

Treatment of Relapsed/Refractory Acute Myeloid Leukemia

Prithviraj Bose, MD¹
Pankit Vachhani, MD²
Jorge E. Cortes, MD^{1,*}

Address

¹Department of Leukemia, University of Texas MD Anderson Cancer Center, 1400 Holcombe Blvd, FC4.3062, Houston, TX, 77030, USA
Email: jcortes@mdanderson.org

²Department of Medicine, Roswell Park Cancer Institute, Buffalo, NY, USA

Published online: 13 March 2017

© Springer Science+Business Media New York 2017

This article is part of the Topical Collection on *Leukemia*

Keywords AML · Epigenetic therapy · Targeted therapy · FLT3 inhibitors · IDH inhibitors · Antibody-drug conjugates

Opinion statement

Approximately 40–45% of younger and 10–20% of older adults with acute myeloid leukemia (AML) will be cured with current standard chemotherapy. The outlook is particularly gloomy for patients with relapsed and/or refractory disease (cure rates no higher than 10%). Allogeneic hematopoietic stem cell transplantation (HSCT), the only realistic hope of cure for these patients, is an option for only a minority. In recent years, much has been learned about the genomic and epigenomic landscapes of AML, and the clonal architecture of both de novo and secondary AML has begun to be unraveled. These advances have paved the way for rational drug development as new “drugable” targets have emerged. Although no new drug has been approved for AML in over four decades, with the exception of gemtuzumab ozogamycin, which was subsequently withdrawn, there is progress on the horizon with the possible regulatory approval soon of agents such as CPX-351 and midostaurin, the Food and Drug Administration “breakthrough” designation granted to venetoclax, and promising agents such as the IDH inhibitors AG-221 and AG-120, the smoothed inhibitor glasdegib and the histone deacetylase inhibitor pracinostat. In our practice, we treat most patients with relapsed/refractory AML on clinical trials, taking into consideration their prior treatment history and response to the same. We utilize targeted sequencing of genes frequently mutated in AML to identify “actionable” mutations, e.g., in *FLT3* or *IDH1/2*, and incorporate small-molecule inhibitors of these oncogenic kinases into our therapeutic regimens whenever possible. In the absence of actionable mutations, we rationally combine conventional agents with other novel therapies such as monoclonal antibodies and other targeted drugs. For fit patients up to the

age of 65, we often use high-dose cytarabine-containing backbone regimens. For older or unfit patients, we prefer hypomethylating agent-based therapy. Finally, all patients with relapsed/refractory AML are evaluated for allogeneic HSCT.

Introduction

The treatment of relapsed/refractory (R/R) acute myeloid leukemia (AML) remains one of the most formidable challenges in oncology today. With a long-term disease-free survival of only approximately 30 to 40% after standard chemotherapy, and a paucity of treatment options besides the standard cytarabine and anthracyclines, disease persistence or recurrence occurs in most patients with AML. Outcomes for patients with relapsed or refractory AML are poor, with overall survival (OS) estimated at no more than 10% at 3 years [1, 2]. In physically fit patients, intensive chemotherapy (IC) with the goal of achieving a second complete remission (CR2) so as to proceed to allogeneic hematopoietic stem cell transplantation (HSCT) is often given, but long-term survival rates remain low, because of the common failure to achieve the necessary response required to achieve the desired “bridge to transplant” and the comorbidities, age considerations, residual toxicities, limited availability of donors, and other factors that limit this approach for a large majority of patients. Thus, the notion that patients can frequently be salvaged with this approach has been termed a “myth” by some experts [3]. Low-intensity therapies, e.g., epigenetic modifiers, are typically offered to patients who are deemed ineligible to receive IC because of age or comorbidities, as in the

frontline setting, but in the salvage setting are palliative at best in most instances.

In recent years, better understanding of the molecular complexity and biology of AML has led to the development of new strategies to treat AML. Many new agents from different therapeutic classes have been investigated in patients with R/R AML, with particular interest in nonchemotherapeutic strategies. Although no new drugs have been approved for the treatment of AML for over 50 years (with the exception of gemtuzumab ozogamycin, GO, Mylotarg which was eventually withdrawn from the market) [4], the improved OS seen in phase III trials of CPX-351 (Vyxeos™) [5••] and the *fms*-like tyrosine kinase 3 (FLT3) inhibitor, midostaurin, [6••] in secondary and *FLT3*-mutated AML, respectively, in the frontline setting, have made approval of these agents likely. In this manuscript, we review some of the most notable recent advances in the treatment of R/R AML, with a focus on those with the most mature or promising clinical data. It is important to emphasize that the list of agents included is by necessity partial and incomplete. This deficiency, however, highlights the very active and promising ongoing research that will undoubtedly break the impasse that has characterized the treatment of AML in the last 50 years.

Cytotoxic Chemotherapy: Novel Agents and Combinations

Vosaroxin

Vosaroxin is a first-in-class, anti-cancer quinolone derivative that inhibits topoisomerase II and intercalates DNA, inducing cell cycle arrest and p53-independent apoptosis [7]. Preclinical evidence of synergism with cytarabine [8] and encouraging results in an early phase combination trial [9] led to the phase III, placebo-controlled, VALOR study in 711 patients with AML in first relapse after or refractory to 1 or 2 cycles of previous anthracycline-containing induction chemotherapy [10••]. Patients were randomized 1:1 to receive cytarabine (1 g/m² on days 1–5) in combination with vosaroxin (90 mg/m² on days 1 and 4 in cycle 1; 70 mg/m² in subsequent cycles) or placebo. Median OS (the primary endpoint) was 7.5 months in the vosaroxin group and 6.1 months in the placebo group ($p = 0.061$) [10]. Complete remission (CR) rates were significantly higher in the vosaroxin group (30 versus 16%, $p < 0.0001$).

Thirty- and 60-day mortality rates were virtually identical in the two groups, although grade ≥ 3 adverse events (AEs) such as febrile neutropenia (FN), neutropenia, stomatitis, hypokalemia, bacteremia, sepsis, and pneumonia were all more frequent in the vosaroxin cohort [10]. Treatment-related serious AEs (SAEs) occurred in 33 and 17% of patients in the vosaroxin and placebo cohorts, respectively. A number of prespecified secondary and subgroup analyses showed significantly improved OS in the vosaroxin group. Older patients and those with early relapse derived the greatest benefit from vosaroxin and cytarabine [10]. The survival benefit seen in patients ≥ 60 years of age (median 7.1 versus 5 months, $p = 0.003$) was maintained after a median of 39.9 months of follow-up and was particularly pronounced among those with refractory disease or early relapse [11]. Additionally, post hoc, exploratory analyses showed OS benefits in patients with unfavorable cytogenetic abnormalities and those with *FLT3* mutations. The trial did not meet its primary endpoint; thus, vosaroxin has not gained regulatory approval. However, vosaroxin remains an attractive agent for patients with AML. Promising results were reported in a single-arm, nonrandomized study in combination with decitabine in older patients in the frontline setting. This study identified 70 mg/m² of vosaroxin on days 1 and 4 in combination with standard dose decitabine as the preferred dose [12•]. However, the combination of vosaroxin with low-dose cytarabine (LDAC) did not improve CR/complete remission with incomplete count recovery (CRi) rates or survival in a randomized comparison with LDAC alone in older, untreated adults with AML, mainly due to excess early mortality in the combination arm [13]. In this “pick a winner” trial, single-agent vosaroxin was also separately compared to LDAC; CR/CRi rates were similar, and 12-month OS was inferior in the vosaroxin group (12 versus 31%, $p = 0.003$) [13].

CPX-351 (Vyxeos™)

CPX-351 is a liposomal formulation of cytarabine and daunorubicin in a fixed molar ratio of 5:1 that may be particularly effective in secondary AML [14]. A phase II study randomized 125 patients with AML in first relapse after a first complete remission (CR1) of ≥ 1 month 2:1 to CPX-351 or investigators' choice of first salvage chemotherapy [15]. CR/CRi rates were numerically higher with CPX-351, but 1-year OS was not significantly different (36 versus 27%, $p = 0.33$). However, patients categorized as poor risk by the European Prognostic Index ($n = 85$) appeared to benefit from CPX-351 (12-month OS 28 versus 9%, $p = 0.02$) [15]. These findings led to a randomized trial in which 309 patients aged 60 to 75 years with newly diagnosed secondary AML were randomized to receive CPX-351 (Vyxeos™; 100 units/m² on days 1, 3, and 5) or standard chemotherapy (“7 + 3”; cytarabine 100 mg/m²/day on days 1–7 and daunorubicin 60 mg/m²/day on days 1–3). Treatment with CPX-351 resulted in improved OS (the primary endpoint of the study), with a median of 9.56 versus 5.95 months with standard chemotherapy ($p = 0.005$), as well as significantly higher response rates (CR/CRi rate 47.7 versus 33.3%; $p = 0.016$) [5••]. Thirty-day (5.9 versus 10.6%) and 60-day (13.7 versus 21.2%) mortality also favored CPX-351 [5••]. A more detailed discussion of this important first-line trial is outside the scope of this review focused on the treatment of R/R AML.

FLAM

Flavopiridol (alvocidib) is a pan-cyclin-dependent kinase (CDK) inhibitor that induces cell cycle arrest and apoptosis and also transcriptionally down-regulates the antiapoptotic protein, MCL-1, via inhibition of CDK9 [16]. Based on preclinical studies showing synergism with standard chemotherapeutic agents, the regimen of “timed sequential therapy” with flavopiridol, cytarabine, and mitoxantrone (FLAM) was developed [17, 18]. FLAM yielded significantly superior CR rates in comparison to 7 + 3 (70 versus 46%, $p = 0.003$) in a randomized, phase II study in newly diagnosed, younger adults with intermediate and poor risk AML, although no differences in OS or event-free survival (EFS) were observed [19]. Because of flavopiridol’s effects on MCL-1, which is critical to the development and maintenance of AML [20••], an ongoing phase II trial (NCT02520011) of the FLAM regimen in patients with R/R AML is restricted to patients who have evidence of MCL-1 dependence of their AML by a specific BH3-profiling assay that uses the proapoptotic “BH3-only” protein, Noxa, which antagonizes MCL-1 [21]. Flavopiridol has received “orphan drug” designation from the FDA for AML [22].

Epigenetic Therapy: New Agents and Targets

Guadecitabine

Guadecitabine (SGI-110) is a second-generation hypomethylating agent (HMA) that is a dinucleotide of decitabine and deoxyguanosine, making it resistant to deamination by cytidine deaminase [23•]. In a phase 1 trial that enrolled patients with either R/R AML ($n = 74$) or myelodysplastic syndrome (MDS, $n = 19$) and tested various schedules of guadecitabine administered subcutaneously, a maximal tolerated dose (MTD) was not identified in patients with AML, while among those with MDS, 90 mg/m²/day for five consecutive days every 4 weeks was defined as the MTD (DLTs at the 125 mg/m²/day dose level included grade 4 cytopenias and FN, resulting in one case in grade 5 sepsis) [23•]. Potent dose-related DNA demethylation occurred on the daily \times 5 regimen, reaching a plateau at 60 mg/m², which was thus the recommended phase 2 dose (RP2D) [23•]. Six patients each with AML and MDS responded. In a phase II trial, patients with R/R AML received either 10 days ($n = 53$) or 5 days ($n = 50$) of guadecitabine per cycle; additionally, those receiving 5 days were randomized to receive either 60 or 90 mg/m²/day [24]. Patients receiving the drug for 10 days per cycle received 60 mg/m²/day. CR rates were 19 and 6%, respectively, in the 10- and 5-day groups ($p = 0.074$). When comparing the sum of CR, CRi, and complete remission with incomplete platelet recovery (CRp) rates (designated the composite CR (CRc) rate), the percentages were 30 and 16% in the 10- and 5-day groups, respectively ($p = 0.106$). The longer 10-day dosing induced significantly more potent and longer demethylation and led also to significantly more anemia and thrombocytopenia. Median OS was 7.1 months with 10-day dosing and 5.7 months with 5-day dosing ($p = 0.505$). There were no significant differences in 30- or 60-day

mortality between the two schedules [24]. Significantly worse OS was observed among patients with performance status 2, poor risk cytogenetics, and those enrolled within 6 months from their previous therapy [25]. An identically designed phase II trial of guadecitabine was conducted in the frontline setting in patients with AML who were not candidates for IC [26]. There were no major differences in clinical responses, demethylation, OS, or safety between 60 and 90 mg/m²/day on the 5-day regimen [27], or between the 5- and 10-day cohorts for the composite primary endpoint (CR + CRi + CRp), median OS, grade ≥ 3 AEs, or 30-, 60-, and 90-day all-cause mortality; response rates and median OS were numerically higher in the 5-day cohort [26]. Based upon these findings, guadecitabine is currently being tested in treatment-naive patients with AML in a pivotal phase III trial using the 60 mg/m²/day for 5 days schedule (ASTRAL-1, NCT02348489). Based on the upregulation by HMAs of tumor-associated antigens and of programmed death ligand-1/2 (PD-L1/2) as a possible mechanism of escape from immune surveillance, an ongoing trial combines guadecitabine with the anti-PD-L1 monoclonal antibody atezolizumab for the treatment of patients with treatment-naive or R/R AML (NCT02892318).

Other Epigenetic Modifiers

A major focus in epigenetics research in recent years has been on the development of targeted therapies against epigenetic “readers” called bromodomains [28, 29]. Their mechanism of action in AML appears to involve induction of hexamethylene bisacetamide-inducible protein 1 (HEXIM1) [30]. Several small-molecule inhibitors of bromodomain and extra-terminal (BET) proteins have been developed, e.g., FT1101, GSK525762, and MK-8628 (OTX015). In a phase I trial of OTX015 (MK-8628), 3 of 36 patients with R/R AML achieved CR/CRp lasting 2 to 5 months [31]. Preclinical studies have suggested synergism between BET and FLT3 or histone deacetylase (HDAC) inhibitors [32, 33]. Such combinations will be entering the clinic in the near future. Another epigenetic target of interest, especially in mixed lineage leukemia (*MLL*)-rearranged AML, is the histone lysine (H3K79) methyltransferase disruptor of telomeric silencing 1-like (DOT1L) [34••, 35••]. Pinometostat (EPZ-5676) is a potent and highly selective inhibitor of DOT1L currently in early stages of clinical development [36]. In a first-in-human (FIH) phase I trial, an MTD was not identified and pharmacodynamic evidence of target inhibition and clinical responses (including CRs) were observed in heavily pretreated patients with *MLL*-rearranged leukemias across a range of doses [36]. Interestingly, DOT1L may also be a therapeutic target in *DNMT3A*-mutated AML [37], which has been shown to carry an adverse prognosis [38•]. Inhibitors of lysine-specific demethylase 1 (LSD1) may reactivate the all-trans-retinoic acid (ATRA) differentiation pathway in AML, and there is preclinical evidence of synergism between ATRA and the LSD1 inhibitor, tranilcypramine, a regimen that may also target leukemia-initiating cells [39]. A number of clinical trials are exploring ATRA/LSD1 inhibitor

combinations in R/R AML (NCT02842827, NCT02273102, NCT02261779, NCT02717884). Synergism between LSD1 inhibitors and HDAC inhibitors against AML has also been demonstrated, both in vitro and in vivo [40]. In a FIH phase I trial, the first-in-class LSD1 inhibitor, Ory-1001, was found to be well-tolerated; objective responses were seen in 5 of 14 (36%) patients with R/R AML in an extension cohort at a dose of 140 $\mu\text{g}/\text{m}^2/\text{day}$ [41].

Antibody-Drug Conjugates, Bispecific T-Cell Engagers, and Dual Affinity Retargeting Molecules

CD33-Targeted ADCs

GO (Mylotarg®) is an antibody drug conjugate (ADC) targeted to CD33 that consists of a monoclonal antibody against CD33 linked to a semisynthetic derivative of the cytotoxic antibiotic, calicheamicin. It was approved by the FDA in May 2000 for older (≥ 60) patients with CD33-expressing AML in first relapse, based on a 26% overall response rate (ORR) in this population [42•]. Among 142 patients in untreated first relapse (median age 61), the CR/CRp rate was 30% (16% CRs) [42•]. Unfortunately, however, a confirmatory phase III trial conducted by the Southwest Oncology Group (SWOG) found no benefit to the addition of GO to standard “7 + 3” induction chemotherapy in 595 younger (18–60) patients with newly diagnosed AML, in terms of CR rate, relapse-free or overall survival [43•]. Of note, the dose of daunorubicin used in this trial during induction was only 45 $\text{mg}/\text{m}^2/\text{day}$ on days 1–3. There was also no improvement in disease-free survival with the use of GO post-consolidation among patients remaining in CR after consolidation [43•]. In contrast to the findings of the SWOG trial [43•], a number of other phase III trials have documented a survival benefit for this agent with little to no increase in toxicity [44••, 45••, 46••]. The improved OS seen with the addition of GO to IC appears restricted to patients with nonadverse cytogenetics [47•]. An in-depth discussion of these trials in the upfront setting is not presented here, but GO is expected to become available again soon, based on these data.

