



The diet of *Stereocyclops incrassatus* Cope, 1870 (Anura: Microhylidae) tadpoles in Atlantic Forest of southern Bahia, Brazil

Caio Vinícius de Mira-Mendes^{1,2} · Edvaldo Moreira da Silva Neto³ · Jeniffer Cabral^{3,4} · Kaoli Pereira Cavalcante⁵ · Sylvia Maria Moreira Susini Ribeiro² · Iuri Ribeiro Dias³ · Yvonnick Le Pendu^{2,3}

Received: 1 November 2022 / Accepted: 19 April 2023 / Published online: 2 May 2023

© The Author(s), under exclusive licence to Plant Science and Biodiversity Centre, Slovak Academy of Sciences (SAS), Institute of Zoology, Slovak Academy of Sciences (SAS), Institute of Molecular Biology, Slovak Academy of Sciences (SAS) 2023

Abstract

Tadpole diet studies based in gut content analysis provide information on the diversity of prey consumed, foraging behavior and ecological function of tadpoles. Although trophic ecology studies have increased, tadpole diet is still understudied compared to the high diversity of anurans species, especially in some families such as Microhylidae, which present great morphological and ecological diversity. In this study we describe the diet of the tadpole of *Stereocyclops incrassatus*, from a population in southern Bahia, Brazil. Gut contents of 15 tadpoles were analyzed under light microscope (microscopic items) and stereoscopic microscope (macroscopical items). We found 14 categories of food items in the diet of the *S. incrassatus*, with detritus, composed primarily of degraded plant material, representing the most important food source (69%), followed by microalgae of the phylum Bacillariophyta (18%). The other items of the diet of *S. incrassatus* were composed mainly of Euglenophyta, Placidozoa, Testacea, Fungi, Nematoda, animal and protozoan fragments. Thus, tadpoles of *S. incrassatus* are classified as detritivorous, feeding especially on organic matter. The results of the present study show that suspension-feeder tadpoles besides being primary consumers are also detritivorous, and can play an important role in nutrient cycles in freshwater trophic webs.

Keywords Anuran larvae · Feeding ecology · Gut content analysis · Detritus · Microalgae

Introduction

Studies on feeding ecology help to understand the ecological function, resource partitioning, and trophic levels of consumers in a community (Paine 1980), and are essential to understand trophic interactions and the dynamics of food webs (Whiles and Altig 2010). Among the great diversity of species in continental aquatic environments, amphibian larvae (tadpoles) are among the least studied groups with respect to trophic relationships (Petranka and Kennedy 1999). Due to their high density, tadpoles exert great influence on the function and structure of aquatic ecosystems, shaping the density of algal communities and modifying primary production patterns and organic matter dynamics (e.g. Kupferberg 1997; Flecker et al. 1999). In addition, tadpoles serve as important food items for other consumers, participating directly in the transfer of energy and nutrients between aquatic and terrestrial ecosystems (Capps et al. 2015; Whiles et al. 2006).

✉ Caio Vinícius de Mira-Mendes
caiomendes@professor.uema.br

¹ Department of Biology, Universidade Estadual do Maranhão, São Luís, Maranhão 65055-310, Brazil

² Graduate program in Tropical Aquatic Systems, Universidade Estadual de Santa Cruz, Rodovia Jorge Amado, km 16, Ilhéus 45662-900, Bahia, Brazil

³ Graduate program in Zoology, Universidade Estadual de Santa Cruz, Rodovia Jorge Amado, km 16, Ilhéus 45662-900, Bahia, Brazil

⁴ Laboratory of Aquatic Organisms, Department of Biological Sciences, Universidade Estadual de Santa Cruz, Km 16, Ilhéus 45662-900, Bahia, Brazil

⁵ Center of Agricultural and Biological Sciences, Universidade Estadual Vale do Acaraú. Avenida da Universidade, 850, Campus Betânia, Sobral 62040-370, Ceará, Brazil

Although studies on the feeding ecology of tadpoles have increased in recent years (e.g. Asrafuzzaman et al. 2018; Mira-Mendes et al. 2020; Protázio et al. 2020), tadpoles diet is still understudied compared to the high diversity of anurans species (Altig et al. 2007; Frost 2021). A recent review on tadpole diets showed that most studies have been conducted on species from the families Hylidae, Bufonidae and Ranidae from Neotropic and Nearctic regions, while species in tropical regions lack trophic ecological data (Montaña et al. 2019). The Microhylidae family has a wide geographic distribution, and has one of the highest species richness among the anurans (> 740 spp.), standing out for its great morphological and ecological diversity (Frost 2023). However, studies of the tadpole trophic ecology of only eight species of microhylids are documented (Montaña et al. 2019). This gap may be related to the fossorial habit and the explosive breeding pattern in adults, observed in most species of microhylids (Duellman and Trueb 1994; Wells 2007), which cause tadpoles of this family to remain in aquatic environments for short periods.

