CLINICAL RESEARCH





High Survivorship With Cementless Stems and Cortical Strut Allografts for Large Femoral Bone Defects in Revision THA

Young-Hoo Kim MD, Jang-Won Park MD, Jun-Shik Kim MD, Devarshi Rastogi MD

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Abstract

Background Numerous studies have investigated the clinical and radiographic results of revision THAs with use of cementless stems and cortical strut allografts. However, to our knowledge, no long-term followup studies have evaluated patients undergoing revision THA with use of cortical strut allografts where the allografts provided the primary stability for extensively coated femoral stems in the presence of extensive femoral diaphyseal bone defects. *Question/purposes* We performed this study to determine (1) validated outcomes scores; (2) radiographic signs of fixation and allograft healing; (3) frequency of complications; and (4) survivorship of the components after use of cortical strut onlay allografts in Types IIIB and IV femoral diaphyseal bone defects.

Methods Between 1994 and 2003, we performed 140 revision THAs in 130 patients with Paprosky Types IIIB and IV femoral diaphyseal defects. The patients were treated using extensively coated femoral stems and cortical strut allografts because primary axial or rotational stability could not be achieved without grafting. Ten of the patients (10 hips; 7.7%) were lost to followup or died before 10 years; the remaining 120 patients (130 hips) represent the study group in this retrospective study. There were 66 men and 54 women. Their mean age at the time of index surgery was 59 ± 18 years (range, 36–67 years). The primary diagnosis was predominantly osteonecrosis of the femoral head (53%). The most common reason for revision was aseptic loosening (97%), followed by periprosthetic fracture (3%). The mean time from primary to revision THA was 12 years (range, 8-27 years). The mean duration of followup was 16.1 years (range, 12-20 years).

Results The mean Harris hip score was 39 ± 10 points before revision and improved to 86 ± 14 points at 16 years followup (p = 0.02). The mean preoperative WOMAC score was 62 ± 29 (41–91) points and improved to 22 ± 19 (11–51) points at 16 years followup (p = 0.003). Of the 130 stems, 113 (87%) had bone ingrowth, five (4%) had stable fibrous ingrowth, and 12 (9%) were unstable. All allografts were incorporated. Four hips (3%) had a displaced femoral shaft fracture at the stem tip; four (3%) had a postoperative dislocation; and six (5%) had early postoperative infection. Kaplan-Meier survivorship analysis, with revision or radiographic failure as the endpoint, revealed that the 16-year rate of survival of the components was 91% (95% CI, 0.88%–0.96%).

Conclusion Supportive cortical strut onlay allografts provided high survivorship beyond 12 years of followup in revision THAs. Future studies might compare this

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approach with allograft-prosthesis composites, proximal femoral replacement, or modular fluted, tapered stems. *Level of Evidence* Level IV, therapeutic study.

Introduction

Revision of a failed THA is challenging because of the difficulty in obtaining and maintaining implant stability with poor bone quantity and quality [4]. Severe acetabular and femoral bone loss (Paprosky Types IIIB and IV) [28] can be seen in patients who have had multiple revision arthroplasties and in patients who have severe osteolysis. Large bulk allografts (such as proximal femoral allografts) or proximal femoral or total femoral replacement prostheses such as those used in tumor cases have been proposed as suitable reconstructive techniques for more severe femoral bone deficiencies [6, 9, 28, 29, 32, 33]. As an alternative, the use of structural cortical strut onlay allografts, in combination with cementless revision femoral stems, is a technique used for reconstruction of large femoral bone defects [3, 18, 23–25].

Successful results with cortical strut onlay allografts have been reported for Paprosky Type IIIA bone loss [18, 25-27] (severe femoral diaphyseal bone loss with less than 4 cm of intact cortical bone at isthmus). In these cases, tight press-fit fixation of the revision femoral stem was achieved in the femur before application of cortical strut onlay allografts. The application of cortical strut onlay allografts was intended to provide only supplementary augmentation of the femur [9, 18, 27]. However, to our knowledge, no previously published studies have explicitly reported on the long-term results of THAs, in which axial and torsional stability of the femoral stem was provided not by the host femoral bone, but by the cortical strut allografts in Types IIIB and IV femoral bone defects [27]. Since this is a technique we have used for femoral bone defects [3], we wished to evaluate the results at a minimum followup of 12 years.

The purpose of this study was to determine (1) validated outcome scores; (2) radiographic signs of fixation and allograft healing; (3) frequency of complications; and (4) survivorship of the components after use of cortical strut onlay allografts in Paprosky Types IIIB and IV femoral diaphyseal bone defects.

