**ORIGINAL ARTICLE**



# **Evidence from Co‑expression Analysis for the Involvement of Amidase and INS in the Tryptophan‑Independent Pathway of IAA Synthesis in** *Arabidopsis*

YousefM. Abu-Zaitoon<sup>1</sup><sup>1</sup> • Ahmed Abu-Zaiton<sup>2</sup> • Abdel Rahman Al Tawaha<sup>2</sup> • Khalid Ghazi Fandi<sup>1</sup> • Sulaiman M. Alnaimat<sup>1</sup> • Siddhartha Pati<sup>3</sup> • Fouad A. Almomani<sup>4</sup>

Accepted: 26 June 2022 / Published online: 8 July 2022 © The Author(s), under exclusive licence to Springer Science+Business Media, LLC, part of Springer Nature 2022

### **Abstract**

The reverse genetic approach has uncovered indole synthase (INS) as the frst enzyme in the tryptophan (trp)-independent pathway of IAA synthesis. The importance of INS was reevaluated suggesting it may interact with tryptophan synthase B (TSB) and therefore involved in the trp-dependent pathway. Thus, the main aim of this study was to clarify the route of INS through the analysis of *Arabidopsis* genome. Analysis of the top 2000 co-expression gene lists in general and specifc conditions shows that *TSA* is strongly positively co-expressed with *TSB* in general, hormone, and abiotic conditions with mutual ranks of 89, 38, and 180 respectively. Moreover, *TSA* is positively correlated with *TSB* (0.291). However, *INS* was not found in any of these coexpressed gene lists and negatively correlated with *TSB* (−0.046) suggesting unambiguously that these two routes are separately and independently operated. So far, the remaining steps in the INS pathway have remained elusive. Among all enzymes reported to have a role in IAA synthesis, *amidase* was found to strongly positively co-expressed with *INS* in general and light conditions with mutual ranks of 116 and 141 respectively. Additionally, *amidase1* was found to positively correlate with *INS* (0.297) and negatively coexpressed with *TSB* concluding that amidase may exclusively involve in the trp-independent pathway.

**Keywords** Amidase · Co-expression · Indole synthase · Tryptophan synthase A · Tryptophan synthase B

 $\boxtimes$  Yousef M. Abu-Zaitoon yousefaz@yahoo.com

<sup>&</sup>lt;sup>1</sup> Department of Biology, Al-Hussein Bin Talal University, Maan, Jordan

<sup>&</sup>lt;sup>2</sup> Department of Biology, Faculty of Science, Al-Albayt University, Mafraq, Jordan

<sup>3</sup> NatNov Bioscience Private Limited, Balasore 756001, India

<sup>4</sup> Department of Applied Biology, Jordan University of Science and Technology, Irbid, Jordan

## **Introduction**

Auxin is a key signaling phytohormone regulating almost all aspects of plant growth and development from embryogenesis to senescence [\[1\]](#page-8-0). Indole-3-acetic acid (IAA), the main naturally occurring auxin in the plant kingdom, has long been proposed to be synthesized from both tryptophan through trp-dependent pathways (Fig. [1\)](#page-1-0) and directly from tryptophan precursors through a trp-independent pathway [\[2,](#page-8-1) [3](#page-8-2)].

Biochemical and genetic experiments have recently revealed the indole pyruvic acid (IPA) pathway as the frst complete route of IAA synthesis via the trp-dependent pathway. In this simple two-step pathway, tryptophan aminotransferase (TAA) converts tryptophan into IPA, whereas YUCCA, a favin-containing monooxygenase, catalyzes the ratelimiting reaction by converting IPA into IAA [\[4](#page-8-3), [5](#page-8-4)]. Additionally, indole-3-acetaldoxime (IAOx) has been demonstrated as a second trp-dependent pathway [[6,](#page-8-5) [7\]](#page-8-6). Even though the cytochrome P450 monooxygenase CYP79B2/CYP79B3 has been suggested to produce IAOx from tryptophan, the remaining intermediates and enzymes are still largely unidentifed. Furthermore, aldehyde oxidase (AAO), nitrilase (NIT), and amidase convert indole-3-acetaldehyde (IAAld), indole-3-acetonitrile (IAN), and indole-3-acetamide (IAM) to IAA. They all were recognized as naturally occurring enzymes. However, pathways including these enzymes are yet to be characterized  $[8-11]$  $[8-11]$ .

