los Alerces Lake Futalaufquen Summer 10 4.0 1.5 0.40 4.1 National Park Range of 10 individuals Summer 8 4.0 1.5 0.40 1.1 National Park Lake Ri			Winter	5	577	5.0	0.83	0.78	21	1.7
Winter 8 510 4.0 0.70 0.18 23 Lake Nahuel Huapi-Huenul Brach Summer 2 571 5.0 0.70 0.18 23 Winter 5 550 4.8 0.68 0.13 33 Lake Nahuel Huapi-Puerto Cisne Summer 8 4.20 3.0 0.74 0.11 27 Lake Nahuel Huapi-Dina Huapi Winter 10 530 4.0 0.71 0.35 38 Lake Nahuel Huapi-Dina Huapi Summer 8 4.20 3.0 0.71 0.35 38 Lake Nahuel Huapi-Dina Huapi Summer 7 4.38 3.0 0.67 0.51 25 Lake Futalaufquen Nuinter 8 307 2.0 0.67 0.51 25 Los Alerces Lake Futalaufquen Summer 10 4.05 1.5 24 0.11 27 Range of 10 individuals Ninter 8 4.0 1.5 0.40 7.3 National Park Lake Rivadavia Summer 8 4.0 1.6 <		Lake Nahuel Huapi-López Bay	Summer	9	380	3.0	0.73	0.21	14	0.19
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$			Winter	8	510	4.0	0.70	0.18	23	0.53
Winter 5 550 4.8 0.68 0.13 32 Lake Nahuel Huapi-Puerto Cisne Summer 8 420 3.0 0.74 0.11 27 Winter 10 530 4.0 0.71 0.35 38 Lake Nahuel Huapi-Dina Huapi Winter 10 530 4.0 0.71 0.35 38 Lake Nahuel Huapi-Dina Huapi Summer 7 438 3.0 0.67 0.51 25 Lake Futalatiquen Summer 7 438 3.0 0.67 0.51 25 Los Alerces Lake Futalatiquen Summer 10 405 0.67 0.51 25 Range of 10 individuals Summer 10 405 4.0 1.5 0.40 11 National Park Lake Rivadavia Summer 8 402 4.0 1.6 0.38 31 National Park Lake Rivadavia Summer 8 402 4.0 1.6 0.38 31 Overall average (wild frout only) Overall 0.45 5.0 <t< td=""><td></td><td>Lake Nahuel Huapi-Huemul Branch</td><td>Summer</td><td>2</td><td>571</td><td>5.0</td><td>0.70</td><td>0.18</td><td>23</td><td>0.53</td></t<>		Lake Nahuel Huapi-Huemul Branch	Summer	2	571	5.0	0.70	0.18	23	0.53
Lake Nahuel Huapi-Duerto Cisne Summer 8 420 3.0 0.74 0.11 27 Kendel Huapi-Dina Huapi Winter 10 530 4.0 0.71 0.35 38 Lake Nahuel Huapi-Dina Huapi Winter 10 530 4.0 0.71 0.35 38 Los Alerces Lake Futalaufquen Winter 8 307 2.0 0.67 0.51 25 Range of 10 individuals National Park Lake Rvadavia Summer 10 4.0 1.5 0.40 41 National Park Lake Rvadavia Summer 8 402 4.0 1.6 0.36 17-0 0.20 17- National Park Lake Rvadavia Summer 8 402 4.0 1.6 0.40 11 Overall average (widt frout only) Overall average (widt frout only) 0.45 35 31			Winter	5	550	4.8	0.68	0.13	32	0.49
Winter 10 530 4,0 0.71 0.35 38 Lake Nahuel Huapi-Dina Huapi Summer 7 438 3.0 0.67 0.51 25 38 Los Alerces Lake Futalaufquen Winter 8 307 2.0 0.67 0.51 25 7.3 Los Alerces Lake Futalaufquen Summer 10 405 4.0 1.5 0.40 41 National Park Lake Rivadavia Summer 8 402 4.0 1.6 0.12-0.92 17- National Park Lake Rivadavia Summer 8 402 4.0 1.6 0.12-0.92 17- Overall averase (wild trout only) Summer 8 402 4.0 1.6 0.38 31		Lake Nahuel Huapi-Puerto Cisne	Summer	8	420	3.0	0.74	0.11	27	0.37
Lake Nahuel Huapi-Dina Huapi Summer 7 438 3.0 0.67 0.51 25 Los Alerces Lake Futalaufquen Winter 8 307 2.0 0.63 0.20 7.3 Los Alerces Lake Futalaufquen Summer 10 405 4.0 1.5 0.40 41 National Park Lake Rivadavia Summer 8 402 4.0 1.2-0.92 17- National Park Lake Rivadavia Summer 8 402 4.0 1.6 0.38 31 Overall average (wild front only) Overall average (wild trout only) 0.45 35			Winter	10	530	4.0	0.71	0.35	38	0.36
Winter 8 307 2.0 0.63 0.20 7.3 Los Alerces Lake Futalaufquen Summer 10 405 4.0 1.5 0.40 41 Range of 10 individuals Summer 10 405 4.0 1.5 0.40 41 National Park Lake Rivadavia Summer 8 402 4.0 1.6 0.38 31 Overall average (wild trout only) Overall average (wild trout only) Summer 8 402 4.0 1.6 0.45 35		Lake Nahuel Huapi–Dina Huapi	Summer	7	438	3.0	0.67	0.51	25	0.18
Los Alerces Lake Futalaufquen Summer 10 405 4.0 1.5 0.40 41 Range of 10 individuals Range of 10 individuals Summer 10 1.2-1.7 0.12-0.92 17- National Park Lake Rivadavia Summer 8 402 4.0 1.6 0.38 31 Overall average (wild trout only) Overall average (wild trout only) 8 4.0 1.0 0.45 35			Winter	8	307	2.0	0.63	0.20	7. 3	0.45
Range of 10 individuals Range of 10 individuals 1.2–1.7 0.12–0.92 17– National Park Lake Rivadavia Summer 8 402 4.0 1.6 0.38 31 Overall average (wild trout only) Overall average (wild trout only) 0.45 35	Los Alerces	Lake Futalaufquen	Summer	10	405	4.0	1.5	0.40	41	<0.9
National Park Lake Rivadavia Summer 8 402 4.0 1.6 0.38 31 Overall average (wild trout only) Overall average (wild trout only) 0.45 35		Range of 10 individuals					1.2-1.7	0.12 - 0.92	17-73	<0.9
Overall average (wild trout only) 1.0 0.45 35	National Park	Lake Rivadavia	Summer	8	402	4.0	1.6	0.38	31	0.28
		Overall average (wild trout only)					1.0	0.45	35	0.61

49

° Age of the individual (if n=1) or average of the n individuals; winter-non-stratified