Currently, Brazil is home to 59 species of microhylids, which represents 8% of the global diversity (Segalla et al. 2021). *Stereocyclops incrassatus* Cope, 1870 is distributed in the coastal forests of eastern Brazil from Northeast (Pernambuco Alagoas and Bahia) to Southeast Regions (Minas Gerais and Espírito Santo) (Frost 2023). Adults of this species can be found in the leaf-litter on the floor of primary and secondary forests, as well as in rubber and cocoa agroforestry systems, especially after periods of heavy rains (Dias

et al. 2014; Mira-Mendes et al. 2018; Peixoto et al. 2010). It is an explosive breeder using temporary waters within or at the edge of forests, where the tadpoles develop. In this study we describe the diet composition of *S. incrassatus* tadpoles from a population in southern Bahia.

Materials and methods

Specimen collection and preparation

The tadpoles of *S. incrassatus* were collected on August 30, 2019 in a semi-permanent pond located in an agroforestry system, named “cabruca”, in which cacao grows under the shade of native tree species [14°47′45.8″S, 39°10′20.9″W, 34 m above sea level], municipality of Ilhéus, state of Bahia, Brazil. The pond measured about 140 square meters, was ~ 1 m deep at the deepest area, and was mostly covered by aquatic vegetation (Fig. 1). Tadpoles were collected using a 20 cm diameter hand net with 2 mm steel mesh. They were immediately euthanized in 5% lidocaine solution and then preserved in 10% formalin. In order to identify the tadpoles, comparisons were made with specimens kept in the herpetological collection of Museu de Zoologia da Universidade Estadual de Santa Cruz, Ilhéus, State of Bahia, Brazil (MZUESC) and we consulted literature describing the tadpole species (Wogel et al. 2000; Dubeux et al. 2020). The tadpoles collected were between development stages 31 and 37 (Gosner 1960).

Fig. 1 A semi-permanent pond inside “cabruca” in the campus of the Universidade Estadual de Santa Cruz (UESC), municipality of Ilhéus, state of Bahia, Brazil



Diet analysis

We dissected digestive tracts of each specimen ($n = 15$) through a ventral incision. Then, we separated the first third of the intestine of each individual and placed the contents in a microtube with 2 mL of Transeau solution. The remaining part of the intestine was placed in another microtube in the same solution. Then, we homogenized the solution with the first third of the intestine, and with a plastic pipette we deposited approximately 0.5 mL of each sample on a glass slide of 75×24 mm covered with a coverslip (50×20 mm) for analysis under a light microscope. The items found in the entire area of glass slide were identified under a ZEISS Axio Scope light microscope. If there was any uncertainty during the identification process of an item, we consulted specific literature (Bicudo and Menezes 2017) and utilized a Canon Powershot G5 digital camera attached to a microscope to take a photograph of it, which was then sent to an expert (one of the authors, KPC). As suggested by Mira-Mendes et al. (2020) we analyzed the remaining intestinal content of each sample under a stereoscopic microscope in order to verify the presence of larger food items such as macroinvertebrates. For this purpose, the remaining intestinal contents were spread with 5 mL of water in a petri dish. The food items were identified at the lowest possible taxonomic level.

We calculated numeric percentage (NP), percentage of the frequency of occurrence (FO) and the importance index “I” proposed by Kawakami and Vazzoler (1980), with numerical percentage replaced by volume as suggested by Huckembeck et al. (2016):

$$I = F_i \times NP_i / \sum (F_i / NP_i) \times 100$$

where F_i is the frequency of occurrence of item i in the sample; and NP_i is the numeric percentage of the item.

