ORIGINAL RESEARCH

Comparative Evaluation of the Properties of Deep‑Frozen Blueberries Dried by Vacuum Infrared Freeze Drying with the Use of CO₂ Laser Perforation, Ultrasound, and Freezing–Thawing as Pretreatments

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Abstract

Existence of cuticle wax layer not only reduces the drying rate of blueberries but also causes the fruit to burst during drying. Such a phenomenon results in undesirable appearance as well as in losses of bioactive compounds responsible for health benefts and in sugars adhering to the surface and hence increased moisture adsorption capability of the dried fruit. In this study, uses of CO₂ laser perforation, ultrasound and freezing–thawing as skin pretreatments prior to infrared freeze drying and their efects on drying characteristics as well as selected properties, i.e., shrinkage, color, rehydration capacity, as well as total anthocyanins and phenolics contents, of blueberries were investigated. Fourier transform infrared (FTIR) spectra of the dried fruit were also determined. Pretreatments increased drying rate and rehydration capacity of blueberries; shrinkage reduced from around 57 to 25%. Laser perforation and freezing–thawing but not ultrasonic pretreatments exhibited no signifcant efects on color of blueberries. Laser perforation and ultrasonic pretreatments exerted positive efects on the total phenolic and anthocyanin contents of the dried samples, respectively. FTIR spectra illustrated that all pretreatments did not alter the chemical fngerprints of the dried fruit powder.

Keywords Anthocyanins · Drying rate · Freezing–thawing · Fourier transform infrared spectroscopy · Phenolic compounds · Physical properties

Introduction

Blueberry *(Vaccinium corymbosum L.)* is a popularly consumed fruit that belongs to the genus Vaccinium in the family Ericaceae. The fruit is sometimes regarded as king of

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berries due to its wide array of health benefts, which include antioxidant, antibacterial, and antiviral activities (Aranaz et al., [2017](#page-10-0)). However, blueberry has very short shelf life of only 2–4 days at a typical ambient temperature. An appropriate means to preserve and hence prolong the shelf life of the fruit is therefore needed (López et al., [2010](#page-10-1); Wang et al., [2013](#page-11-0); Zhang et al., [2006,](#page-11-1) [2017a](#page-11-2)).

Hot air drying, microwave drying, and infrared drying can be used to dry blueberry. Compared with freeze drying, however, these drying methods tend to more extensively reduce the sensory quality and nutritional values of the fruit; freeze drying is therefore an attractive alternative for the preservation of blueberry (Huang, Zhang, Yan et al., [2009](#page-10-2); Kim & Kerr, [2013](#page-10-3); Pei et al., [2014](#page-11-3); Saxena et al., [2012;](#page-11-4) Zielinska et al., [2015](#page-11-5)). An important problem related to freeze drying of blueberry nevertheless exists. Since blueberry epidermis is covered with a layer of wax, water transport from inner to outer parts of the fruit is restricted. This results in prolonged drying time and increased energy consumption. Moreover, when the outward movement of internally sublimed water, which takes place when the fruit temperature reaches the phase transition temperature of water, is restricted, there would be a pressure build-up within the fruit. At a point when such a pressure exceeds the strength of the epidermis, the fruit skin would crack or even burst to release the pressure. This leads to collapse and deformation, which adversely afect the appearance of the dried fruit. Internal juice may also leak out, sometimes adhering to the surface of the fruit (Zielinska et al., [2015](#page-11-5)). Studies have nevertheless confrmed that appropriate pretreatments to modify the skin structure can efectively improve the water transport capability through the fruit skin (Shi et al., [2008\)](#page-11-6). Currently utilized pretreatments for blueberry include chemical and mechanical pretreatments. Chemical residue and possible alteration of the taste of the fruit are among the limitations of chemical pretreatments (Ketata et al., [2013](#page-10-4)). Mechanical pretreatment, on the other hand, involves the use of no chemicals, so it is more health and environmentally friendly than chemical pretreatment. However, mechanical pretreatment still has limitations, including juice loss from the fruit.

