

Serum Amyloid A: The “Other” Inflammatory Protein

Kevin D. O'Brien, MD, and Alan Chait, MD

Corresponding author

Kevin D. O'Brien, MD
Division of Cardiology, Box 356422, University of Washington,
Seattle, WA 98195-6422, USA.
E-mail: cardiac@u.washington.edu

Current Atherosclerosis Reports 2006, 8:62–68
Current Science Inc. ISSN 1523-3804
Copyright © 2006 by Current Science Inc.

Inflammation long has been recognized as a hallmark of atherosclerotic lesions, but more recently attention has focused on chronic low-level elevations of specific plasma inflammatory proteins such as C-reactive protein (CRP) and serum amyloid A (SAA), which may not only represent markers of atherosclerosis risk but also participate directly in atherogenesis. This article briefly reviews evidence for and against potential roles of CRP as an atherosclerosis risk marker and in atherogenesis. The remainder of the article focuses on SAA, an inflammatory protein that is carried on, and may fundamentally alter the function of, high-density lipoprotein. Data are reviewed regarding the regulation of SAA by dietary cholesterol, obesity, and insulin resistance, and its potential role as an atherosclerosis mediator. Lying at the intersection of inflammation, dyslipidemia, obesity, and insulin resistance, SAA may play a key role in regulating the contributions of these processes to atherogenesis.

Introduction

Inflammation long has been recognized as a hallmark of atherosclerotic lesions [1,2], but recently substantial attention has been paid to the role of serum proteins that are increased in response to inflammation. These proteins have been studied primarily as markers of atherosclerotic risk, but some also have been implicated as potential mediators of the atherosclerotic process. This review briefly discusses studies evaluating one of these “inflammatory” proteins, namely C-reactive protein (CRP), but then focuses primarily on serum amyloid A (SAA), which is a less well-studied protein that may serve as a link between atherosclerosis, inflammation, and obesity/insulin resistance.

The Acute-phase Response

In acute inflammation, whether triggered by infection, direct tissue injury, or other diseases, circulating cytokines induce the liver to synthesize a number of proteins that attempt to respond to the insult. Many of these proteins play key roles, not only in the inflammatory and immune response to injury, but also in activating the fibrinolytic and complement systems. Some acute-phase proteins are carried in the plasma on circulating lipoproteins. Though the functions of many acute-phase proteins are well understood, this is not true of all proteins involved in the acute-phase response, particularly CRP and SAA, which may increase by as much as 1000-fold above their baseline levels in acute inflammation [3]. More recently, it has become apparent that levels of many acute-phase proteins, including CRP and SAA, may be chronically elevated in some individuals, albeit at levels much lower than those seen in the acute-phase response. Although acute elevations of these proteins are likely to have beneficial effects, chronic elevations might be deleterious. It is, therefore, of interest that levels of CRP and SAA may be chronically elevated in individuals with atherosclerosis, diabetes, obesity, insulin resistance, and rheumatologic diseases, all of which are associated with an increased risk of cardiovascular disease. Therefore, the potential significance of these proteins in chronic disease states, both as clinical markers and as pathologic mediators of cardiovascular disease, has undergone substantial scrutiny. This is particularly true for CRP, which several studies have associated with increased risk for ischemic heart disease events and stroke. In addition, while the liver classically has been considered the primary source of most acute-phase proteins, recent studies have demonstrated that other tissues might also be sources of these proteins, especially SAA. These observations have raised the exciting possibility that specific acute-phase proteins might be exploited for diagnostic or therapeutic benefits in chronic disease states such as atherosclerosis.

C-Reactive Protein: The “Orthodox” Inflammatory Protein

C-reactive protein is the most studied of the acute-phase proteins and was named for its property of binding to the

C-polysaccharide of *Streptococcus pneumoniae*. CRP is a pentraxin, composed of five identical protein molecules noncovalently linked around a central protein core. CRP circulates in plasma unbound to lipoproteins, but it can bind to oxidized lipoproteins and apoptotic cells [4] as well as to aggregated low-density lipoprotein (LDL) in vitro [5,6]. In addition, CRP can bind complement and can bind to the Fc-gamma-II (CD32) receptor [6].

C-reactive protein as a marker of cardiovascular risk

Recently, CRP has been studied extensively as a marker for atherosclerosis risk, and elevated levels have been associated with increased risk for important atherosclerosis endpoints, including myocardial infarction and stroke [7-10]. CRP levels are increased in a number of conditions that are associated with increased cardiovascular risk, including obesity [11,12•,13•], insulin resistance [13•,14-17], type 2 diabetes [16], and smoking [18]. More recently, increased dietary cholesterol intake has been shown to increase CRP levels in lean, insulin-sensitive subjects [13•]. However, CRP has been less firmly associated with atherosclerosis severity, at least as measured by electron beam computed tomography [19,20] or coronary angiography [21]. Recent studies have suggested that, following acute coronary syndromes, CRP levels give prognostic information beyond that provided by LDL levels alone [22,23]. However, these studies do not take into account the relationships of obesity and insulin resistance to CRP levels [13•,14,15], nor do they prove that CRP has incremental prognostic value over a composite of traditional risk factors that, in aggregate, may account for up to 94% of the population-attributable risk for coronary events [24]. In addition, a recent study by Danesh et al. [25], which included data from the Reykjavik Prospective Study as well as an updated meta-analysis of CRP levels in over 20,000 subjects, concluded that the additional risk associated with elevated CRP levels was less than that for hypercholesterolemia, systolic hypertension, or smoking. Finally, one recent study demonstrated that, even in the absence of changes in either medication or clinical status, nearly 40% of ischemic heart disease patients changed their CRP-based risk category [26] over 1 month of follow-up [27]. Thus, despite its apparent utility as a risk predictor in large groups, the marked inter-individual variability in CRP levels may limit its clinical utility as a tool for either predicting risk or following response to therapy in individual patients.

