MICROVASCULAR COMPLICATIONS-NEPHROPATHY (T ISAKOVA, SECTION EDITOR)

Genetics of Diabetic Nephropathy: a Long Road of Discovery

Amy Jayne McKnight¹ · Seamus Duffy¹ · Alexander P. Maxwell^{1,2}

Published online: 14 May 2015 © Springer Science+Business Media New York 2015

Abstract The global prevalence of diabetic nephropathy is rising in parallel with the increasing incidence of diabetes in most countries. Unfortunately, up to 40 % of persons diagnosed with diabetes may develop kidney complications. Diabetic nephropathy is associated with substantially increased risks of cardiovascular disease and premature mortality. An inherited susceptibility to diabetic nephropathy exists, and progress is being made unravelling the genetic basis for nephropathy thanks to international research collaborations, shared biological resources and new analytical approaches. Multiple epidemiological studies have highlighted the clinical heterogeneity of nephropathy and the need for better phenotyping to help define important subgroups for analysis and increase the power of genetic studies. Collaborative genomewide association studies for nephropathy have reported unique genes, highlighted novel biological pathways and suggested new disease mechanisms, but progress towards clinically relevant risk prediction models for diabetic nephropathy has been slow. This review summarises the current status, recent

This article is part of the Topical Collection on *Microvascular Complications—Nephropathy*

 Alexander P. Maxwell a.p.maxwell@qub.ac.uk
 Amy Jayne McKnight

a.j.mcknight@qub.ac.uk

Seamus Duffy sduffy26@qub.ac.uk

¹ Nephrology Research Group, Centre for Public Health, Queen's University Belfast, c/o Regional Genetics Centre, Level A, Tower Block, Belfast City Hospital, Lisburn Road, Belfast BT9 7AB, UK

² Regional Nephrology Unit, Belfast City Hospital, Belfast BT9 7AB, UK developments and ongoing challenges elucidating the genetics of diabetic nephropathy.

Keywords Albuminuria · Association · Diabetic nephropathy · Genetics · GWAS · Meta-analysis · SNP

Introduction

Diabetes is a significant public health problem, placing increasing human and financial pressures on already overburdened healthcare systems. Six percent of the UK population have been diagnosed with diabetes [1, 2]. Diabetes now accounts for 10 % of the UK National Health Service budget, with 80 % of those costs spent managing secondary complications, such as blindness, amputation, heart disease, stroke and kidney disease, which may be potentially preventable or have their onset delayed with earlier management of the risk factors for these disorders [3, 4]. In the USA, during 2012, it was reported that 11.8 % of adults were living with diabetes resulting in estimated costs of \$245 billion from a combination of lost productivity and direct healthcare expenditure; these costs are 41 % higher than those estimated in 2007 [5, 6]. In the UK, in 2013, diabetic nephropathy accounted for over 25 % of the incident patients with endstage renal disease (ESRD) [7] whereas in the USA, in 2012, over 40 % of incident patients needing dialysis had diabetic nephropathy [8]. In several countries including Malaysia, South Korea and Mexico, the incidence of diabetic nephropathy causing ESRD exceeds 50 % [9]. The global population of individuals living with ESRD is increasing steadily; in the USA, one estimate indicates that the ESRD population may exceed two million by 2030 [10]. Among populations with chronic kidney disease, the risk of ESRD and premature mortality is higher for individuals diagnosed

with diabetes [11, 12]. The increasing global prevalence of diabetes, combined with the fact that up to 40 % of affected individuals will develop kidney complications [13], is a major incentive to develop tools for earlier diagnosis of diabetic kidney disease, to improve prediction models and to identify novel therapeutic targets. Key priorities include strategies to reduce the incidence of diabetic nephropathy and development of treatments to minimise progression to ESRD.

Clinical Epidemiology of Diabetic Nephropathy

Recent research has provided a number of challenges to our understanding of the natural history of 'diabetic nephropathy' [14, 15••]. The classical description of diabetic nephropathy was developed in the 1980s based on clinical observations in longitudinal studies of individuals with type 1 diabetes. The earliest clinical indication of nephropathy was moderately increased albuminuria (usually referred to as 'microalbuminuria') with the later development of persistent and severely increased albuminuria (often described as 'macroalbuminuria') with peak incidence after approximately 15 years duration of diabetes followed by decline in glomerular filtration rate (GFR) and progression to ESRD [16]. Diabetic nephropathy was associated with a poor prognosis, and the majority of deaths were due to cardiovascular disease or ESRD [17–19].

Earlier identification and treatment of diabetic nephropathy proved possible by repeated and sensitive measurements of the urinary albumin excretion rate [20] (used as a marker of diabetic kidney injury) [16, 21]. Although screening for microalbuminuria contributed to improved clinical practice, permitting earlier and more aggressive treatment of diabetic nephropathy, its use as a screening tool is confounded by the fact that microalbuminuria may spontaneously regress in many patients with diabetes [17, 18]. Microalbuminuria therefore does not always predict future risk of kidney failure, and this limits its utility as a biomarker for diabetic nephropathy [22, 23]. Nonetheless, microalbuminuria remains a powerful predictor for the future risk of cardiovascular complications [22].

Defining the clinical phenotype of diabetic nephropathy in type 1 diabetes has become more problematic. For instance, persistent and severely increased albuminuria may accompany rather than precede the fall in GFR whereas in other patients with type 1 diabetes, the associated proteinuria may actually regress despite progressive kidney failure [24]. Of interest, some patients with type 1 diabetes and a low GFR, but no proteinuria, may still have typical pathological features of diabetic nephropathy on renal biopsy [25].

Accurately defining clinical phenotypes remains a crucial starting point for studies of the genetics of diabetic nephropathy. Arguably, the natural history of diabetic nephropathy is easier to study in persons with type 1 diabetes versus type 2 diabetes because the age at onset of diabetes is more accurately determined in type 1 diabetes. There is considerable debate as to whether the underlying genetic or pathological mechanisms responsible for diabetic nephropathy in type 1 diabetes versus type 2 diabetes overlap or are distinct [14, 26, 27].

Studying the genetic susceptibility to diabetic nephropathy in persons with type 2 diabetes is particularly challenging since the clinical phenotype is more difficult to define. For individuals with type 2 diabetes, it cannot be assumed that proteinuria and a low GFR indicate the presence of diabetic nephropathy. Renal biopsy studies have highlighted the broad range of renal pathologies present in persons with type 2 diabetes [28]. The discordance between type 2 diabetes and diabetic nephropathy, as a cause of ESRD, was further emphasised in a recent national registry study from Scotland which reported that only 58 % of individuals with ESRD and type 2 diabetes had a diagnosis of diabetic nephropathy [29]. This contrasted with the data from the same registry which indicated that 91 % of individuals with type 1 diabetes and ESRD had diabetic nephropathy. Clinical phenotyping is further complicated by current analyses suggesting that different gene variants contribute to the risks for proteinuria and ESRD in both type 1 and type 2 diabetes [30, 31], with a little overlap observed between genes identified for individual measurements of renal function such as urinary albumin:creatinine ratio (uACR), serum creatinine and serum cystatin C [32, 33, 34••].