Patients and Methods

Between 1994 and 2003, we performed 140 revision THAs in 130 patients with Paprosky Types IIIB and IV femoral diaphyseal defects. The prerevision diagnoses were aseptic loosening in 136 hips (97%) and periprosthetic fracture in

four hips (3%). The patients were treated using extensively coated femoral stems and cortical strut allografts because primary axial or rotational stability could not be achieved without grafting. We routinely used this approach in all patients who had no axial and torsional stability of the femoral stem provided by host femoral bone. Data for 130 patients were entered in the database. However, 10 of the 130 patients were later excluded from the database because they were lost to followup (seven patients) or died (three patients) before 2 years of followup. Therefore, 120 patients (130 hips) composed the study. The study was approved by the institutional review board, and all patients were reported in a previous study at a mean of 10 years of followup [23].

The study group included 66 men and 54 women who had a mean age (\pm SD) of 59 \pm 18 years (range, 36-67 years) at the time of revision surgery. The mean height of the patients was 171 ± 9 cm (range, 155–185 cm), mean weight was 82 ± 10 kg (range, 65–125 kg), and mean BMI was $28 \pm 5 \text{ kg/m}^2$ (range, 27–37 kg/m²). The minimum time from the primary to revision THA was 12 years (mean, 16 years; range, 8-27 years). Seventy hips (54%) had a Paprosky Type IIIB defect and 60 (46%) had a Paprosky Type IV defect. Reasons for revision were aseptic loosening in 126 hips (97%) and periprosthetic fracture in four hips (3%). Thirteen hips (10%) required one strut graft, 98 (75%) required two strut grafts, and 19 (15%) required three strut grafts. The minimum followup was 12 years (mean, (16 years; range, 12–20 years) (Table 1). The records of the 120 patients were entered in an ongoing computerized registry that was updated continuously. All of the hips were reviewed by a research associate (DRK) not involved with the revision surgery. Followups were scheduled at postoperative intervals of 3 months, 1 year, and yearly thereafter. At these intervals, the patients were evaluated and radiographs were obtained. Preoperative and postoperative review data were recorded according to the systems of Harris hip score [17] and WOMAC score [5]. Clinical, hematologic, and radiographic data were recorded, including any documented history of pyogenic or tuberculous infection at the site of a previous surgical procedure on the involved hip, relevant laboratory findings, and clinical features such as draining sinuses.

Seventy-nine hips (61%) had a first revision surgery, 27 (21%) had a second revision, 12 (9%) had a third revision, and 12 (9%) had a fourth revision. Although 79 hips had a first revision surgery, they had a significant femoral bone defect attributable to osteolysis and/or loosening. Bone defects were substantially worsened during removal of the cementless or cemented stems. There were no differences in bone defects between cementless and cemented stems.

