Stress Signaling in the Heart by AMP-activated Protein Kinase

Raymond Russell III, MD, PhD

Corresponding author

Raymond Russell III, MD, PhD Section of Cardiovascular Medicine, Yale University School of Medicine, 333 Cedar Street, FMP 3, New Haven, CT 06510, USA. E-mail: raymond.russell@yale.edu

Current Hypertension Reports 2006, **8:**446–450 Current Science Inc. ISSN 1522-6417 Copyright © 2006 by Current Science Inc.

The stress-signaling protein, adenosine monophosphate–activated protein kinase (AMPK), regulates a variety of pathways in cells that 1) increase the provision and utilization of energy-providing substrates such as glucose and fatty acids, 2) inhibit energy-requiring pathways such as cholesterol biosynthesis and protein synthesis, and 3) increase the transcription of genes involved in energy metabolism and mitochondrial biogenesis. In the heart, AMPK therefore becomes very important in protecting against ischemia-reperfusion injury and regulating substrate metabolism in the face of changes in workload. This review summarizes the regulation of AMPK activity in the heart and discusses the effects of AMPK activation.

Introduction

Adenosine monophosphate–activated protein kinase (AMPK) is increasingly recognized as a vital signaling molecule in the cellular response to stress. Originally two proteins were described: 3-hydroxy-3-methylglutaryl-CoA (HMG-CoA) reductase kinase, inhibiting cholesterol synthesis, and acetyl-CoA carboxylase (ACC) kinase, stimulating fatty acid metabolism by decreasing the conversion of acetyl-CoA to malonyl-CoA. The structure of the two proteins was found to be identical and it was revealed that this protein, which is sensitive to changes in the energy state of the cell, plays a central role in regulating metabolism [1].

Metabolic and Molecular Regulation of AMPK

AMPK is a serine/threonine kinase that plays a major role in the response of mammalian cells to metabolic stress (Fig. 1). As the name indicates, AMPK activity increases in response to increases in the intracellular content of AMP, which generally occur as a result of hydrolysis of adenosine triphosphate (ATP). As noted above, AMPK inhibits both cholesterol synthesis and protein synthesis [2,3], which are energy-requiring cellular functions. AMPK also activates energy-producing metabolic pathways, including glycolysis [4,5••,6,7] and fatty acid oxidation [8,9]. Therefore, the overall effect of AMPK activation is to shift the balance from ATP consumption to ATP production to compensate for cellular metabolic stress. As a result, AMPK has been referred to as a "metabolic fuel gauge" [10] or "metabolic master switch" [11].

AMPK consists of three subunits, the catalytic α subunit and the noncatalytic β and γ subunits, which regulate the activity of AMPK and its sensitivity to activation by AMP [12]. There are two isoforms of the α subunit, α 1 and $\alpha 2$, which might be responsible for the regulation of different downstream targets [13]. In addition, the $\alpha 2$ subunit is present in the nucleus as well as in the cytosol, suggesting a role for AMPK in gene expression [14]. It has been demonstrated that chronic chemical activation of AMPK with the nucleoside 5-aminoimidazole-4-carboxyamide-1-ribofuranoside (AICAR) increases the expression of key metabolic genes in skeletal muscle [15-19]. In addition, stimulation of AMPK with the AMP analog precursor AICAR in isolated cardiac myocytes and perfused hearts increases the expression of two fatty acid transporters, FABPpm and FAT/CD36 [20]. Interestingly, chronic stimulation of AMPK improves insulin sensitivity in animal models of diabetes and obesity [21,22]. Although little work has been done to demonstrate the effect of chronic activation of AMPK in the heart, mutations in the $\gamma 2$ subunit of AMPK in humans result in increased AMPK activity and a cardiomyopathy characterized by intracellular glycogen accumulation and pre-excitation syndrome [23,24].