Genetic Epidemiology

Several lines of evidence support an inherited genetic predisposition to diabetic nephropathy: only a subset of individuals with type 1 or type 2 diabetes will develop diabetic nephropathy [15••], diabetic nephropathy and ESRD both cluster in families [35-37], and the prevalence of diabetic nephropathy varies between ethnic groups [10, 38]. The risk of developing diabetic nephropathy can be reduced but not eliminated by improved control of known risk factors such as hypertension and poor glycaemic control [39-41]. The genetic component for diabetic nephropathy (heritability) has been estimated between 0.2 and 0.46 [42-45], with one notable study in White individuals with type 2 diabetes reporting an estimated glomerular filtration rate (eGFR) heritability (estimated h^2) of 0.75 after adjusting for age, gender, mean arterial blood pressure, medication and HbA1c [44]. Modifiable, traditional risk factors for diabetic nephropathy include blood pressure, glycaemic control, lipid levels, chronic inflammation, smoking, weight and physical exercise, and it should be highlighted that many of these modifiable risk factors are also influenced by a person's genetic profile [46]. Other risk factors for diabetic nephropathy are not modifiable such as age, gender, age at onset and duration of diabetes, but these may still

influence the future risk of developing diabetic kidney disease, e.g. via gender-specific genetic mechanisms [34••] or longer term epigenetic reprogramming of gene expression associated with age and duration of diabetes [47–49].

Candidate Genes In common with many multifactorial diseases, early genetic studies for diabetic nephropathy focused on candidate genes that had biologically plausible roles in the pathogenesis of this disease. Many genes and singlenucleotide polymorphisms (SNPs) were reported to be significantly associated with diabetic nephropathy; however despite, 'best practice' experiments in the 1980s and 1990s, few of these genetic associations were supported by independent replication [50, 51]. Candidate gene studies remain important and are being published, although typically with more comprehensive analysis of biological and/or positional candidate genes, including non-coding regions with putative regulatory functions. Improved efforts have been made to try and identify associated genes that influence diabetic nephropathy with these studies incorporating more stringent quality control, larger samples sizes, discovery with multiple replication cohorts, matched cases and controls, consideration of relevant covariates and ideally genome-wide significance values. Meta-analyses may also help confirm or refute genetic association findings, but these are often challenging to undertake with different statistical tests performed between studies, multiethnic cohorts, genetic and phenotypic heterogeneity, inability to contact authors for primary data and insufficient information reported in publications. More than 200 meta-analyses have been published for diabetic nephropathy, but these often generate conflicting results and are predominantly composed of smaller studies where it is challenging to standardise quality control across all participating studies. Sizeable meta-analyses published in the last 5 years have been recently reviewed in depth, revealing only one gene associated with diabetic nephropathy where P < 0.0001 from targeted studies [50]; the functional SNP rs1617640 in the promoter region of the erythropoietin (EPO) gene, located on chromosome 7q21-q22, was associated with both proliferative diabetic retinopathy and ESRD in multiple populations with diabetes [52–54].

Linkage Studies Taking a genome-wide approach, multiple linkage studies were conducted using multigenerational families or discordant sib pairs to try and localise genetic risk factors for diabetic nephropathy to specific chromosome regions. These microsatellite and SNP-based linkage studies have been previously reviewed, with combined analysis revealing that every autosome (any chromosome that is not a sex or mitochondrial chromosome) has been highlighted with a kidney-related phenotype [51, 55, 56], although the evidence for robust linkage is typically low. Commonly reported genetic regions include the following: 3q13-26, 7p, 6q22-27, 10p11-15, 15q21, 16p11-13 (UMOD), 18q22 (CNDP1, CNDP2), 20q11, 22q

(MYH9) [50]. A PubMed search for [diabetic AND (nephropathy or kidney) AND linkage], conducted on 24 January 2015 returns two linkage studies in the past 5 years, both of which involved the multiethnic, multicentre, American Family Investigation of Nephropathy and Diabetes (FIND) collection. In 2011, a genome-wide linkage scan for diabetic nephropathy and uACR was conducted using approximately 4400 autosomal SNPs from Illumina's Linkage IVb panel in each African-American, American-Indian, European-American and Mexican-American group [57]. Not unexpectedly, results were inconsistent across all ethnicities, but evidence for linkage with diabetic nephropathy, where logarithm of odds (LOD)>2.5, was observed at chromosome 6p24.3 (LOD 2.84) for European Americans and 7p21.3 for American-Indians (LOD 2.81) [57]. Evidence of linkage with uACR was observed at chromosome region 7q21.2 for European-Americans (LOD 2.96) and 3p13 for African-Americans (LOD 2.76) [57]. A subsequent publication evaluated linkage for eGFR in this population using the same linkage IVb panel, revealing linkage with chromosome 20q11 (LOD 3.34) in Mexican-Americans and 15q12 (LOD 2.84) in European-Americans [58].

Genome-Wide Association Studies Genome-wide association studies (GWAS) have had a pivotal role identifying SNPs associated with common complex diseases such as diabetes [59, 60], cancer [61, 62] and Alzheimer's disease [63]. They are relatively cost-effective, readily amenable to automation, technically easy to perform in a high-throughput manner, and software has been developed to facilitate combining data sets genotyped on different platforms from multiple centres. The primary advantage of GWAS is their flexibility to systematically screen common variants across the genome with no prior biological assumptions, although many GWAS arrays now provide an option to add selected SNPs and rare variants to the panel for no or low extra cost. There are a range of arrays available to perform GWAS, with Illumina's most comprehensive (January 2015) HumanOmni5Exome array providing simultaneous analysis of up to five million SNPs with minor allele frequency >1 % and including exonic variants identified from >12,000 sequenced exomes. A more costeffective option for large-scale population-based genotyping projects is one of the smaller arrays, such as Illumina's customisable HumanCoreExome-24 BeadChip, which analyses more than half a million carefully selected SNPs, including 265,919 exome-focused markers. Affymetrix Axiom genotyping arrays also offer competitively priced arrays [64]. Exploiting linkage disequilibrium for efficiently tagged SNPs by subsequent imputation will provide information on more markers and help compare genotype-phenotype data sets across different centres. Carefully designed studies and reporting genetic association results in line with STREGA guidelines [65], and standardised GWAS quality control [66], help improve transparency, interpretation of results and

inform downstream studies (Fig. 1). GWAS have proved successful identifying SNPs associated with kidney phenotypes including IgA nephropathy [67, 68], membranous nephropathy [69], focal segmental glomerulosclerosis (FSGS) [70, 71], chronic kidney disease (CKD) [32, 72, 73] and ESRD [74, 75••]. However, progress identifying susceptibility genes from GWAS for diabetic nephropathy has been slow.

Multiple GWAS have been performed exploring risk factors for kidney disease in populations with type 2 diabetes, but there has been less enthusiasm compared to type 1 diabetes to perform these studies, largely due to challenges identifying a 'true' diabetic nephropathy clinical phenotype (for cases) and appropriate controls despite the much larger number of individuals diagnosed with type 2 diabetes compared to type 1 diabetes (Table 1). Association studies have been combined with the linkage studies previously described, but the genes involved have yet to be identified in the broad, localised chromosome regions [58]. The first large-scale GWAS for diabetic nephropathy was conducted in Japanese individuals with type 2 diabetes in 2005; the *ELMO1* gene was identified (P= 000008, odds ratio 2.67, 95 % CI 1.71–4.16) from evaluation

of 80,000 gene-based SNPs [76]. There is functional support for *ELMO1* associated with diabetic nephropathy, but subsequent studies have generated inconsistent results and a comprehensive meta-analysis has not yet been published for this gene [54, 77–81]. Subsequent studies have highlighted *PVT1*, *LIMK2*, *SFI1*, *WFS1*, *FTO*, *KCNJ11* and *TCF7L2* genes, but none approached conventional genome-wide significance [82–84].