Table 1. Demographic data

Number of patients (hips)120 (130)Male:female $66:54$ Age (years) (mean \pm SD [range]) 59 ± 18 (36-67)Height (cm) (mean \pm SD [range]) 82 ± 10 (65-125)BMI (kg/m ²) (mean \pm SD [range]) 28 ± 5 (27-37)Primary diagnosis (number [%] of patients/number of hips) 28 ± 5 (27-37)Osteonecrosis of femoral head78 (65)/85Osteonecrosis of femoral head78 (65)/85Osteonecrosis of femoral head78 (65)/7Type of bone defect (Paprosky [24])IIIBIIIB70 hips (54%)IV60 hips (46%)Reasons for revision (number of hips)Aseptic losening126 (97%)Periprosthetic fracture4 (3%)Number of revisionsFirst79 hips (61%)Second27 hips (21%)Third12 hips (9%)Fourth12 hips (9%)Fourth12 hips (10%)Two98 hips (75%)Three19 hips (15%)Zones of allograft (modified Gruen zone [24])Lateral proximal 1/32Lateral proximal 1/33Lateral distal 1/34Around the stem tip5Medial distal 1/36S0% of allograft thickness in 1 or 2 zonesModerate $<50\%$ of thickness of allograft tin 2 zonesFollowup (years)16.1 (12-20)	Parameters	Numbers
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1Lateral proximal $1/3$ 2Lateral middle $1/3$ 3Lateral distal $1/3$ 4Around the stem tip5Medial distal $1/3$ 6Medial middle $1/3$ Grading of allograft resorptionMildMild< 50% of allograft thickness in 1 or 2 zonesModerate< 50% of thickness of allograft in > 2 zones or > 50% of thickness, but not full thickness in 1 or 2 zonesSevereFull thickness in any zone or > 50% of the thickness in > 2 zonesFollowup (years)16.1 (12–20)	Zones of allograft (modified Gruen zone [24])	
2Lateral middle $1/3$ 3Lateral distal $1/3$ 4Around the stem tip5Medial distal $1/3$ 6Medial middle $1/3$ Grading of allograft resorptionMildMild< 50% of allograft thickness in 1 or 2 zonesModerate< 50% of thickness of allograft in > 2 zones or > 50% of thickness, but not full thickness in 1 or 2 zonesSevereFull thickness in 1 or 2 zonesFollowup (years)16.1 (12–20)	1	Lateral proximal 1/3
3Lateral distal 1/34Around the stem tip5Medial distal 1/36Medial middle 1/3Grading of allograft resorptionMildMild< 50% of allograft thickness in 1 or 2 zonesModerate< 50% of thickness of allograft in > 2 zones or > 50% of thickness, but not full thickness in 1 or 2 zonesSevereFull thickness in any zone or > 50% of the thickness in > 2 zonesFollowup (years)16.1 (12–20)	2	Lateral middle 1/3
4Around the stem tip5Medial distal 1/36Medial middle 1/3Grading of allograft resorptionMildMild< 50% of allograft thickness in 1 or 2 zonesModerate< 50% of thickness of allograft in > 2 zones or > 50% of thickness, but not full thickness in 1 or 2 zonesSevereFull thickness in any zone or > 50% of the thickness in > 2 zonesFollowup (years)16.1 (12–20)	3	Lateral distal 1/3
5Medial distal 1/36Medial middle 1/3Grading of allograft resorptionMildMild< 50% of allograft thickness in 1 or 2 zonesModerate< 50% of thickness of allograft in > 2 zones or > 50% of thickness, but not full thickness in 1 or 2 zonesSevereFull thickness in 1 or 2 zonesFollowup (years)16.1 (12–20)	4	Around the stem tip
6Medial middle 1/3Grading of allograft resorptionMild< 50% of allograft thickness in 1 or 2 zonesModerate< 50% of thickness of allograft in > 2 zones or > 50% of thickness, but not full thickness in 1 or 2 zonesSevereFull thickness in any zone or > 50% of the thickness in > 2 zonesFollowup (years)16.1 (12–20)	5	Medial distal 1/3
Grading of allograft resorptionMild< 50% of allograft thickness in 1 or 2 zonesModerate< 50% of thickness of allograft in > 2 zones or > 50% of thickness, but not full thickness in 1 or 2 zonesSevereFull thickness in any zone or > 50% of the thickness in > 2 zonesFollowup (years)16.1 (12–20)	6	Medial middle 1/3
Mild< 50% of allograft thickness in 1 or 2 zonesModerate< 50% of thickness of allograft in > 2 zones or > 50% of thickness, but not full thickness in 1 or 2 zonesSevereFull thickness in any zone or > 50% of the thickness in > 2 zonesFollowup (years)16.1 (12–20)	Grading of allograft resorption	
Moderate< 50% of thickness of allograft in > 2 zones or > 50% of thickness, but not full thickness in 1 or 2 zonesSevereFull thickness in any zone or > 50% of the thickness in > 2 zonesFollowup (years)16.1 (12–20)	Mild	< 50% of allograft thickness in 1 or 2 zones
SevereFull thickness in any zone or $> 50\%$ of the thickness in > 2 zonesFollowup (years)16.1 (12–20)	Moderate	< 50% of thickness of allograft in > 2 zones or > 50% of thickness, but not full thickness in 1 or 2 zones
Followup (years) 16.1 (12–20)	Severe	Full thickness in any zone or > 50% of the thickness in > 2 zones
	Followup (years)	16.1 (12–20)

The femoral components, which were in situ at the time of the current revision, were cementless femoral components in 107 hips (82%) and cemented femoral components in 23 hips (18%). The acetabular components, which were in situ at the time of the current revision, were cementless acetabular components in 114 hips (88%) and cemented acetabular components in 16 hips (12%).