Like GO, vadastuximab talirine (SGN-33A) is also a CD33-directed ADC that employs pyrrolbenzodiazepine instead of calicheamicin, the toxin attached to the anti-CD33 antibody in GO. Pyrrolbenzodiazepine is not known to be associated with liver toxicity, thus eliminating the potential for hepatic veno-occlusive disease (VOD, also known as sinusoidal obstruction syndrome (SOS)) seen with GO [48•]. Thirty-four patients with AML that had relapsed after achievement of CR1 and 52 who had declined IC (40 of whom had received 1–2 prior low-intensity therapies, predominantly HMAs) participated in a phase I trial of SGN-33A [48•]. Most patients had intermediate (51%) or adverse (31%) cytogenetics, and 54% had AML with MDS-related changes. DLTs included grade 4 hematologic toxicity, grade 3 mucositis and pulmonary embolism, and grade 5 sepsis. The most frequent grade ≥ 3 AEs included febrile neutropenia (FN, 69%), thrombocytopenia (29%), and anemia (23%). Fatigue (48%), decreased appetite (28%), constipation, diarrhea, dyspnea, nausea (26% each), and peripheral edema (25%) were common. Thirty-day mortality was 6%, and 40 mcg/kg every 4 weeks was declared the RP2D. Of 21 patients treated at this dose and evaluable for efficacy, 7 (33%) (3 treatment-naive; 14%) achieved CR/CRi [48•]. A phase I study of vadastuximab talirine

(10 mcg/kg) in combination with an HMA (azacitidine or decitabine) for 5 days every 4 weeks in previously untreated patients with AML who declined or were unfit for IC reported a CR/CRi rate of 73% among evaluable patients ($n = 49$) with 30- and 60-day mortality of 2 and 8%, respectively [49]. This agent has also been combined safely with IC in both the induction [50] and consolidation [51] settings in younger, newly diagnosed and post-remission [51, 54•] patients, respectively, as well as administered alone as maintenance after IC and/or HSCT [51]. In combination with 7 + 3 induction chemotherapy, vadas-tuximab talirine produced a CR/CRi rate of 78%, 94% of which occurred with 1 cycle and 74% of which were MRD-negative, among 40 evaluable, newly diagnosed patients with AML, with 30- and 60-day mortality of 0 and 7%, respectively [50]. IMGN779 is yet another CD33-targeted ADC that utilizes a different toxin (DGN462, a novel alkylating agent); this drug is currently in a phase I trial for patients with R/R AML (NCT02674763).

Anti-CD33 BiTE®s

A novel approach to targeting CD33, expression of which is nearly universal on myeloblasts, uses bispecific T-cell engager (BiTE®) technology. AMG 330 is a CD33/CD3 BiTE antibody that recruits and expands T-cells that efficiently lyse autologous blasts in primary AML samples [52•]. At least in some cell lines, epigenetic modifiers, e.g., panobinostat (HDAC inhibitor) and azacitidine increase CD33 expression and augment AMG 330-induced cytotoxicity [53•]. AMG 330, currently in a FIH phase I trial (NCT02520427), has been shown to significantly prolong survival of xenograft mouse models of human *FLT3*-mutated AML [54•]. Interestingly, in preclinical studies, AMG 330 cytotoxicity against primary AML cells is higher in specimens from newly diagnosed patients and those with favorable-risk disease [55]. AMG 330 strongly upregulates programmed death ligand 1 (PD-L1) on primary AML cells in a cytokine-dependent manner, and blockade of the programmed death 1 (PD-1)/PD-L1 axis significantly enhances AMG 330-mediated cell lysis, T-cell proliferation, and interferon- γ secretion, making a compelling argument for combining this agent with anti-PD-1/PD-L1 monoclonal antibodies [56].

Targeting CD123

The interleukin-3 (IL-3) receptor alpha chain, CD123, is highly and differentially (but not exclusively) expressed in leukemic progenitors compared with normal hematopoietic stem and progenitor cells. JNJ-63709178 is a humanized CD123 x CD3 "DuoBody" that is being studied in a FIH phase I trial in patients with R/R AML (NCT02715011). A conceptually similar but structurally distinct strategy to target CD123 involves the use of dual affinity re-targeting (DART) molecules that are generated from antibodies to CD3 and CD123 and redirect T-cells against AML blasts [57•]. MGD006 is a CD3 x CD123 DART currently in a phase I trial in R/RAML and higher risk MDS (NCT02152956). Xmab®14045 is a bispecific monoclonal antibody that binds to both CD3 on T-cells and CD123 on tumor cells via separate antigen recognition and binding sites, thereby cross-linking them to induce potent cell lysis of CD123⁺ myeloblasts in AML; this agent is now in a broad, phase I trial in patients with R/R CD123⁺ hematologic malignancies (NCT02730312). SL-401 is a fusion protein consisting of a truncated diphtheria toxin conjugated to IL-3 that represents another

mechanism of therapeutically targeting CD123. SL-401 is under phase I/II evaluation in AML, both in the R/R setting (NCT02113982) and as consolidation for patients in CR1 (NCT02270463). In the limited experience reported to date, best response in the R/R AML population has been stable disease (SD) [58]. CSL362 (JNJ-56022473) is an anti-CD123 monoclonal antibody engineered to bind with high affinity to CD16 on natural killer (NK) cells, leading to enhanced antibody-dependent cellular cytotoxicity (ADCC). This agent is now in a phase II clinical trial in combination with decitabine (versus decitabine alone) for previously untreated patients with AML for whom IC is not considered appropriate (NCT02472145).

Molecularly Targeted Therapy: Isocitrate Dehydrogenase 1/2 (IDH1/2) Inhibitors

Mutations in *IDH1* and *IDH2* each occur in approximately 8–12% of patients with AML; are generally associated with older age, intermediate-risk cytogenetics, *FLT3* and nucleophosmin 1 (*NPM1*) mutations; and appear not to have an adverse impact on prognosis [59]. However, these mutant enzymes produce an abnormal oncometabolite, 2-hydroxyglutarate (2-HG), instead of the normal metabolite, α -ketoglutarate (α -KG). 2-HG inhibits the α -KG-dependent enzyme, ten-eleven translocation 2 (TET2), leading to a hypermethylated genome with a resultant block in differentiation [60••]. A number of small-molecule inhibitors of mutant IDH1 and/or IDH2 have been developed, of which AG-120 (IDH1 inhibitor) and AG-221 (IDH2 inhibitor) are the most advanced. IDHENTIFY is a phase III clinical trial of AG-221 versus standard of care for older patients with R/R AML harboring an *IDH2* mutation (NCT02577406). In a phase I/II trial (NCT01915498), 198 patients with an *IDH2*-mutated hematologic malignancy (70% R/R AML, 17% untreated AML, 7% MDS, 6% other) received AG-221 once or twice daily in continuous 28-day cycles [61•]. Most (64%) patients with R/R AML had received ≥ 2 prior therapies. An MTD was not reached. The most common AG-221-related AEs were indirect hyperbilirubinemia (19%) and nausea (18%). Objective responses were seen in 52 of 128 (41%) efficacy-evaluable R/R AML patients (CR in 18%, CRp and CRi in 1% each, morphologic CR in 6% and partial remission (PR) in 15%), with a median response duration of 6 months [61•]. Improvements in the absolute neutrophil count (ANC) occurred in 56% of R/R AML patients. Responses may occur as early as the first cycle and in many instances have been sustained for 24+ months. However, the allelic burden of mutant *IDH2* did not decrease in responders [61•]. A similar trial was conducted in patients with advanced, *IDH1*-mutated, hematologic malignancies using the IDH1 inhibitor AG-120 (NCT02074839) [62•]. A MTD was not reached in the dose escalation phase up to a dose of 1200 mg daily. Most AEs were grade 1 or 2, the most common being diarrhea (23%), fatigue (22%), and fever (22%); FN (11%) was the most frequent grade ≥ 3 AE. The overall response rate (ORR) was 36% with a median duration of 5.6 months, and the CR rate 18% [62•]. Three dose expansion cohorts are currently enrolling at 500 mg daily: R/R AML ($n = 125$), untreated AML ($n = 25$), and other *IDH1*-mutated advanced hematologic malignancies ($n = 25$) [62•]. Plasma levels of 2-HG have been identified as a useful biomarker

of the biologic and clinical activity of these inhibitors as they typically go down significantly upon administration of the IDH inhibitor. An important phenomenon is the development of a differentiation syndrome in the first few weeks of therapy, characterized by a rapid increase in blasts that is followed by maturation with subsequent decrease and a corresponding increase in neutrophils. Both AG-221 and AG-120 are being explored in combination with standard IC (NCT02632708) and azacitidine (NCT02677922), respectively, for fit and unfit adults with newly diagnosed *IDH1/2*-mutated AML. IDH305 and FT-2102 are other inhibitors of mutant IDH1 that are currently undergoing phase I testing in advanced malignancies (including AML, NCT02381886) and AML (NCT02719574) with *IDH1* R132 mutations, respectively. Preliminary data from an ongoing phase I trial in patients with advanced, *IDH1*-mutated malignancies suggest that IDH305 has a favorable safety profile and promising antitumor activity in *IDH1*-mutated R/R AML (ORR = 33%, $n = 21$) [63]. AG-881 is an inhibitor of both mutant IDH1 and IDH2 that exhibits excellent blood-brain barrier penetration and is under investigation in separate phase I trials in *IDH1/2*-mutated advanced hematologic malignancies (NCT02492737) and solid tumors (e.g., gliomas, cholangiocarcinoma, NCT02481154).

Molecularly Targeted Therapy: FLT3 Inhibitors

FLT3 mutations are encountered in approximately 30–35% of cases of AML. These include internal tandem duplications (ITD) that occur in approximately 25% of patients with AML, and point mutations, most frequently at residue D835 affecting the tyrosine kinase domain (TKD), in 5–10% [64]. While *FLT3*-ITD is clearly associated with high relapse rates and poor outcomes, the prognostic significance of TKD mutations is less well understood and inconsistent across studies [64]. Remarkably, although the subject of intense investigation for many years, no *FLT3* inhibitor is specifically approved for AML, although this is expected to change in light of positive OS data with midostaurin from the RATIFY trial (discussed below) [6••]. A number of *FLT3* tyrosine kinase inhibitors (TKIs) have been developed, and these may be subdivided into type I (e.g., crenolanib, midostaurin) or type II (e.g., sorafenib, quizartinib, ponatinib) inhibitors depending on the conformation of the kinase to which they bind (Table 1) [65]. The five *FLT3* inhibitors with the most clinical data available are discussed individually below. Other *FLT3* inhibitors, including FLX925 and E6201, are in earlier phases of clinical testing (NCT02335814, NCT02418000).

Sorafenib

The multi-kinase inhibitor sorafenib (Nexavar®), approved for the treatment of advanced hepatocellular and renal cell carcinoma, is a potent inhibitor of *FLT3*-ITD but not wild-type *FLT3* or D835-mutated *FLT3* [66]. Its efficacy was first reported in the phase 1 setting, demonstrating rapid reduction in blast count and some instances of CRi, but rarely CR, and with typically short-lived responses. Studies of sorafenib in conjunction with LDAC in the upfront setting in unselected patients with AML resulted in modest clinical benefit [67–69]. Encouraging results were reported in a phase I/II trial ($n = 43$; 40 of them with *FLT3*-ITD, 6 not previously treated) of sorafenib, 400 mg twice daily

Table 1. Major FLT-3 inhibitors, along with their kinase and efficacy and safety data in relapsed/refractory AML

FLT3 inhibitor	Kinases inhibited	Efficacy in major phase I/II trials	Major adverse events
Sorafenib (BAY 43-9006, Bayer/Onyx)	FLT3 ITD; VEGFR, RAF, RET, c-KIT, PDGFR- β	Sorafenib with azacitidine in AML (93% FLT3-ITD ⁺ , 84% of evaluable pts had R/R AML): ORR = 46% (CR + CRi = 43%, PR = 3%), OS in responders 7.8 m, in nonresponders 6.0 m; 16% bridged to HSCT [70•].	All grades: fatigue (47%), rash and diarrhea (20–30%), hyperbilirubinemia (~60%), AST/ALT elevation (30–40%). Most common grade 3/4: cytopenias, FN[70•].
Midostaurin (PKC412, Novartis)	FLT3 WT, ITD, TKD; VEGFR-2, PKC, c-KIT, PDGFR- β	Midostaurin with azacitidine in AML/MDS (94% with AML; 74% FLT3-ITD ⁺ , 76% previously treated, 24% had received prior FLT3 inhibitor): ORR = 26% (CR + CRi + MLFS = 24%, PR = 2%), ORR = 33% in FLT3-ITD ⁺ FLT3 inhibitor-naïve pts. OS = 22 weeks [78].	Grade 3/4: Neutropenia (96%), thrombocytopenia (94%), anemia (61%), infections (56%), LVEF reduction (11%) [78].
Quizartinib (AC-220, Daiichi Sankyo)	FLT3 WT, ITD; c-KIT, PDGFR- α	Quizartinib in R/R AML: (1) \geq 60 yrs., relapsed in <1 year or refractory to first-line chemo: for FLT3-ITD ⁺ , ORR = 66%, OS = 25.3 weeks; for FLT3-ITD ⁻ , ORR = 32%, OS = 19 weeks [82]. (2) \geq 18 years, R/R to second-line chemo or HSCT: for FLT3-ITD ⁺ , ORR = 68%, OS = 23.1 weeks; for FLT3-ITD ⁻ , ORR = 47%, OS = 25.6 weeks. ~1/3 of pts successfully bridged to HSCT [83].	Grade 3/4: anemia (~25%), neutropenia (12%), FN (20–25%), thrombocytopenia (12–15%), QT prolongation (10–13%; 3% at 30 or 60 mg daily dose in another study) [82, 83, 85].
Grenolanib (CP-868-596, AROG)	FLT3 WT, ITD, TKD; PDGFR	Crenolanib in R/R FLT3 ⁺ AML: (1) No prior FLT3 TKI: CRi = 39%, PR = 11%, OS = 234 days. (2) Prior FLT3 TKI: ORR = 31%, OS = 94 days [94•].	All grades: nausea/vomiting, transaminitis, and fluid retention [94•].
Gilteritinib (ASP2215, Astellas)	FLT3 ITD, TKD; AXL	Gilteritinib in R/R FLT3 ⁺ AML: for doses \geq 80 mg, overall ORR = 52% (CRc = 41%, PR = 11%). ORR = 56% in FLT3 inhibitor-naïve pts vs. 42% in prior FLT3 inhibitor-treated pts. ORR = 55% in FLT3-ITD AML. OS ~31 weeks [99•].	All grades: diarrhea (16%), fatigue (15%) [99•].

CR complete remission, CRi CR with incomplete bone marrow recovery, CRc composite CR, LVEF left ventricular ejection fraction, HSCT hematopoietic stem cell transplant, MLFS morphologic leukemia-free state, ORR objective response rate, OS overall survival, PR partial remission, R/R relapsed/refractory, FN febrile neutropenia, ALT alanine aminotransferase, AST aspartate aminotransferase, ITD internal tandem duplication, TKD tyrosine kinase domain, VEGFR vascular endothelial growth factor receptor, PDGFR platelet-derived growth factor receptor, WT wild type, PKC protein kinase C, TKI tyrosine kinase inhibitor, OS overall survival, AML acute myeloid leukemia, MDS myelodysplastic syndromes, pts patients

continuously, administered in combination with azacitidine, 75 mg/m²/day on days 1–7 of each monthly cycle [70•]. The ORR was 46%, with 16% CR, 27% CRi, and 3% PR and a median time to response of 2 months (2 cycles). The median duration of response (DOR) was 2.3 months, and six patients were bridged to allogeneic HSCT [70•]. The commercial availability of both sorafenib and azacitidine makes this a commonly used regimen for patients with R/R (or newly diagnosed if unfit for IC) *FLT3*-ITD⁺ AML who are unable to enroll on a clinical trial of a *FLT3* inhibitor. While the addition of sorafenib to IC has been reported to yield CR/CRp rates as high as 95% among newly diagnosed younger patients with *FLT3*-ITD⁺ AML [71], two placebo-controlled trials of the addition of sorafenib to IC in unselected patients with previously untreated AML reached opposite conclusions. Serve et al. found no benefit of adding sorafenib to 7 + 3 induction chemotherapy followed by up to 2 cycles of intermediate-dose cytarabine consolidation among elderly patients with AML, including in the *FLT3*-ITD⁺ subgroup, and higher toxicity during induction in the sorafenib arm [72]. In contrast, Rollig et al. reported significantly improved EFS, at the expense of increased toxicity, with the addition of sorafenib to 7 + 3 induction followed by 3 cycles of high-dose cytarabine (HiDAC) consolidation in a trial in younger subjects with AML that also used “maintenance” sorafenib for 12 months [73••]. Maintenance sorafenib may have a role in decreasing the risk of relapse after allogeneic HSCT for patients with R/R *FLT3*-ITD⁺ AML [74].

