



Impaired Exercise Tolerance in Heart Failure: Role of Skeletal Muscle Morphology and Function

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Abstract

Purpose of Review To discuss the impact of deleterious changes in skeletal muscle morphology and function on exercise intolerance in patients with heart failure with reduced ejection fraction (HFrEF) and heart failure with preserved ejection fraction (HFpEF), as well as the utility of exercise training and the potential of novel treatment strategies to preserve or improve skeletal muscle morphology and function.

Recent Findings Both HFrEF and HFpEF patients exhibit a reduction in percent of type I (oxidative) muscle fibers and oxidative enzymes coupled with abnormal mitochondrial respiration. These skeletal muscle abnormalities contribute to impaired oxidative metabolism with an earlier shift towards glycolytic metabolism during exercise that is strongly associated with exercise intolerance. In both HFrEF and HFpEF patients, peripheral “non-cardiac” factors are important determinants of the improvement in exercise tolerance following aerobic exercise training. Adjunctive strategies that include nutritional supplementation with amino acids and/or anabolic drugs to stimulate anabolic molecular pathways in skeletal muscle show great promise for improving exercise tolerance and treating heart failure-associated sarcopenia, but these efforts remain early in their evolution, with no immediate clinical applications.

Summary There is consistent evidence that heart failure is associated with multiple skeletal muscle abnormalities which impair oxygen uptake and utilization and contribute greatly to exercise intolerance. Exercise training induces favorable adaptations in skeletal muscle morphology and function that contribute to improvements in exercise tolerance in patients with HFrEF. The contribution of skeletal muscle adaptations to improved exercise tolerance following exercise training in HFpEF remains unknown and warrants further investigation.

Keywords Cardiorespiratory fitness · Exercise training · Oxidative metabolism · Mitochondrial function · Magnetic resonance spectroscopy · Amino acids

Abbreviations

a-vO₂diff Arterial-venous oxygen content difference
CO Cardiac output
HF Heart failure

HFpEF Heart failure with preserved ejection fraction
HFrEF Heart failure with reduced ejection fraction
NYHA New York Heart Association
PCr Phosphocreatine
VO_{2peak} Cardiorespiratory fitness

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Introduction

Heart failure (HF) is a major healthcare problem associated with high morbidity and mortality [1]. Approximately 50% of HF patients have reduced ejection fraction (HFrEF) while the remainder of patients have preserved ejection fraction (HFpEF) [2•]. While both HFrEF and HFpEF increase with age, incidence of HFpEF is particularly prominent, doubling

in incidence with each decade after age 65 [3]. The cardinal symptom in clinically stable HFrEF and HFpEF patients is reduced exercise tolerance [4, 5, 6••, 7, 8]. Heart failure patients' peak cardiorespiratory fitness (VO_{2peak}) is ~65% of age-matched healthy controls [2•, 6••, 7, 9••, 10]. Moreover, declines in cardiorespiratory fitness in older HF patients are compounded by comorbidity, sarcopenia, malnutrition, and other aging sequelae that exacerbate declines in cardiorespiratory fitness as well as strength and balance, and progressively jeopardize quality of life and functional independence [2•, 11].

The reduced VO_{2peak} is secondary to impaired cardiac and pulmonary performance, as well as peripheral vascular, and skeletal muscle abnormalities that result in reduced convective and diffusive O_2 transport coupled with decreased O_2 utilization by exercising muscle [2•, 6••, 12, 13]. A number of invasive hemodynamic studies have shown that both HFrEF and HFpEF have reduced maximal cardiac output (CO) secondary to a lower heart rate and stroke volume response [7, 9••, 10, 14]. However, non-cardiopulmonary peripheral factors also contribute to the lower VO_{2peak} [6••, 7, 9••, 10, 15, 16••].

The aim of this brief review is to discuss the impact that abnormal skeletal muscle morphology and function play in limiting exercise tolerance in HF patients, and the role of both exercise training and novel treatment strategies to improve skeletal muscle morphology and function.

Role of Skeletal Muscle Abnormalities in HFrEF

It has been long known that HFrEF patients have multiple histological and metabolic skeletal muscle abnormalities including skeletal muscle atrophy [17–21], decreased oxidative (type I) fibers and enzymes [22–26], mitochondrial volume density [22], and capillary-fiber ratio [23, 26] (Table 1). Prior studies demonstrate that skeletal muscle atrophy and reduced lower extremity muscle mass contribute to decreased VO_{2peak} and muscle strength in HFrEF [18, 20, 21, 27, 28]. Moreover, a reduction in the percent of type I fibers and oxidative enzymes coupled with abnormal mitochondrial respiration contributes to impaired oxidative metabolism with an earlier shift towards glycolytic metabolism resulting in decreased aerobic endurance (Table 1).

Weiss et al. [16••], using ^{31}P skeletal muscle spectroscopy, examined skeletal muscle energetics (PCr depletion and inorganic phosphate accumulation rates) at rest and during graded (plantar flexion) exercise test in healthy subjects as well as those with HFrEF and HFpEF. A novel finding was that both NYHA class II and III HF patients had significantly faster rates of exercise-induced PCr depletion compared with healthy individuals and NYHA class I HFrEF patients. Finally, the rate of PCr decline during plantar flexion exercise was correlated ($r^2 = 0.83$) with overall exercise time indicating that a rapid exercise-induced depletion of PCr in symptomatic

HFrEF and HFpEF patients is closely related to exercise intolerance.

Role of Skeletal Muscle Abnormalities in HFpEF

Emerging evidence demonstrates that peripheral “non-cardiopulmonary” factors are important determinants of reduced VO_{2peak} in HFpEF [6••, 9••], mirroring many of the concepts previously only associated with HFrEF. Haykowsky et al. [6••] reported that the strongest independent predictor of VO_{2peak} was the change in $a-vO_2diff$ from rest to peak exercise in elderly HFpEF patients. The mechanisms responsible for this impaired ability to augment $a-vO_2diff$ during peak exercise were not studied; however, it was hypothesized that it may relate to intrinsic skeletal muscle abnormalities that underlie reduced skeletal muscle oxygen delivery and/or impaired oxygen utilization.