C-reactive protein as a mediator of atherogenesis

A number of recent studies also have suggested mechanisms by which CRP might participate directly in atherogenesis. CRP has been shown to have a number of potentially proatherogenic effects on endothelial effects in vitro. These include induction of leukocyte adhesion molecules [28], monocyte chemoattractant protein-1 [29], interleukin-8 [30], and plasminogen activator

inhibitor-1 [31], as well as inhibition of tissue plasminogen activator [32] and of the vasodilators prostacyclin [33] and nitric oxide (NO) [34,35]. However, a more recent study reported that purified, native-form human CRP actually increased NO bioavailability in endothelial cells and isolated arterial rings [36]. Thus, some authorities have raised the concern that proinflammatory effects identified in some previous in vitro studies might have been due to use of CRP with low levels of contaminants, such as endotoxin, sodium azide, or CRP in non-native forms [37].

In vivo evidence of a direct proatherogenic role of CRP consists of one study in the apoE-deficient mouse model of atherogenesis, demonstrating a modest increase in atherosclerotic lesion area in male, but not in female, mice expressing a human CRP transgene. In contrast, three recent studies found no effect on atherosclerosis of overexpression of either a human [38••,39•] or rabbit [40•] CRP transgene. Thus, more recent in vivo studies do not support a role for CRP as a mediator of atherogenesis.

Serum Amyloid A: The "Other" Inflammatory Protein

Serum amyloid A is a family of four homologous, amphipathic, alpha-helical proteins encoded for by genes located on chromosome 7 in mice and on chromosome 11 in humans. SAA includes SAA1 and SAA2, which are acute-phase proteins, and SAA4, which is expressed constitutively. Like CRP, hepatic expression of SAA1 and SAA2 is increased markedly in response to a variety of inflammatory stimuli [3,41]. In contrast, SAA4 is constitutively expressed by the liver [41]. Though classically thought to be produced primarily by hepatocytes, both the acute-phase and constitutive forms of SAA have been shown to be expressed by endothelial cells, macrophages, and smooth muscle cells in human atherosclerotic plaques [42], and by cytokine-stimulated smooth muscle cells in vitro [42]. SAA3 is a truncated protein primarily expressed in mice by extrahepatic cells, including adipocytes and macrophages. SAA3 typically is not expressed in humans due to the presence of a premature stop codon in exon 2 [3,41]. SAA are transported in the plasma primarily on HDL particles, but also may be carried on triglyceride-rich very low-density lipoprotein (VLDL) particles, particularly in circumstances where SAA levels are elevated [3,41]. In general, serum levels of CRP and SAA are highly correlated in humans [12•,13•,43] and, like CRP, SAA levels are elevated in obesity [12•,13•,16,21,44], insulin resistance [12•,13•,45], and diabetes [16,45,46]. In multivariate analyses, CRP levels generally have correlated better with atherosclerosis risk than have levels of SAA [43], though a recent study has demonstrated better correlation of SAA levels than of CRP levels with angiographic coronary artery disease severity in women [21]. Thus, whereas SAA may not be as sensitive a marker of

atherosclerosis risk as CRP, it might prove to be a better marker of atherosclerosis severity. An important caveat is that SAA levels are subject to substantial inter-individual variation over time, as are levels of CRP [12•].

Regulation of serum amyloid A

As noted previously, SAA levels generally correlate with levels of CRP in a variety of disease states in humans. However, in mice, only SAA levels, but not those of CRP, are regulated by inflammatory stimuli. For example, it was shown several years ago that feeding of an atherogenic diet containing fat, cholesterol, and cholate induces SAA expression in mice [47] through induction of nuclear factor- κ B. More recently, it has been demonstrated that mild, chronic elevations of SAA can be found in both the apoE-deficient and LDL receptor-deficient mouse models of atherogenesis, even in mice fed a chow diet [48•]. Another recent study has demonstrated in LDL receptor-deficient mice that SAA levels are elevated by adding fat to a chow diet, and are elevated even more dramatically by the further addition of cholesterol [49••]. Interestingly, both studies demonstrated, either by immunoprecipitation in chow-fed mice [48•] or by fast protein liquid chromatography in chow-, fat-, or fat and cholesterol-fed mice [49••], that SAA was detected not only on high-density lipoprotein (HDL), but also on VLDL. These findings suggest that SAA may influence the roles of both HDL and VLDL in atherogenesis.

