ORIGINAL ARTICLE



The expression and clinical significance of GPR39 in colon cancer

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Received: 27 August 2021 / Accepted: 21 September 2021 / Published online: 29 September 2021 © The Author(s), under exclusive licence to Royal Academy of Medicine in Ireland 2021

Abstract

Background Colorectal cancer is the third most common cancer and requires more prognostic biomarkers for precise treatment. GPR39 is a GPCR which can interact with Zn and modulate the colonocytes' survival. The clinical significance of GPR39 in colon cancer has never been reported.

Materials In our study, we compared GPR39 expression between colon cancers and tumor-adjacent tissues by retrieving TCGA data and detected the expression of GPR39 in colon cancers with qPCR and immunohistochemistry. The clinical significance of GPR39 was evaluated by analyzing the correlations with clinicopathological factors with the chi-square test. The prognostic significance of GPR39 was estimated with univariate and multivariate analyses. The expression of several other biomarkers including PPARG, EPCAM, and PD-L1 was investigated by re-analyzing TCGA data, qPCR, and IHC. The prognostic value of PPARG, EPCAM, and PD-L1 was also estimated with univariate analysis.

Results In both TCGA database and our 15 colon cancer pairs, GPR39 expression was significantly upregulated in colon cancer tissues. GPR39 was an independent prognostic biomarker in colon cancer for poor prognosis. With TCGA data reanalysis, qPCR, and IHC, we showed that GPR39 expression was significantly correlated with the expression of EPCAM and PD-L1, but not PPARG. EPCAM and PD-L1 were also unfavorable prognostic biomarkers of colon cancer.

Conclusions GPR39 was upregulated in colon cancer tissues compared with tumor-adjacent tissues. GPR39 was an independent prognostic biomarker in colon cancer for poor prognosis. EPCAM and PD-L1 were substantially associated with GPR39 expression, and they were also identified as prognostic biomarkers in colon cancers.

Keywords Biomarker \cdot Colon cancer \cdot GPR39 \cdot PD-L1 \cdot Prognosis

Introduction

Colorectal cancer (CRC), including colon cancer and rectal cancer, is the third most common cancer and the second leading cause of cancer-related death worldwide [1, 2]. Although the standard chemotherapy and new targeted therapy have been improved in recent years, problems such as multidrug resistance remain unresolved and the prognosis

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of CRC is still unsatisfactory. In China, the 5-year survival rate for CRC is approximately 30% [3–5]. The high heterogeneity of CRC is a main reason of its poor response to some treatments and dismal prognosis [4, 5]. In this era of precise treatment, many breakthroughs have been made to classify CRC based on the molecular signatures, but few results of these studies were used in clinical practice to predict patients' prognoses. The identification of new biomarkers is the basis of new treatment options and target therapies, so more effective biomarkers are required and more individualized treatment strategies of CRC are needed.

G protein-coupled receptors (GPCRs), also known as 7TM receptors, are a large group of cell surface receptors which respond to a variety of external signals and play essential roles in numerous cellular processes [6]. There are more than 600 members of GPCR, and GPR39 is a distinguished GPCR which can sense changes in extracellular Zinc (Zn²⁺) [7]. Zn²⁺ is an essential micronutrient participating in numerous vital physiological processes as a structural cofactor of numerous zinc finger transcription factors or

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enzymes [8]. In epithelial cells, Zn^{2+} can promote epithelial cell proliferation and survival and facilitate tumor progression originated from epithelial cells [9]. In many tissue types including neurons, colon epithelial cells, skin epidermal cells, pancreatic cells, prostate cancer cells, salivary gland cells, and in bones, Zn^{2+} has been reported to regulate the activity of GPR39.

GPR39 mainly couples Gq and activates the IP3 pathway to release intracellular Ca²⁺ from the endoplasmic reticulum [10]. Other reports showed that GPR39 can activate MAPK and AKT and thus increase cell proliferation and migration [10], which suggested a possible role for GPR39 in cancer progression. GPR39 was reported to be overexpressed in breast cancer and to promote breast cancer cell migration and proliferation [11]. The overexpression of GPR39 was also observed in several tumor types such as esophageal squamous cell carcinomas and gastric cancer [12, 13]. Moreover, the activation of GPR39 was also reported in androgen-independent prostate cancer cells [14]. GPR39 is a key receptor of Zn²⁺ which regulates Zn²⁺ signaling especially in epithelial cells including keratinocytes, colonocytes (colon epithelium cells), and salivary gland cells [15]. In colonocytes, GPR39 can reduce cell death by upregulating clusterin [15]. However, the expression and clinical significance of GPR39 in colon cancer are still unknown.