Three decades ago, a combination of labeling, genetic, and metabolic approaches led to the proposal that IAA is biosynthesized from tryptophan precursors via the



<span id="page-1-0"></span>**Fig. 1** Potential IAA biosynthesis pathways in *Arabidopsis.* CHA, chorismate; ANT, anthranilate; IGP, indole-3-glycerol phosphate; TSA, tryptophan synthase A; TSB, tryptophan synthase B; Trp, tryptophan; INS, indole synthase; IAOx, indole-3 acetaldoxime; IAN, indole-3-acetonitrile; IAM, indole-3-acetamide; TAA, tryptophan aminotransferase of *Arabidopsis*; IPA, indole-3-pyruvic acid; YUCCA, a Flavin monooxygenase; IAA, indole-3-acetic acid. Enzymes are indicated in blue color, whereas intermediates are in black. Arrows in red colors refer to undefned or unconfrmed enzymes, whereas black colors refer to identifed enzymes

trp-independent pathway [[12](#page-8-9), [13\]](#page-8-10). This pathway was suggested to be ubiquitous as it was proposed to synthesize IAA in broad taxonomic groups of plants, including *Arabidopsis*, tomato, maize, carrot, *Lemnagibba*, and bean [\[14](#page-8-11)–[18](#page-8-12)]. Recently, indole severe sensitive 1 mutant *(iss1)* was described to display trp-independent IAA synthesis as well as related physiological activities including longer petioles, adventitious roots, and leaf narrowing [\[19](#page-9-0)]. Six years ago, feeding and genetic approaches were uncovered INS (At4g02610) as the frst enzyme in the trp-independent pathway and an essential role for this pathway in early embryogenesis was suggested [[20\]](#page-9-1). The cytosolic INS was demonstrated to regulate hypocotyl length at high temperature in concert with the IPA pathway in *Arabidopsis* based on the observation that *ins-1* mutants are not 100% penetrant and have an additive efect with *wei8* (weakly ethylene insensitive8) mutants. Wang et al. [[20\]](#page-9-1) hypothesis has been questioned by Nonhebel bioinformatics work. The possibility for the INS or its product, indole, to interact with TSB and therefore with the trp-dependent pathway was not excluded [[21](#page-9-2)]. The additive efect of *wei8* and *ins* was further used to support this hypothesis. Therefore, the involvement of INS in either trp-dependent or independent pathways was considered in this article. In this pathway, indole-3-glycerol phosphate is converted to indole by the indole synthase in the cytosol [\[20](#page-9-1)]. However, enzymes catalyzing the remaining steps have not been identifed so far. Therefore, this piece of work was aimed to fnd out enzymes potentially act with INS in the INS pathway of IAA synthesis. All previously reported genes to have roles in IAA synthesis were investigated in the list of genes most strongly positively co-expressed with *INS*. Additionally, a list of genes negatively co-expressed with *INS* was also studied to shorten the list of genes most probably act with INS in the trpindependent pathway.

Coexpression analysis is a powerful tool that has been used for identifying genes related to a specific metabolic pathway  $[22]$  $[22]$ . For instance, this approach has been adopted to identify genes regulating cellulose synthesis and transcription factors involved in the glucosinolate synthesis pathway [[23,](#page-9-4) [24](#page-9-5)]. In this research, ATTED-II, a web-based tool, was used as a secondary database to analyze publicly available microarray data for the dicot model plant *Arabidopsis thaliana* [[25](#page-9-6)]*.* Unlike Hirai et al. [[24](#page-9-5)], who used Pearson's correlation coefficient for combinations of *Arabidopsis* genes available in ATTED-II, mutual rank (MR) as a powerful measure was used in the latest release of the database. Co-expressed gene relationships are primarily based on the vast amount of microarray data stored in TAIR as well as NASCArrays [\[26](#page-9-7)].