Midostaurin

Midostaurin (formerly PKC412) is another multi-kinase inhibitor that inhibits *FLT3* and is active in patients with both *FLT3*-mutated and wild-type AML [75, 76]. Midostaurin 75 mg three times daily induced a ≥50% reduction in circulating and/or bone marrow (BM) blast count in 14 of 20 patients (70%) with *FLT3*-mutated R/R AML or high-grade MDS who were not candidates for IC [75]. In a phase IIb trial, 95 patients with AML or MDS with either wild-type ($n = 60$) or mutated ($n = 35$) *FLT3* were randomized to receive oral midostaurin, 50 or 100 mg twice daily [76]. Seventy-one percent of patients with *FLT3*-mutant and 42% of those with *FLT3* wild-type disease achieved ≥50% reduction in circulating or BM blasts [76]. Midostaurin was well-tolerated, and there were no differences in toxicity or response rate according to the dose of midostaurin.

Preclinical studies showed synergism between hypomethylating agents and midostaurin against *FLT3*-ITD AML [77]. The combination of midostaurin and azacitidine was evaluated in a phase I/II study ($n = 54$) in patients with AML (95%) or MDS (5%) [78]. Although patients were eligible regardless of their mutation status, 74% of patients had a *FLT3* mutation (68% ITD alone, 6% with both ITD and D835). Seventy-six percent had received prior therapy (median, 2 prior regimens; 43% had received a HMA and 24% a *FLT3* TKI). The ORR was 26% (CR and PR in 1 patient each, CRi in 6 patients and a morphologic leukemia-free state (MLFS) in 6). Among patients with *FLT3*-ITD not previously treated with *FLT3* inhibitors, the ORR was 33%. Overall, 79% of patients had a reduction (median 68% reduction) in BM blast percentage from baseline, with 53% of patients experiencing a ≥50% reduction. The combination was well-tolerated, with some reduction (median 15%) in cardiac ejection fraction (EF) seen in 6 (11%) patients, all of whom had predisposing factors

[78]. A phase I study was also performed in 16 patients with AML (8 newly diagnosed, 8 relapsed, 2 with *FLT3*-ITD) of the combination of midostaurin and decitabine [77]. Based on the clinical activity observed in a phase IB trial evaluating the combination of midostaurin with 7 + 3 induction and HiDAC consolidation in younger, newly diagnosed patients with AML (80% CR rate with midostaurin 50 mg twice daily, 74% in *FLT3*-wild type and 92% in *FLT3*-mutated patients) [79], the phase III RATIFY (Cancer and Leukemia Group B 10603) trial ($n = 717$) was conducted and recently reported. All patients had *FLT3* mutations, and midostaurin (50 mg twice daily) or placebo was administered on days 8–22 during both 7 + 3 induction (daunorubicin, 60 mg/m² on days 1–3, and cytarabine, 200 mg/m² on days 1–7) and HiDAC consolidation (3 g/m² every 12 h on days 1, 3, and 5), as well as maintenance for a year [6••]. Despite similar CR rates in the two arms, the trial showed statistically significant benefits in both EFS and OS for patients treated with midostaurin across *FLT3* mutation subtypes. While a detailed discussion of this frontline study is outside the scope of this review, these results establish midostaurin in combination with standard chemotherapy as standard therapy for younger, newly diagnosed patients with *FLT3*-ITD AML [6••].

Quizartinib

Quizartinib (formerly AC220) is a potent *FLT3* inhibitor with increased selectivity for this kinase [80]. Inhibition of other kinases such as c-KIT requires approximately tenfold higher concentrations in preclinical studies. In a FIH phase I study, 76 patients with R/R AML received quizartinib, irrespective of *FLT3* mutation status [81]. 23 (30%) patients responded, with 2 achieving CR, 3 CRp, 5 CRi and 13 PR. The ORR was 53% among patients with *FLT3*-ITD patients (one CR, one CRp, two CRis, five PRs), 14% among *FLT3*-ITD⁻ negative patients and 22% among those with unknown *FLT3* status. As with other *FLT3* inhibitors, responses to single-agent quizartinib are typically transient. The DLT was QT prolongation, and the MTD was 200 mg/day. Drug-related AEs that occurred at a frequency of >10% included nausea (16%), prolonged QT interval (12%), vomiting (11%), and dysgeusia (11%); most were grade ≤2 [81]. In a large phase II study ($n = 333$), the safety and efficacy of quizartinib monotherapy was examined in two cohorts: older (≥60) patients with AML that had relapsed within a year or was refractory to first-line chemotherapy [82], and younger adults with AML R/R to second-line salvage chemotherapy or relapsed after HSCT [83]. In both cohorts, high rates of composite CR (CRc = CR + CRp + CRi) were observed for *FLT3*-ITD⁺ patients (54% in the older cohort and 44% in the younger cohort); importantly, approximately one third of patients in the younger cohort (8% in the older cohort) were successfully bridged to HSCT [82, 83]. A comparison of the outcomes of 97 *FLT3*-ITD⁺ patients from the younger cohort with those of 183 matched patients from the UK Medical Research Council/National Cancer Research Institute database suggested an improvement in OS in the patients receiving quizartinib [84]. In an effort to minimize the incidence of grade ≥3 QT prolongation with this agent (13 and 10%, respectively, in the older and younger cohorts above), 76 patients with R/R *FLT3*-ITD⁺ AML were randomized 1:1 to receive either 30 or 60 mg daily of quizartinib in another phase II study. These reduced doses yielded response rates nearly identical to those reported with higher doses, with CRc of

approximately 50% in both arms, while the incidence of grade 3 QT prolongation decreased significantly to 3% at the lowest dose [85]. In this trial, 29% of patients treated at 30 mg/day and 37% of those receiving 60 mg/day (33% overall) were successfully bridged to HSCT. Quizartinib is now being investigated in a pivotal phase III trial versus salvage chemotherapy in subjects with *FLT3*-ITD AML refractory to prior therapy or in early first relapse (QUANTUM-R, NCT02039726), and a placebo-controlled, phase III study of quizartinib in combination with IC for previously untreated, younger (18–75) patients with *FLT3*-ITD AML (QuANTUM-First, NCT02668653). It is also being studied in a phase II study in combination with azacitidine or LDAC in older (≥ 60) patients with newly diagnosed, *FLT3*-ITD⁺ myeloid leukemias or in patients of any age receiving first salvage treatment (NCT01892371). The phase I portion of this study enrolled patients with R/R AML, MDS, or chronic myelomonocytic leukemia (CMML) irrespective of *FLT3* mutation and salvage status. Results in the first 52 patients (12 in phase I, 40 in phase II; 38 in the azacitidine arm, 14 in the LDAC arm) included an ORR of 67% overall and 73% in patients with *FLT3*-ITD ($n = 48$) [86]. Like sorafenib, quizartinib has been studied in the post-transplant maintenance setting in patients with *FLT3*-ITD⁺ AML and reported to reduce relapse rates in comparison to historical controls [87].

Crenolanib

The emergence of resistance-conferring point mutations in the kinase domain of *FLT3*, e.g., at the D835 residue, under the selective pressure of type II *FLT3* inhibitors (e.g., sorafenib, quizartinib) occurs in approximately 20–25% of patients [88, 89••, 90]. Molecular docking studies have suggested that D835 mutations primarily confer resistance by stabilizing an active Asp-Phe-Gly in (“DFG-in”) kinase conformation unfavorable to the binding of type II *FLT3* TKIs, which target a “DFG-out” inactive conformation [65]. Crenolanib is a next-generation, type I, pan-*FLT3* inhibitor with activity against TKD point mutations, including D835 and F691, that confer resistance to highly potent type II inhibitors like quizartinib [91•, 92•, 93•]. Crenolanib (100 mg three times daily or 200 mg/m²/day in three divided doses) was tested in a phase II study in patients with R/R *FLT3*-mutated AML [94•]. Among 18 de novo AML patients who were *FLT3*-TKI-naïve, the ORR was 50% (39% CRi + 11% PR) and median OS was 234 days. Thirty-six de novo AML patients had received prior *FLT3* inhibitor therapy; the ORR in this cohort was 31% (17% CRi + 14% PR) and median OS 94 days. Crenolanib provided only transient benefit to patients with secondary *FLT3*-mutated AML ($n = 11$, median OS 55 days) [94•]. The safety of adding crenolanib, 100 mg three times daily, to both idarubicin/HiDAC (in the R/R setting) and 7 + 3 induction plus HiDAC consolidation (in the upfront setting) has been demonstrated [95, 96]. In the former study, 13 patients received idarubicin (12 mg/m²/day for 3 days) plus HiDAC (1.5 g/m²/day for 4 days (for 3 days if over 60 years of age), followed by crenolanib [95]. Responding patients could proceed to HSCT or receive consolidation with cytarabine (750 mg/m²/day for 3 days) and idarubicin (8 mg/m²/day for 2 days), followed by crenolanib. Patients could then continue on maintenance crenolanib unless they received an allograft. No DLTs were observed at any of the dose levels explored, and no dose reductions were required [95]. All non-hematologic AEs were grade 1 in severity. The ORR among 11 evaluable patients

was 36%. Among the six patients who had received ≤ 2 prior AML therapies, the ORR was 67% (all CR or CRi) and median OS was 259 days [95]. Several studies are ongoing investigating crenolanib in combination with IC in newly diagnosed (NCT02283177) and R/R (NCT02400281, NCT02298166) *FLT3*-mutated AML. NCT02400281 also contains an arm in which crenolanib is combined with azacitidine for patients with R/R *FLT3*-mutated AML. Additionally, single-agent crenolanib is being investigated as maintenance post-HSCT (NCT02400255).

Gilteritinib

Gilteritinib (formerly ASP-2215) is a highly selective *FLT3* and *AXL* inhibitor that displays activity against both *FLT3*-ITD and D835 mutations, as well as the “gatekeeper” mutation F691 with minimal activity against wild-type *FLT3* [97]. In a FIH phase I/II trial (NCT02014558) in patients with R/R AML, the drug was found to be well-tolerated across a range of doses (20–300 mg), and antileukemic activity was seen in patients with *FLT3*-mutated AML treated at doses ≥ 80 mg daily [98]. Several dose levels were then expanded with subjects with *FLT3*-mutated AML. Out of a total of 252 patients with R/R AML in this trial (Chrysalis), 159 had *FLT3*-ITD, 13 had a D835 mutation, and 16 had both. Diarrhea (16%) and fatigue (15%) were the most common treatment-related AEs of any grade [99•]. Only 11 patients (<5%) had a maximum post-baseline-corrected QT interval >500 ms. Among 169 patients with *FLT3*-mutated AML who received doses ≥ 80 mg/day, the ORR was 52% (55% in *FLT3*-ITD⁺ subjects, 17% in D835 mutant subjects, and 62% in subjects with both mutations). The ORR was higher (56%) in patients without prior TKI exposure than among those with prior TKI exposure (42%). Median OS was ~ 31 weeks [99•]. There are a number of ongoing studies of gilteritinib in *FLT3*-mutant AML, including a phase III, randomized trial which is investigating the efficacy of gilteritinib versus salvage chemotherapy in the R/R setting (NCT02421939). In newly diagnosed AML patients, gilteritinib is being investigated in combination with IC without regard to *FLT3* mutational status (NCT02236013, NCT02310321) and with azacitidine (NCT02752035) for those with mutant *FLT3* not eligible for IC. Finally, for patients with *FLT3* ITD AML, it is also being studied as maintenance therapy in first CR after induction and consolidation therapy (NCT02927262) and after HSCT (NCT02997202).

Other Targeted Therapies

Venetoclax

Venetoclax (Vendexta™) is a “BH3-mimetic” antagonist of the antiapoptotic protein, BCL-2, with high efficacy in relapsed/refractory chronic lymphocytic leukemia, including in patients with a 17p deletion, for whom it is currently approved [100••]. Venetoclax efficiently caused on-target cell death through induction of the mitochondrial pathway of apoptosis in AML cell lines, primary patient samples, and murine primary xenografts [101••]. In a phase II study in patients with R/R AML or deemed unfit for IC, venetoclax produced an ORR of 19% and demonstrated antileukemic activity in an additional 19% of patients [102•]. Common AEs included nausea, diarrhea and vomiting (all grades), FN, and hypokalemia (grade 3/4) [102•]. The FDA has granted venetoclax

“breakthrough” designation in AML. In a phase IB study in treatment-naive older (≥ 65) patients with cytogenetically intermediate or poor risk AML ineligible for IC, the combination of venetoclax with an HMA (azacitidine or decitabine) yielded an ORR of 76% [103••]. It was higher (82%) in patients with *IDH1/2* mutations, an abnormality that had been shown in preclinical studies to confer enhanced susceptibility to BCL-2 inhibition [104••]. The most common treatment-emergent AEs (TEAEs) were nausea (54%), FN (41%), diarrhea (44%), decreased appetite (33%), and peripheral edema (31%). FN (41%) and neutropenia (33%) were the most common grade 3/4 TEAEs, but no DLT were reported [103••]. A similar trial has been conducted with venetoclax in combination with LDAC (NCT02287233). The ORR was 75% in this 20-patient study, with most responses (14 of 15) being CR/CRi [105]. The 12-month OS was 74.7% for all patients and 86.7% among the responders. Febrile neutropenia (35%), hypertension (20%), and hypophosphatemia (20%) constituted the most common nonhematologic grade 3/4 AEs [105].

MDM2 (HDM2) Inhibitors

Pharmacologic inhibition of murine double minute 2 (MDM2, or its human homolog, HDM2), a physiologic antagonist of p53, activates p53 and induces apoptosis or cell cycle arrest in wild-type p53-expressing AML; however, this strategy is not useful against mutant p53 [106]. The level of pretreatment MDM2 protein expression in leukemic blasts appears to be an important biomarker for prediction of response to the MDM2 inhibitor, idasanutlin (RG7388), among patients with R/R AML [107]. A phase IB trial in 75 patients with R/R AML [108] explored the combination of idasanutlin once or twice a day for 5 days and cytarabine 1 g/m²/day for 6 days. In the dose escalation phase ($n = 23$), patients could be R/R or untreated but not candidates for IC. The CR rate was 25%, the CRC rate was 29%, and when patients achieving MLFS were also considered, the rate was 33% [108]. The median DOR was 6.4 months [108]. A phase III placebo-controlled trial in patients with R/R AML in combination with cytarabine (NCT02545283) is currently underway. DS-3032b is another MDM2 inhibitor currently in a phase I trial (NCT02319369) in advanced hematological malignancies. The MTD was determined to be 160 mg daily on a 21/28 days schedule [109]. Reduction in bone marrow blasts by the end of cycle 1 (4 weeks) was seen in 15 of 38 patients with R/R AML or high-risk MDS. CR was achieved in two patients with AML, and one patient with MDS achieved a marrow CR with platelet improvement [109]. Ninety-three percent of patients experienced a grade ≥ 3 TEAE. The most common TEAEs (any grade) were nausea (73%), diarrhea (57%), vomiting (33%), fatigue (37%), anemia (33%), thrombocytopenia (33%), neutropenia (20%) hypotension (30%), hypokalemia (23%), and hypomagnesemia (20%) [109].

