



Biomarkers of vascular disease in diabetes: the adipose-immune system cross talk

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Abstract

Experimental and clinical studies aimed at investigating the mechanism(s) underlying vascular complications of diabetes indicate that a great number of molecules are involved in the pathogenesis of these complications. Most of these molecules are inflammatory mediators or markers generated by immune or adipose tissue. Some of them, i.e. resistin and sortilin, have been shown to be involved in the cross talk between adipocytes and inflammatory cells. This interaction is an attractive area of research, particularly in type 2 diabetes and obesity. Other proteins, such as adiponectin and visfatin, appear to be more promising as possible vascular markers. In addition, some molecules involved in calcium/phosphorus metabolism, such as klotho and FGF23, have an involvement in the pathogenesis of diabetic vasculopathy, which appears to be dependent on the degree of vascular impairment. Inflammatory markers are a promising tool for treatment decisions while measuring plasma levels of adipokines, sortilin, Klotho and FGF23 in adequately sized longitudinal studies is expected to allow a more precise characterization of diabetic vascular disease and the optimal use of personalized treatment strategies.

Keywords Diabetes · Vascular complications · Biomarkers · Atherosclerosis

Introduction

The increasing prevalence of type 2 diabetes mellitus (T2DM) is associated with an overall rise of vascular diseases [1], such as coronary artery disease (CAD), peripheral artery disease (PAD) and stroke. Taken together, these conditions are the leading causes of morbidity and mortality in western countries. In addition, they induce a major impact on public health, in term of quality of life and economic burden. Assessing the risk of diabetic complications may

facilitate the use of preventive strategies and innovative treatments.

To this end, over the last two decades, plasma levels of several molecules were investigated as potentially associated with the pathogenesis and the evolution of vascular damage, particularly in diabetes mellitus. In clinical practice glycated hemoglobin, albuminuria, serum creatinine and estimated glomerular filtration rate (eGFR) are now routinely measured in diabetic patients. Despite their utility in the prediction of cardiovascular outcomes [2–6], these molecules remain important, being the result of the vascular damage rather than being involved in its pathogenesis [4, 7]. Some data seem to underline the role of T2DM as a promoter of inflammation with an apparently causal link between high levels of inflammatory markers and the occurrence of vascular complications. The adipose tissue is emerging as a new source of inflammatory molecules with an attitude to self-potentiating effects due to a cross talk between adipocytes and inflammatory cells. In addition, the inflammation worsens the insulin resistance while hyperglycemia seems to increase the incidence of major vascular events per se [8].

In this scenario, a growing number of plasma biomarkers are being tested as prognostic and, in some case, therapeutic tools for diabetic vascular complications.

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The aim of this review is retracing the role of molecules identified in the last years, to facilitate an overview of the main results and encourage further investigations in this field.

Potential diagnostic and prognostic biomarkers of vascular diabetes complications

Inflammation-related biomarkers

C-reactive protein (CRP) is widely used as a reliable marker of inflammation. This protein has a key role in the modulation of innate immunity since it promotes the activation of phagocytes and of the complement pathway and the production of cytokines [9]. These effects may account for CRP involvement in atherosclerosis and diabetes mellitus [10]. High sensitivity CRP (hs-CRP), increased plasma levels are considered an independent cardiovascular risk factor which can increase the accuracy of Framingham score in risk prediction [11]. The pro-inflammatory effect of CRP has been attributed to its binding to lectin-like oxidized LDL receptor-1 (LOX-1), expressed by endothelial cells. In fact, this process promotes the expression of more LOX-1 scavenger receptors, with a positive feedback loop. Moreover, it directly determines an augmented intake of proatherogenic and proinflammatory molecules inside endothelial cells, such as oxidized LDL (OxLDL) or electronegative L5 LDL whose plasma levels are particularly increased in patients with high cardiovascular risk, in smokers and in patients affected by dyslipidemia and diabetes mellitus [12]. Furthermore, high levels of OxLDL and L5 LDL increase the CRP levels in atheromatic plaque thus sustaining the vicious cycle between inflammation and atherosclerosis [13]. CRP facilitates the development of foam cells and cause platelet adhesion [10]. Further studies have also demonstrated an enhanced CRP-mediated uptake of OxLDL uptake by macrophages through FCyRs that leads to an overexpression of NF- κ B [13]. Furthermore, high hs-CRP is markedly related to endothelial dysfunction in diabetic patients [14]. The very low specificity of CRP limits its use, since hepatic and local production of CRP constitutes general response to any kind of inflammatory trigger [15].