#### **Methods**

The co-expression analysis tool (CoExSearch: co-expressed gene list from multiple query genes) available on the ATTED-II (<http://atted.jp>) database version 9.2 was used to fnd genes positively and negatively co-expressed with the *Arabidopsis INS* and *TSB* genes. Genes previously reported to have a possible role in IAA synthesis are listed in Table [1](#page-3-0). The available top 2000-gene lists positively and 300-gene lists negatively co-expressed with seed genes were studied and analyzed. The four *TSB* genes in *Arabidopsis* (At5g38530, At5g28237, At4g27070, and At5g54810) were uploaded into the CoExSearch tool of ATTED-II. Afy data were only available to *TSB* (At5g38530).

<span id="page-3-0"></span>**Table 1** Summary of IAA-synthesis genes previously reported having a role in IAA synthesis. *INS*, indole synthase; *TSA*, tryptophan synthase A; *TSB*, tryptophan synthase B; *YUCCA*, a favin monooxygenase; *TAA*, tryptophan aminotransferase of *Arabidopsis*; *TAR*, tryptophan aminotransferase related; *AAO*, aldehyde oxidase; *NIT*, nitrilase

Gene	Locus number	Gene	Locus number	Gene	Locus number
INS	At $4g02610$	AAO <sub>2</sub>	At3g43600	TAR4	At1g34060
<b>TSA</b>	At3g54640	AAO3	At2g27150	YUCCA1	At4g32540
<b>TSB1</b>	At5g54810	AAO4	At1g04580	YUCCA2	At4g13260
TSB <sub>2</sub>	At4g27070	NIT 1	At3g44310	YUCCA3	At1g04610
TSB3	At5g28237	NIT <sub>2</sub>	At3g44300	<i>YUCCA4</i>	At5g11320
TSB4	At5g38530	NIT3	At3g44320	YUCCA5	At5g43890
Amidase1	At1g08980	NIT4	At5g22300	<i>YUCCA6</i>	At5g25620
Amidase2	At5g07360	CYP79B2	At4g39950	<i>YUCCA7</i>	At2g33230
Amidase3	At5g64440	CYP79B3	At2g22330	YUCCA8	At4g28720
Amidase4	At5g09420	TAA1	At1g70560	YUCCA9	At1g04180
Amidase5	At4g34880	TAR 1	At1g23320	<i>YUCCA10</i>	At1g48910
Amidase6	At3g25660	TAR <sub>2</sub>	At4g24670	YUCCA11	At1g21430
AAO1	At5g20960	TAR3	At1g34040		

# **Results and Discussion**

## **Tryptophan Synthase A but Not Indole Synthase Interacts with Tryptophan Synthase B to Produce Tryptophan**

A signifcant role for the involvement of indole synthase in the trp-independent pathway of IAA synthesis has been quite recently reported. The cytosolic INS was suggested to produce indole from the indole-3-glycerol phosphate [[20](#page-9-1), [27\]](#page-9-8). However, this suggestion was challenged by a proposal that INS may interact with TSB to produce tryptophan and therefore with the trp-dependent pathway [\[21\]](#page-9-2). In this work and through the implementing co-expression analysis of *Arabidopsis* genome obtained from the ATTED-II secondary database [\[25\]](#page-9-6), we excluded the possibility of INS's interaction with TSB. Among the 21,000 genes analyzed, the available 2000 gene list correspond to 90% cut-of value positively signifcantly co-expressed with TSB in general conditions (condition-independent manner) were obtained and studied. Furthermore, co-expressed gene lists obtained from specifc samples under certain conditions, including hormone, tissue, biotic, and abiotic stress, were also obtained and investigated. Table [2](#page-4-0) shows a short list of all genes related to IAA synthesis signifcantly positively co-expressed with *TSB* under general and specifc conditions.