In humans, the relationship of dietary composition to SAA levels is more complex and appears to be mediated, at least in part, by both insulin resistance and obesity. The relationship of obesity [16,21,44] and insulin resistance [45] to elevated levels of SAA is well established. However, four recent studies have extended these observations. Three of these studies have demonstrated that dietary weight loss is associated with reduction in SAA levels [12•,50••,51••]. However, in one of these studies, the correlation of weight loss with reduction in SAA was found even in a group of obese women receiving a weight-loss diet that contained high levels of fat and cholesterol [12•]. Interestingly, in that study, decrease in SAA also correlated with improvement in basal insulin resistance [12•]. The fourth study [13•] differed in that it was not a weight-loss study but rather examined the effect of feeding four eggs per day for 1 month on plasma levels in insulin-sensitive and insulin-resistant groups of subjects. Surprisingly, egg feeding had no effect on SAA levels in the insulin-resistant groups, but dramatically increased SAA levels in insulin-sensitive subjects [13•]. Thus, in contrast to mice, in which feeding dietary fat and/or cholesterol raises SAA levels, short-term increases in dietary fat and cholesterol appear to raise SAA levels only in insulin-sensitive individuals. In contrast, whereas obese subjects and insulin-resistant subjects have elevated SAA levels at baseline, short-term increases in dietary fat and/or cholesterol do not further raise their plasma SAA levels. Specific mechanisms that might account for why obesity

and insulin resistance inhibit diet-induced elevations in SAA levels are not known. One possibility is that dietary cholesterol absorption is inhibited in the presence of obesity [52] and/or insulin resistance [53]. Alternatively, it may be that the presence of adipose tissue macrophages, which recently have been shown to accumulate in obesity [54,55], may blunt the inflammatory response to dietary cholesterol by mechanisms as yet unknown.

Extrahepatic sources of serum amyloid A

As noted previously, with the exception of SAA3, the liver classically has been considered the primary source of SAA protein expression in both normal and disease states. However, several years ago, Meek et al. [42] demonstrated that acute-phase and constitutive SAA also may be expressed by plaque cells, especially macrophage and smooth muscle foam cells. More recently, two important studies have challenged the commonly held notion that the liver is the major source of SAA in obesity, by demonstrating that adipocytes represent a major site of SAA expression in obese individuals [50••,51••]. In the first study, investigators found that 1) SAA expression is 20-fold higher in mature adipocytes than in stromal vascular cells of subcutaneous white adipose tissue (sWAT); 2) mRNA and adipocyte immunoreactivity for SAA were much higher in sWAT of obese as compared with lean subjects; and 3) sWAT mRNA levels (and plasma SAA levels) were decreased following dietary weight loss [50••]. In the second study, SAA mRNA and protein were detected in subcutaneous and omental adipose tissue, and SAA protein was localized to adipocytes by immunohistochemistry. Also, similar to the first study, diet-induced weight loss was associated with a reduction in adipose tissue SAA expression that correlated with reduction in plasma SAA levels [51••]. Finally, the second study also demonstrated by microarray analysis that omental and subcutaneous adipose tissue had substantially higher levels of SAA mRNA expression than did any other tissue studied, including liver [51••]. Taken together, these findings suggest that adipocyte SAA expression (and its regulation by obesity) may account for the strong correlation of SAA levels with obesity and insulin resistance. A summary of the potential sources for the increased levels of CRP and SAA seen in response to dietary cholesterol and obesity/insulin resistance are shown in Figure 1.

Serum amyloid A in atherogenesis

The potential links between SAA, obesity, and insulin resistance, as well as the association of SAA with specific plasma lipoproteins, have stimulated recent interest in a potential role for SAA as a mediator of atherogenesis. In addition, SAA has been shown *in vitro* to have a number of effects that could potentially promote atherosclerosis, including mediating HDL binding to differentiated macrophages [56,57] and endothelial cells [57], and impairing the capacity of HDL to promote cholesterol efflux from macrophages [58]. Free SAA also has been shown *in vitro* to induce

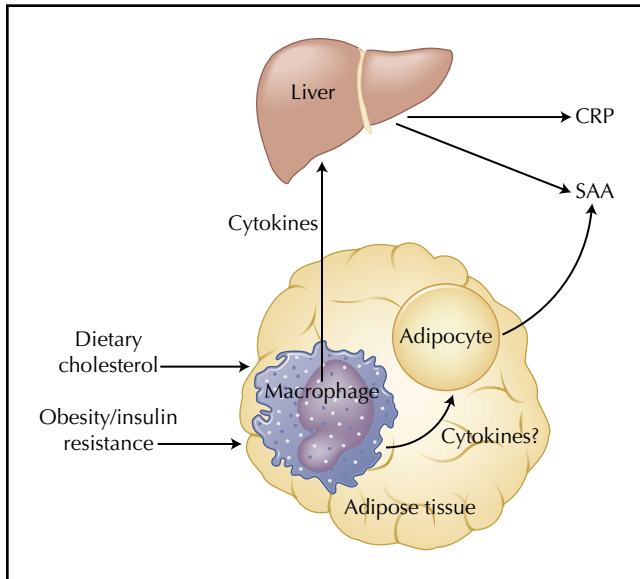


Figure 1. Sources of C-reactive protein (CRP) and serum amyloid A (SAA). Under the influence of dietary cholesterol and/or insulin resistance, adipose tissue accumulates macrophages, which in turn secrete cytokines that induce the liver to express both CRP and SAA. However, adipose tissue adipocytes may also themselves secrete SAA, possibly under the influence of locally secreted cytokines.

expression of extracellular matrix-degrading metalloproteinases [59,60] and to promote chemotaxis and adhesion of both monocytes [61] and T lymphocytes [62].

