Anti-inflammatory and Immunomodulatory Effects of Marine n-3 Polyunsaturated Fatty Acids on Human Health and Diseases

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Abstract The pharmaceutical effects of n-3 polyunsaturated fatty acids (n-3 PUFAs) as dietary nutrients on human health and diseases have gained much attention and are investigated for decades. Docosahexaenoic acid (DHA), eicosapentaenoic acid (EPA) and docosapentaenoic acid (DPA) are the three major n-3 PUFAs enriched in marine organisms, such as fish, shrimp, algae, and so on. It has been well known that n-3 PUFAs, especially DHA and EPA, are beneficial in reducing the risk of cardiovascular and cerebrovascular diseases. Accumulating evidence suggests that n-3 PUFAs might cure inflammatory diseases through several mechanisms, such as plasma membrane remodeling of lymphocytes, down-regulating pro-inflammatory cytokines, and alternating adhesion molecule expressions. Several molecular targets of n-3 PUFAs on immune-regulation have also been identified, such as GPR120 (FFA4), protein kinase C (PKC), and PPAR-γ. However, it remains inconclusive if dietary n-3 PUFAs function the same both *in vitro* and *in vivo* based on cohort studies. This review will focus on the molecular targets and mechanisms of anti-inflammatory and immunomodulatory effects of n-3 PUFAs on human health and diseases, such as obesity, tumor, diabetes, and autoimmune diseases.

Key words n-3 polyunsaturated fatty acids; immune-regulation; anti-inflammation; mechanism

1 Introduction

In 1978, Dyerberg and Bang first reported that the incidence and mortality of cardiovascular and cerebrovascular diseases in Eskimos are significantly lower than that of people living in other parts of the world based on the epidemiological study. The reasonable explanation is the high content of n-3 PUFAs (DHA and EPA) in arctic fish consumed in their diets (Dyerberg *et al.*, 1978; Dyerberg and Bang, 1978). Since then, scholars have paid much attention to n-3 PUFAs for their potential therapeutic role in treating various human diseases, such as cardiovascular diseases, thrombosis, atherosclerosis, stroke, and diabetes (Dawczynski *et al.*, 2013; Jacobo-Cejudo *et al.*, 2017; Nosaka *et al.*, 2017; Phang *et al.*, 2011).

Marine fish, shellfish and microalgae are enriched in n-3 PUFAs. For example, the contents of EPA plus DHA are $717-1533 \text{ mg}(100 \text{ g})^{-1}$ in salmon, $236 \text{ mg}(100 \text{ g})^{-1}$ in Alaskan pollock, $44 \text{ mg}(100 \text{ g})^{-1}$ in channel catfish, and $253 \text{ mg}(100 \text{ g})^{-1}$ in Atlantic cod (Cladis *et al.*, 2014). It is estimated that demand for n-3 PUFAs world widely would

be 241 thousand metric tons with a value of US \$4.96 billion. However, debates over the actual effects of n-3 PUFAs on improving human health have never stopped and contradictory findings are reported periodically (Djousse *et al.*, 2010; Poreba *et al.*, 2017; Wu *et al.*, 2012; Zhang *et al.*, 2013). It seems that the source and dosage of n-3 PUFAs, trial duration, sex, and age could influence the effects and conclusions (Abbott, 2016; Chen *et al.*, 2017). In addition, the ratio of different PUFAs (n-3: n-6) and fat-soluble small molecules, such as vitamins and other unknown molecules, may contribute to the discrepancies.

Many reviews and meta-analysis about n-3 PUFAs have been reported. However, most of them are related to the effects of n-3 PUFAs on human health and diseases such as cardiovascular diseases, non-alcoholic fatty liver, and atherothrombosis (Alexander *et al.*, 2017; Auger *et al.*, 2016; Chen *et al.*, 2016; Chiesa *et al.*, 2016; de Castro and Calder, 2017; Fialkow, 2016a; Kelley, 2016; Rizos and Elisaf, 2017; Siscovick *et al.*, 2017). The benefits of dietary n-3 PUFAs in neuron development are also discussed (Luchtman and Song, 2013; Marszalek and Lodish, 2005; Wurtman, 2008). This review article will be focused on the effects of n-3 PUFAs on inflammation and immune regulation including their potential targets and molecular mechanisms in the following human diseases:

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obesity, cancer, type I diabetes mellitus, and autoimmune diseases. The intrinsic immune association of these human diseases is a promising starting point to understand the effects of n-3 PUFAs on regulating inflammation and immunity and to evaluate both *in vitro* and *in vivo* findings about n-3 PUFAs.

1.1 N-3 Polyunsaturated Fatty Acids (n-3 PUFAs)

Fatty acids are the fundamental components of the cell membrane and the starting materials for the biosynthesis of a variety of important biological molecules. However, the term 'essential fatty acids' are applied only to those polyunsaturated fatty acids (PUFAs) that are necessary for maintaining human health and cannot be synthesized by the cells in the human body (Spector, 1999). PUFAs can be divided into two classes: n-3 and n-6. The nomenclature is based on the location of the farthest ethylenic linkage from the carboxyl group. Strictly speaking, except for alpha-linolenic acid (ALA), other n-3 PUFAs are not 'essential fatty acids', because they can be transformed and synthesized from ALA in the human body (Fig.1) (Arnoldussen and Kiliaan, 2014; Edwards and O'Flaherty, 2008). Nevertheless, since the rate of conversion of ALA to other n-3 PUFAs, like EPA and DHA, is far from meeting daily needs, dietary intake of PUFAs are necessary (Williams and Burdge, 2006).

So far, there is no uniform dosage for the daily intake of dietary n-3 PUFAs. Based on current studies, different research institutions have given their own recommendations based on age, gender and health status (Tables 1–2).

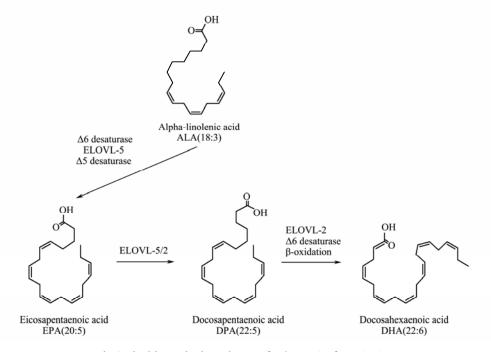


Fig.1 The biosynthetic pathway of n-3 PUFAs from ALA.

Table 1 Daily	y recommended	dietary int	ake of n-3	PUFA in adults

Organization	n-3 PUFA (EPA and DHA) daily recommendation	Subjects
Department of Health (DOH, UK)	200 mg	Healthy individuals
International society for the study of fatty acids and lipids (ISSFAL)	500 mg	Healthy individuals
Dietary Guidelines Advisory Committee	496 mg	Healthy individuals
World Health Organization (WHO)	200 – 500 mg	Healthy individuals
Chinese Nutrition Society (CNS)	Fish dish $(280-525 \text{ g week}^{-1})$	Healthy individuals
National Institutes of Health (NIH)	300 mg	pregnant and lactating women
British Nutrition Foundation (BNF)	500 – 1000 mg	Cardiovascular patients
American Heart Association (AHA)	1000 mg	Cardiovascular patients
	Fish dish (twice/week)	Healthy individuals
European Society of Cardiology (ESC)	1000 mg	Cardiovascular patients
Belgian Superior Health Council	1000 mg	Cardiovascular patients
Food and Agriculture Organization of	250 mg	Healthy individuals
the United Nations (FAO)	2000 mg	Cardiovascular patients
	500 mg	Healthy individuals
The Heart Foundation	1000 mg	Cardiovascular patients
	1200 mg	Hyperlipemia patients

Notes: Dietary guidelines for Americans: Washington 2005, position statement on fish, fish oils, n-3 polyunsaturated fatty acids and cardiovascular health, AIFST conference 2008, fats and fatty acids in human nutrition 2010; Kris-Etherton *et al.*, 2002a; Van de Werf *et al.*, 2003.

Organization	Recommendation (mgd^{-1})		A
Organization	Male	Female	Age
	900	900	4 - 8
Institute of Medicine (IOM, USA)	1200	1000	9-13
	1600	1100	14 - 18
	900	900	4 - 8
Dietary Guidelines Advisory Committee (USA)	1200	1000	9-13
	1600	1100	14 - 18
	1300	1100	3-5
	1400	1300	6-7
	1700	1400	8-9
Ministry of Health, Labor and Welfare (MHLW, Japan)	1700	1500	10 - 11
	2100	1800	12-14
	2300	1700	15-17

Table 2 Daily recommended dietary intake of n-3 PUFA in childhoods

Notes: 'Population nutrient intake goals for preventing diet-related chronic diseases', 2014, http://www.who.int/nutrition/topics/ 5_population_nutrient/en/index25.html. 'Overview of dietary reference intakes for Japanese', 2015, http://www.mhlw.go.jp/bunya/ kenkou/sessyu-kijun.html. Trumbo *et al.*, 2002.