Results

We found 2401 microscopic food items in the diet of the *S. incrassatus* tadpoles, distributed in 14 categories. Detritus, composed primarily of degraded plant material, was present in large amounts in all samples ($N\% = 64.5$; $FO\% = 100$), and represented the most important food source ($I = 69.23$). The other items were composed mainly of algae, although Placidozoa, Testacea, Fungi, Nematoda, animal and protozoan fragments were also found (Table 1).

After the detritus, microalgae of the phylum Bacillariophyta accounted for the highest numerical percentage ($NP = 18.9\%$) and frequency of occurrence ($FO = 93.3\%$), and also were the second most important food source ($I = 18.98$) to *S. incrassatus* tadpoles. Bacillariophyta was

composed of seven families, and eight genera, of which *Gomphonema* and *Pinnularia* were the most representative, corresponding to 35.2 and 25.9% from the remaining genera of the phylum, respectively.

The importance index of food of Euglenophyta was 4.65, with a predominance of the genera *Trachelomonas* ($NP = 52.5\%$) and *Phacus* ($NP = 36\%$). Cyanobacteria ($I = 3.37$) was represented mostly by the genus *Oscillatoria* ($NP = 77\%$). The other food items (Streptophyta, Chlorophyta, Ciliophora, Cryptophyta, Placidozoa, Testacea, Protozoa, Fungi, Nematoda and animal fragments), represent together the remaining value of the importance index of food ($I = 3.77$). We did not identify macroscopic food items in the diet of *S. incrassatus* tadpoles.

Discussion

Tadpoles are generally present in high densities in aquatic environments, usually have a high rate of ingestion of a variety of food items and can be considered the major consumers of freshwater aquatic ecosystems, particularly in temporary ponds (Alford 1999). Tadpoles of *S. incrassatus* are in the suspension-feeder ecomorphological guild (*sensu* Altig and Johnston 1989) by presenting the following characteristics: keratinized mouth parts absent, presence of tail flagellum; inhabit the water column; lateral eyes, body usually strongly depressed and low fins similar. Suspension-feeder tadpoles have been classified as primary consumers, as they occupy lower trophic levels (Altig et al. 2007; Whiles and Altig 2010; Montaña et al. 2019). Our results, based on the analysis of the tadpole gut contents, shows the diet of *S. incrassatus* is composed primarily of detritus (organic matter).

Detritus is frequently observed in the diet of tadpoles (Altig et al. 2007; Souza-Filho et al. 2007) and has been considered a primary food source for aquatic organisms in the trophic webs of several tropical freshwater ecosystems. Some studies have shown that the consumption by detritivorous tadpoles of the most nutrient-rich detritus fractions can increase their growth and survival rates (Kupferberg et al. 1994; Barrett et al. 2017). Furthermore, associated microbes and fungal biomass contribute significantly more to the nutritional value of detritus than the particles themselves (Cummins and Klug 1979; Methvin and Suberkropp 2003). Stable isotope analyses of some centrolenid tadpoles have corroborated this approach, indicating that they are primarily assimilating microbes rather than the detritus they consume (Whiles et al. 2006). Although it is difficult to differentiate between morphic (nutritionally poor) and amorphic detritus (nutritionally rich) in tadpole guts (Montaña et al. 2019), detritus may represent an important

