

# **The Role of the Gut Microbiome in Health and Disease in the Elderly**

**Lea Ann Chen1 · Kaitlyn Boyle2**

Accepted: 5 April 2024 / Published online: 20 April 2024 © The Author(s) 2024

#### **Abstract**

**Purpose of Review** Growing evidence supports the contribution of age in the composition and function of the gut microbiome, with specifc fndings associated with health in old age and longevity.

**Recent Findings** Current studies have associated certain microbiota, such as *Butyricimonas*, *Akkermansia*, and *Odoribacter,* with healthy aging and the ability to survive into extreme old age. Furthermore, emerging clinical and pre-clinical research have shown promising mechanisms for restoring a healthy microbiome in elderly populations through various interventions such as fecal microbiota transplant (FMT), dietary interventions, and exercise programs.

**Summary** Despite several conceptually exciting interventional studies, the feld of microbiome research in the elderly remains limited. Specifcally, large longitudinal studies are needed to better understand causative relationships between the microbiome and healthy aging. Additionally, individualized approaches to microbiome interventions based on patients' comorbidities and the underlying functional capacity of their microbiomes are needed to achieve optimal results.

**Keywords** Microbiota · Aging · Geriatric · Probiotics · Frailness · Aged, 80 and older

# **Introduction**

Over the past several decades, scientists have established an extensive relationship between the gastrointestinal (GI) microbiome and host health. For example, commensal microbiota contribute to host immune system development and function [[1](#page-9-0)], with disruptions potentially contributing to immune-mediated diseases such as systemic lupus erythematosus (SLE) and infammatory bowel disease (IBD) [\[2](#page-9-1), [3\]](#page-10-0). Microbiome composition and function further infuence the metabolism of nutrients and drugs [\[4](#page-10-1), [5](#page-10-2)]. Growing research also suggests an important role for microbes in the gut-brain axis that modulates neuropsychological and sensory disorders, such as autism and irritable bowel syndrome (IBS) [\[6](#page-10-3), [7](#page-10-4)].

Scientists have yet to identify one specifc healthy microbiome, and it is generally agreed upon that there is no singular "normal" composition [[8\]](#page-10-5). This is likely due to the vast

 $\boxtimes$  Lea Ann Chen leaann.chen@rutgers.edu spectrum of factors that infuence the gut microbiome, including variations in diet [[9\]](#page-10-6), genetics [[10](#page-10-7)], and environment [[11\]](#page-10-8). As such, changes in the microbiome also occur as part of the natural aging process. Microbiome development begins at birth as soon as newborns exit the vaginal canal and encounter the mother's vaginal fluids [\[12\]](#page-10-9), or alternatively through the skin and the environment for babies delivered by Cesarean section [[13\]](#page-10-10). For breast-fed infants, breastmilk contains several prebiotics (e.g., human milk oligosaccharides) which selectively support the growth of benefcial bacteria in the GI tract [\[13](#page-10-10)]. As children transition to solid foods, they encounter new dietary components such as starches and cell wall polysaccharides; their microbiomes must then shift to select for bacteria capable of metabolizing these nutrients [[14\]](#page-10-11). While changes in infant and early childhood microbiomes have been studied extensively, there is less information regarding alterations in the gut microbiome of the elderly. Accordingly, there is growing interest in understanding the efect of aging on the host microbiome and whether aging and its associated features, such as frailty and declined cognition, can be modulated by gut bacteria. According to the United Nations, the number of people over the age of 65 worldwide in 2021 was 761 million, with that number expected to rise to 1.6 billion by 2050 [\[15\]](#page-10-12). A greater understanding of this population's microbiome is thus of growing relevance in addressing human health and disease.

<sup>&</sup>lt;sup>1</sup> Division of Gastroenterology and Hepatology, Department of Medicine, Rutgers, New Brunswick, NJ, USA

<sup>2</sup> Rutgers Robert Wood Johnson Medical School, New Brunswick, NJ, USA

Increasing research suggests that natural, or "healthy", aging leads to specifc changes in gut microbiome composition, such as the loss of certain commensal genera, including *Prevotella*, *Faecalibacterium*, and *Bifdobacterium*, and the species *Eubacterium rectale* [[16,](#page-10-13) [17\]](#page-10-14). These taxa are instead replaced at older age by other commensal organisms, such as *Butyricimonas*, *Akkermansia*, and *Odoribacter* [\[12](#page-10-9), [16](#page-10-13)[–18](#page-10-15)]. *Akkermansia muciniphila*, in particular, has been widely studied in aging and disease and is known to contribute to mucin degradation in the intestines [[19](#page-10-16)]. Some have speculated that *Akkermansia* levels can indicate health status, with an increased relative abundance (above that seen in healthy aging) associated with excellent health among centenarians and a decreased relative abundance associated with thinning of the gut mucus layer and decreased acylglycerol, [[18,](#page-10-15) [20](#page-10-17)] an endocannabinoid that regulates gut permeability and decreases intestinal infammation [\[18](#page-10-15), [20\]](#page-10-17).

Scientists have also identifed several pathobionts, or conditionally pathogenic microorganisms, that are increased in "unhealthy" aging [\[17](#page-10-14), [18](#page-10-15)], a process characterized by rapid physical and mental decline and associated with disease progression and physical frailty. Some of these pathobionts include *Eggerthella*, *Actinomyces*, and *Enterobacteriaceae*, the presence and quantity of which may help physicians predict lifespan and disease outcomes [[17](#page-10-14), [18](#page-10-15)].

One challenge in conducting and interpreting microbiome studies in the elderly is distinguishing results attributable solely to age from those due to diferent states of health. Unique subjects by which to study these questions are those who are of "extreme" old age, such as centenarians ( $\geq$  100 years old) and supercentenarians ( $\geq$  110 years old). Microbiome features of these extremely aged individuals presumably confer longevity rather than any deleterious aspects of aging. For example, bacterial strains that are often decreased in the elderly, such as *Christensenella* and *Bifdobacterium*, are actually increased in semi-supercentenarians (i.e.,  $105-109$  years old) [[18](#page-10-15)], suggesting their beneficial efect. Additionally, the highly studied *Akkermansia* taxon, which is abundant in healthy aging, is even more dramatically increased in extreme aging [[18](#page-10-15)].

To further clarify the independent contributions of age and health, Wilmanski et al. cross-sectionally evaluated gut microbiome compositions by decade of life in 3653 U.S. adults, aged 18–87 years old. The authors found that starting around 40–60 years old, individuals become more "unique" in their microbiomes as measured by the Bray–Curtis dissimilarity matrix, in which individuals' microbiomes were compared to the one most similar from the remaining subjects  $[21\bullet]$ . More microbiome dissimilarity was seen with each passing decade, a finding that was consistent regardless of sex, body mass index (BMI), and alpha diversity (i.e., within-sample diversity). Samples in this analysis were evaluated in two separate

groups ( $n = 2539$  and  $n = 1114$ ) as there was a change in microbiome vendors and sample processing during the study; nevertheless, the finding of greater microbiome dissimilarity with increasing age was present in each cohort, lending further strength to the results. In the same publication, the investigators analyzed a different cohort of only men, aged > 78 yo (*n* = 599 discovery cohort and  $n = 308$  validation cohort). Again, microbiome uniqueness, as measured by Bray–Curtis dissimilarity, was positively correlated with age. Notably, the strength of this correlation increased among healthy participants, as determined by medication use, self-perceived health, life-space score (LSC) [[22\]](#page-10-19), and walking speed. When sub-analyzing the community dwellers of this male cohort (i.e., those who did not reside in nursing homes, assisted living, or were hospitalized in the past 12 months;  $n = 706$ ), investigators identified a correlation between the relative abundance of *Bacteroides* and all-cause mortality, independent of multiple potential confounders (e.g., age, BMI, and self-perceived health). This association between *Bacteroides* abundance and mortality was even stronger among community dwelling subjects in this cohort  $85 + \text{years}$  old, suggesting the potential use of decreased *Bacteroides* as a longevity biomarker [[21](#page-10-18)••]. In contrast, a large 2023 cohort study  $(n = 1575$ , including 297 centenarians) found a relative abundance of *Bacteroides* in centenarians and furthermore identified several other "youth-associated" features, such as decreased pathobionts, in centenarians [[23](#page-10-20)]. Thus, the role of *Bacteroides spp.* and their utility as a gut microbial predictor of long life requires additional study. Another growing area of research is exploring the centenarian virome, with early results suggesting increased viral diversity and unique genera enrichment that distinguish centenarians from other older adults  $(> 60)$  [[24•](#page-10-21)].

The logical next steps in this line of research will be longitudinal studies of the microbiome to determine whether microbes associated with longevity are present earlier in life (suggesting their role in predicting or promoting long life) or if their increase occurs only upon old age, suggesting that these changes are relevant only in older age or are secondary to other factors of extreme age.

# **Geriatric Health and the Microbiome**

Although we know that microbiome compositions shift throughout the aging process, the exact mechanisms for this are unclear. Below, we explore some common life changes and medical conditions among the elderly in which intestinal microbiomes are altered (Fig. [1\)](#page-2-0).



<span id="page-2-0"></span>**Fig. 1** Lifestyle changes and medical conditions associated with alterations in the intestinal microbiome of the elderly

### **Infamm‑aging**

Studies from as early as the 1960's have indicated a decrease in immune function in aging adults [[25](#page-10-22)]. This process, now known as immunosenescence, is associated with a decline in immune system function that leads to an accumulation of pro-infammatory cytokines. The increased infammatory state in elderly populations is now commonly referred to as "infamm-aging" [[26](#page-10-23)]. Pro-infammatory states place patients at higher risk for a variety of conditions such as autoimmune and cardiovascular diseases, as well as infections [[27–](#page-10-24)[29\]](#page-10-25).