Among alternative pretreatment methods for blueberry, use of carbon dioxide-based laser is an attractive one. This is because the laser beam emitted at the working wavelength of $CO₂$ (10.6 µm) is strongly absorbed by water; the laser can also produce spots (micropores) on the surface of the fruit in a much controllable manner. In addition to $CO₂$ -based laser, ultrasonic pretreatment is also of interest (Fan et al., [2020;](#page-10-5) Zhang et al., [2019\)](#page-11-7). Ultrasound has been shown to help modify the skins (or peels) of an array of fruit, hence improving its water transport behavior (Kek et al., [2013](#page-10-6)). Nowacka et al. ([2014\)](#page-11-8), for instance, observed that after 10 min of ultrasonic pretreatment, cell walls of kiwifruit slices started to exhibit microscopic channels that promoted water transport. However, no study is so far available on comprehensive comparison of $CO₂$ -based laser and ultrasound to modify the skin structure of blueberry to enhance its freeze drying rate and to reduce skin rupture and hence internal juice leakage upon drying.

The purpose of this study was to compare the efects of $CO₂$ -based laser perforation and ultrasonic skin pretreat– ments on drying rate and selected quality characteristics of dried frozen blueberries prepared by infrared freeze drying (IRFD), which has been shown to be capable of accelerating the rate of conventional freeze drying (Hnin et al., [2019](#page-10-7)). Freezing–thawing pretreatment was also conducted and compared with laser and ultrasonic pretreatments. Shrinkage, color, rehydration capacity, as well as total anthocyanin and phenolic contents were determined; the contents of these bioactive compounds were determined as they are well recognized to be the key compounds responsible for the health benefts of blueberry. Fourier transform infrared (FTIR) spectra were also determined to identify possible chemical structure alterations and moisture adsorption of the fruit upon pretreatments.

Materials and Methods

Materials and Chemical Reagents

Individually quick frozen blueberries were supplied by Shanghao Biotech Co., Ltd. (Qingdao, China). Blue‑ berries with uniform size and diameter between 12 and 17 mm were stored in a cryogenic refrigerator (Haier, DM-86L626, Qingdao, China) at −65 °C until the time of an experiment. A diagram of the whole experimental program is shown in Fig. [1](#page-2-0). Note that the initial moisture content of blueberries was 89.54% (w.b.) and the total soluble solids content of the frozen fruit was 12.94 °Brix.

Methanol, hydrochloric acid, folinol, sodium carbonate, potassium chloride, sodium acetate, gallic acid monohydrate, ferrous sulfate heptahydrate, salicylic acid, and 30% hydrogen peroxide solution were obtained from Sinopharm Chemical Reagent Co., Ltd (Shanghai, China).

Skin Pretreatments

CO₂ laser device (Yiwang Laser Equipment, CO2-30L, Wuxi, China) was used for laser skin pretreatment. The emission wavelength was noted to be $10.64 \mu m$, while the marking speed, frequency, and power were 100 mm/s, 20 kHz, 30 W, respectively. Perforations were made in a square grid pattern with the dimensions of 2.0×2.0 mm; 25 perforations were made per one blueberry fruit. The distance between the laser head and blueberry surface was maintained at 150 mm; a fruit sample was perforated in its frozen state. After perforation, the sample was returned to the freezer and stayed for at least 12 h prior to subsequent drying. The reason for the refreezing process is that the pretreatment caused some ice crystals in the fruit to melt, so refreezing was needed prior to subsequent IRFD.

In the case of ultrasonic pretreatment, individually frozen blueberries were frst thawed at 4 °C for 5 h. The sample was introduced into an ultrasonic bath (Ultrasonic Instruments, KQ-300VDV, Kunshan, China) for 20 min; the frequency and specifc power of ultrasound were 45 kHz and 20 W/g, respectively. Temperature during ultrasonic pretreatment was noted to be lower than 30 °C at all times. After the treatment, excess water on the surface of the sample was removed by a stream of cool air. The sample was then returned to the freezer and stayed for at least 12 h prior to subsequent drying.

In the case of freezing–thawing pretreatment, individually frozen blueberries were thawed at 4° C for 4 h, and then refrozen in the same cryogenic refrigerator. The refreezing time was again at least 12 h prior to subsequent drying.

Fig. 1 Diagram of the whole experimental program

Amount of 60 g of individual fruit was treated in each case. All the pretreatment experiments were performed in triplicate. Individually frozen blueberries without any pre‑ treatment and dried by IRFD were used as the control sample.