Regulation of AMPK occurs through both covalent and allosteric mechanisms. With respect to the covalent regulation of AMPK, phosphorylation of the catalytic α subunit at threonine 172 (Thr172) results in increased activity. This phosphorylation occurs through the activity of upstream kinases, known collectively as AMPK kinases (AMPKKs). The family of AMPKKs includes the tumor suppressor LKB1 [25••,26] and calmodulin-dependent



Figure 1. The adenosine monophosphate– activated protein kinase (AMPK) signaling pathway and the downstream metabolic effects of AMPK activation. ACC—acetyl-CoA carboxylase; ATP—adenosine triphosphate; FABP—fatty acid binding protein; GLUT—facilitative glucose transporter; PFK—phosphofructokinase; UCP—uncoupling protein.

protein kinase kinase- β [27,28], although there may be other members of this family of kinases that are responsible for Thr172 phosphorylation and ultimately AMPK activation. This is supported by recent findings that loss of LKB1 function in the heart prevents the Thr172 phosphorylation of the α 2 subunit but does not affect phosphorylation and activation of the α 1 subunit [29].

5'-Adenosine monophosphate regulates AMPK through noncovalent binding with the γ subunit. This noncovalent interaction has several effects that increase AMPK activity. First, the AMP- γ subunit complex interacts with an autoinhibitory region in the catalytic α subunit, increasing the interaction of the α subunit with target proteins [12]. Second, the binding of AMP to the γ subunit makes AMPK less susceptible to the actions of protein phosphatases, which can desphosphorylate Thr172 of the α subunit [30].

As expected, stimuli that cause changes in the energy charge of the cell will increase the activity of AMPK. These include ischemia [5.,8], increased contractile work [6,31,32], oxidant stress [33,34], and osmotic stress [35]. In addition, the chemical agent AICAR stimulates AMPK activity by being metabolized to an AMP analog, ZMP, albeit at concentrations in the millimolar range, in which there may be nonspecific effects [36,37]. However, AMPK can be activated by other stimuli that do not affect the AMP/ATP ratio. Specifically, the hypoglycemic agents rosiglitazone and metformin can cause Thr172 phosphorylation and AMPK activation [38,39], although at concentrations well above the serum concentrations that are observed with clinical use of the drugs. Nitric oxide (NO) also can stimulate AMPK activation [40], which is interesting given the fact that AMPK can phosphorylate and activate endothelial NO synthase (eNOS) [41], and NO production has been implicated in the stimulation of glucose uptake by AMPK [42•]. Furthermore, the adipokines leptin [43,44] and adiponectin [45,46] can stimulate AMPK activity, although some of these effects may be tissue-specific [47].

Role of AMPK in the Cardiovascular System The first (and still the most studied) stimulus for AMPK activation in the heart is myocardial ischemia. Myocardial ischemia results in a switch in the main source of energy production from fatty acid oxidation to glycolysis. During reperfusion, with restored provision of oxygen to the heart, the rate of fatty acid oxidation increases over the rate observed under basal conditions. These metabolic switches during ischemia and reperfusion can be explained by activation of AMPK. AMPK is activated very rapidly with the onset of ischemia [48••]. This increase in AMPK activity is maintained during ischemia and for at least 30 minutes during reperfusion $[5 \bullet \bullet]$. Early studies using AICAR demonstrated that AMPK activation causes translocation of the facilitative glucose transporter GLUT4 from an intracellular storage pool to the cell surface, where it is biologically active, increasing glucose uptake [4]. Subsequent studies using transgenic mice in which AMPK was rendered inactive demonstrated that there is an absolute requirement for AMPK activation for increased glucose uptake in the setting of myocardial ischemia [5••,49]. Furthermore, AMPK activation enhances glycolysis by activating phosphofructokinase (PFK)-2, which generates fructose 2,6-bisphosphate, an activator of the glycolytic enzyme PFK-1 [7].

As noted above, AMPK phosphorylates and thereby inactivates ACC, which results in increased fatty acid oxidation. The phosphorylation of ACC decreases its ability to convert acetyl-CoA to malonyl-CoA, which is an allosteric inhibitor of carnitine palmitoyltransferase-1 (CPT-1). CPT-1 regulates the transport of fatty acids into the mitochondria and is therefore the rate-limiting enzyme of fatty acid β -oxidation by the mitochondria. Although AMPK activation during ischemia will result in the phosphorylation of ACC, the oxidation of fatty acids does not increase; instead, it **decreases** because of the lack of sufficient oxygen to maintain β -oxidation. During reperfusion, however, when adequate blood flow and oxygenation are reestablished, the inhibition of ACC results in enhanced fatty acid oxidation $[5 \bullet , 8, 9]$.