In our study, we retrieved the data from TCGA database and detected the expression of GPR39 in colon cancers with quantitative real-time PCR (qPCR) and immunohistochemistry (IHC). Moreover, we compared GPR39 expression between colon cancers and tumor-adjacent tissues. The clinical significance of GPR39 was evaluated by analyzing the correlations with clinicopathological factors such as histological grade, tumor size, vascular invasion, tumor infiltration, lymphatic invasion, and metastasis. The prognostic significance of GPR39 was also estimated with univariate and multivariate analyses. By analyzing the TCGA database and literature review, we investigated several biomarkers which may be related with GPR39 expression including peroxisome proliferator-activated receptor gamma (PPARG), epithelial cell adhesion molecule (EPCAM), and programmed cell death 1 ligand 1 (PD-L1) and further investigated their expression and prognostic significance in colon cancer.

Materials and methods

Patients and follow-up

Our study was approved by the committee of Yidu Central Hospital and Qilu Hospital of Shandong University. A total of 273 patients were diagnosed with colon cancer and underwent radical surgery from 2010 to 2012 in Yidu Central Hospital and Qilu Hospital. A total of 161 patients were further selected into the final cohort if they had available follow-ups. All the specimens were obtained after the consents of patients. The whole study was approved and supervised by the Ethics Committee of Yidu Central Hospital and Qilu Hospital of Shandong University. The TNM stage was according to the 8th American Joint Committee on Cancer/Union for International Cancer Control (AJCC/ UICC) staging system.

In silico analysis

The data from 275 colon cancers and 349 tumor-adjacent colon tissues were retrieved from the Cancer Genome Atlas (TCGA, http://cancergenome-nih.gov/) for data re-analysis. Transcripts Per Kilobase of exon model per Million mapped reads (TPMs) were applied to evaluate the expression of GPR39, PPARG, EPCAM, and PD-L1. The correlations between GPR39, PPARG, EPCAM, and PD-L1 were also assessed by TPMs. An online website GEPIA was used for data retrieval and analysis (http://gepia.cancer-pku.cn/detail. php).

RNA extraction and qPCR

The mRNA levels of GPR39, PPARG, EPCAM, and PD-L1 in the 15 pairs of colon cancers and the corresponding tumor-adjacent normal tissues were detected with qPCR. Firstly, TRIzol reagent (Thermo Fisher) and RNeasy protect mini kit (Qiagen, Hilden, Germany) were used to extract the total RNAs of these tissues. After that, Primescript RT reagent kit (Takara BIO INC.) was used for reverse transcription PCR. The quantification of qPCR was finally achieved by Thermo Fisher 7500 PCR System. The results were analyzed with the GAPDH as the internal control in a $2^{-\Delta\Delta Ct}$ method. The qPCR primers were designed as follows: GPR39, forward: 5'-CAGGTCCCCGACAAGATC ATA-3', reverse: 5'-TGAGACCGTGTGGTACTTGAG-3'; PPARG, forward: 5'-GATGCCAGCGACTTTGACTC-3', reverse: 5'-ACCCACGTCATCTTCAGGGA-3'; EPCAM, forward: 5'-AATCGTCAATGCCAGTGTACTT-3', reverse: 5'-TCTCATCGCAGTCAGGATCATAA-3'; PD-L1, forward: 5'-TGGCATTTGCTGAACGCATTT-3'. reverse: 5'-TGCAGCCAGGTCTAATTGTTTT-3'; GADPH, forward: 5'-TGTGGGCATCAATGGATTTGG-3', reverse: 5'-ACA CCATGTATTCCGGGTCAAT-3'.