The first GWAS for individuals with type 1 diabetes followed the GAMES approach [85] using microsatellites and multiple DNA case-control pools to explore association with diabetic nephropathy [86]. Several genetic regions were highlighted in this low-resolution screen, but no marker provided strong evidence of association [86]. Two GWAS were published in 2009 reporting associations with *FRMD3*, *CARS*, *CHN2*, *CPVL*, *ZMIZ1* and *MSC* genes, although none reached genome-wide significance and replication has proved challenging [87, 88]. Suggestive trends towards association for *FRMD3* have been supported by independent groups [54, 89] and an *in silico* functional mechanism of action proposed through which a *FRMD3* promoter polymorphism influences

Fig. 1 Genetic association designs to investigate diabetic nephropathy for either a targeted approach or genome-wide association studies. These study designs are employed to discover genetic risk factors for diabetic nephropathy including discrete traits associated with kidney disease, e.g. proteinuria, rate of decline in eGFR. DN diabetic nephropathy, eGFR estimated glomerular filtration rate, eOTL [134] expression quantitative trait loci, ESRD end-stage renal disease, HaploReg [135] a tool for exploring annotations of the noncoding genome, QC quality control, SNP single-nucleotide polymorphism, SrCr serum creatinine, srCysC serum cystatin C, uACR urinary albumin:creatinine ratio

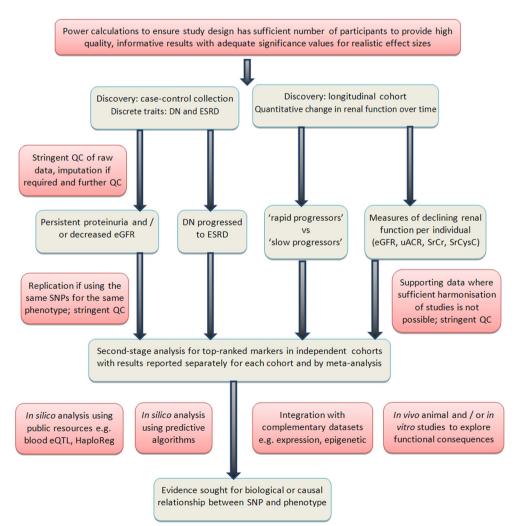


 Table 1
 Key genes identified by GWAS and meta-analysis demonstrating association with diabetic nephropathy

Gene	Locus	Evidence for association	Functional role
AFF3 (AF4/FMR2 family, member 3)	2q11	Meta-analysis of GWAS in European population with T1DM identified association with ESRD and rs7583877 (OR 1.29, 95 % CI 1.18–1.40, $P=1.2 \times 10^{-8}$) [75••].	Codes for a nuclear transcriptional activator. Functional analysis suggests $AFF3$ may play a role in TGF- β 1 induced renal tubule fibrosis [75••].
<i>COX6A1</i> (cytochrome c oxidase subunit VIa polypeptide 1)	12q24	European populations with T1DM revealed association for rs12310837 with ESRD (P=0.000002) and DN $(P=0.00003)[102].$	<i>COX6A1</i> codes for a subunit of cytochrome c oxidase, and this may increase oxidative stress within the kidney [102].
<i>ELMO1</i> (engulfment and cell motility 1)	7p14.1	 GWAS for Japanese patients with T2DM <i>intron 18+9170</i> (OR 2.67, 95 % CI 1.71–4.16, P=0.000008) associated with DN [76]. rs741301 (OR 1.89, P=0.004) and rs10951509 (OR 1.76, P=0.02) associated with DN in Chinese population with T2DM [80]. Meta-analysis of Asian subgroup showed rs741301 associated with T2DM (OR 1.58, 95 % CI 1.28–1.94) [136]. No significant association in meta-analysis of European populations for any SNP within 20 kb of <i>ELMO1</i> [54]. 	Involved in phagocytosis and cellular motility. Increased expression in kidneys of diabetic mice compared to control mice [76]. Overexpressed in cells cultured in high glucose conditions. Increased <i>ELMO1</i> expression may have a possible role in progression of glomerulosclerosis [76].
EPO (erythropoietin)	7q21-q22	rs1617640 associated with DN in meta-analysed T1DM & T2DM populations (OR 1.47, 95 % CI 1.31–1.65) [52]. Meta-analysis of European populations with T1DM showed significant association with DN (OR 1.31, $P=2\times10^{-9}$) [54].	Encodes erythropoietin, a hormone expressed in the kidney with roles in erythropoiesis and angiogenesis. The SNP reported here has also been significantly associated with diabetic retinopathy and ESRD in three European-American cohorts [53]. Recombinant erythropoietin is associated with increased renal damage in rats [137].
<i>ERBB4</i> (erb-b2 receptor tyrosine kinase 4)	2q33.3-q34	Meta-analysis of GWAS in European population with T1DM identified association with DN and rs7588550 (OR 0.66, 95 % CI 0.56–0.77, $P=2.1\times10^{-7}$) [75••]. Japanese population with T2DM (OR 0.79, 95 % CI 0.65–0.95, $P=0.01$) [92].	
GLRA3 (glycine receptor, alpha 3)	4q34.1	GWAS in European population with T1DM rs10011025 (OR 0.21, 95 % CI 0.14–0.27, $P=1.5\times10^{-9}$) associated with uAER in discovery, rs11725853 from meta-analysis (OR 0.11, 95 % CI: 0.07–0.16, $P=7.9\times10^{-7}$ [33].	Encodes a glycine receptor subunit, which is involved in glucagon secretion [33].Low levels of mRNA expression are reported in both glomeruli and tubules [140] and in tubule-enriched kidney biopsies [141] of diabetic and non-diabetic subjects.
Intergenic	15q26	Meta-analysis of GWAS in European population with T1DM identified association with ESRD and rs12437854 (OR 1.8, 95 % CI 1.48–2.17, $P=2.0\times\times10^{-9}$) [75••].	-
SORBS1 (sorbin and SH3 domain containing 1)	10q23.33	Multistage GWAS in population with T1DM in four populations. rs1326934 was associated with DN (OR=0.83, 95 % CI=0.72–0.96, P=0.009) [94].	May have a role in insulin signalling and resistance [142]. Differential expression in glomeruli of DN rats [143] and renal tubule overexpression observed in T2D patients compared with controls [94].

DN diabetic nephropathy, TIDM type 1 diabetes, T2DM type 2 diabetes, uAER urinary albumin excretion rate

transcriptional regulation of the bone morphogenetic protein (BMP) signalling pathway [90].

Using the same inclusion/exclusion criteria for diabetic nephropathy phenotype, the GEnetics of Nephropathy—an International Effort (GENIE) consortium performed two novel GWAS on independent collections from the UK and Ireland (UK-ROI) and Finland (FinnDiane) [75••]. These novel GWAS data were combined with GWAS data from the US-GoKinD collection, which was available through dbGAP [91]. All three GWAS underwent consistent quality control and imputation, with meta-analysis of GWAS so that approximately 2.4 million SNPs were evaluated in a total of 6691 individuals for diabetic nephropathy and ESRD [75..]. Selected SNPs that demonstrated preliminary evidence of association in the discovery phase were followed up in 5873 individuals who had similar phenotypic characteristic to the discovery cohorts. SNP rs7588550 in ERBB4 showed the most evidence for association with the primary phenotype of diabetic nephropathy, and this was supported by gene expression data and plausible biological relevance whereby ERBB4 is co-expressed with collagen genes associated with renal fibrosis in the tubulointerstitial compartment of the kidney, although has not yet been widely replicated [75..]. Two independent Japanese groups studying diabetic nephropathy in type 2 diabetes have reported nominal association with this SNP in ERBB4 gene [92], although in the opposite direction of effect to that observed in the GENIE GWAS. Considering the more extreme phenotype of ESRD, rs12437854 was observed with genome-wide significance $(P=2\times 10^{-9})$ [75...], but this SNP is located between RGMA and MCTP2 genes, with unknown function, albeit the association has been supported by additional statistical approaches [93]. Of particular interest were several SNPs in the AFF3 gene (rs7583877, $P=1.2\times$ 10^{-8} was the most significant), which were associated with ESRD, supported by functional data demonstrating increased gene expression and protein levels in cell models of kidney fibrosis, as well as being involved with the transforming growth factor beta pathway [75..]. A key strength of the effective GENIE consortium was use of harmonised clinical phenotypes that facilitated pooling resources to generate a larger discovery sample size with relatively extensive replication and active engagement by all teams.

