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A Cassava Starch–Chitosan Edible Coating Enriched with *Lippia sidoides* Cham. Essential Oil and Pomegranate Peel Extract for Preservation of Italian Tomatoes (*Lycopersicon esculentum* Mill.) Stored at Room Temperature

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Abstract

The effect of edible cassava starch–chitosan coatings incorporated with rosemary pepper (*Lippia sidoides* Cham.) essential oil and pomegranate peel extract on the shelf-life of tomatoes during storage at 25 °C for 12 days was investigated. Sixteen formulations, containing 10 g L⁻¹ cassava starch and various concentrations of chitosan (5, 10, 20, 30 g L⁻¹), essential oil (0, 2.5, 5, 10 mL L⁻¹) and pomegranate peel extract (0, 5, 10, 20 mL L⁻¹) were prepared and applied to tomatoes. Physical–chemical and microbiological analyses were performed on days 1, 4, 8 and 12. Most of the coatings delayed the ripening of tomatoes, lowering the total soluble solids (38–44 g sucrose kg⁻¹) and weight loss (93–128 g kg⁻¹) and maintaining constant firmness compared to the uncoated tomatoes (45 g sucrose kg⁻¹, 175 g kg⁻¹) at 12 days of storage. Conversely, except red intensity (*a**), which was higher for the uncoated samples, the colour parameters (*L**, *b**) of the coating showed potential to maintain the quality of tomatoes during storage at 25 °C for 12 days. In this context, tomatoes coated with the formulation comprising 10 g L⁻¹ cassava starch, 10 g L⁻¹ pomegranate peel extract showed the lowest weight loss and reduced total soluble solids content compared with uncoated ones.

Keywords Natural compounds · Fruit residue · Shelf-life · Postharvest treatment · Quality control

Introduction

Edible coatings have proven to be highly useful in food preservation, acting as a barrier to gases and water vapour, improving food appearance and controlling microbial

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contamination (Aider 2010). Chitosan (CH) is of interest as an edible film component, as it is a natural, non-toxic, biocompatible and biodegradable polymer, with antimicrobial activity and excellent oxygen barrier properties (Elsabee and Abdou 2013). One of the most important polysaccharides used in the formulation of biodegradable films and edible coatings is starch. Cassava starch (CS) is particularly useful in coatings as it is tasteless, odourless and transparent, thereby retaining the original taste, aroma and appearance of the product (Garcia et al. 2010; Chiumarelli and Hubinger 2012; Chiumarelli and Hubinger 2014). Besides, this product is cheap and abundant in Brazil, for this reason, it was considered the use of cassava starch as one of components of edible coatings in this work. In this context, several researchers have developed coatings or films based on cassava starch and chitosan blends (Azevedo et al. 2014; Aquino et al. 2015; Frazão et al. 2017; Valencia-Sullca et al. 2018; Zhao et al. 2018).

The tomato (Solanum lycopersicon L.) is a climacteric fruit, consumed mainly in a natural or processed form (in sauces, purees, ketchup and others). However, this fruit has a relatively short postharvest life due to several factors, such as transpiration, postharvest diseases and increased ripening and senescence, which accelerate the deterioration during storage and transport, causing a serious problem for commercialisation (Ali et al. 2010; Nawab et al. 2017). Moreover, in tropical regions, where the temperature is high, tomatoes show rapid ripening and deterioration, due to the increased respiration of the fruit (Bailén et al. 2006). Consequently, numerous researchers have developed various edible coating formulations to extend the shelf-life of tomatoes (Mehyar et al. 2011; Ramos-García et al. 2012; Das et al. 2013; Fagundes et al. 2014; Fagundes et al. 2015; Guerra et al. 2015; Rodriguez-Garcia et al. 2016; Nawab et al. 2017).

Bioactive compounds, such as plant extracts, are increasingly being incorporated in edible coatings to further extend the shelf-life and facilitate the processing and consumption of food (Sung et al. 2013; Azevedo et al. 2014; He et al. 2014; Aquino et al. 2015; Frazão et al. 2017; Wang et al. 2017). Among compounds with potential use in edible coatings, the essential oil (EO) from rosemary peeper (Lippia sidoides Cham.) has shown high pharmacological and commercial value, attributed to the presence of thymol and carvacrol monoterpenes, as major constituents in the leaves. These compounds are responsible for insecticidal, fungicidal, leishmanicidal, larvicidal, acaricidal and anti-inflammatory activities (Cavalcanti et al. 2010; Veras et al. 2012; Lima et al. 2013). Thymol and carvacrol have been accepted as flavouring agents in Europe, as they are not considered to present any health risk to consumers at the estimated intake level (Hyldgaard et al. 2012). However, the incorporation of EOs in active food packaging requires additional research to establish the effective and safe concentrations of these ingredients (Seow et al. 2014).

Another useful source of plant bioactives is pomegranate peel extracts (PPEs), which have been highlighted as potent, natural inhibitors of several bacteria (e.g. Bacillus subtilis, Bacillus coagulans, Bacillus cereus, Escherichia coli, Klebsiella pneumoniae) and fungi (e.g. Aspergillus niger, Mucor indicus, Penicillium citrinum, Rhizopus oryzae, Trichoderma reesei, Rhizopus stolonifer, Botrytis cinerea) (Dahham et al. 2010; Tehranifar et al. 2011; Akhtar et al. 2015; Nicosia et al. 2016). Pomegranate extracts have been applied to meat surfaces, chicken breasts and fish, and have also been implicated as antibacterial therapeutics to ameliorate diseases and topical infections, such as those in the oral cavity or on the skin surface (Howell and D'Souza 2013; Tanveer et al. 2015). The extracts of pomegranate peels have demonstrated wound healing properties, immunomodulatory activity, antibacterial activity and antiatherosclerotic and antioxidative capacities. Although PPE has been used in CH- and alginate-based coatings, to extend the postharvest quality of guava (Nair et al. 2017), there are currently no reports about the effects of PPE incorporated in active packagings on consumer health (Goula and Lazarides 2015).

In this context, the current work evaluated the effect of edible CS-CH coatings enriched with *L. sidoides* EO and PPE on the physical–chemical and microbiological quality of tomatoes stored at 25 °C for 12 days.

Materials and Methods

Materials

L. sidoides leaves, collected at Mossoró county, Rio Grande do Norte, Brazil, were cultivated in the Research Farm of the Federal University of Sergipe, Department of Agronomical Engineering, São Cristóvão, Brazil. For extraction of EO. the leaves were pre-dried at 40 °C in a forced air oven (Marconi MA 037, Piracicaba, São Paulo, Brazil) for 48 h and pulverised using a mill (Grindomix GM 200, Haan, Germany) (Cavalcanti et al. 2010). The EO (major components, thymol (0.7 mL L^{-1}) and *p*-cymene (0.1 mL L^{-1})) was obtained by hydrodistillation using Clevenger equipment and donated by the Department of Agronomical Engineering of Federal University of Sergipe, São Cristóvão, Brazil. Powdered CH (80 mesh particle size, 859 g kg⁻¹ deacetylation degree, pH 8.2) was obtained from Polymar (Fortaleza, Brazil), and CS (Dinha Bá brand) (850 g kg⁻¹ carbohydrates, 0.45 g kg^{-1} calcium and 0.03 g kg^{-1} iron) was purchased from "Cachoeira Alimentos e Bebidas Ltda" company (São Cristóvão, Sergipe, Brazil). Italian tomatoes, without peduncles and sepals (tomatoes designated as the Italian group cultivated in São Paulo and Minas Gerais, Brazil, characterised by elongated fruits of 7 to 10 cm, with a transverse diameter of 3 to 5 cm, intense red colour, bilocular, thick wall and sweet taste), were purchased from Antônio Franco market of Aracaju (Brazil) and transferred to the laboratory in sterile boxes at room temperature within 1 h. The number of tomatoes used for each formulation was purchased separately, and the experiments were started on the same day. The mature fruits (with green traces) were selected for size uniformity, without visible physical damage and microbial deterioration.