^b Length of the individual (if n=1) or average of the n individuals

	Brook Trout	Season ^a	Number of	Length ^b	Age ^c	Muscl	e	Liver	
			Individuals (n)	(mm)	(years)	Se	Hg	Se	Hg
Los Alerces National Park	Lake Futalaufquen	Summer	1	433	-	1.8	0.76	7.4	0.63
Nahuel Huapi	Lake Espejo	Summer				_	_	_	_
National Park	Chico	Winter	3	437	_	1.8	0.71	9.5	1.2
	Lake Traful	Summer	9	371	3.7	1.4	0.66	3.9	0.22
		Winter	10	358	3.5	1.7	0.24	5.5	0.14
	Lake Guillelmo	Winter	10	308		2.2	0.29	5.5	1.6
	Range of 10 individuals					1.7– 3.1	0.12– 0.57	4.1– 7.6	0.23– 5.3
	Lake Moreno-	Summer	1	240	2.0	0.96	0.25	4.5	0.31
	West ^d	Winter	6	417	3.8	0.26– 1.9	0.94– 1.3	3.0– 6.7	0.2 - 1.07
	Lake Moreno- East ^d	Summer	4	319	2.8	0.93– 1.04	0.60– 1.08	3.1– 5.3	0.42– 1.6
		Winter	10	408	3.8	0.80– 1.03	0.38– 4.0	3.3– 7.6	0.49– 7.1
	Lake Nahuel Huapi–Rincón	Summer	1	340	_	0.87	0.37	2.6	0.29
	Branch Overall average	Winter	4	361	-	1.0 1.3	0.45 0.65	3.8 5.1	0.42 0.81

Table 5 Concentrations of Se and Hg in Muscle and Liver of Brook Trout ($\mu g g^{-1} DW$)

^a Summer-stratified; winter-non-stratified

^b Length of the individual (if n=1) or average of the n individuals

^c Age of the individual (if n=1) or average of the *n* individuals

^d Range of values of the different seasons

samples, and about 16% for Hg in muscle samples and 18% in liver samples. Figure 2a shows the plots of Hg contents in liver vs Hg contents in muscle, and Fig. 2b shows the plots of Se contents in liver vs Se contents in muscle for all the analyzed samples, indicating the lake of provenance. Table 8 shows the results of the analysis of diet items of wild fish from lake Moreno and commercial pelletized food for fish.

The analysis of the data of the individuals of each species collected in one site showed that the standard deviation of the Se contents in muscle is similar or lower than the analytical uncertainty (which is around 11%), indicating that there are not significant differences in Se contents in muscle among individuals of the same species in one site (see Fig. 2). Se liver contents of salmonids and perch have much higher variability, reaching maximum to minimum value ratios of up to ten for brown and rainbow trout.

Selenium contents in brown and rainbow trout muscle samples range between 0.56 and 1.1 μ g g⁻¹ DW, for fish from lakes Moreno and Nahuel Huapi, whereas the samples of the same species of all the remaining lakes range between 1.2 and 1.7 μ g g⁻¹ DW, which are included within the expected range of values for non-contaminated lakes [1]. Se contents in creole perch muscle are higher but show a similar pattern concerning the lake of origin (see Fig. 2).

The differences in Se contents in muscle of each species in the different lakes cannot be related directly to the geographic setting of the lakes: lakes Futalaufquen and Rivadavia