Given that the majority of oxygen consumed during exercise occurs in the exercising muscle [2•, 10, 42], a decline in metabolically active tissue may limit exercise tolerance. Using dual-energy X-ray absorptiometry and maximal exercise testing, Haykowsky et al. [30] measured lean body mass and VO_{2peak} in older HFpEF patients and age-matched healthy controls. Older HFpEF patients had significantly reduced percent total and leg lean mass, and decreased peak VO_2 indexed to lean body mass versus healthy controls [30] (Table 1). Also, the change in VO_{2peak} with increasing percent leg lean mass was blunted in HFpEF compared to healthy controls [30].

These investigators also reported significantly increased intermuscular adipose tissue and ratio of intermuscular adipose to skeletal muscle area in HFpEF patients [43, 44] (Table 1). Both intermuscular adipose area and intermuscular adipose to skeletal muscle area were independent predictors of VO_{2peak} in HFpEF [43], suggesting it is not only the loss of lean body mass, but the quality of muscle that determines VO_{2peak} . Notably, skeletal muscle atrophy and increased intermuscular adipose tissue detected in HFpEF is similar to skeletal muscle changes that occur as part of aging physiology [45]. This raises important considerations regarding the overlap of HFpEF and aging physiology.

Additional histological and metabolic skeletal muscle abnormalities [31••, 33••, 35] (Table 1) have also been demonstrated in HFpEF patients. Kitzman et al. [31••] showed a shift in skeletal muscle fiber type distribution towards a higher percentage of glycolytic (type II) fibers, with a subsequent reduction in percent type I (oxidative) fibers, type I/type II fiber ratio, and capillary-to-fiber ratio compared to age-matched healthy controls. Molina et al. [33••] extended those findings by demonstrating that skeletal muscle oxidative capacity, mitochondrial content, and mitochondrial fusion were abnormal in older patients with HFpEF, and that they were associated with reduced VO_{2peak} and 6-min walk distance. Cumulatively, these findings suggest that a fiber type shift

Table 1 Summary of skeletal muscle abnormalities that contribute to exercise intolerance in patients with heart failure with reduced or preserved ejection fraction

Variable	HFrEF vs control	HFpEF vs control
Morphology		
Percent lean body mass	↓ [17–20, 27, 28] ↔ [29]	↓ [30]
% type I fibers	↓ [22–24, 26]	↓ [31••]
% type II fibers	↑ [22–24, 26]	↑ [31••]
Capillary density	↓ [22, 23, 25, 26, 32] ↔ [24]	↓ [31••]
Mitochondrial volume density	↓ [22–26]	↓ [33••]
Mitochondrial enzyme density	↓ [22]	↓ [33••]
Function		
Peak exercise a-vO ₂ diff	↔ [7, 8, 9••, 10] ↑ [34]	↓ [6••, 9••, 35] ↔ [14, 36]
Sub-maximal exercise oxidative metabolism	↓ [15, 16••, 21, 37–41]	↓ [16••, 35]

↑ increase, ↓ decrease, ↔ stays the same, a-vO₂diff arterial-venous oxygen content difference, HFpEF heart failure with preserved ejection fraction, HFrEF heart failure with reduced ejection fraction

from oxidative to glycolytic fibers coupled with abnormal mitochondrial function also contributes to impaired oxidative metabolism during exercise in HFpEF. Consistently, prior studies assessing skeletal muscle metabolism during small muscle mass exercise with ³¹P magnetic resonance spectroscopy showed a marked reduction in leg muscle oxidative metabolism in HFpEF patients compared to healthy individuals [16••, 35] (Table 1). Overall, impaired mitochondrial oxidative metabolism appears to be an important contributor to reduced exercise tolerance in HFpEF.

Exercise Interventions Improve Exercise Tolerance and Skeletal Muscle Function in HFrEF

Aerobic exercise training has been shown to increase VO_{2peak} by 2.6–5.4 ml/kg/min [46, 47] compared to usual care in patients with HFrEF. Whereas many cardiovascular experts assumed this was determined by cardiac performance, in fact much of this performance improvement is mediated by favorable adaptations in skeletal muscle morphology and function [42, 48–52] (Table 2). Hambrecht et al. [48–50] found that 6 months of aerobic exercise training (walking and cycling) significantly increased skeletal muscle mitochondrial and cytochrome c oxidase volume density, percentage of type I (oxidative) fibers, and number of capillaries that supply each of these fibers in HFrEF. Cytochrome c oxidase volume density also increased, and was associated with improved VO_{2peak} [50].

Improvements in skeletal muscle oxidative capacity, capillary density, and mitochondrial volume density have also been demonstrated after small muscle mass exercise training in patients with HFrEF [42, 51, 59] (Table 2). Esposito et al. [42] showed that 8 weeks of unilateral knee extension exercise significantly increased vastus lateralis muscle fiber cross-sectional area, percent type I fibers, muscle capillarity, and

mitochondrial volume density. The improvement in skeletal muscle morphology with training correlated with the increase in VO_{2peak} assessed during maximal cycle exercise [42]. Overall, these studies highlight the ability of aerobic exercise training to induce rapid adaptations in skeletal muscle morphology and function in patients with HFrEF, and to improve exercise tolerance and VO_{2peak}.

Improvements in exercise tolerance and VO_{2peak} have also been observed following resistance training performed alone [53, 67–69] or combined with aerobic exercise training [68, 70] in HFrEF. Despite the paucity of studies investigating the peripheral adaptations associated with resistance exercise [53], it appears that increases in oxidative muscle fiber cross-sectional area and oxidative enzyme capacity are likely contributors. Pu et al. [53] demonstrated that 10 weeks of high-intensity progressive resistance exercise training in older women with HFrEF increased skeletal muscle type I fiber area and citrate synthase activity, and were predictive of improvements in functional capacity (assessed by 6-min walk distance).

Exercise Interventions Improve Exercise Tolerance and Skeletal Muscle Function in HFpEF

Similar to HFrEF, aerobic exercise training has been shown to increase VO_{2peak} by 2.1–3.0 ml/kg/min [71–74, 75•] compared to usual care in patients with HFpEF [58••, 62, 76–79]. However, in contrast to HFrEF, this form of training is not associated with increased peak exercise cardiac output [62, 63••]. Specifically, Haykowsky et al. [63••] demonstrated that 84% of the improvement in VO_{2peak} following 16 weeks of aerobic exercise training was attributed to increases in peak exercise a-vO₂diff (Table 2). Similarly, Fu et al. [62] recently reported that 12 weeks of high-intensity interval exercise training significantly increased VO_{2peak}, with the

improvements in VO_{2peak} driven by increases in estimated peak exercise $a-vO_2diff$ and leg muscle oxygenation, with little or no change in peak exercise cardiac output. The mechanisms responsible for these exercise-mediated peripheral adaptations that underlie improvements in peak exercise $a-vO_2diff$ seem likely to relate to improved peripheral muscle perfusion and/or enhanced mitochondrial function.