PPE Preparation

The fruit was sanitised by immersion in 0.2 g L⁻¹ chlorinated solution for 15 min. The peels were then manually separated from the pulp and dried at 50 °C for 24 ± 2 h before they were triturated and stored in sterile glass bottles, until use. To obtain the PPE, the peels (5 g) were mixed with 10 mL methanol (600 mL L⁻¹) under agitation (Solab SL222 orbital shaker, Brazil) at 250 rpm/45 °C for 35 min, based on Gullon et al. (2016), with modifications.

Preparation of Edible Coatings

Sixteen formulations of edible coatings were prepared, as described by Azevedo et al. (2014), adapted from Vasconez et al. (2009), using CS (10 g L⁻¹) and various concentrations of CH (5, 10, 20, 30 g L⁻¹), *L. sidoides* EO (0, 2.5, 5, 10 mL L⁻¹) and PPE (0, 5, 10, 20 mL L⁻¹) (Table 1). CH was dissolved (w/v) in acetic acid solution (0.26 mol L⁻¹) at 25 °C, and glycerol (12.8 g L⁻¹) was then added, to complete the volume to 100 mL. A CS solution (100 mL) was prepared in glycerol (6.4 g L⁻¹). The starch solution was stirred in a heated water bath not exceeding 70 °C. After cooling to 25 °C, the solution was added to the CH solution and stirred until it was completely homogeneous. Then, the PPE and *L. sidoides* EO were added to the mixture. All experiments were performed in triplicate.

Application of Edible Coatings to Tomatoes

The tomatoes were sanitised by immersion in sodium hypochlorite solution (0.2 g L^{-1}) for 15 min, rinsed with water and dried at room temperature on plastic trays. Then, groups of 10 tomatoes were immersed in each formulation, dried at room temperature for 30 min (all tomatoes showed visually uniform texture) and stored on aluminium trays in biochemical oxygen demand chambers (SP Labor, Brazil) with internal forced air circulation by micro-ventilators, at 25 °C and 86–89% relative humidity for 1, 4, 8 and 12 days (i.e. 10 tomatoes per day of storage for each formulation, totalling 40 tomatoes per formulation). A total of 640 tomatoes for 16 formulations, plus 40 uncoated tomatoes, were used. The physical–chemical and microbiological analyses were performed at 1, 4, 8 and 12 days of storage, for each applied formulation. Uncoated tomatoes were used as the control.

pH, Titratable Acidity and Total Soluble Solids Analyses

Five grams of each fruit was homogenised with 50 mL distilled water and the pH value recorded (HI 9219 pH meter, Hanna Instruments, Woonsocket, RI, USA). The titratable acidity (TA) was measured using 5 g of each tomato macerated and homogenised with 50 mL distilled water, with 0.1 N NaOH as the titrant, and the results expressed as g citric acid kg⁻¹, as described by Bassetto et al. (2005), with modifications. The total soluble solids (TSS) was determined on 5 g of macerated fruit sample, using a bench refractometer and the results were expressed as g sucrose kg⁻¹ (Guerra et al. 2015). The measurements were taken from two tomatoes at 1, 4, 8 and 12 days of storage at 25 °C.

Weight Loss

For determining the weight loss, the fruit was weighed at 0, 1, 4, 8 and 12 days of storage (10 fruit per interval) on a balance with 0.001 g accuracy (Radwag brand, Poland). The total weight loss was calculated as a percentage of the initial weight (Ali et al. 2010).

Tomato Firmness

The firmness of uncoated and coated tomatoes was analysed using a Brookfield CT3 Texture Analyser (Middleboro, USA), equipped with a 2-mm diameter probe, and maximum load cell of 25 kg. The central region (three points) of each tomato (three fruit per interval) was compressed at a rate of 1 mm s⁻¹, using a 4.0 to 8.6 N load cell. Firmness (N) was defined as the maximum force required to penetrate to a depth of 10 mm. The analysis was performed at 1, 4, 8 and 12 days of storage (Aquino et al. 2015).

Colour Parameters

Superficial colour alterations were monitored using a CR-400 Minolta colourimeter (Osaka, Japan) on three tomatoes per interval of storage. The International Commission on Illumination (CIE 1986) colour parameters, a^* (redness), b^* (yellowness) and L^* (luminosity), were directly recorded on the surface of the tomatoes from three regions, at 1, 4, 8 and 12 days of storage at 25 °C.

Microbiological Analyses

For microbiological analyses, tomatoes stored at 1, 4, 8 and 12 days were placed in a sterile plastic bag (five fruit per interval) with saline solution (8.5 g L^{-1}) (corresponding to a 10^{-1} dilution) and homogenised. Then, successive dilutions $(10^{-2} \text{ and } 10^{-3})$ were obtained. The yeasts and moulds were enumerated, by the surface-seeding technique with potato dextrose agar and incubation at 35 °C for 5 days. The results were expressed as colony-forming units (CFU) per gram, as detailed by Rojas-Grau et al. (2007), with modifications. The most probable number (MPN) of thermotolerant coliforms was determined as described in the Bacteriological Analytical Manual (FDA 2010). First, tubes containing lauryl sulphate tryptose (LST) broth were incubated at 35 °C for 48 h in a bacteriological incubator, for a presumptive test. Then, tubes that exhibited gas production in the LST broth were transferred to tubes containing E. coli broth and incubated at 44.5 °C for 48 h (confirmatory test). The presence of thermotolerant coliforms was verified in tubes that exhibited growth with gas production in E. coli broth. The MPN of thermotolerant coliforms was calculated with the use of the Hoskins table (Vanderzant and Splittstoesser 2001).