The most recently published GWAS for diabetic nephropathy in type 1 diabetes used a discovery collection of 683 cases compared to 779 controls, with first-stage replication in US-GoKinD followed by second-stage replication in FinnDiane and UK-ROI collections [94]. Top-ranked SNPs following discovery and initial replication were in the *SORBS1* gene, although no association was observed in FinnDiane and a non-significant trend in the same direction as the discovery cohort was observed in the UK-ROI collection resulting in the most significant SNP from this metaanalysis rs1326934 C allele (P=0.009, odds ratio 0.83, 95 % CI 0.72–0.96) [94].

Sample size is critically important to ensure GWAS studies are adequately powered to identify risk alleles. For example, an early GWAS in 4921 individuals identified common variants in the *HMGA2* gene associated with human height. Metaanalysis of GWAS in 2008 identified twelve loci that explain approximately 2 % of phenotypic variation in height [95]. Just 2 years later, analysis of 183,727 individuals revealed 180 loci, which influence human height and explain ~10 % of population differences [96]. Most recently (November 2014), 697 variants clustered in 423 loci were identified at genome-wide significance by analysing GWAS data from 253,288 individuals; together, these common variants explain 60 % of the heritability for height [97]. To improve power in identifying variants influencing diabetic nephropathy, a larger JDRF Diabetic Nephropathy Collaborative Research Initiative is currently underway typing Illumina's HumanCoreExome array with imputation to 1000 genomes and meta-analysis of ~25,000 individuals with type 1 diabetes for association with diabetic nephropathy.

Extending Typical GWAS GWAS studies typically analyse autosomes, with the initial quality control steps excluding analysis of the sex chromosomes (X, Y) and the mitochondrial genome. Gender-specific differences are apparent for renal phenotypes, including an increased incidence and prevalence of diabetic nephropathy and ESRD in men [98]. This has led to gender-specific genetic association analyses being performed in men and women separately. Based on metaanalysis of GWAS data, a key SNP, rs4972593 on chromosome 2q31, was associated in women with an odds ratio of 1.81 (95 % confidence interval 1.47–2.24, $P=3.85\times10^{-8}$), yet showed no association in men (P=0.77), despite 99 % power to identify this association in men [34..]. The observation that chromosome Y is associated with coronary artery disease in British men [99], primarily connected to inflammatory and immunity genes, suggests sex chromosomes are worthy of investigation for diabetic nephropathy.

Mitochondrial dysfunction is evident for diabetic nephropathy [100, 101], with multiple SNPs in genes related to mitochondrial function recently observed to be associated with diabetic nephropathy based on 6819 individuals with type 1 diabetes [102]; of particular interest is the *COX6A1* gene located on chromosome 12q24, which was independently identified among top-ranked signals from independent genetic and methylation studies focused on mitochondrial-related genes [102, 103]. Next-generation sequencing of the mitochondrial genome has revealed genetic variants associated in an ESRD population [104], and this approach for the single, <17 kbp mitochondrial chromosome may identify further risk factors for diabetic nephropathy.

Omics Genetic variation does not function in isolation, and increasingly, researchers are combining multiomic data sets to identify and help explain risk factors for diabetic nephropathy [75••, 105, 106]. Gene expression studies are moving from targeted microarrays towards more detailed RNA-seq approaches [107], which provide very rich data sets that may be exploited in cell-based models, animal studies and human studies. Epigenetic analysis for diabetic nephropathy is increasingly being studied at a genome-wide level [50], with associations reported for post-translational chromatin modifications [108–110], non-protein-coding RNAs [111–113] and

DNA methylation features [47, 103, 114–116]. Metabolomic [117–119] and proteomic [120–122] profiles are revealing intriguing biomarker signatures, but a lack of standardisation in terms of subject recruitment, experimental platforms or analytical approaches makes such multicentre studies challenging. A standardised approach to integrate diverse data sets has not yet been established, but recent large-scale studies suggest that Data-driven Enrichment-Prioritized Integration for Complex Traits (DEPICT) outperformed GRAIL and MAGENTA when prioritising associated SNPs and finding the most likely causal gene(s) based on integrated data, which included extensive expression, protein interactome, reactome, gene ontology and pathway-based analyses [123–125].

Next-Generation Sequencing A complementary approach to increasing sample size is more comprehensive exploration of the genome, primarily to identify less common variants that may have relatively large effect sizes for diabetic nephropathy. Several years ago, the Wellcome Trust initiated a project to sequence 1000 genomes [126], and in 2014, a new UK project commenced, which plans to sequence 100,000 genomes by 2017 and integrate this data with NHS medical records [127]. The primary focus of the 100,000 genomes project by Genomics England is sequencing the genomes of patients with rare diseases and cancer to enhance research and help progress genomic medicine within the NHS; however, multiple groups are taking advantage of the low-cost whole-genome genotyping option to have population-based cohorts sequenced. These resources offer promising opportunities for productive diabetic nephropathy research, although challenges remain in terms of effectively managing the sheer volume of data and how to deal with medically actionable results from an individual's genome. To date, few next-generation sequencing projects have been conducted for diabetic nephropathy. Initial microarrays, focused on non-synonymous SNPs, did not generate exciting results for diabetic nephropathy [128, 129], but whole-exome sequencing approaches have enabled the identification of additional exonic SNPs and development of high-density exome arrays. Using publicly available whole-exome sequencing data, 31 coding SNPs across selected genes with prior evidence for association with kidney disease were genotyped to reveal exonic SNPs associated with ESRD in individuals with type 2 diabetes (P < 0.05) [130].

Future Directions

Large-scale epidemiological studies have underscored the need for more extensive clinical characterisation of kidney disease in individuals with diabetes to improve the precision of phenotyping for diabetic nephropathy and increase the power of genetic association studies. There is however a trade-off between the gains from precise phenotypes that minimise bioclinical complexity, and reduction in the potential sample sizes that fulfil inclusion criteria. There are three primary approaches by which this may be achieved: (1) recruitment of carefully phenotyped cohorts of individuals with diabetes with long-term follow-up for diabetic nephropathy; (2) utilising other case-control or longitudinal cohorts, which were not specifically designed to examine diabetic nephropathy, but collected information on relevant phenotypes including blood glucose levels, diabetes status and measures of renal function; and (3) large-scale population-based registers such as the 100,000 genomes project [127], the UK Biobank [131], Generation Scotland [132], the Health Retirement Study [133] and GERA cohort (dbGaP Study Accession: phs000674.v1.p1).