Table 1 Microscopic diet of *Stereocyclops incrassatus* tadpoles

Food items	N	N%	FO	FO%	I
Bacillariophyta	455	18.95	14	93.33	18.98
Pinnulariaceae					
<i>Pinnularia</i>	118	4.91	8	53.33	
Gomphonemataceae					
<i>Gomphonema</i>	160	6.66	8	53.33	
Naviculaceae					
<i>Navicula</i>	21	0.87	2	13.33	
Eunotiaceae					
<i>Eunotia</i>	23	0.96	3	20.00	
Bacillariaceae					
<i>Nitzschia</i>	48	2.00	3	20.00	
Diadesmidaceae					
<i>Luticola</i>	13	0.54	1	6.67	
<i>Diadesmis</i>	5	0.21	1	6.67	
Sellaphoraceae					
<i>Sellaphora</i>	67	2.79	1	6.67	
Streptophyta	47	1.96	8	53.33	1.12
Zygnemataceae					
<i>Mougeotia</i>	26	1.08	3	20.00	
Closteriaceae					
<i>Closterium</i>	21	0.87	6	40.00	
Chlorophyta	36	1.50	5	33.33	1.07
Oedogoniaceae					
<i>Oedogonium</i>	13	0.54	3	20.00	
Chlamydomonadaceae					
<i>Chlamydomonas</i>	11	0.46	3	20.00	
Chlorophytes (NI)	12	0.50	5	33.33	
Ciliophora	20	0.83	6	40.00	0.36
Vorticellidae					
<i>Vorticela</i>	3	0.12	3	20.00	
Ciliates (NI)	17	0.71	3	20.00	
Cryptophyta	2	0.08	1	6.67	0.01
Cryptomonadaceae	2	0.08	1	6.67	
Cyanobacteria	87	3.62	13	86.67	3.37
Borziaceae					
<i>Borzia</i>	1	0.04	1	6.67	
Spirulinaceae					
<i>Spirulina</i>	14	0.58	5	33.33	
Oscillatoriaceae					
<i>Oscillatoria</i>	67	2.79	9	60.00	
Aphanizomenonaceae					
<i>Dolichospermum</i>	3	0.12	1	6.67	
Pseudanabaenaceae					
<i>Pseudoanabaena</i>	2	0.08	2	13.33	
Euglenophyta	120	5.00	13	86.67	4.65
Euglenaceae					
<i>Trachelomonas</i>	63	2.62	13	86.67	
<i>Euglena</i>	6	0.25	3	20.00	
Phacaceae					
<i>Lepocinclis</i>	15	0.62	5	33.33	
<i>Phacus</i>	36	1.50	12	80.00	
Placidozoa	3	0.12	1	6.67	0.01
Opalinidae					

Table 1 (continued)

Food items	N	N%	FO	FO%	I
<i>Opalina</i>	3	0.12	1	6.67	
Testacea	27	1.12	7	46.67	0.56
Arcellidae					
<i>Arcella</i>	7	0.29	3	20.00	
Lesquereusiidae					
<i>Lesquereusia</i>	2	0.08	1	6.67	
Trinematidae					
<i>Trinema</i>	12	0.50	6	40.00	
Euglyphidae					
<i>Euglypha</i>	6	0.25	3	20.00	
Protozoa	18	0.75	1	6.67	0.05
Fungi	22	0.92	6	40.00	0.39
Nematoda	6	0.25	5	33.33	0.09
Animal fragments	9	0.37	4	26.67	0.11
Detritus (organic matter)	1549	64.51	15	100.00	69.23

N = total number per item; NP (%) = numerical percentage; F = total frequency per item; FO = frequency of occurrence in percentage; I = importance index

item in the diet of *S. incrassatus* because it was present in large amounts in the gut of all examined tadpoles.

Most of the remaining items present in the diet of *S. incrassatus* tadpoles are periphytic taxa loosely attached to substrate or associated with the matrix of their own or other algae mucilage (e.g. *Pinnularia* spp., Bacillariophyta; *Mougeotia* spp., Streptophyta; *Oscillatoria* spp., Cyanophyta) (Burliga and Schwarzbald 2013). Although these organisms are not essentially planktonic, this means that depending on the hydrodynamics of the environment (e.g. shallower or deeper ponds), these taxa detach from the substrate and become available in the water column and can then be consumed by suspension-feeder species. Such hydrodynamic fluctuations often occur in shallow environments, as is the case in the semi-permanent pond where *S. incrassatus* tadpoles were collected. Furthermore, about 18% of the food items are represented by genera of flagellates or ciliates (e.g. *Chlamydomonas* spp., Chlorophyta; *Vorticella* spp., Ciliophora; *Trachelomonas* spp., Euglenophyta), which have structures that allow their movement in the water column for consumption by suspension-feeder species.

After detritus, Bacillariophyta, Cyanophyta and Euglenophyta also represented important items in the diet of *S. incrassatus* tadpoles. Algae are considered one of the most common food items present in the diet of tadpoles of different trophic guilds (e.g., benthic as *Macrogenioglottus alipioi* Carvalho, 1946, nektonic as *Scinax similis* (Cochran, 1952), suspension-rasper as *Pithecopus nordestinus* (Caramaschi, 2006)) because they represent a very abundant group in aquatic environments (Rossa-Feres et al. 2004; Mira-Mendes et al. 2020; Protázio et al. 2020). Some

studies have demonstrated that the consumption of algae with diatoms epiphytes can accelerate the development and growth of tadpoles (e.g. Kupferberg et al. 1994; Kupferberg 1997). This is very important for microhylid tadpoles, which exhibit an explosive breeding behaviour and need to leave the ephemeral/temporary ponds in a short period of time. Studies describing the diet of microhylids tadpoles, such as *Elachistocleis bicolor* (Guérin-Méneville, 1838), *Dermatonotus muelleri* (Boettger, 1885), *Kaloula pulchra* Gray, 1831 and *Microhyla ornata* (Duméril and Bibron, 1841) (Rossa-Feres et al. 2004; Candiotti 2007; Dey and Goswami 2015; Lalremsanga et al. 2017), showed that diatoms and filamentous algae also accounted for a significant proportion of consumed items.