Edible coatings containing CS (10 g L ⁻¹) PPE Formulations CH EO PPE 1 Formulations CH EO PPE 1 1 5 0 0 5.04^{aAB} \pm 0.05 2 5 2.5 5 4.84^{cdB} \pm 0.01 3 5 5 10 4.79^{eB} \pm 0.02 4 5 10 20 4.87^{bcC} \pm 0.02 5 10 20 4.87^{bcC} \pm 0.02 6 10 20 4.88^{bcC} \pm 0.01 7 10 2.5 5 4.68^{BB} \pm 0.01 7 10 2.5 5 4.68^{BB} \pm 0.01 8 10 2.5 5 4.79^{eBB} \pm 0.01 9 20 0 0 4.93^{bcC} \pm 0.02 10 20 5 4.93^{bcC} \pm 0.02 10 8 10 20 4.93^{bcC} \pm 0.02 10 10 20 5 4.93^{bcC} \pm 0.02 10	$\begin{array}{c c} 1 & & & \\ & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ &$	~					
$ \begin{array}{cccc} CH & EO & PPE \\ (g L^{-1}) & (mL L^{-1}) & (mL L^{-1}) \\ 5 & 0 & 0 \\ 5 & 2.5 & 5 \\ 5 & 10 & 20 \\ 10 & 0 & 0 \\ 10 & 2.5 & 5 \\ 10 & 2.5 & 5 \\ 10 & 10 & 20 \\ 10 & 10 & 20 \\ 20 & 0 & 0 \\ 20 & 5 & 10 \\ 20 & 5 & 10 \\ 20 & 5 & 10 \\ 20 & 5 & 10 \\ 20 & 10 & 20 \\ 20 & 5 & 10 \\ 20 & 10 & 20 \\ 20 & 10 & 20 \\ 20 & 10 & 20 \\ 20 & 20 & 20 \\ 20 & 20 & 20 \\ 20 & 20 &$		8		Titratable aci	Titratable acidity (g citric acid kg ⁻¹)	d kg ⁻¹)	
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5 2.5 5 5 5 5 10 5 10 0 0 10 2.5 5 10 10 2.5 5 5 10 2.5 5 5 10 2.5 5 5 20 0 0 20 20 2.5 5 5 20 2.5 5 5 20 5 10 20		$4.93^{efB}\pm0.03$	$5.07^{deA}\pm0.09$	$21^{hB} \pm 1$	$29^{fgA}\pm 2$	$25^{efA} \pm 2$	$17.3^{fB}\pm2$
5 5 5 10 5 10 0 20 10 2.5 5 5 10 2.5 5 10 10 2.5 5 5 10 10 20 20 20 2.5 5 5 20 2.5 5 5 20 5 10 20 20 5 10 20		$4.97^{efA}\pm0.02$	$4.92^{\mathrm{fA}}\pm0.05$	$29^{efA}\pm0$	$29^{fgA}\pm2$	$25^{efb} \pm 2$	$26^{bcAB}\pm2$
5 10 20 10 0 0 0 10 2.5 5 10 10 5 10 20 20 0 0 20 20 2.5 5 5 20 2.5 5 5 20 2.5 5 5 20 5 10 20	$79^{efb} \pm 0.08$ $5.08^{bA} \pm 0.05$	$4.98^{deA}\pm0.05$	$5.00^{\mathrm{efA}}\pm0.03$	$31^{deAB}\pm0$	$33^{deA}\pm1$	$30^{bcAB} \pm 1$	$29^{abB}\pm3$
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$37^{bcC} \pm 0.02$ $4.95^{cdB} \pm 0.03$	$4.98^{deB}\pm0.03$	$5.09^{\rm dA}\pm0.03$	$31^{deA}\pm0$	$32^{deA}\pm2$	$32^{abcA}\pm1$	$33^{aA}\pm 2$
10 2.5 5 10 5 10 10 10 20 20 0 0 20 2.5 5 20 5 10 20 5 10 20 5 10 20 10 20	$36^{\text{cdC}} \pm 0.01$ $4.99^{\text{cA}} \pm 0.01$	$5.01^{\mathrm{cdA}}\pm0.01$	$4.92^{fB}\pm0.01$	$26^{fgB}\pm1$	$32^{deA}\pm3$	$29^{\mathrm{cdAB}}\pm0$	$28^{abAB}\pm1$
10 5 10 10 10 20 20 0 0 20 5 5 20 10 20	$58^{hB} \pm 0.01$ $4.78^{gAB} \pm 0.06$	$4.91^{\rm fA}\pm0.08$	$4.90^{\mathrm{fA}}\pm0.10$	$34^{cdA}\pm 1$	$34^{cdA}\pm2$	$20^{fgB}\pm0$	$18^{fC}\pm3$
10 10 20 20 0 0 20 20 2.5 5 20 5 10 20 10 20	$32^{\text{ceD}} \pm 0.06$ 5.11 ^{bB} ± 0.03	$4.91^{\rm fC}\pm0.04$	$5.26^{bcA}\pm0.03$	$33^{cdB}\pm2$	$32^{deB}\pm1$	$38^{aA}\pm 1$	$22^{deC}\pm3$
20 0 0 20 2.5 5 20 5 10 20 10 20	$33^{bcC} \pm 0.01$ $4.93^{deC} \pm 0.04$	$5.07^{bcB}\pm0.04$	$5.19^{\mathrm{cA}}\pm0.02$	$29^{efB}\pm2$	$44^{\mathrm{aA}}\pm0$	$29^{bcB} \pm 3$	$32^{aB}\pm3$
20 2.5 5 20 5 10 20 10 20	$92^{bcC} \pm 0.06$ $5.00^{cB} \pm 0.02$	$4.97^{efBC}\pm0.02$	$5.24^{bcA}\pm0.02$	$30^{efAB}\pm3$	$24.8^{hB}\pm1$	$32^{bcA} \pm 4$	$31^{abA}\pm2$
20 5 10 20 10 20	$72^{ghC} \pm 0.02$ $4.92^{efB} \pm 0.02$	$5.00^{ m deA}\pm0.02$	$4.69^{hC}\pm0.06$	$36^{cA}\pm3$	$38^{bcA}\pm1$	$20^{\mathrm{gB}}\pm4$	$24^{cdB}\pm3$
20 10 20	$88^{bcC} \pm 0.02$ $5.07^{bB} \pm 0.01$	$5.22^{\mathrm{aA}}\pm0.04$	$5.23^{bcA}\pm0.02$	$44^{aA} \pm 3$	$27^{ghB}\pm2$	$28^{\mathrm{cdB}}\pm4$	$20^{efC}\pm0$
20 10 20	$4.91^{bcC} \pm 0.02 \qquad 4.93^{deC} \pm 0.02$	$5.12^{\mathrm{bB}}\pm0.04$	$5.24^{bcA}\pm0.03$	$34^{cdB} \pm 3$	$44^{aA} \pm 4$	$29^{cdB} \pm 2$	$31^{abB}\pm2$
13 30 0 0 $5.05^{aB} \pm 0.02$	$05^{aB} \pm 0.02$ $5.00^{cBC} \pm 0.04$	$4.98^{deC}\pm0.02$	$5.23^{bcA}\pm0.05$	$27^{fgC}\pm 2$	$38^{bA}\pm 2$	$34^{abAB}\pm4$	$30^{abBC}\pm3$
14 30 2.5 5 $4.85^{cdB} \pm 0.01$	$35^{cdB} \pm 0.01$ $5.11^{bA} \pm 0.06$	$5.05^{ m cdA}\pm0.03$	$4.91^{\mathrm{fB}}\pm0.01$	$27^{\rm fgA}\pm4$	$15^{jB} \pm 1$	$30^{bcA}\pm2$	$30^{abA}\pm1$
15 30 5 10 $4.95^{bC} \pm 0.02$	$95^{bC} \pm 0.02$ $5.19^{aB} \pm 0.01$	$4.93^{ m efC}\pm0.01$	$5.28^{\rm bA}\pm0.04$	$24^{\mathrm{gB}}\pm1$	$11^{1C} \pm 1$	$28^{\rm cdAB}\pm1$	$30^{abA}\pm3$
16 $30 10 20 4.85^{cdD} \pm 0.04$	$85^{cdD} \pm 0.04$ $5.19^{aB} \pm 0.04$	$5.07^{ m bcC}\pm0.08$	$5.77^{\rm aA}\pm0.01$	$33^{cdA}\pm 2$	$18^{iC} \pm 1$	$26^{deB}\pm1$	$28^{abB}\pm4$
Uncoated tomatoes $4.70^{\text{fgA}} \pm 0.10$	$70^{fgA} \pm 0.10$ $4.78^{gA} \pm 0.02$	$4.82^{\rm gA}\pm0.05$	$4.78^{\rm gA}\pm0.06$	$40^{bA}\pm1$	$37^{bcB}\pm2$	$30^{bcC}\pm1$	$28^{abC}\pm1$

Statistical Analysis

The composition of edible coatings varied, according to a completely randomised design. All data were analysed using Assistat 7.7 beta software. The analyses were performed using descriptive statistics (mean and standard deviation). The differences among means were determined by Duncan's range test at a 5% significance level.