1.2 Inflammation and Immune Regulation

Inflammation is a complex biological response of the host to infections and injuries from pathogens, damaged cells, or irritants (Ferrero-Miliani *et al.*, 2007). Immunocyte recruitment and inflammatory cytokines secretion at lesions are always involved in the response. The function of inflammation is to eliminate the cause of cell damage and to initiate tissue repair. Cellular signaling pathways play pivotal roles in inflammation. Three classical signal transduction pathways, JAK-STAT, MAPK, and Nf- κ B, are associated with inflammation. They can regulate the inflammation individually or form a regulatory network (Fig.2) (Arbabi and Maier, 2002; Davis, 2000; Dell'Albani *et al.*, 2001; Jiang *et al.*, 1997; Seger and Krebs, 1995).

Immune regulation is the interaction between immune cells and immune molecules within the immune system, as well as with other systems, to make the immune response at a proper level (Boothby and Rickert, 2017; Jin *et al.*, 2017; Thaiss *et al.*, 2016; Verbist *et al.*, 2012). To

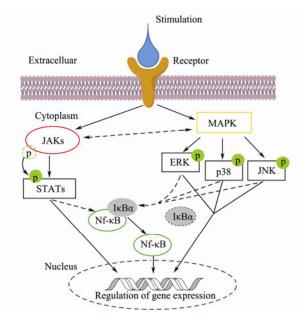


Fig.2 The cross-talk of JAK, MAPK and Nf- κ B signaling pathways.

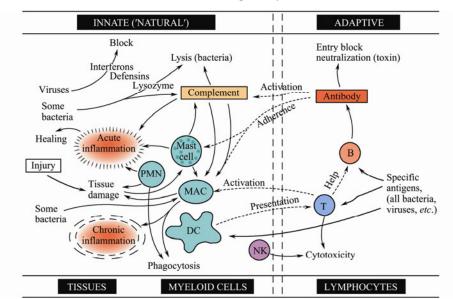


Fig.3 Relationship between inflammation and immune regulation (cited from Immunology at a glance. 10th ed, 2013, with permission from Wiley-Blackwell).

function properly, immune system should have the ability to detect various pathogens from viruses to bacteria, or some other microorganisms like dust mite, and to distinguish them from the body's own tissues. Disorders of the immune system are catastrophic, which would result in autoimmune diseases, inflammatory diseases and cancers (O'Byrne and Dalgleish, 2001). The human immune system can be classified into innate immune system and adaptive immune system. Though, inflammation and immune regulation are different concepts, there is a close relationship between them in the process of host resisting pathogens (Fig.3) (Playfair and Chain, 2013).

2 Function of n-3 PUFAs in Regulating Obesity- and Cancer-Induced Inflammation

Mounting evidence shows that there is a strong association between obesity and cancer (Bandini *et al.*, 2017; Carreras-Torres *et al.*, 2017; Dobbins *et al.*, 2013; Hidayat *et al.*, 2016; Sun *et al.*, 2015; Zhang *et al.*, 2014). Actually, obesity is not only a risk factor for cancer, both obese and cancer patients have some similar physiologic index because they all suffer a long-term disease-associated inflammation (Divella *et al.*, 2016). For example, an elevated levels of pro-inflammatory factors, such as C-reactive protein, TNF- α , and IL-6, can be observed in their blood circulation.

Recently, the benefits of n-3 PUFAs to the obese and cancer patients have gained much attention. Studies show that n-3 PUFAs can suppress adipogenesis, and dietary n-3 PUFAs supplementation is helpful to lose weight for the obese, but show little effect in healthy human subjects (Hooper, 2007; Li et al., 2008). N-3 PUFAs can influence glucose and lipid metabolism through lowering triacylglycerol concentration, boosting high density lipoprotein (HDL) cholesterol levels in plasma, and improving insulin sensitivity (Delarue et al., 2004; Kris-Etherton et al., 2002b). Furthermore, experiments on animal model indicate that the weight loss is not on account of less caloric intake, but the regulation of lipid metabolism-related gene expression, more energy expenditure and modulation of adipocyte differentiation (Buettner et al., 2006; Huang et al., 2016; Ralston et al., 2015).

For cancer patients, n-3 PUFAs have showed anti-tumor effect both *in vitro* and *in vivo* (Xue *et al.*, 2014; Zou *et al.*, 2013). Besides that, n-3 PUFAs can reduce the side effects of chemotherapy and radiotherapy during cancer treatment and also have chemoprevention effects. (Calviello *et al.*, 2009; Fasano *et al.*, 2010). The mechanisms underlying the anti-tumor effects of n-3 PUFAs are multiple including cell cycle arrest and apoptosis induction (So *et al.*, 2015).

Except for the above mentioned function of n-3 PUFAs on obesity and cancer, they also exhibit the ability of alleviating obesity- and cancer-induced inflammation. In patients, cyclooxygenase and 5-lipoxygenase convert membrane-derived arachidonic acid (AA, an n-6 PUFA) into pro-inflammatory eicosanoids, which is an important reason for long-lasting inflammation (Flachs *et al.*, 2009). Different from n-6 PUFAs, the metabolites of n-3 PUFAs are beneficial in anti-inflammation (James *et al.*, 2000a). Additional consumption of n-3 PUFAs can significantly suppress AA conversion to pro-inflammatory eicosanoids (Lee *et al.*, 1985; Lokesh and Kinsella, 1987). Inflammatory cytokine production will be inhibited too. For example, after intervention of n-3 PUFAs, the levels of TNF- α and IL-6 at nidus are decreased (Muurling *et al.*, 2003; Perez-Echarri *et al.*, 2008). In addition, when n-3 PUFAs combine with cancer chemopreventive agents, such as curcumin, the synergistic effects are observed by reducing inflammatory ROS level through downregulating of iNOS, COX-2, and cLPA2 (Saw *et al.*, 2010).

3 Benefits of n-3 PUFAs on Auto-Immune Disease and Type I Diabetes

Auto-immune disease is caused by over response to the self-antigen of the body's immune system. In recent years, many diseases have been classified as autoimmune diseases, such as systematic lupus erythematosus (SLE), rheumatoid arthritis (RA), ankylosing spondylitis (AS), autoimmune hemolytic anemia (AIHA) and others (Emmungil *et al.*, 2014; Giacomelli *et al.*, 2017; Kamesaki, 2017).

Type I diabetes is characterized as absolutely insufficient in insulin production, and it usually occurs in children and adolescent. The assumption that type I diabetes is an auto-immune disease was firstly proposed in 1965 (Eisenbarth *et al.*, 1988). To date, some evidence supports the hypothesis: 1) when treating leukemia patients through bone marrow transplantation from type I diabetes donor, the acceptor develops diabetes mellitus; 2) insulitis is mainly caused by lymphocyte, macrophage or monocyte infiltration; 3) immunosuppressive agents can slow the pathogenesis of pancreatic islet beta cell injury in newly diagnosed type I diabetes patients (Bottazzo *et al.*, 1985; Domenick and Ildstad, 2001; Hyslop *et al.*, 2016; Orban *et al.*, 2011).

Mounting work suggests that long-term consumption of n-3 PUFAs can provide symptom relief of auto-immune diseases and type I diabetes. In autoimmune-mediated glomerulonephritis, intake of n-3 PUFAs can decrease proteinuria and improve glomerular filtration rate (GFR) (Holman et al., 1994; Pestka, 2010). The study of DAISY (Diabetes Autoimmunity Study in the Young) found that dietary supplementation of n-3 PUFAs from 1 year old could reduce the risk of islet autoimmunity in children with type I diabetes (Norris et al., 2007). Another case-control study in Norway showed that by supplying cod liver oil for 1 year old, significantly reduced incidence of childhood-onset type 1 diabetes is observed (Stene and Joner, 2003). Besides that, n-3 PU-FAs can improve glucose metabolism through enhanced insulin secretion and more resistance to cytokine-induced β cell death (Wei *et al.*, 2010), as well as reshaped Th1 and Th2 balance through modulation of T cell differentiation. Studies showed that the oxidation of n-3 PUFAs is

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partially responsible for the adverse effect observed in human and animal models (Albert *et al.*, 2016; Grimm *et al.*, 2016; Mason and Sherratt, 2017). And antioxidants, such as vitamin E, have been used along with n-3 PUFAs to overcome the oxidation-related side effects (Meydani, 1996).