Within the GI tract, the maintenance of functioning epithelial and mucus barriers is essential for protection against infection and disease [\[30\]](#page-10-26). Increased intestinal permeability can lead to translocation of microbes into host circulation, exacerbating a pro-infammatory state [\[30\]](#page-10-26). A study in wildtype C57BL/6 mice showed that age-associated disruption of the small intestine mucosal barrier led to increased interaction between gut microbiota and the host immune system, as determined by fuorescent in situ hybridization using the bacterial probe EUB338-Alexa Fluor 488, as well as enlargement of solitary intestinal lymphoid tissue (SILT), which are hypertrophied upon interaction with gut microbes [\[31,](#page-10-27) [32](#page-10-28)]. Barrier defects were also associated with relative decreases in *Akkermansia* [\[31](#page-10-27)].

Experiments in germ-free mouse models by Thevaranjan et al. suggest that it is the changing microbiome itself in aging populations that leads to a pro-infammatory state, with germ-free mice living much longer than their conventional counterparts [[33\]](#page-10-29). Furthermore, young, germ-free mice gavaged with the microbiome of older mice developed greater intestinal permeability and circulating TNF than mice gavaged with microbiomes of other young mice [\[33](#page-10-29)]. However, additional research, especially from longitudinal studies in humans, will be necessary to confrm a causal relationship between the gut microbiome and infamm-aging.

### **Diet and Environment**

Elderly individuals requiring greater assistance with activities of daily living (ADLs) may transition from community living to long-term care facilities. This relocation has been shown to produce microbiome shifts due to presumed changes in environmental, dietary, and medical factors [[34\]](#page-11-0). For instance, in general adult population studies, the microbes residing on household surfaces correlate with gut microbiome composition, which bears consideration in the transition to long-term care environments. Furthermore, both aging and exposure to healthcare facilities, such as long-term care facilities, are associated with an increased risk for *Clostridioides difficile* infection (CDI), a major cause of healthcare-associated, infammatory diarrhea [\[35](#page-11-1)].

Regardless of age, there is strong evidence to suggest that specifc diets can cause unique alterations in the microbiome [\[36](#page-11-2), [37](#page-11-3)], as well as corresponding serum and fecal metabolites [[38](#page-11-4)]. A well-controlled study by Tanes et al. followed 30 subjects who were randomized to vegan (high fber), omnivore (intermediate fber) and formula-based (no fber) diets [\[39](#page-11-5)]. After 6 days, the subjects were given a "gut purge" using a combination of oral antibiotics and polyethylene glycol. Researchers found that the microbiome of vegan subjects recovered more rapidly after the "purge" compared to the other groups, regaining greater diversity in a shorter time span [\[39](#page-11-5)]. Subjects adhering to the formula-based diet, on the other hand, had the most prolonged recovery phase. In another cross-sectional study, the gut microbiomes of previously uncontacted Yanomami Amerindians, who live in the High Orinoco state of Venezuela and eat a largely plant-based, high-fber diet, were compared to microbiomes of individuals residing in the United States and semitransculturated populations, such as Guahibo Amerindians and Malawians. The Yanomami were noted to have a markedly higher gut microbiome diversity compared to those in the United States, with the semi-transculturated populations having an intermediate level of diversity. It is noted, however, that other social and medical factors, rather than diet alone, could also have contributed to this increased diversity [\[40\]](#page-11-6).

Nevertheless, a component of age-related changes in the microbiome appears definitively related to diet and eating, particularly as the elderly are at increased risk for poor dentition or chewing difficulties, decreased appetite, and lack of social support in obtaining nutritious foods [\[41\]](#page-11-7). For example, one of the most dramatic diet changes that has been shown to cause microbiome alterations is the move from independent, community living to assisted living within a long-term care facility. This transition often leads to a change from a high-fber, low-fat diet to a lowfber, high-fat diet, which has been associated with a shift to a lower diversity microbiome in long-term care residents compared to community dwelling counterparts [[34\]](#page-11-0). Of note, the disparity between these long-term care residents and community dwellers correlated with the amount of time spent in long-term care. During digestion, fiber is metabolized into short chain fatty acids (SCFAs), which provide many benefts to the GI tract by serving as an energy source for protective microbiota, assisting with anti-infammatory responses, and maintaining gut barrier integrity [\[42](#page-11-8)]. Thus, SCFA deficiencies caused by dietary changes when moving to long term care facility can indirectly contribute to intestinal dysfunction.

#### **Co‑morbidities**

An area of growing interest is the study of the gut-brain axis via microbes that may infuence cognitive function (Table [1\)](#page-4-0). The topic is of particular relevance as mild cognitive impairment (MCI) is highly prevalent in the elderly, afecting approximately 10% of those aged 70–74 yo and 25% of those 80–84 yo [[43\]](#page-11-9). Furthermore, patients with MCI are far more likely to progress to dementia [[43\]](#page-11-9). Pharmacologic treatments to date can only slow the progression of MCI, but not reverse it [\[44\]](#page-11-10). While there is still disagreement on whether microbiome alterations infuence cognitive function and vice versa [[45,](#page-11-11) [46](#page-11-12)], ongoing long-term projects such as MOTION (Microbiome Of the ageing gut and its efect on human gut health and cogniTION), which studies cognitive and microbiome changes of healthy aging [\[47\]](#page-11-13), provide hope that these interactions will soon be clarifed.

A 2019 study of shotgun metagenomic sequences, comparing 57 nursing home residents with dementia, including Alzheimer's disease (AD), with 51 elderly individuals without AD or other forms of dementia, revealed higher levels of pro-infammatory gut bacteria in those with dementia [\[61](#page-12-0)]. The authors also noted a decrease in butyrate-synthesizing bacterial species, such as those in the genera *Butyrivibrio* and *Eubacteria,* in the AD group when compared to both subjects without dementia and subjects with other dementias besides AD [[61](#page-12-0)]. A subsequent systematic review and metanalysis similarly found decreased alpha diversity in the gut microbiomes of AD patients compared to healthy controls, but not between those with mild cognitive impairment (MCI) and healthy controls. Diferences in microbiome compositions between AD, MCI, and healthy samples (i.e., beta diversity) were not consistently altered [[62](#page-12-1)]. One challenge in studying the gut microbiome as it relates to dementia is the lack of clear, objective, and non-invasive tests to conclusively determine diagnosis and disease stage, thus further complicating the interpretation of study results. While beyond the scope of the gut microbiome, we note with interest that post-mortem studies of AD brain tissue have identifed the presence of microbes within the brain, suggesting the presence of a brain microbiome associated with neurodegenerative disease [\[63](#page-12-2)].

Furthermore, a large genome wide association study identifed several microbiome genera associated with high risk alleles of the apolipoprotein E ε4 (APOE ε4) gene, a well-established risk factor for AD [\[64](#page-12-3)•]. Some of the most signifcant fndings of this study included a strong correlation between the pro-infammatory genus *Collinsella* and APOE risk alleles, as well as a proposed protective role for the genus *Eubacterium fssicatena* [[64](#page-12-3)•].

Parkinson's disease (PD) is another neurological disorder that is more common in the elderly and for which there is growing interest in the gut microbiome as a biomarker or therapy. A 2020 meta-analysis of 16S sequencing data from Japan, the United States, Finland, Russia, and Germany found that patients with PD have relatively decreased *Roseburia* and *Faecalibacterium –* both important producers of the SCFA butyrate [\[65](#page-12-4)]. A 2022 shotgun sequencing study of 490 PD and 234 healthy controls confrmed these fndings and also identifed several other genera that are altered in PD patients, such as an increase in pathogenic species of *Prevotella* [[66](#page-12-5)•]. Interestingly, multiple studies have noted an increase in the *Akkermansia* genus [[65\]](#page-12-4) among those with PD. This is surprising considering *Akkermansia* is generally associated with healthy aging and is particularly abundant in supercentenarians [[18\]](#page-10-15). Some scientists have speculated that *Akkermansia* is an important component of healthy

<span id="page-4-0"></span>



aging, but that increased abundance puts patients at risk for neurocognitive disease [[67](#page-12-6)]. We further hypothesize that changes in *Akkermansia* abundance may be secondary to the development of constipation, a common gastrointestinal complication of PD and a condition that has independently been associated with increased *Akkermansia* in multiple other studies [[68\]](#page-12-7). As the link between PD and *Akkermansia* is an inconsistent fnding [[66](#page-12-5) •], further research is needed to determine the precise role of this genus in PD and in aging more broadly. In a PD mouse model that overexpresses α-synuclein aggregates, a common fnding in the brains of PD patients, mice colonized with the gut microbiome of 6 human PD patients had increased physical motor impair ments and constipation compared to mice colonized with healthy donor microbiota [\[69\]](#page-12-8). Building on these early fndings of altered microbiota in PD, a pilot randomized control trial found that stool from healthy donors, given as lyophilized pills twice a week for 12 weeks, could improve constipation and gut motility as well as transiently improve objective motor skills among patients with mild to moderate PD [\[70](#page-12-9)•]. While significant translational and clinical data development are still needed, these initial fndings maintain the promise that gut microbiome modulation may improve gastrointestinal and/or neurological symptoms of PD and provide deeper insight into disease pathophysiology.

Several early-stage studies have also been conducted on the relationship between the gut microbiome and sarcope nia, the progressive deterioration of muscle mass that occurs with aging and that leads to physical frailty. While these studies have yielded conficting results about which bacterial species are increased or decreased in the condition, study fndings have consistently demonstrated no change in over all microbial diversity between frail and non-frail elderly individuals [\[71](#page-12-10), [72\]](#page-12-11). Pre-clinical experiments have also sug gested a role for gut bacteria in skeletal health, although [deta](#page-12-12)[ils](#page-12-13) of how these effects are mediated have been unclear [[73,](#page-12-12) [74\]](#page-12-13). In correlating human subject research, a relatively large 16S study (i.e., 60 individuals with osteoporosis and 60 age- and gender-matched controls with normal bone mineral density) found a relative abundance of *Actinomy ces, Clostridium XIVa, Eggerthella,* and *Lactobacillus* and a relative decrease in *Veillonella* in those with osteoporosis. There were, however, no changes in overall microbiome alpha diversity between groups [\[75](#page-12-14)].