Indole synthase gene was found neither in the general nor in the specifc co-expression data lists, suggesting that this enzyme may not interact with TSB to produce IAA via the trp-dependent pathway. In contrast to *INS*, *TSA* which its product forms a complex with TSB [[28](#page-9-9)], was strongly co-expressed with the query gene with mutual ranks of 38, 98, and 180 in the hormone, general, and abiotic conditions respectively. Interestingly, *INS* was not also found in the top 300 gene list negatively co-expressed with *TSB*, suggesting that trpdependent and independent pathways may interact to produce IAA in *Arabidopsis*. Another evidence for the involvement of INS in the trp-independent pathway came from the analysis of correlation patterns. Figure [2](#page-4-1) shows the correlation of expression patterns for *INS*



<span id="page-4-0"></span>



<span id="page-4-1"></span>**Fig. 2** Correlation of the expression obtained from developmental and vegetative samples for *INS* and *TSB* (**A** and **B**) and for *TSA* and *TSB* (**C** and **D**)

 $\bigcirc$  Springer

and *TSB* (A and B) as well as *TSA* and *TSB* (C and D) for samples obtained from both developmental and vegetative tissues, respectively. The correlation of expression for INS and TSB is −0.046, whereas for TSA and TSB is 0.291 is further supports the involvement of INS in the trp-independent pathway of IAA synthesis. This suggestion is in agreement with the independent production of indole by TSA and INS suggested in maize [[27](#page-9-8)].

#### **Amidase May Interact with INS in the trp‑Independent Pathway**

Even though Wang et al. [\[20\]](#page-9-1) reported a significant role for INS in IAA production through the trp-independent pathway, the remaining reaction(s) in this pathway needs to be identifed. Functional identifcation of gene(s) expected to work in the same pathway of IAA synthesis with INS was also attempted in this study using the publically available coexpression database ATTED-II [\[25\]](#page-9-6). Indole synthase was used as a seed gene to search for other possible genes in the tryptophan-independent pathway of IAA synthesis. Among all genes previously reported to be involved in IAA synthesis, *amidase1* (At1g08980) was strongly co-expressed with *INS* in general and light conditions with mutual ranks of 116 and 141 respectively (Table [3\)](#page-5-0). Additionally, *Amidase1* was found to positively correlate with *INS* (0.297) (Fig. [3\)](#page-6-0). This correlation value was similar to that found between *TSA* and *TSB*; their products constitute an enzyme complex involved in the production of trp from indole-3-glycerol phosphate. Taken together and in addition to the observation that this gene was not found to positively co-expressed with *TSB*, suggests that amidase1 may be exclusively involved in the trp-independent pathway of IAA synthesis. Finding *amidase1* in the top 300 gene list negatively co-expressed with *TSB* (MR 20,628) further supports this hypothesis. It concludes that amidase may catalyze the rate-limiting step in this pathway. These findings are in agreement with the observation that Amidase1 is specifically convert IAM to IAA (*K*m 972 µM) [\[29\]](#page-9-10). Furthermore, the importance of amidase1 in IAA synthesis has been quite recently suggested by Gao et al. [[30](#page-9-11)].

Amidase5 (At4g34880), an *Arabidopsis* amidase member that functionally not characterized, was co-expressed with *INS* in the general and light conditions with a MR of 1000 and 926 respectively. However, statistical supportability of the co-expression relation for this gene was found to be weak. The same thing was observed for the expression of this gene with  $TS\beta$  in the tissue condition (MR 2471) but again with low statistical supportability suggesting that the gene product may be involved in the production of substance other than IAA. Amidase2 (At5g07360) which was proposed to convert substrate other than indole-3-acetamide (IAM) [\[31\]](#page-9-12) was co-expressed with  $TS\beta$  in the general, tissue, biotic, and abiotic specifc conditions whereas co-expressed with *INS* in the abiotic conditions

<span id="page-5-0"></span>





<span id="page-6-0"></span>**Fig. 3** Correlation of the expression for *INS* and *amidase1* obtained from developmental samples (**A**) and vegetative samples (**B**)

only. Amidase6 (At3g25660), another uncharacterized *Arabidopsis* amidase member, was co-expressed only with *TSβ* in the light condition (MR 1419).