trations of Se and Hg in Muscle and Li	iver of Crec	le Perch ($\mu g g^{-1} DW$)						
Creole Perch	Season ^a	Number of Individuals (n)	Length ^b (mm)	Age ^c (years)	Muscle		Liver	
					Se	Hg	Se	Hg
Lake Espejo Chico	Summer	6	432	6	2.5	1.3	8.5	0.50
	Winter	7	413	6	2.2	1.4	7.3	1.3
Lake Traful	Summer Winter	1	435	6	3.0	0.68	4.4	0.14
Lake Moreno-West	Summer	21	394	7	1.2	1.3	5.4	0.66
Range of 10 individuals					0.97 - 1.4	0.70 - 2.0		
1	Winter	42	400	7	1.2	1.0	4.2	0.37
Lake Moreno-East	Summer	21	390	6.6	1.2	1.2	5.2	0.44
Range of 11 individuals					0.86 - 1.5	0.71 - 3.2	3.8-8.0	0.19-0.64
	Winter	36	411	8	1.3	1.3	5.2	0.40
Lake Nahuel Huapi-Rincón Branch	Summer	10	442	9.5	1.1	1.4	4.8	0.46
	Winter				I	I	Ι	I
Lake Nahuel Huapi-López Bay	Summer	8	367	7	1.1	0.43	2.3	0.18
	Winter	5	406	8	1.2	0.61	3.0	0.073
Lake Nahuel Huapi-Huemul Branch	Summer	12	394	7	0.97	0.53	5.0	0.36
	Winter				Ι	I	I	I
Lake Nahuel Huapi-Puerto Cisne	Summer	5	422	6	0.94	0.89	5.1	0.21
	Winter	13	423	6	0.93	1.2	3.3	0.25
Lake Nahuel Huapi-Dina Huapi	Summer	2	450	10	0.79	2.5	5.5	1.5
	Winter	10	410	8	0.93	0.87	4.6	0.47
Lake Futalaufquen	Summer	0	261	4	2.2	0.47	6.8	0.22
Lake Rivadavia	Summer	6	315	4	3.2	0.25	7.6	0.23
Overall average					1.5	1.0	5.2	0.46
	Trations of Se and Hg in Muscle and L Creole Perch Lake Espejo Chico Lake Traful Lake Moreno-West Range of 10 individuals Lake Moreno-East Range of 11 individuals Lake Moreno-East Range of 11 individuals Lake Nahuel Huapi-Rincón Branch Lake Nahuel Huapi-López Bay Lake Nahuel Huapi-López Bay Lake Nahuel Huapi-Lopez Bay Lake Nahuel Huapi-Lopez Bay Lake Nahuel Huapi-Dina Huapi Lake Rivadavia Overall average	Trations of Se and Hg in Muscle and Liver of Crec Creole Perch Season ^a Lake Espejo Chico Summer Lake Traful Summer Lake Traful Summer Minter Summer Lake Moreno-West Summer Range of 10 individuals Winter Range of 11 individuals Winter Lake Moreno-East Summer Range of 11 individuals Winter Lake Nahuel Huapi-Rincón Branch Summer Lake Nahuel Huapi-López Bay Summer Lake Nahuel Huapi-Dína Huapi Summer Lake Nahuel Huapi-Dína Huapi Summer Lake Futalaufquen<	Itrations of Se and Hg in Muscle and Liver of Creole Perch (µg g ⁻¹ DW) Creole Perch Season ^a Number of Individuals (n) Lake Espejo Chico Summer 9 Lake Traful Winter 7 Lake Traful Summer 1 Minter 7 1 Lake Moreno-West Summer 1 Range of 10 individuals Winter 21 Range of 11 individuals Winter 21 Range of 11 individuals Winter 36 Lake Moreno-West Summer 10 Range of 11 individuals Winter 36 Lake Nahuel Huapi-Rincón Branch Summer 36 Lake Nahuel Huapi-López Bay Summer 36 Lake Nahuel Huapi-López Bay Summer 36 Lake Nahuel Huapi-Lúpez Bay Summer 3 Lake Nahuel Huapi-Lúpez Bay Summer 12 Lake Nahuel Huapi-Dina Huapi Summer 3 Lake Nahuel Huapi-Dina Huapi Summer 3 Lake Nahuel Huapi-Dina Huapi Summer 1 Lake Futalaufquen Summer 3 <	Itrations of Se and Hg in Muscle and Liver of Creole Perch ($\log g^{-1}$ DW)Creole PerchSeason ^a Number of Individuals (n)Length ^b (mm)Lake Espejo ChicoSummer9432Lake TraftulWinter7413Lake Moreno-WestSummer21394Lake Moreno-WestSummer21394Range of 10 individualsWinter21396Lake Moreno-EastSummer21396Range of 11 individualsWinter21390Lake Moreno-EastSummer21390Lake Moreno-EastSummer21390Lake Moreno-EastSummer21390Lake Moreno-EastSummer21390Lake Moreno-EastSummer21390Lake Nahuel Huapi-Rincón BranchSummer8367Lake Nahuel Huapi-López BayWinter5400Lake Nahuel Huapi-López BaySummer8367Lake Nahuel Huapi-Dina HuapiSummer5420Lake Nahuel Huapi-Dina HuapiSummer2420Uake FutalaufquenSummer2400Lake RivadaviaSummer230Lake RivadaviaSummer23Lake RivadaviaSummer23Lake RivadaviaSummer3261Lake RivadaviaSummer33Lake RivadaviaSummer33Lake RivadaviaSummer33	Itrations of Se and Hg in Muscle and Liver of Creole Perch (ug g^{-1} DW)Age ⁶ (years)Creole PerchSeason ^a Number of Individuals (n)Length ^b (mm)Age ⁶ (years)Lake Espejo ChicoSummer74139Lake TrafulWinter74139Lake TrafulSummer14359Lake TrafulSummer213947Lake Moreno-WestSummer213997Lake Moreno-EastSummer213997Lake Moreno-EastSummer104429.5Lake Moreno-EastSummer104429.5Lake Moreno-EastSummer104429.5Lake Moreno-EastSummer104429.5Lake Nahuel Huapi-López BaySummer83677Lake Nahuel Huapi-López BaySummer124429.5Lake Nahuel Huapi-López BaySummer54008Minter12Adot77Lake Nahuel Huapi-Lúpez BaySummer54007Lake Nahuel Huapi-Dina HuapiSummer54008	trations of Se and Hg in Muscle and Liver of Creole Perch (ug g^{-1} DW)Creole PerchSeason*Number of Individuals (n)Age ⁴ (yeans)MuscleLake Espejo ChicoSummer743292.5Lake TrafulWinter741392.2Lake TrafulSummer143593.0Lake TrafulSummer143593.0Lake TrafulSummer139471.2Lake Moreno-WestSummer2139471.2Lake Moreno-WestSummer2139471.2Lake Moreno-EastSummer2139471.2Lake Moreno-EastSummer2139471.2Lake Nahuel Huapi-Rincón BranchSummer344181.3Lake Nahuel Huapi-Huenul BranchWinter53471.1Lake Nahuel Huapi-Huenul BranchSummer136771.1Lake Nahuel Huapi-Dina H	transmission of Se and Hg in Muscle and Liver of Creole Petch (tug g ⁻¹ DW)Creole PetchSeason ^a Number of Individuals (n)Age ⁶ (years)MuscleLake Espejo ChicoSummer92.51.3Lake Espejo ChicoSummer92.21.4Ninter741392.21.4Lake Espejo ChicoSummer143592.21.4Lake TraftilSummer2143592.21.4Ninter741392.21.40.00-2.0Ninter2139471.21.21.2Lake Moreno-WestSummer2139471.21.3Range of 10 individualsWinter3644290.97-1.40.00-2.0Lake Moreno-EastSummer104429.51.11.4Lake Nahuel Huapi-Liopez BayWinter364429.51.10.43Vinter540081.30.90.90.90.6Lake Nahuel Huapi-Liopez BayWinter544070.970.30.6Lake Nahuel Huapi-Liopez BayWinter14429.51.10.43Lake Nahuel Huapi-Liopez BayWinter14429.51.10.43Lake Nahuel Huapi-Puerto CisueSummer12.21.10.43Lake Nahuel Huapi-Puerto CisueSummer12.21.10.43<	transmersion of Se and Hg in Muscle and Liver of Cheole Perch (ug g^{-1} DW)Creole PerchSeasontNumber of Individuals (n)Age ⁶ (verss)MuscleLiverLake Espejo ChicoSummer92.51.38.5Lake TraftilSummer741392.51.4Lake TraftilSummer143392.51.47Lake TraftilSummer2143392.00.684.4Lake TraftilSummer213906.61.21.38.2Lake Moreno-WestSummer213906.61.21.38.2Lake Moreno-WestSummer213906.61.21.38.2Lake Moreno-WestSummer213906.61.21.38.2Lake Moreno-EastSummer1041181.11.44.8Lake Moreno-EastSummer106.61.21.33.2Lake Moreno-EastSummer83.64.44.8Lake Moreno-EastSummer106.61.21.33.2Lake Moreno-EastSummer106.61.21.33.2Lake Moreno-EastSummer1071.21.33.2Lake Moreno-EastSummer106.61.21.33.2Lake Moreno-EastSummer1.21.31.44.8Lake Nahuel Huapi-Lopez BaySummer

^b Length of the individual (if n=1) or average of the n individuals ^c Age of the individual (if n=1) or average of the n individuals

^a Summer-stratified; winter-non-stratified

	Velvet Catfish	Season ^a	Number of	Length ^b	Muscle	e	Liver	
			Individuals (n)	(mm)	Se	Hg	Se	Hg
Nahuel Huapi	Lake Traful	Winter	5	209	1.2	0.21	5.1	0.28
National Park	Range of 5				0.84– 1.6	0.055-	4.2-	0.09-
	Lake Moreno-East	Summer	7	221	0.90	1.3	5.4	1.6
	Range of 7				0.82-	0.76-		
	individuals				0.99	2.3		
	Lake Nahuel Huapi-	Summer	1	155	1.4	1.4	5.8	3.9
	Rincón Branch	Winter			_	_	_	_
	Lake Nahuel Huapi– López Bay	Summer	3	219	0.77	0.44	4.1	0.41
	Range of 3				0.48-	0.32-	3.9-	0.32-
	individuals				1.1	0.52	4.9	0.49
	Lake Nahuel Huapi– Huemul Branch	Summer	5	245	0.73	0.86	5.4	0.61
	Lake Nahuel Huapi– Puerto Cisne	Summer	3	267	0.69	0.46	4.9	0.29
	Range of 3				0.57-	0.26-	4.5-	0.15-
	individuals				0.82	0.85	5.1	0.39
		Winter	1	190	0.63	0.27	4.6	0.58
	Lake Nahuel Huapi– Dina Huapi	Summer	9	196	0.63	0.24	4.1	0.25
	Range of 9				0.48-	0.075-	3.3-	0.10-
	individuals				0.82	0.43	4.6	0.42
		Winter	1	150	0.48	0.19	4.0	0.40
	Overall average				0.83	0.60	4.8	0.92