# **Interventions to Delay or Reverse Aging**

While a more thorough understanding is required of the microbial changes that can be isolated to age specifcally, studies have already begun evaluating how to restore a healthy microbiome in aging populations to promote health and longevity.

### **FMT**

Fecal microbiota transplant (FMT) is a therapy that has been growingly incorporated into the treatment of recurrent CDI [[76\]](#page-12-15) and has furthermore been studied for infammatory bowel disease [\[77](#page-12-16)] and post-antibiotic dysbiosis [[78\]](#page-12-17). During FMT, stool from a healthy donor is transplanted into a recipient via colonoscopy, naso- or oro-enteric tubes, enema, or capsule, with the goal of transferring the corresponding intestinal microbes, as well as their contained functions and metabolic products. This has led some to speculate whether the microbiome from a young, healthy donor can be transplanted into an elderly individual to reverse some of the efects of unhealthy aging (Table [2](#page-7-0)).

A study by Parker et. al demonstrated that transfer of an "aged" microbiome from elderly mice to younger mice caused several age-associated phenotypes including advanced central nervous system deterioration and vision deficits  $[79\bullet]$ . Importantly, in a set of correlating experiments, age-related changes improved in elderly mice after microbiome transplantation with stool of younger mice [\[79•](#page-12-18)]. This work provides strong pre-clinical evidence that microbiome profles between young and aged mice are not only diferent, but that the associated physiological efects of these microbiomes are transferrable. These and similar fndings have been reproduced by other investigators [[59](#page-11-25)], including D'Amato et al., who demonstrated that transferring the microbiome of elderly mice to young ones can lead to cognitive deficits [[80](#page-12-19)].

Progeria is a particularly unique disease by which to study microbiome and senescence, as afected individuals carry a mutation in the gene encoding lamin A which leads to rapid aging. Despite a normal appearance at birth, afected individuals typically develop fatal complications of their disease, predominantly cardiovascular disease, in their teens or early adulthood [[89\]](#page-13-0). Using a mouse model of progeria, Bárcena et al. showed that certain bacterial strains enriched in human centenarians such as *Akkermansia muciniphila* can be transplanted to increase mouse lifespan and to reverse intestinal mucosal thinning [\[81\]](#page-12-20). Although these fndings are still in the preclinical phase, they hold exciting promise for the use of FMT from young donors, or its therapeutic components, to reverse certain aspects of unhealthy aging.

# **Diet and Probiotics**

As discussed previously, elderly populations often have variations in diet as they age, which contribute to microbiome changes. One of the most studied dietary changes associated with aging is reduced fber intake; however clinical trials supplementing fber have yielded conficting results regarding shifts in microbiota composition and infammatory status [\[90,](#page-13-1) [91\]](#page-13-2), with some researchers hypothesizing that the efficacy of dietary interventions and supplements may depend on the host's initial microbiome profle. For example, in a double-blind, crossover trial of 21 healthy volunteers over 60 years old who were given supplemental wheat branderived arabinoxylan-oligosaccharide found that resulting microbiome compositions varied based on subjects' initial *Prevotella* abundance [[92](#page-13-3)]. Although limited, these findings suggest that an individualized approach is required to manipulate the microbiome, with screening of patients' initial microbiomes necessary to tailor the intervention needed for the desired outcome.

In addition to specifc supplements, certain diets have been associated with gut health. The Mediterranean diet, consisting of plant-based foods, whole grains, and healthy fats, has been shown to prevent cardiovascular disease in all age ranges [[93\]](#page-13-4), with the efects of this diet potentially mediated by the gut microbiome. For example, a 2020 study by Ghosh et al. found that adherence to the Mediterranean diet for at least one year corresponded to a relative increase in intestinal *F. prausnitzii*, *R. hominis*, *E. rectale*, *E. eligens*, *E. xylanophilum*, *B. thetaiotaomicron*, *P. copri* and *A. hadrus* [[48](#page-11-14)••]*.* Adherence to the diet furthermore correlated with improved cognitive function, as measured by the BabCock Memory Score and Constructional Praxis, as well as decreased systemic infammatory markers such as highsensitivity C reactive protein (hsCRP) and interleukin 17 (IL-17) levels. Mouse studies have also demonstrated that a Western diet, which is high in fat and sodium, leads to an increased "predicted age" of the gut microbiome based on a Bayesian model trained on male C57BL/6 J mice whose microbiomes were characterized from week 9 to week 112 of life. These microbiome disturbances reversed once the mice returned to a standard diet [[94\]](#page-13-5). Interventional diet studies evaluating both the gut microbiota and clinical outcomes in elderly, human cohorts are therefore of particular interest given these individuals' susceptibility to cognitive decline and unhealthy aging.

Probiotic interventions have been specifcally studied in the aged. Unfortunately, similar to studies in the general population, the generation of clinically actionable data has been dampened by the great heterogeneity of studied products and outcomes as well as the multitude of underpowered studies [[95\]](#page-13-6). While no singular or combination of probiotic organisms have been identifed to defnitively improve or reverse signs of aging [[96\]](#page-13-7), a growing number of studies are evaluating specifc microbial strains and their impact on objective physiological efects. For example, in a double blind, placebo controlled study, *L. reuteri* ATCC PTA 6475 supplementation in elderly women with low bone mineral density improved tibia total volumetric BMD (vBMD) [[97,](#page-13-8) [98](#page-13-9)]. Furthermore, in the Senescence Accelerated Mouse-Prone 8 (SAMP8) mouse model, probiotic *Lactobacillus casei Shirota* administration reduced age-related muscle deterioration



<span id="page-7-0"></span>

 $\overline{1}$ 

 $\overline{1}$ 

and mitochondrial dysfunction [[99\]](#page-13-10). In humans, some small, but double-blinded randomized controlled trials have identifed specifc probiotics that appear to improve cognitive function in elderly adults, especially probiotics that include *Bifdobacterium* and *Lactobacillus spp.* [\[49,](#page-11-15) [50\]](#page-11-16). Thus, as a more rigorous understanding between microbiome manipulation and objective health measures develops, probiotic therapies may entail customized cocktails of microorganisms to target specifc defciencies or conditions in a personalized approach to care.

## **Exercise**

Multiple studies have reported an alteration in the gut microbiome following the implementation of an exercise program [[100\]](#page-13-11), with early results suggesting that this is true in elderly populations as well [\[101–](#page-13-12)[103\]](#page-13-13). A 2020 study by Zhu et al. utilized fecal specimens from the American Gut Project, which also included patient-reported information on BMI and exercise habits [[102\]](#page-13-14). The study included samples from 1,589 adults (aged 18–60 years) with a healthy BMI (18.5  $\leq$  BMI  $\leq$  25) and 897 elderly patients (aged  $>$  60), who were further stratifed by BMI into normal weight  $(n=462)$ , overweight (BMI > 25,  $n=413$ ) and underweight  $(BMI < 18.5, n = 22)$ , as well as by exercise frequency. Investigators found that as the reported frequency of exercise increased in elderly patients, the microbiome of the elderly patients more closely resembled that of the healthy BMI adults based on the relative abundance of specifc taxa and common pathways. For example, the relative abundance of *Actinobacteria* in exercising elderly adults increased compared to non-exercising elderly adults and approached the levels seen in healthy BMI adults. Furthermore, the relative abundance of *Cyanobacteria*, decreased in exercising elderly patients, again approaching levels seen in healthy BMI adults. (Of note, however, *Cyanobacteria* produce toxins such as β-N-Methylamino-l-alanine (BMAA) have been implicated in neurodegenerative diseases such as AD and ALS [[104,](#page-13-15) [105\]](#page-13-16).) In a smaller study by Erlandson et al., 15 sedentary elderly patients (aged 50–75) were recruited for a supervised 24 week, thrice-weekly cardiovascular and resistance exercise program. Stool samples were collected before and after the intervention for 16S sequencing [\[103](#page-13-13)]. Researchers observed an increased relative abundance of *Bifdobacterium* after 24 weeks of the exercise program, as well as increased butyrate levels. Considering the speculated role of *Bifdobacterium* in extreme aging and improved cognitive function, these fndings suggest that the health benefts related to exercise may also be mediated through the gut microbiome.

Despite these results, there is signifcant interpersonal variation in the reported microbiome changes that occur with exercise [\[106\]](#page-13-17). Additionally, many of the current studies do



not have a control arm, lack rigor, and/or have small sample sizes. Future studies are needed to identify if there is in fact a relationship between exercise and healthy aging microbiota, as well as the type of physical activity that can infuence gut microbiomes.

# **Conclusions**

Microbiome research in the elderly is an exciting, rapidly growing feld; however, a major gap in the literature is the lack of longitudinal data by which to distinguish between causative and correlative relationships given the many concomitant changes that occur with age, including altered dentition, diet, sleep, and lifestyle patterns. A challenge in interpreting currently available data is the diference in sequencing methodologies utilized. For instance, 16S study results can difer depending on the portion of the variable region within the 16S gene that is being sequenced  $[107]$  $[107]$ , as well as due to variation in 16S copy number between bacterial species [[107,](#page-13-20) [108](#page-13-21)]. Comparing 16S data across studies is also challenging as the results provide only the relative abundances of taxa in a group [\[109\]](#page-13-22); thus, abundances of one group may appear to change but only because of changes in the abundance of other taxa  $[110]$  $[110]$  $[110]$ . Furthermore, as taxonomy does not necessarily inform microbial function [[111\]](#page-13-24), it is likely that a future shift in focus to metagenomic function may better clarify the mechanisms by which gut microbes infuence their host.