Infrared Freeze Drying

Infrared freeze drying system (Changzhou One Step Drying Equipment, Changzhou, China) shown in Fig. [2](#page-3-0) was used for the experiments. Blueberry sample was loaded into the drying chamber where the infrared radiator was set to maintain the drying temperature at 60 °C. The cold trap temperature was maintained at−40 °C, while the pressure within the drying chamber was 80 Pa. Each drying experiment was performed in triplicate.

Drying Kinetics Determination

The moisture and dry matter contents of frozen, dried, and rehydrated blueberries were gravimetrically determined by vacuum drying at 70 °C as per the ISO 1026–1982 standard. The moisture ratio was then calculated as per Eq. ([1\)](#page-2-1):

$$
MR = \frac{M_t - M_e}{M_0 - M_e} \tag{1}
$$

where M_t , M_0 , and M_e are the instantaneous, initial, and equilibrium moisture contents of the sample, respectively. Evolution of the sample moisture content is shown in terms of the drying curve.

Fig. 2 Schematic diagram of infrared radiation freeze drying equipment. 1=Control panel; 2 =thermocouples; 3 =infrared lamps; $4 =$ sample; $5 =$ drying chamber; $6 =$ observation window; $7 =$ chamber door; $8 =$ vac $$ uum pump; 9=purge valve; $10=$ condenser; $11=$ refrigerat– ing and fan systems

Physical Property Determination

Shrinkage Determination

The solid displacement method of Huang, Zhang, Mujumdar et al. ([2009](#page-10-8)) was used to determine the shrinkage of a sample. Shrinkage was calculated as:

$$
Shrinkage = \frac{V_0 - V_1}{V_0}
$$
\n⁽²⁾

where V_0 and V_1 are the volumes of the sample before and after drying, respectively. Each measurement was performed in triplicate.

Color Determination

Color of a sample was measured using a spectrophotometer (Konica Minolta, CM-3600d, Tokyo, Japan). The meter was calibrated against a standard white plate prior to each measurement using D65 as the light source at 10° standard observer. Lightness (L^*) , redness/greenness (a^*) , and yellowness/blueness (b^*) values were measured. Total color difference (ΔE) was then calculated as per Eq. ([2\)](#page-3-1). Each measure– ment was performed in triplicate.

$$
\Delta E = \sqrt{\left(L^* - L_0\right)^2 + \left(a^* - a_0\right)^2 + \left(b^* - b_0\right)^2}
$$
\n(3)

Rehydration Capacity Determination

The rehydration capacity is defned by Lewicki ([1998](#page-10-9)) as the ratio of the mass of water adsorbed during rehydration to the dry mass of the fruit. Two grams of dried blueberries was placed in 40 g distilled water at 37 °C to reconstitute; the sample was kept in the water for 2, 7, 17, 32, and 52 min (Feng et al., [1999](#page-10-10)). After rehydration, the sample was wiped with filter paper to remove excess water and then weighed. A rehydration curve of each sample was prepared by plotting the amount of adsorbed water per unit dry matter against time. Each measurement was performed in triplicate.

Bioactive Compound Content Determination

Blueberry powder (0.1 g) (or 5 g fresh mass) was accurately weighed and placed in a 25-mL tube. Then, 25 mL of 80% (v/v) methanol was added; 1% (v/v) HCl was also added to ensure that the pH of the solution was in the range of 2–3. Extraction was performed under ultrasonic condition (frequency of 45 kHz and specifc power of 12 W/g sam‑ ple+solvent) for 30 min. Clarifed extract was obtained by fltering through a flter paper; extraction was performed in triplicate for each sample. The extract was stored in a refrigerator at 4 °C until further analysis (but not longer than 24 h).