Table 7 Concentrations of Se and Hg in Muscle and Liver of Velvet Catfish ($\mu g^{-1} DW$)

^a Summer-stratified; winter-non-stratified

^b Length of the individual (if n=1) or average of the *n* individuals

are located about 150 km to the South of lake Nahuel Huapi; Traful is located to the North without a direct connection to the Nahuel Huapi-Moreno lake system, and Espejo Chico is close to lake Nahuel Huapi (its waters discharge into lake Correntoso, which drains into Nahuel Huapi through a short river); and Guillelmo is about 40 km South of Nahuel Huapi, and it belongs to a basin that drains to the Pacific ocean. Furthermore, the results of a survey on heavy metal contents in soft tissues and digestive gland of a widespread patagonian mussel, *Diplodon chilensis*, performed in the four lakes of Nahuel Huapi National Park, showed that Se content in all sites was analytically equivalent [22].

Some authors ascribe differences in body burden to dilution effects with growth. We did not observe any correlation between Se with fish length or age. Regarding the length at a certain age, salmonids from lake Nahuel Huapi tend to attain larger sizes. It was possible to obtain the growth curves for rainbow trout of four lakes of Nahuel Huapi National Park (see Fig. 3). The larger fish sizes at lake Nahuel Huapi could explain for the lower Se concentrations. However, the growth curves for lakes Moreno and Traful are similar, whereas the Se concentrations are different.

No fish exceeded the Argentinian Se advisory (Código Alimentario Argentino and Res. SENASA N°533–10.05.94) for human adult consumption without restrictions (2.4 μ g g⁻¹



Fig. 2 a Hg contents in liver tissue vs Hg contents in muscle for brown trout, rainbow trout, brook trout, creole perch, and velvet catfish from lakes Nahuel Huapi (N), Moreno (M), Traful (T), Espejo Chico (E), Guillelmo (G), Futalaufquen (F) and Rivadavia (R). Farmed rainbow trout from lake Moreno are included (O). **b** Se contents in liver tissue vs Se contents in muscle for the same fish species and sites. The least square linear fit of the set of data of each graph is shown. All data are shown in DW basis

DW, assuming a dry weight to wet weight ratio of 0.25). The overall Se mean contents in muscle for each species is below the recommended value for consumption without restriction by children (0.3 μ g g⁻¹ FW, 1.2 μ g g⁻¹ DW, dry weight to wet weight ratio of 0.25), as well as fish from lakes Moreno and Nahuel Haupi. Fish from lakes Traful, Espejo Chico, Guillelmo, Rivadavia, and Futalaufquen are slightly above this guideline.

	Food prey	Se	Hg
Wild fish	Insect larvae	3.2	1.4
	Salmonid juveniles		
	Length=6.8–8.5 cm	1.23	0.077
	Length=13 cm	0.94	0.206
	Galaxid juveniles		
	Length= $3.7-6.8$ cm	1.8	0.385
	Aegla sp. (small crabs)		
	Length= $1.4-1.8$ cm	1.0	0.14
	Length=1.8–2.9 cm	1.0	0.14
	Gasteropods		
	Length= $1.0-1.7$ cm	0.34	0.14
	Length=1.7-3.0 cm	0.21	0.046
	Samastacus sp. (shrimp)		
	Length=2.5–3.7 cm	1.3	< 0.1
	Length=4.5-5.8 cm	0.81	0.25
Farmed fish	Pelletized food for younger fish	1.6-2.6	0.02-1.0
	Pelletized food for adult fish	1.0-1.9	0.02-0.81

Table 8 Se and Hg Contents in Food Preys for Rainbow Trout and Perch from Lake Moreno, and Pelletized Food for Farmed Fish ($\mu g g^{-1} DW$)

Conversely, Se concentrations in liver of brown and rainbow trout, up to 279 μ g g⁻¹ DW, greatly exceeded 20 μ g g⁻¹ DW, the concentration in liver associated with Se toxicity [4]. The values we measured are closer to the 21 μ g g⁻¹ FW level mentioned in [5], which could be an indication that salmonids from the lakes studied are having Se rich diets. Table 9 shows the selenium contents measured in this work compared to contents in salmonids, northern hemisphere perciformes, and catfishes, from Se contaminated and non-contaminated lakes. Se contents in muscle of the salmonids included in this study are similar or slightly above the values measured for salmonids of non-contaminated boreal lakes; however, they are clearly below the Se contents in muscle of fish from sites with Se rich waters. The same





		1			
Species	Reference	Site	Specific characteristics	Tissue ^a	Se µg g ⁻¹
Salmonids (3 spp.)	This work	Northern Patagonian lakes		М	0.56–3.1 DW
				L	2.6–279 DW
Native perch (P. trucha)				Μ	0.79–3.2 DW
				L	2.3–8.5 DW
Native catfish (D. viedmensis)				Μ	0.48–1.6 DW
-				L	0.055–2.3 DW
Satimontds Rainbow trout (<i>O. mykiss</i>)	[34]	Lake in an artificially irrigated area: the Riverton Reclamation Project, Fremont County, Wyoming.	Middle Depression lake-1988	X	9.2–15.2 DW
			Middle Depression lake-1994	Μ	5.0–9.8 DW
Trout, 2 spp.	[35]	Wyoming	Water containing 12.3-13.3 ppb Se	L	50.0-70.0 FW
				Μ	<2.0 FW
Char (Salvelinus alpinus)	[36]	Canadian Subartic and	Mackenzie river basin	Μ	0.14-1.65FW
Whitefish (Coregonus hueneaformis)		Artic freshwater fish	and other boreal lakes	М	0.07–0.79 FW
Lake trout (<i>S. namaycush</i>) Perciformes				М	0.08-0.34 FW
Walleye (S. vitreum)	[36]	Canadian Subartic and Artic freshwater fish	Mackenzie river basin and other boreal lakes	Μ	0.14-0.26 FW
	[37]	Lake Oahe, South Dakota, Foster Bay (FB), Grand River (GR), Moreau River (MR)	Suspected contaminated waters	Μ	0.49-0.56 FW (FB)
				Μ	0.43–0.83 FW (GR)
				Μ	0.42–0.60 FW (MR)
Perch (P. fluviatilis)	[7]	Lake Öltertjärn, N Sweden	With controlled addition of sodium selenite to the	Μ	0.21–0.57 FW t
			Walci (11 y 12)		
					1.72-10.16 F W tl
					1.92-8.46 FW t2