#### **Role of Aldehyde Oxidase and Nitrilase in IAA Synthesis**

Condition-dependent and independent co-expression lists were also searched for previously genes reported to be involved in IAA biosynthesis. A group of IAA synthesis genes including *NIT*, *AAO*, *YUC*, and *CYP79B2/B3* is positively co-expressed with *TSB*. The nitrilase isoform (*NIT3)*, which was reported exclusively in Brassicaceae [[32](#page-9-13), [33](#page-9-14)], is co-expressed with *TSB* in all but biotic conditions (Table [2](#page-4-0)). Through the analysis of *NIT2*RNAi lines, it has been shown that the level of IAA is considerably reduced due to non-detected NIT1-3 [[34](#page-9-15)] suggesting that NIT3 may participate in IAA synthesis through the trp-dependent pathway. Additionally, *NIT4* found in monocot and dicot plants is strongly co-expressed with *TSB* in hormone conditions in addition to general and biotic conditions (Table [2](#page-4-0)). NIT4 is primarily involved in cyanide detoxifcation [[35](#page-9-16)]. These controversial data need to be clarifed for a possible role of NIT4 in IAA synthesis, particularly in the trp-dependent pathway. None of the *NIT* isoforms was found to co-expressed with *INS* suggesting that this enzyme may not have any possible role in IAA production through the INS pathway.

Aldehyde oxidase  $\alpha$  and  $\gamma$  are homodimers of AAO1 and AAO2 respectively whereas aldehyde oxidase  $\beta$  is a heterodimer of AAO1 and AAO2. They have shown to have limited preferences for abscisic aldehyde [\[36\]](#page-9-17). In this work, co-expression of *INS* was noted only with  $AAO2$  in the hormone-specific condition, suggesting a possible role for  $AO\gamma$  in IAA biosynthesis through the trp-independent pathway. However, aldehyde oxidase1, 2, and 3 were found to co-express with *TSB*. Aldehyde oxidase1 is co-expressed with *TSB* in all analyzed general and specifc conditions, whereas *AAO2* co-expressed with *TSB* in the tissue, abiotic and light conditions with mutual ranks of 328, 1456, and 302, respectively. Therefore, a possible role for these enzymes in IAA synthesis through the trp-dependent pathway is not excluded. Additionally, *AAO3* product which was reported to oxidize

abscisic aldehyde efficiently is co-expressed with *TSB* in general, hormone, and light conditions. This fnding may explain the crosstalk between IAA and ABA hormones.

# **Conclusion**

The trp-independent pathway was earlier proposed as a major route of IAA biosynthesis in *Arabidopsis* based on metabolites quantifcation and feeding experiments in trp biosynthesis mutants. More recent evidence supporting the involvement of the trp-independent pathway in auxin biosynthesis through INS was further illustrated. However, the interaction of INS with TSB to produce trp and, as a result, a possible role for this enzyme in the trpdependent pathway was also suggested based mainly on bioinformatics studies. Therefore, the main aim of this work was to collect evidence through the implementation of coexpression analysis of the *Arabidopsis* genome for the specifc role of INS in auxin biosynthesis. Among the 21,000 genes of the *Arabidopsis* genome available on the ATTED-II ([http://](http://atted.jp) [atted.jp](http://atted.jp)) database, *TSA* was found to be strongly positively coexpressed with *TSB* in the top 2000 gene lists of general, hormone, and abiotic conditions. Mutual ranks under these conditions were found to be 89, 38, and 180, respectively. Furthermore, *TSA* was found to positively correlate with *TSB* (0.291) in a large group of vegetative and developmental *Arabidopsis* samples. The correlation was clearer in developmental samples comparing to vegetative ones. However, *INS* was co-expressed with *TSB* neither in general nor in specifc conditions and was found to negatively correlate with *TSB* (−0.046), suggesting that INS is operated in the trp-independent pathway. Absence of co-expression and correlation between *INS* and *TSB* may suggest that trp-dependent and independent pathways may separately and independently operate to synthesize IAA.