Total Anthocyanin Content Determination

pH diferential assay as suggested by Giusti and Wrolstad (2001) (2001) was used to determine the total anthocyanin content (TAC) of a sample extract. One milliliter of crude extract was diluted to 10 mL, either with a buffer solution of pH 1.0 or pH 4.5, and stored in dark for 15 min. One milliliter of distilled water and 9 mL of the corresponding bufer solutions were used as blanks. Absorbances were measured at 520 nm and 700 nm using a spectrophotometer (Unico Instruments, UV-2600, Shanghai, China). Monomeric anthocyanins content is expressed as mg cyanidin 3-glucoside equivalent (C-3-G equivalent) and was calculated as follows:

$$
A = (A_{530} - A_{700})_{\text{pH 1.0}} - (A_{530} - A_{700})_{\text{pH 4.5}}
$$

$$
A \times V \times DF \times MW
$$

TAC(mg C-3-G equivalent/g) = $\frac{A \times V \times DF \times MW}{\varepsilon \times m}$

where *V* is the total volume of the extract (mL), DF is the dilution factor, MW is the relative molecular weight of cyanidin-3-glucoside (449.2), ε is the extinction coefficient (26,900), and *m* is the mass of the blueberry powder (g). TAC is expressed in terms of mg cyanidin 3-glucoside equivalent per g dry fruit (mg C-3-G equivalent/g, d.b.). Each measurement was performed in triplicate.

Total Phenolic Content Determination

Total phenolic content (TPC) of a sample was determined by the Folin–Ciocalteu assay as recommended by Lao et al. [\(2019\)](#page-10-12). One milliliter of crude extract was combined with 5 mL of Folin–Ciocalteu reagent in a test tube; 4 mL of sodium carbonate (7.5% v/v) was then mixed. After 1 h of incubation at room temperature, the absorbance was measured at 765 nm using the same spectrophotometer. TPC is expressed in terms of mg gallic acid equivalent per g of dry fruit (mg GAE/g, d.b.). Each determination was performed in triplicate.

Fourier Transform Infrared Spectroscopy

FTIR spectra of freeze-dried powdery samples were obtained using an FTIR spectrometer (Thermo Fisher Scientific, Nicolet[™] iS[™] 10, Waltham, MA, USA). The measurement was performed at 4000–400 cm−1 with a resolution of 2 cm−1 and scan speed of 128. Twenty milligrams of a sample was ground with 2 g of potassium bromide and made into a tablet prior to each measurement.

Statistical Analysis

The experimental data were subject to one-way analysis of variance (ANOVA) and are reported as mean values with standard deviations. Signifcances among mean values were established using Duncan's new multiple range tests at a confdence level of 95%. SPSS 23 (IBM, Chicago, IL, USA) was used for all statistical computations.

Results and Discussion

Drying Curves of Blueberries

Figure [3](#page-5-0) shows the drying curves in terms of the moisture ratio of blueberries undergone diferent pretreatments. The moisture ratio not only indicates the remaining moisture inside a raw material but also refects the drying rate (Vega-Galvez et al., [2012](#page-11-9); Wu et al., [2020\)](#page-11-10). The moisture content (wet basis) of the samples right after $CO₂$ laser perforation, ultrasonic, and freezing–thawing pretreatments were 89.04%, 91.52%, and 88.45%, respectively. The typical decreasing trend of moisture with time was observed in all cases.

CO₂ laser perforation and ultrasonic pretreatments could, in descending order, help accelerate the drying process when compared with freezing–thawing pretreatment. $CO₂$ laser perforation created pores on the skin of the fruit. This provided channels for water transport from inside of the fruit to its exterior. Munzenmayer et al. [\(2020](#page-10-13)) arrived at a similar conclusion; these investigators observed that laser perforation could reduce the freeze drying time of blueberries from 17 to 13 h. In the case of ultrasonic pretreatment, the enhancement is attributed to the cavitation and mechanical efects of ultrasound on the tissue structure of blueberries. This is in accordance with the results of Ziying et al. ([2018\)](#page-11-11) who reported that the drying rate of ultrasonically pretreated apple was 1.25 times higher than that of the control sample. Gao et al. [\(2018](#page-10-14)) reported indeed that ultrasonically induced cavitation bubbles helped destroy the stratum corneum structure of the epidermis.

Freezing–thawing treatment, while might being capable of reducing the skin thickness and damaging the internal structure of blueberries (Zielinska et al., [2015](#page-11-5)), did not exert any signifcant enhancement efect in this case. This is probably because ice crystals produced by rapid freezing (−65 °C) were smaller, thus resulting in negligible damage to blueberry structure during thawing.