4 Potential Targets and Mechanism of n-3 PUFAs in Anti-Inflammation and Immune Regulation

Potential targets and molecular mechanisms of n-3 PUFAs in anti-inflammation and immune regulation are described in the following sections.

4.1 Influencing the Metabolism of Arachidonic Acid

PUFAs are important components of the phospholipids in cell membranes. The most common PUFA in phospholipid membrane of immunocytes is arachidonic acid (AA, n-6 PUFA) (Calder, 2013). When inflammation occurs, AA will be converted into eicosanoids, including prostaglandins (PGs) and leukotrienes (LTs), through the respective cyclooxygenase (COX), isomerase and 5-lipooxygenase pathway. The eicosanoid PGE2 is a COX metabolite of AA, which can be pro-inflammatory and modulates cytokine production. The 4-series LTs, lipoxygenase metabolites of AA, with chemotactic ability to promote leucocytes move towards a site of inflammation and upregulate pro-inflammatory cytokine production (Sperling et al., 1993a). The n-3 PUFAs (DHA and EPA) can competitively inhibit oxygenation of AA due to EPA is also a substrate for both COX-2 and 5-lipoxygenase. However, the metabolites of EPA have a very different structure from those made from AA (James et al., 2000b). Recent studies have discovered a novel series of lipid mediators from n-3 PUFAs: resolvins and protectins

(Bannenberg *et al.*, 2005; Serhan *et al.*, 2004). The biological function of resolvins and protectins have been examined in a number of models and have shown antiinflammatory effects. For example, resolvin E1 and protectin D1 both inhibit transendothelial migration of neutrophils, therefore interrupting the infiltration of neutrophils into sites of inflammation (Schwab *et al.*, 2007; Serhan *et al.*, 2008).

4.2 Remodeling of Lipid Membrane

As indicated above, AA is a key structural and functional component of lipid membrane in cells, especially in immunocytes, such as macrophages, neutrophils, and lymphocytes (Calder, 2013). It is reported that fatty acid composition of inflammatory cells and immunocytes is sensitive to dietary fatty acids (Pollock *et al.*, 2016; Yaqoob *et al.*, 2000a). Further studies show that more consumption of n-3 PUFAs will change fatty acid profiles of lipid membrane in cells (Chapkin *et al.*, 1991; Sperling *et al.*, 1993b). AA will be displaced by n-3 PUFA at plasma membranes and lead to a suppressed generation of inflammatory precursors (Wang and Huang, 2015; Yaqoob *et al.*, 2000b).

On the other hand, n-3 PUFAs can alter lipid rafts, which are nanoscale regions of the plasma membrane, enriched in cholesterol, sphingomyelin, and phospholipids (Turk and Chapkin, 2013). Studies show that n-3 PUFAs are inserted into both raft and non-raft regions of the plasma membrane, and result in enhanced clusters of lipid raft region (Fig.4). Then, the cell signaling transduction will be influenced, for example, DHA can suppress ERK1/2 and STAT3 in regulating inflammation both *in vitro* and *in vivo* (Turk *et al.*, 2012). And the lipid raft dependent phosphorylation of PLC γ -1 in T cell can be regulated by n-3 PUFAs as well (Fan *et al.*, 2004).

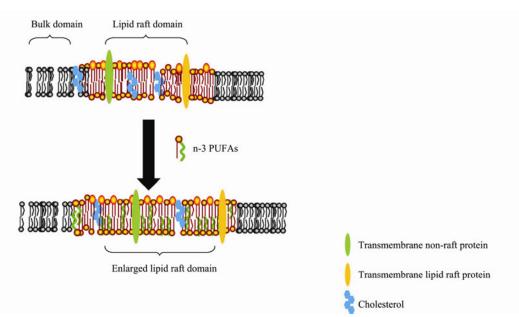


Fig.4 Putative model for the effect of n-3 PUFA on lipid rafts. Lipid rafts are cholesterol and phospholipid-enriched nanoscale region. Upon treatment with n-3 PUFAs, they are incorporated into phospholipids and induce enlarged clustering of lipid raft domains.

4.3 Eicosanoid-Independent Mechanisms

There is increasing insight that eicosanoid-independent mechanisms play an important role of n-3 PUFAs in regulating inflammation and immune response. In other words, n-3 PUFAs can directly regulate signaling pathways, modulate transcription factor activity, and act on gene expression (Calder, 2003; Simopoulos, 2002). For instance, n-3 PUFAs can inhibit MAPK activation in PKC-dependent and independent ways, therefore interfere in MAPK signaling transduction (Denys et al., 2001a). Additionally, alteration of transcription factor activity and gene expression level regulated by n-3 PUFAs has been described for PPAR, LXR, Nf-kB, AP-1, CREB and SREBP (Jia et al., 2006; Jump, 2004; Ntambi and Bene, 2001; Shi and Pestka, 2006). Chen et al found that DHA could induce S-phase cell cycle arrest and inhibit cell proliferation. After treating cells with DHA, the DNA replication site appears far away from upstream of ori- β , which indicates that n-3 PUFAs could influence DNA synthesis (Chen and Istfan, 2001).

4.4 Targeting at Mediators of Inflammation and Immune Response

Last but not the least, n-3 PUFAs have impact on mediators of inflammation and immune response, such as lymphocytes, NK cells, macrophages, T cells, platelet activating factor (PAF), and so on (Lu *et al.*, 2008; Monk *et al.*, 2013; Rombaldova *et al.*, 2017).

GPR120 is a G protein-coupled receptor for n-3 PUFAs in macrophages, and is also known as free fatty acid re-

ceptor 4 (FF4) (Moniri, 2016). The ligand specificity of GPR4 is much higher for PUFAs, such as DHA and EPA, than saturated long-chain fatty acids. However, it is hard to distinguish different PUFAs, for example, linoleic acid (n-6 PUFA) and palmitoleic acid (n-7 PUFA) almost have similar binding affinity with GPR120 (Galindo et al., 2012). Two isoforms of GPR120 have been identified as a long isoform and a short isoform. The long isoform only responds to β -arrestin-induced signaling, but the short isoform can trigger both G protein-mediated Ca2+ signaling and β-arrestin-mediated phosphorylation and internalization (Watson et al., 2012). High GPR120 expression is found in bone marrow-derived macrophages and in primary intraperitoneal macrophages (Oh et al., 2010). The mechanism of n-3 PUFAs in regulating GPR120 signaling has been elucidated that β -arrestin-2 is recruited after GPR120 activation, and the subsequent internalization of the GPR120/\beta-arrestin-2 complex leads to its binding with TAB1, which suppresses TAK1 activation and causes Nf-kB pathway blockade (Oh et al., 2010). X. Li et al. studied the effect of n-3 PUFAs on LPS-induced inflammation in macrophage and found that n-3 PUFAs could reduce COX-2 expression and decrease PGE2 synthesis in a concentration- and time-dependent manner, besides that TLR4 signaling through phosphorylation of Akt/JNK and Nf-kB p65 is repressed by n-3 PUFAs activated GPR120 too (Li et al., 2013). Liu et al. raised another potential mechanism that n-3 PUFAs activate cytosolic phospholipase A2 (cPLA2), which inhibits the NFκB signaling pathway via EP4 receptor and leads to the inhibition of LPS-induced IL-6 secretion in macrophages (Liu et al., 2014).

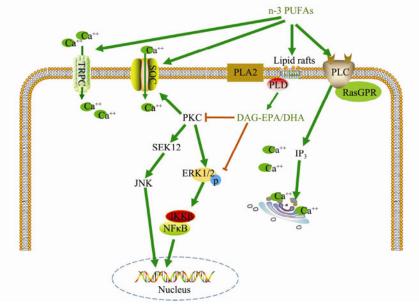


Fig.5 Schematic representation of the regulation of n-3 PUFA in T-cell signaling. N-3 PUFAs activated by PLD will increase the DAG containing n-3 PUFA (DAG-DHA/EPA) which, in turn, will upregulate $[Ca^{2+}]_i$ by TRPC channels, modulate RasGRP, and act on different isoforms of PKC. All of these effects of n-3 PUFAs will inhibit inflammatory gene transcription and lead to immunosuppression.

T cell is another immunocyte highly related to inflammation, especially Th1, Th2, Th17 and Treg subgroups. T cells play a central role in the immune system because they are involved in both innate immunity and adaptive immunity (Bedoui *et al.*, 2016; Eisenbarth and Flavell, 2009). In the process of host defends against pathogens, T

cells mediate the occurrence and development of inflammation. For example, in rheumatoid arthritis (RA) patients HIF-1 transcription is induced by Th1 cytokines TNF α and IL-1 β , which can lead to angiogenesis and inflammation in hypoxia (Larsen *et al.*, 2012). Recently, Th17 and Treg identified as two new subsets of T cells are tightly associated with autoimmune and inflammatory diseases. It is reported that Th17 cells represent a proinflammatory subgroup; however, Treg cells have a suppressive effect (Noack and Miossec, 2014).