Surgical Technique

It often was difficult to determine the severity of the bone loss from preoperative radiographs and CT scans; however, after implant removal, the available bone stock could be clearly defined by intraoperative findings. In all cases in the current series, bone loss was too severe (Paprosky defect Type IIIB or IV [27]) to provide satisfactory axial and torsional stability of a revision total hip prosthesis. The decision to use a cortical strut onlay allograft was made at surgery by a senior surgeon (YHK) if it was determined that axial and torsional stability of the trial femoral stem was not able to be obtained before application of the cortical strut onlay grafts. We never used other techniques such as circumferential proximal femoral allograft, impaction bone grafting, proximal femoral allograftprosthesis composite, proximal femur replacement and modular fluted tapered stem. Through a posterolateral approach, the acetabulum and the proximal 1/3 of the femur were exposed. The loose femoral stem and acetabular component were removed. If the acetabular component was well fixed in an unsatisfactory position, a curved osteotome (DePuy, Warsaw, IN, USA) was used to remove the cup. An extended greater trochanteric osteotomy was performed in 81 hips (62%) to remove the cementless or cemented stem with surrounding cement. Less than 2 cm of diaphyseal-isthmic bone was available after removal of the stems in all hips.

All patients had implantation of fresh-frozen femoral cortical strut onlay allografts (LifeNet Health, Virginia Beach, VA, USA). One cortical strut allograft was used when one side of the femur had a bone defect, two were used when two sides of the femur had bone defects, and three were used when three sides of the femur had bone defects. One cortical strut onlay allograft was used in 13 hips (10%), two were used in 98 hips (75%), and three were used in 19 hips (15%).

The width and length of the cortical strut onlay allografts were selected to bridge the bone defects without interfering with the knee. Proximally, impingement of the strut onlay grafts against the pelvis was avoided. The mean width of the individual strut graft was 4 cm (range, 3– 6 cm), and the mean length was 25 cm (range, 16–32 cm). To maintain femoral blood supply, only limited dissection of the soft tissues from the femur, especially at the linea aspera, was done. If possible, the remaining periosteum and residual shells of host bone were left in place.



Fig. 1A–C A 45-year-old man had Paprosky Type IV bone defects of both hips. (**A**) His AP radiograph shows a loose porous-coated anatomic cementless femoral component and dislocated cement spacer of the left hip before revision surgery. (**B**) An intraoperative photographs shows the allografts are fixed with four Dall-MilesTM

(Stryker Orthopaedics, Mahwah, NJ, USA) cables. (C) An AP radiograph of both hips taken 2 weeks after revision surgery shows the femoral and acetabular components are well fixed in a satisfactory position. The allografts are attached to the host bone with Dall-MilesTM cables.

The allografts were secured to the host bone with an average of five cables (range, 4–8 cables). Definitive intraoperative stability of the implants was achieved by complete tightening of the cortical strut onlay allografts. Supplemental cancellous bone grafts around the strut grafts were applied in all patients. Vascularized muscle tissue was placed over all of the grafts. For all cases, long extensively porous-coated cementless femoral prostheses were used (Solution System[®]; DePuy). The mean diameter of Solution System[®] stem was 17.25 mm (range, 15–19.5 mm) (Fig. 1). Alternatives such as allograft-prosthesis composites, modular fluted, tapered stems were not used, because these stems did not appear to be able to be fixed in the damaged diaphyseal-isthmic bone.

All of the patients began standing at the bedside or walking with assistance of crutches or a walker twice daily for 30 minutes each time under the supervision of a therapist on the second postoperative day. The patients used crutches or a walker with toe-touch weightbearing for 3 months and used a cane with full weightbearing when needed thereafter.

The interpretation of all radiographs was done in a blinded fashion by one observer (JWP) who was not involved in treatment of any of the patients. Implant subsidence was measured using constant reference points for each patient such as a tip of the greater trochanter and the upper margin of the shoulder of the prosthesis [33]. The radiographic stability was classified according to the techniques of Engh and Massin [12].

The radiographic criterion of incorporation of cortical strut onlay allograft to host bone was complete bridging

between strut allograft and host bone. The allograft was divided into modified Gruen zones [13]. Resorption of cortical strut onlay allograft was graded as follows [24]: mild, less than 50% of allograft thickness in one or two zones; moderate, less than 50% of thickness of allograft in more than two zones or greater than 50% of thickness but not fully thickness in one or two zones; or severe, full thickness in any zone or greater than 50% of thickness in more than two zones. Data from all radiographs showed that for angular measurements, the mean intraobserver difference was 1.5° (range, $0.8^{\circ}-2.5^{\circ}$) and the intraclass correlation coefficient was 0.95. For linear measurements (width of radiolucent line and resorption of allograft), the mean intraobserver difference was 1.5 mm (range, 1.0-1.8 mm) and the intraclass correlation coefficient was 0.96.

Statistical Analysis

Descriptive analysis was performed using the mean and frequency for continuous and categorical variables, respectively. Cumulative survival rates and associated 95% CIs were calculated using a Kaplan-Meier survival curve [22]. Comparisons of clinical characteristics and preoperative and postoperative values used ANOVA for continuous variables or chi-square test or Fisher's exact test for categorical variables. Statistical analysis was performed using SPSS software (Version 19.0; IBM, Armonk, NY, USA), and a p value less than 0.05 was considered significant.