Immune Checkpoint Inhibitors

Immune checkpoints refer to a number of inhibitory pathways that are critical for maintaining self-tolerance and modulating physiological immune responses in order to minimize collateral tissue damage. These pathways are often co-opted by tumor cells to escape immune-mediated destruction [110]. The immune checkpoint pathways consisting of cytotoxic T-lymphocyte-associated antigen 4 (CTLA-4) and its ligands CD80 and CD86, as well as the

programmed cell death protein 1 (PD-1) and its ligands PD-L1 and PD-L2 have come to the forefront given that their blockade has been clinically efficacious in numerous different tumor types such as melanoma, lung cancer, head and neck cancer, Hodgkin's lymphoma, and urothelial and renal cell carcinoma (reviewed in [111]).

In a phase I/IB study, the anticytotoxic T-lymphocyte-associated protein 4 (CTLA-4) antibody, ipilimumab, was administered to 28 patients with hematologic malignancies that had relapsed post-allogeneic HSCT [112••]. CRs occurred in four patients with extramedullary AML and one patient with MDS evolving into AML, all at a dose of 10 mg/kg (every 3 weeks for 4 doses, then every 12 weeks for up to 60 weeks) [112••]. Immune-related AEs were reported in six patients (21%; one of them grade 5), and graft-versus-host disease (GVHD) precluding further administration of ipilimumab in four (14%) [112••]. There is preclinical evidence for upregulation of CTLA-4 and both PD-1 and its ligands, PD-L1 and PD-L2, on T-cells and neoplastic cells by azacitidine in AML and MDS [113, 114]. Based on this, the combination of azacitidine and the anti-PD-1 monoclonal antibody, nivolumab, is under investigation in patients with R/R AML and older patients (>65) with newly diagnosed AML (NCT02397720). Preliminary results on the first 51 patients, all with R/R AML, enrolled to this trial were recently presented [115•]. Of 35 response-evaluable patients, 6 (17%) obtained CR/CRI, 5 (14%) had hematologic improvement (HI), 9 (26%) had $\geq 50\%$ BM blast reduction, and 3 (9%) had SD for >6 months. The median OS of 9.3 months observed in this study compared favorably to that reported for historical patients treated with azacitidine-based salvage protocols at the MD Anderson Cancer Center (4.1 months). Immune-related AEs (grade 3/4 in 14% and grade 2 in 12%) responded rapidly to steroids in 12 of 13 episodes and these patients were successfully re-challenged with nivolumab, but one patient died from grade 4 pneumonitis/epiglottitis [115•]. There is also preclinical evidence that blockade of the inhibitory killer-cell immunoglobulin receptors (KIRs), which negatively regulate NK cell-mediated killing of human leukocyte antigen (HLA) class I-expressing tumors, may have antileukemic effects [116]. Twenty-one patients with relapsed AML have been treated with the combination of the anti-KIR monoclonal antibody lirilumab and azacitidine in an ongoing phase IB/II study [117]. No DLTs were observed and lirilumab 3 mg/kg was established as the RP2D in combination with standard dose azacitidine. Of 12 patients evaluable for response, 1 achieved CR and 1 CRI, 2 had $\geq 50\%$ BM blast reduction, and 2 HI lasting >6 months. Immune-related AEs occurred in three (14%) patients, all of whom responded rapidly to steroids and were re-challenged safely with lirilumab [117].

Rational Combinations

While targeted approaches to specific molecular abnormalities are attractive, AML is a molecularly complex disease with multiple aberrant pathways involved. It is therefore unlikely that blockade of any one specific pathway will lead to prolonged remissions and cures in more than a fraction of the patient population. Thus, combination strategies are likely to be the required to improve outcome. To date, most of the combinations have used a targeted agent

combined with standard agents such as anthracyclines, cytarabine, or hypomethylating agents. Rationally designed combinations that will more specifically and directly alter the biology of the disease are required. The number of such rational combinations possible is virtually limitless. The discussion that follows will focus on some promising approaches that have been shown to be synergistic in laboratory studies, some of which are being pursued in clinical trials (Table 2). This presentation should be considering only a sampling of what is to come rather than an exhaustive discussion of such approaches or a declaration of partiality for specific combinations.

As noted previously, *IDH1/2* mutations induce BCL-2 dependence in AML [104••]. The production by mutant IDH enzymes of 2-HG instead of the normal metabolite, α -KG, creates a dependence of the *IDH*-mutated cells on glutamine as their primary source of α -KG, which can be exploited therapeutically by pharmacologic inhibitors of glutaminase. Thus, *IDH*-mutated AML may be particularly sensitive to glutaminase inhibition [118•]. Glutamine levels also control mitochondrial oxidative phosphorylation (OXPHOS) in AML cells, and glutaminase inhibitors, e.g., CB-839, can reduce OXPHOS, leading to leukemic cell proliferation arrest and apoptosis [119•]. Inhibition of glutaminase by CB-839 activates mitochondrial apoptosis and sensitizes leukemic cells to venetoclax [119•]. These preclinical observations provide a compelling rationale for combining glutaminase inhibitors with BCL-2 antagonists in the clinic.

The major mediator of resistance to BCL-2/-xL inhibition in AML is the antiapoptotic protein, MCL-1 [120]. There is, thus, significant interest in combining venetoclax with drugs that downregulate MCL-1, such as mitogen-activated protein kinase (MEK) inhibitors [121] or sorafenib [122]. Synergism has also been demonstrated between venetoclax and idasanutlin, both in vitro and in vivo [123]. These findings form the basis of an ongoing phase I/II clinical trial (NCT02670044) investigating the combination of venetoclax with cobimetinib, a MEK inhibitor, and with idasanutlin in older patients (≥ 60) with R/R AML who are not eligible for IC. While MEK inhibitors have limited single-agent activity in R/R AML (with perhaps some preference for patients with *RAS* mutations) [124], synergistic induction of apoptosis is seen in AML cells with combined blockade of MEK and MDM2 [125]. *FLT3* signals downstream to members of the BCL-2 family, and synergistic induction of cell death in AML cell lines by the combination of sorafenib and the BCL-2/-xL inhibitor, ABT-737, has been shown [126]. Accordingly, combining sorafenib with venetoclax might improve the outcome observed with *FLT3* inhibitors alone in patients with *FLT3*-ITD, and may increase the observed responses to *FLT3* inhibitors such as quizartinib and gilteritinib seen among patients with wild-type *FLT3*. MDM2 inhibitors also potentiate apoptosis induction by *FLT3* inhibitors through multiple mechanisms [127, 128]. Similar to MDM2 inhibition, another therapeutic strategy to stabilize wild-type p53 is pharmacologic inhibition of chromosomal region maintenance 1 (CRM1), a nuclear export receptor, blockade of which results in nuclear retention of p53 [129]. Selective inhibitors of nuclear export (SINEs) synergize with MDM2 inhibitors in apoptosis induction of patient-derived AML cells [129]. NCT02530476 is an ongoing phase I/II study of sorafenib plus the SINE, selinexor (KPT-330), in patients with R/R *FLT3*-mutated AML. Studies of selinexor in combination with quizartinib and with MDM2 inhibitors in R/R AML are planned. Sequential treatment of AML

Table 2. Selected rational combinations in early phase trials for relapsed/refractory AML

Combination	Mechanism/rationale	Clinicaltrials.gov identifier
Venetoclax (BCL-2 inhibitor) + Idasanutlin (MDM2 antagonist)	Venetoclax inhibits anti-apoptotic BCL-2. Idasanutlin antagonizes MDM2 and activates p53, thereby enhancing the proapoptotic effects of venetoclax.	NCT02670044
Venetoclax (BCL-2 inhibitor) + Cobimetinib (MEK inhibitor)	Venetoclax inhibits antiapoptotic BCL-2, but not MCL-1. Cobimetinib, by blocking the MAPK pathway, downregulates MCL-1. MCL-1 is critical to the development and maintenance of AML.	NCT02670044
Sorafenib (FLT3 inhibitor) + Selinexor (SINE)	CRM1 (XPO1) mediates nuclear export of proteins, including p53. CRM1 expression is increased in FLT3-ITD ⁺ AML and correlates with worse outcomes. SINEs inhibit CRM1 and induce blast differentiation, besides down-regulating FLT3 and c-KIT. Sorafenib inhibits FLT3-ITD. Combination treatment has synergistic/additive pro-apoptotic effects in FLT3-mutant AML cells and suppresses the MAPK and Akt pathways.	NCT02530476
Fludarabine, Cytarabine (chemotherapy) + prexasertib (Chk1 inhibitor)	Nucleoside analogs cause DNA damage. This activates Chk1, which then causes cell cycle arrest, stabilizes stalled replication forks, activates DNA repair, and suppresses apoptosis. Chk1 inhibitors increase cytotoxicity of DNA-damaging chemotherapy by inducing cell death through "mitotic catastrophe."	NCT02649764
Belinostat (HDACI) + AZD1775 (Wee1 inhibitor)	HDACIs induce DNA damage and inhibit DNA repair. HDACIs also downregulate Chk1, thus abrogating its activation in response to Wee1 inhibition. Together, Wee1 inhibitors and HDACIs interact reciprocally to disrupt both the G1/S and G2/M cell cycle checkpoints.	NCT02381548
Idarubicin, Cytarabine + LY2510924 (CXCR4 antagonist)	Inhibition of the CXCR4-CXCL12 interaction blocks homing of AML blasts to protective bone marrow microenvironmental niches and makes them more susceptible to killing by chemotherapy.	NCT02652871
Azacitidine (HMA) + Pevonedistat (NAE Inhibitor)	Pevonedistat inhibits NAE, a critical step in the neddylation process, leading to impaired CRL function and causing accumulation of many CRL substrates including RRM2, a mediator of resistance to many cytotoxic anticancer agents. Azacitidine antagonizes RRM2.	NCT02610777
Azacitidine (HMA) + Nivolumab (anti-PD1 antibody)	Azacitidine upregulates CTLA-4, PD-1, PD-L1, and PD-L2. PD-1/PD-L1 interaction contributes to T-cell inactivation, decreased proliferation and effector function, and apoptosis. Nivolumab interferes with the PD-1/PD-L1 interaction.	NCT02397720

BCL-2 B-cell lymphoma 2, *MCL-1* myeloid cell leukemia 1, *MAPK* mitogen-activated protein kinase, *MEK* MAPK kinase, *FLT3* fms-like tyrosine kinase 3, *ITD* internal tandem duplication, *CRL* cullin-RING ligase, *HDACI* histone deacetylase inhibitor, *HMA* hypomethylating agent, *NAE* NEDD8-activating enzyme, *SINE* selective inhibitor of nuclear export, *PD-1* programmed death-1, *PD-L1* programmed death ligand-1, *PD-L2* programmed death ligand-2, *RRM2* ribonucleotide reductase regulatory subunit M2, *CTLA-4* cytotoxic T-lymphocyte antigen 4, *Chk1* checkpoint kinase 1, *CXCR4* C-X-C chemokine receptor type 4, *CXCL12* C-X-C motif chemokine 12, *AML* acute myeloid leukemia

blasts with decitabine followed by selinexor (decitabine “priming”) enhances the antileukemic effects of selinexor, possibly through re-expression of tumor suppressor proteins, e.g., CDKN1A and FOXO3A, that are epigenetically silenced by DNA methylation [130]. These preclinical findings were translated to a phase I clinical trial in 24 patients with R/R ($n = 19$) or previously untreated ($n = 5$, age ≥ 60 , unfit) AML (NCT02093403) [131]. The rate of CR/CRi/marrow CR was 80% in older, untreated and 21% in R/R AML patients. However, selinexor has considerable toxicities, which were somewhat ameliorated by twice weekly, flat (60 mg) dosing [131].

Pevonedistat (MLN4924) is a first-in-class inhibitor of protein neddylation that is active against AML cell lines, primary patient specimens, and xenograft models [132], and also exhibits modest single-agent efficacy in patients with R/R AML [133]. Pevonedistat synergizes with azacitidine in AML cells [134] and is currently being evaluated in combination with azacitidine (versus azacitidine alone) in a randomized, phase II clinical trial in patients with low blast count AML, MDS, or CMML (NCT02610777). In a separate, dose-escalation study in treatment-naive patients with AML ≥ 60 years of age, the MTD of the combination was found to be 20 mg/m²/day of pevonedistat, administered on days 1, 3, and 5, along with 75 mg/m²/day of azacitidine, administered on days 1–5, 8, and 9 in 4-week cycles [135]. The ORR was 60% among 52 evaluable patients (18 CR + 5 CRi + 8 PR) with a median DOR of 8.3 months; 6-month survival was 52% after a median follow-up of 16.4 months [135]. A clinical trial of pevonedistat in combination with the HDAC inhibitor, belinostat, in R/R AML is planned based on *in vitro* and *in vivo* evidence of synergism [136].

Cell cycle checkpoint dysfunction is universal in neoplastic cells, and a synthetic lethal therapeutic strategy that exploits this phenomenon is the combination of checkpoint kinase (e.g., Chk1, Wee1) inhibitors with DNA-damaging chemotherapy to trigger cell death by “mitotic catastrophe” [137]. The development of some Chk1 inhibitors, e.g., MK-8776, has been discontinued [138], but newer, potentially more potent ones, such as LY2606368, are in clinical trials in R/R AML in combination with IC (NCT02649764). There also exists strong preclinical rationale to combine inhibitors of Chk1 or Wee1 with HDAC inhibitors [139, 140], and a phase I clinical trial, primarily in R/R AML patients, of the combination of the Wee1 inhibitor, MK-1775, and belinostat is ongoing (NCT02381548).

Conclusion

After a long drought of new drug approvals in AML, regulatory approval of several new agents is expected in 2017. The OS benefit seen with some of these agents has brought renewed hope to investigators, physicians and patients. A comprehensive discussion of all the drugs in development for R/R AML is beyond the scope of this review. Table 3 lists some additional agents not covered in the text that are currently in clinical trials for patients with R/R AML. There are yet other agents, such as the histone deacetylase inhibitor pracinostat [141], and the smoothened (Hedgehog pathway) inhibitor glasdegib [142], which have shown great promise in previously untreated patients and are expected to be active in salvage settings as well. It is expected that with our improved understanding of the biology of AML and our ability to

Table 3. Miscellaneous agents currently in early phase clinical development for relapsed/refractory AML

Agent	Agent class	Phase of development	Clinicaltrials.gov identifier
Ibrutinib	BTK inhibitor	II	NCT02351037, NCT02635074
Ulixertinib (BVD-523, VRT752271)	ERK1/2 inhibitor	I/II	NCT02296242
BGB324	Axl inhibitor	I	NCT02488408
INCB053914	pan-PIM kinase inhibitor	I/II	NCT02587598
ONC201 (TIC10)	TRAIL pathway-inducer, triggers p53-independent apoptosis through ATF4 induction and atypical integrated stress response	I/II	NCT02392572
IACS-010759	Oxidative phosphorylation inhibitor (selectively inhibits complex I of the electron transport chain)	I	NCT02882321
BP-100-1.01	Liposomal Grb-2 (adaptor protein) antisense oligonucleotide	I	NCT01159028
FF-10501-01	Inosine 5'-monophosphate dehydrogenase (IMPDH) inhibitor	I/II	NCT02193958
APTO-253 (LOR-253, LT-253)	Induces expression of tumor suppressor transcription factor KLF4	I	NCT02267863
ADCT-301	CD25-directed antibody-drug conjugate (toxin: pyrrolbenzodiazepine)	I	NCT02588092
AGS67E	CD37-directed antibody-drug conjugate (toxin: monomethyl Auristatin E)	I	NCT02610062
<i>BTK</i> Bruton's tyrosine kinase, <i>ERK</i> extracellular signal-regulated kinase, <i>TRAIL</i> tumor necrosis factor alpha-related apoptosis inducing ligand, <i>ATF4</i> activating transcription factor 4, <i>KLF4</i> Kruppel-like factor 4			

translate this knowledge into effective therapies, the outcome of patients with AML will improve in a more definitive way in the coming years. Challenges in the definition of relevant endpoints, adequate clinical trial design, access to ever more costly drugs, and others still have to be addressed to make this reality more universal.

Acknowledgements

This work was supported in part by the MD Anderson Cancer Center Support Grant No. P30 CA016672 from the National Institutes of Health.

Compliance with Ethical Standards

Conflict of Interest

Prithviraj Bose has received research funding through a grant from Celgene Corporation.

Pankit Vachhani declares that he has no conflict of interest.

Jorge E. Cortes has received research funding through grants from Arog, Celator, Pfizer, Novartis, Tolero, Janssen, FORMA Therapeutics, Daiichi, Astellas, and Bristol-Myers Squibb; and has received compensation from Celator, Pfizer, Novartis, Janssen, Aegios, Astellas, and Bristol-Myers Squibb for service as a consultant.