Additional challenges specifc to clinical microbiome studies in the elderly and the extremely elderly include diffculties with mobility, the tendency for increased medical co-morbidities, as well as difficulties determining capacity for decision-making, and fnding a proxy for subjects who may not be able to provide their own consent to participate in research.

Nonetheless, early fndings suggest that there is the potential to reverse microbiome aging with interventions such as FMT, exercise, and dietary modifcations. Several large, longitudinal cohort studies are now underway that aim to characterize microbiome changes throughout aging, including the MOTION study and a new branch of the Wisconsin Longitudinal study (WLS) [[47,](#page-11-13) [112\]](#page-13-25). As mentioned previously, the MOTION study is a longitudinal, prospective cohort study of 360 healthy individuals over the age of 60, that is specifcally interested in the relationship between cognitive function and gut microbiome in elderly populations [[47](#page-11-13)]. The WLS, a longitudinal study of one-third of the Wisconsin high-school graduates in 1957, recently incorporated a microbiota branch to the project through the collection of 429 stool specimens (74% response rate), which will analyze the gut microbiome as it relates to environmental conditions and disease development [\[112](#page-13-25)].

As the feld of medicine becomes more individualized with the growth of genetics, epigenetics, and other biomarkers, we must also consider the importance of a unique microbiome profle in diagnosis and treatment. A common theme throughout much of the research is the signifcance of individualized care, with treatments based on the initial host microbiome composition. Although many of these studies are still in the early stages and require additional evidence to confrm a true causative relationship between illness and dysbiosis, elucidation of a unique microbiome disease profle opens the door to new avenues of treatment for these diseases.

**Acknowledgements** Akshata Shukla assisted with manuscript review.

**Author Contributions** LA.C. and K.B. wrote the main manuscript text and prepared accompanying fgures and tables.

**Funding** No funding was received to assist with the preparation of this manuscript.

**Data Availability** No datasets were generated or analysed during the current study.

#### **Declarations**

**Competing Interests** The authors declare no competing interests.

**Human and Animal Rights and Informed Consent** This article does not contain any studies with human or animal subjects performed by any of the authors.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit<http://creativecommons.org/licenses/by/4.0/>.

## **References**

Papers of particular interest, published recently, have been highlighted as:

- Of importance
- •• Of major importance
- <span id="page-9-0"></span>1. Zheng D, Liwinski T, Elinav E. Interaction between microbiota and immunity in health and disease. Cell Res. 2020;30(6):492– 506.<https://doi.org/10.1038/s41422-020-0332-7>.
- <span id="page-9-1"></span>2. Ma Y, Xu X, Li M, Cai J, Wei Q, Niu H. Gut microbiota promote the infammatory response in the pathogenesis of systemic

lupus erythematosus. Mol Med. 2019;25(1):35. [https://doi.org/](https://doi.org/10.1186/s10020-019-0102-5) [10.1186/s10020-019-0102-5](https://doi.org/10.1186/s10020-019-0102-5).