The vital role that AMPK plays in the ischemic heart is underscored by studies assessing the effect of loss of AMPK function on the response of the heart to ischemia. In the setting of no-flow ischemia, loss of AMPK function results in a more rapid onset of ischemic contracture [49]. With low-flow ischemia followed by reperfusion, loss of AMPK function results in poorer postischemic recovery of function [5••]. Furthermore, this contractile dysfunction is associated with greater myocyte damage and increased apoptosis. It remains to be determined if this antiapoptotic effect of AMPK is related to the metabolic effects of this stress protein or if there is some direct effect on the apoptotic pathways that is mediated by AMPK.

Because of the rapid hydrolysis of ATP to adenosine diphosphate (ADP) and AMP in the setting of myocardial ischemia [50], rapid activation of AMPK is to be expected, and detectable changes in AMPK activity have been demonstrated within 1 minute of the onset of ischemia [48••]. This activation is mirrored by an increase in AMPKK activity, as determined by Thr172 phosphorylation of AMPK. Interestingly, in contrast to AMPK, which demonstrates clear allosteric activation by AMP, AMPKK activity is not affected by AMP concentrations [48••]. Furthermore, in vitro activity of one of the putative AMP-KKs, LKB1, is not increased by myocardial ischemia, suggesting either that LKB1 does not play a significant role in the regulation of AMPK in the heart [51] or that other mechanisms, such as the association of LKB1 with the accessory subunits MO25 and STRAD, are responsible for increased AMPKK activity [25••].

The effects of AMPK on fatty acid metabolism are rather direct, with phosphorylation and inactivation of ACC being responsible for increased fatty acid oxidation, but the downstream mechanisms responsible for GLUT4 translocation are only beginning to be characterized. Recent studies have demonstrated that inhibition of p38 mitogen-activated protein kinase (MAPK) results in partial inhibition of the increase in glucose uptake caused by either hypoxia or AICAR stimulation and decreased translocation of GLUT4 to the cell surface [52•]. As mentioned above, AMPK activation results in phosphorylation of eNOS. However, treatment of heart muscle with NO inhibitors incompletely attenuates the increase in glucose uptake in response to AICAR stimulation [42•]. It is not clear whether the incomplete nature of the attenuation of the effects of AMPK stimulation on glucose metabolism by NO inhibitors is due to incomplete inactivation of p38 MAPK or NO production or whether p38 MAPK and eNOS are only some of the parallel mediators of the AMPK signaling cascade.

The role of AMPK in the heart's response to ischemia discussed above represents an acute response with little effect on gene transcription and protein expression. However, several studies have investigated the effects of chronic stimuli on AMPK activity in the heart. Chronic pressure overload, induced by ascending aortic banding, increases both α 1- and α 2-isoform–specific AMPK activity, although the expression of the $\alpha 1$ isoform increases while expression of the $\alpha 2$ isoform decreases, suggesting that there may be greater Thr172 phosphorylation of the $\alpha 2$ isoform to explain the increased $\alpha 2$ activity [53]. Interestingly, volume overload inhibits the developmental increases in AMPK expression [54]. Metabolic signals, such as prolonged exposure of isolated cardiac myocytes to the saturated fatty acid palmitate, cause a decrease in AMPK expression and increase apoptosis [55]. Furthermore, perfusion of isolated hearts with high concentrations of free fatty acids has been shown to increase Thr172 phosphorylation of the α subunits of AMPK, as does 24 hours of fasting [56]. Chronic caloric restriction did not change myocardial AMPK activity in another study, however [57]. Furthermore, in insulin-resistant rats with increased circulating free fatty acid concentrations, no change in myocardial AMPK activity was noted [58]. The conflicting results of the above studies demonstrate the need for further research in the area of chronic regulation of AMPK.

Although most of this review has focused on the role of AMPK in cardiac myocytes, it is important to point out that AMPK also plays a critical role in endothelial cells. Endothelial cells derive the bulk of their ATP from glycolysis, although glucose and fatty acid oxidation can contribute variable amounts to ATP production [59]. As mentioned above, AMPK activation leads to eNOS phosphorylation, which may be responsible, in part, for the increase in glucose utilization by endothelial cells [40,41]. Endothelialcell AMPK activation by AICAR can increase the rates of oxidation of both glucose and fatty acids [59,60]. The significance of endothelial-cell AMPK activation by metabolic stress in vivo remains to be determined, although AMPK activation can decrease reactive oxygen species generation and apoptosis induced by hyperglycemia in endothelial cells [61,62]. Furthermore, loss of AMPK activity inhibits the expression of vascular endothelial growth factor and angiogenesis in response to hypoxic stress [63,64].