Results and Discussion

Effect of Edible Coatings on pH, TA, TSS and Weight Loss of Tomatoes

The pH, weight loss, TA and TSS content of uncoated and coated tomatoes were determined during the storage at 25 °C for 1, 4, 8 and 12 days. Citric acid is the most abundant acid in tomatoes and the largest contributor to the TA (90 g citric acid kg^{-1}). During maturation, there is an initial increase in the citric acid content, followed by a decrease over time until full maturity (Ali et al. 2010). Thus, an increased pH and decreased TA in tomatoes can be attributed to the decline in, primarily, citric acid concentration, as the fruit ripens (Das et al. 2013). The majority of the coated tomatoes (pH 4.62–5.77) showed a slight pH increase, with a significant difference (p < p0.05) and no difference (p > 0.05) in TA on days 1 and 12 of storage, respectively. Only tomatoes coated with formulations 6 (10 g L⁻¹ CS, 10 g L⁻¹ CH, 2.5 mL L⁻¹ EO, 5 mL L⁻¹ PPE), 7 (10 g L^{-1} CS, 10 g L^{-1} CH, 5 mL L^{-1} EO, 10 mL L^{-1} PPE), 10 (10 g L^{-1} CS, 20 g L^{-1} CH, 2.5 mL L^{-1} EO, 5 mL L^{-1} PPE), 11 (10 g L^{-1} CS, 20 g L^{-1} CH, 5 mL L^{-1} EO, 10 mL L^{-1} PPE) and 16 (10 g L^{-1} CS, 30 g L^{-1} CH, 10 mL L^{-1} EO, 20 mL L^{-1} PPE), as well as the uncoated tomatoes, showed a significant difference (p < 0.05) in TA from 1 to 12 days of storage, with reductions of 47, 33, 32, 55, 15 and 29 g citric acid kg^{-1} , respectively, at the end of storage, meaning they exhibited faster ripening than the others. However, comparing coated with uncoated tomatoes, there was no significant difference (p > 0.05) in TA on day 12 of storage, with the values maintained at around 30 g citric acid kg⁻¹ (Table 1), except tomatoes coated with formulations 1, 6, 7, 10 and 11.

The TSS content of various tomato varieties commonly ranges from 40 to 80 g sucrose kg⁻¹ (Cramer et al. 2001). According to Tigist et al. (2013), an initial increase of TSS content during maturation of fresh market tomato varieties, followed by a decrease over time, can be attributed to a gradual decline in the rate of hydrolysis of carbohydrates at the end of storage. In the present work, the TSS content ranged from 31 to 42 g sucrose kg⁻¹, and the coated tomatoes tended to show a slight but significant difference (p < 0.05) in TSS, throughout storage (Table 2). The TSS of coated and uncoated tomatoes differed statistically (p < 0.05) at 12 days of storage, with the uncoated fruit displaying the higher TSS content (45 g sucrose kg^{-1}), implying a relatively faster ripening.

The weight loss primarily results from transpiration and the loss of carbon atoms from fruit, in each cycle of respiration (Das et al. 2013). Most of the coated tomatoes showed a weight loss between 105 and 128 g kg⁻¹ on day 12 of storage, without significant differences (p > 0.05) among them (Table 2). The highest weight loss (175 g kg^{-1}) was recorded for the uncoated tomatoes, and the lowest (93 and 94 g kg⁻¹) corresponded to those coated with formulation 8 (10 g L^{-1} CS, 10 g L^{-1} CH, 5 mL L^{-1} EO, 10 mL L^{-1} PPE) and 11 $(20 \text{ g L}^{-1} \text{ CH}, 5 \text{ mL L}^{-1} \text{ EO}, 10 \text{ mL L}^{-1} \text{ PPE})$, at 12 days of storage. The weight losses were 4.2 to 7.7% less for coated tomatoes than those for uncoated ones (at 12 days of storage), demonstrating the efficacy of the coatings as moisture barriers. When stored at 24 °C for 20 days, Das et al. (2013) obtained a weight loss of 3.53% less for tomatoes coated with rice starch-based edible coating containing coconut oil and tea leaf extract than the control. For the same duration at 20 °C. Ali et al. (2010) detected 2% less weight loss for tomatoes coated with 10% gum arabic compared to the control. In comparison, the results in the present work show greater efficiency in reducing the weight loss of tomatoes during storage.

Most of the coated tomatoes maintained a constant firmness (p > 0.05) between 1 and 12 days of storage (Table 2). However, a higher firmness loss (77%) was noted in tomatoes coated with 10 g L⁻¹ CS, 30 g L⁻¹ CH, 2.5 mL L⁻¹ EO and 5 mL L⁻¹ PPE (formulation 14) than the others, while uncoated tomatoes showed a 22% loss of firmness. Probably, the high percentage of CH in the edible coatings contributed to the softening of the tomatoes. Ali et al. (2010) mentioned the softening of fruit during maturation reflects changes in the structure/composition of the cell wall and intracellular components, involving the enzymatic hydrolysis of pectin and starch.

In this work, the edible coatings contributed to a lower weight loss and firmness but maintained the pH and TSS of the tomatoes compared to the uncoated fruit. In contrast, Guerra et al. (2015) obtained no differences in TA, TSS and firmness among cherry tomato fruit treated with coatings comprising CH and Mentha piperita L. or Mentha \times villosa than the uncoated fruit, at 12 or 25 °C during 12 days of storage. Fagundes et al. (2014) also verified that a coating application (edible composite coatings based on hydroxypropyl methylcellulose (HPMC), beeswax and food preservatives) did not significantly affect the TA, pH and firmness of cherry tomatoes stored at 5 °C for 15 days, followed by 20 °C for 5 days. Dávila-Aviña et al. (2011) noticed tomatoes coated with carnauba and mineral oil edible coatings had a relatively constant TA and TSS content, but reduced weight loss and firmness. Ali et al. (2010) coated tomatoes with various concentrations of aqueous arabic gum and documented a delay in changes of weight, firmness, TA and TSS compared to the