In addition, SAA may play major roles in lipid transport. As noted previously, SAA associates with both HDL and VLDL in plasma. The teleologic reason why SAA might associate with HDL in inflammatory states is not known, but one theory [56,58] is based on the observations that 1) SAA can displace apolipoprotein A-I from HDL particles [63]; and 2) as compared with HDL without SAA, SAA-containing HDL has decreased affinity for hepatocytes [56] and increased affinity for macrophages [56,58]. In this scheme, the presence of SAA changes HDL from a particle that removes cholesterol from peripheral tissues to the liver (so-called reverse cholesterol transport) to a particle that delivers cholesterol to peripheral tissues, in particular to sites of inflammation [56,58]. In contrast, a recent study has demonstrated that SAA2 (but not SAA1) contains a lipid transport activity in its amino-terminal region that promotes cholesterol efflux from cholesterol-laden macrophages *in vitro* [64].

Two recent studies [48•,49••] also have demonstrated that SAA might play a role in retention of HDL particles in atherosclerotic tissue by acting as a "bridging" molecule mediating binding of HDL to vascular proteoglycans. SAA has a number of positively charged amino acids in its carboxy-terminal region that have been implicated in proteoglycan binding [65]. The recent studies have shown that the presence of SAA on HDL particles increases its binding *in vitro* to perlecan [48•], a proteoglycan that

accumulates in murine atherosclerotic lesions [66], and to biglycan [49••], another proteoglycan that accumulates in both murine [66] and human [67] atherosclerosis. Moreover, both studies co-localized SAA and apoA-I to perlecan-rich regions of murine atherosclerotic lesions, but not in perlecan-free, nonlesioned areas, providing *in vivo* evidence for a role for SAA in plaque HDL retention [48•,49••]. Importantly, one of these studies also showed that plasma levels of SAA, but not of cholesterol, correlated strongly with atherosclerotic lesion area [49••], further supporting a potential direct role for SAA in atherogenesis. However, firm confirmation of a role for SAA in atherogenesis will require direct testing in both transgenic and SAA-deficient animal models.

The retention of HDL on atherosclerotic extracellular matrix by SAA could promote atherogenesis in several ways. Firstly, HDL trapped in the atherosclerotic plaque would be unavailable for transport of cholesterol out of the plaque in the reverse cholesterol transport pathway. Secondly, HDL retained in plaque might be more susceptible to oxidation as well as to other chemical modifications such as nitrosylation [68], chlorination [69], and acrolein adducts [70], all of which may render HDL more atherogenic. In addition, SAA might also play a role in mediating plaque retention of VLDL, as both murine studies have confirmed that SAA also is present on VLDL particles [48•,49••].

Thus, the association of SAA with HDL (and VLDL) in chronic disease states, including insulin resistance, obesity, and diabetes could account, at least in part, for the association of these diseases with increased risk for atherosclerosis. Some of the multiple potential mechanisms by which SAA might stimulate atherogenesis are shown in Figure 2.

Serum amyloid A as a therapeutic target

If SAA is a mediator of atherosclerosis, what therapies may decrease its expression? In general, many therapies that decrease circulating CRP levels also decrease those of SAA, including weight loss [12•,50••,51••], statin treatment [71,72], and improvement in insulin resistance through weight loss [12•] or treatment with peroxisome proliferator activated receptor gamma (PPAR γ) activators [45,73]. In addition, because dietary fat and cholesterol appear to increase SAA levels in lean subjects [13•] but not in individuals who are obese [12•,13•] and/or insulin-resistant [13•], decreasing intake of these dietary components might reduce SAA levels in lean individuals. However, the effects of these therapies are not exclusive to SAA. Therapies that specifically target SAA, such as inhibiting its proteoglycan binding domain (as has been done for apoB) [74], have not been tested.

Conclusions

Acute-phase proteins, in particular CRP and SAA, appear to be good predictors of cardiovascular disease in large