Conclusions

Significant progress has been made improving the clinical care of persons with diabetes to reduce an individual's personal risk of developing diabetic nephropathy. Nevertheless, the rising global burden of diabetes will continue to drive an increased incidence of diabetic nephropathy. Ideally, a combination of clinical characteristics, renal functional measurements and relevant biomarkers would permit a more accurate prediction of the risk of developing of nephropathy and its rate of progression. A key benefit of clinically useful, predictive genetic biomarkers is their potential to identify those individuals at highest (or lowest) risk of diabetic nephropathy before it is clinically apparent, enabling a stratified medicine approach. Cost-effective and individualised clinical care could then be directed to those individuals at the highest lifetime risk of diabetic nephropathy. To realise this ambitious goal, there is an urgent need to improve our understanding of the genetic architecture underlying diabetic nephropathy. An unanswered question for researchers is why some individuals with diabetes develop nephropathy whereas others are protected from this complication. The answers are within the complex and dynamic interactions between genomic risk factors, behavioural traits and environmental stressors.

Compliance with Ethics Guidelines

Conflict of Interest Amy Jayne McKnight, Seamus Duffy and Alexander P. Maxwell declare that they have no conflict of interest.

Human and Animal Rights and Informed Consent All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.

References

Papers of particular interest, published recently, have been highlighted as:

- •• Of major importance
 - 1. Diabetes UK: State of the Nation Report 2014. http://www. diabetes.org.uk/Documents/About%20Us/What%20we%20say/ State%20of%20the%20nation%202014.pdf.
 - Quality and Outcomes Framework: 2012–13. www.hscic.gov.uk/ catalogue/PUB12262.
 - Hex N, Bartlett C, Wright D, et al. Estimating the current and future costs of type 1 and type 2 diabetes in the UK, including direct health costs and indirect societal and productivity costs. Diabet Med. 2012;29(7):855–62.
 - 4. Kerr, M. Inpatient care for people with diabetes—the economic case for change. http://www.diabetes.org.uk/upload/News/ Inpatient%20Care%20for%20People%20with%20Diabetes% 20%20The%20Economic%20Case%20for%20Change% 20Nov%202011.pdf.
 - Ali MK, Bullard KM, Gregg EW, et al. A cascade of care for diabetes in the United States: visualizing the gaps. Ann Intern Med. 2014;161(10):681–9.
 - 6. American Diabetes Association. Economic costs of diabetes in the U.S. in 2012. Diabetes Care. 2013;36(4):1033–46.
 - Gilg J, Pruthi R, Fogarty D. UK renal registry 17th annual report: chapter 1 UK renal replacement therapy incidence in 2013: national and centre-specific analyses. Nephron Physiol. 2015;129 Suppl 1:1–29.
 - Chapter 1, United States Renal Data System, 2014 annual data report: an overview of the epidemiology of kidney disease in the United States. www.usrds.org/2014/download/V2_Ch_01_ ESRD Incidence Prevalence 14.pdf.
 - Chapter 10, United States Renal Data System, 2014 annual data report: an overview of the epidemiology of kidney disease in the United States. www.usrds.org/2014/download/V2_Ch_10_ International 14.pdf.
- 10. United States Renal Data System, 2014 annual data report: an overview of the epidemiology of kidney disease in the United States. http://www.usrds.org/adr.aspx.
- Coresh J, Turin TC, Matsushita K, et al. Decline in estimated glomerular filtration rate and subsequent risk of end-stage renal disease and mortality. JAMA. 2014;311(24):2518–31.
- Fox CS, Matsushita K, Woodward M, et al. Associations of kidney disease measures with mortality and end-stage renal disease in individuals with and without diabetes: a meta-analysis. Lancet. 2012;380(9854):1662–73.
- Hill CJ, Cardwell CR, Patterson CC, et al. Chronic kidney disease and diabetes in the national health service: a cross-sectional survey of the U.K. National Diabetes Audit. Diabet Med. 2014;31(4): 448–54.
- Ho K, McKnight AJ. The changing landscape of diabetic kidney disease: new reflections on phenotype, classification, and disease progression to influence future investigative studies and therapeutic trials. Adv Chronic Kidney Dis. 2014;21(3): 256–9.
- 15.•• Marshall SM. Natural history and clinical characteristics of CKD in type 1 and type 2 diabetes mellitus. Adv Chronic Kidney Dis. 2014;21(3):267–72. Comprehensive review explaining the challenges of assigning clinical phenotypes for kidney disease in individuals with diabetes.
- Mogensen CE, Christensen CK. Predicting diabetic nephropathy in insulin-dependent patients. N Engl J Med. 1984;311(2):89–93.

- Andersen AR, Christiansen JS, Andersen JK, et al. Diabetic nephropathy in type 1 (insulin-dependent) diabetes: an epidemiological study. Diabetologia. 1983;25(6):496–501.
- Borch-Johnsen K, Andersen PK, Deckert T. The effect of proteinuria on relative mortality in type 1 (insulin-dependent) diabetes mellitus. Diabetologia. 1985;28(8):590–6.
- Marshall SM. Diabetic nephropathy in type 1 diabetes: has the outlook improved since the 1980s? Diabetologia. 2012;55(9): 2301–6.
- Mogensen CE, Keane WF, Bennett PH, et al. Prevention of diabetic renal disease with special reference to microalbuminuria. Lancet. 1995;346(8982):1080–4.
- Viberti GC, Hill RD, Jarrett RJ, et al. Microalbuminuria as a predictor of clinical nephropathy in insulin-dependent diabetes mellitus. Lancet. 1982;1(8287):1430–2.
- 22. Parving HH, Persson F, Rossing P. Microalbuminuria: a parameter that has changed diabetes care. Diabetes Res Clin Pract. 2015;107(1):1–8.
- Perkins BA, Ficociello LH, Silva KH, et al. Regression of microalbuminuria in type 1 diabetes. N Engl J Med. 2003;348(23):2285-93.
- Perkins BA, Ficociello LH, Roshan B, et al. In patients with type 1 diabetes and new-onset microalbuminuria the development of advanced chronic kidney disease may not require progression to proteinuria. Kidney Int. 2010;77(1):57–64.
- Caramori ML, Fioretto P, Mauer M. Low glomerular filtration rate in normoalbuminuric type 1 diabetic patients: an indicator of more advanced glomerular lesions. Diabetes. 2003;52(4):1036–40.
- Najafian B, Alpers CE, Fogo AB. Pathology of human diabetic nephropathy. Contrib Nephrol. 2011;170:36–47.
- Tervaert TW, Mooyaart AL, Amann K, et al. Pathologic classification of diabetic nephropathy. J Am Soc Nephrol. 2010;21(4): 556–63.
- Mazzucco G, Bertani T, Fortunato M, et al. Different patterns of renal damage in type 2 diabetes mellitus: a multicentric study on 393 biopsies. Am J Kidney Dis. 2002;39(4):713–20.
- Bell S, Fletcher EH, Brady I, et al. End-stage renal disease and survival in people with diabetes: a national database linkage study. QJM. 2015;108(2):127–34.
- Chan Y, Lim ET, Sandholm N, et al. An excess of risk-increasing low-frequency variants can be a signal of polygenic inheritance in complex diseases. Am J Hum Genet. 2014;94(3):437–52.
- Placha G, Canani LH, Warram JH, et al. Evidence for different susceptibility genes for proteinuria and ESRD in type 2 diabetes. Adv Chronic Kidney Dis. 2005;12(2):155–69.
- Kottgen A, Pattaro C, Boger CA, et al. New loci associated with kidney function and chronic kidney disease. Nat Genet. 2010;42(5):376–84.
- Sandholm N, Forsblom C, Makinen VP, et al. Genome-wide association study of urinary albumin excretion rate in patients with type 1 diabetes. Diabetologia. 2014;57(6):1143–53.
- 34.•• Sandholm N, McKnight AJ, Salem RM, et al. Chromosome 2q31.1 associates with ESRD in women with type 1 diabetes. J Am Soc Nephrol. 2013;24(10):1537–43. First identification of gender-specific SNP with genome-wide significance for diabetic nephropathy.
- Canani LH, Gerchman F, Gross JL. Familial clustering of diabetic nephropathy in Brazilian type 2 diabetic patients. Diabetes. 1999;48(4):909–13.
- Skrunes R, Svarstad E, Reisaeter AV, et al. Familial clustering of ESRD in the Norwegian population. Clin J Am Soc Nephrol. 2014;9(10):1692–700.
- Spray BJ, Atassi NG, Tuttle AB, et al. Familial risk, age at onset, and cause of end-stage renal disease in white Americans. J Am Soc Nephrol. 1995;5(10):1806–10.