Some studies have showed that n-3 PUFAs can inhibit T cell proliferation in vitro (Calder et al., 1991; Calder et al., 1992; Camps et al., 1992). The animals treated with n-3 PUFAs also show a down-regulated T cell proliferation (Fowler et al., 1993; Peterson et al., 1998), suppressive cytokine production (Jolly et al., 1997), and modified Th1 and Th2 subsets (Wallace et al., 2001). The underlying mechanisms of n-3 PUFAs-regulated T cell activity have been interpreted as reduced generation of diacylglycerol (Fowler et al., 1993), inhibition of PKC activation (Denys et al., 2005), and repressed ERK/MAPK signaling transduction (Denys et al., 2001b). Calcium signaling as the second messenger also is the target of n-3 PUFAs in regulating T cell function (Yog et al., 2010). Chow and Jondal found that n-3 PUFAs could increase $[Ca^{2+}]_i$ in Jurkat T-cells (Chow and Jondal, 1990), and a similar effect of n-3 PUFAs on monocytes is observed too (Aires et al., 2007). The upregulation of $[Ca^{2+}]_i$ is due to the activation of calcium release-activated calcium (CRAC) channels after treated with n-3 PUFAs, and a stimulated production of diacylglycerol (DAG)-n- 3PUFAs by activating phospholipase D (PLD) can be observed. Moreover, n-3 PUFAs seem to inhibit PKC activation via diminished ERK1/2 phosphorylation (Denys et al., 2004). Furthermore, immunologic synapse involved in APC presenting antigens to T cells is dependent on Ca²⁺ mediated mitochondrial redistribution (Quintana et al., 2007). Studies have shown that when T cells are treated with n-3 PUFAs the formation of immunologic synapse will be inhibited, which is associated with the change of $[Ca^{2+}]_i$ and is consistent with immune suppression caused by n-3 PUFAs in autoimmune disease (Yog et al., 2010). Based on current studies, multiple signaling pathways are interlinked to each other as the targets of n-3 PUFAs in regulating T cell function (Fig.5).

To briefly summarize the discovered mechanisms of n-3 PUFAs in immune regulation, firstly, n-3 PUFAs can influence the composition of lipid membrane and change lipid raft regions, then the phosphorylation of PKC which is an integral part of lipid microdomains will be down-regulated (Akhtar, 2010); secondly, metabolites of inflammatory precursor are transformed into anti-inflammatory agents; thirdly, n-3 PUFAs will interfere in down-stream cell signaling cascade, including PKC, MAPK, Nf- κ B, $[Ca^{2+}]_i$ and so on.

5 Perspectives

Since the first publication by Dyerberg, many studies

have been carried out to understand physiological function and health care effect of n-3 PUFAs. The epidemiology, clinical and experimental researches in recent years have shown that n-3 PUFAs are essential fatty acids for human health (Brantsaeter et al., 2017; Marushka et al., 2017). It has been well accepted that n-3 PUFAs as dietary supplementation and animal feed should be used with care because they are easily oxidized (Ballou and De-Peters, 2008). However, whether n-3 PUFAs can be used as drug in clinics is still a controversial issue (Calder, 2013; Fialkow, 2016b). Although the negative results are reported occasionally, the overall impact of n-3 PUFAs on human health is still positive. In the future, more detailed research, including sex, age, ethnicity, trial duration and health conditions, may be able to answer the different roles of n-3 PUFAs being played in human. There is no doubt that more in-depth studies of n-3 PUFAs are called for to meet the goal of improving human health and preventing inflammatory diseases.

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References

- Abbott, K. A., 2016. Do ω-3 PUFAs affect insulin resistance in a sex-specific manner? A systematic review and meta-analysis of randomized controlled trials. *American Journal of Clinical Nutrition*, **104**: 1470-1484.
- Aires, V., Hichami, A., Filomenko, R., Ple, A., Rebe, C., Bettaieb, A., and Khan, N. A., 2007. Docosahexaenoic acid induces increases in [Ca²⁺]_i via inositol 1,4,5-triphosphate production and activates protein kinase C gamma and -delta via phosphatidylserine binding site: Implication in apoptosis in U937 cells. *Molecular Pharmacology*, **72**: 1545-1556.
- Akhtar, K. N., 2010. Polyunsaturated fatty acids in the modulation of T-cell signalling. *Prostaglandins Leukotrienes Essential Fatty Acids*, 82: 179-187.
- Albert, B. B., Vickers, M. H., Gray, C., Reynolds, C. M., Segovia, S. A., Derraik, J. G., Lewandowski, P. A., Garg, M. L., Cameron-Smith, D., Hofman, P. L., and Cutfield, W. S., 2016. Oxidized fish oil in rat pregnancy causes high newborn mortality and increases maternal insulin resistance. *American Journal of Physiology-Regulatory Integrative and Comparative Physiology*, **311**: R497-504.
- Alexander, D. D., Miller, P. E., Van Elswyk, M. E., Kuratko, C. N., and Bylsma, L. C., 2017. A meta-analysis of randomized controlled trials and prospective cohort studies of eicosapentaenoic and docosahexaenoic long-chain omega-3 fatty acids and coronary heart disease risk. *Mayo Clinic Proceedings*, 92: 15-29.
- Arbabi, S., and Maier, R. V., 2002. Mitogen-activated protein kinases. *Critical Care Medicine*, 30: S74-S79.
- Arnoldussen, I. A., and Kiliaan, A. J., 2014. Impact of DHA on metabolic diseases from womb to tomb. *Marine Drugs*, 12: 6190-6212.
- Auger, C., Said, A., Nguyen, P. N., Chabert, P., Idris-Khodja,

N., and Schini-Kerth, V. B., 2016. Potential of food and natural products to promote endothelial and vascular health. *Journal of Cardiovascular Pharmacology*, **68**: 11-18.

- Ballou, M. A., and DePeters, E. J., 2008. Supplementing milk replacer with omega-3 fatty acids from fish oil on immunocompetence and health of Jersey calves. *Journal of Dairy Science*, 91: 3488-3500.
- Bandini, M., Gandaglia, G., and Briganti, A., 2017. Obesity and prostate cancer. *Current Opinion in Urology*, 27 (5): 415-421.
- Bannenberg, G. L., Chiang, N., Ariel, A., Arita, M., Tjonahen, E., Gotlinger, K. H., Hong, S., and Serhan, C. N., 2005. Molecular circuits of resolution: Formation and actions of resolvins and protectins. *Journal of Immunology*, **174**: 4345-4355.
- Bedoui, S., Heath, W. R., and Mueller, S. N., 2016. CD4(+) T-cell help amplifies innate signals for primary CD8(+) T-cell immunity. *Immunological Reviews*, **272**: 52-64.
- Boothby, M., and Rickert, R. C., 2017. Metabolic regulation of the immune humoral response. *Immunity*, **46**: 743-755.
- Bottazzo, G. F., Dean, B. M., McNally, J. M., MacKay, E. H., Swift, P. G., and Gamble, D. R., 1985. *In situ* characterization of autoimmune phenomena and expression of HLA molecules in the pancreas in diabetic insulitis. *New England Journal of Medicine*, 313: 353-360.
- Brantsaeter, A. L., Englund-Ogge, L., Haugen, M., Birgisdottir, B. E., Knutsen, H. K., Sengpiel, V., Myhre, R., Alexander, J., Nilsen, R. M., Jacobsson, B., and Meltzer, H. M., 2017. Maternal intake of seafood and supplementary long chain n-3 poly-unsaturated fatty acids and preterm delivery. *BMC Pregnancy and Childbirth*, 17: 41.
- Buettner, R., Parhofer, K. G., Woenckhaus, M., Wrede, C. E., Kunz-Schughart, L. A., Scholmerich, J., and Bollheimer, L. C., 2006. Defining high-fat-diet rat models: Metabolic and molecular effects of different fat types. *Journal of Molecular Endocrinology*, **36**: 485-501.
- Calder, P. C., 2003. N-3 polyunsaturated fatty acids and inflammation: From molecular biology to the clinic. *Lipids*, **38**: 343-352.
- Calder, P. C., 2013. Omega-3 polyunsaturated fatty acids and inflammatory processes: Nutrition or pharmacology? *British Journal of Clinical Pharmacology*, **75**: 645-662.
- Calder, P. C., Bond, J. A., Bevan, S. J., Hunt, S. V., and Newsholme, E. A., 1991. Effect of fatty acids on the proliferation of concanavalin A-stimulated rat lymph node lymphocytes. *International Journal of Biochemistry*, 23: 579-588.
- Calder, P. C., Bevan, S. J., and Newsholme, E. A., 1992. The inhibition of T-lymphocyte proliferation by fatty acids is *via* an eicosanoid-independent mechanism. *Immunology*, 75: 108-115.
- Calviello, G., Serini, S., Piccioni, E., and Pessina, G., 2009. Antineoplastic effects of n-3 polyunsaturated fatty acids in combination with drugs and radiotherapy: Preventive and therapeutic strategies. *Nutrition and Cancer*, 61: 287-301.
- Camps, J., Bargallo, T., Gimenez, A., Alie, S., Caballeria, J., Pares, A., Joven, J., Masana, L., and Rodes, J., 1992. Relationship between hepatic lipid peroxidation and fibrogenesis in carbon tetrachloride-treated rats: Effect of zinc administration. *Clinical Science (London)*, 83: 695-700.
- Carreras-Torres, R., Johansson, M., Haycock, P. C., Wade, K. H., Relton, C. L., Martin, R. M., Davey, S. G., Albanes, D., Aldrich, M. C., Andrew, A., Arnold, S. M., Bickeboller, H., Bojesen, S. E., Brunnstrom, H., Manjer, J., Bruske, I., Caporaso, N. E., Chen, C., Christiani, D. C., Christian, W. J., Doherty, J. A., Duell, E. J., Field, J. K., Davies, M., Marcus, M.