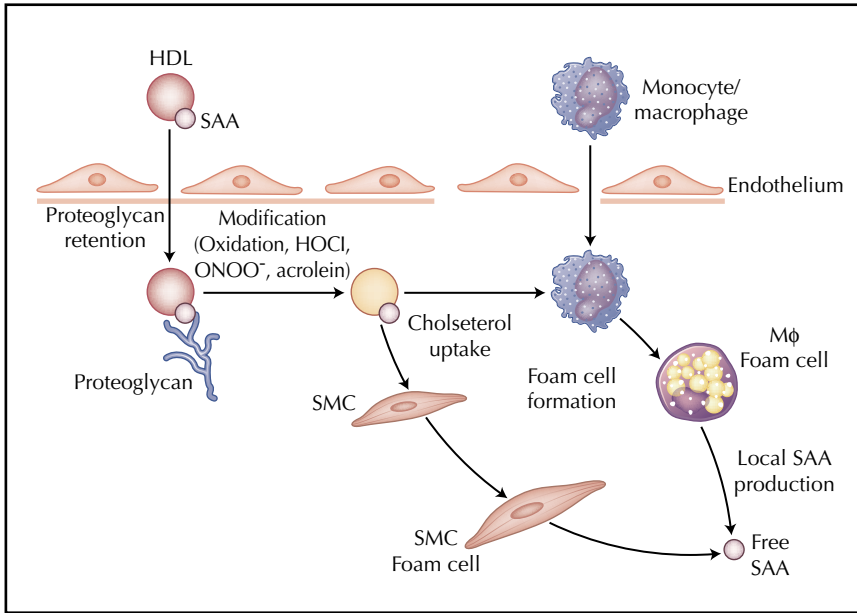


Figure 2. Potential roles of serum amyloid A (SAA) in atherogenesis. When it associates with high-density lipoprotein (HDL), SAA can mediate binding of HDL to vessel wall proteoglycans. The trapped SAA containing HDL then could undergo oxidation or other chemical modifications by locally produced hypochlorous acid (HOCl), peroxynitrite (ONOO⁻), and/or acrolein, which impair the ability of HDL to remove cellular cholesterol. SAA also can directly increase HDL binding to plaque cells. Together, these changes could result in a marked increase in net cellular cholesterol accumulation, thereby leading to macrophage (M ϕ) and smooth muscle cell (SMC) foam cell formation. These activated plaque cells then could secrete free SAA, which in turn may further accelerate plaque development by inducing expression of matrix-degrading metalloproteinases and promoting monocyte and T lymphocyte chemotaxis and adhesion.

epidemiologic studies, though SAA has been less well studied than CRP. The roles of these molecules as mediators of atherogenesis are less clear, particularly for CRP, as three recent transgenic animal studies of CRP have failed to demonstrate any proatherogenic effect. SAA, through its effects on HDL metabolism and regulation by dietary cholesterol, obesity, and insulin resistance, lies at the intersection of inflammation, dyslipidemia, and metabolic syndrome. As a consequence, the potential role of SAA as an atherosclerosis mediator is provocative, though it still needs to be rigorously tested in animal models. If SAA is shown definitively to increase atherosclerosis risk, specific targeting of this inflammatory molecule may offer therapeutic approaches beyond classical risk reduction.

Acknowledgment

Dr. Chait can be contacted at the Division of Metabolism, Endocrinology and Nutrition at University of Washington in Seattle. His e-mail address is achait@u.washington.edu. This manuscript was supported in part by grant HL30086 from the National Institutes of Health, Bethesda, MD.

References and Recommended Reading

Papers of particular interest, published recently, have been highlighted as:

- Of importance
- Of major importance

1. Ross R: **The pathogenesis of atherosclerosis: a perspective for the 1990s.** *Nature* 1993, 362:801–809.
2. Ross R: **Mechanisms of disease. Atherosclerosis—an inflammatory disease.** *N Engl J Med* 1999, 340:115–126.

3. Malle E, De Beer FC: **Human serum amyloid A (SAA) protein: a prominent acute-phase reactant for clinical practice.** *Eur J Clin Invest* 1996, 26:427–435.
4. Chang MK, Binder CJ, Torzewski M, Witztum JL: **C-reactive protein binds to both oxidized LDL and apoptotic cells through recognition of a common ligand: phosphorylcholine of oxidized phospholipids.** *Proc Natl Acad Sci U S A* 2002, 99:13043–13048.
5. De Beer FC, Soutar AK, Baltz ML, et al.: **Low density lipoprotein and very low density lipoprotein are selectively bound by aggregated C-reactive protein.** *J Exp Med* 1982, 156:230–242.
6. Zwaka TP, Hombach V, Torzewski J: **C-reactive protein-mediated low density lipoprotein uptake by macrophages: implications for atherosclerosis.** *Circulation* 2001, 103:1194–1197.
7. Ridker PM, Cushman M, Stampfer MJ, et al.: **Inflammation, aspirin, and the risk of cardiovascular disease in apparently healthy men.** *N Engl J Med* 1997, 336:973–979.
8. Kuller LH, Tracy RP, Shaten J, Meilahn EN: **Relation of C-reactive protein and coronary heart disease in the MRFIT nested case-control study. Multiple Risk Factor Intervention Trial.** *Am J Epidemiol* 1996, 144:537–547.
9. Koenig W, Sund M, Frohlich M, et al.: **C-Reactive protein, a sensitive marker of inflammation, predicts future risk of coronary heart disease in initially healthy middle-aged men: results from the MONICA (Monitoring Trends and Determinants in Cardiovascular Disease) Augsburg Cohort Study, 1984 to 1992.** *Circulation* 1999, 99:237–242.
10. Ridker PM, Rifai N, Rose L, et al.: **Comparison of C-reactive protein and low-density lipoprotein cholesterol levels in the prediction of first cardiovascular events.** *N Engl J Med* 2002, 347:1557–1565.
11. Visser M, Bouter LM, McQuillan GM, et al.: **Elevated C-reactive protein levels in overweight and obese adults.** *JAMA* 1999, 282:2131–2135.
12. O'Brien KD, Brehm BJ, Seeley RJ, et al.: **Diet-induced weight loss is associated with decreases in plasma serum amyloid A and C-reactive protein independent of dietary macronutrient composition in obese subjects.** *J Clin Endocrinol Metab* 2005, In press.

This is one of three recent papers to demonstrate that weight loss is associated with reduction in SAA levels, and also is notable for demonstrating that in obese subjects weight loss is a more important regulator of plasma SAA levels than are dietary fat and cholesterol content.

- 13.● Tannock LR, O'Brien KD, Knopp RH, et al.: **Cholesterol feeding increases C-reactive protein and serum amyloid A levels in lean insulin-sensitive subjects.** *Circulation* 2005, **111**:3058–3062.