- Thameem F, Kawalit IA, Adler SG, et al. Susceptibility gene search for nephropathy and related traits in Mexican-Americans. Mol Biol Rep. 2013;40(10):5769–79.
- Adler AI, Stevens RJ, Manley SE, et al. Development and progression of nephropathy in type 2 diabetes: the United Kingdom Prospective Diabetes Study (UKPDS 64). Kidney Int. 2003;63(1): 225–32.
- DCCT Research. Group. The effect of intensive treatment of diabetes on the development and progression of long-term complications in insulin-dependent diabetes mellitus. The Diabetes Control and Complications Trial Research Group. N Engl J Med. 1993;329(14):977–86.
- Gaede P, Vedel P, Larsen N, et al. Multifactorial intervention and cardiovascular disease in patients with type 2 diabetes. N Engl J Med. 2003;348(5):383–93.
- Forsblom CM, Kanninen T, Lehtovirta M, et al. Heritability of albumin excretion rate in families of patients with type II diabetes. Diabetologia. 1999;42(11):1359–66.
- Fogarty DG, Rich SS, Hanna L, et al. Urinary albumin excretion in families with type 2 diabetes is heritable and genetically correlated to blood pressure. Kidney Int. 2000;57(1):250–7.
- Langefeld CD, Beck SR, Bowden DW, et al. Heritability of GFR and albuminuria in Caucasians with type 2 diabetes mellitus. Am J Kidney Dis. 2004;43(5):796–800.
- MacCluer JW, Scavini M, Shah VO, et al. Heritability of measures of kidney disease among Zuni Indians: the Zuni Kidney Project. Am J Kidney Dis. 2010;56(2):289–302.
- Harjutsalo V, Groop PH. Epidemiology and risk factors for diabetic kidney disease. Adv Chronic Kidney Dis. 2014;21(3):260–6.
- Bell CG, Teschendorff AE, Rakyan VK, et al. Genome-wide DNA methylation analysis for diabetic nephropathy in type 1 diabetes mellitus. BMC Med Genomics. 2010;3:33.
- 48. Gu T, Gu HF, Hilding A, et al. Increased DNA methylation levels of the insulin-like growth factor binding protein 1 gene are associated with type 2 diabetes in Swedish men. Clin Epigenetics. 2013;5(1):21.
- Rakyan VK, Beyan H, Down TA, et al. Identification of type 1 diabetes-associated DNA methylation variable positions that precede disease diagnosis. PLoS Genet. 2011;7(9):e1002300.
- McKnight AJ, McKay GJ, Maxwell AP. Genetic and epigenetic risk factors for diabetic kidney disease. Adv Chron Kidney Dis. 2014;21(3):287–96.
- McKnight AJ, O'Donoghue D, Peter MA. Annotated chromosome maps for renal disease. Hum Mutat. 2009;30(3):314–20.
- Nazir N, Siddiqui K, Al-Qasim S, et al. Meta-analysis of diabetic nephropathy associated genetic variants in inflammation and angiogenesis involved in different biochemical pathways. BMC Med Genet. 2014;15(1):103.
- Tong Z, Yang Z, Patel S, et al. Promoter polymorphism of the erythropoietin gene in severe diabetic eye and kidney complications. Proc Natl Acad Sci U S A. 2008;105(19):6998–7003.
- Williams WW, Salem RM, McKnight AJ, et al. Association testing of previously reported variants in a large case-control metaanalysis of diabetic nephropathy. Diabetes. 2012;61(8):2187–94.
- McKnight AJ, Currie D, Maxwell AP. Unravelling the genetic basis of renal diseases; from single gene to multifactorial disorders. J Pathol. 2010;220(2):198–216.
- 56. Pezzolesi MG, Krolewski AS. The genetic risk of kidney disease in type 2 diabetes. Med Clin N Am. 2013;97(1):91–107.
- 57. Igo Jr RP, Iyengar SK, Nicholas SB, et al. Genomewide linkage scan for diabetic renal failure and albuminuria: the FIND study. Am J Nephrol. 2011;33(5):381–9.
- Thameem F, Igo Jr RP, Freedman BI, et al. A genome-wide search for linkage of estimated glomerular filtration rate (eGFR) in the Family Investigation of Nephropathy and Diabetes (FIND). PLoS One. 2013;8(12):e81888.

- Mahajan A, Sim X, Ng HJ, et al. Identification and functional characterization of G6PC2 coding variants influencing glycemic traits define an effector transcript at the G6PC2-ABCB11 locus. PLoS Genet. 2015;11(1):e1004876.
- 60. Palmer ND, Goodarzi MO, Langefeld CD et al. Genetic variants associated with quantitative glucose homeostasis traits translate to type 2 diabetes in Mexican Americans: the GUARDIAN (Genetics Underlying Diabetes in Hispanics) Consortium. Diabetes 2014.
- Kuchenbaecker KB, Ramus SJ, Tyrer J, et al. Identification of six new susceptibility loci for invasive epithelial ovarian cancer. Nat Genet. 2015;47(2):164–71.
- 62. Al Olama AA, Kote-Jarai Z, Berndt SI, et al. A meta-analysis of 87,040 individuals identifies 23 new susceptibility loci for prostate cancer. Nat Genet. 2014;46(10):1103–9.
- 63. Lambert JC, Ibrahim-Verbaas CA, Harold D, et al. Meta-analysis of 74,046 individuals identifies 11 new susceptibility loci for Alzheimer's disease. Nat Genet. 2013;45(12):1452–8.
- 64. Ha NT, Freytag S, Bickeboeller H. Coverage and efficiency in current SNP chips. Eur J Hum Genet. 2014;22(9):1124–30.
- 65. Little J, Higgins JP, Ioannidis JP, et al. STrengthening the REporting of Genetic Association Studies (STREGA): an extension of the STROBE statement. PLoS Med. 2009;6(2):e22.
- Winkler TW, Day FR, Croteau-Chonka DC, et al. Quality control and conduct of genome-wide association meta-analyses. Nat Protoc. 2014;9(5):1192–212.
- Feehally J, Farrall M, Boland A, et al. HLA has strongest association with IgA nephropathy in genome-wide analysis. J Am Soc Nephrol. 2010;21(10):1791–7.
- Kiryluk K, Li Y, Scolari F, et al. Discovery of new risk loci for IgA nephropathy implicates genes involved in immunity against intestinal pathogens. Nat Genet. 2014;46(11):1187–96.
- Stanescu HC, Arcos-Burgos M, Medlar A, et al. Risk HLA-DQA1 and PLA(2)R1 alleles in idiopathic membranous nephropathy. N Engl J Med. 2011;364(7):616–26.
- Genovese G, Tonna SJ, Knob AU, et al. A risk allele for focal segmental glomerulosclerosis in African Americans is located within a region containing APOL1 and MYH9. Kidney Int. 2010;78(7):698–704.
- Kopp JB, Smith MW, Nelson GW, et al. MYH9 is a major-effect risk gene for focal segmental glomerulosclerosis. Nat Genet. 2008;40(10):1175–84.
- Kottgen A, Glazer NL, Dehghan A, et al. Multiple loci associated with indices of renal function and chronic kidney disease. Nat Genet. 2009;41(6):712–7.
- Liu CT, Garnaas MK, Tin A, et al. Genetic association for renal traits among participants of African ancestry reveals new loci for renal function. PLoS Genet. 2011;7(9):e1002264.
- Kao WH, Klag MJ, Meoni LA, et al. MYH9 is associated with nondiabetic end-stage renal disease in African Americans. Nat Genet. 2008;40(10):1185–92.
- 75.•• Sandholm N, Salem RM, McKnight AJ, et al. New susceptibility loci associated with kidney disease in type 1 diabetes. PLoS Genet. 2012;8(9):e1002921. Largest meta-analysis of GWAS conducted for diabetic nephropathy with replication confirming significant SNPs.
- Shimazaki A, Kawamura Y, Kanazawa A, et al. Genetic variations in the gene encoding ELMO1 are associated with susceptibility to diabetic nephropathy. Diabetes. 2005;54(4):1171–8.
- Hanson RL, Millis MP, Young NJ, et al. ELMO1 variants and susceptibility to diabetic nephropathy in American Indians. Mol Genet Metab. 2010;101(4):383–90.
- Leak TS, Perlegas PS, Smith SG, et al. Variants in intron 13 of the ELMO1 gene are associated with diabetic nephropathy in African Americans. Ann Hum Genet. 2009;73(2):152–9.