Novel Therapies to Target Skeletal Muscle Abnormalities in HF

As the key role of skeletal muscle in exercise tolerance has been recognized, multiple initiatives have been underway to improve skeletal muscle performance. Supplementing nutrition has demonstrated benefit as it responds to the hypercatabolic and malnourished state of typical HF patients [80]. Paradoxically, it has also been demonstrated that caloric restriction is also beneficial [81]. In the latter, benefits are mediated by healthful molecular signaling that stimulates clinical benefits [82]. In addition to dietary manipulations, a multitude of pharmacological-based research efforts are underway in which novel therapies are being studied to promote skeletal muscle growth in adults with sarcopenia, and which can presumably be applied to those with HF.

Nutrition

Several HFpEF studies have substantiated the premise of amino acid supplementation to improve exercise tolerance [83•, 84•, 85]. In a randomized controlled trial by Aquilani et al. [83•], the benefits of an oral nutritional mixture of amino acids (4 g twice daily) versus a placebo were compared in 95 stable elderly HFpEF patients (NYHA Class II–III). VO_{2peak} improved in the nutrition supplemented group. More recently, Lombardi et al. [84•] demonstrated that supplementing HFpEF patients (NYHA Class II–III) with one specific amino acid (L-carnosine) every day (500 mg dosage) for 6 months significantly improved exercise tolerance and functional capacity. These findings suggest that amino acid supplementation may improve exercise tolerance in patients with HFpEF as a result of correcting an amino acid deficiency either within cardiac or skeletal muscle. While it seems probable that similar nutritional supplementation would benefit patients with HFpEF as much as those with HFrEF, studies in this population have not yet been completed.

In contrast to nutritional supplements, nutritionally balanced caloric restriction has been demonstrated to trigger vital subcellular benefits in older adults through a very different mechanism of action [86]. Key molecular signaling pathways (e.g., mTor and AMPkinase) are suppressed or stimulated, with downstream clinical benefits [86]. In older, obese individuals without HF, caloric restriction has been shown to

improve LV mass and diastolic function, exercise capacity, body composition, and skeletal muscle function [87–90].

Kitzman et al. [58••] studied similar principles in older, obese HFpEF patients, comparing the effects of 20 weeks of caloric restriction or aerobic exercise training alone, or in combination, on VO_{2peak} and quality of life. Aerobic exercise training (+1.2 ml/kg/min) and caloric restriction (+1.3 ml/kg/min) both yielded similar improvements in VO_{2peak} and functional capacity, while a combination of both (aerobic exercise + caloric restriction) caused an additive effect on VO_{2peak} (+2.5 ml/kg/min). Both aerobic exercise training and caloric restriction reduced body weight and fat mass, while caloric restriction improved muscle leg muscle quality, and reduced abdominal and thigh subcutaneous fat. In addition, the change in VO_{2peak} was positively correlated with both the change in percent lean mass and the change in thigh muscle to intermuscular fat ratio. These findings demonstrate that caloric restriction alone or combined with aerobic exercise yield favorable improvements in body composition (including improved muscle quality).

Nonetheless, the long-term efficacy of caloric restriction for improving clinical outcomes in HF patients entails many aspects of clinical complexity that are inherently problematic. Older adults who are prone to developing HF are also susceptible to sarcopenia and frailty. Benefits of caloric restriction must be counterbalanced by the risks it may impose as it undercuts vital nutrition in patients who are relatively more enfeebled. Furthermore, observational studies report an obesity paradox in this patient population [91, 92], with overweight and obese HFpEF patients having better survival outcomes than those who are normal or underweight according to body mass index.

Novel Pharmacological Approaches

Pharmacological approaches to skeletal muscle growth remain an active area of research. Initiatives primarily target age-related sarcopenia, but with a common presumption that older HF patients may benefit disproportionately due to the additive effects of aging and disease on skeletal muscle atrophy and weakening. Pharmacotherapies targeting myostatin inhibition remain a particularly compelling consideration [93]. Myostatin is a highly conserved member of the transforming growth factor-beta superfamily that signals through the activin receptor type IIB (ActRIIB). Myostatin stimulates catabolic processes, and inhibits transcription of genes that underlie proliferation of skeletal muscle precursor cells. Thus, myostatin inhibition may moderate or reverse skeletal muscle loss and functional decline. Nonetheless, trials of myostatin inhibitors have revealed many side effects that heretofore have diminished enthusiasm for clinical application (e.g., aseptic meningitis, diarrhea, confusion, fatigue, involuntary muscle contractions) [93]. Nonetheless, ongoing studies with the anti-ActRII

Table 2 Summary of exercise training-mediated skeletal muscle adaptations that contribute to improved exercise tolerance in patients with heart failure with reduced or preserved ejection fraction

Variable	HFrEF	HFpEF
Morphology		
Lean body mass	↔ [53–57]	↔ [58••]
% type I fibers	↑ [42, 49, 53] ↔ [51, 59]	Not studied.
% type II fibers	↓ [42, 49] ↔ [51, 59]	Not studied.
Capillary density	↑ [42, 48, 51]	Not studied.
Mitochondrial volume density	↑ [42, 49, 50]	Not studied.
Mitochondrial enzyme density	↑ [42, 49–53, 59]	Not studied.
Function		
Peak exercise a-vO ₂ diff	↑ [42, 50, 60, 61] ↔ [62]	↑ [62, 63••]
Sub-maximal exercise oxidative metabolism	↑ [56, 64–66]	Not studied.

↑ increase, ↓ decrease, ↔ stays the same, a-vO₂diff arterial-venous oxygen content difference, HFpEF heart failure with preserved ejection fraction, HFrEF heart failure with reduced ejection fraction

antibody “Bimagrumab” (BYM338) remain an eagerly anticipated focus of investigation [94].