Table 9 (continued)					
Species	Reference	Site	Specific characteristics	Tissue ^a	Se µg g ⁻¹
Yellow perch (P. flavescens)	[34]	Lake in an artificially irrigated area: the Riverton Reclamation Project, Fremont County, Wyoming.	Lake Cameahwait-1988	W	8.40–10.4 DW
			Lake Cameahwait – 1994	Μ	6.1–9.5 DW
			Sand Mesa pond – 1994	M X	1.8–3.3 DW
T	1012	Ē		M	2 1 1117
Largemouth bass	[13]	Kogers Quarry in Anderson	From 196/ to 1989 for the disposal	Μ	3-1 F W
(Micropterus salmoides)		County, Tennessee	of fly ash from a coal fired steam plant. Fish deformed of high Se		
Largemouth bass	[1]	Upper Mississippi River	From water containing <5 ppb Se		
I	1			Μ	0.05 FW
				Г	0.8 FW
			From water containing 22.6 ppb Se	М	1.7 FW
				L	10.2 FW
Bluegill (Lepomis macrochirus)	Ξ	Upper Mississippi River	From water containing <5 ppb Se	W	0.04 FW
				L	0.7 FW
			From water containing 22.6 ppb Se	М	2.0–3.1 FW
				Г	11.2 FW
Green sunfish	[2]	Lake Belews, North Carolina	Receives Se as fly ash wastes from	Г	Up to 21.4 FW
(Lepomis cyanellus) Other fishes			coal fired power station		
Several species	[38]	Madeira river, Brazil	The river, tributary of the Amazon, is located in a gold mining area	W	0.012-0.37 FW
Several species	[39]	San Francisco Bay	Receives Se rich irrigation drainage	М	3-20 DW
Several species	[40]	Lake Macquarie, NSW,	located near 3 coal-fired	М	4-10 DW
		Australia	power stations and several collieries		
^a M muscle; L liver; FW fresh weight	t; DW dry weight. Dry	/ to wet mass ratios for this work muscle sar	nples range from 0.20 to 0.27.		

56

pattern is observed when perch is compared with other perciformes and velvet catfish to other catfishes of the world.

Although Se liver contents present a high degree of variability, there is a correlation between Se contents in liver and Se contents in muscle (ρ between 0.46 and 0.66, Pbetween 0.0001 and 0.0050) for the whole set of data. The linear fits are shown in Fig. 2b. Because there were not significant differences in Se contents in fish muscle for each species and lake, these correlations indicate that Se contents in liver show the same geographic pattern as muscle. These fits give an average liver to muscle Se content ratio of 73 ± 23 for brown trout, 27 ± 8 for rainbow trout, 1.3 ± 0.4 for brook trout, 1.6 ± 0.4 for creole perch, and 1.4 ± 0.3 for velvet catfish.

Mercury contents do not exceed 1.0 μ g g⁻¹ FW, which is expected in biota from locations not directly affected by direct anthropic sources but higher than what is expected in remote lakes [6]. However, similar contents have been reported in trout from several wilderness lakes. The lower values measured in this study are similar or lower to those measured in low impacted lakes of the Northern hemisphere, whereas the higher values are about twice the maximum values measured in those lakes [12, 41–47].

For brown trout, rainbow trout, perch, and catfish, the longest specimens were, respectively, 25, 40, 34, and 67% longer than the smallest one; no correlation was found between Hg contents and length, except for the latter species (ρ =0.66).

Bio-accumulation of Hg occurs in the species and lakes studied in this project. Although the ranges of Hg contents for each species overlap, the species considered as the top predator, brown trout, has lower Hg contents in muscle than the other species in contrast with the observations in other water bodies. Moreover, brown trout from lake Nahuel Huapi, where this species behaves exclusively as a piscivore, has the lowest Hg contents (see Fig. 2). Because organic Hg compounds, not total Hg, are the ones prone of being biomagnified in the trophic web, no conclusions can be drawn from the total mercury data alone regarding this phenomenon. A better knowledge of the trophic web structure of the lakes is needed to draw a conclusion on the low Hg burden of the species considered as the top predator.

Temperature is one of the more influential parameters in Hg bio-accumulation [48–50]. A rise in temperature could increase the accumulation of Hg in fish by increasing the metabolic rate. We analyzed the ratio between Hg contents in summer to contents in winter for each tissue at each sampling site. We found that Hg contents in both tissues were consistently equal or higher in summer than in winter of brown trout for all sites but not for the other species.

No significant correlation was found between Hg contents in liver vs muscle for a particular lake; however, when considering the whole data set, there is correlation between Hg contents in liver and Hg contents in muscle for each species (ρ between 0.55 and 0.89, P between <0.0001 and 0.0018). The linear fits are shown in Fig. 2a. These fits give an average liver to muscle Hg content ratio of 1.0 ± 0.1 for brown trout, 0.7 ± 0.2 for rainbow trout, 1.2 ± 0.3 for brook trout, 0.27 ± 0.06 for creole perch, and 1.8 ± 0.3 for velvet catfish. Salmonids have, in average, the same Hg concentration in liver as in muscle, whereas perch has four times more Hg contents in muscle than in liver, and catfishes have two times more Hg concentration in liver than in muscle.

Table 10 shows the ranges of concentrations measured in this work compared with Hg contents of fish found in the literature. Hg concentrations in the same species from other works are compared to present research when data is available; in other cases, species with similar habitat or diet are compared (creole perch is compared to other perciformes and velvet catfish with other carnivore catfishes). Mercury contents in fish tissues have a larger degree of variability than Se; however, the values reported in this work range from the

Species	Ref.	Site	Specific characteristics	Tissue ^a	${\rm Hg}_{\rm T},\mu gg^{-1}$	MeHg ($\mu g \ g^{-1}$)
Brown trout (S. trutta)	This work	Northern Patagonian lakes	No direct anthropogenic Hg inputs	W .	<0.05-2.4 DW	
	[12]	25 lakes in Norway	No direct Hg inputs	Ч	<0.01-2.7 DW 0.01-0.24 FW	
	[43]	Gulf of Bothnia	•	Μ	0.10-0.19 FW	
Rainbow trout (O. mykiss)	This work	Northern Patagonian lakes	No direct anthropogenic Hg inputs	М	0.1–1.7 DW	
				Г	0.18-2.1 DW	
	[45]	Lakes Okareka, Okaro and	Lakes receiving geothermal waters	Μ	(~0.24-2 FW)	0.22–1.84 FW
		Rotomaha, New Zealand				
	[44]	Lakes Okareka, Okaro and Potomoha Nawi Zaoland	Lakes receiving geothermal waters	Μ	<u>~</u>	69–96%
		Kotomana, INEW Zealand		;		
	[51]	Owyhee River drainage basin	No direct Hg inputs. Likely sources:	Μ	0.25–0.42 FW	~90%
		at Owyhee Reservoir, SW	mercuriferous soil/rock, geothermal			
		Oregon, USA.	venting and historic gold			
			and suver mining.			
Brook trout (S. fontinalis)	This work	Northern Patagonian lakes	No direct anthropogenic Hg inputs	M 1	0.12-4.0 DW	
				L	0.14-7.1 DW	
	[52]	Herrington Creek (HC) tributary	(BR) has high acid neutralizing	M		0.04 FW (BR)
		and Blacklick Run (BR) streams,	capacity=high pH system, (HC)			
		W Maryland	low pH system.			0 06 EW (HC)
						0.00 F W (FLC)
Lake trout (Salvelinus namaycush)	[53]	British Columbia, Ontario, Quebec, New York		W	1.1–10.5 FW	
				Μ	0.3–1.3 FW	
	[36]	Canadian Subartic and Artic	Mackenzie river basin and other	Μ	0.17–0.79 FW	
		freshwater fish	boreal lakes			
Trout	[41]	Wilderness lakes, northern Maine		Μ	0.1–0.5 FW	
Creole Perch (P. trucha)	This work	Northern Patagonia	No direct anthropogenic Hg inputs	Μ	0.25–3.2 DW	
				L	0.07–1.5 DW	
Walleye (Stizostedion vitreum)	[37]	Lake Oahe, South Dakota, Foster	Suspected contaminated waters	Μ	0.33-0.50 FW (FB)	
		Bay (FB), Grand River (GR), Moreau River (MR)				
				Γ	0.05-0.20 FW(FB)	
				M	0.27–0.54 FW (GR)	
				L	0.09-0.22 FW (GR)	
				М	0.25-0.42 FW (MR)	