W., Goodman, G. E., Grankvist, K., Haugen, A., Hong, Y. C., Kiemeney, L. A., van der Heijden, E., Kraft, P., Johansson, M. B., Lam, S., Landi, M. T., Lazarus, P., Le Marchand, L., Liu, G., Melander, O., Park, S. L., Rennert, G., Risch, A., Haura, E. B., Scelo, G., Zaridze, D., Mukeriya, A., Savic, M., Lissowska, J., Swiatkowska, B., Janout, V., Holcatova, I., Mates, D., Schabath, M. B., Shen, H., Tardon, A., Teare, M. D., Woll, P., Tsao, M. S., Wu, X., Yuan, J. M., Hung, R. J., Amos, C. I., McKay, J., and Brennan, P., 2017. Obesity, metabolic factors and risk of different histological types of lung cancer: A Mendelian randomization study. *PLoS One*, 12: e0177875.

- Chapkin, R. S., Akoh, C. C., and Miller, C. C., 1991. Influence of dietary n-3 fatty acids on macrophage glycerophospholipid molecular species and peptidoleukotriene synthesis. *Journal* of Lipid Research, **32**: 1205-1213.
- Chen, C., Yang, Y., Yu, X., Hu, S., and Shao, S., 2017. Association between omega-3 fatty acids consumption and the risk of type 2 diabetes: A meta-analysis of cohort studies. *Journal* of Diabetes Investigation, 8: 480-488.
- Chen, L. H., Wang, Y. F., Xu, Q. H., and Chen, S. S., 2016. Omega-3 fatty acids as a treatment for non-alcoholic fatty liver disease in children: A systematic review and metaanalysis of randomized controlled trials. *Clinical Nutrition*, DOI: 10.1016/j.clnu.2016.12.009.
- Chen, Z. Y., and Istfan, N. W., 2001. Docosahexaenoic acid, a major constituent of fish oil diets, prevents activation of cyclin-dependent kinases and S-phase entry by serum stimulation in HT-29 cells. *Prostaglandins Leukotrienes Essential Fatty Acids*, 64: 67-73.
- Chiesa, G., Busnelli, M., Manzini, S., and Parolini, C., 2016. Nutraceuticals and bioactive components from fish for dyslipidemia and cardiovascular risk reduction. *Marine Drugs*, DOI: 10.3390/md14060113.
- Chow, S. C., and Jondal, M., 1990. Polyunsaturated free fatty acids stimulate an increase in cytosolic Ca²⁺ by mobilizing the inositol 1,4,5-trisphosphate-sensitive Ca²⁺ pool in T cells through a mechanism independent of phosphoinositide turn-over. *Journal of Biological Chemistry*, **265**: 902-907.
- Cladis, D. P., Kleiner, A. C., Freiser, H. H., and Santerre, C. R., 2014. Fatty acid profiles of commercially available finfish fillets in the United States. *Lipids*, **49**: 1005-1018.
- Davis, R. J., 2000, Signal transduction by the JNK group of MAP kinases. *Cell*, **103**: 239-252.
- Dawczynski, C., Massey, K. A., Ness, C., Kiehntopf, M., Stepanow, S., Platzer, M., Grun, M., Nicolaou, A., and Jahreis, G., 2013. Randomized placebo-controlled intervention with n-3 LC-PUFA-supplemented yoghurt: Effects on circulating eicosanoids and cardiovascular risk factors. *Clinical Nutrition*, **32**: 686-696.
- de Castro, G. S., and Calder, P. C., 2017. Non-alcoholic fatty liver disease and its treatment with n-3 polyunsaturated fatty acids. *Clinical Nutrition*, Epub ahead.
- Delarue, J., LeFoll, C., Corporeau, C., and Lucas, D., 2004. N-3 long chain polyunsaturated fatty acids: A nutritional tool to prevent insulin resistance associated to type 2 diabetes and obesity? *Reproduction Nutrition Development*, 44: 289-299.
- Dell'Albani, P., Santangelo, R., Torrisi, L., Nicoletti, V. G., de Vellis, J., and Giuffrida, S. A., 2001. JAK/STAT signaling pathway mediates cytokine-induced iNOS expression in primary astroglial cell cultures. *Journal of Neuroscience Research*, 65: 417-424.
- Denys, A., Hichami, A., and Khan, N. A., 2001. Eicosapentaenoic acid and docosahexaenoic acid modulate MAP kinase

(ERK1/ERK2) signaling in human T cells. *Journal of Lipid Research*, **42**: 2015-2020.

- Denys, A., Hichami, A., and Khan, N. A., 2005. n-3 PUFAs modulate T-cell activation via protein kinase C-alpha and -epsilon and the NF-kappaB signaling pathway. *Journal of Lipid Research*, 46: 752-758.
- Denys, A., Aires, V., Hichami, A., and Khan, N. A., 2004. Thapsigargin-stimulated MAP kinase phosphorylation via CRAC channels and PLD activation: Inhibitory action of docosahexaenoic acid. FEBS Letters, 564: 177-182.
- Divella, R., De Luca, R., Abbate, I., Naglieri, E., and Daniele, A., 2016. Obesity and cancer: The role of adipose tissue and adipo-cytokines-induced chronic inflammation. *Journal of Cancer*, 7: 2346-2359.
- Djousse, L., Gaziano, J. M., Buring, J. E., and Lee, I. M., 2010. Dietary omega-3 fatty acids and fish consumption and risk of type 2 diabetes. *American Journal of Clinical Nutrition*, 93: 143-150.
- Dobbins, M., Decorby, K., and Choi, B. C. K., 2013. The association between obesity and cancer risk: A meta-analysis of observational studies from 1985 to 2011. *ISRN Preventive Medicine*, 2013: 680536.
- Domenick, M. A., and Ildstad, S. T., 2001. Impact of bone marrow transplantation on type I diabetes. *World Journal of Sur*gery, 25: 474-480.
- Dyerberg, J., Bang, H. O., Stoffersen, E., Moncada, S., and Vane, J. R., 1978. Eicosapentaenoic acid and prevention of thrombosis and atherosclerosis? *Lancet*, 2: 117-119.
- Dyerberg, J., and Bang, H. O., 1978. Dietary fat and thrombosis. *Lancet*, 1: 152.
- Edwards, I. J., and O'Flaherty, J. T., 2008. Omega-3 fatty acids and PPARgamma in cancer. *PPAR Research*, **2008**: 358052.
- Eisenbarth, G. S., Nayak, R. C., and Rabinowe, S. L., 1988, Type I diabetes as a chronic autoimmune disease. *Journal of Diabet Complications*, 2: 54-58.
- Eisenbarth, S. C., and Flavell, R. A., 2009, Innate instruction of adaptive immunity revisited: The inflammasome. *EMBO Molecular Medicine*, 1: 92-98.
- Emmungil, H., Erdogan, M., Kalfa, M., Karabulut, G., Kocanaogullari, H., Inal, V., Aksu, K., Oksel, F., Kabasakal, Y., and Keser, G., 2014. Autoimmune thyroid disease in ankylosing spondylitis. *Clinical Rheumatology*, **33**: 955-961.
- Fan, Y. Y., Ly, L. H., Barhoumi, R., McMurray, D. N., and Chapkin, R. S., 2004. Dietary docosahexaenoic acid suppresses T cell protein kinase C theta lipid raft recruitment and IL-2 production. *Journal of Immunology*, **173**: 6151-6160.
- Fasano, E., Serini, S., Piccioni, E., Innocenti, I., and Calviello, G., 2010. Chemoprevention of lung pathologies by dietary n-3 polyunsaturated fatty acids. *Current Medicinal Chemistry*, 17: 3358-3376.
- Ferrero-Miliani, L., Nielsen, O. H., Andersen, P. S., and Girardin, S. E., 2007. Chronic inflammation: Importance of NOD2 and NALP3 in interleukin-1beta generation. *Clinical* and Experimental Immunology, 147: 227-235.
- Fialkow, J., 2016. Omega-3 fatty acid formulations in cardiovascular disease: Dietary supplements are not substitutes for prescription products. *American Journal of Cardiovascular Drugs*, 16: 229-239.
- Flachs, P., Rossmeisl, M., Bryhn, M., and Kopecky, J., 2009. Cellular and molecular effects of n-3 polyunsaturated fatty acids on adipose tissue biology and metabolism. *Clinical Science (London)*, **116**: 1-16.
- Fowler, K. H., McMurray, D. N., Fan, Y. Y., Aukema, H. M., and Chapkin, R. S., 1993. Purified dietary n-3 polyunsatu-