Physical Properties of Blueberries

Color of Blueberries

It is seen in Table [1](#page-5-1) that, compared with the control sample, ultrasonic pretreatment exhibited a signifcant impact on the color of blueberries; *L**, *a**, and *b** values of the fruit increased when ultrasonic pretreatment was applied. On the other hand, no signifcant diferences in color were observed among the control sample and those undergone $CO₂$ laser perforation and freezing–thawing pretreatments. Note that *L**, *a**, and *b** values of the frozen blueberries were 26.35 ± 0.67 , 3.51 ± 0.52 , and -1.27 ± 0.23 , respectively.

Compared with frozen blueberries, *L** value of all dried samples decreased, except for those undergone ultrasonic pretreatment. It has been reported that shrinkage and structural deformation during drying may have shifted photons or absorbed more light, thus resulting in the decreased L^* values (da Silva Júnior et al., [2018\)](#page-10-15). Ultrasonically pretreated dried blueberries became lighter, as shown in Fig. [4.](#page-6-0) Δ*E* value of freeze-dried blueberries that undergone diferent pretreatments could be summarized as follows: ultrasonically treated $>$ CO₂ laser perforation treated $>$ freezing–thawing treated>control. Except for the ultrasonically treated fruit, blueberries rather retained their original color. This observation was related to the loss of anthocyanins during the ultrasonic pretreatment process. This observation is in accordance with that of Siucińska et al. ([2016\)](#page-11-12) who reported that Δ*E* value of dried sour cherries that undergone ultrasonic pretreatment was higher than that of the untreated sample. Cavitation-induced damage was noted to be responsible for such a change.

Shrinkage of Blueberries

Shrinkage of the samples ranged from 25.45 to 56.67%. Samples undergone ultrasonic, freezing–thawing, and $CO₂$ laser perforation suffered shrinkage in ascending order; control sample sufered most extensive shrinkage (see Table [1](#page-5-1) and Fig. [4](#page-6-0)). Pretreatment could reduce the degree of shrinkage when comparing the pretreated samples with the control one. This is due to the fact that pretreatments could help enhance the moisture transport capability of the fruit during drying. Ultrasonic pretreatment maintained the original volume of blueberries to the highest extent, with the degree of shrinkage of 25.45%. This trend of results is in accordance with that of Guo et al. [\(2020\)](#page-10-16) who observed that ultrasonic pretreatment not only changed the internal tissue structure of the berries to form microporous channels but also destroyed the fruit cell walls to increase the rate of water transport during the drying process, thereby reducing the deformation of the fruit during drying. In the case of freezing–thawing

Data are given as means \pm standard deviations. Different lowercase letters 3 within the same column indicate that mean values are significantly different $(p < 0.05)$

Fig. 4 Dried blueberries treated by diferent pretreatments

 $CO₂$ laser perforation

Freezing-thawing

Ultrasonic

pretreatment, Tatemoto et al. (2016) reported that this pretreatment could help prevent carrot tissue deformation and shrinkage during drying. Laser-perforated blueberries, on the other hand, sufered more extensive shrinkage. Mayor and Sereno ([2004\)](#page-10-17) suggested that shrinkage of plant tissues during drying is caused by the imbalance of pressures inside and outside of the tissues. The more rapidly the water is removed, the higher the moisture gradients and the more extensive shrinkage. The higher rate of drying caused by the laser-perforation pretreatment might be responsible for the observed more extensive shrinkage.

Rehydration Capacity of Blueberries Rehydration capacity is an important quality attribute of a dried product; rehydration characteristics of dried foods are associated with changes in the cellular structure of the foods induced by pretreatment or dehydration process (Xie et al., [2017\)](#page-11-14). Figure [5](#page-6-1) shows the rehydration kinetics of dried blueberries that undergone different pretreatments. The rehydration capacities of blueberries expectedly increased with the rehydration time; the rate of increase nevertheless decreased with time. Rehydration capacities of differently pretreated samples were noted to be signifcantly diferent. The rehydration rate of ultrasonically pretreated blueberries was the highest; the rehydration curve tended to reach the plateau only after 10 min of rehydration, indicating that the water adsorption had reached the maximum. Rapid and complete rehydration is the desired property of a dried product as faster rehydration and higher rehydration ratio illustrate the superior water reconstitution characteristics of the product; such superiority indicates less damage of the cellular structure of the product (Wang et al., [2018\)](#page-11-15).