Production of IAA through INS in the trp-independent pathway is unclear in terms of other enzymes involved in this pathway. Therefore, all previously reported enzymes to have a possible role in IAA synthesis were analyzed in this research. Among all these enzymes, amidase was proposed to interact with INS in the trp-independent pathway. Amidase is the only gene found to be strongly positively coexpressed with *INS*. Mutual ranks for coexpression between *INS* and *amidase* (in particular *amidase1*) were 116, and 141 in general and light conditions respectively. Mutual ranks for these two genes were similar to that found between *TSA* and *TSB*, in which their products form an active complex. *INS* and *amidase* were positively correlated with each other. The value of correlation was similar to that found between *TSA* and *TSB*. Taken together and in addition to the observation that *amidase* is negatively correlated with *TSB*, we suggested that amidase1 is solely involved in the trp-independent pathway of IAA synthesis.

**Author Contribution** All authors contributed to the study conception and design. YMA-Z designed the study and supervised the work. Introduction was written by AA-Z and ARAT. Materials and methods were designed and written by KGF and SMA. The fnal results, discussion and conclusion were written by YMA-Z and FAA. SP revised and commented the fnal draft of the manuscript. All authors commented on previous versions of the manuscript. All authors read and approved the fnal manuscript.

**Data Availability** Not applicable.

### **Declarations**

**Ethical Approval** Not applicable.

**Consent to Participate** Not applicable.

**Consent for Publication** Not applicable.

**Competing Interests** The authors declare no competing interests.

# **References**

- <span id="page-8-0"></span>1. Olanrewaju, O. S., Ayilara, M. S., Ayangbenro, A. S., & Babalola, O. O. (2021). Genome mining of three plant growth-promoting Bacillus species from maize rhizosphere. *Applied Biochemistry and Biotechnology, 193*(12), 3949–3969.
- <span id="page-8-1"></span>2. Woodward, A. W., & Bartel, B. (2005). Auxin: Regulation, action, and interaction. *Annals of Botany, 95*(5), 707–735.
- <span id="page-8-2"></span>3. Mano, Y., & Nemoto, K. (2012). The pathway of auxin biosynthesis in plants. *Journal of Experimental Botany, 63*(8), 2853–2872.
- <span id="page-8-3"></span>4. Won, C., Shen, X., Mashiguchi, K., Zheng, Z., Dai, X., Cheng, Y., & Zhao, Y. (2011). Conversion of tryptophan to indole-3-acetic acid by tryptophan aminotransferases of Arabidopsis and YUCCAs in Arabidopsis. *Proceedings of the National Academy of Sciences, 108*(45), 18518–18523.
- <span id="page-8-4"></span>5. Stepanova, A. N., Yun, J., Robles, L. M., Novak, O., He, W., Guo, H., & Alonso, J. M. (2011). The Arabidopsis YUCCA1 favin monooxygenase functions in the indole-3-pyruvic acid branch of auxin biosynthesis. *The Plant Cell, 23*(11), 3961–3973.
- <span id="page-8-5"></span>6. Zhao, Y., Hull, A. K., Gupta, N. R., Goss, K. A., Alonso, J., Ecker, J. R., & Celenza, J. L. (2002). Trp-dependent auxin biosynthesis in Arabidopsis: Involvement of cytochrome P450s CYP79B2 and CYP79B3. *Genes & Development, 16*(23), 3100–3112.
- <span id="page-8-6"></span>7. Sugawara, S., Hishiyama, S., Jikumaru, Y., Hanada, A., Nishimura, T., Koshiba, T., & Kasahara, H. (2009). Biochemical analyses of indole-3-acetaldoxime-dependent auxin biosynthesis in Arabidopsis. *Proceedings of the National Academy of Sciences, 106*(13), 5430–5435.