- Pezzolesi MG, Katavetin P, Kure M, et al. Confirmation of genetic associations at ELMO1 in the GoKinD collection supports its role as a susceptibility gene in diabetic nephropathy. Diabetes. 2009;58(11):2698–702.
- Wu HY, Wang Y, Chen M, et al. Association of ELMO1 gene polymorphisms with diabetic nephropathy in Chinese population. J Endocrinol Invest. 2013;36(5):298–302.
- Yadav AK, Kumar V, Dutta P, et al. Variations in CCR5, but not HFE, ELMO1, or SLC12A3, are associated with susceptibility to kidney disease in north Indian individuals with type 2 diabetes CCR5HFEELMO1SLC12A32. J Diabetes. 2014;6(6):547–55.
- Franceschini N, Shara NM, Wang H, et al. The association of genetic variants of type 2 diabetes with kidney function. Kidney Int. 2012;82(2):220–5.
- Hanson RL, Craig DW, Millis MP, et al. Identification of PVT1 as a candidate gene for end-stage renal disease in type 2 diabetes using a pooling-based genome-wide single nucleotide polymorphism association study. Diabetes. 2007;56(4):975–83.
- McDonough CW, Palmer ND, Hicks PJ, et al. A genome-wide association study for diabetic nephropathy genes in African Americans. Kidney Int. 2011;79(5):563–72.
- Ebers GC, Sadovnik AD. Re: GAMES issue. J Neuroimmunol. 2004;153(1–2):4–5.
- McKnight AJ, Maxwell AP, Sawcer S, et al. A genome-wide DNA microsatellite association screen to identify chromosomal regions harboring candidate genes in diabetic nephropathy. J Am Soc Nephrol. 2006;17(3):831–6.
- Craig DW, Millis MP, DiStefano JK. Genome-wide SNP genotyping study using pooled DNA to identify candidate markers mediating susceptibility to end-stage renal disease attributed to type 1 diabetes. Diabet Med. 2009;26(11):1090–8.
- Pezzolesi MG, Poznik GD, Mychaleckyj JC, et al. Genome-wide association scan for diabetic nephropathy susceptibility genes in type 1 diabetes. Diabetes. 2009;58(6):1403–10.
- Palmer ND, Ng MC, Hicks PJ, et al. Evaluation of candidate nephropathy susceptibility genes in a genome-wide association study of African American diabetic kidney disease. PLoS One. 2014;9(2):e88273.
- 90. Martini S, Nair V, Patel SR, et al. From single nucleotide polymorphism to transcriptional mechanism: a model for FRMD3 in diabetic nephropathy. Diabetes. 2013;62(7):2605–12.
- Tryka KA, Hao L, Sturcke A, et al. NCBI's database of Genotypes and Phenotypes: dbGaP. Nucleic Acids Res. 2014;42(Database issue):D975–9.
- 92. Maeda S, Imamura M, Kurashige M, et al. Replication study for the association of 3 SNP loci identified in a genome-wide association study for diabetic nephropathy in European type 1 diabetes with diabetic nephropathy in Japanese patients with type 2 diabetes. Clin Exp Nephrol. 2013;17(6):866–71.
- Sambo F, Malovini A, Sandholm N, et al. Novel genetic susceptibility loci for diabetic end-stage renal disease identified through robust naive Bayes classification. Diabetologia. 2014;57(8):1611– 22.
- Germain M, Pezzolesi MG, Sandholm N, et al. SORBS1 gene, a new candidate for diabetic nephropathy: results from a multi-stage genome-wide association study in patients with type 1 diabetes. Diabetologia. 2015;58(3):543–8.
- Lettre G, Jackson AU, Gieger C, et al. Identification of ten loci associated with height highlights new biological pathways in human growth. Nat Genet. 2008;40(5):584–91.
- Lango AH, Estrada K, Lettre G, et al. Hundreds of variants clustered in genomic loci and biological pathways affect human height. Nature. 2010;467(7317):832–8.
- 97. Wood AR, Esko T, Yang J, et al. Defining the role of common variation in the genomic and biological architecture of adult human height. Nat Genet. 2014;46(11):1173–86.