This paper confirms that obesity and insulin resistance appear to protect from the effect of dietary cholesterol in raising SAA levels, but is important because it demonstrates that lean patients are particularly susceptible to the effect of dietary cholesterol in raising SAA (and CRP levels).

14. Festa A, D'Agostino R, Howard G, et al.: **Chronic subclinical inflammation as part of the insulin resistance syndrome: the Insulin Resistance Atherosclerosis Study (IRAS).** *Circulation* 2000, **102**:42–47.
15. McLaughlin T, Abbasi F, Lamendola C, et al.: **Differentiation between obesity and insulin resistance in the association with C-reactive protein.** *Circulation* 2002, **106**:2908–2912.
16. Leinonen E, Hurt-Camejo E, Wiklund O, et al.: **Insulin resistance and adiposity correlate with acute-phase reaction and soluble cell adhesion molecules in type 2 diabetes.** *Atherosclerosis* 2003, **166**:387–394.
17. Weiss R, Dziura J, Burgert TS, et al.: **Obesity and the metabolic syndrome in children and adolescents.** *N Engl J Med* 2004, **350**:2362–2374.
18. Fredrikson GN, Hedblad B, Nilsson JA, et al.: **Association between diet, lifestyle, metabolic cardiovascular risk factors, and plasma C-reactive protein levels.** *Metabolism* 2004, **53**:1436–1442.
19. Redberg RF, Rifai N, Gee L, Ridker PM: **Lack of association of C-reactive protein and coronary calcium by electron beam computed tomography in postmenopausal women: implications for coronary artery disease screening.** *J Am Coll Cardiol* 2000, **36**:39–43.
20. Hunt ME, O'Malley PG, Vernalis MN, et al.: **C-reactive protein is not associated with the presence or extent of calcified subclinical atherosclerosis.** *Am Heart J* 2001, **141**:206–210.
21. Johnson BD, Kip KE, Marroquin OC, et al.: **Serum amyloid A as a predictor of coronary artery disease and cardiovascular outcome in women—The National Heart, Lung, and Blood Institute-sponsored Women's Ischemia Syndrome Evaluation (WISE).** *Circulation* 2004, **109**:726–732.
22. Ridker PM, Cannon CP, Morrow D, et al.: **C-reactive protein levels and outcomes after statin therapy.** *N Engl J Med* 2005, **352**:20–28.
23. Nissen SE, Tuzcu EM, Schoenhagen P, et al.: **Statin therapy, LDL cholesterol, C-reactive protein, and coronary artery disease.** *N Engl J Med* 2005, **352**:29–38.
24. Yusuf S, Hawken S, Ounpuu S, et al.: **Effect of potentially modifiable risk factors associated with myocardial infarction in 52 countries (the INTERHEART study): case-control study.** *Lancet* 2004, **364**:937–952.
25. Danesh J, Wheeler JG, Hirschfield GM, et al.: **C-reactive protein and other circulating markers of inflammation in the prediction of coronary heart disease.** *N Engl J Med* 2004, **350**:1387–1397.
26. Pearson TA, Mensah GA, Alexander RW, et al.: **Markers of inflammation and cardiovascular disease: application to clinical and public health practice: A statement for healthcare professionals from the Centers for Disease Control and Prevention and the American Heart Association.** *Circulation* 2003, **107**:499–511.
27. Bogaty P, Brophy JM, Boyer L, et al.: **Fluctuating inflammatory markers in patients with stable ischemic heart disease.** *Arch Intern Med* 2005, **165**:221–226.
28. Pasceri V, Willerson JT, Yeh ET: **Direct proinflammatory effect of C-reactive protein on human endothelial cells.** *Circulation* 2000, **102**:2165–2168.
29. Pasceri V, Cheng JS, Willerson JT, Yeh ET: **Modulation of C-reactive protein-mediated monocyte chemoattractant protein-1 induction in human endothelial cells by anti-atherosclerosis drugs.** *Circulation* 2001, **103**:2531–2534.
30. Devaraj S, Kumaresan PR, Jialal I: **Effect of C-reactive protein on chemokine expression in human aortic endothelial cells.** *J Mol Cell Cardiol* 2004, **36**:405–410.