rated fatty acids alter diacylglycerol mass and molecular species composition in concanavalin A-stimulated murine splenocytes. *Biochimica et Biophysica Acta*, **1210**: 89-96.

- Galindo, M. M., Voigt, N., Stein, J., van Lengerich, J., Raguse, J. D., Hofmann, T., Meyerhof, W., and Behrens, M., 2012. G protein-coupled receptors in human fat taste perception. *Chemical Senses*, 37: 123-139.
- Giacomelli, R., Afeltra, A., Alunno, A., Baldini, C., Bartoloni Bocci, E., Berardicurti, O., Carubbi, F., Cauli, A., Cervera, R., Ciccia, F., Cipriani, P., Conti, F., De Vita, S., Di Benedetto, P., Doria, A., Drosos, A. A., Favalli, E. G., Gandolfo, S., Gatto, M., Grembiale, R. D., Liakouli, V., Lories, R., Lubrano, E., Lunardi, C., Margiotta, D., Massaro, L., Meroni, P., Minniti, A., Navarini, L., Pendolino, M., Perosa, F., Pers, J. O., Prete, M., Priori, R., Puppo, F., Quartuccio, L., Ruffatti, A., Ruscitti, P., Russo, B., Sarzi-Puttini, P., Shoenfeld, Y., Somarakis, G. A., Spinelli, F. R., Tinazzi, E., Triolo, G., Ursini, F., Valentini, G., Valesini, G., Vettori, S., Vitali, C., and Tzioufas, A. G., 2017. International consensus: What else can we do to improve diagnosis and therapeutic strategies in patients affected by autoimmune rheumatic diseases (rheumatoid arthritis, spondyloarthritides, systemic sclerosis, systemic lupus erythematosus, antiphospholipid syndrome and Sjogren's syndrome)?: The unmet needs and the clinical grey zone in autoimmune disease management. Autoimmunity Reviews, DOI: 10.1016/j.autrev.2017.07.012.
- Grimm, M. O., Haupenthal, V. J., Mett, J., Stahlmann, C. P., Blumel, T., Mylonas, N. T., Endres, K., Grimm, H. S., and Hartmann, T., 2016. Oxidized docosahexaenoic acid species and lipid peroxidation products increase amyloidogenic amyloid precursor protein processing. *Neurodegenerative Diseases*, 16: 44-54.
- Hidayat, K., Du, X., Chen, G., Shi, M., and Shi, B., 2016. Abdominal obesity and lung cancer risk: Systematic review and meta-analysis of prospective studies. *Nutrients*, DOI: 10. 1042/BSR20160474.
- Holman, R. T., Johnson, S. B., Bibus, D., Spencer, D. C., and Donadio, J. J., 1994. Essential fatty acid deficiency profiles in idiopathic immunoglobulin A nephropathy. *American Journal* of Kidney Diseases, 23: 648-654.
- Hooper, L., 2007. Primary prevention of CVD: Diet and weight loss. *BMJ Clinical Evidence*, **10** (1): 0219.
- Huang, C. W., Chien, Y. S., Chen, Y. J., Ajuwon, K. M., Mersmann, H. M., and Ding, S. T., 2016. Role of n-3 Polyunsaturated fatty acids in ameliorating the obesity-induced metabolic syndrome in animal models and humans. *International Journal of Molecular Sciences*, DOI: 10.3390/ijms17101689.
- Hyslop, C. M., Tsai, S., Shrivastava, V., Santamaria, P., and Huang, C., 2016. Prolactin as an adjunct for type 1 diabetes immunotherapy. *Endocrinology*, **157**: 150-165.
- Jacobo-Cejudo, M. G., Valdes-Ramos, R., Guadarrama-Lopez, A. L., Pardo-Morales, R. V., Martinez-Carrillo, B. E., and Harbige, L. S., 2017. Effect of n-3 polyunsaturated fatty acid supplementation on metabolic and inflammatory biomarkers in type 2 diabetes mellitus patients. *Nutrients*, DOI: 10.3390/ nu9060573.
- James, M. J., Gibson, R. A., and Cleland, L. G., 2000. Dietary polyunsaturated fatty acids and inflammatory mediator production. *American Journal of Clinical Nutrition*, 71: 343S-348S.
- Jia, Q., Zhou, H. R., Shi, Y., and Pestka, J. J., 2006. Docosahexaenoic acid consumption inhibits deoxynivalenol-induced CREB/ATF1 activation and IL-6 gene transcription in mouse macrophages. *Journal of Nutrition*, **136**: 366-372.

- Jiang, Y., Gram, H., Zhao, M., New, L., Gu, J., Feng, L., Padova, F. D., Ulevitch, R. J., and Han, J., 1997. Characterization of the structure and function of the fourth member of p38 group mitogen-activated protein kinases, p38delta. *Journal of Biological Chemistry*, **272**: 30122-30128.
- Jin, H. S., Suh, H. W., Kim, S. J., and Jo, E. K., 2017. Mitochondrial control of innate immunity and inflammation. *Immune Network*, 17: 77-88.
- Jolly, C. A., Jiang, Y. H., Chapkin, R. S., and McMurray, D. N., 1997. Dietary (n-3) polyunsaturated fatty acids suppress murine lymphoproliferation, interleukin-2 secretion, and the formation of diacylglycerol and ceramide. *Journal of Nutrition*, 127: 37-43.
- Jump, D. B., 2004. Fatty acid regulation of gene transcription. *Critical Reviews in Clinical Laboratory Sciences*, 41: 41-78.
- Kamesaki, T., 2017. Progress in diagnosis and treatment of autoimmune hemolytic disorders. *Rinsho Ketsueki*, 58: 329-335.
- Kelley, N. S., 2016. Treatment of nonalcoholic fatty liver disease with long-chain n-3 polyunsaturated fatty acids in humans. *Metabolic Syndrome and Related Disorders*, 14: 417-430.
- Kris-Etherton, P. M., Harris, W. S., and Appel, L. J., 2002. Fish consumption, fish oil, omega-3 fatty acids, and cardiovascular disease. *Circulation*, **106**: 2747-2757.
- Larsen, H., Muz, B., Khong, T. L., Feldmann, M., and Paleolog, E. M., 2012. Differential effects of Th1 *versus* Th2 cytokines in combination with hypoxia on HIFs and angiogenesis in RA. *Arthritis Research and Therapy*, 14: R180.
- Lee, T. H., Hoover, R. L., Williams, J. D., Sperling, R. I., Ravalese, J. R., Spur, B. W., Robinson, D. R., Corey, E. J., Lewis, R. A., and Austen, K. F., 1985. Effect of dietary enrichment with eicosapentaenoic and docosahexaenoic acids on *in vitro* neutrophil and monocyte leukotriene generation and neutrophil function. *New England Journal of Medicine*, 312: 1217-1224.
- Li, J. J., Huang, C. J., and Xie, D., 2008. Anti-obesity effects of conjugated linoleic acid, docosahexaenoic acid, and eicosapentaenoic acid. *Molecular Nutrition and Food Research*, 52: 631-645.
- Li, X., Yu, Y., and Funk, C. D., 2013. Cyclooxygenase-2 induction in macrophages is modulated by docosahexaenoic acid via interactions with free fatty acid receptor 4 (FFA4). FASEB Journal, 27: 4987-4997.
- Liu, Y., Chen, L. Y., Sokolowska, M., Eberlein, M., Alsaaty, S., Martinez-Anton, A., Logun, C., Qi, H. Y., and Shelhamer, J. H., 2014. The fish oil ingredient, docosahexaenoic acid, activates cytosolic phospholipase A(2) via GPR120 receptor to produce prostaglandin E(2) and plays an anti-inflammatory role in macrophages. *Immunology*, **143**: 81-95.
- Lokesh, B. R., and Kinsella, J. E., 1987. Modulation of prostaglandin synthesis in mouse peritoneal macrophages by enrichment of lipids with either eicosapentaenoic or docosahexaenoic acids in vitro. *Immunobiology*, **175**: 406-419.
- Lu, J., Caplan, M. S., Li, D., and Jilling, T., 2008. Polyunsaturated fatty acids block platelet-activating factor-induced phosphatidylinositol 3 kinase/Akt-mediated apoptosis in intestinal epithelial cells. *American Journal of Physiology Gastrointestinal and Liver Physiology*, **294**: 1181-1190.
- Luchtman, D. W., and Song, C., 2013. Cognitive enhancement by omega-3 fatty acids from child-hood to old age: Findings from animal and clinical studies. *Neuropharmacology*, 64: 550-565.