Human and Animal Rights and Informed Consent

This article does not contain any studies with human or animal subjects performed by any of the authors.

References

Papers of particular interest, published recently, have been highlighted as:

- Of importance
 - Of major importance
1. Dohner H, Weisdorf DJ, Bloomfield CD. Acute myeloid leukemia. *N Engl J Med*. 2015;373:1136–52.
 2. Rowe JM, Tallman MS. How I treat acute myeloid leukemia. *Blood*. 2010;116:3147–56.
 3. Forman SJ, Rowe JM. The myth of the second remission of acute leukemia in the adult. *Blood*. 2013;121:1077–82.
 4. Ravandi F, Estey EH, Appelbaum FR, Lo-Coco F, Schiffer CA, Larson RA, Burnett AK, Kantarjian HM. Gemtuzumab ozogamicin: time to resurrect? *J Clin Oncol*. 2012;30:3921–3.
 5. •• Lancet JE, Uy GL, Cortes JE, Newell LF, Lin TL, Ritchie EK, Stuart RK, Strickland SA, Hogge D, Solomon SR, Stone RM, Bixby D, Koltz JE, Schiller GJ, Wieduwilt MJ, Ryan D, Hoering A, Chiarella M, Louie AC, Medeiros BC. Final results of a phase III randomized trial of CPX-351 versus 7 + 3 in older patients with newly diagnosed high risk (secondary) AML. *J Clin Oncol*. 2016;34:7000.
 6. •• Results of the pivotal trial of CPX-351 in older patients with newly diagnosed secondary AML showing a survival benefit for this agent.
 6. •• Stone RM, Mandrekar S, Sanford BL, Geyer S, Bloomfield CD, Dohner K, Thiede C, Marcucci G, Lo-Coco F, Klisovic RB, Wei A, Sierra J, Sanz MA, Brandwein JM, de Witte T, Niederwieser D, Appelbaum FR, Medeiros BC, Tallman MS, Krauter J, Schlenk RF, Ganser A, Serve H, Ehninger G, Amadori S, Larson RA, Dohner H. The multi-kinase inhibitor midostaurin (M) prolongs survival compared with placebo (P) in combination with daunorubicin (D)/cytarabine (C) induction (ind), high-dose C consolidation (consol), and as maintenance (maint) therapy in newly diagnosed acute myeloid leukemia (AML) patients (pts) age 18-60 with

- FLT3* mutations (mut): an international prospective randomized (rand) P-controlled double-blind trial (CALGB 10603/RATIFY [alliance]). *Blood*. 2015;126:6.
- Results of a cooperative group study that showed a survival benefit for the addition of midostaurin to standard chemotherapy and as maintenance in younger, newly diagnosed patients with *FLT3*-mutated AML.
7. Hawtin RE, Stockett DE, Byl JA, McDowell RS, Nguyen T, Arkin MR, Conroy A, Yang W, Osheroff N, Fox JA. Voreloxin is an anticancer quinolone derivative that intercalates DNA and poisons topoisomerase II. *PLoS One*. 2010;5:e10186.
 8. Walsby EJ, Coles SJ, Knapper S, Burnett AK. The topoisomerase II inhibitor voreloxin causes cell cycle arrest and apoptosis in myeloid leukemia cells and acts in synergy with cytarabine. *Haematologica*. 2011;96:393–9.
 9. Lancet JE, Roboz GJ, Cripe LD, Michelson GC, Fox JA, Leavitt RD, Chen T, Hawtin R, Craig AR, Ravandi F, Maris MB, Stuart RK, Karp JE. A phase 1b/2 study of vosaroxin in combination with cytarabine in patients with relapsed or refractory acute myeloid leukemia. *Haematologica*. 2015;100:231–7.
 - 10.●● Ravandi F, Ritchie EK, Sayar H, Lancet JE, Craig MD, Vey N, Strickland SA, Schiller GJ, Jabbour E, Erba HP, Pigneux A, Horst HA, Recher C, Klimek VM, Cortes J, Roboz GJ, Odenike O, Thomas X, Havelange V, Maertens J, Derigs HG, Heuser M, Damon L, Powell BL, Gaidano G, Carella AM, Wei A, Hogge D, Craig AR, Fox JA, Ward R, Smith JA, Acton G, Mehta C, Stuart RK, Kantarjian HM. Vosaroxin plus cytarabine versus placebo plus cytarabine in patients with first relapsed or refractory acute myeloid leukaemia (VALOR): a randomised, controlled, double-blind, multinational, phase 3 study. *Lancet Oncol*. 2015;16:1025–36.
- Results of the VALOR trial, one of the largest trials ever conducted in the salvage setting in AML, comparing vosaroxin plus cytarabine to placebo plus cytarabine.
11. Ravandi F, Ritchie EK, Sayar H, Lancet JE, Craig M, Vey N, Strickland SA, Schiller GJ, Jabbour EJ, Erba HP, Pigneux A, Horst HA, Recher C, Klimek VM, Cortes JE, Roboz GJ, Craig AR, Ward R, Smith J, Kantarjian HM, Stuart RK. Durable overall survival benefit in patients ≥ 60 Years with relapsed or refractory AML treated with vosaroxin/cytarabine vs placebo/cytarabine: updated results from the valor trial. *Blood*. 2016;128:903.
 - 12.● Daver NG, Kantarjian HM, Garcia-Manero G, Jabbour EJ, Borthakur G, Pierce SR, Vaughan K, Ning J, Gonzalez G, Pemmaraju N, Kadia TM, Konopleva MY, Andreeff M, Dinardo CD, Cortes JE, Ward R, Craig AR, Ravandi F. Phase I/II study of vosaroxin and decitabine in newly diagnosed older patients (pts) with acute myeloid leukemia (AML) and high-risk myelodysplastic syndrome (MDS). *Haematologica*. 2016;S505
- Promising results with the combination of decitabine and vosaroxin in older, newly diagnosed patients with AML.
13. Dennis M, Russell N, Hills RK, Hemmaway C, Panoskaltis N, McMullin MF, Kjeldsen L, Dignum H, Thomas IF, Clark RE, Milligan D, Burnett AK. Vosaroxin and vosaroxin plus low-dose Ara-C (LDAC) vs low-dose Ara-C alone in older patients with acute myeloid leukemia. *Blood*. 2015;125:2923–32.
 14. Lancet JE, Cortes JE, Hogge DE, Tallman MS, Kovacsovics TJ, Damon LE, Komrokji R, Solomon SR, Koltz JE, Cooper M, Yeager AM, Louie AC, Feldman EJ. Phase 2 trial of CPX-351, a fixed 5:1 molar ratio of cytarabine/daunorubicin, vs cytarabine/daunorubicin in older adults with untreated AML. *Blood*. 2014;123:3239–46.
 15. Cortes JE, Goldberg SL, Feldman EJ, Rizzeri DA, Hogge DE, Larson M, Pigneux A, Recher C, Schiller G, Warzocha K, Kantarjian H, Louie AC, Koltz JE. Phase II, multicenter, randomized trial of CPX-351 (cytarabine:daunorubicin) liposome injection versus intensive salvage therapy in adults with first relapse AML. *Cancer*. 2015;121(2):234–42.
 16. Bose P, Simmons GL, Grant S. Cyclin-dependent kinase inhibitor therapy for hematologic malignancies. *Expert Opin Investig Drugs*. 2013;22:723–38.
 17. Bible KC, Kaufmann SH. Cytotoxic synergy between flavopiridol (NSC 649890, L86-8275) and various antineoplastic agents: the importance of sequence of administration. *Cancer Res*. 1997;57:3375–80.
 18. Karp JE, Ross DD, Yang W, Tidwell ML, Wei Y, Greer J, Mann DL, Nakanishi T, Wright JJ, Colevas AD. Timed sequential therapy of acute leukemia with flavopiridol: in vitro model for a phase I clinical trial. *Clin Cancer Res*. 2003;9:307–15.
 19. Zeidner JF, Foster MC, Blackford AL, Litzow MR, Morris LE, Strickland SA, Lancet JE, Bose P, Levy MY, Tibes R, Gojo I, Gocke CD, Rosner GL, Little RF, Wright JJ, Doyle LA, Smith BD, Karp JE. Randomized multicenter phase II study of flavopiridol (alvocidib), cytarabine, and mitoxantrone (FLAM) versus cytarabine/daunorubicin (7 + 3) in newly diagnosed acute myeloid leukemia. *Haematologica*. 2015;100:1172–9.
 - 20.●● Glaser SP, Lee EF, Trounson E, Bouillet P, Wei A, Fairlie WD, Izon DJ, Zuber J, Rappaport AR, Herold MJ, Alexander WS, Lowe SW, Robb L, Strasser A. Anti-apoptotic Mcl-1 is essential for the development and sustained growth of acute myeloid leukemia. *Genes Dev*. 2012;26:120–5.
- Elegant preclinical work showing the critical role of MCL-1 in the development and maintenance of AML.
21. Del Gaizo Moore V, Letai A. BH3 profiling—measuring integrated function of the mitochondrial apoptotic pathway to predict cell fate decisions. *Cancer Lett*. 2013;332(2):202–5.
 22. Bose P, Grant S. Orphan drug designation for pracinostat, volasertib and alvocidib in AML. *Leuk Res*. 2014;38:862–5.
 - 23.● Issa JP, Roboz G, Rizzieri D, Jabbour E, Stock W, O'Connell C, Yee K, Tibes R, Griffiths EA, Walsh K, Daver N, Chung W, Naim S, Taverna P, Oganessian A, Hao Y, Lowder JN, Azab M, Kantarjian H. Safety and tolerability of guadecitabine (SGI-110) in patients with myelodysplastic syndrome and acute

- myeloid leukaemia: a multicentre, randomised, dose-escalation phase 1 study. *Lancet Oncol.* 2015;16:1099–110.
- Phase I results with a novel, second generation hypomethylating agent, guadecitabine, in MDS and AML.
24. Roboz GJ, Ravandi F, Kropf P, Yee K, O'Connell C, Griffiths EA, Stock W, Garcia-Manero G, Jabbour EJ, Daver N, Pemmaraju N, Issa JP, Walsh K, Rizzieri D, Lunin S, Naim S, Hao Y, Azab M, Kantarjian HM. Comparison of efficacy and safety of 5-day and 10-day schedules of SGI-110, a novel subcutaneous (SC) hypomethylating agent (HMA), in the treatment of relapsed/refractory acute myeloid leukemia (r/r AML). *Ann Oncol.* 2014;25(suppl_4):iv327–39.
 25. Daver N, Kantarjian HM, Roboz GJ, Kropf PL, Yee KWL, O'Connell C, Griffiths EA, Jabbour EJ, Stock W, Walsh K, Rizzieri DA, Berdeja JG, Su XY, Azab M, Issa JP. Long term survival and clinical complete responses of various prognostic subgroups in 103 relapsed/refractory acute myeloid leukemia (r/r AML) patients treated with guadecitabine (SGI-110) in phase 2 studies. *Blood.* 2016;128:904.
 26. Kantarjian HM, Roboz GJ, Kropf PL, KWL Y, O'Connell C, Tibes R, Walsh K, Podeltsev NA, Griffiths EA, Jabbour EJ, Garcia-Manero G, Rizzieri DA, Stock W, Savona MR, Rosenblat T, Berdeja JG, Wilson L, Lowder JN, Taverna P, Hao Y, Azab M, Issa JP. Comparison of efficacy and safety results in 103 treatment-naïve acute myeloid leukemia (TN-AML) patients not candidates for intensive chemotherapy using 5-day and 10-day regimens of guadecitabine (SGI-110), a novel hypomethylating agent (HMA). *Blood.* 2015;126:458.
 27. Yee K, Daver N, Kropf P, Tibes R, O'Connell C, Roboz G, Walsh K, Pemmaraju N, Rosenblat T, Berdeja J, Lunin S, Chung W, Issa JP, Naim S, Taverna P, Hao Y, Azab M, Kantarjian H. Results of a randomized multicenter phase 2 study of a 5-day regimen of SGI-110, a novel hypomethylating agent, in treatment naïve elderly acute myeloid leukemia not eligible for intensive therapy. 2014:S647-S647.
 28. Belkina AC, Denis GV. BET domain co-regulators in obesity, inflammation and cancer. *Nat Rev Cancer.* 2012;12:465–77.
 29. Dawson MA, Kouzarides T, Huntly BJ. Targeting epigenetic readers in cancer. *N Engl J Med.* 2012;367:647–57.
 30. Devaraj SG, Fiskus W, Shah B, Qi J, Sun B, Iyer SP, Sharma S, Bradner JE, Bhalla KN. HEXIM1 induction is mechanistically involved in mediating anti-AML activity of BET protein bromodomain antagonist. *Leukemia.* 2016;30:504–8.
 31. Berthon C, Raffoux E, Thomas X, Vey N, Gomez-Roca C, Yee K, Taussig DC, Rezai K, Roumier C, Herait P, Kahatt C, Quesnel B, Michallet M, Recher C, Lokiec F, Preudhomme C, Dombret H. Bromodomain inhibitor OTX015 in patients with acute leukaemia: a dose-escalation, phase 1 study. *Lancet Haematol.* 2016;3:e186–95.
 32. Fiskus W, Sharma S, Qi J, Shah B, Devaraj SG, Leveque C, Portier BP, Iyer SP, Bradner JE, Bhalla KN. BET protein antagonist JQ1 is synergistically lethal with FLT3 tyrosine kinase inhibitor (TKI) and overcomes resistance to FLT3-TKI in AML cells expressing FLT-ITD. *Mol Cancer Ther.* 2014.
 33. Fiskus W, Sharma S, Qi J, Valenta JA, Schaub LJ, Shah B, Peth K, Portier BP, Rodriguez M, Devaraj SG, Zhan M, Sheng J, Iyer SP, Bradner JE, Bhalla KN. Highly active combination of BRD4 antagonist and histone deacetylase inhibitor against human acute myelogenous leukemia cells. *Mol Cancer Ther.* 2014;13:1142–54.
 - 34.●● Bernt KM, Zhu N, Sinha AU, Vempati S, Faber J, Krivstov AV, Feng Z, Punt N, Daigle A, Bullinger L, Pollock RM, Richon VM, Kung AL, Armstrong SA. *MLL*-rearranged leukemia is dependent on aberrant H3K79 methylation by DOT1L. *Cancer Cell.* 2011;20:66–78.
- Preclinical work identifying DOT1L as a novel therapeutic target in *MLL*-rearranged leukemia.
- 35.●● Daigle SR, Olhava EJ, Therkelsen CA, Majer CR, Sneeringer CJ, Song J, Johnston LD, Scott MP, Smith JJ, Xiao Y, Jin L, Kuntz KW, Chesworth R, Moyer MP, Bernt KM, Tseng JC, Kung AL, Armstrong SA, Copeland RA, Richon VM, Pollock RM. Selective killing of mixed lineage leukemia cells by a potent small-molecule DOT1L inhibitor. *Cancer Cell.* 2011;20:53–65.
- Preclinical activity of a novel, small-molecule antagonist of DOT1L against *MLL*-rearranged leukemia.
36. Stein EM, Garcia-Manero G, Rizzieri DA, Tibes R, Berdeja JG, Jongen-Lavrencic M, Altman JK, Dohner H, Thomson B, Blakemore SJ, Daigle S, Fine G, Waters NJ, Krivstov AV, Koche R, Armstrong SA, Ho PT, Lowenberg B, Tallman MS. A phase 1 study of the DOT1L inhibitor, pinometostat (EPZ-5676), in adults with relapsed or refractory leukemia: safety, clinical activity. *Exposure Target Inhibition Blood.* 2015;126:2547.
 37. Rau RE, Rodriguez BA, Luo M, Jeong M, Rosen A, Rogers JH, Campbell CT, Daigle SR, Deng L, Song Y, Sweet S, Chevassut T, Andreeff M, Kornblau SM, Li W, Goodell MA. DOT1L as a therapeutic target for the treatment of DNMT3A-mutant acute myeloid leukemia. *Blood.* 2016;128:971–81.
 - 38.● Ley TJ, Ding L, Walter MJ, McLellan MD, Lamprecht T, Larson DE, Kandoth C, Payton JE, Baty J, Welch J, Harris CC, Lichti CF, Townsend RR, Fulton RS, Dooling DJ, Koboldt DC, Schmidt H, Zhang Q, Osborne JR, Lin L, O'Laughlin M, McMichael JF, Delehaunty KD, McGrath SD, Fulton LA, Magrini VJ, Vickery TL, Hundal J, Cook LL, Conyers JJ, Swift GW, Reed JP, Alldredge PA, Wylie T, Walker J, Kalicki J, Watson MA, Heath S, Shannon WD, Varghese N, Nagarajan R, Westervelt P, Tomasson MH, Link DC, Graubert TA, DiPersio JF, Mardis ER, Wilson RK. DNMT3A mutations in acute myeloid leukemia. *N Engl J Med.* 2010;363:2424–33.
- Important paper highlighting the prognostic importance of *DNMT3A* mutations in AML.
39. Schenk T, Chen WC, Gollner S, Howell L, Jin L, Hebestreit K, Klein HU, Popescu AC, Burnett A, Mills K, Casero Jr RA, Marton L, Woster P, Minden MD, Dugas M, Wang JC, Dick JE, Muller-Tidow C, Petrie K, Zelent