31. Devaraj S, Xu DY, Jialal I: **C-reactive protein increases plasminogen activator inhibitor-1 expression and activity in human aortic endothelial cells: implications for the metabolic syndrome and atherothrombosis.** *Circulation* 2003, **107**:398–404.
32. Singh U, Devaraj S, Jialal I: **C-reactive protein decreases tissue plasminogen activator activity in human aortic endothelial cells. evidence that c-reactive protein is a procoagulant.** *Arterioscler Thromb Vasc Biol* 2005, In press.
33. Venugopal SK, Devaraj S, Jialal I: **C-reactive protein decreases prostacyclin release from human aortic endothelial cells.** *Circulation* 2003, **108**:1676–1678.
34. Venugopal SK, Devaraj S, Yuhanna I, et al.: **Demonstration that C-reactive protein decreases eNOS expression and bioactivity in human aortic endothelial cells.** *Circulation* 2002, **106**:1439–1441.
35. Verma S, Wang CH, Li SH, et al.: **A self-fulfilling prophecy: C-reactive protein attenuates nitric oxide production and inhibits angiogenesis.** *Circulation* 2002, **106**:913–919.
36. Clapp BR, Hirschfield GM, Storry C, et al.: **Inflammation and endothelial function: direct vascular effects of human C-reactive protein on nitric oxide bioavailability.** *Circulation* 2005, **111**:1530–1536.
37. Pepys MB: **CRP or not CRP? That is the question.** *Arterioscler Thromb Vasc Biol* 2005, **25**:1091–1094.
- 38.●● Hirschfield GM, Gallimore JR, Kahan MC, et al.: **Transgenic human C-reactive protein is not proatherogenic in apolipoprotein E-deficient mice.** *Proc Natl Acad Sci U S A* 2005, **102**:8309–8314.
- This paper is the first of a series of three recent studies demonstrating no effect on atherosclerosis by over-expression of CRP in mouse models of atherosclerosis. This observation seriously calls into question whether CRP mediates atherogenesis in vivo.
- 39.● Trion A, de Maat MP, Jukema JW, et al.: **No effect of C-reactive protein on early atherosclerosis development in apolipoprotein E*3-leiden/human C-reactive protein transgenic mice.** *Arterioscler Thromb Vasc Biol* 2005, **25**:1635–1640.
- This paper finds no effect of transgenic expression of human CRP on atherosclerotic lesion size in the apoE*3-Leiden mouse model of atherosclerosis. This extends the observation that CRP does not promote atherosclerosis in vivo to a mouse atherosclerosis model other than apoE-deficient mice.
- 40.● Reifenberg K, Lehr HA, Baskal D, et al.: **Role of C-reactive protein in atherogenesis: can the apolipoprotein E knock-out mouse provide the answer?** *Arterioscler Thromb Vasc Biol* 2005, **25**:1641–1646.
- This paper finds no effect of transgenic expression of rabbit CRP on atherosclerotic lesion size in apoE-deficient mice.
41. Uhlar CM, Whitehead AS: **Serum amyloid A, the major vertebrate acute-phase reactant.** *Eur J Biochem* 1999, **265**:501–523.
42. Meek RL, Urieli-Shoval S, Benditt EP: **Expression of apolipoprotein serum amyloid A mRNA in human atherosclerotic lesions and cultured vascular cells: implications for serum amyloid A function.** *Proc Natl Acad Sci U S A* 1994, **91**:3186–3190.
43. Ridker PM, Hennekens CH, Buring JE, Rifai N: **C-reactive protein and other markers of inflammation in the prediction of cardiovascular disease in women.** *N Engl J Med* 2000, **342**:836–843.
44. Jousilahti P, Salomaa V, Rasi V, et al.: **The association of c-reactive protein, serum amyloid A and fibrinogen with prevalent coronary heart disease—baseline findings of the PAIS project.** *Atherosclerosis* 2001, **156**:451–456.
45. Ebeling P, Teppo AM, Koistinen HA, et al.: **Troglitazone reduces hyperglycaemia and selectively acute-phase serum proteins in patients with Type II diabetes.** *Diabetologia* 1999, **42**:1433–1438.