pomegranate peet extract during storage at 22 °C 101 12 days	I extract duri	ng storage at 2.	2 -C IOL 17 0	lays										
			Time of sto	Time of storage (days)										
Edible coatings containing CS (10 g L^{-1})	ontaining CS	(10 g L^{-1})	Total soluble solid	ole solid cont	content (g sucrose kg^{-1})	e kg ⁻¹)	Weight loss (g kg ⁻¹)	ss (g kg ⁻¹)			Firmness (N)			
Formulations CH (g I	$\begin{array}{c} H \\ EO \\ L^{-1} \end{array} (mL I) \end{array}$	CH EO PPE $(g L^{-1})$ (mL L^{-1}) (mL L^{-1})	-	4	8	12	-	4	8	12	1	4	8	12
5	0	0	$38^{\rm fA}\pm0$	$35^{fB}\pm1$	$38^{bcdeA} \pm 0$	$39^{fgA}\pm1$	$12^{cdD}\pm0$	$34^{\text{eC}} \pm 1$	$65^{deB} \pm 7$	$106^{bcA}\pm 5$	$6.4^{abA}\pm0.7$	$6.0^{abAB}\pm0.9$	$5.4^{efB}\pm0.7$	$5.0^{\mathrm{deB}}\pm0.9$
2 5	2.5	5	$39^{efB}\pm1$	$40^{cdA}\pm1$	$40^{abcA}\pm0$	$41^{deA}\pm1$	$15^{bC}\pm1$	$65^{abB}\pm7$	$107^{bA}\pm3$	$126^{bcA}\pm 5$	$6.4^{abA}\pm0.7$	$6.3^{abA}\pm0.7$	$6.4^{abA}\pm0.6$	$6.2^{abA}\pm0.9$
3 5	5	10	$39^{deB}\pm0$	$40^{cdA}\pm0$	$40^{abcA}\pm0$	$38^{ghC}\pm1$	$12^{bcC}\pm 2$	$37^{cC}\pm4$	$79^{cdB}\pm 5$	$110^{bcA}\pm9$	$5.9^{cdA}\pm0.8$	$4.9^{\rm deB}\pm0.4$	$5.6^{cdAB}\pm0.6$	$5.7^{\text{bcAB}} \pm 0.6$
4 5	10	20	$38^{efB}\pm1$	$45^{aA}\pm3$	$42^{aAB}\pm2$	$41^{cdB}\pm1$	$10^{cdD}\pm1$	$40^{bcC}\pm1$	$70^{cdB}\pm 5$	$125^{bA}\pm7$	$5.4^{\rm fgAB}\pm0.8$	$5.8^{abA}\pm0.6$	$5.7^{cdAB}\pm0.8$	$4.0^{\text{efB}}\pm1.0$
5 1(0 (0	$41^{abB}\pm1$	$45^{abA}\pm1$	$35^{efD}\pm1$	$38^{hC}\pm0$	$8^{efD}\pm 2$	$37^{cC}\pm1$	$76^{cdB}\pm 8$	$105^{bcA}\pm3$	$6.1^{bcA}\pm0.7$	$5.7^{bcA}\pm0.6$	$5.9^{\mathrm{cdA}}\pm0.5$	$5.8^{bcA} \pm 0.8$
6 1() 2.5	5	$37^{gC}\pm1$	$42^{bcA}\pm0$	$39^{bcdB} \pm 1$	$39^{fgB}\pm1$	$9^{deD}\pm 1$	$42^{bcC} \pm 2$	$72^{cdB}\pm 5$	$110^{bcA}\pm6$	$7.0^{aA}\pm0.4$	$6.5^{aAB}\pm0.6$	$6.8^{abA}\pm0.5$	$5.9^{ m bcB}\pm0.9$
7 1() 5	10	$39^{efA}\pm1$	$31^{\rm gB}\pm1$	$33^{fB}\pm1$	$40^{efA}\pm0$	$\gamma^{fgC}\pm 1$	$40^{cBC}\pm9$	$70^{efAB}\pm7$	$115^{cdA}\pm7$	$5.6^{\rm deA}\pm0.8$	$5.1^{cdA}\pm0.6$	$5.5^{\rm efA}\pm0.8$	$5.0^{cdA}\pm1.0$
8 1() 10	20	$39^{efb} \pm 1$	$44^{\rm abA}\pm0$	$43^{aA}\pm2$	$40^{efB}\pm0$	$5^{gD}\pm 0$	$31^{cC}\pm 2$	$59^{fB}\pm1$	$93^{\rm dA}\pm7$	$6.1^{bcA}\pm0.5$	$6.0^{bcA} \pm 1.0$	$6.0^{bcA}\pm1.0$	$6.0^{bcA}\pm1.0$
9 2(0	0	$31^{hC}\pm1$	$34^{fgB}\pm0$	$36^{deB}\pm2$	$41^{cdA}\pm1$	$11^{cdD} \pm 1$	$50^{abC}\pm5$	$77^{cdB} \pm 6$	$113^{bcA}\pm 6$	$6.0^{bcA}\pm0.9$	$5.8^{abA}\pm0.6$	$5.3^{efA}\pm0.6$	$5.6^{bcA} \pm 0.9$
10 20) 2.5	5	$38^{fB}\pm0$	$42^{bcA}\pm0$	$38^{bcdeB}\pm0$	$43^{bcA}\pm1$	$9^{efD}\pm 1$	$34^{\text{cC}} \pm 3$	$71^{cdB}\pm2$	$105^{cdA}\pm2$	$7.0^{abA}\pm1.0$	$6.0^{abAB}\pm1.0$	$6.0^{deB}\pm1.0$	$6.3^{abAB}\pm0.4$
11 2() 5	10	$39^{cdD}\pm1$	$42^{bcB}\pm0$	$41^{abBC}\pm1$	$44^{abA}\pm0$	$11^{cdD} \pm 1$	$35^{cC}\pm0$	$65^{efB}\pm1$	$94^{\mathrm{dA}}\pm1$	$6.0^{bcA}\pm0.9$	$5.0^{cdA}\pm1.0$	$6.0^{bcA}\pm0.8$	$6.0^{bcA}\pm1.0$
12 20) 10	20	$42^{aA}\pm0$	$42^{bcA}\pm0$	$42^{aA}\pm0$	$42^{cdA}\pm0$	$9^{deD}\pm 0$	$36^{\text{cC}}\pm6$	$65^{efB}\pm7$	$105^{cdA}\pm7$	$5.3^{\rm gA}\pm0.5$	$5.7^{bcA}\pm0.8$	$6.0^{bcA}\pm0.6$	$5.4^{cdA} \pm 0.7$
13 30	0	0	$32^{hC}\pm0$	$32^{gC}\pm0$	$36^{deB}\pm0$	$38^{ghA}\pm1$	$10^{cdD}\pm0$	$39^{bcC} \pm 2$	$83^{cdB} \pm 4$	$128^{bcA}\pm 5$	$5.5^{efA}\pm0.6$	$4.8^{eAB}\pm0.6$	$5.0^{\mathrm{f}}\pm1.0$	$4.3^{fB}\pm0.7$
14 30) 2.5	5	$40^{cdAB}\pm0$	$39^{cdeB}\pm0$	$40^{abcAB}\pm0$	$41^{\text{deA}}\pm0$	$13^{bcD}\pm1$	$44^{bcC} \pm 4$	$86^{cB}\pm5$	$113^{bcA} \pm 3$	$6.0^{abA}\pm1.0$	$5^{cdA} \pm 1.0$	$6.5^{abA}\pm0.8$	$1.4^{gB}\pm0.2$
15 3() 5	10	$36^{\rm gA}\pm0$	$37^{efA}\pm1$	$38^{bcdeA}\pm0$	$38^{hA}\pm2$	$15^{bD}\pm3$	$49^{bcC} \pm 1$	$72^{cdB} \pm 3$	$113^{bcA} \pm 3$	$6.6^{abA}\pm0.9$	$6.4^{abA}\pm0.9$	$6.1^{bcA}\pm0.7$	$7.0^{aA}\pm1.0$
16 30) 10	20	$38^{\rm hA}\pm0$	$39^{\text{deA}}\pm1$	$37^{cdeA} \pm 1$	$35^{iB}\pm1$	$15^{bD}\pm1$	$47^{bcC} \pm 7$	$75^{cdB} \pm 6$	$114^{bcA}\pm2$	$7.0^{abA}\pm1.0$	$5.8^{abB}\pm0.4$	$7.1^{\rm aA}\pm0.5$	$6.4^{abAB}\pm0.6$
D	Uncoated tomatoes	itoes	$40^{bcB}\pm2$	$39^{cdeB} \pm 1$	$40^{abcB}\pm0$	$45^{aA}\pm1$	$21^{aD}\pm3$	$60^{aC}\pm2$	$129^{aB}\pm 6$	$175^{aA}\pm7$	$6.4^{abA}\pm0.6$	$6.5^{ab}\pm0.6$	$5.6^{\mathrm{cdAB}}\pm0.9$	$5.0^{\mathrm{deB}}\pm1.0$
^{a-i} For each parameter, different lower case letters in the same column or different upper case letters in the same row indicate significant differences ($p < 0.05$) between the mean values according to Duncan's test. <i>M</i> , mean; <i>SD</i> , standard deviation; CS, cassava starch; CH, chitosan; EO, <i>Lippia sidoides</i> essential oil; PPE, pomegranate peel extract. Values are means from three analytical experiments	neter, differen), standard de	t lower case let eviation; CS, ci	ters in the san assava starch,	ne column or ; CH, chitos:	: different upp m; EO, <i>Lippi</i>	er case lette a sidoides e	rs in the san ssential oil;	ne row indic ; PPE, pome	ate significal sgranate peel	nt differences l extract. Val	(p < 0.05) bet ues are means	ween the mean from three ana	values accordin Ilytical experime	g to Duncan's ents

uncoated control fruit. Das et al. (2013) detected a reduction in weight loss and TA, but an increase in TSS and a slight increase in pH of tomatoes coated with a rice starch-based edible film containing coconut oil and tea leaf extract. The same authors concluded that the coated fruit exhibited delayed ripening effects compared to the uncoated controls.

Most of the coated tomatoes maintained a constant pH and TA and lower TSS content, weight loss and firmness loss than uncoated ones at day 12 of storage, characterising a delay in fruit ripening.