Zooplankton items, composed of Placidozoa, Testacea, Protozoa (NI), Nematoda and Animal fragments parts, represented only 2.7 of the importance index and are of lesser importance in the diet of *S. incrassatus* tadpoles. The percentage of zooplankton items in the diet of other microhylid species is also low (e.g. Rossa-Feres et al. 2004; Echeverría et al. 2007; Asrafuzzaman et al. 2018). According to Altig et al. (2007) the presence of animal items in the diet of tadpoles can lead to an accelerated growth rate, even if in small proportions (Alford 1999).

Our results show that the *S. incrassatus* tadpoles can be classified as detritivores, feeding especially on organic matter. The results provided in the present study show that suspension-feeder tadpoles besides being primary consumers (Altig et al. 2007; Whiles and Altig 2010; Montaña et al. 2019) are also detritivores, and can play an important role in nutrient cycles in freshwater trophic webs. Gut content analysis studies are important because they provide information

on the diversity of prey consumed, foraging behavior and ecological function of tadpoles. However, complementary techniques such as stable isotope analysis are essential to validate the results of gut contents studies and to provide a better understanding of the resources assimilated by tadpoles.

Acknowledgements This study was financed in part by the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior –(CAPES). CVMM is grateful to CAPES for a fellowship (process PNPd/1682788) and give particular thanks to his wife Fernanda Tonolli and son Francisco Tonolli Mendes for their patience, affection and eternal loving support. IRD acknowledges the Conselho Nacional de Desenvolvimento Científico e Tecnológico – CNPq fellowship (PQ 315362/2021-9).

Author Contribution Conceptualization: CVMM; Methodology: CVMM, EMSN, JC; Formal analysis and investigation: CVMM, EMSN, JBC, KPC, SMMSR, IRD; Writing - original draft preparation: CVMM; Supervision: CVMM, YLP. All authors approved the final version of the manuscript.

Declarations

Conflict of interest There are no conflicts of interest related to this work.

Ethics approval No approval of research ethics committees was required to accomplish the goals of this study because no experimental work was conducted. Tadpoles were collected under the permanent permit n 13708-1 issued to Mirco Solé by Instituto Brasileiro do Meio Ambiente e dos Recursos Naturais Renováveis (IBAMA) and Instituto Chico Mendes de Conservação da Biodiversidade (ICMBio).