Marszalek, J. R., and Lodish, H. F., 2005. Docosahexaenoic

acid, fatty acid-interacting proteins, and neuronal function: Breastmilk and fish are good for you. *Annual Review of Cell and Developmental Biology*, **21**: 633-657.

- Marushka, L., Batal, M., David, W., Schwartz, H., Ing, A., Fediuk, K., Sharp, D., Black, A., Tikhonov, C., and Chan, H. M., 2017. Association between fish consumption, dietary omega-3 fatty acids and persistent organic pollutants intake, and type 2 diabetes in 18 First Nations in Ontario. *Environmental Research*, **156**: 725-737.
- Mason, R. P., and Sherratt, S. C., 2017. Omega-3 fatty acid fish oil dietary supplements contain saturated fats and oxidized lipids that may interfere with their intended biological benefits. *Biochemical and Biophysical Research Communications*, 483: 425-429.
- Meydani, S. N., 1996. Effect of (n-3) polyunsaturated fatty acids on cytokine production and their biologic function. *Nutrition*, **12**: S8-S14.
- Moniri, N. H., 2016. Free-fatty acid receptor-4 (GPR120): Cellular and molecular function and its role in metabolic disorders. *Biochemical Pharmacology*, **110-111**: 1-15.
- Monk, J. M., Hou, T. Y., Turk, H. F., McMurray, D. N., and-Chapkin, R. S., 2013. n3 PUFAs reduce mouse CD4+ T-cell ex vivo polarization into Th17 cells. *Journal of Nutrition*, 143: 1501-1508.
- Muurling, M., Mensink, R. P., Pijl, H., Romijn, J. A., Havekes, L. M., and Voshol, P. J., 2003. A fish oil diet does not reverse insulin resistance despite decreased adipose tissue TNF-alpha protein concentration in ApoE-3*Leiden mice. *Journal of Nutrition*, 133: 3350-3355.
- Noack, M., and Miossec, P., 2014. Th17 and regulatory T cell balance in autoimmune and inflammatory diseases. *Autoimmunity Reviews*, **13**: 668-677.
- Norris, J. M., Yin, X., Lamb, M. M., Barriga, K., Seifert, J., Hoffman, M., Orton, H. D., Baron, A. E., Clare-Salzler, M., Chase, H. P., Szabo, N. J., Erlich, H., Eisenbarth, G. S., and Rewers, M., 2007. Omega-3 polyunsaturated fatty acid intake and islet autoimmunity in children at increased risk for type 1 diabetes. *JAMA*, **298**: 1420-1428.
- Nosaka, K., Miyoshi, T., Iwamoto, M., Kajiya, M., Okawa, K., Tsukuda, S., Yokohama, F., Sogo, M., Nishibe, T., Matsuo, N., Hirohata, S., Ito, H., and Doi, M., 2017. Early initiation of eicosapentaenoic acid and statin treatment is associated with better clinical outcomes than statin alone in patients with acute coronary syndromes: 1-year outcomes of a randomized controlled study. *International Journal of Cardiology*, 228: 173-179.
- Ntambi, J. M., and Bene, H., 2001. Polyunsaturated fatty acid regulation of gene expression. *Journal of Molecular Neuroscience*, 16: 273-278.
- O'Byrne, K. J., and Dalgleish, A. G., 2001. Chronic immune activation and inflammation as the cause of malignancy. *British Journal of Cancer*, **85**: 473-483.
- Oh, D. Y., Talukdar, S., Bae, E. J., Imamura, T., Morinaga, H., Fan, W., Li, P., Lu, W. J., Watkins, S. M., and Olefsky, J. M., 2010. GPR120 is an omega-3 fatty acid receptor mediating potent anti-inflammatory and insulin-sensitizing effects. *Cell*, 142: 687-698.
- Orban, T., Bundy, B., Becker, D. J., DiMeglio, L. A., Gitelman, S. E., Goland, R., Gottlieb, P. A., Greenbaum, C. J., Marks, J. B., Monzavi, R., Moran, A., Raskin, P., Rodriguez, H., Russell, W. E., Schatz, D., Wherrett, D., Wilson, D. M., Krischer, J. P., and Skyler, J. S., 2011. Co-stimulation modulation with abatacept in patients with recent-onset type 1 diabetes: A randomised, double-blind, placebo-controlled trial. *Lancet*, 378:

412-419.

- Perez-Echarri, N., Perez-Matute, P., Marcos-Gomez, B., Baena, M. J., Marti, A., Martinez, J. A., and Moreno-Aliaga, M. J., 2008. Differential inflammatory status in rats susceptible or resistant to diet-induced obesity: Effects of EPA ethyl ester treatment. *European Journal of Nutrition*, **47**: 380-386.
- Pestka, J. J., 2010. n-3 polyunsaturated fatty acids and autoimmune-mediated glomerulonephritis. *Prostaglandins Leukot Essent Fatty Acids*, 82: 251-258.
- Peterson, L. D., Thies, F., Sanderson, P., Newsholme, E. A., and Calder, P. C., 1998. Low levels of eicosapentaenoic and docosahexaenoic acids mimic the effects of fish oil upon rat lymphocytes. *Life Science*, 62: 2209-2217.
- Phang, M., Lazarus, S., Wood, L. G., and Garg, M., 2011. Diet and thrombosis risk: Nutrients for prevention of thrombotic disease. *Seminars in Thrombosis and Hemostasis*, 37: 199-208.
- Playfair, J. H. L., and Chain, B. M., 2013. *Immunology at a Glance: Chichester*. Wiley-Blackwell, West Sussex, 118pp.
- Pollock, A. H., Tedla, N., Hancock, S. E., Cornely, R., Mitchell, T. W., Yang, Z., Kockx, M., Parton, R. G., Rossy, J., and Gaus, K., 2016. Prolonged intake of dietary lipids alters membrane structure and T cell responses in LDLr-/-mice. *Journal of Immunology*, **196**: 3993-4002.
- Poreba, M., Mostowik, M., Siniarski, A., Golebiowska-Wiatrak, R., Malinowski, K. P., Haberka, M., Konduracka, E., Nessler, J., Undas, A., and Gajos, G., 2017. Treatment with high-dose n-3 PUFAs has no effect on platelet function, coagulation, metabolic status or inflammation in patients with atherosclerosis and type 2 diabetes. *Cardiovascular Diabetology*, DOI: 10.1186/s12933-017-0523-9.
- Quintana, A., Schwindling, C., Wenning, A. S., Becherer, U., Rettig, J., Schwarz, E. C., and Hoth, M., 2007. T cell activation requires mitochondrial translocation to the immunological synapse. *Proceedings of the National Academy of Sciences of the United States of America*, **104**: 14418-14423.
- Ralston, J. C., Matravadia, S., Gaudio, N., Holloway, G. P., and Mutch, D. M., 2015. Polyunsaturated fatty acid regulation of adipocyte FADS1 and FADS2 expression and function. *Obesity (Silver Spring)*, 23: 725-728.
- Rizos, E. C., and Elisaf, M. S., 2017. Does supplementation with omega-3 PUFAs add to the prevention of cardiovascular disease? *Current Cardiology Reports*, **19**: 47.
- Rombaldova, M., Janovska, P., Kopecky, J., and Kuda, O., 2017. Omega-3 fatty acids promote fatty acid utilization and production of pro-resolving lipid mediators in alternatively activated adipose tissue macrophages. *Biochemical and Biophysical Research Communications*, DOI: 10.1016/j.bbrc.2017.06. 170.
- Saw, C. L., Huang, Y., and Kong, A. N., 2010. Synergistic anti-inflammatory effects of low doses of curcumin in combination with polyunsaturated fatty acids: Docosahexaenoic acid or eicosapentaenoic acid. *Biochemical Pharmacology*, 79: 421-430.
- Schwab, J. M., Chiang, N., Arita, M., and Serhan, C. N., 2007. Resolvin E1 and protectin D1 activate inflammation-resolution programmes. *Nature*, 447: 869-874.
- Seger, R., and Krebs, E. G., 1995. The MAPK signaling cascade. FASEB Journal, 9: 726-735.
- Serhan, C. N., Gotlinger, K., Hong, S., and Arita, M., 2004. Resolvins, docosatrienes, and neuroprotectins, novel omega-3-derived mediators, and their aspirin-triggered endogenous epimers: An overview of their protective roles in catabasis. *Prostaglandins & Other Lipid Mediators*, **73**: 155-172.