- <span id="page-10-0"></span>3. Chassaing B, Darfeuille-Michaud A. The commensal microbiota and enteropathogens in the pathogenesis of infammatory bowel diseases. Gastroenterology. 2011;140(6):1720–8. [https://doi.org/](https://doi.org/10.1053/j.gastro.2011.01.054) [10.1053/j.gastro.2011.01.054.](https://doi.org/10.1053/j.gastro.2011.01.054)
- <span id="page-10-1"></span>4. Pedersen HK, Gudmundsdottir V, Nielsen HB, Hyotylainen T, Nielsen T, Jensen BA, et al. Human gut microbes impact host serum metabolome and insulin sensitivity. Nature. 2016;535(7612):376–81. [https://doi.org/10.1038/nature18646.](https://doi.org/10.1038/nature18646)
- <span id="page-10-2"></span>5. Zimmermann M, Zimmermann-Kogadeeva M, Wegmann R, Goodman AL. Mapping human microbiome drug metabolism by gut bacteria and their genes. Nature. 2019;570(7762):462–7. [https://doi.org/10.1038/s41586-019-1291-3.](https://doi.org/10.1038/s41586-019-1291-3)
- <span id="page-10-3"></span>6. Kang DW, Adams JB, Gregory AC, Borody T, Chittick L, Fasano A, et al. Microbiota transfer therapy alters gut ecosystem and improves gastrointestinal and autism symptoms: an openlabel study. Microbiome. 2017;5(1):10. [https://doi.org/10.1186/](https://doi.org/10.1186/s40168-016-0225-7) [s40168-016-0225-7.](https://doi.org/10.1186/s40168-016-0225-7)
- <span id="page-10-4"></span>7. Jacobs JP, Gupta A, Bhatt RR, Brawer J, Gao K, Tillisch K, et al. Cognitive behavioral therapy for irritable bowel syndrome induces bidirectional alterations in the brain-gut-microbiome axis associated with gastrointestinal symptom improvement. Microbiome. 2021;9(1):236. [https://doi.org/10.1186/](https://doi.org/10.1186/s40168-021-01188-6) [s40168-021-01188-6](https://doi.org/10.1186/s40168-021-01188-6).
- <span id="page-10-5"></span>Lloyd-Price J, Abu-Ali G, Huttenhower C. The healthy human microbiome. Genome Med. 2016;8(1):51. [https://doi.org/10.](https://doi.org/10.1186/s13073-016-0307-y) [1186/s13073-016-0307-y.](https://doi.org/10.1186/s13073-016-0307-y)
- <span id="page-10-6"></span>9. Rinninella E, Cintoni M, Raoul P, Lopetuso LR, Scaldaferri F, Pulcini G, et al. Food components and dietary habits: keys for a healthy gut microbiota composition. Nutrients. 2019;11(10). <https://doi.org/10.3390/nu11102393>.
- <span id="page-10-7"></span>10. Hall AB, Tolonen AC, Xavier RJ. Human genetic variation and the gut microbiome in disease. Nat Rev Genet. 2017;18(11):690– 9. <https://doi.org/10.1038/nrg.2017.63>.
- <span id="page-10-8"></span>11. Ahn J, Hayes RB. Environmental Infuences on the Human Microbiome and Implications for Noncommunicable Disease. Annu Rev Public Health. 2021;42:277–92. [https://doi.org/10.](https://doi.org/10.1146/annurev-publhealth-012420-105020) [1146/annurev-publhealth-012420-105020](https://doi.org/10.1146/annurev-publhealth-012420-105020).
- <span id="page-10-9"></span>12. Song SJ, Wang J, Martino C, Jiang L, Thompson WK, Shenhav L, et al. Naturalization of the microbiota developmental trajectory of Cesarean-born neonates after vaginal seeding. Med (N Y). 2021;2(8):951-64.e5. [https://doi.org/10.1016/j.medj.2021.](https://doi.org/10.1016/j.medj.2021.05.003) [05.003.](https://doi.org/10.1016/j.medj.2021.05.003)
- <span id="page-10-10"></span>13. Azad MB, Konya T, Maughan H, Guttman DS, Field CJ, Chari RS, et al. Gut microbiota of healthy Canadian infants: profles by mode of delivery and infant diet at 4 months. CMAJ. 2013;185(5):385–94. [https://doi.org/10.1503/cmaj.121189.](https://doi.org/10.1503/cmaj.121189)
- <span id="page-10-11"></span>14. Dominguez-Bello MG, Godoy-Vitorino F, Knight R, Blaser MJ. Role of the microbiome in human development. Gut. 2019;68(6):1108–14. [https://doi.org/10.1136/](https://doi.org/10.1136/gutjnl-2018-317503) [gutjnl-2018-317503.](https://doi.org/10.1136/gutjnl-2018-317503)
- <span id="page-10-12"></span>15. Affairs UNDoEaS. Leaving No One Behind In An Ageing World. World Social Report. 2023;2023:17–34. [https://doi.org/](https://doi.org/10.18356/9789210019682) [10.18356/9789210019682](https://doi.org/10.18356/9789210019682).
- <span id="page-10-13"></span>16. Jefery IB, Lynch DB, O'Toole PW. Composition and temporal stability of the gut microbiota in older persons. Isme J. 2016;10(1):170–82. [https://doi.org/10.1038/ismej.2015.88.](https://doi.org/10.1038/ismej.2015.88)
- <span id="page-10-14"></span>17 Ghosh TS, Das M, Jefery IB, O'Toole PW. Adjusting for age improves identifcation of gut microbiome alterations in multiple diseases. eLife. 2020;9:e50240. [https://doi.org/10.7554/eLife.](https://doi.org/10.7554/eLife.50240) [50240](https://doi.org/10.7554/eLife.50240).
- <span id="page-10-15"></span>18. Biagi E, Franceschi C, Rampelli S, Severgnini M, Ostan R, Turroni S, et al. Gut microbiota and extreme longevity. Curr Biol. 2016;26(11):1480–5. [https://doi.org/10.1016/j.cub.2016.04.016.](https://doi.org/10.1016/j.cub.2016.04.016)
- 
- <span id="page-10-16"></span>19. Cani PD, Depommier C, Derrien M, Everard A, de Vos WM. Akkermansia muciniphila: paradigm for next-generation beneficial microorganisms. Nat Rev Gastroenterol Hepatol. 2022;19(10):625–37. [https://doi.org/10.1038/](https://doi.org/10.1038/s41575-022-00631-9) [s41575-022-00631-9](https://doi.org/10.1038/s41575-022-00631-9).
- <span id="page-10-17"></span>20. Everard A, Belzer C, Geurts L, Ouwerkerk JP, Druart C, Bindels LB, et al. Cross-talk between Akkermansia muciniphila and intestinal epithelium controls diet-induced obesity. Proc Natl Acad Sci U S A. 2013;110(22):9066–71. [https://doi.org/10.1073/](https://doi.org/10.1073/pnas.1219451110) [pnas.1219451110.](https://doi.org/10.1073/pnas.1219451110)
- <span id="page-10-18"></span>21.•• Wilmanski T, Diener C, Rappaport N, Patwardhan S, Wiedrick J, Lapidus J, et al. Gut microbiome pattern refects healthy ageing and predicts survival in humans. Nat Metab. 2021;3(2):274–86. [https://doi.org/10.1038/s42255-021-00348-0.](https://doi.org/10.1038/s42255-021-00348-0) **Microbiome uniqueness, as measured by Bray-Curtis dissimilarity, is positively correlated with age.**
- <span id="page-10-19"></span>22. Peel C, Sawyer Baker P, Roth DL, Brown CJ, Brodner EV, Allman RM. Assessing mobility in older adults: the UAB Study of Aging Life-Space Assessment. Phys Ther. 2005;85(10):1008–119.
- <span id="page-10-20"></span>23. Pang S, Chen X, Lu Z, Meng L, Huang Y, Yu X, et al. Longevity of centenarians is refected by the gut microbiome with youthassociated signatures. Nat Aging. 2023. [https://doi.org/10.1038/](https://doi.org/10.1038/s43587-023-00389-y) [s43587-023-00389-y.](https://doi.org/10.1038/s43587-023-00389-y)
- <span id="page-10-21"></span>24.• Johansen J, Atarashi K, Arai Y, Hirose N, Sørensen SJ, Vatanen T, et al. Centenarians have a diverse gut virome with the potential to modulate metabolism and promote healthy lifespan. Nat Microbiol. 2023;8(6):1064–78. [https://doi.org/10.1038/s41564-](https://doi.org/10.1038/s41564-023-01370-6) [023-01370-6.](https://doi.org/10.1038/s41564-023-01370-6) **Early work suggests that like the microbiome, the centenarian virome may also have distinct features compared to aging adults such as increased diveristy and unique genera**.
- <span id="page-10-22"></span>25. Walford RL. The immunologic theory of aging1. Gerontologist. 1964;4(4):195–7.<https://doi.org/10.1093/geront/4.4.195>.
- <span id="page-10-23"></span>26. Franceschi C, BonafÈ M, Valensin S, Olivieri F, De Luca M, Ottaviani E, et al. Infamm-aging: an evolutionary perspective on immunosenescence. Ann N Y Acad Sci. 2000;908(1):244–54. <https://doi.org/10.1111/j.1749-6632.2000.tb06651.x>.
- <span id="page-10-24"></span>27. Jaiswal S, Libby P. Clonal haematopoiesis: connecting ageing and infammation in cardiovascular disease. Nat Rev Cardiol. 2020;17(3):137–44. [https://doi.org/10.1038/s41569-019-0247-5.](https://doi.org/10.1038/s41569-019-0247-5)
- 28. Zhao TV, Sato Y, Goronzy JJ, Weyand CM. T-Cell agingassociated phenotypes in autoimmune disease. Front Aging. 2022;3:867950.<https://doi.org/10.3389/fragi.2022.867950>.
- <span id="page-10-25"></span>29. Goronzy JJ, Weyand CM. Mechanisms underlying T cell ageing. Nat Rev Immunol. 2019;19(9):573–83. [https://doi.org/10.1038/](https://doi.org/10.1038/s41577-019-0180-1) [s41577-019-0180-1.](https://doi.org/10.1038/s41577-019-0180-1)
- <span id="page-10-26"></span>30. Choi W, Yeruva S, Turner JR. Contributions of intestinal epithelial barriers to health and disease. Exp Cell Res. 2017;358(1):71– 7. [https://doi.org/10.1016/j.yexcr.2017.03.036.](https://doi.org/10.1016/j.yexcr.2017.03.036)
- <span id="page-10-27"></span>31. Sovran B, Hugenholtz F, Elderman M, Van Beek AA, Graversen K, Huijskes M, et al. Age-associated impairment of the mucus barrier function is associated with profound changes in microbiota and immunity. Sci Rep. 2019;9(1):1437. [https://doi.org/10.](https://doi.org/10.1038/s41598-018-35228-3) [1038/s41598-018-35228-3.](https://doi.org/10.1038/s41598-018-35228-3)
- <span id="page-10-28"></span>32. Pabst O, Herbrand H, Friedrichsen M, Velaga S, Dorsch M, Berhardt G, et al. Adaptation of solitary intestinal lymphoid tissue in response to microbiota and chemokine receptor CCR7 signaling. J Immunol. 2006;177(10):6824–32. [https://doi.org/10.4049/](https://doi.org/10.4049/jimmunol.177.10.6824) [jimmunol.177.10.6824.](https://doi.org/10.4049/jimmunol.177.10.6824)
- <span id="page-10-29"></span>33. Thevaranjan N, Puchta A, Schulz C, Naidoo A, Szamosi JC, Verschoor CP, et al. Age-associated microbial dysbiosis promotes intestinal permeability, systemic infammation, and macrophage dysfunction. Cell Host Microbe. 2017;21(4):455-66.e4. [https://](https://doi.org/10.1016/j.chom.2017.03.002) [doi.org/10.1016/j.chom.2017.03.002.](https://doi.org/10.1016/j.chom.2017.03.002)
- <span id="page-11-0"></span>34. Claesson MJ, Jefery IB, Conde S, Power SE, O'Connor EM, Cusack S, et al. Gut microbiota composition correlates with diet and health in the elderly. Nature. 2012;488(7410):178–84. <https://doi.org/10.1038/nature11319>.
- <span id="page-11-1"></span>35. Asempa TE, Nicolau DP. Clostridium difficile infection in the elderly: an update on management. Clin Interv Aging. 2017;12:1799–809.<https://doi.org/10.2147/cia.S149089>.
- <span id="page-11-2"></span>36. Wastyk HC, Fragiadakis GK, Perelman D, Dahan D, Merrill BD, Yu FB, et al. Gut-microbiota-targeted diets modulate human immune status. Cell. 2021;184(16):4137-53.e14. [https://doi.org/](https://doi.org/10.1016/j.cell.2021.06.019) [10.1016/j.cell.2021.06.019.](https://doi.org/10.1016/j.cell.2021.06.019)
- <span id="page-11-3"></span>37. Wu GD, Chen J, Hofmann C, Bittinger K, Chen YY, Keilbaugh SA, et al. Linking long-term dietary patterns with gut microbial enterotypes. Science. 2011;334(6052):105–8. [https://doi.org/10.](https://doi.org/10.1126/science.1208344) [1126/science.1208344](https://doi.org/10.1126/science.1208344).
- <span id="page-11-4"></span>38. Wu GD, Compher C, Chen EZ, Smith SA, Shah RD, Bittinger K, et al. Comparative metabolomics in vegans and omnivores reveal constraints on diet-dependent gut microbiota metabolite production. Gut. 2016;65(1):63–72. [https://doi.org/10.1136/](https://doi.org/10.1136/gutjnl-2014-308209) [gutjnl-2014-308209.](https://doi.org/10.1136/gutjnl-2014-308209)
- <span id="page-11-5"></span>39. Tanes C, Bittinger K, Gao Y, Friedman ES, Nessel L, Paladhi UR, et al. Role of dietary fber in the recovery of the human gut microbiome and its metabolome. Cell Host Microbe. 2021;29(3):394-407.e5. [https://doi.org/10.1016/j.chom.2020.](https://doi.org/10.1016/j.chom.2020.12.012) [12.012.](https://doi.org/10.1016/j.chom.2020.12.012)
- <span id="page-11-6"></span>40. Clemente JC, Pehrsson EC, Blaser MJ, Sandhu K, Gao Z, Wang B, et al. The microbiome of uncontacted Amerindians. Sci Adv. 2015;1(3). [https://doi.org/10.1126/sciadv.1500183.](https://doi.org/10.1126/sciadv.1500183)
- <span id="page-11-7"></span>41 Salazar N, Valdés-Varela L, González S, Gueimonde M, de los Reyes-Gavilán CG. Nutrition and the gut microbiome in the elderly. Gut Microbes. 2017;8(2):82–97. [https://doi.org/10.1080/](https://doi.org/10.1080/19490976.2016.1256525) [19490976.2016.1256525.](https://doi.org/10.1080/19490976.2016.1256525)
- <span id="page-11-8"></span>42. Ragonnaud E, Biragyn A. Gut microbiota as the key controllers of "healthy" aging of elderly people. Immun Ageing. 2021;18(1):2. [https://doi.org/10.1186/s12979-020-00213-w.](https://doi.org/10.1186/s12979-020-00213-w)
- <span id="page-11-9"></span>43. Petersen RC, Lopez O, Armstrong MJ, Getchius TSD, Ganguli M, Gloss D, et al. Practice guideline update summary: mild cognitive impairment: report of the guideline development, dissemination, and implementation subcommittee of the American academy of neurology. Neurology. 2018;90(3):126–35. [https://](https://doi.org/10.1212/wnl.0000000000004826) [doi.org/10.1212/wnl.0000000000004826.](https://doi.org/10.1212/wnl.0000000000004826)
- <span id="page-11-10"></span>44 Russ TC, Morling JR. Cholinesterase inhibitors for mild cognitive impairment. Cochrane Database Syst Rev. 2012;2012(9):Cd009132. [https://doi.org/10.1002/14651858.](https://doi.org/10.1002/14651858.CD009132.pub2) [CD009132.pub2.](https://doi.org/10.1002/14651858.CD009132.pub2)
- <span id="page-11-11"></span>45. van Soest APM, Hermes GDA, Berendsen AAM, van de Rest O, Zoetendal EG, Fuentes S, et al. Associations between Pro- and Anti-Infammatory gastro-intestinal microbiota, diet, and cognitive functioning in Dutch Healthy older adults: The NU-AGE study. Nutrients. 2020;12(11). [https://doi.org/10.3390/nu121](https://doi.org/10.3390/nu12113471) [13471](https://doi.org/10.3390/nu12113471).
- <span id="page-11-12"></span>46. Fam J, Sun Y, Qi P, Lau RC, Feng L, Kua EH, et al. Mindfulness practice alters brain connectivity in community-living elders with mild cognitive impairment. Psychiatr Clin Neurosci. 2020;74(4):257–62. [https://doi.org/10.1111/pcn.12972.](https://doi.org/10.1111/pcn.12972)
- <span id="page-11-13"></span>47. Phillips S, Watt R, Atkinson T, Rajan S, Hayhoe A, Savva GM, et al. A protocol paper for the MOTION Study-A longitudinal study in a cohort aged 60 years and older to obtain mechanistic knowledge of the role of the gut microbiome during normal healthy ageing in order to develop strategies that will improve lifelong health and wellbeing. PLoS ONE. 2022;17(11):e0276118. [https://doi.org/10.1371/journal.pone.](https://doi.org/10.1371/journal.pone.0276118) [0276118](https://doi.org/10.1371/journal.pone.0276118).
- <span id="page-11-14"></span>48.•• Ghosh TS, Rampelli S, Jefery IB, Santoro A, Neto M, Capri M, et al. Mediterranean diet intervention alters the gut microbiome in older people reducing frailty and improving health status: the