 Table 10
 Mercury Contents in Fish from the Present Work Compared to Other Researches

				L	0.04-0.17 FW (MR)	
	[36]	Canadian Subartic and Artic	Mackenzie river basin and other	Μ	0.07–1.4 FW	
		freshwater fish	boreal lakes			
Yellow perch (Perca flavescens)	[54]	Ontario, Canada	Uncontaminated	Μ	0.031-0.233 FW	
	[46]	Lake survey,	24 not heavily impacted lakes		0.01-0.75 FW	
		Massachusetts, USA.				
	[51]	Owyhee River drainage basin	No direct Hg inputs. Likely sources:	Μ	0.29–1.74 FW	~90%
		at Owyhee Reservoir, SW	mercuriferous soil/rock, geothermal			
		Oregon, USA.	venting and historic gold and			
			silver mining.			
	[55]	Several US Mid West lakes		Μ	0.16 - 0.26	%66
	[56]	Lillinonah lake, Connecticut		Μ	0.23–0.74 FW	
	[57]	Plastic lake (acidic), Ontario,	Remote from urban centers and	Μ	0.59±0.20 DW (PL)	
		Canada	point-source HG emitting industries			
Perch (Perca fluviatilis)	[2]	Lake Öltertjärn, N Sweden	With controlled addition of sodium	М	0.09-0.29 FW t0	
			selenite to the water (t1)			
					0.02-0.17 FW tl	
Velvet catfish (D.viedmensis)	This work	Northern Patagonian lakes	No direct anthropogenic Hg inputs	Μ	0.055–2.3 DW	
				L	0.09–3.9 DW	
Channel catfish (Ictalurus punctatus)	[53]	Lake Erie		Μ	0.3–1.8 FW	
		Lake St. Clair, Ohio, Illinois,		Μ	0.02–2.5 FW	
		Oregon, Georgia, Texas				
Channel catfish	[51]	Owyhee River drainage basin at	No direct Hg inputs. Likely Hg	M	0.30–1.47 FW	~90%
		Owyhee Reservoir, SW	sources: mercuriferous soil/rock,			
		Oregon, USA.	geothermal venting and historic			
			Au anu Ag mung.			
Omnivore catfish (<i>Hepteropneusptes fossilis</i>)	[58]	Rivers, estuaries and lakes from Bangladesh	Sites with absence of mining and industrial activities	W	0.018-0.083 FW	$80 \pm 10\%$
Piscivore catfishes (Mystus					0.029–0.427 FW	$90 \pm 18\%$
seenghala, Silonia silondia, Wallago attu)						

^a M muscle; L liver, W whole, FW fresh weight, DW dry weight. Dry to wet mass ratios for this work muscle samples range from 0.20 to 0.27.

lowest measured in other parts of the world. It is noteworthy to mention that Walleye, a perciforme from the North hemisphere, shows Hg liver to muscle content ratios smaller than 1, as is the case with the Patagonian creole perch. Catfish Hg contents in muscle are similar to other catfishes of the world living in water bodies without direct Hg inputs.

Considering that most Hg in muscle is in the organic form, few fish (creole perch, velvet catfish, and brook trout) exceeded Argentinian recommendation for methylmercury intake of freshwater fisheries consumption (0.5 μ g g⁻¹ FW, or approximately 2 μ g g⁻¹ DW) and the US Environmental Protection Agency's health advisory for freshwater fish limited consumption.

Selenium and Mercury

There is evidence that selenites of Hg are insoluble and reduce Hg and Se bioavailability. Se and Hg contents in muscle were not correlated (ρ ranging from -0.36 to 0.46), as well as Se and Hg contents in livers from rainbow trout, brook trout, and creole perch (ρ <0.3), and weakly correlated for livers of brown trout and catfish (ρ equal to 0.65 and 0.51, respectively). This lack of correlation has previously been observed in omnivorous and piscivorous fish [38].

Total Hg to Se molar ratios in the present work range from 0.04 to 0.4 for brown trout, 0.06 to 0.9 for rainbow trout, 0.03 to 1.2 for creole perch, and 0.07 to 0.4 for velvet catfish. Except for one unique muscle sample, Se always exceeded Hg, a fact which is consistent with the findings in other uncontaminated freshwater ecosystems [59–61]. In rivers contaminated by gold mining activities, Hg is typically in excess of Se [38].

Concentrations of Se in all food items from lake Moreno range between 0.21 and 3.2 $\mu g g^{-1}$ DW, with only insect larvae above 3 $\mu g g^{-1}$ DW, the aquatic ecological risk threshold for diet. Se contents in pelletized food lies between these limits. Both wild and farmed trout have similar Se contents in muscle, although Se content in wild trout liver is at least four times higher. Contents of Hg in food items was variable, from 0.08 to 1.4 $\mu g g^{-1}$ DW, as well as Hg contents in different aliquots of pelletized food (ranging from 0.02 to 1 $\mu g g^{-1}$ DW, see Table 8). The previous results do not allow to draw conclusions on Hg antagonism or synergism based on the two different diets of rainbow trout.

It should be noted that the species that have the lowest Hg contents in muscle, brown trout and rainbow trout, have the highest Se contents in liver, which could be an indication that these fish are protected from Hg by a Se-rich diet.

Conclusions

The following conclusions were drawn from this study:

- Hg contents in liver and muscle tissue are higher than those expected for pristine lakes; however, the values are not unusual. The most remarkable result is that the species considered as the top predator does not have the highest contents of Hg in muscle.
- The higher Hg contents in muscle from creole perch are in coincidence with the lower Se in liver. Evidence would indicate that brown trout and rainbow trout (with higher Se contents in liver) have Se-rich diets, which could be preventing Hg accumulation in muscle.
- Selenium contents in liver of brown trout and rainbow trout are well above the toxicity threshold; however, all fish analyzed belonged to populations which did not appear stunted. More studies are necessary to assess the impact on the fish population.