Tissue microarray (TMA) and immunohistochemistry

The 161 cases of formalin-fixed and paraffin-embedded colon cancer tissues were used to construct a TMA with 1 mm cylinders representing each sample. In the TMA, the expressions of GPR39, PPARG, EPCAM, and PD-L1 were detected with IHC. In brief, the TMAs were first

deparaffinized with xylene and rehydrated with graded ethanol. Three percent of hydrogen peroxide was applied to inactivate the endogenous peroxidase activity. Slides were boiled in citrate buffer (pH = 6.0) for 10 min for optimal antigen retrieval, and then in 5% bovine serum albumin for 30 min to eliminate unspecific antigen binding. The primary antibodies of GPR39 (Novus Biologicals, catalog: NLS139), PPARG (Santa Cruz Biotechnology, catalog: sc-7273), EPCAM (BioLegend, catalog: 324,202), and PD-L1 (Invitrogen, catalog: 14-5983-80) were used to incubate the specimen at 4 °C overnight. The biotinlabeled secondary antibodies (Sangon, Shanghai, China) and streptavidin-peroxidase (Sangon) were used to incubate the slides. Finally, the visualization of slides was achieved by incubation 3, 3'-diaminobenzidine substrate for 10 min.

Evaluation of IHC results

The results of IHC were semi-quantified by evaluation with two senior pathologists who were unaware of the clinical information. The IHC score was defined as the score of staining intensity multiplied by the score of IHC positive cell percentage. The scores of staining intensity were set as score 0 representing negative staining, 1 representing weak staining, 2 representing moderate staining, and 3 representing strong staining. The scores of IHC positive cell percentage were defined as score 1 representing less than 25% of positive cells, score 2 representing 25–50% of positive cells, score 3 representing 50-75% of positive cells, and score 4 representing more than 75% of positive cells. So the final IHC scores ranged from 0 to 12. The patients were then divided into subgroups with low and high IHC scores of these biomarkers by the cut-off, which was identified by the receiver operating characteristic (ROC) curve.

Statistical analysis

SPSS 22.0 (IBM cooperation, Chicago, USA) was used to analyze the data. One-way ANOVA was used to compare TPMs of GPR39 in colon cancers and adjacent tissues. Paired t test was used to compare GPR39 expression in 15 pairs of colon cancer tissues. The correlations between the expression of GPR39 and the clinicopathological variables were analyzed with the chi-square test. Kaplan–Meier method was used to plot the overall survival (OS) curves, and the log-rank test was used to calculate the statistical difference between different groups. The Cox-regression hazard model was applied to identify the independent prognostic factors. P < 0.05 was considered as statistically significant.

Results

GPR39 was overexpressed in colon cancer compared with adjacent tissues

The expression of GPR39 was first evaluated by retrieved the data from TCGA database. TPMs were used to evaluate GPR39 mRNA level in 275 colon tumors and 349 tumoradjacent colon tissues (Fig. 1A). The TPMs of GPR39 in tumor tissues were significantly higher than TPMs in adjacent tissues. Moreover, we collected 15 pairs of colon tumors and tumor-adjacent colon tissues and detected the GPR39 mRNA levels with qPCR. The qPCR results were in consistent with the mRNA sequencing results, which indicated that GPR39 was overexpressed in colon cancer tissues (Fig. 1B). Furthermore, we detected the expression of GPR39 with IHC in 161 colon cancer tissues (Fig. 1C).



Fig. 1 GPR39 expression in colon cancers. **A** The TPMs of GPR39 in 275 cases of colon cancers and 349 cases of adjacent tissues were compared with one-way ANOVA test. **B** GPR39 mRNA levels in 15 pairs of colon cancers and tumor-adjacent tissues were detected with qPCR. Statistical significance was evaluated with paired *t* test, and data were from 3 independent experiments. **C** IHC was used to detect GPR39 expression in 161 cases of colon cancer. **D** The magnified images of the boxes in **C**