- <span id="page-8-7"></span>8. Brumos, J., Alonso, J. M., & Stepanova, A. N. (2014). Genetic aspects of auxin biosynthesis and its regulation. *Physiologia Plantarum, 151*(1), 3–12.
- 9. Abu-Zaitoon, Y. M. (2014). Phylogenetic analysis of putative genes involved in the tryptophandependent pathway of auxin biosynthesis in rice. *Applied Biochemistry and Biotechnology, 172*(5), 2480–2495.
- 10. Kasahara, H. (2016). Current aspects of auxin biosynthesis in plants. *Bioscience, Biotechnology, and Biochemistry, 80*(1), 34–42.
- <span id="page-8-8"></span>11. Abu-Zaitoon, Y. M., Al Tawaha, A. R., Alnaimat, S. M., Al-Rawashdeh, I. M., Abu-Zaiton, A., & Khalifat, A. (2019). Investigation of the potential role of aldehyde oxidase in indole-3-acetic acid synthesis of developing rice grains. *Plant Cell Biotechnology and Molecular Biology, 20*(1), 6–13.
- <span id="page-8-9"></span>12. Wright, A. D., Sampson, M. B., Neufer, M. G., Michalczuk, L., Slovin, J. P., & Cohen, J. D. (1991). Indole-3-acetic acid biosynthesis in the mutant maize orange pericarp, a tryptophan auxotroph. *Science, 254*(5034), 998–1000.
- <span id="page-8-10"></span>13. Normanly, J., Cohen, J. D., & Fink, G. R. (1993). Arabidopsis thaliana auxotrophs reveal a tryptophan-independent biosynthetic pathway for indole-3-acetic acid. *Proceedings of the National Academy of Sciences, 90*(21), 10355–10359.
- <span id="page-8-11"></span>14. Michalczuk, L., Ribnicky, D. M., Cooke, T. J., & Cohen, J. D. (1992). Regulation of indole-3-acetic acid biosynthetic pathways in carrot cell cultures. *Plant Physiology, 100*(3), 1346–1353.
- 15. Östin, A., Ilić, N., & Cohen, J. D. (1999). An in vitro system from maize seedlings for tryptophanindependent indole-3-acetic acid biosynthesis. *Plant Physiology, 119*(1), 173–178.
- 16. Epstein, E., Cohen, J. D., & Slovin, J. P. (2002). The biosynthetic pathway for indole-3-acetic acid changes during tomato fruit development. *Plant Growth Regulation, 38*(1), 15–20.
- 17. Rapparini, F., Tam, Y. Y., Cohen, J. D., & Slovin, J. P. (2002). Indole-3-acetic acid metabolism in *Lemna gibba* undergoes dynamic changes in response to growth temperature. *Plant Physiology, 128*(4), 1410–1416.
- <span id="page-8-12"></span>18. Sztein, A. E., Ilić, N., Cohen, J. D., & Cooke, T. J. (2002). Indole-3-acetic acid biosynthesis in isolated axes from germinating bean seeds: The efect of wounding on the biosynthetic pathway. *Plant Growth Regulation, 36*(3), 201–207.
- <span id="page-9-0"></span>19. Pieck, M., Yuan, Y., Godfrey, J., Fisher, C., Zolj, S., Vaughan, D., & Celenza, J. L. (2015). Auxin and tryptophan homeostasis are facilitated by the ISS1/VAS1 aromatic aminotransferase in Arabidopsis. *Genetics, 201*(1), 185–199.
- <span id="page-9-1"></span>20. Wang, B., Chu, J., Yu, T., Xu, Q., Sun, X., Yuan, J., & Li, J. (2015). Tryptophan-independent auxin biosynthesis contributes to early embryogenesis in Arabidopsis. *Proceedings of the National Academy of Sciences, 112*(15), 4821–4826.
- <span id="page-9-2"></span>21. Nonhebel, H. M. (2015). Tryptophan-independent indole-3-acetic acid synthesis: Critical evaluation of the evidence. *Plant Physiology, 169*(2), 1001–1005.
- <span id="page-9-3"></span>22. Gupta, C., & Pereira, A. (2019). Recent advances in gene function prediction using context-specifc coexpression networks in plants. *F1ooo Research, 8*, 153.
- <span id="page-9-4"></span>23. Persson, S., Wei, H., Milne, J., Page, G. P., & Somerville, C. R. (2005). Identifcation of genes required for cellulose synthesis by regression analysis of public microarray data sets. *Proceedings of the National Academy of Sciences, 102*(24), 8633–8638.