- Serhan, C. N., Chiang, N., and Van Dyke, T. E., 2008. Resolving inflammation: Dual anti-inflammatory and pro-resolution lipid mediators. *Nature Review Immunology*, 8: 349-361.
- Shi, Y., and Pestka, J. J., 2006. Attenuation of mycotoxin-induced IgA nephropathy by eicosapentaenoic acid in the mouse: Dose response and relation to IL-6 expression. *Journal of Nutritional Biochemistry*, **17**: 697-706.
- Simopoulos, A. P., 2002. Omega-3 fatty acids in inflammation and autoimmune diseases. *Journal of the American College of Nutrition*, 21: 495-505.
- Siscovick, D. S., Barringer, T. A., Fretts, A. M., Wu, J. H., Lichtenstein, A. H., Costello, R. B., Kris-Etherton, P. M., Jacobson, T. A., Engler, M. B., Alger, H. M., Appel, L. J., and Mozaffarian, D., 2017. Omega-3 polyunsaturated fatty acid (fish oil) supplementation and the prevention of clinical cardiovascular disease: A science advisory from the american heart association. *Circulation*, **135**: e867-e884.
- So, W. W., Liu, W. N., and Leung, K. N., 2015. Omega-3 polyunsaturated fatty acids trigger cell cycle arrest and induce apoptosis in human neuroblastoma LA-N-1 cells. *Nutrients*, 7: 6956-6573.
- Spector, A. A., 1999. Essentiality of fatty acids. *Lipids*, **34**: S1-S3.
- Sperling, R. I., Benincaso, A. I., Knoell, C. T., Larkin, J. K., Austen, K. F., and Robinson, D. R., 1993. Dietary omega-3 polyunsaturated fatty acids inhibit phosphoinositide formation and chemotaxis in neutrophils. *Journal of Clinical Investigation*, **91**: 651-660.
- Stene, L. C., and Joner, G., 2003. Use of cod liver oil during the first year of life is associated with lower risk of childhoodonset type 1 diabetes: A large, population-based, case-control study. *The American Journal of Clinical Nutrition*, **78**: 1128-1134.
- Sun, J. W., Zhao, L. G., Yang, Y., Ma, X., Wang, Y. Y., and Xiang, Y. B., 2015. Obesity and risk of bladder cancer: A dose-response meta-analysis of 15 cohort studies. *PLoS One*, **10**: e0119313.
- Thaiss, C. A., Zmora, N., Levy, M., and Elinav, E., 2016. The microbiome and innate immunity. *Nature*, 535: 65-74.
- Trumbo, P., Schlicker, S., Yates, A. A., and Poos, M., 2002. Dietary reference intakes for energy, carbohydrate, fiber, fat, fatty acids, cholesterol, protein and amino acids. *Journal of the American Dietetic Association*, **102**: 1621-1630.
- Turk, H. F., Barhoumi, R., and Chapkin, R. S., 2012. Alteration of EGFR spatiotemporal dynamics suppresses signal transduction. *PLoS One*, 7: e39682.
- Turk, H. F., and Chapkin, R. S., 2013. Membrane lipid raft organization is uniquely modified by n-3 polyunsaturated fatty acids. *Prostaglandins, Leukotrienes and Essential Fatty Acids* (*PLEFA*), 88: 43-47.
- Van de Werf, F., Ardissino, D., Betriu, A., Cokkinos, D. V., Falk, E., Fox, K. A., Julian, D., Lengyel, M., Neumann, F. J., Ruzyllo, W., Thygesen, C., Underwood, S. R., Vahanian, A., Verheugt, F. W., and Wijns, W., 2003. Management of acute myocardial infarction in patients presenting with ST-segment elevation. The task force on the management of acute myocardial infarction of the European Society of Cardiology. *European Heart Journal*, 24: 28-66.
- Verbist, K. C., Wang, R., and Green, D. R., 2012. T cell metabolism and the immune response. *Seminars in Immunology*, 24: 399-404.
- Wallace, F. A., Miles, E. A., Evans, C., Stock, T. E., Yaqoob, P., and Calder, P. C., 2001. Dietary fatty acids influence the production of Th1- but not Th2-type cytokines. *Journal of*

Leukocyte Biology, 69: 449-457.

- Wang, Y., and Huang, F., 2015. N-3 Polyunsaturated fatty acids and inflammation in obesity: Local effect and systemic benefit. *BioMed Research International*, DOI: 10.1155/2015/ 581469.
- Watson, S. J., Brown, A. J., and Holliday, N. D., 2012. Differential signaling by splice variants of the human free fatty acid receptor GPR120. *Molecular Pharmacology*, 81: 631-642.
- Wei, D., Li, J., Shen, M., Jia, W., Chen, N., Chen, T., Su, D., Tian, H., Zheng, S., Dai, Y., and Zhao, A., 2010. Cellular production of n-3 PUFAs and reduction of n-6-to-n-3 ratios in the pancreatic beta-cells and islets enhance insulin secretion and confer protection against cytokine-induced cell death. *Diabetes*, **59**: 471-478.
- Williams, C. M., and Burdge, G., 2006. Long-chain n-3 PUFA: Plant v. marine sources. *Proceedings of the Nutrition Society*, 65: 42-50.
- Wu, J. H. Y., Micha, R., Imamura, F., Pan, A., Biggs, M. L., Ajaz, O., Djousse, L., Hu, F. B., and Mozaffarian, D., 2012. Omega-3 fatty acids and incident type 2 diabetes: A systematic review and meta-analysis. *British Journal of Nutrition*, 107: S214-S227.
- Wurtman, R. J., 2008. Synapse formation and cognitive brain development: Effect of docosahexaenoic acid and other dietary constituents. *Metabolism*, 57: 6-10.
- Xue, M., Wang, Q., Zhao, J., Dong, L., Ge, Y., Hou, L., Liu, Y., and Zheng, Z., 2014. Docosahexaenoic acid inhibited the Wnt/beta-catenin pathway and suppressed breast cancer cells *in vitro* and *in vivo*. *Journal of Nutritional Biochemistry*, 25: 104-110.
- Yaqoob, P., Pala, H. S., Cortina-Borja, M., Newsholme, E. A.,

and Calder, P. C., 2000a. Encapsulated fish oil enriched in alpha-tocopherol alters plasma phospholipid and mononuclear cell fatty acid compositions but not mononuclear cell functions. *European Journal of Clinical Investigation*, **30**: 260-274.

- Yaqoob, P., Pala, H. S., Cortina-Borja, M., Newsholme, E. A., and Calder, P. C., 2000b. Encapsulated fish oil enriched in alpha-tocopherol alters plasma phospholipid and mononuclear cell fatty acid compositions but not mononuclear cell functions. *European Journal of Clinical Investigation*, **30**: 260-274.
- Yog, R., Barhoumi, R., McMurray, D. N., and Chapkin, R. S., 2010. n-3 polyunsaturated fatty acids suppress mitochondrial translocation to the immunologic synapse and modulate calcium signaling in T cells. *Journal of Immunology*, **184**: 5865-5873.
- Zhang, M., Picard-Deland, E., and Marette, A., 2013. Fish and marine omega-3 polyunsatured fatty acid consumption and incidence of type 2 diabetes: A systematic review and metaanalysis. *International Journal of Endocrinology*, **10**: 1-11.
- Zhang, W., Bai, X., Ge, H., Cui, H., Wei, Z., and Han, G., 2014. Meta-analysis in the association between obesity and risk of thyroid cancer. *International Journal of Clinical and Experimental Medicine*, 7: 5268-5274.
- Zou, Z., Bellenger, S., Massey, K. A., Nicolaou, A., Geissler, A., Bidu, C., Bonnotte, B., Pierre, A. S., Minville-Walz, M., Rialland, M., Seubert, J., Kang, J. X., Lagrost, L., Narce, M.. and Bellenger, J., 2013. Inhibition of the HER2 pathway by n-3 polyunsaturated fatty acids prevents breast cancer in fat-1 transgenic mice. *Journal of Lipid Research*, 54: 3453-3463.

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