Patients affected by diabetic PAD often undergo endovascular revascularization procedures. This approach is frequently affected by a higher rate of post-procedural complications such as major adverse limb events (MALE) and major adverse cardiovascular events (MACE). Elevated plasmatc levels of CRP show a linear association with worse vascular outcomes in diabetic PAD [10] being an independent predictor for limb amputation in diabetic patients [16].

Pentraxin 3

Pentraxin 3 (PTX3) is an acute phase reactant of the same family of CRP. It seems to act as a local inflammation marker produced by endothelial, smooth muscle and dendritic cells, macrophages, neutrophils when they are activated by tumor necrosis factor (TNF)- α , interleukin (IL)-1 β and other toll-like receptors agonists [15]. PTX3 has been detected at high concentrations in human carotid plaques and coronary atherosclerotic lesions of patients with acute myocardial infarction and unstable angina [17].

A causal link between PTX3 and endothelial damage has been hypothesized in vascular complications of diabetic patients [18].

Tumor necrosis factor- α

There is evidence suggesting that TNF- α may play an important role in the pathogenesis of atherothrombosis of diabetic patients [10]. Higher levels of TNF- α are associated with higher failure rates of lower extremity endovascular revascularization and increased risk of MACE and MALE [10]. Among diabetic patients, those who are affected by diabetic ulcer foot present a higher concentration of TNF- α [19]. However, the causative link between high TNF values and adverse events still needs to be demonstrated. Indeed, anti-TNF strategies have been thought to be beneficial in several inflammatory diseases including atherosclerosis [20]. However, the negative effect on lipid profile and the lack of positive outcome detected in patients with heart failure [20] limited further studies on the therapeutic approach targeting TNF- α [11].

Potential benefits have already been shown with other anti-inflammatory drugs.

Colchicine, a classic old drug used for chronic inflammatory pathologies, is correlated, in these diseases, with better cardiovascular outcomes [21]. Similar benefits were also documented in patients at risk who have been treated with low doses of methotrexate [11].

Interleukins

Interleukin-1 (IL-1) family is constituted by IL-1 α and IL-1 β , which have a pro-inflammatory effect, and by the IL-1 receptor antagonist, IL1-Ra that counteracts the effect of IL-1.

IL-1 pro-inflammatory properties result from the induction of leukocyte adhesion molecules, from the expression of prostaglandin-E2 and from cellular release of histamine, which promotes vasodilatation and permeability [22].

In diabetic patients, IL-1 contributes to pancreatic cell apoptosis leading to hyperglycemia, which results in an inflammatory burden on endothelium [22]. Moreover, the

therapeutic reduction of IL-1 concentrations (through the administration of IL-1Ra) improves glycemic control, reduces CRP and decreases the incidence of cardiovascular disease [23].

IL-1 α promotes inflammation, through the induction of IL-6 with an indirect pro-inflammatory effect that is also shared by the isoform IL-1 β [11].

The role of IL-1 β in diabetic atherosclerosis and its modulation as possible therapeutic strategy is receiving increasing attention. In fact, IL-1 β is implicated in the development of the inflammasome of atherosclerotic plaques [11]. Moreover, IL-1 β production enhances by “autoinduction” both the expression of its own gene and of several other genes in cells involved in atherosclerosis [24]. This molecule decreases cardiac contractility by worsening cell injury associated with reperfusion and remodeling [25]. This cytokine stimulates smooth muscle cell proliferation [26] and regulates leukocyte adhesion molecules as ICAM-1 and VCAM-1; furthermore, it promotes the induction of cyclooxygenase-2 (COX-2) nitric oxide synthase (iNOS). Finally, IL-1 β activates cells belonging to the innate immunity system [11]. The role of IL-1 β in the development and progression of atherosclerosis led to test a specific treatment to antagonize its effects [11]. Canakinumab is a human monoclonal antibody which selectively inhibits IL-1 β and has a positive effect in IL-1 β mediated pathologies such as juvenile inflammatory arthritis [27], and acute gouty arthritis [11]. Specifically, in cardiovascular diseases this monoclonal antibody allows a reduction in plasma concentrations of hs-CRP, IL-6 and fibrinogen, with possible benefits for patients who present a residual inflammatory risk, defined as an hs-CRP greater than 2 mg/L, after the optimization of standard therapy [11].