References

- Alford RA (1999) Ecology: resource use, competition, and predation. In: McDiarmid RW, Altig R (eds) Tadpoles: The Biology of Anuran Larvae. University of Chicago Press, Chicago, pp 240–278
- Altig R, Johnston GF (1989) Guilds of anuran larvae: relationships among developmental modes, morphologies, and habitats. *Herpetol Monogr* 3:81–109. <https://doi.org/10.2307/1466987>
- Altig R, Whiles MR, Taylor CL (2007) What do tadpoles really eat? Assessing the trophic status of an understudied and imperiled group of consumers in freshwater habitats. *Freshw Biol* 52:386–395. <https://doi.org/10.1111/j.1365-2427.2006.01694.x>
- Asrafuzzaman S, Mahapatra S, Rout J, Sahoo G (2018) Dietary assessment of five species of anuran tadpoles from northern Odisha, India. *J Threat Taxa* 10:12382–12388. <https://doi.org/10.11609/jott.3902.10.10.12382-12388>
- Barrett K, Crawford JA, Reinstein Z, Milanovich JR (2017) Detritus quality produces species-specific tadpole growth and survivorship responses in experimental wetlands. *J Herpetol* 51:227–231. <https://doi.org/10.1670/16-091>
- Bicudo CEM, Menezes M (2017) Gênero de algas de águas continentais do Brasil. Chave de identificação e descrições. 3a. edição. RiMa, São Carlos
- Burliga AL, Schwarzbald A (2013) Perifiton: diversidade taxonômica e morfológica. In: Schwarzbald A, Burliga AL, Torgan LC (eds) *Ecologia do perifiton*. Rima, São Carlos, pp 1–6
- Candiotti MFV (2007) Anatomy of anuran tadpoles from lentic water bodies: systematic relevance and correlation with feeding habits. *Zootaxa* 1600:1–175. <https://doi.org/10.11646/zootaxa.1600.1.1>
- Capps KA, Berven KA, Tiegs SD (2015) Modelling nutrient transport and transformation by pool-breeding amphibians in forested landscapes using a 21-year dataset. *Freshw Biol* 60:500–511. <https://doi.org/10.1111/fwb.12470>
- Cummins KW, Klug MJ (1979) Feeding ecology of stream invertebrates. *Annu Rev Ecol Evol Syst* 10:147–172. <https://www.jstor.org/stable/2096788>
- Dey M, Goswami S (2015) Diet of tadpoles of *Microhyla ornata* (Dumeril and Bibron, 1841) from a freshwater system in Rosekandy Tea Estate, Cachar, Assam and significance of conservation of aquatic habitats. *Curr World Environ* 10:238–244. <https://doi.org/10.12944/CWE.10.1.28>
- Dias IR, Mira-Mendes CV, Solé M (2014) Rapid inventory of herpetofauna at the APA (Environmental Protection Area) of the Lagoa Encantada and Rio Almada, Southern Bahia, Brazil. *Herpetology Notes* 7:627–637
- Dubeux MJM, Nascimento FAC, Lima LR, Magalhães FM, Silva IRS, Gonçalves U, Almeida JPF, Correia LL, Garda AA, Mesquita DO, Rossa-Feres DC, Mott T (2020) Morphological characterization and taxonomic key of tadpoles (Amphibia: Anura) from the northern region of the Atlantic Forest. *Biota Neotrop* 20:1–24. <https://doi.org/10.1590/1676-0611-BN-2018-0718>
- Duellman WE, Trueb L (1994) *Biology of Amphibians*. The Johns Hopkins University Press, Baltimore
- Echeverria DD, Volpedo AV, Mascitti VI (2007) Diet of tadpoles from a pond in Iguazu National Park, Argentina. *Gayana* 71:8–14
- Flecker AS, Feifarek BP, Taylor BW (1999) Ecosystem engineering by a tropical tadpole: density-dependent effects on habitat structure and larval growth rates. *Copeia* 1999:495–500. <https://doi.org/10.2307/1447498>
- Frost, DR. *Amphibian Species of the World: An Online Reference*. Version 6.1 (23 April). Electronic Database accessible at <https://amphibiansoftheworld.amnh.org/index.php>. American Museum of Natural History, New York, USA., <https://doi.org/10.5531/db.vz.0001> (2021).
- Gosner KL (1960) A simplified table for staging anuran embryos and larvae with notes on identification. *Herpetologica* 16:183–190. <https://www.jstor.org/stable/3890061>
- Huckembeck S, Alves LT, Loebmann D, Garcia AM (2016) What the largest tadpole feeds on? A detailed analysis of the diet composition of *Pseudis minuta* tadpoles (Hylidae, Dendropsophini). *An Acad Bras Ciênc* 88:1397–1400. <https://doi.org/10.1590/0001-3765201620150345>
- Kawakami E, Vazzole G (1980) Método gráfico e estimativa de índice alimentar aplicado no estudo de alimentação de peixes. *Bol Inst Oceanogr* 29:205–207. <https://doi.org/10.1590/S0373-55241980000200043>
- Kupferberg SJ (1997) Facilitation of periphyton production by tadpole grazing: functional differences between species. *Freshw Biol* 37:427–439. <https://doi.org/10.1046/j.1365-2427.1997.00170.x>
- Kupferberg SJ, Marks JC, Power ME (1994) Effects of variation in natural algal and detrital diets on larval anuran (*Hyla regilla*) life-history traits. *Copeia* 1994:446–457. <https://doi.org/10.2307/1446992>
- Lalremsanga HT, Sailo S, Hooroo RNK (2017) External morphology, oral structure and feeding behaviour of *Kaloula pulchra* tadpoles Gray, 1831 (Amphibia: Anura: Microhylidae). *Sci Technol J* 5:97–103
- Methvin BR, Suberkropp K (2003) Annual production of leaf-decaying fungi in 2 streams. *J North Am Benthol Soc* 22:554–564. <https://doi.org/10.2307/1468352>
- Mira-Mendes CV, Ruas DS, Oliveira RM, Castro IM, Dias IR, Baumgarten JE, Juncá JA, Solé M (2018) Amphibians of the