| Factors | | GPR39 | GPR39 | |
|---------------------|-----|-------|-------|-------|
| | | Low | | |
| Sex | | | | |
| Male | 80 | 38 | 42 | 0.048 |
| Female | 81 | 51 | 30 | |
| Age | | | | |
| <60 | 62 | 32 | 30 | 0.459 |
| ≥60 | 99 | 57 | 42 | |
| Tumor diameter (cm) | | | | |
| ≤5 | 68 | 35 | 33 | 0.406 |
| >5 | 93 | 54 | 39 | |
| Histological grade | | | | |
| Ι | 41 | 26 | 15 | 0.402 |
| II | 79 | 43 | 36 | |
| III | 41 | 20 | 21 | |
| T stage | | | | |
| T1 + T2 | 19 | 16 | 3 | 0.007 |
| T3 + T4 | 142 | 73 | 69 | |
| Lymph node invasion | | | | |
| No(N0) | 105 | 59 | 46 | 0.75 |
| Yes(N1/2) | 56 | 30 | 26 | |
| Metastasis | | | | |
| Negative | 150 | 86 | 64 | 0.064 |
| Positive | 11 | 3 | 8 | |
| Vascular invasion | | | | |
| Negative | 102 | 60 | 42 | 0.234 |
| Positive | 59 | 29 | 30 | |
| TNM stage | | | | |
| Ι | 34 | 25 | 9 | 0.034 |
| II | 69 | 33 | 36 | |
| III | 53 | 30 | 23 | |
| IV | 5 | 1 | 4 | |

 Table 1 Correlations between GPR39 expression and the clinicopathological factors

*Chi-square test

As a GPCR, GPR39 was expressed in the cell membrane in our study as expected (Fig. 1D). The 161 colon cancer tissues were further divided into subgroups which had low or high GPR39 expression, accounting for 55.28% (89/161) and 44.72% (72/161), respectively.

GPR39 was associated with patients' sex, T, and TNM stage

In the 161 patients, we performed the chi-square test to screen the potential tumor-associated progression which GPR39 may be involved in. In our study, GPR39 was significantly associated with T stage (P = 0.007) and TNM stage (P = 0.034). These results suggested that high expression of GPR39 in colon tumors indicated the advanced T stage and

TNM stage (Table 1). Interestingly, we also observed that GPR39 correlated with patients' sex in colon cancer. Female patients tended to have high GPR39 expression (P = 0.048).

GPR39 expression was correlated with the OS in colon cancers

In addition, we analyzed the correlation between the clinicopathological factors, GPR39 expression, and the OS with univariate analysis (Table 2). In our cohort, clinicopathological factors including histological grade (P < 0.001), lymphatic invasion (P < 0.001), metastasis (P = 0.002), vascular invasion (P = 0.016), and TNM stage (P = 0.001) were all prognostic factors. High histological grade, positive

Table 2 The univariate analysis to identify the prognostic factors

| Factors | 5-year OS rate | <i>P</i> * | |
|---------------------|----------------|------------|--|
| Sex | | | |
| Male | 60.7 | 0.658 | |
| Female | 58.9 | | |
| Age | | | |
| <60 | 54.3 | 0.347 | |
| ≥60 | 63.3 | | |
| Tumor diameter (cm) | | | |
| ≤5 | 63.0 | 0.671 | |
| >5 | 57.3 | | |
| Histological grade | | | |
| Ι | 84.4 | < 0.001 | |
| II | 72.7 | | |
| III | 38.6 | | |
| T stage | | | |
| T1 + T2 | 68.4 | 0.389 | |
| T3 + T4 | 58.4 | | |
| Lymph node invasion | | | |
| No(N0) | 68.0 | < 0.001 | |
| Yes(N1/2) | 44.2 | | |
| Metastasis | | | |
| Negative | 61.5 | 0.002 | |
| Positive | 36.4 | | |
| Vascular invasion | | | |
| Negative | 67.3 | 0.016 | |
| Positive | 46.6 | | |
| TNM stage | | | |
| Ι | 76.5 | 0.001 | |
| II | 62.8 | | |
| III | 46.7 | | |
| IV | 40 | | |
| GPR39 | | | |
| Low | 67.9 | 0.002 | |
| High | 49.7 | | |

*Log-rank test

lymphatic invasion, metastasis and vascular invasion, and advanced TNM stage were all significantly associated with low OS rates in colon cancer (Fig. 2A–E). Moreover, GPR39 was identified as a significant biomarker predicting the poor prognosis of colon cancer (P=0.002). The 5-year OS rates of patients with low and high expression of GPR39 were 67.9% and 49.7%, respectively (Fig. 2F). Other clinicopathological factors including T stage (P=0.389) and tumor size (P=0.671) exhibited no significant statistical difference towards the OS rate in colon cancer(Fig. 2G, H).