46. Haffner SM, Agostino RD Jr, Saad MF, et al.: Carotid artery atherosclerosis in type-2 diabetic and nondiabetic subjects with and without symptomatic coronary artery disease (The Insulin Resistance Atherosclerosis Study). *Am J Cardiol* 2000, 85:1395-1400.
47. Liao F, Andalibi A, deBeer FC, et al.: Genetic control of inflammatory gene induction and NF-kappa B-like transcription factor activation in response to an atherogenic diet in mice. *J Clin Invest* 1993, 91:2572-2579.
- 48.● O'Brien KD, McDonald TO, Kunjathoor V, et al.: Serum amyloid A and lipoprotein retention in murine models of atherosclerosis. *Arterioscler Thromb Vasc Biol* 2005, In press. This paper demonstrates that SAA co-localizes with HDL apolipoproteins (and with VLDL apolipoproteins) in atherosclerotic lesions at all stages of lesion development in two mouse models of atherosclerosis. It further demonstrates that SAA containing HDL binds more avidly to vascular proteoglycans in vitro. Taken together, these observations support the possibility that SAA may promote atherosclerosis by mediating plaque lipoprotein retention.
- 49.●● Lewis KE, Kirk EA, McDonald TO, et al.: Increase in serum amyloid A evoked by dietary cholesterol is associated with increased atherosclerosis in mice. *Circulation* 2004, 110:540-545. This paper demonstrates not only that SAA co-localizes with apolipoprotein A-I in murine atherosclerotic lesions, but also that plasma SAA levels correlate with atherosclerotic lesion size. These are probably the strongest in vivo data to date supporting a possible role for SAA as an atherosclerosis mediator.
- 50.●● Poitou C, Viguier N, Canello R, et al.: Serum amyloid A: production by human white adipocyte and regulation by obesity and nutrition. *Diabetologia* 2005, 48:519-528. This is one of two recent papers establishing convincingly that adipocytes produce SAA. It also demonstrates that adipose tissue SAA expression correlates with plasma SAA levels and that weight loss is associated with decrease in adipose tissue and plasma SAA.
- 51.●● Sjöholm K, Palming J, Olofsson LE, et al.: A microarray search for genes predominantly expressed in human omental adipocytes: adipose tissue as a major production site of serum amyloid A. *J Clin Endocrinol Metab* 2005, 90:2233-2239. This is the second of two recent papers demonstrating that adipose tissue can produce SAA. The authors also demonstrate that diet-induced weight loss is associated with a decrease in plasma SAA levels.
52. Miettinen TA, Kesaniemi YA: Cholesterol absorption: regulation of cholesterol synthesis and elimination and within-population variations of serum cholesterol levels. *Am J Clin Nutr* 1989, 49:629-635.
53. Simonen P, Gylling H, Howard AN, Miettinen TA: Introducing a new component of the metabolic syndrome: low cholesterol absorption. *Am J Clin Nutr* 2000, 72:82-88.
54. Weisberg SP, McCann D, Desai M, et al.: Obesity is associated with macrophage accumulation in adipose tissue. *J Clin Invest* 2003, 112:1796-1808.
55. Xu H, Barnes GT, Yang Q, et al.: Chronic inflammation in fat plays a crucial role in the development of obesity-related insulin resistance. *J Clin Invest* 2003, 112:1821-1830.
56. Kisilevsky R, Subrahmanyam L: Serum amyloid A changes high density lipoprotein's cellular affinity. A clue to serum amyloid A's principal function [published erratum appears in *Lab Invest* 1992 Jul;67(1):151]. *Lab Invest* 1992, 66:778-785.
57. Hayat S, Raynes JG: Acute phase serum amyloid A protein increases high density lipoprotein binding to human peripheral blood mononuclear cells and an endothelial cell line. *Scand J Immunol* 2000, 51:141-146.
58. Artl A, Marsche G, Lestavel S, et al.: Role of serum amyloid A during metabolism of acute-phase HDL by macrophages. *Arterioscler Thromb Vasc Biol* 2000, 20:763-772.
59. Migita K, Kawabe Y, Tominaga M, et al.: Serum amyloid A protein induces production of matrix metalloproteinases by human synovial fibroblasts. *Lab Invest* 1998, 78:535-539.
60. Strissel KJ, Girard MT, West-Mays JA, et al.: Role of serum amyloid A as an intermediate in the IL-1 and PMA-stimulated signaling pathways regulating expression of rabbit fibroblast collagenase. *Exp Cell Res* 1997, 237:275-287.
61. Badolato R, Wang JM, Murphy WJ, et al.: Serum amyloid A is a chemoattractant: induction of migration, adhesion, and tissue infiltration of monocytes and polymorphonuclear leukocytes. *J Exp Med* 1994, 180:203-209.
62. Xu L, Badolato R, Murphy WJ, et al.: A novel biologic function of serum amyloid A. Induction of T lymphocyte migration and adhesion. *J Immunol* 1995, 155:1184-1190.
63. Husebekk A, Skogen B, Husby G: Characterization of amyloid proteins AA and SAA as apolipoproteins of high density lipoprotein (HDL). Displacement of SAA from the HDL-SAA complex by apo AI and apo AII. *Scand J Immunol* 1987, 25:375-381.
64. Tam SP, Flexman A, Hulme J, Kisilevsky R: Promoting export of macrophage cholesterol: the physiological role of a major acute-phase protein, serum amyloid A 2.1. *J Lipid Res* 2002, 43:1410-1420.
65. Ancsin JB, Kisilevsky R: The heparin/heparan sulfate-binding site on apo-serum amyloid A. Implications for the therapeutic intervention of amyloidosis. *J Biol Chem* 1999, 274:7172-7181.
66. Kunjathoor VV, Chiu DS, O'Brien KD, LeBoeuf RC: Accumulation of biglycan and perlecan, but not versican, in lesions of murine models of atherosclerosis. *Arterioscler Thromb Vasc Biol* 2002, 22:462-468.
67. O'Brien KD, Olin KL, Alpers CE, et al.: Comparison of apolipoprotein and proteoglycan deposits in human coronary atherosclerotic plaques: colocalization of biglycan with apolipoproteins. *Circulation* 1998, 98:519-527.
68. Pennathur S, Bergt C, Shao B, et al.: Human atherosclerotic intima and blood of patients with established coronary artery disease contain high density lipoprotein damaged by reactive nitrogen species. *J Biol Chem* 2004, 279:42977-42983.
69. Bergt C, Pennathur S, Fu X, et al.: The myeloperoxidase product hypochlorous acid oxidizes HDL in the human artery wall and impairs ABCA1-dependent cholesterol transport. *Proc Natl Acad Sci U S A* 2004, 101:13032-13037.
70. Shao B, Fu X, McDonald TO, et al.: Acrolein impairs ABCA1-dependent cholesterol export from cells through site-specific modification of apolipoprotein A-I. *J Biol Chem* 2005, In press.
71. Kinlay S, Schwartz GG, Olsson AG, et al.: High-dose atorvastatin enhances the decline in inflammatory markers in patients with acute coronary syndromes in the MIRACL study. *Circulation* 2003, 108:1560-1566.
72. Schillinger M, Exner M, Mlekusch W, et al.: Statin therapy improves cardiovascular outcome of patients with peripheral artery disease. *Eur Heart J* 2004, 25:742-748.
73. Marx N, Froehlich J, Siam L, et al.: Antidiabetic PPAR gamma-activator rosiglitazone reduces MMP-9 serum levels in type 2 diabetic patients with coronary artery disease. *Arterioscler Thromb Vasc Biol* 2003, 23:283-288.
74. Skalen K, Gustafsson M, Rydberg EK, et al.: Subendothelial retention of atherogenic lipoproteins in early atherosclerosis. *Nature* 2002, 417:750-754.