Conclusions

Recognition of the importance of AMPK in the response to metabolic and hemodynamic stressors in the cardiovascular system continues to increase, with AMPK found to be responsible for metabolic adaptations to ischemia, changes in workload, and alterations in circulating substrate concentrations, as well as for genomic-level adaptations with changes in expression of key proteins and protection against cell injury and death. Based on the growing body of information concerning the role of AMPK in the heart and vasculature, increased interest will be focused on pharmacologic manipulation of the AMPK signaling pathway. There may also be interest in possible methods of noninvasively assessing the activity of this pathway in disease states.

Acknowledgment

The author's laboratory is supported by grants (K08 HL04438 and R01 HL077310) from the National Heart, Lung, and Blood Institute of the US Public Health Service.

References and Recommended Reading

Papers of particular interest, published recently, have been highlighted as:

- Of importance
- •• Of major importance
- 1. Carling D, Clarke PR, Zammit VA, Hardie DG: Purification and characterization of the AMP-activated protein kinase. Copurification of acetyl-CoA carboxylase kinase and 3-hydroxy-3-methylglutaryl-CoA reductase kinase activities. Eur J Biochem 1989, 186:129–136.
- 2. Bolster DR, Crozier SJ, Kimball SR, Jefferson LS: AMPactivated protein kinase suppresses protein synthesis in rat skeletal muscle through down-regulated mammalian target of rapamycin (mTOR) signaling. J Biol Chem 2002, 277:23977–23980.
- 3. Horman S, Beauloye C, Vertommen D, et al.: Myocardial ischemia and increased heart work modulate the phosphorylation state of eukaryotic elongation factor-2. *J Biol Chem* 2003, 278:41970–41976.
- 4. Russell RR III, Bergeron R, Shulman GI, Young LH: Translocation of myocardial GLUT-4 and increased glucose uptake through activation of AMPK by AICAR. *Am J Physiol* 1999, 277:H643–H649.
- 5.•• Russell RR III, Li J, Coven DL, et al.: AMP-activated protein kinase mediates ischemic glucose uptake and prevents postischemic cardiac dysfunction, apoptosis, and injury. J Clin Invest 2004, 114:465-468.

In this study, transgenic mice expressing a dominant negative form of AMPK were used to demonstrate that loss of AMPK function blocks the ischemia-mediated increase in glycolysis in the heart. The inability to increase glucose uptake in response to ischemia resulted in greater myocyte damage and cell death and greater postischemic contractile dysfunction.

- Hayashi T, Hirshman MF, Kurth EJ, et al.: Evidence for 5' AMP-activated protein kinase mediation of the effect of muscle contraction on glucose transport. *Diabetes* 1998, 47:1369–1373.
- 7. Marsin AS, Bertrand L, Rider MH, et al.: Phosphorylation and activation of heart PFK-2 by AMPK has a role in the stimulation of glycolysis during ischaemia. *Curr Biol* 2000, 10:1247–1255.
- 8. Kudo N, Barr AJ, Barr RL, et al.: High rates of fatty acid oxidation during reperfusion of ischemic hearts are associated with a decrease in malonyl-CoA levels due to an increase in 5'-AMP-activated protein kinase inhibition of acetyl-CoA carboxylase. J Biol Chem 1995, 270:17513–17520.
- Kudo N, Gillespie JG, Kung L, et al.: Characterization of 5'AMP-activated protein kinase activity in the heart and its role in inhibiting acetyl-CoA carboxylase during reperfusion following ischemia. *Biochim Biophys Acta* 1996, 1301:67–75.
- 10. Hardie DG, Carling D: The AMP-activated protein kinase—fuel gauge of the mammalian cell? *Eur J Biochem* 1997, 246:259–273.
- 11. Winder WW, Hardie DG: AMP-activated protein kinase, a metabolic master switch: possible roles in type 2 diabetes. *Am J Physiol* 1999, 277:E1–E10.
- 12. Cheung PC, Salt IP, Davies SP, et al.: Characterization of AMP-activated protein kinase gamma-subunit isoforms and their role in AMP binding. *Biochem J* 2000, 346:659–669.
- 13. Woods A, Salt I, Scott J, et al.: The alpha1 and alpha2 isoforms of the AMP-activated protein kinase have similar activities in rat liver but exhibit differences in substrate specificity in vitro. *FEBS Lett* 1996, **397**:347–351.