Related research is focused on integrated regulatory mechanisms that determine muscle metabolism and growth. In part, this also relates to myostatin pathways, as myostatin also inhibits anabolic pathways in skeletal muscle in response to pro-growth signals (e.g., insulin and insulin-like growth factor-1). Moreover, in addition to myostatin inhibition, supplementation of anabolic agents (e.g., testosterone) has been an active area of investigation. While persistent concerns regarding secondary risks of testosterone (e.g., fluid retention, gynecomastia, prostate tumors, and adverse lipid profiles) have diminished enthusiasm for clinical applications, there is still strong interest in its therapeutic potential. In contrast to early studies that utilized high-dose testosterone, those using more physiological testosterone doses achieve greater safety and benefit [95]. Furthermore, as compared to oral formulations, transdermal or intramuscular administration is safer and better tolerated [96]. Furthermore, interest in selective androgen-receptor modulators (SARMs) like enobosarm has advanced as an alternative means of treating muscle and bone disorders, with relatively fewer side effects than testosterone [97].

Conclusions

Heart failure is associated with multiple skeletal muscle abnormalities (reduced lean mass, oxidative fiber percentage, capillarity, oxidative enzyme capacity, and mitochondrial volume), which impair oxygen uptake and utilization and contribute greatly to exercise intolerance. Large and small muscle mass exercise training induces favorable adaptations in skeletal muscle morphology and function (increased oxidative fiber percentage, capillarity, oxidative enzyme capacity, and mitochondrial

volume) in patients with HFrEF. Further, these adaptations are associated with increased exercise tolerance. In patients with HFpEF, improvements in exercise tolerance following aerobic exercise training are primarily mediated through peripheral “non-cardiac” factors with little to no change in cardiac output. The contribution of skeletal muscle adaptations to improved exercise tolerance in HFpEF remains unknown and warrants further investigation. Furthermore, adjunctive strategies, both to supplement nutrition with amino acids, and to stimulate anabolic molecular pathways in skeletal muscle with caloric restriction are beneficial, with synergy observed when combined with exercise training. Parallel investigations are exploring the utility of pharmacological strategies to similarly stimulate healthful molecular signaling and anabolic cell metabolism for older patients who have both sarcopenia and HF, but these efforts remain early in their evolution, and no immediate clinical applications.

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Compliance with Ethical Standards

Conflict of Interest The authors declare that they have no conflict of interest.

Human and Animal Rights and Informed Consent This article does not contain any studies with human or animal subjects performed by any of the authors.

References

Papers of particular interest, published recently, have been highlighted as:

- Of importance
- Of major importance

1. Benjamin EJ, Virani SS, Callaway CW, Chamberlain AM, Chang AR, Cheng S, et al. Heart Disease and Stroke Statistics-2018 update: a report from the American Heart Association. *Circulation*. 2018;137(12):e67–e492.
2. • Haykowsky MJ, Tomczak CR, Scott JM, Paterson DI, Kitzman DW. Determinants of exercise intolerance in patients with heart failure and reduced or preserved ejection fraction. *J Appl Physiol* (1985). 2015;119(6):739–44. **This review covers the mechanisms responsible for exercise intolerance in patients with heart failure and reduced or preserved ejection fraction.**
3. Upadhyya B, Kitzman DW. Heart Failure with preserved ejection fraction in older adults. *Heart Fail Clin*. 2017;13(3):485–502.
4. Borlaug BA, Melenovsky V, Russell SD, Kessler K, Pacak K, Becker LC, et al. Impaired chronotropic and vasodilator reserves limit exercise capacity in patients with heart failure and a preserved ejection fraction. *Circulation*. 2006;114(20):2138–47.
5. Borlaug BA, Olson TP, Lam CS, Flood KS, Lerman A, Johnson BD, et al. Global cardiovascular reserve dysfunction in heart failure with preserved ejection fraction. *J Am Coll Cardiol*. 2010;56(11):845–54.
6. •• Haykowsky MJ, Brubaker PH, John JM, Stewart KP, Morgan TM, Kitzman DW. Determinants of exercise intolerance in elderly heart failure patients with preserved ejection fraction. *J Am Coll Cardiol*. 2011;58(3):265–74. **This cross-sectional study investigated the mechanisms responsible for reduced exercise tolerance in heart failure with preserved ejection fraction. Reduced peak exercise VO_2 was associated with reduced peak cardiac output and reduced peak arteriovenous oxygen content difference ($a\text{-vO}_2\text{diff}$); however, the change in $a\text{-vO}_2\text{diff}$ was the strongest independent predictor of peak VO_2 during exercise.**
7. Sullivan MJ, Knight JD, Higginbotham MB, Cobb FR. Relation between central and peripheral hemodynamics during exercise in patients with chronic heart failure. Muscle blood flow is reduced with maintenance of arterial perfusion pressure. *Circulation*. 1989;80(4):769–81.
8. Wilson JR, Mancini DM, Dunkman WB. Exertional fatigue due to skeletal muscle dysfunction in patients with heart failure. *Circulation*. 1993;87(2):470–5.
9. •• Dhakal BP, Malhotra R, Murphy RM, Pappagianopoulos PP, Baggish AL, Weiner RB, et al. Mechanisms of exercise intolerance in heart failure with preserved ejection fraction: the role of abnormal peripheral oxygen extraction. *Circ Heart Fail*. 2015;8(2):286–94. **First study to directly measure arteriovenous oxygen content ($a\text{-vO}_2\text{diff}$) throughout exercise in heart failure with reduced ejection fraction (HFrEF), preserved ejection fraction (HFpEF), or normal individuals. Reduced peak exercise $a\text{-vO}_2\text{diff}$ was the major determinant of exercise capacity in HFpEF.**
10. Esposito F, Mathieu-Costello O, Shabetai R, Wagner PD, Richardson RS. Limited maximal exercise capacity in patients with chronic heart failure: partitioning the contributors. *J Am Coll Cardiol*. 2010;55(18):1945–54.
11. Forman DE, Arena R, Boxer R, Dolansky MA, Eng JJ, Fleg JL, et al. Prioritizing functional capacity as a principal end point for therapies oriented to older adults with cardiovascular disease: a scientific statement for healthcare professionals from the American Heart Association. *Circulation*. 2017;135(16):e894–918.
12. Poole DC, Richardson RS, Haykowsky MJ, Hirai DM, Musch TI. Exercise limitations in heart failure with reduced and preserved ejection fraction. *J Appl Physiol* (1985). 2018;124(1):208–24.
13. Houstis NE, Eisman AS, Pappagianopoulos PP, Wooster L, Bailey CS, Wagner PD, et al. Exercise intolerance in heart failure with preserved ejection fraction: diagnosing and ranking its causes using personalized O_2 pathway analysis. *Circulation*. 2018;137(2):148–61.
14. Kitzman DW, Higginbotham MB, Cobb FR, Sheikh KH, Sullivan MJ. Exercise intolerance in patients with heart failure and preserved left ventricular systolic function: failure of the Frank-Starling mechanism. *J Am Coll Cardiol*. 1991;17(5):1065–72.
15. Sullivan MJ, Green HJ, Cobb FR. Altered skeletal muscle metabolic response to exercise in chronic heart failure. Relation to skeletal muscle aerobic enzyme activity. *Circulation*. 1991;84(4):1597–607.
16. •• Weiss K, Schar M, Panjath GS, Zhang Y, Sharma K, Bottomley PA, et al. Fatigability, exercise intolerance, and abnormal skeletal muscle energetics in heart failure. *Circ Heart Fail*. 2017;10(7). **This cross-sectional study showed that patients with heart failure with reduced or preserved ejection fraction with exercise intolerance exhibit an accelerated rate of phosphocreatine depletion during plantar flexion exercise compared with healthy controls and HFpEF patients without exercise intolerance.**
17. Anker SD, Ponikowski P, Varney S, Chua TP, Clark AL, Webb-Peploe KM, et al. Wasting as independent risk factor for mortality in chronic heart failure. *Lancet*. 1997;349(9058):1050–3.
18. Cicoira M, Zanolla L, Franceschini L, Rossi A, Golia G, Zamboni M, et al. Skeletal muscle mass independently predicts peak oxygen consumption and ventilatory response during exercise in noncachectic patients with chronic heart failure. *J Am Coll Cardiol*. 2001;37(8):2080–5.
19. Fulster S, Tacke M, Sandek A, Ebner N, Tschope C, Doehner W, et al. Muscle wasting in patients with chronic heart failure: results from the studies investigating co-morbidities aggravating heart failure (SICA-HF). *Eur Heart J*. 2013;34(7):512–9.
20. Mancini DM, Walter G, Reichel N, Lenkinski R, McCully KK, Mullen JL, et al. Contribution of skeletal muscle atrophy to exercise intolerance and altered muscle metabolism in heart failure. *Circulation*. 1992;85(4):1364–73.
21. Minotti JR, Christoph I, Oka R, Weiner MW, Wells L, Massie BM. Impaired skeletal muscle function in patients with congestive heart failure. Relationship to systemic exercise performance. *J Clin Invest*. 1991;88(6):2077–82.
22. Drexler H, Riede U, Munzel T, Konig H, Funke E, Just H. Alterations of skeletal muscle in chronic heart failure. *Circulation*. 1992;85(5):1751–9.
23. Magnusson G, Kaijser L, Rong H, Isberg B, Sylven C, Saltin B. Exercise capacity in heart failure patients: relative importance of heart and skeletal muscle. *Clin Physiol*. 1996;16(2):183–95.
24. Massie BM, Simonini A, Sahgal P, Wells L, Dudley GA. Relation of systemic and local muscle exercise capacity to skeletal muscle characteristics in men with congestive heart failure. *J Am Coll Cardiol*. 1996;27(1):140–5.
25. Schaufelberger M, Andersson G, Eriksson BO, Grimby G, Held P, Swedberg K. Skeletal muscle changes in patients with chronic heart failure before and after treatment with enalapril. *Eur Heart J*. 1996;17(11):1678–85.
26. Sullivan MJ, Green HJ, Cobb FR. Skeletal muscle biochemistry and histology in ambulatory patients with long-term heart failure. *Circulation*. 1990;81(2):518–27.
27. Minotti JR, Pillay P, Oka R, Wells L, Christoph I, Massie BM. Skeletal muscle size: relationship to muscle function in heart failure. *J Appl Physiol* (1985). 1993;75(1):373–81.
28. Piepoli MF, Kaczmarek A, Francis DP, Davies LC, Rauchhaus M, Jankowska EA, et al. Reduced peripheral skeletal muscle mass and