Effect of Edible Coatings on Colour Parameters (*L**, *a**, *b**) of Tomatoes

The colour is an important quality parameter of tomato fruit, with redness predominant, being indicative of lycopene content, followed by carotenes (yellow to orange) and xanthophylls (yellow) (López et al. 2001; Ali et al. 2010). In contrast to the tomatoes coated with formulations 13-16, which contained 30 g L^{-1} CH and showed a constant luminosity, a reduced brightness was observed in the uncoated fruit and remaining coated fruit, which exhibited a significant difference between 1 and 12 days of storage (p < 0.05) (Table 3). Thus, the tomatoes coated with formulations 13 -16 displayed higher luminosity values than the control samples. Probably, the higher percentage of CH contributed to these results because a higher luminosity has previously been described for CH-coated fruits, which could be associated with the luminosity and high transparency of CH films (Oliveira et al. 2014).

Coated and uncoated tomatoes recorded increased a^* values during storage, with a significant difference (p < p)0.05) between days 1 and 12 while the b^* values remained predominantly constant, with no significant differences among the samples (p > 0.05) (Table 3). The uncoated tomatoes had higher a^* values than those coated with formulations 1 (10 g L^{-1} CS, 5 g L^{-1} CH), 2 $(10 \text{ g } \text{L}^{-1} \text{ CS}, 5 \text{ g } \text{L}^{-1} \text{ CH}, 2.5 \text{ mL } \text{L}^{-1} \text{ EO}, 5 \text{ mL } \text{L}^{-1}$ PPE), 5 (10 g L^{-1} CS, 10 g L^{-1} CH), 7 (10 g L^{-1} CS, 10 g L^{-1} CH, 5 mL L^{-1} EO, 10 mL L^{-1} PPE) and 16 $(10 \text{ g } \text{L}^{-1} \text{ CS}, 30 \text{ g } \text{L}^{-1} \text{ CH}, 10 \text{ mL } \text{L}^{-1} \text{ EO}, 20 \text{ mL } \text{L}^{-1}$ PPE), which showed no significant difference (p > 0.05)on the last day of storage. The results meant that these coatings were more efficacious at delaying the ripening of tomatoes. Overall, the colour parameters indicated that the applied coatings delayed ripening of the fruit relative to the control samples. Dávila-Aviña et al. (2011) also verified that mineral oil coatings delayed colour changes in tomatoes at the end of storage. Fagundes et al. (2014) stated that HPMC-beeswax edible coatings did not alter the colour parameters of cold-stored cherry tomatoes. Guerra et al. (2015) stated that the application of coatings comprising CH and M. piperita L. or Mentha \times villosa Huds. EO presented no adverse effect on lycopene formation in red cherry tomatoes. Das et al. (2013) delayed the colour (L^* , $a^* b^*$) change in tomatoes during 20-day storage, by coating the fruit with a film containing starch, glycerol, coconut oil and tea leaf extract. Ali et al. (2010) observed that uncoated tomatoes turned from green to red within 4–8 days of storage while fruits coated with 50 and 100 g L⁻¹ gum arabic were red at 12–16 days, and those coated with 150 and 200 g L⁻¹ gum arabic remained green, even at 20 days of storage.

The colour parameters of the majority of the coated tomatoes showed similar behaviour to the uncoated fruit, revealing decreased L^* , constant b^* and increased a^* values during storage.

Effect of Edible Coatings on Microbiological Quality of Tomatoes

The microbiological quality of uncoated and coated tomatoes was evaluated during storage at 25 °C for 12 days. The mould and yeast counts were typically maintained at < 10 CFU g⁻¹ for the coated and uncoated tomatoes until day 12 of storage, except for formulations 1, 3 and 9 (Table 4). Overall, all samples remained within the mould and yeast count limit of 1.0×10^6 CFU g⁻¹, considered acceptable for fruit-based products (Bierhals et al. 2011). On the first and last day of storage, the thermotolerant coliforms were present at 3.0 MPN g⁻¹ for uncoated and coated tomatoes. These results comply with the Brazilian legislation (RDC Resolution No. 12; Brasil 2001), where fresh fruit prepared 'in nature' (peeled, selected or fractionated), sanitised, refrigerated or frozen for direct consumption should contain a maximum of 5.0×10^2 MPN g⁻¹.

Few authors have studied the microbiological quality of tomatoes. Among them, Das et al. (2013) affirmed that a film containing starch, glycerol, coconut oil and tea leaf extract acted as an antimicrobial barrier, reducing the number of microorganisms in tomatoes during storage for 20 days. Fagundes et al. (2015) established that edible coatings based on HPMC, beeswax and sodium benzoate did not prevent the appearance of the spores of Alternaria alternate in the artificially inoculated fruit but reduced the incidence and severity of the fungus on cherry tomatoes stored at 5 °C for 21 days, followed by 20 °C for 4 days. Finally, Guerra et al. (2015) demonstrated that coatings comprising CH and M. piperita L. or Mentha \times villosa Huds. delayed the growth of decaycausing A. niger, B. cinerea, Penicillium expansum and R. stolonifer in artificially contaminated tomato fruit during storage at either room temperature or low temperature.

The coatings did not influence the microbiological quality, given that both uncoated and coated tomatoes maintained the mould and yeast counts below the limit recommended by Brazilian legislation, until day 12 of storage.