- 14. Salt I, Celler JW, Hawley SA, et al.: AMP-activated protein kinase: greater AMP dependence, and preferential nuclear localization, of complexes containing the alpha2 isoform. *Biochem J* 1998, 334:177–187.
- 15. Holmes BF, Kurth-Kraczek EJ, Winder WW: Chronic activation of 5'-AMP-activated protein kinase increases GLUT-4, hexokinase, and glycogen in muscle. *J Appl Physiol* 1999, 87:1990–1995.
- 16. Winder WW, Holmes BF, Rubink DS, et al.: Activation of AMP-activated protein kinase increases mitochondrial enzymes in skeletal muscle. J Appl Physiol 2000, 88:2219–2226.
- Bergeron R, Ren JM, Cadman KS, et al.: Chronic activation of AMP kinase results in NRF-1 activation and mitochondrial biogenesis. Am J Physiol Endocrinol Metab 2001, 281:E1340-E1346.
- 18. Putman CT, Kiricsi M, Pearcey J, et al.: AMPK activation increases uncoupling protein-3 expression and mitochondrial enzyme activities in rat muscle without fibre type transitions. *J Physiol* 2003, 551:169–178.
- 19. Bamford JA, Lopaschuk GD, MacLean IM, et al.: Effects of chronic AICAR administration on the metabolic and contractile phenotypes of rat slow- and fast-twitch skeletal muscles. *Can J Physiol Pharmacol* 2003, 81:1072–1082.
- 20. Chabowski A, Momken I, Coort SL, et al.: Prolonged AMPK activation increases the expression of fatty acid transporters in cardiac myocytes and perfused hearts. *Mol Cell Biochem* 2006, 288:210–212.
- 21. Buhl ES, Jessen N, Pold R, et al.: Long-term AICAR administration reduces metabolic disturbances and lowers blood pressure in rats displaying features of the insulin resistance syndrome. *Diabetes* 2002, 51:2199–2206.
- 22. Halseth AE, Ensor NJ, White TA, et al.: Acute and chronic treatment of ob/ob and db/db mice with AICAR decreases blood glucose concentrations. *Biochem Biophys Res Commun* 2002, **294**:798–805.
- Zou L, Shen M, Arad M, et al.: N488I mutation of the gamma2-subunit results in bidirectional changes in AMPactivated protein kinase activity. *Circ Res* 2005, 97:323–328.
- 24. Ahmad F, Arad M, Musi N, et al.: Increased alpha2 subunit-associated AMPK activity and PRKAG2 cardiomyopathy. *Circulation* 2005, **112**:3140–3148.
- 25.•• Hawley S, Boudeau J, Reid J, et al.: Complexes between the LKB1 tumor suppressor, STRAD alpha/beta and MO25 alpha/beta are upstream kinases in the AMP-activated protein kinase cascade. J Biol 2003, 2:28.

This study identified the first of several upstream kinases of AMPK. These kinases, collectively known as AMPK kinases, are responsible for the phosphorylation and activation of AMPK.

- 26. Shaw RJ, Kosmatka M, Bardeesy N, et al.: The tumor suppressor LKB1 kinase directly activates AMP-activated kinase and regulates apoptosis in response to energy stress. *Proc Natl Acad Sci U S A* 2004, 101:3329–3335.
- 27. Woods A, Dickerson K, Heath R, et al.: Ca2+/calmodulindependent protein kinase kinase-beta acts upstream of AMP-activated protein kinase in mammalian cells. Cell Metab 2005, 2:21–33.
- Hawley SA, Pan DA, Mustard KJ, et al.: Calmodulin-dependent protein kinase kinase-beta is an alternative upstream kinase for AMP-activated protein kinase. Cell Metab 2005, 2:9–19.
- 29. Sakamoto K, Zarrinpashneh E, Budas GR, et al.: Deficiency of LKB1 in heart prevents ischemia-mediated activation of AMPKalpha2 but not AMPKalpha1. Am J Physiol Endocrinol Metab 2006, 290:E780-E788.
- 30. Davies SP, Helps NR, Cohen PT, Hardie DG: 5'-AMP inhibits dephosphorylation, as well as promoting phosphorylation, of the AMP-activated protein kinase. Studies using bacterially expressed human protein phosphatase-2C alpha and native bovine protein phosphatase-2AC. FEBS Lett 1995, 377:421-425.
- Coven DL, Hu X, Cong L, et al.: Physiological role of AMP-activated protein kinase in the heart: graded activation during exercise. Am J Physiol Endocrinol Metab 2003, 285:E629–E636.