- abnormal reflex physiology in chronic heart failure. *Circulation*. 2006;114(2):126–34.
29. Lang CC, Chomsky DB, Rayos G, Yeoh TK, Wilson JR. Skeletal muscle mass and exercise performance in stable ambulatory patients with heart failure. *J Appl Physiol* (1985). 1997;82(1):257–61.
 30. Haykowsky MJ, Brubaker PH, Morgan TM, Kritchevsky S, Eggebeen J, Kitzman DW. Impaired aerobic capacity and physical functional performance in older heart failure patients with preserved ejection fraction: role of lean body mass. *J Gerontol A Biol Sci Med Sci*. 2013;68(8):968–75.
 31. •• Kitzman DW, Nicklas B, Kraus WE, Lyles MF, Eggebeen J, Morgan TM, et al. Skeletal muscle abnormalities and exercise intolerance in older patients with heart failure and preserved ejection fraction. *Am J Physiol Heart Circ Physiol*. 2014;306(9):H1364–70. **Using needle biopsies of the vastus lateralis, this cross-sectional study demonstrated that older patients with heart failure and preserved ejection fraction exhibit multiple skeletal muscle abnormalities characterized by a shift in muscle fiber type distribution with reduced type I oxidative muscle fibers and a reduced capillary-to-fiber ratio that are both associated with reduced VO_{2peak}.**
 32. Duscha BD, Kraus WE, Keteyian SJ, Sullivan MJ, Green HJ, Schachar FH, et al. Capillary density of skeletal muscle: a contributing mechanism for exercise intolerance in class II-III chronic heart failure independent of other peripheral alterations. *J Am Coll Cardiol*. 1999;33(7):1956–63.
 33. •• Molina AJ, Bharadwaj MS, Van Horn C, Nicklas BJ, Lyles MF, Eggebeen J, et al. Skeletal muscle mitochondrial content, oxidative capacity, and mfn2 expression are reduced in older patients with heart failure and preserved ejection fraction and are related to exercise intolerance. *JACC Heart Fail*. 2016;4(8):636–45. **Using needle biopsies of the vastus lateralis, this cross-sectional study demonstrates that skeletal muscle oxidative capacity, mitochondrial content, and mitochondrial fusion are abnormal in older patients with heart failure and preserved ejection fraction and may contribute to severe exercise intolerance in this patient population.**
 34. Martin WH 3rd, Berman WI, Buckley JC, Snell PG, Blomqvist CG. Effects of active muscle mass size on cardiopulmonary responses to exercise in congestive heart failure. *J Am Coll Cardiol*. 1989;14(3):683–94.
 35. Bhella PS, Prasad A, Heinicke K, Hastings JL, Arbab-Zadeh A, Adams-Huet B, et al. Abnormal haemodynamic response to exercise in heart failure with preserved ejection fraction. *Eur J Heart Fail*. 2011;13(12):1296–304.
 36. Abudiyab MM, Redfield MM, Melenovsky V, Olson TP, Kass DA, Johnson BD, et al. Cardiac output response to exercise in relation to metabolic demand in heart failure with preserved ejection fraction. *Eur J Heart Fail*. 2013;15(7):776–85.
 37. Chati Z, Zannad F, Robin-Lherbier B, Escanye JM, Jeandel C, Robert J, et al. Contribution of specific skeletal muscle metabolic abnormalities to limitation of exercise capacity in patients with chronic heart failure: a phosphorus 31 nuclear magnetic resonance study. *Am Heart J*. 1994;128(4):781–92.
 38. Lipkin DP, Jones DA, Round JM, Poole-Wilson PA. Abnormalities of skeletal muscle in patients with chronic heart failure. *Int J Cardiol*. 1988;18(2):187–95.
 39. Mancini DM, Coyle E, Coggan A, Beltz J, Ferraro N, Montain S, et al. Contribution of intrinsic skeletal muscle changes to 31P NMR skeletal muscle metabolic abnormalities in patients with chronic heart failure. *Circulation*. 1989;80(5):1338–46.
 40. Massie BM, Conway M, Yonge R, Frostick S, Sleight P, Ledingham J, et al. 31P nuclear magnetic resonance evidence of abnormal skeletal muscle metabolism in patients with congestive heart failure. *Am J Cardiol*. 1987;60(4):309–15.
 41. van der Ent M, Jeneson JA, Remme WJ, Berger R, Ciampricotti R, Visser F. A non-invasive selective assessment of type I fibre mitochondrial function using 31P NMR spectroscopy. Evidence for impaired oxidative phosphorylation rate in skeletal muscle in patients with chronic heart failure. *Eur Heart J*. 1998;19(1):124–31.
 42. Esposito F, Reese V, Shabetai R, Wagner PD, Richardson RS. Isolated quadriceps training increases maximal exercise capacity in chronic heart failure: the role of skeletal muscle convective and diffusive oxygen transport. *J Am Coll Cardiol*. 2011;58(13):1353–62.
 43. Haykowsky MJ, Kouba EJ, Brubaker PH, Nicklas BJ, Eggebeen J, Kitzman DW. Skeletal muscle composition and its relation to exercise intolerance in older patients with heart failure and preserved ejection fraction. *Am J Cardiol*. 2014;113(7):1211–6.
 44. Haykowsky MJ, Nicklas BJ, Brubaker PH, Hundley WG, Brinkley TE, Upadhyaya B, et al. Regional adipose distribution and its relationship to exercise intolerance in older obese patients who have heart failure with preserved ejection fraction. *JACC Heart Fail*. 2018.
 45. Delmonico MJ, Harris TB, Visser M, Park SW, Conroy MB, Velasquez-Mieyer P, et al. Longitudinal study of muscle strength, quality, and adipose tissue infiltration. *Am J Clin Nutr*. 2009;90(6):1579–85.
 46. Haykowsky MJ, Liang Y, Pechter D, Jones LW, McAlister FA, Clark AM. A meta-analysis of the effect of exercise training on left ventricular remodeling in heart failure patients: the benefit depends on the type of training performed. *J Am Coll Cardiol*. 2007;49(24):2329–36.
 47. Weston KS, Wisloff U, Coombes JS. High-intensity interval training in patients with lifestyle-induced cardiometabolic disease: a systematic review and meta-analysis. *Br J Sports Med*. 2014;48(16):1227–34.
 48. Erbs S, Holtriegel R, Linke A, Beck EB, Adams V, Gielen S, et al. Exercise training in patients with advanced chronic heart failure (NYHA IIIb) promotes restoration of peripheral vasomotor function, induction of endogenous regeneration, and improvement of left ventricular function. *Circ Heart Fail*. 2010;3(4):486–94.
 49. Hambrecht R, Fiehn E, Yu J, Niebauer J, Weigl C, Hilbrich L, et al. Effects of endurance training on mitochondrial ultrastructure and fiber type distribution in skeletal muscle of patients with stable chronic heart failure. *J Am Coll Cardiol*. 1997;29(5):1067–73.
 50. Hambrecht R, Niebauer J, Fiehn E, Kalberer B, Offner B, Hauer K, et al. Physical training in patients with stable chronic heart failure: effects on cardiorespiratory fitness and ultrastructural abnormalities of leg muscles. *J Am Coll Cardiol*. 1995;25(6):1239–49.
 51. Magnusson G, Gordon A, Kaijser L, Sylven C, Isberg B, Karpakka J, et al. High intensity knee extensor training, in patients with chronic heart failure. Major skeletal muscle improvement. *Eur Heart J*. 1996;17(7):1048–55.
 52. Wisloff U, Stoylen A, Loennechen JP, Bruvold M, Rognum O, Haram PM, et al. Superior cardiovascular effect of aerobic interval training versus moderate continuous training in heart failure patients: a randomized study. *Circulation*. 2007;115(24):3086–94.
 53. Pu CT, Johnson MT, Forman DE, Hausdorff JM, Roubenoff R, Foldvari M, et al. Randomized trial of progressive resistance training to counteract the myopathy of chronic heart failure. *J Appl Physiol* (1985). 2001;90(6):2341–50.
 54. Bouchla A, Karatzanos E, Dimopoulos S, Tasoulis A, Agapitou V, Diakos N, et al. The addition of strength training to aerobic interval training: effects on muscle strength and body composition in CHF patients. *J Cardiopulm Rehabil Prev*. 2011;31(1):47–51.
 55. Jankowska EA, Wegrzynowska K, Superlak M, Nowakowska K, Lazorczyk M, Biel B, et al. The 12-week progressive quadriceps resistance training improves muscle strength, exercise capacity and quality of life in patients with stable chronic heart failure. *Int J Cardiol*. 2008;130(1):36–43.