NU-AGE 1-year dietary intervention across fve European countries. Gut. 2020;69(7):1218–28. [https://doi.org/10.1136/gutjnl-](https://doi.org/10.1136/gutjnl-2019-319654)[2019-319654.](https://doi.org/10.1136/gutjnl-2019-319654) **Elderly patient adherence to a Mediterranean diet for one year led to a decrease in frailty and infammation, with an increase in cognitive function**.

- <span id="page-11-15"></span>49. Kim C-S, Cha L, Sim M, Jung S, Chun WY, Baik HW, et al. Probiotic supplementation improves cognitive function and mood with changes in gut microbiota in community-dwelling older adults: a randomized, double-blind, placebo-controlled, multicenter trial. J Gerontol: Ser A. 2021;76(1):32–40. [https://](https://doi.org/10.1093/gerona/glaa090) [doi.org/10.1093/gerona/glaa090](https://doi.org/10.1093/gerona/glaa090).
- <span id="page-11-16"></span>50. Hwang Y-H, Park S, Paik J-W, Chae S-W, Kim D-H, Jeong D-G, et al. Efficacy and safety of lactobacillus plantarum c29-fermented soybean (DW2009) in individuals with mild cognitive impairment: A 12-week, multi-center, randomized, double-blind, placebo-controlled clinical trial. Nutrients. 2019. [https://doi.org/](https://doi.org/10.3390/nu11020305) [10.3390/nu11020305](https://doi.org/10.3390/nu11020305).
- <span id="page-11-17"></span>51. Asaoka D, Xiao J, Takeda T, Yanagisawa N, Yamazaki T, Matsubara Y, et al. Efect of Probiotic Bifdobacterium breve in improving cognitive function and preventing brain atrophy in older patients with suspected mild cognitive impairment: results of a 24-week randomized, double-blind. Placebo-Controlled Trial J Alzheimers Dis. 2022;88(1):75–95. [https://doi.org/10.](https://doi.org/10.3233/jad-220148) [3233/jad-220148.](https://doi.org/10.3233/jad-220148)
- <span id="page-11-18"></span>52. Komanduri M, Savage K, Lea A, McPhee G, Nolidin K, Deleuil S, et al. The relationship between gut microbiome and cognition in older Australians. Nutrients. 2021;14(1). [https://doi.org/10.](https://doi.org/10.3390/nu14010064) [3390/nu14010064.](https://doi.org/10.3390/nu14010064)
- <span id="page-11-19"></span>53. Pan Q, Li YQ, Guo K, Xue M, Gan Y, Wang K, et al. Elderly patients with mild cognitive impairment exhibit altered gut microbiota profles. J Immunol Res. 2021;2021:5578958. [https://](https://doi.org/10.1155/2021/5578958) [doi.org/10.1155/2021/5578958.](https://doi.org/10.1155/2021/5578958)
- <span id="page-11-20"></span>54. Zhang X, Wang Y, Liu W, Wang T, Wang L, Hao L, et al. Diet quality, gut microbiota, and microRNAs associated with mild cognitive impairment in middle-aged and elderly Chinese population. Am J Clin Nutr. 2021;114(2):429–40. [https://doi.org/10.](https://doi.org/10.1093/ajcn/nqab078) [1093/ajcn/nqab078.](https://doi.org/10.1093/ajcn/nqab078)
- <span id="page-11-21"></span>55. Li B, He Y, Ma J, Huang P, Du J, Cao L, et al. Mild cognitive impairment has similar alterations as Alzheimer's disease in gut microbiota. Alzheimers Dement. 2019;15(10):1357–66. [https://](https://doi.org/10.1016/j.jalz.2019.07.002) [doi.org/10.1016/j.jalz.2019.07.002.](https://doi.org/10.1016/j.jalz.2019.07.002)
- <span id="page-11-22"></span>56. Park SH, Lee JH, Kim JS, Kim TJ, Shin J, Im JH, et al. Fecal microbiota transplantation can improve cognition in patients with cognitive decline and Clostridioides difficile infection. Aging (Albany NY). 2022;14(16):6449–66. [https://doi.org/10.](https://doi.org/10.18632/aging.204230) [18632/aging.204230.](https://doi.org/10.18632/aging.204230)
- <span id="page-11-23"></span>57. Liu P, Wu L, Peng G, Han Y, Tang R, Ge J, et al. Altered microbiomes distinguish Alzheimer's disease from amnestic mild cognitive impairment and health in a Chinese cohort. Brain Behav Immun. 2019;80:633–43. [https://doi.org/10.1016/j.bbi.2019.05.](https://doi.org/10.1016/j.bbi.2019.05.008) [008.](https://doi.org/10.1016/j.bbi.2019.05.008)
- <span id="page-11-24"></span>58. Ueda A, Shinkai S, Shiroma H, Taniguchi Y, Tsuchida S, Kariya T, et al. Identifcation of Faecalibacterium prausnitzii strains for gut microbiome-based intervention in Alzheimer's-type dementia. Cell Rep Med. 2021;2(9):100398. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.xcrm.2021.100398) [xcrm.2021.100398.](https://doi.org/10.1016/j.xcrm.2021.100398)
- <span id="page-11-25"></span>59. Lee J, Venna VR, Durgan DJ, Shi H, Hudobenko J, Putluri N, et al. Young versus aged microbiota transplants to germ-free mice: increased short-chain fatty acids and improved cognitive performance. Gut Microbes. 2020;12(1):1–14. [https://doi.org/](https://doi.org/10.1080/19490976.2020.1814107) [10.1080/19490976.2020.1814107](https://doi.org/10.1080/19490976.2020.1814107).
- <span id="page-11-26"></span>60. Yang X, Yu D, Xue L, Li H, Du J. Probiotics modulate the microbiota-gut-brain axis and improve memory deficits in aged SAMP8 mice. Acta Pharm Sin B. 2020;10(3):475–87. [https://](https://doi.org/10.1016/j.apsb.2019.07.001) [doi.org/10.1016/j.apsb.2019.07.001.](https://doi.org/10.1016/j.apsb.2019.07.001)
- <span id="page-12-0"></span>61. Haran JP, Bhattarai SK, Foley SE, Dutta P, Ward DV, Bucci V, et al. Alzheimer's disease microbiome is associated with dysregulation of the anti-infammatory P-Glycoprotein Pathway. mBio. 2019;10(3). <https://doi.org/10.1128/mBio.00632-19>.
- <span id="page-12-1"></span>62. Hung CC, Chang CC, Huang CW, Nouchi R, Cheng CH. Gut microbiota in patients with Alzheimer's disease spectrum: a systematic review and meta-analysis. Aging (Albany NY). 2022;14(1):477–96. <https://doi.org/10.18632/aging.203826>.
- <span id="page-12-2"></span>63. Westfall S, Dinh DM, Pasinetti GM. Investigation of potential brain microbiome in Alzheimer's disease: implications of study bias. J Alzheimers Dis. 2020;75(2):559–70. [https://doi.org/10.](https://doi.org/10.3233/jad-191328) [3233/jad-191328.](https://doi.org/10.3233/jad-191328)
- <span id="page-12-3"></span>64.• Cammann D, Lu Y, Cummings MJ, Zhang ML, Cue JM, Do J, et al. Genetic correlations between Alzheimer's disease and gut microbiome genera. Sci Rep. 2023;13(1):5258. [https://doi.org/](https://doi.org/10.1038/s41598-023-31730-5) [10.1038/s41598-023-31730-5.](https://doi.org/10.1038/s41598-023-31730-5) **Distinct microbiome profles are associated with high-risk alleles of the apolipoprotein E ε4 (APOE ε4) gene, a well-established risk factor for AD.**
- <span id="page-12-4"></span>65. Nishiwaki H, Ito M, Ishida T, Hamaguchi T, Maeda T, Kashihara K, et al. Meta-analysis of gut dysbiosis in Parkinson's disease. Mov Disord. 2020;35(9):1626–35. [https://doi.org/10.1002/mds.](https://doi.org/10.1002/mds.28119) [28119](https://doi.org/10.1002/mds.28119).
- <span id="page-12-5"></span>66.• Wallen ZD, Demirkan A, Twa G, Cohen G, Dean MN, Standaert DG, et al. Metagenomics of Parkinson's disease implicates the gut microbiome in multiple disease mechanisms. Nat Commun. 2022;13(1):6958. [https://doi.org/10.1038/s41467-022-34667-x.](https://doi.org/10.1038/s41467-022-34667-x) **Researchers identifed altered microbiome signatures in PD patients, such as an increase in pathogenic** *Prevotella* **species.**
- <span id="page-12-6"></span>67. Zhang N, Zhang Y, Wang Z, Pan F, Ren R, Li Z, et al. Regular fecal microbiota transplantation to Senescence Accelerated Mouse-Prone 8 (SAMP8) mice delayed the aging of locomotor and exploration ability by rejuvenating the gut microbiota. Front Aging Neurosci. 2022;14. [https://doi.org/10.3389/fnagi.](https://doi.org/10.3389/fnagi.2022.991157) [2022.991157](https://doi.org/10.3389/fnagi.2022.991157).
- <span id="page-12-7"></span>68. Cao H, Liu X, An Y, Zhou G, Liu Y, Xu M, et al. Dysbiosis contributes to chronic constipation development via regulation of serotonin transporter in the intestine. Sci Rep. 2017;7(1):10322. <https://doi.org/10.1038/s41598-017-10835-8>.
- <span id="page-12-8"></span>69. Sampson TR, Debelius JW, Thron T, Janssen S, Shastri GG, Ilhan ZE, et al. Gut microbiota regulate motor defcits and neuroinfammation in a model of parkinson's disease. Cell. 2016;167(6):1469-80.e12. [https://doi.org/10.1016/j.cell.2016.](https://doi.org/10.1016/j.cell.2016.11.018) [11.018.](https://doi.org/10.1016/j.cell.2016.11.018)
- <span id="page-12-9"></span>70.• DuPont HL, Suescun J, Jiang ZD, Brown EL, Essigmann HT, Alexander AS, et al. Fecal microbiota transplantation in Parkinson's disease-A randomized repeat-dose, placebo-controlled clinical pilot study. Front Neurol. 2023;14:1104759. [https://doi.](https://doi.org/10.3389/fneur.2023.1104759) [org/10.3389/fneur.2023.1104759.](https://doi.org/10.3389/fneur.2023.1104759) **Early research suggest FMT could decrease symptoms in PD patients.**
- <span id="page-12-10"></span>71. Picca A, Ponziani FR, Calvani R, Marini F, Biancolillo A, Coelho-Júnior HJ, et al. Gut microbial, infammatory and metabolic signatures in older people with physical frailty and sarcopenia: results from the BIOSPHERE Study. Nutrients. 2020. [https://](https://doi.org/10.3390/nu12010065) [doi.org/10.3390/nu12010065](https://doi.org/10.3390/nu12010065).
- <span id="page-12-11"></span>72. Almeida HM, Sardeli AV, Conway J, Duggal NA, Cavaglieri CR. Comparison between frail and non-frail older adults' gut microbiota: A systematic review and meta-analysis. Ageing Res Rev. 2022;82:101773. [https://doi.org/10.1016/j.arr.2022.101773.](https://doi.org/10.1016/j.arr.2022.101773)
- <span id="page-12-12"></span>73. Quach D, Collins F, Parameswaran N, McCabe L, Britton RA. Microbiota reconstitution does not cause bone loss in Germ-Free Mice. mSphere. 2018;3(1). [https://doi.org/10.1128/mSphereDir](https://doi.org/10.1128/mSphereDirect.00545-17) [ect.00545-17](https://doi.org/10.1128/mSphereDirect.00545-17).
- <span id="page-12-13"></span>74. Yan J, Herzog JW, Tsang K, Brennan CA, Bower MA, Garrett WS, et al. Gut microbiota induce IGF-1 and promote bone formation and growth. Proc Natl Acad Sci U S A.