GPR39 as an independent prognostic factor in colon cancer

All the prognostic factors in univariate analysis were enrolled into the multivariate analysis for the identification of independent prognostic factors of colon cancer in our study. The enrolled factors included histological grade, lymphatic invasion, metastasis, vascular invasion, and GPR39 expression (Table 3). TNM stage was a consequent factor constituted by T, N, and M stage, so it was excluded from the multivariate analysis. In the Cox-regression model for multivariate analysis, GPR39 expression was identified as an independent risk for unfavorable prognosis (P = 0.002). The cancer-related death odds of patients with high GPR39 expression was 2.17-fold higher than the patients with low GPR39 expression (HR = 2.17). In addition, high



Fig. 2 Survival significance of clinicopathological factors and GPR39 expression. **A**–**F** Histological grade, lymph node invasion, metastasis, vascular invasion, TNM stage, and GPR39 expression were all prognostic factors in colon cancers. Kaplan–Meier method was used to plot the OS curves and the log-rank test was used to calculate the statistical differences. **G** and **H** The correlations between T stage, tumor size, and the OS rate had no significant difference

 Table 3
 The Cox-regression model to identify independent prognostic factors

| Factors | HR | 95%CI | P^* |
|---------------------|------|-----------|---------|
| Histological grade | | | |
| I+II | 1.00 | | |
| III | 2.67 | 1.59-4.47 | < 0.001 |
| Lymph node invasion | | | |
| No(N0) | 1.00 | | |
| Yes(N1/2) | 2.12 | 1.30-3.44 | 0.002 |
| Metastasis | | | |
| Negative | 1.00 | | |
| Positive | 2.50 | 1.23-5.07 | < 0.001 |
| Vascular invasion | | | |
| Negative | 1.00 | | |
| Positive | 1.33 | 0.81-2.19 | 0.263 |
| GPR39 | | | |
| Low | 1.00 | | |
| High | 2.17 | 1.33–3.53 | 0.002 |

*Cox-regression model

histological grade (P < 0.001), positive lymph node invasion (P = 0.002), and positive metastasis (P < 0.001) all indicated the poor prognosis independently in colon cancer (Table 3).

GPR39 expression was significantly associated with EPCAM and PDL1 expression

From the TCGA database, we screened the potential genes which may be associated with GPR39. PPARG and EPCAM were selected because they had possible positive correlations with GPR39 in TCGA database (Fig. 3A), and they were reported to be involved in tumor progression and prognosis in colon cancer [16-18]. In our study, PD-L1 was also detected because of its essential role in the response to immunotherapy. We detected their mRNA correlations with GPR39 mRNA in the 15 patients with colon cancer. Interestingly, there were no substantial correlations between GPR39 and PPARG, though PPARG was shown to be associated with GPR39 with mRNA sequencing in TCGA database. EPCAM and PD-L1 mRNA expression had significant correlations with GPR39 in the consecutive cohort consisting of 15 colon cancers (Fig. 3B). Moreover, we detected PPARG, EPCAM, and PD-L1 in the 161 cases of colon cancer and divided them into subsets according to the different expressions of PPARG, EPCAM, and PD-L1 (Fig. 3C). In colon cancer, PPARG was mainly localized in cell nucleus, EPCAM was localized in both cell nucleus and cytoplasm, and PD-L1 was mainly expressed in membrane (Fig. 3D). In consistent with the qPCR results in Fig. 3B, patients with high GPR39 expression had a high expression level of EPCAM and PD-L1, but not PPARG (Fig. 3E).



Fig.3 GPR39 expression correlated with EPCAM and PD-L1 expression. **A** In TCGA database, the GPR39 expression exhibited significant correlation with PPARG and EPCAM in 275 colon cancers. *R* value was calculated by the Spearman analysis. **B** The correlations between PPARG, EPCAM, and PD-L1, and GPR39 expression were analyzed in the 15 colon cancer tissues with the Spearman analysis. **C** and **D** PPARG, EPCAM, and PD-L1 expressions in 161 colon cancer tissues were detected with IHC. **E** Patients with high GPR39 expression had more expression of EPCAM and PD-L1, but not PPARG. * represents P < 0.05, ** represents P < 0.01 with *t* test