56. Minotti JR, Johnson EC, Hudson TL, Zuroske G, Murata G, Fukushima E, et al. Skeletal muscle response to exercise training in congestive heart failure. *J Clin Invest*. 1990;86(3):751–8.
57. Senden PJ, Sabelis LW, Zonderland ML, Hulzebos EH, Bol E, Mosterd WL. The effect of physical training on workload, upper leg muscle function and muscle areas in patients with chronic heart failure. *Int J Cardiol*. 2005;100(2):293–300.
58. •• Kitzman DW, Brubaker P, Morgan T, Haykowsky M, Hundley G, Kraus WE, et al. Effect of caloric restriction or aerobic exercise training on peak oxygen consumption and quality of life in obese older patients with heart failure with preserved ejection fraction: a randomized clinical trial. *JAMA*. 2016;315(1):36–46. **This randomized, attention-controlled, 2 × 2 factorial trial investigated whether 20 weeks of caloric restriction, exercise, or both improves exercise capacity in obese older patients with heart failure and preserved ejection fraction. Exercise training and caloric restriction both yielded similar improvements in VO_{2peak}, while a combination of both caused an additive effect on VO_{2peak}. Both exercise training and caloric restriction reduced body weight and fat mass, while caloric restriction improved muscle leg muscle quality. In addition, the change in VO_{2peak} was positively correlated with both the change in percent lean mass and the change in thigh muscle to intermuscular fat ratio.**
59. Tyni-Lenne R, Gordon A, Jensen-Urstad M, Dencker K, Jansson E, Sylven C. Aerobic training involving a minor muscle mass shows greater efficiency than training involving a major muscle mass in chronic heart failure patients. *J Card Fail*. 1999;5(4):300–7.
60. Dubach P, Myers J, Dziekan G, Goebbels U, Reinhart W, Muller P, et al. Effect of high intensity exercise training on central hemodynamic responses to exercise in men with reduced left ventricular function. *J Am Coll Cardiol*. 1997;29(7):1591–8.
61. Sullivan MJ, Higginbotham MB, Cobb FR. Exercise training in patients with severe left ventricular dysfunction. Hemodynamic and metabolic effects. *Circulation*. 1988;78(3):506–15.
62. Fu TC, Yang NI, Wang CH, Cheng WJ, Chou SL, Pan TL, et al. Aerobic interval training elicits different hemodynamic adaptations between heart failure patients with preserved and reduced ejection fraction. *Am J Phys Med Rehabil*. 2016;95(1):15–27.
63. •• Haykowsky MJ, Brubaker PH, Stewart KP, Morgan TM, Eggebeen J, Kitzman DW. Effect of endurance training on the determinants of peak exercise oxygen consumption in elderly patients with stable compensated heart failure and preserved ejection fraction. *J Am Coll Cardiol*. 2012;60(2):120–8. **This single-blind, randomized controlled trial in elderly heart failure with preserved ejection fraction patients showed that the improvement in peak exercise VO₂ (exercise capacity) following 4 months of exercise training was driven primarily by peripheral adaptations with little to no changes in peak exercise cardiac output.**
64. Adamopoulos S, Coats AJ, Brunotte F, Arnolda L, Meyer T, Thompson CH, et al. Physical training improves skeletal muscle metabolism in patients with chronic heart failure. *J Am Coll Cardiol*. 1993;21(5):1101–6.
65. Ohtsubo M, Yonezawa K, Nishijima H, Okita K, Hanada A, Kohya T, et al. Metabolic abnormality of calf skeletal muscle is improved by localised muscle training without changes in blood flow in chronic heart failure. *Heart*. 1997;78(5):437–43.
66. Stratton JR, Dunn JF, Adamopoulos S, Kemp GJ, Coats AJ, Rajagopalan B. Training partially reverses skeletal muscle metabolic abnormalities during exercise in heart failure. *J Appl Physiol* (1985). 1994;76(4):1575–82.
67. Giuliano C, Karahalios A, Neil C, Allen J, Levinger I. The effects of resistance training on muscle strength, quality of life and aerobic capacity in patients with chronic heart failure - a meta-analysis. *Int J Cardiol*. 2017;227:413–23.
68. Jewiss D, Ostman C, Smart NA. The effect of resistance training on clinical outcomes in heart failure: a systematic review and meta-analysis. *Int J Cardiol*. 2016;221:674–81.
69. Maiorana A, O'Driscoll G, Cheetham C, Collis J, Goodman C, Rankin S, et al. Combined aerobic and resistance exercise training improves functional capacity and strength in CHF. *J Appl Physiol* (1985). 2000;88(5):1565–70.
70. Tucker W, Beaudry RI, Liang Y, Clark AM, Tomczak CR, Nelson MD, et al. Meta-analysis of exercise training on left ventricular ejection fraction in heart failure with reduced ejection fraction: a 10-year update. *Prog Cardiovasc Dis*. 2018. Accepted for publication: In press.
71. Dieberg G, Ismail H, Giallauria F, Smart NA. Clinical outcomes and cardiovascular responses to exercise training in heart failure patients with preserved ejection fraction: a systematic review and meta-analysis. *J Appl Physiol* (1985). 2015;119(6):726–33.
72. Pandey A, Parashar A, Kumbhani D, Agarwal S, Garg J, Kitzman D, et al. Exercise training in patients with heart failure and preserved ejection fraction: meta-analysis of randomized control trials. *Circ Heart Fail*. 2015;8(1):33–40.
73. Taylor RS, Davies EJ, Dalal HM, Davis R, Doherty P, Cooper C, et al. Effects of exercise training for heart failure with preserved ejection fraction: a systematic review and meta-analysis of comparative studies. *Int J Cardiol*. 2012;162(1):6–13.
74. Tucker WJ, Nelson MD, Beaudry RI, Halle M, Sarma S, Kitzman DW, et al. Impact of exercise training on peak oxygen uptake and its determinants in heart failure with preserved ejection fraction. *Card Fail Rev*. 2016;2(2):95–101.
75. • Tucker WJ, Lijauco CC, Hearon CM Jr, Angadi SS, Nelson MD, Sarma S, et al. Mechanisms of the Improvement in peak VO₂ with exercise training in heart failure with reduced or preserved ejection fraction. *Heart Lung Circ*. 2018;27(1):9–21. **This review covers the central and peripheral mechanisms responsible for exercise intolerance in patients with heart failure and reduced or preserved ejection fraction.**
76. Angadi SS, Mookadam F, Lee CD, Tucker WJ, Haykowsky MJ, Gaesser GA. High-intensity interval training vs. moderate-intensity continuous exercise training in heart failure with preserved ejection fraction: a pilot study. *J Appl Physiol* (1985). 2015;119(6):753–8.
77. Edelmann F, Gelbrich G, Dungen HD, Frohling S, Wachter R, Stahrenberg R, et al. Exercise training improves exercise capacity and diastolic function in patients with heart failure with preserved ejection fraction: results of the Ex-DHF (Exercise training in Diastolic Heart Failure) pilot study. *J Am Coll Cardiol*. 2011;58(17):1780–91.
78. Kitzman DW, Brubaker PH, Morgan TM, Stewart KP, Little WC. Exercise training in older patients with heart failure and preserved ejection fraction: a randomized, controlled, single-blind trial. *Circ Heart Fail*. 2010;3(6):659–67.
79. Smart NA, Haluska B, Jeffriess L, Leung D. Exercise training in heart failure with preserved systolic function: a randomized controlled trial of the effects on cardiac function and functional capacity. *Congest Heart Fail*. 2012;18(6):295–301.
80. Lin H, Zhang H, Lin Z, Li X, Kong X, Sun G. Review of nutritional screening and assessment tools and clinical outcomes in heart failure. *Heart Fail Rev*. 2016;21(5):549–65.
81. de Lucia C, Gambino G, Petraglia L, Elia A, Komici K, Femminella GD, et al. Long-term caloric restriction improves cardiac function, remodeling, adrenergic responsiveness, and sympathetic innervation in a model of postischemic heart failure. *Circ Heart Fail*. 2018;11(3):e004153.
82. Longo VD, Antebi A, Bartke A, Barzilai N, Brown-Borg HM, Caruso C, et al. Interventions to slow aging in humans: are we ready? *Aging Cell*. 2015;14(4):497–510.
83. • Aquilani R, Viglio S, Iadarola P, Opasich C, Testa A, Dioguardi FS, et al. Oral amino acid supplements improve exercise capacities in