The aforementioned rehydration behavior is believed to be related to the blueberry tissue structure. Ultrasonic cavitation exhibited signifcant efect on the blueberry tissue and resulted in microporous structure prior to drying. This led to

Fig. 5 Rehydration characteristics of dried blueberries treated by different pretreatments

increased water transport (reconstitution) capability of the dried sample. This fnding is consistent with that of Alolga et al. ([2020](#page-10-18)) on garlic slices. The trend of the rehydration capacity was opposite to that of shrinkage. Less extensive shrinkage implies that a dried sample sufers less damage and can easier adsorb water and hence rehydrate more rapidly. On the contrary, a sample with a higher degree of shrinkage would exhibit a lower rehydration capability. Jayaraman et al. [\(1990\)](#page-10-19) also observed that the contraction of capillaries in plant tissues led to the restricted rehydration of caulifowers.

Exception to the abovementioned argument was observed in the case of $CO₂$ laser perforation. Although shrinkage of the laser-treated sample was more extensive than that of the sample treated by freezing–thawing pretreatment, the rehydration capability of the former was higher. This is most probably ascribed to the pores that formed on the skin of the fruit. Perforation is indeed often used as an effective method to improve the rehydration capacity of a dried product.

Bioactive Compound Contents

Total Anthocyanin Content

Figure [6](#page-7-0) shows the TAC of the samples that undergone different pretreatments. As expected, fresh and dried blueberries

contents of blueberries treated by different pretreatments. Different letters over the bars imply that values are signifcantly different $(p < 0.05)$

Fig. 6 Total anthocyanin

had significantly different TAC, indicating that anthocyanins in the blueberries degraded during drying; anthocyanins are indeed known to be sensitive to heat (Wojdylo et al. [2014\)](#page-11-16). Comparing with the control sample, only the CO₂ laser perforation treatment significantly reduced TAC; there was no signifcant diference among the ultrasonically treated, freezing–thawing treated, and control samples. Note that TACs of mature blueberry peels were 7 and 192 times of those of the whole fruit and pulp, respectively (Ribera et al., [2010](#page-11-17)). Heat involved in the laser treatment of the blueberry epidermis is then believed to result in the extensive loss of anthocyanins.

Although not significantly different, the content of anthocyanins in the sample treated with ultrasound was slightly higher than that of the control sample. Zhang et al. ([2020](#page-11-18)) observed similar results; the content of anthocyanins in ultrasonically treated strawberries was higher than that in the untreated control sample. This may be related to the extraction of anthocyanins from the treated blueberries. Tiwari et al. ([2010](#page-11-19)) illustrated that ultrasound could promote dissolution of anthocyanins; anthocyanin extraction (for quantification purpose) was therefore more efficient. In the case of freezing–thawing pretreatment, ice crystals might have damaged the cellular structure of the fruit, resulting in juice leakage and hence the slightly more extensive loss

of anthocyanins, which is water-soluble in nature (Nowak et al., [2019\)](#page-11-20).

Total Phenolic Content

The effects of different pretreatments on the TPC of blueberries are shown in Fig. [7.](#page-8-0) Phenolics are among the major health benefcial compounds in blueberries; phenolics are also sensitive to heat and oxygen. TPC of blueberries significantly reduced upon drying. No signifcant diferences were noted among the control sample as well as those undergone CO₂ laser perforation and ultrasonic pretreatment; freez– ing–thawing pretreatment, however, exerted a signifcant adverse infuence on TPC of the sample. TPC of blueberries treated with ultrasound was signifcantly higher than that treated with freezing–thawing pretreatment, again probably because ultrasonic pretreatment helped promote release of phenolic compounds from the matrix of blueberries.

Among the three pretreatment methods, $CO₂$ laser perforation and ultrasound could efectively prevent bursting of blueberries during drying. Interestingly, at the end of drying, except for the sample pretreated by ultrasound, surface of blueberries appeared sticky; the sticky appearance was due to sugar crystallization. Similar phenomenon was observed by Zielinska et al. [\(2015\)](#page-11-5) who reported that the form and number of crystals in blueberries were different when different drying methods, which led to diferent degrees of sugar crystallization, were employed.