EPCAM and PD-L1 were also prognostic biomarkers of colon cancer

After detection of EPCAM, PD-L1, and GPR39 expression with IHC, we analyzed the correlation between EPCAM,

PD-L1, and GPR39 expression with the chi-square test. EPCAM (P = 0.001) and PD-L1 (P = 0.001) expression were all positively associated with GPR39 expression (Table 4). The prognostic value of PPARG, EPCAM, and PD-L1 was evaluated with univariate analysis. Intriguingly, PPARG exhibited no correlation with the OS in colon cancer (P = 0.350) (Fig. 4A), while both EPCAM (P = 0.019) and PD-L1 (P = 0.033) indicated the poor prognoses of colon cancer (Fig. 4B, C).

Discussion

 Zn^{2+} is a critical metal elements in human and the deficiency of Zn^{2+} is linked to many diseases mainly in the digestive, immune, nervous, endocrine, and integumentary systems, such as impaired learning and memory, diarrhea and taste disorders [19, 20]. Zn^{2+} is an essential structural element and cofactor in enzymes and transcription factors, and there are approximate 3000 proteins which have Zn^{2+} binding sites and their activities were regulated by Zn^{2+} interaction [21–23]. Although it has been decades since the essential role of Zn^{2+} has been defined, the mechanisms underlying Zn²⁺-involved cellular processes and tumor progression are still poorly understood [24]. GPR39 is known as Zn^{2+} sensing receptor can interact with extracellular Zn²⁺ at physiological concentrations, couples Gq-protein and therefore triggers the IP3 pathway to release intracellular Ca^{2+} [10]. In colonocytes, GPR39 was showed to activate downstream signaling pathways such as MAPK-ERK signaling and PI3K-AKT pathway [25–27]. In our study, we for the first time investigated the expression of GPR39 in colon cancer and the corresponding tumor-adjacent tissues. Furthermore, we identified GPR39 as an independent prognostic biomarker of colon cancers. This is an important supplement of the oncogenic role of GPR39 because there are quite few studies on the function of GPR39 in cancer. The expression of

| Factors | | GPR39 | | <i>P</i> * | Univariate analysis | |
|---------|----|-------|------|------------|---------------------|-----------------------|
| | n | Low | High | | 5-year OS rate | <i>P</i> [#] |
| PPARG | | | | | | |
| Low | 84 | 42 | 42 | 0.212 | 56.4 | 0.350 |
| High | 77 | 47 | 30 | | 63.5 | |
| EPCAM | | | | | | |
| Low | 89 | 61 | 28 | 0.001 | 66.7 | 0.019 |
| High | 72 | 28 | 44 | | 51.3 | |
| PDL1 | | | | | | |
| Low | 79 | 55 | 24 | 0.001 | 68.1 | 0.033 |
| High | 82 | 34 | 48 | | 52.3 | |

*Chi-square test

[#]Log-rank test

Table 4The correlationbetween GPR39 expression,PPARG, EPCAM, and PDL1

expressions



Fig. 4 Survival significance of PPARG, EPCAM, and PD-L1 expression. A–C PPARG had no significant correlation with OS rate (A), but high expression of EPCAM (B) and PD-L1 (C) was correlated with low OS rate in colon cancer. P value was generated by the log-rank test

GPR39 was only studied in prostate cancer, breast cancer and gastric cancer till now.

The AJCC TNM system is the only well-accepted system to predict the prognosis and select patients for adjuvant therapy until now [28]. In recent years, several studies tried to classify CRC according to the molecular characteristics [29]. Personalized medicine is relied on the stratification of tumors into subtypes by molecular fingerprints associated with different prognoses or therapeutic responses. For example, in colorectal cancer, KRAS mutations correlates with poor prognosis and guide therapeutic decisions [30, 31]. However, the high-throughput methods such as mRNA sequencing need the validation of protein level detection, and the results need further verification of larger cohorts and multi-centers. In our study, we identified GPR39 as an independent prognostic biomarker in colon cancer for the first time, which suggested that detection of GPR39 expression could stratify the high-risk patients more precisely. This is an important supplement to the post-operational detection of colon cancer, and may be a new elicitation to the precise treatment of patients with colon cancer.