Results

Preoperative outcome scores were improved significantly (p = 0.002) after revision surgery. The Harris hip scores improved from a mean of 39 ± 10 points (range, 21–54 points) before revision surgery to a mean of 86 ± 14 points (range, 50–92 points) at the latest followup. Mean improvement was 47 points (95% CI, 39.1–52.3 points) (p = 0.002). Before revision surgery, the mean pain score was 14 points (range, 9–19 points) and the mean function

score was 25 points (range, 12–35 points). At the latest followup, the mean pain score was 42 points (range, 26–44 points) and the mean function score was 44 points (range, 24–48 points) (p = 0.002). Limping, dependence on a walking aid, and walking distance were improved markedly after revision surgery (p = 0.004). The WOMAC score improved from a mean of 62 ± 29 points before revision surgery to a mean of 22 ± 19 points at followup. Mean improvement was 40 points (30–40 points) (95% CI, 36.5–44.1) (p = 0.003) (Table 2).

Table 2. Clinical results

Parameters	Before revision surgery	Final followup (16.1 years)	Improvement	p Value (ANOVA)
Total Harris	Mean 39 ± 10 (21–54)	Mean 86 ± 14 (50–92)	Mean 47 (29-38)	0.002
hip score (points)*			[39.1–52.3]	
Pain score	Mean 14 (9–19)	Mean 42 (26-44)	28 (17-25)	
Function score	Mean 25 (12-35)	Mean 44 (24–48)	19 (12–13)	
Degree of pain				
None	0	40 patients (33%)		< 0.001
Slight	0	32 patients (27%)		
Mild	0	28 patients (23%)		
Moderate	85 patients (71%)	9 patients (8%)		
Marked	35 patients (29%)	11 patients (9%)		
Totally disabled	0	0		
Limp				
None	0	30 patients (25%)		0.004
Slight	0	42 patients (35%)		
Moderate	85 patients (71%)	37 patients (31%)		
Severe	35 patients (29%)	11 patients (9%)		
Unable to walk	0	0		
Support				
None	0	87 patients (72%)		0.004
Cane, long walks	0	13 patients (11%)		
Cane, full time	60 patients (50%)	9 patients (8%)		
1 crutch	7 patients (6%)	0		
2 crutches	53 patients (44%)	11 patients (9%)		
Unable to walk	0	0		
Distance walked				
Unlimited	0	40 patients (33%)		0.004
6 blocks	0	60 patients (50%)		
2-3 blocks	85 patients (71%)	9 patients (8%)		
Indoors only	35 patients (29%)	11 patients (9%)		
Bed and chair	0	0		
WOMAC score				
Total	Mean 62.9 \pm 29 (41–91)	Mean 22 \pm 19 (11–51)	Mean 40 (30-40)	0.003
Satisfaction				
Fully satisfied	0 (0%)	40 patients (33%)		
Satisfied	0 (0%)	60 patients (50%)		
Somewhat dissatisfied	0 (0%)	9 patients (8%)		
Fully dissatisfied	130 patients (100%)	11 patients (9%)		

* Ranges in parentheses. 95% CI in brackets.

Of the 130 stems, 113 (87%) had bone ingrowth, five (4%) had stable fibrous ingrowth, and 12 (9%) were unstable. Eleven of 12 stems were revised or rererevised with a larger cementless Solution System[®] stem and one hip was rerevised with the largest Wagner stem (Zimmer, Warsaw, IN, USA) on the market. In this particular case, the largest Solution System[®] stem was not able to provide axial and torsional stability: therefore, the largest Wagner stem was used to provide axial and torsional stability (Fig. 2). We did not detect bone resorption of the proximal host femur related to stress shielding in the current series, because the proximal host femur was covered by cortical strut allograft. Allografts were incorporated in the host femur consistently and reliably by 1 year, on average, in all hips. Although bone ingrowth cannot occur from a cortical strut allograft to a porous-coated stem (because the cortical strut allograft has no vital cells), there was no intervening radiolucent line at the interface between the cortical strut allograft and the host bone in 120 hips (92%). In the remaining 10 hips, there was a radiolucent line smaller than 1 mm between the host bone and allograft. There was slight loss of length of the cortical strut onlay allografts by remodeling at the ends of the allografts. The resorption of allografts was graded as mild in 110 hips (85%) and moderate in 20 hips (15%). The resorption of allograft was noticed 1 year after revision and it progressed slowly. The resorption usually abated by 3 years. Minor resorption of the allograft was seen around cables, but there were no failures of any of the cortical strut onlay allografts (Table 3). The previous allografts provided enhanced bone stock in all hips.