Edible coatings containing CS (10 g L ⁻¹) Luminosity (L*) Red intensity (a*) Formulations CH EO PPE 1 4 (gL^{-1}) $(mL L^{-1})$ $(mL L^{-1})$ $(mL L^{-1})$ $(mL L^{-1})$ $(mL L^{-1})$ $(mL L^{-1})$ 1 5 0 0 $4g^{8hh} \pm 1$ $43^{6hB} \pm 1$ $43^{6hB} \pm 1$ $23^{6hB} \pm 2$ $26^{hhB} \pm 2$ 2 5 2.5 4q^{9hh} \pm 1 $43^{6hB} \pm 1$ $42^{6hB} \pm 1$ $27^{6h} \pm 2$ $26^{hhB} \pm 2$ 3 5 5 $qq^{9hh} \pm 2$ $4q^{6hB} \pm 1$ $42^{6hB} \pm 1$ $27^{6h} \pm 2$ $26^{hhB} \pm 2$ 4 5 10 2.0 $q^{8hh} \pm 3$ $44^{6hh} \pm 3$ $41^{6hB} \pm 3$ $27^{hB} \pm 2$ $26^{hhB} \pm 2$ 7 10 5 4 $47^{hh} \pm 3$ $44^{hh} \pm 3$ $41^{ehB} \pm 3$ $27^{hB} \pm 3$ $27^{hB} \pm 3$ 8 10 10 2.5 5 $44^{hh} \pm 3$ $47^{hB} \pm 3$ $27^{hB} \pm 3$ $27^{hB} \pm 3$ $27^{hB} \pm 3$ <t< th=""><th>Time of storage (days)</th><th></th><th></th><th></th></t<>	Time of storage (days)			
CH EO PPE 1 4 8 12 1 $(g L^{-1})$ $(mL L^{-1})$		(*)	Yellow intensity (b^*)	
500 $4g^{abh}\pm1$ $4g^{cdB}\pm1$ $4g^{cdB}\pm1$ $2q^{cdB}\pm1$ $20^{Ac}\pm1$ 52.55 $4g^{abh}\pm4$ $4g^{bcAB}\pm1$ $4g^{cdB}\pm3$ $4f^{cdB}\pm3$ $2g^{ab-A}\pm2$ 51020 $4g^{bh}\pm0$ $4g^{bcB}\pm1$ $4g^{cdB}\pm1$ $2g^{ab-A}\pm2$ 10000 $4g^{abh}\pm3$ $4f^{abh}\pm1$ $4g^{cdB}\pm1$ $2q^{ab}\pm2$ 102.55 $4g^{abh}\pm2$ $4q^{abh}\pm4$ $4g^{cdB}\pm1$ $2d^{ab}\pm1$ $2q^{aB}\pm4$ 102.55 $4g^{abh}\pm2$ $4g^{abh}\pm4$ $4g^{bcB}\pm1$ $41^{cdB}\pm1$ $2q^{aB}\pm3$ 102.55 $4g^{abh}\pm2$ $4g^{bcB}\pm1$ $41^{cdB}\pm1$ $2q^{aB}\pm3$ 10102.64 $4g^{bch}\pm2$ $4g^{bcB}\pm1$ $41^{cdB}\pm1$ $2q^{aB}\pm3$ 2000 $4g^{abh}\pm2$ $4g^{bch}\pm3$ $4g^{bcB}\pm1$ $41^{cdB}\pm1$ $2q^{aB}\pm3$ 2010102.6 $4g^{bbh}\pm2$ $4g^{bch}\pm3$ $2g^{aB}\pm3$ $2g^{aB}\pm3$ 20000 $4g^{bbh}\pm2$ $4g^{bch}\pm3$ $2g^{aB}\pm3$ 20102.6 $4g^{bbh}\pm3$ $4g^{bbh}\pm3$ $4g^{bbh}\pm3$ $2g^{aB}\pm3$ 202000 $4g^{bbh}\pm3$ $4g^{bbh}\pm3$ $4g^{cbh}\pm3$ $2g^{aB}\pm3$ 202000 $4g^{bbh}\pm3$ $4g^{bbh}\pm3$ $4g^{bbh}\pm3$ $2g^{aB}\pm4$ 201020 $4g^{bh}\pm3$ $4g^{bh}\pm4$ $4g^{ch}\pm4$ $4g^{ch}\pm4$ $4g^{ah}\pm4$	∞	8 12	1 4	8 12
5 2.5 5 $49^{abh}\pm4$ $45^{bcAB}\pm1$ $4c^{abB}\pm3$ $41^{abB}\pm3$ $25^{aB}\pm2$ 510 25 $45^{abh}\pm2$ $44^{cbAB}\pm1$ $22^{cB}\pm0$ $41^{abC}\pm1$ $24^{ah}\pm2$ 51020 $46^{abh}\pm3$ $44^{abAB}\pm1$ $41^{aB}\pm0$ $41^{abE}\pm1$ $23^{aB}\pm2$ 10000 $48^{abh}\pm3$ $44^{abAB}\pm1$ $41^{aB}\pm1$ $24^{aB}\pm1$ $23^{aB}\pm4$ 102.55 $49^{abh}\pm2$ $44^{abAB}\pm1$ $41^{aB}\pm1$ $41^{aB}\pm1$ $24^{aB}\pm3$ 10102.647^{abh}\pm2 $44^{bcAB}\pm4$ $45^{bcB}\pm1$ $41^{cB}\pm1$ $24^{aB}\pm3$ 10102.047^{abh}\pm2 $44^{bcAB}\pm4$ $45^{cB}\pm1$ $41^{cB}\pm1$ $24^{aB}\pm3$ 200047^{abh}\pm2 $44^{bcAB}\pm1$ $41^{cB}\pm1$ $24^{aB}\pm3$ 2010102.6 $47^{abh}\pm2$ $44^{bcAB}\pm1$ $41^{cB}\pm1$ $24^{aB}\pm2$ 202.55 $51^{abh}\pm2$ $47^{abh}\pm2$ $47^{abh}\pm2$ $24^{aB}\pm2$ $24^{aB}\pm3$ 20000 $47^{abh}\pm3$ $42^{cB}\pm1$ $47^{cB}\pm2$ $24^{aB}\pm2$ 202.65.010 $47^{abh}\pm3$ $47^{cB}\pm2$ $24^{aB}\pm2$ 202.65.010 $47^{abh}\pm3$ $47^{cB}\pm2$ $47^{cB}\pm2$ 202.65.010 $20^{aB}\pm2$ $47^{cB}\pm2$ $47^{cB}\pm2$ $24^{aB}\pm2$ 20102.6 $48^{abh}\pm3$ $42^{cB}\pm2$ $47^{cB}\pm2$ 28	$43^{cdB} \pm 1 43^{cdB} \pm 1 42^{cdB} \pm 1 20^{Ac} \pm 1$	$32^{aA}\pm1$	$34^{abA}\pm 1 31^{abAB}\pm 1$	$30^{bcB} \pm 3$ $28^{bcB} \pm 2$
5510 $45^{b}\Lambda_{\pm}2$ $44^{cd}\Lambda_{B}\pm 1$ $42^{cd}B\pm 0$ $43^{abc}\Lambda_{B}\pm 1$ $24^{ah}\pm 2$ 51020 $46^{ab}\Lambda_{\pm}3$ $48^{bb}\pm 1$ $42^{cd}\pm 0$ $41^{cd}\pm 1$ $23^{aB}\pm 2$ 1000 $48^{ab}\Lambda_{\pm}3$ $44^{ab}\Lambda_{B}\pm 1$ $41^{cd}B\pm 1$ $24^{aB}\pm 1$ 102.55 $49^{ab}\Lambda_{\pm}2$ $44^{bc}\Lambda_{B}\pm 1$ $41^{cd}B\pm 1$ $24^{aB}\pm 3$ 102.510 $47^{ab}\Lambda_{\pm}2$ $44^{bc}\Lambda_{B}\pm 1$ $41^{cd}B\pm 1$ $24^{aB}\pm 3$ 10102.0 $47^{ab}\Lambda_{\pm}2$ $44^{bc}\Lambda_{B}\pm 1$ $41^{cd}B\pm 1$ $24^{aB}\pm 3$ 2000 $49^{ab}\Lambda_{\pm}2$ $44^{bc}\Lambda_{B}\pm 1$ $41^{cd}B\pm 1$ $24^{aB}\pm 3$ 2010102.0 $47^{ab}\Lambda_{\pm}2$ $44^{bc}\Lambda_{B}\pm 1$ $41^{cd}B\pm 1$ $24^{aB}\pm 2$ 20000 $49^{ab}\Lambda_{\pm}2$ $44^{bc}\Lambda_{B}\pm 1$ $41^{cd}B\pm 1$ $24^{aB}\pm 2$ 202.000 $49^{ab}\Lambda_{\pm}2$ $44^{bc}\Lambda_{\pm}2$ $44^{bc}B\pm 3$ $24^{aB}\pm 2$ 202.0102.0 $47^{ab}\Lambda_{\pm}3$ $44^{bc}\Lambda_{B}\pm 1$ $47^{cd}\Lambda_{B}\pm 2$ $24^{aB}\pm 2$ 202.0102.0 $47^{ab}\Lambda_{\pm}3$ $44^{bc}\Lambda_{\pm}2$ $44^{bc}\Lambda_{\pm}3$ $24^{aB}\pm 2$ 202.02.02.02.02.02.02.0 $24^{ab}\Lambda_{\pm}3$ $24^{ab}\Lambda_{\pm}2$ 202.02.02.02.02.02.02.0 $24^{ab}\Lambda_{\pm}3$ $24^{ab}\Lambda_{\pm}4$ $26^{ab}\Lambda_{\pm}4$	$45^{bcAB} \pm 3$ $44^{cdAB} \pm 3$ $41^{cdB} \pm 3$ $25^{aB} \pm 2$	$^{AB}\pm2~26^{deAB}\pm2~29^{abA}\pm1$	$33^{abA}\pm2 31^{abA}\pm3$	$30^{bcA}\pm3 28^{cdA}\pm4$
51020 $46^{abh}\pm0$ $48^{bbh}\pm1$ $42^{cdC}\pm0$ $41^{cdC}\pm1$ $23^{aB}\pm2$ 1000048^{abh}\pm3 $44^{abh}\pm1$ $41^{dB}\pm0$ $41^{cdE}\pm1$ $24^{aB}\pm1$ 102.55 $49^{abh}\pm2$ $44^{bch}\pm1$ $41^{cB}\pm1$ $41^{cdB}\pm1$ $24^{aB}\pm3$ 102.510 $47^{abh}\pm2$ $44^{bch}\pm1$ $43^{cdB}\pm1$ $41^{cdB}\pm1$ $24^{aB}\pm3$ 10102.00047^{abh}\pm2 $44^{bch}\pm1$ $41^{cdB}\pm1$ $24^{aB}\pm3$ 2000047^{abh}\pm2 $44^{bch}\pm1$ $41^{cdB}\pm1$ $41^{cdB}\pm1$ $24^{aB}\pm3$ 2000049^{abh}\pm4 $45^{bch}\pm2$ $45^{abh}\pm3$ $24^{ab}\pm3$ $24^{ab}\pm3$ 2000049^{abh}\pm3 $44^{bch}\pm2$ $44^{bch}\pm1$ $47^{cdB}\pm2$ $29^{aB}\pm4$ 20102.049^{abh}\pm3 $44^{bch}\pm1$ $47^{cdB}\pm2$ $24^{aB}\pm1$ $24^{aB}\pm3$ 20102.049^{abh}\pm3 $44^{bch}\pm1$ $47^{cdB}\pm2$ $29^{aB}\pm4$ 20102.049^{abh}\pm3 $46^{bch}\pm4$ $45^{abh}\pm2$ $29^{aB}\pm4$ 20102.049^{abh}\pm3 $46^{bch}\pm4$ $45^{abh}\pm2$ $29^{aB}\pm4$ 202.555 $69^{abh}\pm3$ $46^{bch}\pm4$ $45^{abh}\pm4$ $27^{aB}\pm3$ 20102.00000 $20^{abh}\pm3$ $46^{bch}\pm4$ $46^{abh}\pm4$ $21^{aB}\pm3$ 202.5 </td <td>$44^{cdAB} \pm 1$ $42^{cdB} \pm 0$ $43^{abcAB} \pm 1$ $24^{aA} \pm 2$</td> <td>$^{A}\pm 1$ $27^{bcA}\pm 2$ $27^{deA}\pm 2$</td> <td>$29^{bA}\pm3 30^{bcA}\pm1$</td> <td>$28^{bcA}\pm 1 \qquad 28^{cdA}\pm 1$</td>	$44^{cdAB} \pm 1$ $42^{cdB} \pm 0$ $43^{abcAB} \pm 1$ $24^{aA} \pm 2$	$^{A}\pm 1$ $27^{bcA}\pm 2$ $27^{deA}\pm 2$	$29^{bA}\pm3 30^{bcA}\pm1$	$28^{bcA}\pm 1 \qquad 28^{cdA}\pm 1$
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301020 $51^{ab} \pm 5$ $47^{bcA} \pm 5$ $46^{abcA} \pm 4$ $46^{abA} \pm 4$ $21^{aB} \pm 3$ Uncoated tomatoes $47^{abA} \pm 3$ $43^{cdAB} \pm 3$ $41^{dB} \pm 2$ $40^{dB} \pm 1$ $25^{aB} \pm 2$	$52^{aA} \pm 7$ 49 $^{aA} \pm 4$ 49 $^{aA} \pm 3$ $25^{aA} \pm 2$	$^{B}\pm 2$ $25^{eA}\pm 2$ $25^{efA}\pm 1$	$36^{aA} \pm 3$ $37^{aA} \pm 4$	$37^{aA} \pm 6$ $37^{aA} \pm 4$
$47^{abA} \pm 3$ $43^{cdAB} \pm 3$ $41^{dB} \pm 2$ $40^{dB} \pm 1$ $25^{aB} \pm 2$	$47^{bcA} \pm 5$ $46^{abcA} \pm 4$ $46^{abA} \pm 4$ $21^{aB} \pm 3$	$^{B}\pm 2$ $26^{deA}\pm 2$ $31^{aA}\pm 2$	$34^{abA}\pm3$ $33^{abA}\pm6$	$32^{abA}\pm 5$ $30^{bcA}\pm 3$
	$43^{cdAB}\pm 3 41^{dB}\pm 2 \qquad 40^{dB}\pm 1 \qquad 25^{aB}\pm 2$	$^{A}\pm 1$ $28^{bcAB}\pm 1$ $31^{aA}\pm 2$	$34^{abA}\pm 5$ $32^{abA}\pm 4$	$29^{bcA}\pm2$ $28^{bcA}\pm1$