- 32. Wojtaszewski JF, Nielsen P, Hansen BF, et al.: Isoformspecific and exercise intensity-dependent activation of 5'-AMP-activated protein kinase in human skeletal muscle. J Physiol 2000, 528:221–226.
- Choi SL, Kim SJ, Lee KT, et al.: The regulation of AMPactivated protein kinase by H₂O₂. Biochem Biophys Res Commun 2001, 287:92–97.
- 34. Leon H, Atkinson LL, Sawicka J, et al.: Pyruvate prevents cardiac dysfunction and AMP-activated protein kinase activation by hydrogen peroxide in isolated rat hearts. *Can J Physiol Pharmacol* 2004, **82**:409–416.
- Barnes K, Ingram JC, Porras OH, et al.: Activation of GLUT1 by metabolic and osmotic stress: potential involvement of AMP-activated protein kinase (AMPK). J Cell Sci 2002, 115(Pt 11):2433-2442.
- Corton JM, Gillespie JG, Hawley SA, Hardie DG: 5-aminoimidazole-4-carboxamide ribonucleoside. A specific method for activating AMP-activated protein kinase in intact cells? *Eur J Biochem* 1995, 229:558–565.
- 37. Zhang L, Frederich M, He H, Balschi JA: Relationship between 5-aminoimidazole-4-carboxamide-ribotide and AMP-activated protein kinase activity in the perfused mouse heart. Am J Physiol Heart Circ Physiol 2006, 290: H1235-H1243.
- Zhou G, Myers R, Li Y, et al.: Role of AMP-activated protein kinase in mechanism of metformin action. J Clin Invest 2001, 108:1167–1174.
- 39. Fryer LG, Parbu-Patel A, Carling D: The anti-diabetic drugs rosiglitazone and metformin stimulate AMP-acti-vated protein kinase through distinct signaling pathways. J Biol Chem 2002, 277:25226-25232.
- Fryer LG, Hajduch E, Rencurel F, et al.: Activation of glucose transport by AMP-activated protein kinase via stimulation of nitric oxide synthase. *Diabetes* 2000, 49:1978–1985.
- 41. Chen ZP, Mitchelhill KI, Michell BJ, et al.: AMP-activated protein kinase phosphorylation of endothelial NO synthase. *FEBS Lett* 1999, 443:285–289.
- 42.• Li J, Hu X, Selvakumar P, et al.: Role of the nitric oxide pathway in AMPK-mediated glucose uptake and GLUT4 translocation in heart muscle. Am J Physiol Endocrinol Metab 2004, 287:E834-841.

This study identified nitric oxide synthesis by eNOS as one of the downstream signals of AMPK-mediated increases in glucose uptake.

- 43. Minokoshi Y, Kim YB, Peroni OD, et al.: Leptin stimulates fatty-acid oxidation by activating AMP-activated protein kinase. *Nature* 2002, **415**:339–343.
- 44. Steinberg GR, Rush JW, Dyck DJ: AMPK expression and phosphorylation are increased in rodent muscle after chronic leptin treatment. Am J Physiol Endocrinol Metab 2003, 284:E648–E654.
- 45. Yamauchi T, Kamon J, Minokoshi Y, et al.: Adiponectin stimulates glucose utilization and fatty-acid oxidation by activating AMP-activated protein kinase. *Nat Med* 2002, 8:1288–1295.
- Shibata R, Sato K, Pimentel DR, et al.: Adiponectin protects against myocardial ischemia-reperfusion injury through AMPK- and COX-2-dependent mechanisms. *Nat Med* 2005, 11:1096-1103.
- 47. Atkinson LL, Fischer MA, Lopaschuk GD: Leptin activates cardiac fatty acid oxidation independent of changes in the AMP-activated protein kinase-acetyl-CoA carboxylasemalonyl-CoA axis. J Biol Chem 2002, 277:29424–29430.
- 48.•• Baron SJ, Li J, Russell RR III, et al.: Dual mechanisms regulating AMPK kinase action in the ischemic heart. *Circ Res* 2005, **96:**337–345.