FTIR Spectra

Figure [8](#page-9-0) shows FTIR spectra of the powders prepared from dried blueberries undergone diferent pretreatments. The main characteristic peaks of the samples appeared at 3400 cm^{-1} , 2927 cm^{-1} , 1640 cm^{-1} , 1734 cm^{-1} , and 1060 cm−1. The strong absorption bands of blueberries were between 3000 and 3500 cm⁻¹, which correspond to the tensile vibrations of O–H and N–H (Radzki et al., [2016\)](#page-11-21). The peak at 2927 cm−1 is related to the C-H stretching vibration (Li et al., [2010\)](#page-10-20), while the absorption peak near 1640 cm^{-1} represents the stretching vibration of $C = C$ bond of the aromatic group of anthocyanins (Beullens et al., [2006](#page-10-21)). The peak at 1734 cm^{-1} is related to the stretching vibra-tion of C=O (Tao et al., [2017\)](#page-11-22), while the one at 1060 cm⁻¹ is related to the stretching vibration of C-O. The peak at $950 \sim 750$ cm⁻¹ is mainly the absorption peak of polysaccharides, while the absorption peaks of inorganic elements should be at wavenumbers lower than 700 cm^{-1} .

Peak positions were similar for both the control and pretreated samples, indicating that the pretreatments did not

Fig. 7 Total phenolics contents of blueberries treated by dif‑ ferent pretreatments. Diferent letters over the bars imply that values are signifcantly diferent $(p < 0.05)$

Table 2 Ratio of each peak after normalization at 1424 cm⁻¹

Data are given as means \pm standard deviations. Different lowercase letters 7 within the same column indicate that mean values are significantly different $(p < 0.05)$

alter the molecular structure of the fruit powder. The peaks of the infrared spectrum were normalized at 1424 cm−1 and then semi-quantitatively analyzed. The ratio of each peak is shown in Table [2](#page-9-1). There were diferences in the absorption peak intensities belonging to the control and pretreated samples, indicating that the pretreatments exhibited some efects on the vibration intensity of the original functional groups of blueberries. Note that the O–H stretching vibration was more intensive than the C-H stretching one, indicating that blueberries contained a relatively large number of OH group (Tang et al., [2019](#page-11-23)). It is indeed generally believed that hygroscopicity is related to the number of OH group (Bichot et al., [2020](#page-10-22)). Tang et al. [\(2019](#page-11-23)), for example, illustrated that hygroscopicity of eosinophilic pollen was determined by the OH group of organic compounds contained within such a pollen; FTIR spectra revealed good correlation between hygroscopicity of the pollen and its number of OH group. In our case, it can be said that the pretreatments are benefcial to the alleviation of the hygroscopicity of blueberries. Further research is required to determine the moisture adsorption characteristics of blueberries, however.

Conclusion

The effects of $CO₂$ laser perforation, ultrasonic, and freez– ing–thawing pretreatments on the drying characteristics and selected properties of infrared freeze-dried individually quick frozen blueberries were investigated. Laser perforation was noted to efectively improve the drying rate of blueberries by causing physical damage to the skin of the fruit. While exerting no signifcant impact on the color of the dried product, laser perforation helped reduce shrinkage,

increase rehydration capability, and enhance total phenolics content of the dried fruit. Ultrasonic pretreatment, on the other hand, significantly affected the color of dried blueberries but, at the same time, reduced shrinkage, improved rehydration capability and enhanced total anthocyanins content of the dried fruit. The three pretreatments may have the potential to reduce hygroscopicity of freeze-dried blueberries. Interestingly, however, only ultrasonic pretreatment was capable of preventing blueberries from cracking during drying, thus avoiding undesirable adhesion between individually dried fruit and hence its enhanced appearance and prolonged storage life. Ultrasonic pretreatment is therefore recommended as the most appropriate pretreatment for blueberries prior to infrared freeze drying. Further study on how to avoid or reduce the loss of blueberry skin color during ultrasonic pretreatment is recommended.

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Data Availability The data are available from the corresponding author upon suitable request.

Declarations

Conflict of Interest The authors declare no competing interests.

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