- elderly patients with chronic heart failure. *Am J Cardiol.* 2008;101(11a):104e–10e. **This randomized, double-blind, placebo-controlled study showed that 30 days of supplementation with amino acids increased exercise capacity in elderly patients with heart failure.**
84. Lombardi C, Carubelli V, Lazzarini V, Vizzardi E, Bordonali T, Ciccarese C, et al. Effects of oral administration of orodispersible levo-carnosine on quality of life and exercise performance in patients with chronic heart failure. *Nutrition.* 2015;31(1):72–8. **This prospective, open-label, randomized controlled, parallel group study showed that 6 months of amino acid supplementation (levo-carnosine) improved exercise capacity and quality of life in heart failure patients with reduced ejection fraction.**
 85. Lombardi C, Carubelli V, Lazzarini V, Vizzardi E, Quinzani F, Guidetti F, et al. Effects of oral amino acid supplements on functional capacity in patients with chronic heart failure. *Clin Med Insights Cardiol.* 2014;8:39–44.
 86. Lopez-Otin C, Galluzzi L, JMP F, Madeo F, Kroemer G. Metabolic control of longevity. *Cell.* 2016;166(4):802–21.
 87. Beavers KM, Miller ME, Rejeski WJ, Nicklas BJ, Kritchevsky SB. Fat mass loss predicts gain in physical function with intentional weight loss in older adults. *J Gerontol A Biol Sci Med Sci.* 2013;68(1):80–6.
 88. de las Fuentes L, Waggoner AD, Mohammed BS, Stein RI, Miller BV 3rd, Foster GD, et al. Effect of moderate diet-induced weight loss and weight regain on cardiovascular structure and function. *J Am Coll Cardiol.* 2009;54(25):2376–81.
 89. Haufe S, Utz W, Engeli S, Kast P, Bohnke J, Pofahl M, et al. Left ventricular mass and function with reduced-fat or reduced-carbohydrate hypocaloric diets in overweight and obese subjects. *Hypertension.* 2012;59(1):70–5.
 90. Normandin E, Houston DK, Nicklas BJ. Caloric restriction for treatment of geriatric obesity: Do the benefits outweigh the risks? *Curr Nutr Rep.* 2015;4(2):143–55.
 91. Haass M, Kitzman DW, Anand IS, Miller A, Zile MR, Massie BM, et al. Body mass index and adverse cardiovascular outcomes in heart failure patients with preserved ejection fraction: results from the Irbesartan in Heart Failure with Preserved Ejection Fraction (I-PRESERVE) trial. *Circ Heart Fail.* 2011;4(3):324–31.
 92. Sharma A, Lavie CJ, Borer JS, Vallakati A, Goel S, Lopez-Jimenez F, et al. Meta-analysis of the relation of body mass index to all-cause and cardiovascular mortality and hospitalization in patients with chronic heart failure. *Am J Cardiol.* 2015;115(10):1428–34.
 93. Jasuja R, LeBrasseur NK. Regenerating skeletal muscle in the face of aging and disease. *Am J Phys Med Rehabil.* 2014;93(11 Suppl 3):S88–96.
 94. Morvan F, Rondeau JM, Zou C, Minetti G, Scheufler C, Scharenberg M, et al. Blockade of activin type II receptors with a dual anti-ActRIIA/IIB antibody is critical to promote maximal skeletal muscle hypertrophy. *Proc Natl Acad Sci U S A.* 2017;114(47):12448–53.
 95. Bianchi VE. Testosterone, myocardial function, and mortality. *Heart Fail Rev.* 2018.
 96. Borst SE, Shuster JJ, Zou B, Ye F, Jia H, Wokhlu A, et al. Cardiovascular risks and elevation of serum DHT vary by route of testosterone administration: a systematic review and meta-analysis. *BMC Med.* 2014;12:211.
 97. Springer J, Springer JI, Anker SD. Muscle wasting and sarcopenia in heart failure and beyond: update 2017. *ESC Heart Fail.* 2017;4(4):492–8.