This study demonstrated the rapid time course of activation of AMPK and its upstream kinase, AMPKK, in response to myocardial ischemia. The study also demonstrated that, in contrast to AMPK, the activity of AMPKK is not affected by AMP concentrations.

- 49. Xing Y, Musi N, Fujii N, et al.: Glucose metabolism and energy homeostasis in mouse hearts overexpressing dominant negative alpha2 subunit of AMP-activated protein kinase. J Biol Chem 2003, 278:28372–28377.
- Williamson JR: Glycolytic control mechanisms. II. Kinetics of intermediate changes during the aerobicanoxic transition in perfused rat heart. J Biol Chem 1966, 241:5026-5036.
- 51. Altarejos JY, Taniguchi M, Clanachan AS, Lopaschuk GD: Myocardial ischemia differentially regulates LKB1 and an alternate 5'-AMP-activated protein kinase kinase. J Biol Chem 2005, 280:183–190.
- 52.• Li J, Miller EJ, Ninomiya-Tsuji J, et al.: AMP-activated protein kinase activates p38 mitogen-activated protein kinase by increasing recruitment of p38 MAPK to TAB1 in the ischemic heart. *Circ Res* 2005, 97:872–879.

This study demonstrated that, in addition to increased NO production, AMPK activation increases p38 MAPK activity as part of the pathway that enhances glucose uptake.

- 53. Tian R, Musi N, D'Agostino J, et al.: Increased adenosine monophosphate-activated protein kinase activity in rat hearts with pressure-overload hypertrophy. *Circulation* 2001, 104:1664-1669.
- 54. Kantor PF, Robertson MA, Coe JY, Lopaschuk GD: Volume overload hypertrophy of the newborn heart slows the maturation of enzymes involved in the regulation of fatty acid metabolism. J Am Coll Cardiol 1999, 33:1724–1734.
- Hickson-Bick DL, Buja ML, McMillin JB: Palmitatemediated alterations in the fatty acid metabolism of rat neonatal cardiac myocytes. J Mol Cell Cardiol 2000, 32:511-519.
- Clark H, Carling D, Saggerson D: Covalent activation of heart AMP-activated protein kinase in response to physiological concentrations of long-chain fatty acids. *Eur J Biochem* 2004, 271:2215–2224.
- 57. Gonzalez AA, Kumar R, Mulligan JD, et al.: Metabolic adaptations to fasting and chronic caloric restriction in heart, muscle, and liver do not include changes in AMPK activity. Am J Physiol Endocrinol Metab 2004, 287: E1032–E1037.
- Atkinson LL, Kozak R, Kelly SE, et al.: Potential mechanisms and consequences of cardiac triacylglycerol accumulation in insulin-resistant rats. Am J Physiol Endocrinol Metab 2003, 284:E923–E930.
- Dagher Z, Ruderman N, Tornheim K, Ido Y: Acute regulation of fatty acid oxidation and AMP-activated protein kinase in human umbilical vein endothelial cells. Circ Res 2001, 88:1276-1282.
- 60. Dagher Z, Ruderman N, Tornheim K, Ido Y: The effect of AMP-activated protein kinase and its activator AICAR on the metabolism of human umbilical vein endothelial cells. Biochem Biophys Res Commun 1999, 265:112–115.
- 61. Ido Y, Carling D, Ruderman N: Hyperglycemia-induced apoptosis in human umbilical vein endothelial cells: inhibition by the AMP-activated protein kinase activation. *Diabetes* 2002, 51:159–167.
- 62. Kukidome D, Nishikawa T, Sonoda K, et al.: Activation of AMP-activated protein kinase reduces hyperglycemiainduced mitochondrial reactive oxygen species production and promotes mitochondrial biogenesis in human umbilical vein endothelial cells. *Diabetes* 2006, 55:120–127.
- 63. Nagata D, Mogi M, Walsh K: AMP-activated protein kinase (AMPK) signaling in endothelial cells is essential for angiogenesis in response to hypoxic stress. J Biol Chem 2003, 278:31000–31006.
- 64. Ouchi N, Shibata R, Walsh K: AMP-activated protein kinase signaling stimulates VEGF expression and angiogenesis in skeletal muscle. *Circ Res* 2005, 96:838–846.