- A. Inhibition of the LSD1 (KDM1A) demethylase reactivates the all-trans-retinoic acid differentiation pathway in acute myeloid leukemia. *Nat Med.* 2012;18:605–11.
40. Fiskus W, Sharma S, Shah B, Portier BP, Devaraj SG, Liu K, Iyer SP, Bearss D, Bhalla KN. Highly effective combination of LSD1 (KDM1A) antagonist and pan-histone deacetylase inhibitor against human AML cells. *Leukemia.* 2014;28:2155–64.
41. Somerville T, Salamero O, Montesinos P, Willekens C, Simon JAP, Pigneux A, Recher C, Popat R, Molinero C, Mascaró C, Maes T, Bosch F. Safety, pharmacokinetics (PK), pharmacodynamics (PD) and preliminary activity in acute leukemia of Ory-1001, a first-in-class inhibitor of lysine-specific histone demethylase 1A (LSD1/KDM1A): initial results from a first-in-human phase 1 study. *Blood.* 2016;128:4060.
- 42.● Sievers EL, Larson RA, Stadtmayer EA, Estey E, Lowenberg B, Dombret H, Karanes C, Theobald M, Bennett JM, Sherman ML, Berger MS, Eten CB, Loken MR, van Dongen JJ, Bernstein ID, Appelbaum FR, Mylotarg Study Group. Efficacy and safety of gemtuzumab ozogamicin in patients with CD33-positive acute myeloid leukemia in first relapse. *J Clin Oncol.* 2001;19:3244–54.
- The study that led to the original FDA approval of gemtuzumab ozogamicin for AML.
- 43.● Petersdorf SH, Kopecky KJ, Slovak M, Willman C, Nevill T, Brandwein J, Larson RA, Erba HP, Stiff PJ, Stuart RK, Walter RB, Tallman MS, Stenke L, Appelbaum FR. A phase 3 study of gemtuzumab ozogamicin during induction and postconsolidation therapy in younger patients with acute myeloid leukemia. *Blood.* 2013;121:4854–60.
- The study that led to the withdrawal of gemtuzumab ozogamicin.
- 44.●● Burnett AK, Hills RK, Milligan D, Kjeldsen L, Kell J, Russell NH, Yin JA, Hunter A, Goldstone AH, Wheatley K. Identification of patients with acute myeloblastic leukemia who benefit from the addition of gemtuzumab ozogamicin: results of the MRC AML15 trial. *J Clin Oncol.* 2011;29:369–77.
- Demonstration that the survival advantage with gemtuzumab ozogamicin is restricted to patients with favorable and non-adverse cytogenetics.
- 45.●● Burnett AK, Russell NH, Hills RK, Kell J, Freeman S, Kjeldsen L, Hunter AE, Yin J, Craddock CF, Dufva IH, Wheatley K, Milligan D. Addition of gemtuzumab ozogamicin to induction chemotherapy improves survival in older patients with acute myeloid leukemia. *J Clin Oncol.* 2012;30:3924–31.
- Phase III trial from the UK demonstrating improved survival with the addition of gemtuzumab ozogamicin to chemotherapy in older patients with newly diagnosed AML.
- 46.●● Castaigne S, Pautas C, Terre C, Raffoux E, Bordessoule D, Bastie JN, Legrand O, Thomas X, Turlure P, Reman O, de Revel T, Gastaud L, de Gunzburg N, Contentin N, Henry E, Marolleau JP, Aljijakli A, Rouselot P, Fenaux P, Preudhomme C, Chevret S, Dombret H. Acute Leukemia French Association. Effect of gemtuzumab ozogamicin on survival of adult patients with de-novo acute myeloid leukaemia (ALFA-0701): a randomised, open-label, phase 3 study. *Lancet.* 2012;379:1508–16.
- Randomized phase III trial from France demonstrating improved survival with the addition of gemtuzumab ozogamicin to chemotherapy in patients with newly diagnosed AML.
- 47.● Hills RK, Castaigne S, Appelbaum FR, Delaunay J, Petersdorf S, Othus M, Estey EH, Dombret H, Chevret S, Ifrah N, Cahn JY, Recher C, Chilton L, Moorman AV, Burnett AK. Addition of gemtuzumab ozogamicin to induction chemotherapy in adult patients with acute myeloid leukaemia: a meta-analysis of individual patient data from randomised controlled trials. *Lancet Oncol.* 2014;15:986–96.
- Meta-analysis (including the negative SWOG trial) documenting the survival advantage conferred by the addition of gemtuzumab ozogamicin to induction chemotherapy in patients with AML without adverse cytogenetics.
- 48.● Stein AM, Walter RB, Erba HP, Fathi AT, Advani AS, Lancet JE, Ravandi F, Kovacovics TJ, DeAngelo DJ, Bixby D, Faderl S, Jillela AP, O'Meara MM, Zhao B, Stein EM. A phase 1 trial of SGN-CD33A as monotherapy in patients with CD33-positive acute myeloid leukemia (AML). *Blood.* 2015;126:324.
- Single-agent data with a novel, CD33 antibody-drug conjugate in relapsed/refractory AML.
- 49.● Fathi AT, Erba HP, Lancet JE, Stein EM, Ravandi F, Faderl S, Walter RB, Advani AS, DeAngelo DJ, Kovacovics TJ, Jillela AP, Bixby D, Levy M, O'Meara MM, Ho P, Stein AS. Vadastuximab talirine plus hypomethylating agents: a well-tolerated regimen with high remission rate in frontline older patients with acute myeloid leukemia (AML). *Blood.* 2016;128:591.
- 50.● Erba HP, Levy M, Vasu S, Stein AS, Fathi AT, Maris MB, Advani AS, Faderl S, Smith SE, Wood B, Walter RB, Yang J, Donnellan WB, Feldman EJ, Voellinger JL, Ravandi F. A phase 1b study of vadastuximab talirine in combination with 7 + 3 induction therapy for patients with newly diagnosed acute myeloid leukemia (AML). *Blood.* 2016;128:211.
- 51.● Yang J, Ravandi F, Advani AS, Vasu S, Walter RB, Faderl S, Stein AS, Erba HP, Fathi AT, Donnellan WB, Levy MY, Smith SE, Wood B, Feldman EJ, Voellinger JL, Maris MB. A phase 1b study of vadastuximab talirine as maintenance and in combination with standard consolidation for patients with acute myeloid leukemia (AML). *Blood.* 2016;128:340.
- 52.● Krupka C, Kufer P, Kischel R, Zugmaier G, Bogeholz J, Kohnke T, Lichtenegger FS, Schneider S, Metzeler KH, Fiegl M, Spiekermann K, Baeuerle PA, Hiddemann W, Riethmuller G, Subklewe M. CD33 target validation and sustained depletion of AML blasts in long-term cultures by the bispecific T-cell-engaging antibody AMG 330. *Blood.* 2014;123:356–65.
- Preclinical activity of a novel, CD33-targeted BiTE® in AML.

53. • Laszlo GS, Gudgeon CJ, Harrington KH, Dell'Aringa J, Newhall KJ, Means GD, Sinclair AM, Kischel R, Frankel SR, Walter RB. Cellular determinants for preclinical activity of a novel CD33/CD3 bispecific T-cell engager (BiTE) antibody, AMG 330, against human AML. *Blood*. 2014;123:554–61.
- Preclinical activity of a novel, CD33-targeted BiTE® in AML.
54. • Friedrich M, Henn A, Raum T, Bajtus M, Matthes K, Hendrich L, Wahl J, Hoffmann P, Kischel R, Kvesic M, Slootstra JW, Baeuerle PA, Kufer P, Rattel B. Preclinical characterization of AMG 330, a CD3/CD33-bispecific T-cell-engaging antibody with potential for treatment of acute myelogenous leukemia. *Mol Cancer Ther*. 2014;13:1549–57.
- Preclinical activity of a novel, CD33-targeted BiTE® in AML.
55. Harrington KH, Gudgeon CJ, Laszlo GS, Newhall KJ, Sinclair AM, Frankel SR, Kischel R, Chen G, Walter RB. The broad anti-AML activity of the CD33/CD3 BiTE antibody construct, AMG 330, is impacted by disease stage and risk. *PLoS One*. 2015;10:e0135945.
56. Krupka C, Kufer P, Kischel R, Zugmaier G, Lichtenegger FS, Kohnke T, Vick B, Jeremias I, Metzeler KH, Altmann T, Schneider S, Fiegl M, Spiekermann K, Bauerle PA, Hiddemann W, Riethmuller G, Subklewe M. Blockade of the PD-1/PD-L1 axis augments lysis of AML cells by the CD33/CD3 BiTE antibody construct AMG 330: reversing a T-cell-induced immune escape mechanism. *Leukemia*. 2016;30:484–91.
57. • Al-Hussaini M, Rettig MP, Ritchey JK, Karpova D, Uy GL, Eissenberg LG, Gao F, Eades WC, Bonvini E, Chichili GR, Moore PA, Johnson S, Collins L, DiPersio JF. Targeting CD123 in acute myeloid leukemia using a T-cell-directed dual-affinity retargeting platform. *Blood*. 2016;127:122–31.
- Preclinical paper introducing the DART technology for therapy of AML.
58. Sweet K, Pemmaraju N, Lane AA, Stein AM, Vasu S, Blum W, Rizzieri D, Wang ES, Rowinsky EK, Szarek M, Brooks CL, Disalvatore S, Liu D, Duvic M, Schwartz J, Konopleva M. Lead-in stage results of a pivotal trial of SL-401, an interleukin-3 receptor (IL-3R) targeting biologic, in patients with blastic plasmacytoid dendritic cell neoplasm (BPDCN) or acute myeloid leukemia (AML). *Blood*. 2015;126:3795.
59. DiNardo CD, Ravandi F, Agresta S, Konopleva M, Takahashi K, Kadia T, Routbort M, Patel KP, Mark B, Pierce S, Garcia-Manero G, Cortes J, Kantarjian H. Characteristics, clinical outcome, and prognostic significance of IDH mutations in AML. *Am J Hematol*. 2015;90:732–6.
60. •• Figueroa ME, Abdel-Wahab O, Lu C, Ward PS, Patel J, Shih A, Li Y, Bhagwat N, Vasanthakumar A, Fernandez HF, Tallman MS, Sun Z, Wolniak K, Peeters JK, Liu W, Choe SE, Fantin VR, Paietta E, Lowenberg B, Licht JD, Godley LA, Delwel R, Valk PJ, Thompson CB, Levine RL, Melnick A. Leukemic IDH1 and IDH2 mutations result in a hypermethylation phenotype, disrupt TET2 function, and impair hematopoietic differentiation. *Cancer Cell*. 2010;18:553–67.
- Seminal paper describing the role of mutant IDH1/2 enzymes in AML pathogenesis.
61. • Stein EM, Dinardo CD, Altman JK, Collins R, De Angelo DJ, Kantarjian HM, Sekeres MA, Fathi AT, Flinn IW, Frankel A, Levine RL, Medeiros BC, Patel MR, Pollyea DA, Roboz GJ, Stone RM, Swords RT, Tallman MS, Yen K, Attar EC, Xu Q, Tosolini A, Mei JM, Thakurta A, Knight RD, de Botton S. Safety and efficacy of AG-221, a potent inhibitor of mutant IDH2 that promotes differentiation of myeloid cells in patients with advanced hematologic malignancies: results of a phase 1/2 trial. *Blood*. 2015;126:323.
- Promising single-agent data with the IDH2 inhibitor AG221 in patients with R/R AML.
62. • Dinardo CD, de Botton S, Pollyea DA, Stein EM, Fathi AT, Roboz GJ, Collins R, Swords RT, Flinn IW, Altman JK, Tallman MS, Kantarjian HM, Derti A, Goldwasser M, Prahm M, Wu B, Yen K, Agresta S, Stone RM. Molecular profiling and relationship with clinical response in patients with IDH1 mutation-positive hematologic malignancies receiving AG-120, a first-in-class potent inhibitor of mutant IDH1, in addition to data from the completed dose escalation portion of the phase 1 study. *Blood*. 2015;126:1306.
- Promising single-agent data with the IDH1 inhibitor AG120 in patients with R/R AML.
63. Dinardo CD, Schimmer AD, Yee KWL, Hochhaus A, Carvajal RD, Janku F, Bedard P, Carpio C, Wick A, Schwartz GK, Schoffski P, Wen P, van den Bent MJ, Rosenthal M, O'Keefe J, Chen X, Pagliarini R, Schuck V, Myers A, Wei A. A phase I study of IDH305 in patients with advanced malignancies including relapsed/refractory AML and MDS that harbor *IDH1*^{R132} mutations. *Blood*. 2016;128:1073.
64. Kindler T, Lipka DB, Fischer T. FLT3 as a therapeutic target in AML: still challenging after all these years. *Blood*. 2010;116:5089–102.
65. Smith CC, Lin K, Stecula A, Sali A, Shah NP. FLT3 D835 mutations confer differential resistance to type II FLT3 inhibitors. *Leukemia*. 2015;29:2390–2.
66. Zhang W, Konopleva M, Shi YX, McQueen T, Harris D, Ling X, Estrov Z, Quintas-Cardama A, Small D, Cortes J, Andreeff M. Mutant FLT3: a direct target of sorafenib in acute myelogenous leukemia. *J Natl Cancer Inst*. 2008;100:184–98.
67. Borthakur G, Kantarjian H, Ravandi F, Zhang W, Konopleva M, Wright JJ, Faderl S, Verstovsek S, Mathews S, Andreeff M, Cortes JE. Phase I study of sorafenib in patients with refractory or relapsed acute leukemias. *Haematologica*. 2011;96:62–8.
68. Crump M, Hedley D, Kamel-Reid S, Leber B, Wells R, Brandwein J, Buckstein R, Kassiss J, Minden M, Matthews J, Robinson S, Turner R, McIntosh L, Eisenhauer E, Seymour L. A randomized phase I clinical and biologic study of two schedules of sorafenib in patients with myelodysplastic syndrome or acute myeloid leukemia: a NCIC (National Cancer Institute of Canada) Clinical Trials Group Study. *Leuk Lymphoma*. 2010;51:252–60.