Table 4Microorganisms countsin uncoated tomatoes andtomatoes coated with differentedible coatings containingcassava starch, chitosan, *Lippia*sidoides essential oil andpomegranate peel extract duringstorage at 25 °C for 12 days

				Storag	e time (d	ays)			
				(CFU	and yeas g^{-1}) ated value			Thermot coliform (MPN g	s
Edible coatings	s containin	g CS (10 g L	-1)	1	4	8	12	1	12
Formulations	CH (g L ⁻¹)	EO $(mL L^{-1})$	PPE (mL L ⁻¹)						
1	5	0	0	< 10	< 10	< 10	15	< 3.0	< 3.0
2	5	2.5	5	< 10	<10	< 10	< 10	< 3.0	< 3.0
3	5	5	10	< 10	55	15	20	< 3.0	< 3.0
4	5	10	20	< 10	< 10	< 10	< 10	< 3.0	< 3.0
5	10	0	0	< 10	<10	< 10	< 10	< 3.0	< 3.0
6	10	2.5	5	< 10	< 10	< 10	320	< 3.0	< 3.0
7	10	5	10	< 10	<10	< 10	< 10	< 3.0	< 3.0
8	10	10	20	<10	< 10	< 10	< 10	< 3.0	< 3.0
9	20	0	0	<10	<10	26	146	< 3.0	< 3.0
10	20	2.5	5	<10	< 10	< 10	< 10	< 3.0	< 3.0
11	20	5.0	10	<10	< 10	< 10	< 10	< 3.0	< 3.0
12	20	10	20	< 10	< 10	< 10	< 10	< 3.0	< 3.0
13	30	0	0	< 10	<10	< 10	< 10	< 3.0	< 3.0
14	30	2.5	5	< 10	< 10	< 10	< 10	< 3.0	< 3.0
15	30	5	10	< 10	< 10	< 10	< 10	< 3.0	< 3.0
16	30	10	20	< 10	< 10	< 10	< 10	< 3.0	< 3.0
	Uncoated	d tomatoes		< 10	<10	< 10	< 10	< 3.0	< 3.0

CS, cassava starch; CH, chitosan; EO, *Lippia sidoides* essential oil; PPE, pomegranate peel extract. CFU, colony-forming units; MPN, most probable number

Conclusions

In this study, edible coatings based on CS and CH were enriched with L. sidoides EO and PPE, for the first time. The majority of the analysed physical-chemical parameters were not significantly different between the coated and uncoated tomatoes at day 12 of storage. However, the coatings lowered the weight loss and TSS content, demonstrating their effectiveness in delaying fruit ripening. In this context, the tomatoes coated with 10 g L^{-1} CS, 10 g L^{-1} CH, 10 mL L^{-1} EO and 20 mL L^{-1} PPE (formulation 8) showed the best results (lowest weight loss and reduced TSS content). Considering the perishability of Italian tomatoes, the findings proved that these coatings effectively delayed ripening and maintained the physical-chemical quality of the fruit during 12 days of storage at 25 °C. The coatings have a potential to be used in future applications as bioactive packaging. However, further studies are needed to evaluate the sensorial parameters of coated tomatoes and the safety of L. sidoides EO and PPE for human consumption.

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