- Se contents in muscle of fish from lakes Moreno and Nahuel Huapi are about one half of the Se contents in muscle of fish from the other lakes. This geographical pattern, although less obvious, appears for Se contents in liver.
- The use of Se compounds (sodium selenite) as a feeding supplement for the fish at the trout farm at lake Moreno East does not seem to affect the wild fish population.

Acknowledgments The authors wish to thank the RA-6 reactor operation staff for their assistance during the irradiations. This work was carried out within the Technical Cooperation Agreement ARG/7/006 with the International Atomic Energy Agency (IAEA)

References

- Eisler R (1999) Selenium hazards to fish, wildlife, and invertebrates: a synoptic review, in Contaminant Hazard Reviews, Report 5, USGS/BRD/BSR-1999-0002
- Presser T, Sylvester M, Low W (1994) Bioaccumulation of selenium from natural geological sources and its potential consequences. Environ Manage 18:423–436
- 3. Presser T (1994) The Kesterton effect. Environ Manage 18:437-454
- Lemly AD (1998) Pathology of selenium poisoning in fish. In: Frankenberger WT, Engberg RR (eds) Environmental chemistry of selenium. Marcel Dekker, New York
- Sorensen EMB, Cumbie PM, Bauer TL, Bell JS, Harlan CW (1984) Histopathological, hematological, condition-factor, and organ weight changes associated with selenium accumulation in fish from Belews Lake, North Carolina. Arch Environ Contam Toxicol 13:153–162
- Eisler R (1999) Mercury hazards to fish, wildlife, and invertebrates: a synoptic review, in Contaminant Hazard Reviews, Report 10, USGS/BRD/BSR-1999-0002
- Paulsson K, Lundbergh K (1991) Treatment of mercury contaminated fish by selenium addition. Water Air Soil Pollut 56:833–841
- EPA (1997) Mercury report to congress. United States Environmental Protection Agency, EAP-425/R-97-003 and 005
- 9. UNEP (2002) Global mercury assessment. United Nations Environmental Program Report, Geneva
- 10. Sigel A, Sigel H (1997) Mercury and its effects on environment and biology. Marcel Dekker, New York
- Rudd JW, Turner MA (1983) Selenium in lake enclosures: its geochemistry, bioaccumulation and ability to reduce mercury bioaccumulation. Can J Fish Aquat Sci 40:2228–2250
- Fjield E, Rognerud S (1993) Use of path analysis to investigate mercury accumulation in brown trout (Salmo trutta) in Norway and the influence of environmental factors. Can J Fish Aquatic Sci 50:1158–1167
- Southworth GR, Peterson MJ, Ryon MG (2000) Long-term increased bioaccumulation of mercury in largemouth bass follows reduction of waterborne selenium. Chemosphere 41:1101–1105
- Bjerregaard P, Andersen D, Rankin JJ (1999) Retention of methylmercury in rainbow trout, Oncorhynchus mykiss: effect of dietary selenium. Aquatic Toxicol 45:171–180
- Cuvin Aralar MA, Furness R (1991) Mercury and selenium interaction: a review. Ecotox Environ Saf 21:348–364
- Chen YW, Belzile N, Gunn JM (2001) Antagonistic effect of Selenium on mercury assimilation by fish populations near Sudbury metal smelters. Limnol Oceanogr 46(7):1814–1818
- Belzile N, Chen YW, Gunn JM, Tong J, Alarie Y, Delonchamp T, Lang CY (2006) The effect of selenium on mercury assimilation by freshwater organisms. Can J Fish Aquat Sci 63:1–10
- Heinz GH, Hoffman DJ (1998) Methylmercury chloride and selenomethionine interactions on health and reproduction in mallards. Environ Toxicol Chem 17(2):139–145
- Heinz GH, Hoffman DJ (1998) Effects of mercury and selenium on glutathione metabolism and oxidative stress in mallard ducks. Environ Toxicol Chem 17(2):161–166
- Bubach DF, Arribére MA, Ribeiro Guevara S, Calvelo S (2001) Study on the feasibility of using transplanted *Protousnea magellanica* thalli as bioindicators of atmospheric contamination. J Radioanal Nuc Chem 250:63–68
- Ribeiro Guevara S, Massaferro J, Villarosa G, Arribére MA, Rizzo AP (2002) Heavy metal contamination in sediments of lake Nahuel Huapi, Nahuel Huapi National Park, Northern Patagonia, Argentina. Water Air Soil Pollut 137:21–44
- Ribeiro Guevara S, Bubach DF, Vigliano PH, Lippolt G, Arribére MA (2004) Heavy metals and other trace elements in native mussel *Diplodon chilensis* from Northern Patagonian lakes, Argentina. Biol Trace Elem Res 102(1–3):245