69. Macdonald DA, Assouline SE, Brandwein J, Kamel-Reid S, Eisenhauer EA, Couban S, Caplan S, Foo A, Walsh W, Leber B. A phase I/II study of sorafenib in combination with low dose cytarabine in elderly patients with acute myeloid leukemia or high-risk myelodysplastic syndrome from the National Cancer Institute of Canada Clinical Trials Group: trial IND.186. *Leuk Lymphoma*. 2013;54:760–6.
- 70.● Ravandi F, Alattar ML, Grunwald MR, Rudek MA, Rajkhowa T, Richie MA, Pierce S, Daver N, Garcia-Manero G, Faderl S, Nazha A, Konopleva M, Borthakur G, Burger J, Kadia T, Dellasala S, Andreeff M, Cortes J, Kantarjian H, Levis M. Phase 2 study of azacytidine plus sorafenib in patients with acute myeloid leukemia and FLT-3 internal tandem duplication mutation. *Blood*. 2013;121:4655–62.
- A useful combination of commercially available drugs for patients with *FLT3*-ITD AML.
71. Ravandi F, Arana Yi C, Cortes JE, Levis M, Faderl S, Garcia-Manero G, Jabbour E, Konopleva M, O'Brien S, Estrov Z, Borthakur G, Thomas D, Pierce S, Brandt M, Pratz K, Luthra R, Andreeff M, Kantarjian H. Final report of phase II study of sorafenib, cytarabine and idarubicin for initial therapy in younger patients with acute myeloid leukemia. *Leukemia*. 2014;28:1543–5.
72. Serve H, Krug U, Wagner R, Sauerland MC, Heinecke A, Brunnberg U, Schaich M, Ottmann O, Duyster J, Wandt H, Fischer T, Giagounidis A, Neubauer A, Reichle A, Aulitzky W, Noppeney R, Blau I, Kunzmann V, Stuhlmann R, Kramer A, Kreuzer KA, Brandts C, Steffen B, Thiede C, Muller-Tidow C, Ehninger G, Berdel WE. Sorafenib in combination with intensive chemotherapy in elderly patients with acute myeloid leukemia: results from a randomized, placebo-controlled trial. *J Clin Oncol*. 2013;31:3110–8.
- 73.●● Rollig C, Serve H, Huttmann A, Noppeney R, Muller-Tidow C, Krug U, Baldus CD, Brandts CH, Kunzmann V, Einsele H, Kramer A, Schafer-Eckart K, Neubauer A, Burchert A, Giagounidis A, Krause SW, Mackensen A, Aulitzky W, Herbst R, Hanel M, Kiani A, Frickhofen N, Kullmer J, Kaiser U, Link H, Geer T, Reichle A, Jungthans C, Repp R, Heits F, Durk H, Hase J, Klut IM, Illmer T, Bornhauser M, Schaich M, Parmentier S, Gorner M, Thiede C, von Bonin M, Schetelig J, Kramer M, Berdel WE, Ehninger G. Study Alliance Leukaemia. Addition of sorafenib versus placebo to standard therapy in patients aged 60 years or younger with newly diagnosed acute myeloid leukaemia (SORAML): a multicentre, phase 2, randomised controlled trial. *Lancet Oncol*. 2015;16:1691–9.
- A placebo-controlled, phase II, randomized trial showing a survival benefit for the addition of sorafenib to chemotherapy in younger, unselected patients with newly diagnosed AML.
74. Chen YB, Li S, Lane AA, Connolly C, Del Rio C, Valles B, Curtis M, Ballen K, Cutler C, Dey BR, El-Jawahri A, Fathi AT, Ho VT, Joyce A, McAfee S, Rudek M, Rajkhowa T, Verselis S, Antin JH, Spitzer TR, Levis M, Soiffer R. Phase I trial of maintenance sorafenib after allogeneic hematopoietic stem cell transplantation for *fms*-like tyrosine kinase 3 internal tandem duplication acute myeloid leukemia. *Biol Blood Marrow Transplant*. 2014;20:2042–8.
75. Stone RM, DeAngelo DJ, Klimek V, Galinsky I, Estey E, Nimer SD, Grandin W, Lebwohl D, Wang Y, Cohen P, Fox EA, Neuberg D, Clark J, Gilliland DG, Griffin JD. Patients with acute myeloid leukemia and an activating mutation in *FLT3* respond to a small-molecule *FLT3* tyrosine kinase inhibitor, PKC412. *Blood*. 2005;105:54–60.
76. Fischer T, Stone RM, Deangelo DJ, Galinsky I, Estey E, Lanza C, Fox E, Ehninger G, Feldman EJ, Schiller GJ, Klimek VM, Nimer SD, Gilliland DG, Dutreix C, Huntsman-Labed A, Virkus J, Giles FJ. Phase IIB trial of oral midostaurin (PKC412), the *FMS*-like tyrosine kinase 3 receptor (*FLT3*) and multi-targeted kinase inhibitor, in patients with acute myeloid leukemia and high-risk myelodysplastic syndrome with either wild-type or mutated *FLT3*. *J Clin Oncol*. 2010;28:4339–45.
77. Williams CB, Kambhampati S, Fiskus W, Wick J, Dutreix C, Ganguly S, Aljitawi O, Reyes R, Fleming A, Abhyankar S, Bhalla KN, McGuirk JP. Preclinical and phase I results of decitabine in combination with midostaurin (PKC412) for newly diagnosed elderly or relapsed/refractory adult patients with acute myeloid leukemia. *Pharmacotherapy*. 2013;33:1341–52.
78. Strati P, Kantarjian H, Ravandi F, Nazha A, Borthakur G, Daver N, Kadia T, Estrov Z, Garcia-Manero G, Konopleva M, Rajkhowa T, Durand M, Andreeff M, Levis M, Cortes J. Phase I/II trial of the combination of midostaurin (PKC412) and 5-azacytidine for patients with acute myeloid leukemia and myelodysplastic syndrome. *Am J Hematol*. 2015;90:276–81.
79. Stone RM, Fischer T, Paquette R, Schiller G, Schiffer CA, Ehninger G, Cortes J, Kantarjian HM, DeAngelo DJ, Huntsman-Labed A, Dutreix C, del Corral A, Giles F. Phase IB study of the *FLT3* kinase inhibitor midostaurin with chemotherapy in younger newly diagnosed adult patients with acute myeloid leukemia. *Leukemia*. 2012;26:2061–8.
80. Zarrinkar PP, Gunawardane RN, Cramer MD, Gardner MF, Brigham D, Belli B, Karaman MW, Pratz KW, Palares G, Chao Q, Sprankle KG, Patel HK, Levis M, Armstrong RC, James J, Bhagwat SS. AC220 is a uniquely potent and selective inhibitor of *FLT3* for the treatment of acute myeloid leukemia (AML). *Blood*. 2009;114:2984–92.
81. Cortes JE, Kantarjian H, Foran JM, Ghirdaladze D, Zodelava M, Borthakur G, Gammon G, Trone D, Armstrong RC, James J, Levis M. Phase I study of quizartinib administered daily to patients with relapsed or refractory acute myeloid leukemia irrespective of *FMS*-like tyrosine kinase 3-internal tandem duplication status. *J Clin Oncol*. 2013;31:3681–7.
82. Cortes JE, Perl AE, Dombret H, Kayser S, Steffen B, Rousselot P, Martinelli G, Estey EH, Burnett AK, Gammon G, Trone D, Leo E, Levis MJ. Final results of a phase 2 open-label, monotherapy efficacy and safety

- study of quizartinib (AC220) in patients \geq 60 years of age with FLT3 ITD positive or negative relapsed/refractory acute myeloid leukemia. ASH Annual Meeting Abstracts. 2012;120:48.
83. Levis MJ, Perl AE, Dombret H, Dohner H, Steffen B, Rousselot P, Martinelli G, Estey EH, Burnett AK, Gammon G, Trone D, Leo E, Cortes JE. Final results of a phase 2 open-label, monotherapy efficacy and safety study of quizartinib (AC220) in patients with FLT3-ITD positive or negative relapsed/refractory acute myeloid leukemia after second-line chemotherapy or hematopoietic stem cell transplantation. ASH Annual Meeting Abstracts. 2012;120:673.
84. Hills RK, Gammon G, Trone D, Burnett AK. Quizartinib significantly improves overall survival in FLT3-ITD positive AML patients relapsed after stem cell transplantation or after failure of salvage chemotherapy: a comparison with historical AML database (UK NCRI data). *Blood*. 2015;126:2557.
85. Cortes JE, Tallman MS, Schiller G, Trone D, Gammon G, Goldberg S, Perl AE, Marie JP, Martinelli G, Levis M. Results of a phase 2 randomized, open-label, study of lower doses of quizartinib (AC220; ASP2689) in subjects with FLT3-ITD positive relapsed or refractory acute myeloid leukemia (AML). *Blood*. 2013;122:494.
86. Abdelall W, Kantarjian HM, Borthakur G, Garcia-Manero G, Patel KP, Jabbour EJ, Daver NG, Kadia TM, Gborogen R, Konopleva M, Ravandi F, Andreeff M, Cortes J. The combination of quizartinib with azacitidine or low dose cytarabine is highly active in patients (pts) with FLT3-ITD mutated myeloid leukemias: interim report of a phase I/II trial. *Blood*. 2016;128:1642.
87. Sandmaier BM, Khaled SK, Oran B, Gammon G, Trone D, Frankfurt O. Results of a phase 1 study of quizartinib (AC220) as maintenance therapy in subjects with acute myeloid leukemia in remission following allogeneic hematopoietic cell transplantation. *Blood*. 2014;124:428.
88. Bagrintseva K, Schwab R, Kohl TM, Schnittger S, Eichenlaub S, Ellwart JW, Hiddemann W, Spiekermann K. Mutations in the tyrosine kinase domain of FLT3 define a new molecular mechanism of acquired drug resistance to PTK inhibitors in FLT3-ITD-transformed hematopoietic cells. *Blood*. 2004;103:2266–75.
- 89.●● Smith CC, Wang Q, Chin CS, Salerno S, Damon LE, Levis MJ, Perl AE, Travers KJ, Wang S, Hunt JP, Zarrinkar PP, Schadt EE, Kasarskis A, Kuriyan J, Shah NP. Validation of ITD mutations in FLT3 as a therapeutic target in human acute myeloid leukaemia. *Nature*. 2012;485:260–3.
- Preclinical work demonstrating the emergence of resistance-conferring mutations in the FLT3 tyrosine kinase domain under the selective pressure of FLT3-ITD-specific inhibitors.
90. Alvarado Y, Kantarjian HM, Luthra R, Ravandi F, Borthakur G, Garcia-Manero G, Konopleva M, Estrov Z, Andreeff M, Cortes JE. Treatment with FLT3 inhibitor in patients with FLT3-mutated acute myeloid leukemia is associated with development of secondary FLT3-tyrosine kinase domain mutations. *Cancer*. 2014;120:2142–9.
- 91.● Zimmerman EI, Turner DC, Buaboonnam J, Hu S, Orwick S, Roberts MS, Janke LJ, Ramachandran A, Stewart CF, Inaba H, Baker SD. Crenolanib is active against models of drug-resistant FLT3-ITD-positive acute myeloid leukemia. *Blood*. 2013;122:3607–15.
- Preclinical study showing activity of crenolanib against resistance-conferring point mutations in FLT3.
- 92.● Galanis A, Ma H, Rajkhowa T, Ramachandran A, Small D, Cortes J, Levis M. Crenolanib is a potent inhibitor of FLT3 with activity against resistance-conferring point mutants. *Blood*. 2014;123:94–100.
- Preclinical study showing activity of crenolanib against resistance-conferring point mutations in FLT3.
- 93.● Smith CC, Lasater EA, Lin KC, Wang Q, McCreery MQ, Stewart WK, Damon LE, Perl AE, Jeschke GR, Sugita M, Carroll M, Kogan SC, Kuriyan J, Shah NP. Crenolanib is a selective type I pan-FLT3 inhibitor. *Proc Natl Acad Sci U S A*. 2014;111:5319–24.
- Preclinical study showing activity of crenolanib against resistance-conferring point mutations in FLT3.
- 94.● Cortes JE, Kantarjian HM, Kadia TM, Borthakur G, Konopleva M, Garcia-Manero G, Daver NG, Pemmaraju N, Jabbour E, Estrov Z, Ramachandran A, Paradela J, Pond B, Ravandi F, Vusirkala M, Patel PA, Levis MJ, Perl AE, Andreeff M, Collins R. Crenolanib besylate, a type I pan-FLT3 inhibitor, to demonstrate clinical activity in multiply relapsed FLT3-ITD and D835 AML. *J Clin Oncol*. 2016;34:7008.
- Single-agent data with crenolanib in relapsed/refractory FLT3-mutated AML.
95. Ohanian M, Kantarjian HM, Borthakur G, Kadia TM, Konopleva MY, Garcia-Manero G, Estrov Z, Ferrajoli A, Takahashi K, Jabbour EJ, Daver NG, Kornblau SM, Wierda WG, Burger JA, Naqvi K, Benton CB, Bose P, Eckardt J, Ravandi F, Cortes JE. Efficacy of a type I FLT3 inhibitor, crenolanib, with idarubicin and high-dose Ara-C in multiply relapsed/refractory FLT3+ AML. *Blood*. 2016;128:2744.
96. Wang ES, Stone RM, Tallman MS, Walter RB, Eckardt JR, Collins R. Crenolanib, a type I FLT3 TKI, can be safely combined with cytarabine and anthracycline induction chemotherapy and results in high response rates in patients with newly diagnosed FLT3 mutant acute myeloid leukemia (AML). *Blood*. 2016;128:1071.
97. Stein EM, Tallman MS. Emerging therapeutic drugs for AML. *Blood*. 2016;127:71–8.
98. Levis MJ, Perl AE, Altman JK, Cortes JE, Ritchie EK, Larson RA, Smith CC, Wang ES, Strickland SA, Baer MR, Litzow MR, Claxton D, Schiller GJ, Ustun C, Liu C, Gill S, Sargent B, Bahceci E. Results of a first-in-human, phase I/II trial of ASP2215, a selective, potent inhibitor of FLT3/Axl in patients with relapsed or refractory (R/R) acute myeloid leukemia (AML). *J Clin Oncol*. 2015;33:7003.
- 99.● Perl AE, Altman JK, Cortes JE, Smith CC, Litzow MR, Baer MR, Claxton DF, Erba HP, Gill S, Goldberg S, Jurcic JG, Larson RA, Liu C, Ritchie EK, Schiller GJ, Spira