Four hips (3%) had a displaced femoral shaft fracture at the stem tip after revision surgery. The fractures were fixed with two bivalve femoral cortical strut onlay grafts and healed uneventfully. The femoral stems were stabilized by bone ingrowth at the latest followup.

Six hips (5%) had an early postoperative infection. In four of the six hips, the infections were treated successfully

Table 3. Radiographic results at final followup

Parameters	Numbers
Fixation of stem	
Bone ingrowth stable fibrous	113 hips (87%)
Ingrowth	5 hips (4%)
Unstable	12 hips (9%)
Union of allograft	130 hips (100%)
Length of allograft (cm)	
Medial cortical strut*	
Initial	13 cm (10–20 cm)
Final	12 cm (8–18 cm)
Lateral cortical strut*	
Initial	13 cm (9–18 cm)
Final	11 cm (8–17 cm)
Posterior cortical strut*	
Initial	11 cm (9–13 cm)
Final	10 cm (9-13 cm)
Resorption of allograft	
Mild	110 hips (85%)
Moderate	20 hips (15%)

* Ranges in parentheses.



Fig. 2A–C A 65-year-old man had aseptic loosening of the cemented femoral component. (A) His prerevision AP radiograph shows a cement fracture, varus shift, and subsidence of the left femoral component. The bone defect of the left femur is Paprosky Type IV. (B) An AP radiograph of the patient's left hip taken 6 months after surgery shows a fully porous-coated cementless stem and two strut cortical onlay allografts placed in a satisfactory position. Although there is no radiographic loosening of the femoral component, the patient reported severe pain during weightbearing. This patient had

the largest size of Solution System^(R) stem implanted during revision surgery, but he had persistent pain and underwent rerevision surgery using the largest size Wagner stem. (C) The AP radiograph taken 15 years after rerevision surgery shows the Wagner cementless stem is embedded in a satisfactory position with solid bone ongrowth on the entire femoral stem. A two-cortical strut onlay allograft was incorporated in the host bone. The acetabular component is well fixed in a satisfactory position. by open débridement and 6 weeks of intravenous antibiotics. The remaining two hips with failed open débridement underwent two-stage exchange arthroplasty. None of these hips had recurrence of infection. Four hips (3%) had a postoperative dislocation and these were treated by closed reduction and an abduction brace for 3 months. There were no additional dislocations. Five hips (4%) had calf thrombi but there were no pulmonary embolisms after treatment with a mechanical compression device. No patient had wound problems. No hip had Grade III or IV heterotopic ossification [7].

Kaplan-Meier survival analysis revealed 91% survivorship of the femoral components as the endpoint of revision or aseptic loosening at 16 years (95% CI, 0.88-0.96). In the worst-case scenario, Kaplan-Meier survivorship analysis, with revision or mechanical failure as the end point for failure, estimated the 16-year survival rate of the femoral component to be 83% (95% CI, 0.78-0.88), if the 10 patients who did not complete followup or died were assumed to have a failed femoral component. Twelve hips (9%) were revised owing to failure of femoral components. Ten of these 12 hips were rerevised for aseptic loosening, whereas two of the 12 hips were rerevised for an infection. Two of the 10 hips had three more rerevisions for aseptic loosening. In these 10 rerevised hips because of aseptic loosening, four had Paprosky Type IIIB and six had Type IV femoral diaphyseal bone defects.

Discussion

Numerous studies have investigated the clinical and radiographic results of revision THAs with the use of cementless stem and strut allografts [10, 11, 15, 19, 22, 23, 27]. However, to our knowledge, no long-term followup studies have evaluated patients undergoing revision THA with use of cortical strut allografts where the allografts provided the primary stability for extensively coated femoral stems in the presence of extensive femoral diaphyseal bone defects. The purpose of our study was to determine (1) validated outcome scores; (2) radiographic signs of fixation and allograft healing; (3) frequency of complications; and (4) survivorship of the components after use of cortical strut onlay allografts in Paprosky Types IIIB and IV femoral diaphyseal bone defects.