- Harjutsalo V, Maric C, Forsblom C, et al. Sex-related differences in the long-term risk of microvascular complications by age at onset of type 1 diabetes. Diabetologia. 2011;54(8):1992–9.
- Charchar FJ, Bloomer LD, Barnes TA, et al. Inheritance of coronary artery disease in men: an analysis of the role of the Y chromosome. Lancet. 2012;379(9819):915–22.
- Sharma K, Karl B, Mathew AV, et al. Metabolomics reveals signature of mitochondrial dysfunction in diabetic kidney disease. J Am Soc Nephrol. 2013;24(11):1901–12.
- Higgins GC, Coughlan MT. Mitochondrial dysfunction and mitophagy: the beginning and end to diabetic nephropathy? Br J Pharmacol. 2014;171(8):1917–42.
- 102. Swan EJ, Salem RM, Sandholm N et al. Genetic risk factors affecting mitochondrial function are associated with kidney disease in individuals with type 1 diabetes. Diabet Med 2015.
- 103. Swan EJ, Maxwell AP, McKnight AJ. Distinct methylation patterns in genes that affect mitochondrial function are associated with kidney disease in blood-derived DNA from individuals with type 1 diabetes. Diabet Med 2015.
- Douglas AP, Vance DR, Kenny EM, et al. Next-generation sequencing of the mitochondrial genome and association with IgA nephropathy in a renal transplant population. Sci Rep. 2014;4: 7379.
- McKnight AJ, Maxwell AP. Bioinformatic Resources for Diabetic Nephropathy. J Diabetes Bioinforma. 2013;1(1):11–8.
- Martini S, Nair V, Keller BJ, et al. Integrative biology identifies shared transcriptional networks in CKD. J Am Soc Nephrol. 2014;25(11):2559–72.
- 107. Brennan EP, Morine MJ, Walsh DW, et al. Next-generation sequencing identifies TGF-beta1-associated gene expression profiles in renal epithelial cells reiterated in human diabetic nephropathy. Biochim Biophys Acta. 2012;1822(4):589–99.
- Liu R, Zhong Y, Li X, et al. Role of transcription factor acetylation in diabetic kidney disease. Diabetes. 2014;63(7):2440–53.
- Miao F, Chen Z, Genuth S, et al. Evaluating the role of epigenetic histone modifications in the metabolic memory of type 1 diabetes. Diabetes. 2014;63(5):1748–62.
- Reddy MA, Sumanth P, Lanting L, et al. Losartan reverses permissive epigenetic changes in renal glomeruli of diabetic db/db mice. Kidney Int. 2014;85(2):362–73.
- 111. Kato M, Zhang J, Wang M, et al. MicroRNA-192 in diabetic kidney glomeruli and its function in TGF-beta-induced collagen expression via inhibition of E-box repressors. Proc Natl Acad Sci U S A. 2007;104(9):3432–7.
- 112. Zhou Q, Chung AC, Huang XR, et al. Identification of novel long noncoding RNAs associated with TGF-beta/Smad3-mediated renal inflammation and fibrosis by RNA sequencing. Am J Pathol. 2014;184(2):409–17.
- 113. Zhou J, Peng R, Li T, et al. A potentially functional polymorphism in the regulatory region of let-7a-2 is associated with an increased risk for diabetic nephropathy. Gene. 2013;527(2):456–61.
- Smyth LJ, McKay GJ, Maxwell AP, et al. DNA hypermethylation and DNA hypomethylation is present at different loci in chronic kidney disease. Epigenetics. 2014;9(3):366–76.
- 115. Zhang H, Cai X, Yi B, et al. Correlation of CTGF gene promoter methylation with CTGF expression in type 2 diabetes mellitus with or without nephropathy. Mol Med Rep. 2014;9(6):2138–44.
- 116. Sapienza C, Lee J, Powell J, et al. DNA methylation profiling identifies epigenetic differences between diabetes patients with ESRD and diabetes patients without nephropathy. Epigenetics. 2011;6(1):20–8.
- 117. Hirayama A, Nakashima E, Sugimoto M, et al. Metabolic profiling reveals new serum biomarkers for differentiating diabetic nephropathy. Anal Bioanal Chem. 2012;404(10):3101–9.
- 118. Pena MJ, Lambers Heerspink HJ, Hellemons ME, et al. Urine and plasma metabolites predict the development of diabetic

nephropathy in individuals with type 2 diabetes mellitus. Diabet Med. 2014;31(9):1138–47.

- 119. Stec DF, Wang S, Stothers C, et al. Alterations of urinary metabolite profile in model diabetic nephropathy. Biochem Biophys Res Commun. 2015;456(2):610–4.
- Zubiri I, Posada-Ayala M, Sanz-Maroto A, et al. Diabetic nephropathy induces changes in the proteome of human urinary exosomes as revealed by label-free comparative analysis. J Proteomics. 2014;96:92–102.
- Caseiro A, Barros A, Ferreira R, et al. Pursuing type 1 diabetes mellitus and related complications through urinary proteomics. Transl Res. 2014;163(3):188–99.
- Zurbig P, Jerums G, Hovind P, et al. Urinary proteomics for early diagnosis in diabetic nephropathy. Diabetes. 2012;61(12):3304– 13.
- Locke AE, Kahalo B, Berndt SI, et al. Genetic studies of body mass index yield new insights for obesity biology. Nature. 2015;518(7538):197–206.
- Shungin D, Winkler TW, Croteau-Chonka DC, et al. New genetic loci link adipose and insulin biology to body fat distribution. Nature. 2015;518(7538):187–96.
- Pers TH, Karjalainen JM, Chan Y, et al. Biological interpretation of genome-wide association studies using predicted gene functions. Nat Commun. 2015;6:5890.
- Abecasis GR, Auton A, Brooks LD, et al. An integrated map of genetic variation from 1,092 human genomes. Nature. 2012;491(7422):56–65.
- 127. Genonics England. The 100,000 Genomes Project. www. genomicsengland.co.uk/the-100000-genomes-project.
- McKnight AJ, Currie D, Patterson CC, et al. Targeted genomewide investigation identifies novel SNPs associated with diabetic nephropathy. Hugo J. 2009;3(1–4):77–82.
- Savage DA, Patterson CC, Deloukas P, et al. Genetic association analyses of non-synonymous single nucleotide polymorphisms in diabetic nephropathy. Diabetologia. 2008;51(11):1998–2002.
- Cooke Bailey JN, Palmer ND, Ng MC, et al. Analysis of coding variants identified from exome sequencing resources for association with diabetic and non-diabetic nephropathy in African Americans. Hum Genet. 2014;133(6):769–79.
- Collins R. What makes UK Biobank special? Lancet. 2012;379(9822):1173–4.

- 132. Kerr SM, Campbell A, Murphy L, et al. Pedigree and genotyping quality analyses of over 10,000 DNA samples from the Generation Scotland: Scotlish Family Health Study. BMC Med Genet. 2013;14:38.
- Zhang C, Pierce BL. Genetic susceptibility to accelerated cognitive decline in the US Health and Retirement Study. Neurobiol Aging. 2014;35(6):1512–8.
- Westra HJ, Peters MJ, Esko T, et al. Systematic identification of trans eQTLs as putative drivers of known disease associations. Nat Genet. 2013;45(10):1238–43.
- Ward LD, Kellis M. HaploReg: a resource for exploring chromatin states, conservation, and regulatory motif alterations within sets of genetically linked variants. Nucleic Acids Res. 2012;40(Database issue):D930–4.
- Mooyaart AL, Valk EJ, van Es LA, et al. Genetic associations in diabetic nephropathy: a meta-analysis. Diabetologia. 2014;57(3): 650.
- 137. Garcia DL, Anderson S, Rennke HG, Brenner BM. Anemia lessens and its prevention with recombinant human erythropoietin worsens glomerular injury and hypertension in rats with reduced renal mass. Proc Natl Acad Sci U S A. 1988;85(16):6142–6.
- Veikkolainen V, Naillat F, Railo A, et al. ErbB4 modulates tubular cell polarity and lumen diameter during kidney development. J Am Soc Nephrol. 2012;23(1):112–22.
- Zeng F, Zhang MZ, Singh AB, et al. ErbB4 isoforms selectively regulate growth factor induced Madin-Darby canine kidney cell tubulogenesis. Mol Biol Cell. 2007;18(11):4446–56.
- Woroniecka KI, Park AS, Mohtat D, Thomas DB, Pullman JM, Susztak K. Transcriptome analysis of human diabetic kidney disease. Diabetes. 2011;60(9):2354–69.
- Schmid H, Boucherot A, Yasuda Y, et al. Modular activation of nuclear factor-kappaB transcriptional programs in human diabetic nephropathy. Diabetes. 2006;55(11):2993–3003.
- 142. Lin WH, Huang CJ, Liu MW, et al. Cloning, mapping, and characterization of the human sorbin and SH3 domain containing 1 (SORBS1) gene: a protein associated with c-Abl during insulin signaling in the hepatoma cell line Hep3B. Genomics. 2001;74(1): 12–20.
- 143. Nakatani S, Kakehashi A, Ishimura E et al. Targeted proteomics of isolated glomeruli from the kidneys of diabetic rats: sorbin and SH3 domain containing 2 is a novel protein associated with diabetic nephropathy. Exp Diabetes Res. 2011.