- Ribeiro Guevara S, Bubach DF, Arribére MA (2004) Mercury in lichens of Nahuel Huapi National Park, Patagonia, Argentina. J Radioanal Nuc Chem 261(3):679–687
- Ribeiro Guevara S, Arribére MA, Bubach DF, Vigliano P, Rizzo A, Alonso M, Sánchez R (2005) Silver contamination on abiotic and biotic compartments of lake Nahuel Huapi National Park lakes, Patagonia, Argentina. Sci Total Environ 336(1–3):119–134
- Ribeiro Guevara S, Rizzo A, Sánchez RS, Arribére MA (2005) Heavy metal inputs in Northern Patagonia lakes from short sediment core analysis. J Radioanal Nuc Chem 265(3):481–493
- 26. Calcagno A, Fioritti MJ, Pedrozo F, Vigliano PH, López H, Rey C, Razquin ME, Quirós R (1995) Catálogo de lagos y embalses de la República Argentina. Ministerio de Economía y Obras y Servicios Públicos, Secretaría de Obras Públicas, Subsecretaría de Recursos Hídricos, Argentina
- Pascual M, Macchi PJ, Urbanski J, Marcos F, Riva Rossi C, Novara M, Dell' Arciprete P (2002) Evaluating potential effects of exotic freshwater fish from incomplete species presence-absence data. Biol Invasions 4:101–113
- 28. Vigliano PH, Alonso MF, Denegri MA, Garcia Asorey MI, Lippolt G, Macchi PJ, Milano D (2001) Estructura de las comunidades de peces de lagos y embalses patagónicos: estado actual del conocimiento y problemática. Proc. I Encuentro Binacional de Ecología, XX Reunión Argentina de Ecología and X Reunión de la Sociedad de Ecología de Chile, Bariloche, Argentina
- Vigliano PH, Alonso MF (2007) Salmonid introductions in Patagonia Argentina: a mixed blessing. In: Bret TM (ed) Ecological and genetic implications of aquaculture. Springer, The Netherlands
- Macchi PJ, Cussac VE, Alonso MF, Denegri MA (1999) Predation relationships between introduced salmonids and the native fish fauna in lakes and reservoirs in Northern Patagonia. Ecol Freshw Fish 8:227–236
- Ribeiro Guevara S, Bubach DF, Macchi PJ, Vigliano PH, Arribére MA, Colombo JC (2006) Rb–Cs ratio as an indicator of fish diet in lakes of the Patagonia, Argentina. Biol Trace Elem Res 110:97–119
- 32. Vigliano PH, Macchi PJ, Denegri MA, Alonso MF, Milano D, Lippolt G, Padilla G (1999) Un diseño modificado y procedimiento de calado de redes agalleras para estudios cuali-cuantitativos de peces por estratos de profundidad en lagos araucanos. Natura Neotropicalis 30(1–2):1–11
- 33. Arribére MA, Ribeiro Guevara S, Bubach DF, Vigliano PH (2006) Trace elements as fingerprint of lake of provenance and of species of some native and exotic fish of northern Patagonian lakes. Biol Trace Elem Res 110:71–95
- Ramirez Jr, P, Dickerson K (1997) Follow-up investigation of selenium and other trace elements in biota from the Riverton Reclamation Project, Fremont County, Wyoming, Contaminant Report Number: R6/ 709C/97
- 35. Kaiser II, Young P, Johnson JD (1979) Chronic exposure of trout to waters with naturally high selenium levels: effects on transfer RNA methylation. J Fish Res Board Canada 36:689–694
- 36. Evans MS, Muir D, Lockhart WL, Stern G, Ryan M, Roach P (2005) Persistent organic pollutants and metals in the freshwater biota of the Canadian Subarctic and Arctic: an overview. Sci Total Environ 351– 352:94–147
- Mauk RJ, Brown ML (2001) Selenium and mercury concentrations in brood-stock walleye collected from three sites on Lake Oahe. Arch Environ Contam Toxicol 40:257–263
- Dorea JG, Moreira MB, East G, Barbosa AC (1998) Selenium and mercury concentrations in some fish species of the Madeira river, Amazon basin, Brazil. Biol Trace Elem Res 65:211–220
- Stewart AR, Luoma SN, Schlekat CE, Doblin MA, Hieb KA (2004) Food web pathway determines how selenium affects aquatic ecosystems: a San Francisco Bay case study. Environ Sci Technol 38:4519–4526
- Barwick M, Maher W (2003) Biotransference and biomagnification of selenium, copper, cadmium, zinc, arsenic and lead in a temperate seagrass ecosystem from Lake Macquarie Estuary, NSW, Australia. Mar Environ Res 56:471–502
- Akielaszak JJ, Haines TA (1981) Mercury in the muscle tissue of fish from three northern Maine lakes. Bull Environ Contam Toxicol 27:201–208
- Sloan R, Schofield C (1983) Mercury levels in brook trout (*Salvelinus fontinalis*) from selected acid and limed Adirondack lakes. Northeast Environ Sci 2:165–170
- Vuorinen PJ, Rantio T, Witick A, Vuorinen M (1994) Organochlorines and heavy metals in sea trout (Salmo trutta) in the gulf of Bothnia off the coast of Finland. Aqua Fennica 24(1):29–35
- 44. Kim JP (1995) Methylmercury in rainbow trout (*Oncorhynchus mykiss*) from Lakes Okareka, Okaro, Rotomahana, Rotorua and Tarawera, North Island, New Zealand. Sci Total Environ 164:209–219
- 45. Kim JP, Burggraaf S (1999) Mercury bioaccumulation in rainbow trout (*Oncorhynchus mykiss*) and the trout food web in lakes Okarera, Okaro, Rotomahana and Rotorua, New Zealand. Water Air Soil Pollut 115:535–546
- Rose J, Hutcheson MS, West CR, Pancorbo O, Hulme K, Cooperman A, DeCesare G, Isaac R, Screpetis A (1999) Fish mercury distribution in Massachusetts, USA lakes. Environ Toxicol Chem 18(7):1370–1379

- Newman MC, Jagoe CH (1994) Ligands and the bioavailability of metals in the aquatic environments. In: Hamelink JM, Landrum PF, Bergman HL, Benson WH (eds) Bioavailability. CRC Press, Boca Raton, USA
- 48. Huckabee JW, Elwood JW, Hildebrand SG (1979) Accumulation of mercury in freshwater biota. In: Nriagu J (ed) The biogeochemistry of mercury in the environment. Elsevier, Amsterdam
- 49. Jobling M (1993) Bioenergetics: Feed intake and energy partitioning. In: Rankin JC, Jensen FB (eds) Fish ecophysiology. Chapman and Hall, USA
- Bridges CR (1993) Ecophysiology of intertidal fish. In: Rankin JC, Jensen FB (eds) Fish ecophysiology. Chapman and Hall, USA
- Neumann CM, Kauffman KW, Gilroy DJ (1997) Methylmercury in fish from Owyhee Reservoir in Southeast Oregon: scientific uncertainty and fish advisories. Sci Total Environ 204:205–214
- Mason RP, Laporte JM, Andres S (2000) Factors controlling the bioaccumulation of mercury, methylmercury, arsenic, selenium and cadmium by freshwater invertebrates and fish. Arch Environ Contam Toxicol 38:283–297
- Jenkins DW (1980) Biological monitoring of toxic trace metals. Vol. 2. Toxic trace metals in plants and animals of the world, Part 1. U.S. Environmental Protection Agency Rep. 600/3-80-090:30–138
- Suns K, Hitchin G (1990) Interrelationships between mercury levels in yearling yellow perch, fish condition and water quality. Water Air Soil Pollut 650:255–265
- 55. Bloom NS (1992) On the chemical form of Hg in edible fish and marine invertebrate tissue. Can J Fish Aquatic Sci 49(5):1010–1017
- Neumann RM, Ward SM (1999) Bioaccumulation and biomagnification of mercury in two warm water fish communities. J Freshw Ecol 14(4):487–497
- Scheuhammer AM, Graham JE (1999) The bioaccumulation of mercury in aquatic organisms from two similar lakes with differing pH. Ecotoxicol 8:49–56
- Holsbeek L, Das HK, Joiris CR (1997) Mercury speciation and accumulation in Bangladesh freshwater and anadromous fish. Sci Total Environ 198:201–210
- Burger J, Cooper K, Gochfeld M (1992) Exposure assessment for heavy metal ingestion from a sport fish in Puerto Rico: estimating risk for local fishermen. J Toxicol Environ Health 36:355–365
- Sindayigaya E, Van Cauwenbergh R, Robberecht H, Deelstra H (1994) Copper, zinc, manganese, iron, lead, cadmium, mercury and arsenic in fish from Lake Tanganika, Burundi. Sci Total Environ 144: 103–115
- Benemariya H, Robberecht H, Deelstra H (1994) Atomic absorption spectrometric determination of Zinc, Copper and Selenium in fish from Lake Tanganika, Burundi, Africa. Sci Total Environ 144:103–115