There are some limitations to our study. First, two concerns common in retrospective and nonrandomized trials were unlikely to have been a factor here: selection bias and lost to followup. We treated 100% of our patients with Paprosky Types IIIB and IV femoral defects using this approach between 1994 and 2003; no patient was treated with another approach. We also were able to account for a large proportion of our patients at a minimum followup of

12 years. This is unusual in long-term studies of this sort, and gives us confidence that few patients could have undergone revision surgery elsewhere for undetected failures. Second, we have no interobserver variability to ensure interpreting hip scores and radiographic findings, including radiolucent line and loosening. However, the intraclass correlation coefficient for intraobserver difference in angular radiographic measurement was 0.95 and in linear radiographic measurement was 0.96, indicating excellent reproducibility in these measurements. Finally, although all allografts appeared to be incorporated radiographically, radiographs are not reliable indicators of graft incorporation. Furthermore, radiolucent lines between the strut allograft and porous implant may be missed on simple AP and lateral views of radiographs.

Large bulk allografts, such as proximal femoral allografts or proximal femoral or total femoral replacement prostheses such as those used in tumor cases, have been proposed as suitable reconstructive techniques for more severe femoral defects [6, 9, 10, 29, 32, 33]. To our knowledge, no long-term clinical results have been reported. Emersen et al. [11] reported that clinical results of femoral revision with strut allografting showed satisfactory results with an average Harris hip score of 79.6 points at average 34 months followup. Barden et al. [3] reported that clinical results of supportive strut grafts for diaphyseal bone defects in revision THAs in 20 patients showed the average Harris hip score was 75.7 points (range, 58-92 points) at mean 4.7 years followup. All of their patients were bedridden or wheelchair bound. They suggested that lower Harris hip score was a result of reduced abductor muscle function rather than pain. Head and Malinin [18] showed that the average Harris hip score improved 45 points at average 9.5 years followup in 251 patients who underwent revision THAs with strut allografts. In the current study, the average hip score improved 47 points at 16 years followup. Our clinical results showed satisfactory results compared with historic controls [3, 11, 18, 19, 25] despite much longer followup. We believe that all of the allografts united to the host bone, and relatively good function of the abductor muscle resulted in satisfactory outcomes.

Barden et al. [3] reported that 17 of 20 cementless femoral revision stems radiographically showed bone ingrowth. Three of 20 revision femoral stems were interpreted as being fixed only by fibrous ingrowth after early subsidence. Head and Malinin [18] reported that 97% of revision THAs with strut allografts had stable fixation with bone ingrowth at an average of 9.5 years followup. However, Emerson et al. [11] reported that eight stems (7%) subsided greater than 1 cm, all of which were clinical failures. In the current study, 91% of femoral stems (118 of 130 stems) had stable fixation at a mean followup of

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CITCUIIII ETEIIUAI PIOXIIIAI JEIIIOIAI	20 hips	Mean 8 years	Allograft resorption	8 hips (40%)	18 hips (90%)
allografts (Roos et al. [30])			Allograft nonunion	1 hip (5%)	
			Allograft partial union	1 hip (5%)	
			Trochanteric migration	1 hip (5%)	
			Total	11 hips (55%)	
Impaction bone grafting with	99 hips	Mean 5.2 years	Intraoperative fracture or perforation	20 hips (20%)	93.1% at 5.2 years as revision end point
cemented stem (Iwase et al.			Resorption of allograft	9 hips (9%)	
([07]			Subsidence of stem $(> 5 \text{ mm})$	2 hips (2%)	
			Total	31 hips (31%)	
Impaction bone grafting with	243 hips	Mean 4.4 years	Aseptic loosening	7 hips (2.9%)	93.8% at 4.4 years as revision end point
cementless stem (Wimmer et al.			Revision rates	21 hips (8.6%)	
			Total	28 hips (12%)	
Proximal femoral allograft-	57 patients	Mean 12 years	Aseptic loosening	4 patients (7%)	69.0% at 10 years as revision end point
prosthesis composite (Babis			Allograft resorption	3 patients (5%)	
et al. [2])			Allograft nonunion	2 patients (4%)	
			Allograft fracture	4 patients (7%)	
			Fracture of stem	1 patient (2%)	
			Deep infection	5 patients (9%)	
			Rerevision	19 patients (44%)	
			Total	38 patients (67%)	
Proximal femoral replacement	27 patients	Mean 55.3 months	Dislocation	2 patients (7%)	Success rate at 55.3 months = 86.4%
(Jaiswal et al. [21])			Change in polyethylene liner and head	1 patient (4%)	
			Cup revision	1 patient (4%)	
			Deep infection	3 patients (11%)	
			Total	7 patients (26%)	
Fluted tapered modular femoral	92 patients	6.4 years	Intraoperative fracture	11 hips (12%)	97% (88 of 92 hips) at 6.4 years
stem (Amanatullah et al. [1])		(2-12 years)	Postoperative periprosthetic fracture	7 hips (8%)	
			Dislocation	17 hips (19%)	
			Deep infection	6 hips (7%)	
			Other	11 hips (12%)	
			Revision	18 hips (20%)	
			Total	70 hips (76%)	