- Reserva Ecológica Michelin: a high diversity site in the lowland Atlantic Forest of southern Bahia, Brazil. *ZooKeys* 753:1–21. <https://doi.org/10.3897/zookeys.753.21438>
- Mira-Mendes CV, Almeida CSS, Junqueira RA, Cavalcante HP, Ribeiro SMMS, Mariano R, Dias IR, Solé M, Le Pendu Y (2020) The diet of *Macrogenioglottus alipioi* Carvalho, 1946 (Anura: Odontophrynidae) tadpoles in Atlantic Forest of southern Bahia, Brazil. *Stud Neotrop Fauna Environ* 57(3):262–266. <https://doi.org/10.1080/01650521.2020.1861888>
- Montaña CG, Silva SDGTM, Hagyar D, Wager J, Tiegls L, Sadeghian C, Schriever TA, Schalk CM (2019) Revisiting “what do tadpoles really eat?” A 10-year perspective. *Freshw Biol* 64:2269–2282. <https://doi.org/10.1111/fwb.13397>
- Paine RT (1980) Food webs: linkage, interaction strength and community infrastructure. *J Anim Ecol* 49:667–685. <https://doi.org/10.2307/4220>
- Peixoto OL, Pimenta B, Carvalho-e-Silva SP (2010) *Stereocyclops incrassatus*. The IUCN Red List of Threatened Species 2010 eT58006A11712656. <https://doi.org/10.2305/IUCN.UK.2010-2.RLTS.T58006A11712656.en>. Accessed 17 October 2022
- Petranka JW, Kennedy CA (1999) Pond tadpoles with generalized morphology: is it time to reconsider their functional roles in aquatic communities? *Oecologia* 120:621–631. <https://doi.org/10.1007/s004420050898>
- Protázio AS, Protázio AS, Gama V, Silva SV, Santos CGC, Oliveira JKG (2020) Diet of tadpoles for five anuran species of north-east Brazil. *J Limnol* 79:180–186. <https://doi.org/10.4081/jlimnol.2020.1912>
- Rossa-Feres DC, Jim J, Fonseca MG (2004) Diets of tadpoles from a temporary pond in southeastern Brazil (Amphibia, Anura). *Rev Bras Zool* 21:745–754. <https://doi.org/10.1590/S0101-81752004000400003>
- Segalla MV, Berneck B, Canedo C, Caramaschi U, Cruz CAG, Garcia PCA, Grant T, Haddad CFB, Lourenço ACC, Mângia S, Mott T, Nascimento LB, Toledo LF, Werneck FP, Langone JA (2021) Brazilian amphibians: list of species. *Herpetologia Brasileira* 10:121–216
- Sousa Filho IF, Branco CC, Carvalho-e-Silva AMPT, Silva GR, Sabagh LT (2007) The diet of *Scinax angrensis* (Lutz) tadpoles in an area of the Atlantic Forest (Mangaratiba, Rio de Janeiro) (Amphibia, Anura, Hylidae). *Rev Bras Zool* 24:965–970. <https://doi.org/10.1590/S0101-81752007000400012>
- Wells KD (2007) *The Ecology and Behavior of Amphibians*. University of Chicago Press, Chicago and London
- Whiles MR, Altig R (2010) Dietary assessment of larval amphibians. In: Dodd CK Jr (ed) *Amphibian Ecology and Conservation*. Oxford University Press, New York, NY
- Whiles MR, Lips KR, Pringle CM, Kilham SS, Bixby RJ, Brenes R, Connelly S, Colon-Gaud JC, Hunte-Brown M, Huryn AD, Montgomery C, Peterson S (2006) The effects of amphibian population declines on the structure and function of neotropical stream ecosystems. *Front Ecol Environ* 4:27–34
- Wogel H, Abrunhosa PA, Pombal JP (2000) Girinos de cinco espécies de anuros do sudeste do Brasil (Amphibia: Hylidae, Leptodactylidae, Microhylidae). *Bol Mus Nac NS Zool* 427:1–16

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.