2016;113(47):E7554–63. [https://doi.org/10.1073/pnas.16072](https://doi.org/10.1073/pnas.1607235113) [35113.](https://doi.org/10.1073/pnas.1607235113)

- <span id="page-12-14"></span>75. Das M, Cronin O, Keohane DM, Cormac EM, Nugent H, Nugent M, et al. Gut microbiota alterations associated with reduced bone mineral density in older adults. Rheumatology. 2019;58(12):2295–304. [https://doi.org/10.1093/rheumatology/](https://doi.org/10.1093/rheumatology/kez302) [kez302.](https://doi.org/10.1093/rheumatology/kez302)
- <span id="page-12-15"></span>76. Kelly CR, Yen EF, Grinspan AM, Kahn SA, Atreja A, Lewis JD, et al. Fecal microbiota transplantation is highly efective in real-world practice: initial results from the FMT national registry. Gastroenterology. 2021;160(1):183-92.e3. [https://doi.org/](https://doi.org/10.1053/j.gastro.2020.09.038) [10.1053/j.gastro.2020.09.038.](https://doi.org/10.1053/j.gastro.2020.09.038)
- <span id="page-12-16"></span>77 Kedia S, Virmani S, Vuyyuru SK, Kumar P, Kante B, Sahu P, et al. Faecal microbiota transplantation with anti-infammatory diet (FMT-AID) followed by anti-infammatory diet alone is efective in inducing and maintaining remission over 1 year in mild to moderate ulcerative colitis: a randomised controlled trial. Gut. 2022;71(12):2401. [https://doi.org/10.1136/](https://doi.org/10.1136/gutjnl-2022-327811) [gutjnl-2022-327811.](https://doi.org/10.1136/gutjnl-2022-327811)
- <span id="page-12-17"></span>78. Suez J, Zmora N, Zilberman-Schapira G, Mor U, Dori-Bachash M, Bashiardes S, et al. Post-antibiotic gut mucosal microbiome reconstitution is impaired by probiotics and improved by Autologous FMT. Cell. 2018;174(6):1406-23.e16. [https://doi.](https://doi.org/10.1016/j.cell.2018.08.047) [org/10.1016/j.cell.2018.08.047.](https://doi.org/10.1016/j.cell.2018.08.047)
- <span id="page-12-18"></span>79.• Parker A, Romano S, Ansorge R, Aboelnour A, Le Gall G, Savva GM, et al. Fecal microbiota transfer between young and aged mice reverses hallmarks of the aging gut, eye, and brain. Microbiome. 2022;10(1):68. [https://doi.org/10.1186/s40168-](https://doi.org/10.1186/s40168-022-01243-w) [022-01243-w](https://doi.org/10.1186/s40168-022-01243-w). **Animal models have demonstrated that the physiological efects of young and aging microbiomes on cognitive function are transferable.**
- <span id="page-12-19"></span>80. D'Amato A, Di Cesare ML, Lucarini E, Man AL, Le Gall G, Branca JJV, et al. Faecal microbiota transplant from aged donor mice afects spatial learning and memory via modulating hippocampal synaptic plasticity- and neurotransmission-related proteins in young recipients. Microbiome. 2020;8(1):140.<https://doi.org/10.1186/s40168-020-00914-w>.
- <span id="page-12-20"></span>81. Bárcena C, Valdés-Mas R, Mayoral P, Garabaya C, Durand S, Rodríguez F, et al. Healthspan and lifespan extension by fecal microbiota transplantation into progeroid mice. Nat Med. 2019;25(8):1234–42. [https://doi.org/10.1038/](https://doi.org/10.1038/s41591-019-0504-5) [s41591-019-0504-5.](https://doi.org/10.1038/s41591-019-0504-5)
- <span id="page-12-21"></span>82. Xu L, Zhang Q, Dou X, Wang Y, Wang J, Zhou Y, et al. Fecal microbiota transplantation from young donor mice improves ovarian function in aged mice. J Genet Genomics. 2022;49(11):1042–52. [https://doi.org/10.1016/j.jgg.2022.05.](https://doi.org/10.1016/j.jgg.2022.05.006) [006](https://doi.org/10.1016/j.jgg.2022.05.006).
- <span id="page-12-22"></span>83. Kim KH, Chung Y, Huh JW, Park DJ, Cho Y, Oh Y, et al. Gut microbiota of the young ameliorates physical ftness of the aged in mice. Microbiome. 2022;10(1):238. [https://doi.org/](https://doi.org/10.1186/s40168-022-01386-w) [10.1186/s40168-022-01386-w.](https://doi.org/10.1186/s40168-022-01386-w)
- <span id="page-12-23"></span>84. Valeri F, Dos Santos Guilherme M, He F, Stoye NM, Schwiertz A, Endres K. Impact of the age of cecal material transfer donors on Alzheimer's disease pathology in 5xFAD Mice. Microorganisms. 2021;9(12). [https://doi.org/10.3390/micro](https://doi.org/10.3390/microorganisms9122548) [organisms9122548](https://doi.org/10.3390/microorganisms9122548).
- <span id="page-12-24"></span>85. Stebegg M, Silva-Cayetano A, Innocentin S, Jenkins TP, Cantacessi C, Gilbert C, et al. Heterochronic faecal transplantation boosts gut germinal centres in aged mice. Nat Commun. 2019;10(1):2443. [https://](https://doi.org/10.1038/s41467-019-10430-7) doi. org/ 10. 1038/ [s41467-019-10430-7](https://doi.org/10.1038/s41467-019-10430-7).
- <span id="page-12-25"></span>86. Zeng X, Li X, Li X, Wei C, Shi C, Hu K, et al. Fecal microbiota transplantation from young mice rejuvenates aged hematopoietic stem cells by suppressing infammation. Blood. 2023. [https://doi.](https://doi.org/10.1182/blood.2022017514) [org/10.1182/blood.2022017514.](https://doi.org/10.1182/blood.2022017514)
- <span id="page-13-18"></span>87. Chen Y, Zhang S, Zeng B, et al. Transplant of microbiota from long-living people to mice reduces aging-related indices and transfers beneficial bacteria. Aging (Albany NY). 2020;12(6):4778–93. <https://doi.org/10.18632/aging.102872>.
- <span id="page-13-19"></span>88. Hu C, Liu M, Sun B, Tang L, Zhou X, Chen L. Young fecal transplantation mitigates the toxicity of perfuorobutanesulfonate and potently refreshes the reproductive endocrine system in aged recipients. Environ Int. 2022;167:107418. [https://doi.org/10.](https://doi.org/10.1016/j.envint.2022.107418) [1016/j.envint.2022.107418.](https://doi.org/10.1016/j.envint.2022.107418)
- <span id="page-13-0"></span>89. Talukder P, Saha A, Roy S, Ghosh G, Dutta Roy D, Barua S. Progeria—a rare genetic condition with accelerated ageing process. Appl Biochem Biotechnol. 2023;195(4):2587–96. [https://](https://doi.org/10.1007/s12010-021-03514-y) [doi.org/10.1007/s12010-021-03514-y.](https://doi.org/10.1007/s12010-021-03514-y)
- <span id="page-13-1"></span>90. Ganda Mall J-P, Fart F, Sabet JA, Lindqvist CM, Nestestog R, Hegge FT, et al. Effects of dietary fibres on acute indomethacininduced intestinal hyperpermeability in the elderly: a randomised placebo controlled parallel clinical trial. Nutrients. 2020. [https://](https://doi.org/10.3390/nu12071954) [doi.org/10.3390/nu12071954](https://doi.org/10.3390/nu12071954).
- <span id="page-13-2"></span>91. Tran TTT, Cousin FJ, Lynch DB, Menon R, Brulc J, Brown JR, et al. Prebiotic supplementation in frail older people afects specifc gut microbiota taxa but not global diversity. Microbiome. 2019;7(1):39.<https://doi.org/10.1186/s40168-019-0654-1>.
- <span id="page-13-3"></span>92. Chung WSF, Walker AW, Bosscher D, Garcia-Campayo V, Wagner J, Parkhill J, et al. Relative abundance of the Prevotella genus within the human gut microbiota of elderly volunteers determines the inter-individual responses to dietary supplementation with wheat bran arabinoxylan-oligosaccharides. BMC Microbiol. 2020;20(1):283. [https://doi.org/10.1186/](https://doi.org/10.1186/s12866-020-01968-4) [s12866-020-01968-4](https://doi.org/10.1186/s12866-020-01968-4).
- <span id="page-13-4"></span>93. Delgado-Lista J, Alcala-Diaz JF, Torres-Peña JD, Quintana-Navarro GM, Fuentes F, Garcia-Rios A, et al. Long-term secondary prevention of cardiovascular disease with a Mediterranean diet and a low-fat diet (CORDIOPREV): a randomised controlled trial. Lancet. 2022;399(10338):1876–85. [https://doi.org/10.](https://doi.org/10.1016/S0140-6736(22)00122-2) [1016/S0140-6736\(22\)00122-2.](https://doi.org/10.1016/S0140-6736(22)00122-2)
- <span id="page-13-5"></span>94. Low A, Soh M, Miyake S, Seedorf H. Host age prediction from fecal microbiota composition in male C57BL/6J Mice. Microbiol Spectr. 2022;10(3):e0073522. [https://doi.org/10.1128/spectrum.](https://doi.org/10.1128/spectrum.00735-22) [00735-22.](https://doi.org/10.1128/spectrum.00735-22)
- <span id="page-13-6"></span>95. Chen LA, Sears CL. 3 - Prebiotics, Probiotics, and Synbiotics. In: Bennett JE, Dolin R, Blaser MJ, editors. Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases. 8th ed. Philadelphia: W.B. Saunders; 2015. p. 19- 25.e1.
- <span id="page-13-7"></span>96. Chenhuichen C, Cabello-Olmo M, Barajas M, Izquierdo M, Ramírez-Vélez R, Zambom-Ferraresi F, et al. Impact of probiotics and prebiotics in the modulation of the major events of the aging process: A systematic review of randomized controlled trials. Exp Gerontol. 2022;164:111809. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.exger.2022.111809) [exger.2022.111809.](https://doi.org/10.1016/j.exger.2022.111809)
- <span id="page-13-8"></span>97 Li P, Ji B, Luo H, Sundh D, Lorentzon M, Nielsen J. Oneyear supplementation with Lactobacillus reuteri ATCC PTA 6475 counteracts a degradation of gut microbiota in older women with low bone mineral density. npj Biofilms Microbiomes. 2022;8(1):84. [https://doi.org/10.1038/](https://doi.org/10.1038/s41522-022-00348-2) [s41522-022-00348-2.](https://doi.org/10.1038/s41522-022-00348-2)
- <span id="page-13-9"></span>98. Nilsson AG, Sundh D, Bäckhed F, Lorentzon M. Lactobacillus reuteri reduces bone loss in older women with low bone mineral density: a randomized, placebo-controlled, double-blind, clinical trial. J Intern Med. 2018;284(3):307–17. [https://doi.org/10.1111/](https://doi.org/10.1111/joim.12805) [joim.12805](https://doi.org/10.1111/joim.12805).
- <span id="page-13-10"></span>99. Chen LH, Chang SS, Chang HY, Wu CH, Pan CH, Chang CC, et al. Probiotic supplementation attenuates age-related