Table 4. continued					
Study	Numbers of hips or patients	Followup	Complications		Survivorship
Fluted tapered modular femoral stem (Brown et al. [8])	58 patients	5.6 years (2–12.6 years)	Subsidence (> 4 mm) Thigh pain Reoperation	6 patients (10%) 11 patients (19%) 10 patients (17%)	98% (95% CI, 89.5–99.5%) at 12 years
Current series	130 hips	Mean 16.1 years	Total Dislocation Deep vein thrombosis (calf vein) Deep infection Femoral shaft Revision Total	27 patients (47%) 4 hips (3%) 5 hips (4%) 6 hips (5%) 4 hips (3%) 12 hips (24%) 31 hips (24%)	91% (95% CI, 0.88–0.96) at 16 years with revision endpoint Worst case scenario: 83% (95% CI, 0.78–0.88) with revision endpoint

16 years. Our results compared well with the results of other series with a short-term followup [11, 18, 19, 25, 27].

A low incidence of failed cortical strut onlay allograft incorporation ranging from 0% to 7% has been reported [10, 12, 18, 27, 33]. These results appear to be confirmed by the apparent incorporation of the cortical strut onlay allografts in the current study. Only small amounts of allograft resorption were observed in all hips. There were no cases of major resorption of the cortical strut allografts used in this study. Buoncristiani et al. [9] found diminution of allografts in 38% of cases, and in one of 32 cases, the allograft appeared to have resorbed completely. In the series of Pak et al. [27], the strut allografts showed severe resorption around femoral stems (7%). Haddad et al. [15] noted a higher rate of resorption (28%; 11 of 40 cases) of the allografts. They also reported that severe resorption of allograft was observed only in cases in which the proximal host femur was discarded for proximal segmental structural allografting [14]. They generated a hypothesis regarding its potential etiology, and the discarded proximal host femur may play a role [15]. The residual host bone may have a physical and immune-protective effect that was not present in their cases. Hamadouche et al. [16] also suggested that when a structural cortical strut allograft is used during revision THA, the host femur should be retained to enhance allograft incorporation. In the current series, the apparent biologic incorporation of all cortical strut onlay allografts could be attributed to the large contact area between the allografts and the host bone [12] and the retained host femur [14]. Although there was no apparent stress shielding in the proximal femur in our series, stress shielding would appear with longer-term followup, because load bearing is transferred more distal to the femur by a fully porouscoated stem. Barden et al. [3] reported one case of peroneal nerve palsy, one case of deep vein thrombosis (DVT), and one case of pneumonia. They had no dislocation of revision THAs. However, Pak et al. [27] reported a 7% dislocation rate (eight of 113 hips). In our series, there were low incidences of DVT and dislocation. We believe that a mechanical compression device for DVT treatment decreased the incidence of DVT. Good alignment of the components and good abductor muscle function were attributed to the low dislocation rate.

The infection rate after revision THA using cortical strut allografts has been reported to range between 0% and 2% [3, 6, 10, 12, 27]. The high postoperative deep infection rate (5%) in our series can be explained by multiple previous surgeries that patients had undergone, long operative time, and the large size of cortical strut onlay allografts. Complication and reoperation rates of alternative techniques, including circumferential proximal femoral allograft [30], impaction bone grafting [20, 31], proximal femoral allograft-prosthesis composites [2], proximal femur replacements [21], and modular fluted tapered stems [1, 8] appear to be greater than in the current study despite that the durations of followup were substantially shorter in those series (Table 4).

Head and Malinin [18] reported the survival rate of revision cementless stems with structural onlay allografts was 97% at 9.5 years followup. Barnes et al. [3] reported the survival rate of revision THA with strut allografts was 100% at 4.7 years followup. Emerson et al. [11] reported the survival rate for revision cementless stems with cortical strut allografts was 93% at 8.4 months followup. They suggested that revision femoral component stability and clinical success was attributable to a combination of implant design, surgical technique, and host-bone factors. In the current series, survivorship of the revision stem was 91% and in the worst-case scenario, it was 83%. We believe that rigid distal fixation of extensively porous-coated long femoral stems with femoral strut allografts was attributable to good results.

Supportive cortical strut onlay allografts provided high survivorship beyond 12 years of followup in revision THAs. Further studies might compare this approach with allograft-prosthesis composites, proximal femoral replacements, or modular fluted, tapered stems.

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