Till now, there is no well-accepted biomarker and criteria to predict the response to immunotherapy, mainly including the immune checkpoint inhibitor such as PD1 antagonist. It is still controversial whether PD-L1 expression is associated with the response to PD1 inhibitor [32]. Here we detected the expression of PD-L1 in colon tumor cells and interestingly demonstrated that PD-L1 was a prognostic biomarker of colon cancer. All the patients in our cohort received no immunotherapy because the establishment of cohort is more than 10 years ago, when the therapy of immune checkpoint inhibitor is not so common. In these patients, high expression of PD-L1 indicated the poor prognosis of colon cancer, which may be explained that high PD-L1 in tumor cells could inhibit the immune cells and promote the immune escape in the microenvironment of colon cancer. However, whether these patients have better response to immunotherapy requires further clinical study to demonstrate.

PPARG is a kind of nuclear receptor which interacts with peroxisome proliferators such as hypolipidemic drugs and fatty acids. PPARG rs3856806 C>T polymorphism increased the risk of colorectal cancer in Eastern Chinese Han population [33], but PPARG was also reported to be correlated with good prognosis in colorectal cancer in a cohort consisting of American patients [16], which was not validated in our cohort. In addition, plenty of evidence showed that EPCAM could promote the progression of colon cancer [17, 18], and indicated that EPCAM antibody can help treat colon cancer [34]. In our stydy, we screened the potential biomarkers which may be associated with GPR39 expression in colon cancer in TCGA database and selected PPARG and EPCAM by literature review. Interestingly, only EPCAM had positive correlation with GPR39 and OS rate, though PPARG and EPCAM exhibited strong correlations with GPR39 in mRNA sequencing results in TCGA. This result showed that the validation of protein level to highthroughput sequencing is necessary because that the probes of mRNA sequencing may be not that special and many post-translational changes after mRNA translation could not be ignored. For the first time, we showed that PD-L1 and EPCAM expressions were significantly associated with GPR39, which is an interesting result and needs further study to elucidate the mechanism. Our results indicated that PD-L1 or EPCAM may be downstream factors responsible for the GPR39-induced prognosis of colon cancer. This may elicit a new direction of GPR39 and colon cancer study.

The importance of GPR39 has been investigated in many diseases, including inflammatory bowel diseases, epilepsy, depression, and cancer [7]. Moreover, GPR39 knock-out mice have heavier body weight, more food intake, and energy expenditure, indicating that GPR39 may be also involved in metabolic processes [35]. Taken together, GPR39 is a promising drug target considering the essentiality of Zn²⁺ in numerous cellular processes and the clinical significance of GPR39. With a variety of high-throughput screenings, several agonists for GPR39 have been proposed but very few are successfully validated by in vivo experiments [36]. Recently, an oral active GPR39 agonist, called Cpd1324, was developed as a weight-lowering agent due to its stimulatory effect on GLP-1 secretion [37]. However, the specific inhibitor of GPR39 has not been identified. To screen and validate the effective inhibitor of GPR39 is a promising but challenging topic to elucidate the underlying mechanism of GPR39-involved cellular processes and help develop new therapeutic strategies to treat GPR30-related diseases.

In summary, we demonstrated that GPR39 was significantly upregulated in colon cancer tissues compared with tumor-adjacent tissues by TCGA database re-analysis and qPCR. Moreover, we detected GPR39 expression in 161 colon cancer tissues and showed that GPR39 was an independent prognostic biomarker. With TCGA data re-analysis, qPCR, and IHC, we showed that GPR39 expression was significantly correlated with the expression of EPCAM and PD-L1, but not PPARG. EPCAM and PD-L1 were also unfavorable prognostic biomarkers of colon cancer.

Abbreviations GPR39: G-protein coupled receptor 39; PPARG: Peroxisome proliferator-activated receptor gamma; EPCAM: Epithelial cell adhesion molecule; PD-L1: Programmed cell death 1 ligand 1; qPCR: Quantitative real-time PCR; IHC: Immunohistochemistry; TCGA: The Cancer Genome Atlas; CRC: Colorectal cancer; MAPK: Mitogen-activated protein kinase; AKT: RAC-alpha serine/ threonine-protein kinase; TPM: Transcripts per million

Declarations

Conflict of interest The authors declare no competing interests.

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