sarcopenia via the gut-muscle axis in SAMP8 mice. J Cachexia Sarcopenia Muscle. 2022;13(1):515–31. [https://doi.org/10.1002/](https://doi.org/10.1002/jcsm.12849) [jcsm.12849.](https://doi.org/10.1002/jcsm.12849)

- <span id="page-13-11"></span>100. Dalton A, Mermier C, Zuhl M. Exercise infuence on the microbiome-gut-brain axis. Gut Microbes. 2019;10(5):555–68. [https://](https://doi.org/10.1080/19490976.2018.1562268) [doi.org/10.1080/19490976.2018.1562268](https://doi.org/10.1080/19490976.2018.1562268).
- <span id="page-13-12"></span>101. Zhong F, Wen X, Yang M, Lai HY, Momma H, Cheng L, et al. Efect of an 8-week exercise training on gut microbiota in physically inactive older women. Int J Sports Med. 2021;42(7):610– 23.<https://doi.org/10.1055/a-1301-7011>.
- <span id="page-13-14"></span>102. Zhu Q, Jiang S, Du G. Efects of exercise frequency on the gut microbiota in elderly individuals. MicrobiologyOpen. 2020;9(8):e1053. [https://doi.org/10.1002/mbo3.1053.](https://doi.org/10.1002/mbo3.1053)
- <span id="page-13-13"></span>103. Erlandson KM, Liu J, Johnson R, Dillon S, Jankowski CM, Kroehl M, et al. An exercise intervention alters stool microbiota and metabolites among older, sedentary adults. Ther Adv Infect Dis. 2021;8:20499361211027068. [https://doi.org/10.1177/20499](https://doi.org/10.1177/20499361211027067) [361211027067.](https://doi.org/10.1177/20499361211027067)
- <span id="page-13-15"></span>104. Cox PA, Kostrzewa RM, Guillemin GJ. BMAA and Neurodegenerative Illness. Neurotox Res. 2018;33(1):178–83. [https://](https://doi.org/10.1007/s12640-017-9753-6) [doi.org/10.1007/s12640-017-9753-6](https://doi.org/10.1007/s12640-017-9753-6).
- <span id="page-13-16"></span>105. Silva DF, Candeias E, Esteves AR, Magalhães JD, Ferreira IL, Nunes-Costa D, et al. Microbial BMAA elicits mitochondrial dysfunction, innate immunity activation, and Alzheimer's disease features in cortical neurons. J Neuroinflammation. 2020;17(1):332. <https://doi.org/10.1186/s12974-020-02004-y>.
- <span id="page-13-17"></span>106. Ramos C, Gibson GR, Walton GE, Magistro D, Kinnear W, Hunter K. Systematic review of the effects of exercise and physical activity on the gut microbiome of older adults. Nutrients. 2022;14(3). [https://doi.org/10.3390/nu14030674.](https://doi.org/10.3390/nu14030674)
- <span id="page-13-20"></span>107. Abellan-Schneyder I, Matchado MS, Reitmeier S, Sommer A, Sewald Z, Baumbach J, et al. Primer, Pipelines, Parameters: Issues in 16S rRNA Gene Sequencing. mSphere. 2021;6(1). [https://doi.org/10.1128/mSphere.01202-20.](https://doi.org/10.1128/mSphere.01202-20)
- <span id="page-13-21"></span>108. Kembel SW, Wu M, Eisen JA, Green JL. Incorporating 16S gene copy number information improves estimates of microbial diversity and abundance. PLoS Comput Biol. 2012;8(10):e1002743. <https://doi.org/10.1371/journal.pcbi.1002743>.
- <span id="page-13-22"></span>109. Jo J-H, Kennedy EA, Kong HH. Research techniques made simple: bacterial 16s ribosomal RNA gene sequencing in cutaneous research. J Investig Dermatol. 2016;136(3):e23–7. [https://doi.](https://doi.org/10.1016/j.jid.2016.01.005) [org/10.1016/j.jid.2016.01.005.](https://doi.org/10.1016/j.jid.2016.01.005)
- <span id="page-13-23"></span>110. Weiss S, Xu ZZ, Peddada S, Amir A, Bittinger K, Gonzalez A, et al. Normalization and microbial diferential abundance strategies depend upon data characteristics. Microbiome. 2017;5(1):27. [https://doi.org/10.1186/s40168-017-0237-y.](https://doi.org/10.1186/s40168-017-0237-y)
- <span id="page-13-24"></span>111. Inkpen SA, Douglas GM, Brunet TDP, Leuschen K, Doolittle WF, Langille MGI. The coupling of taxonomy and function in microbiomes. Biol Philos. 2017;32(6):1225–43. [https://doi.org/](https://doi.org/10.1007/s10539-017-9602-2) [10.1007/s10539-017-9602-2](https://doi.org/10.1007/s10539-017-9602-2).
- <span id="page-13-25"></span>112. Herd P, Schaeffer NC, DiLoreto K, Jacques K, Stevenson J, Rey F, et al. The infuence of social conditions across the life course on the human gut microbiota: a pilot project with the wisconsin longitudinal study. J Gerontol B Psychol Sci Soc Sci. 2017;73(1):124–33. [https://doi.org/10.1093/geronb/gbx029.](https://doi.org/10.1093/geronb/gbx029)

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional afliations.