- AI, Strickland SA, Tibes R, Ustun C, Wang ES, Stuart RK, Rollig C, Neubauer A, Martinelli G, Bahceci E, Levis MJ. Antileukemic activity and tolerability of ASP2215 80 mg and greater in FLT3 mutation-positive subjects with relapsed or refractory acute myeloid leukemia: results from a phase 1/2, open-label, dose-escalation/dose-response study. *Blood*. 2016;128:1069.
- Results from the CHRYSALIS trial of gilteretinib in relapsed/refractory FLT3-mutated AML.
- 100.●● Roberts AW, Davids MS, Pagel JM, Kahl BS, Puvvada SD, Gerecitano JF, Kipps TJ, Anderson MA, Brown JR, Gressick L, Wong S, Dunbar M, Zhu M, Desai MB, Cerri E, Heitner Enschede S, Humerickhouse RA, Wierda WG, Seymour JF. Targeting BCL2 with venetoclax in relapsed chronic lymphocytic leukemia. *N Engl J Med*. 2016;374:311–22.
- Phase I trial of venetoclax in patients with relapsed or refractory CLL showing high efficacy.
- 101.●● Pan R, Hogdal LJ, Benito JM, Bucci D, Han L, Borthakur G, Cortes J, DeAngelo DJ, Debose L, Mu H, Dohner H, Gaidzik VI, Galinsky I, Golfman LS, Haferlach T, Harutyunyan KG, Hu J, Levenson JD, Marcucci G, Muschen M, Newman R, Park E, Ruvolo PP, Ruvolo V, Ryan J, Schindela S, Zweidler-McKay P, Stone RM, Kantarjian H, Andreeff M, Konopleva M, Letai AG. Selective BCL-2 inhibition by ABT-199 causes on-target cell death in acute myeloid leukemia. *Cancer Discov*. 2014;4:362–75.
- Preclinical studies validating BCL-2 as a therapeutic target in AML.
- 102.●● Konopleva M, Pollyea DA, Potluri J, Chyla B, Hogdal L, Busman T, McKeegan E, Salem AH, Zhu M, Ricker JL, Blum W, DiNardo CD, Kadia T, Dunbar M, Kirby R, Falotico N, Levenson J, Humerickhouse R, Mabry M, Stone R, Kantarjian H, Letai A. Efficacy and biological correlates of response in a phase II study of venetoclax monotherapy in patients with acute myelogenous leukemia. *Cancer Discov*. 2016;6(10):1106–17.
- Single-agent data with venetoclax in relapsed/refractory AML.
- 103.●● Pollyea DA, Dinardo CD, Thirman MJ, Letai AG, Wei AH, Jonas BA, Arellano ML, Frattini MG, Kantarjian HM, Chyla B, Zhu M, Potluri J, Humerickhouse R, Mabry MH, Konopleva M, Pratz KW. Results of a phase 1b study of venetoclax plus decitabine or azacitidine in untreated acute myeloid leukemia patients ≥65 years ineligible for standard induction therapy. *J Clin Oncol*. 2016;34:7009.
- High efficacy of venetoclax in combination with hypomethylating agents in AML, leading to FDA “breakthrough” therapy designation.
- 104.●● Chan SM, Thomas D, Corces-Zimmerman MR, Xavy S, Rastogi S, Hong WJ, Zhao F, Medeiros BC, Tyvoll DA, Majeti R. Isocitrate dehydrogenase 1 and 2 mutations induce BCL-2 dependence in acute myeloid leukemia. *Nat Med*. 2015;21:178–84. Seminal preclinical paper showing how *IDH1/2* mutations confer BCL-2 dependence.
105. Wei A, Strickland SA, Roboz GJ, Hou J, Fiedler W, Lin TL, Martinelli G, Walter RB, Enjeti A, Fakouhi K, Darden DE, Dunbar M, Zhu M, Agarwal S, Salem AH, Mabry M, Hayslip J. Safety and efficacy of venetoclax plus low-dose cytarabine in treatment-naive patients aged ≥65 Years with acute myeloid leukemia. *Blood*. 2016;128:102.
106. Kojima K, Konopleva M, Samudio IJ, Shikami M, Cabreira-Hansen M, McQueen T, Ruvolo V, Tsao T, Zeng Z, Vassilev LT, Andreeff M. MDM2 antagonists induce p53-dependent apoptosis in AML: implications for leukemia therapy. *Blood*. 2005;106:3150–9.
107. Reis B, Jukofsky L, Chen G, Martinelli G, Zhong H, So WV, Dickinson MJ, Drummond M, Assouline S, Hashemyan M, Theron M, Blotner S, Lee JH, Kasner M, Yoon SS, Rueger R, Seiter K, Middleton SA, Kelly KR, Vey N, Yee K, Nichols G, Chen LC, Pierceall WE. Acute myeloid leukemia patients’ clinical response to idasanutlin (RG7388) is associated with pre-treatment MDM2 protein expression in leukemic blasts. *Haematologica*. 2016;101:e185–8.
108. Martinelli G, Pappayannidis C, Yee K, Vey N, Drummond M, Kelly K, Dickinson M, Lee J, Seiter K, Yoon SS, Assouline S, Kasner M, Nichols G, Middleton S, Blotner S, Zhi J, Pierceall W, Chen LC. Phase 1b results of idasanutlin + cytarabine (Ara-C) in acute myeloid leukemia (AML) patients (pts). 2016:S504-S504.
109. Dinardo CD, Rosenthal J, Andreeff M, Zernovak O, Kumar P, Gajee R, Chen S, Rosen M, Song S, Kochan J, Limsakun T, Olin R. Phase 1 dose escalation study of MDM2 inhibitor DS-3032b in patients with hematologic malignancies—preliminary results. *Blood*. 2016;128:593.
110. Pardoll DM. The blockade of immune checkpoints in cancer immunotherapy. *Nat Rev Cancer*. 2012;12:252–64.
111. Bousiotis VA. Molecular and biochemical aspects of the PD-1 checkpoint pathway. *N Engl J Med*. 2016;375:1767–78.
- 112.●● Davids MS, Kim HT, Bachireddy P, Costello C, Liguori R, Savell A, Lukez AP, Avigan D, Chen YB, McSweeney P, LeBoeuf NR, Rooney MS, Bowden M, Zhou CW, Granter SR, Hornick JL, Rodig SJ, Hirakawa M, Sevegnini M, Hodi FS, Wu CJ, Ho VT, Cutler C, Koreth J, Alyea EP, Antin JH, Armand P, Streicher H, Ball ED, Ritz J, Bashey A, Soiffer RJ. Leukemia and Lymphoma Society Blood Cancer Research Partnership. Ipilimumab for patients with relapse after allogeneic transplantation. *N Engl J Med*. 2016;375:143–53.
- Important study providing proof of concept for immune checkpoint blockade in patients with relapsed/refractory AML and other hematologic malignancies.
113. Orskov AD, Treppendahl MB, Skovbo A, Holm MS, Friis LS, Hokland M, Gronbaek K. Hypomethylation and up-regulation of PD-1 in T cells by azacytidine in MDS/AML patients: a rationale for combined targeting of PD-1 and DNA methylation. *Oncotarget*. 2015;6:9612–26.

114. Yang H, Bueso-Ramos C, DiNardo C, Estecio MR, Davanlou M, Geng QR, Fang Z, Nguyen M, Pierce S, Wei Y, Parmar S, Cortes J, Kantarjian H, Garcia-Manero G. Expression of PD-L1, PD-L2, PD-1 and CTLA4 in myelodysplastic syndromes is enhanced by treatment with hypomethylating agents. *Leukemia*. 2014;28:1280–8.
115. Daver N, Basu S, Garcia-Manero G, Cortes J, Ravandi F, Jabbour EJ, Hendrickson S, Pierce S, Ning J, Konopleva M, Andreeff M, Kornblau SM, Pemmaraju N, Bueso-Ramos C, Blando J, Lopez JEH, Allison J, Kantarjian H, Sharma P. Phase IB/II study of nivolumab in combination with azacytidine (AZA) in patients (pts) with relapsed acute myeloid leukemia (AML). *Blood*. 2016;128:763.
- Promising results with the combination of azacitidine and nivolumab in patients with relapsed/refractory AML.
116. Romagne F, Andre P, Spee P, Zahn S, Anfossi N, Gauthier L, Capanni M, Ruggeri L, Benson Jr DM, Blaser BW, Della Chiesa M, Moretta A, Vivier E, Caligiuri MA, Velardi A, Wagtmann N. Preclinical characterization of 1-7F9, a novel human anti-KIR receptor therapeutic antibody that augments natural killer-mediated killing of tumor cells. *Blood*. 2009;114:2667–77.
117. Daver N, Garcia-Manero G, Basu S, Cortes JE, Ravandi F, Jabbour EJ, Pierce S, Ning J, Konopleva M, Andreeff M, Kornblau SM, Borthakur G, Pemmaraju N, Bueso-Ramos CE, Lopez JEH, Blando J, Allison J, Kantarjian HM, Sharma P. Phase IB/II study of lirilumab in combination with azacytidine (AZA) in patients (pts) with relapsed acute myeloid leukemia (AML). *Blood*. 2016;128:1641.
118. Emadi A, Jun SA, Tsukamoto T, Fathi AT, Minden MD, Dang CV. Inhibition of glutaminase selectively suppresses the growth of primary acute myeloid leukemia cells with IDH mutations. *Exp Hematol*. 2014;42:247–51.
- Preclinical work showing that IDH-mutated AML may be particularly sensitive to glutaminase inhibition.
119. Jacque N, Ronchetti AM, Larrue C, Meunier G, Birsan R, Willems L, Saland E, Decroocq J, Maciel TT, Lambert M, Poulain L, Hospital MA, Sujobert P, Joseph L, Chapuis N, Lacombe C, Moura IC, Demo S, Sarry JE, Recher C, Mayeux P, Tamburini J, Bouscary D. Targeting glutaminolysis has antileukemic activity in acute myeloid leukemia and synergizes with BCL-2 inhibition. *Blood*. 2015;126:1346–56.
- Preclinical work showing that glutaminase inhibition may synergize with BCL-2 blockade in AML.
120. Konopleva M, Contractor R, Tsao T, Samudio I, Ruvolo PP, Kitada S, Deng X, Zhai D, Shi YX, Sneed T, Verhaegen M, Soengas M, Ruvolo VR, McQueen T, Schober WD, Watt JC, Jiffar T, Ling X, Marini FC, Harris D, Dietrich M, Estrov Z, McCubrey J, May WS, Reed JC, Andreeff M. Mechanisms of apoptosis sensitivity and resistance to the BH3 mimetic ABT-737 in acute myeloid leukemia. *Cancer Cell*. 2006;10:375–88.
121. Konopleva M, Milella M, Ruvolo P, Watts JC, Ricciardi MR, Korchin B, McQueen T, Bornmann W, Tsao T, Bergamo P, Mak DH, Chen W, McCubrey J, Tafuri A, Andreeff M. MEK inhibition enhances ABT-737-induced leukemia cell apoptosis via prevention of ERK-activated MCL-1 induction and modulation of MCL-1/BIM complex. *Leukemia*. 2012;26:778–87.
122. Rahmani M, Davis EM, Bauer C, Dent P, Grant S. Apoptosis induced by the kinase inhibitor BAY 43-9006 in human leukemia cells involves down-regulation of Mcl-1 through inhibition of translation. *J Biol Chem*. 2005;280:35217–27.
123. Lehmann C, Friess T, Birzele F, Kiialainen A, Dangl M. Superior anti-tumor activity of the MDM2 antagonist idasanutlin and the Bcl-2 inhibitor venetoclax in p53 wild-type acute myeloid leukemia models. *J Hematol Oncol*. 2016;9(1):50.
124. Jain N, Curran E, Iyengar NM, Diaz-Flores E, Kunnakkam R, Popplewell L, Kirschbaum MH, Karrison T, Erba HP, Green M, Poire X, Koval G, Shannon K, Reddy PL, Joseph L, Atallah EL, Dy P, Thomas SP, Smith SE, Doyle LA, Stadler WM, Larson RA, Stock W, Odenike O. Phase II study of the oral MEK inhibitor selumetinib in advanced acute myelogenous leukemia: a University of Chicago phase II consortium trial. *Clin Cancer Res*. 2014;20:490–8.
125. Zhang W, Konopleva M, Burks JK, Dywer KC, Schober WD, Yang JY, McQueen TJ, Hung MC, Andreeff M. Blockade of mitogen-activated protein kinase/extracellular signal-regulated kinase and murine double minute synergistically induces apoptosis in acute myeloid leukemia via BH3-only proteins Puma and Bim. *Cancer Res*. 2010;70:2424–34.
126. Zhang W, Konopleva M, Ruvolo VR, McQueen T, Evans RL, Bornmann WG, McCubrey J, Cortes J, Andreeff M. Sorafenib induces apoptosis of AML cells via Bim-mediated activation of the intrinsic apoptotic pathway. *Leukemia*. 2008;22:808–18.
127. Kojima K, Konopleva M, Tsao T, Andreeff M, Ishida H, Shiotsu Y, Jin L, Tabe Y, Nakakuma H. Selective FLT3 inhibitor FI-700 neutralizes Mcl-1 and enhances p53-mediated apoptosis in AML cells with activating mutations of FLT3 through Mcl-1/Noxa axis. *Leukemia*. 2010;24:33–43.
128. Kojima K, McQueen T, Chen Y, Jacamo R, Konopleva M, Shinojima N, Shpall E, Huang X, Andreeff M. p53 activation of mesenchymal stromal cells partially abrogates microenvironment-mediated resistance to FLT3 inhibition in AML through HIF-1alpha-mediated down-regulation of CXCL12. *Blood*. 2011;118:4431–9.
129. Kojima K, Kornblau SM, Ruvolo V, Dilip A, Duvvuri S, Davis RE, Zhang M, Wang Z, Coombs KR, Zhang N, Qiu YH, Burks JK, Kantarjian H, Shacham S, Kauffman M, Andreeff M. Prognostic impact and targeting of CRM1 in acute myeloid leukemia. *Blood*. 2013;121:4166–74.
130. Ranganathan P, Yu X, Santhanam R, Hofstetter J, Walker A, Walsh K, Bhatnagar B, Klisovic R, Vasu S, Phelps MA, Devine S, Shacham S, Kauffman M,

- Marcucci G, Blum W, Garzon R. Decitabine priming enhances the antileukemic effects of exportin 1 (XPO1) selective inhibitor selinexor in acute myeloid leukemia. *Blood*. 2015;125:2689–92.
131. Bhatnagar B, Klisovic RB, Walker A, Vasu S, Mims A, Walsh K, Behbehani GK, Blachly JS, Vittorio M, Zhao Q, Ruppert AS, Orwick S, Ranganathan P, Byrd JC, Blum W, Garzon R. A phase 1 clinical trial of selinexor in combination with decitabine in patients with newly diagnosed and relapsed or refractory acute myeloid leukemia. *Blood*. 2016;128:1651.
132. Swords RT, Kelly KR, Smith PG, Garnsey JJ, Mahalingam D, Medina E, Oberheuser K, Padmanabhan S, O'Dwyer M, Nawrocki ST, Giles FJ, Carew JS. Inhibition of NEDD8-activating enzyme: a novel approach for the treatment of acute myeloid leukemia. *Blood*. 2010;115:3796–800.
133. Swords RT, Erba HP, DeAngelo DJ, Bixby DL, Altman JK, Maris M, Hua Z, Blakemore SJ, Faessel H, Sedarati F, Dezube BJ, Giles FJ, Medeiros BC. Pevonedistat (MLN4924), a first-in-class NEDD8-activating enzyme inhibitor, in patients with acute myeloid leukaemia and myelodysplastic syndromes: a phase 1 study. *Br J Haematol*. 2015;169:534–43.
- Single-agent data with first-in-class neddylation inhibitor pevonedistat in AML.
134. Visconte V, Nawrocki ST, Espitia CM, Kelly KR, Possemato A, Beausoleil SA, Han Y, Carraway HE, Nazha A, Advani AS, Maciejewski JP, Sekeres MA, Carew JS. Comprehensive quantitative proteomic profiling of the pharmacodynamic changes induced by MLN4924 in acute myeloid leukemia cells establishes rationale for its combination with azacitidine. *Leukemia*. 2016;30(5):1190–4.
135. Swords RT, Coutre S, Maris MB, Zeidner JF, Foran JM, Cruz JC, Erba HP, Berdeja JC, Tam W, Vardhanabhuti S, Dobler I, Faessel HM, Dash AB, Sedarati F, Dezube BJ, Savona MR. Results of a clinical study of pevonedistat (Pev), a first-in-class NEDD8-activating enzyme (NAE) inhibitor, combined with azacitidine (Aza) in older patients (pts) with acute myeloid leukemia (AML). *Blood*. 2016;128:98.
136. Zhou L, Chen S, Zhang Y, Kmiecik M, Leng Y, Li L, Lin H, Rizzo KA, Dumur CI, Ferreira-Gonzalez A, Rahmani M, Povirk L, Chalasani S, Berger AJ, Dai Y, Grant S. The NAE inhibitor pevonedistat interacts with the HDAC inhibitor belinostat to target AML cells by disrupting the DDR. *Blood*. 2016;127(18):2219–30.
137. Sakurikar N, Eastman A. Will targeting chk1 have a role in the future of cancer therapy? *J Clin Oncol*. 2015;33:1075–7.
138. Karp JE, Thomas BM, Greer JM, Sorge C, Gore SD, Pratz KW, Smith BD, Flatten KS, Peterson K, Schneider P, Mackey K, Freshwater T, Levis MJ, McDevitt MA, Carraway HE, Gladstone DE, Showel MM, Loechner S, Parry DA, Horowitz JA, Isaacs R, Kaufmann SH. Phase I and pharmacologic trial of cytosine arabinoside with the selective checkpoint 1 inhibitor Sch 900776 in refractory acute leukemias. *Clin Cancer Res*. 2012;18(24):6723–31.
139. Dai Y, Chen S, Kmiecik M, Zhou L, Lin H, Pei XY, Grant S. The novel Chk1 inhibitor MK-8776 sensitizes human leukemia cells to HDAC inhibitors by targeting the intra-S checkpoint and DNA replication and repair. *Mol Cancer Ther*. 2013;12(6):878–89.
140. Zhou L, Zhang Y, Chen S, Kmiecik M, Leng Y, Lin H, Rizzo KA, Dumur CI, Ferreira-Gonzalez A, Dai Y, Grant S. A regimen combining the Wee1 inhibitor AZD1775 with HDAC inhibitors targets human acute myeloid leukemia cells harboring various genetic mutations. *Leukemia*. 2015;29(4):807–18.
141. Garcia-Manero G, Atallah E, Khaled SK, Arellano M, Patnaik MM, Odenike O, Sayar H, Tummala M, Patel PA, Ghalie RG, Medeiros BC. A phase 2 study of pracinostat and azacitidine in elderly patients with acute myeloid leukemia (AML) not eligible for induction chemotherapy: response and long-term survival benefit. *Blood*. 2016;128:100.
- Promising efficacy of HDAC inhibitor pracinostat in combination with azacitidine in previously untreated elderly patients with AML.
142. Cortes JE, Heidel FH, Heuser M, Fiedler W, Smith BD, Robak T, Fernandez PM, Ma WW, Shaik MN, Zeremski M, O'Connell A, Chan G. A phase 2 randomized study of low dose Ara-C with or without glasdegib (PF-04449913) in untreated patients with acute myeloid leukemia or high-risk myelodysplastic syndrome. *Blood*. 2016;128:99.
- Survival benefit with the addition of Hedgehog (smoothened) inhibitor glasdegib to low dose cytarabine in newly diagnosed older patients with AML.