- <span id="page-9-5"></span>24. Hirai, M. Y., Sugiyama, K., Sawada, Y., Tohge, T., Obayashi, T., Suzuki, A., & Saito, K. (2007). Omics-based identifcation of Arabidopsis Myb transcription factors regulating aliphatic glucosinolate biosynthesis. *Proceedings of the National Academy of Sciences, 104*(15), 6478–6483.
- <span id="page-9-6"></span>25. Obayashi, T., Aoki, Y., Tadaka, S., Kagaya, Y., & Kinoshita, K. (2018). ATTED-II in 2018: A plant coexpression database based on investigation of the statistical property of the mutual rank index. *Plant and Cell Physiology, 59*(1), e3–e3.
- <span id="page-9-7"></span>26. Obayashi, T., Kinoshita, K., Nakai, K., Shibaoka, M., Hayashi, S., Saeki, M., & Ohta, H. (2007). ATTED-II: a database of co-expressed genes and cis elements for identifying co-regulated gene groups in Arabidopsis. *Nucleic Acids Research, 35*(suppl\_1), D863–D869.
- <span id="page-9-8"></span>27. Richter, A., Powell, A. F., Mirzaei, M., Wang, L. J., Movahed, N., Miller, J. K., & Jander, G. (2021). Indole-3-glycerolphosphate synthase, a branchpoint for the biosynthesis of tryptophan, indole, and benzoxazinoids in maize. *The Plant Journal, 106*(1), 245–257.
- <span id="page-9-9"></span>28. Nagao, R. T., & Moore, T. C. (1972). Partial purifcation and properties of tryptophan synthase of pea plants. *Archives of Biochemistry and Biophysics, 149*(2), 402–413.
- <span id="page-9-10"></span>29. Hofmann, M., Lehmann, T., Neu, D., Hentrich, M., & Pollmann, S. (2010). Expression of amidase1 (AMI1) is suppressed during the frst two days after germination. *Plant Signaling and Behavior, 5*(12), 1642–1644.
- <span id="page-9-11"></span>30. Gao, Y., Dai, X., Aoi, Y., Takebayashi, Y., Yang, L., Guo, X., & Zhao, Y. (2020). Two homologous indole-3-acetamide (IAM) hydrolase genes are required for the auxin efects of IAM in Arabidopsis. *Journal of Genetics and Genomics, 47*(3), 157–165.
- <span id="page-9-12"></span>31. Pollmann, S., Neu, D., & Weiler, E. W. (2003). Molecular cloning and characterization of an amidase from Arabidopsis thaliana capable of converting indole-3-acetamide into the plant growth hormone, indole-3-acetic acid. *Phytochemistry, 62*(3), 293–300.
- <span id="page-9-13"></span>32. Urbancsok, J., Bones, A. M., & Kissen, R. (2018). Benzyl cyanide leads to auxin-like efects through the action of nitrilases in Arabidopsis thaliana. *Frontiers in Plant Science, 9*, 1240.
- <span id="page-9-14"></span>33. Vorwerk, S., Biernacki, S., Hillebrand, H., Janzik, I., Müller, A., Weiler, E. W., & Piotrowski, M. (2001). Enzymatic characterization of the recombinant Arabidopsis thaliana nitrilase subfamily encoded by the NIT 2/NIT 1/NIT 3-gene cluster. *Planta, 212*(4), 508–516.
- <span id="page-9-15"></span>34. Lehmann, T., Janowitz, T., Sánchez-Parra, B., Alonso, M. M. P., Trompetter, I., Piotrowski, M., & Pollmann, S. (2017). Arabidopsis nitrilase 1 contributes to the regulation of root growth and development through modulation of auxin biosynthesis in seedlings. *Frontiers in Plant Science, 8*, 36.
- <span id="page-9-16"></span>35. Piotrowski, M., Schönfelder, S., & Weiler, E. W. (2001). The Arabidopsis thaliana isogene NIT4 and its orthologs in tobacco encode β-cyano-L-alanine hydratase/nitrilase. *Journal of Biological Chemistry, 276*(4), 2616–2621.
- <span id="page-9-17"></span>36. Seo, M., Akaba, S., Oritani, T., Delarue, M., Bellini, C., Caboche, M., & Koshiba, T. (1998). Higher activity of an aldehyde oxidase in the auxin-overproducing superroot1 mutant of Arabidopsis thaliana. *Plant Physiology, 116*(2), 687–693.

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.