Furthermore, IL-1 inhibition treatment decreases the risks of overall MACE, unstable angina, and breakthrough or recurrence of heart failure [28].

IL-18 is an additional member of the IL-1 superfamily. It works as a starter of the inflammatory process through the induction of interferon gamma (IFN γ) and the over-activation of Nf-kB [22].

Interleukin 18 is activated together with IL-1 β [11]. Higher levels of IL-18 correlate with increased incidence of both diabetes mellitus and diabetic complications [22].

In addition, the role of chemotaxis is being investigated in atherosclerosis. In particular, the monocyte chemoattractant protein-1 (MCP-1/CCL2), a chemoattractant of monocyte, and its receptor C–C chemokine receptor type 2 (CCR2) seem to have an important role in atherosclerosis [29]. Recent studies have focused on monocyte behavior in cardiovascular disease [11].

In diabetes mellitus, there are increased plasma concentrations of MCP-1, which promote monocyte migration into

the vascular wall and foam cell generation. Higher MCP-1 levels are common in patients with high levels of hs-CRP and in those with carotid artery thickness [30].

In addition, epicardial adipose stores of patients with critical CAD overexpress MCP-1 mRNA that could partially account for the increased incidence of myocardial infarction and stroke [31] observed in these patients.

The binding of MCP-1 to CCR2 receptor plays a role in the development of insulin resistance, obesity and diabetic major vessel complications [32].

This recognition of the role of MCP-1 in the inflammatory and atherosclerotic process led to development of new possible therapeutic strategies, such as monoclonal antibodies aimed to inhibit CCR2 and reduce inflammation [11]. Additionally, targeting MCP-1 could become a potential new direct treatment of diabetes mellitus and its major complications [33].

Moreover, since the overexpression of this chemoattractant protein might be involved in the pathophysiological process of diabetes mellitus, it becomes an available marker to guarantee a prompt diagnosis preventing the development of vascular complications [32].

IL-6 is a pro-inflammatory cytokine produced by lymphocytes and macrophages, which is able to stimulate the immunity system. High levels of this interleukin are predictor of MACE and MALE in diabetic patients with PAD as well as of poor outcome of endovascular revascularization procedures [10].

IL-6 is the main starter for inflammasome development and the importance of this cytokine in atherosclerosis is clearly documented by the lower incidence of cardiovascular adverse events occurring in patients with a particular loss-of-function variant of IL-6 receptor that inhibits the proinflammatory downstream pathway [34].

Tocilizumab, a monoclonal antibody that inhibits IL-6, has been shown to have a potential benefit in several diseases and to reduce CRP increase after primary coronary artery procedures [11]. However, blocking both IL-6 signaling pathways (the classic and the trans-signaling one), results in an impairment of immunity as well as of insulin sensitivity [24].

New approaches to selectively inhibit part of the IL-6 pathways are being investigated. The inhibitor of p38 MAP kinase (Iosmapimod) does not seem to affect the IL-6 pathway in a manner resulting in a reduction of MACE [11].

High mobility group box-1

The high mobility group box-1 (HMGB-1) is a nuclear protein that regulates gene expression and active pro-inflammatory responses in damaged and necrotic endothelial cells [35]. It is an inflammatory mediator able to induce the secretion of other inflammatory molecules such

as IL-6 and TNF- α by neutrophils and macrophages [36]. There is evidence of a cross talk among HMGB-1 and pro-inflammatory cytokines, suggesting a contribution of this pathway to the inflammatory process and to atheroma formation [37]. Indeed, several studies have documented a correlation between HMGB-1 levels, diabetes and its complications [38]. In addition, Oozawa et al. found that HMGB1 plasma levels were increased in diabetic patients with PAD [39]. We demonstrated in a large population of T2DM patients that HMGB-1 plasma levels are significantly increased in PAD patients with a positive correlation with clinical severity of vascular impairment [40].

In addition, HMGB-1 increases the expression of osteoprotegerin (OPG) [41] whose role as biomarker of vascular impairment is described below.

Biomarkers generated by adipose tissues

Adipokines

Adipokines are bioactive substances produced by visceral adipose tissue. These molecules have a role in the regulation of the balance between pro-inflammatory and anti-inflammatory effects of adipose tissue (Fig. 1). There are experimental and clinical data showing that this balance is compromised in T2DM [42]. Adipose tissue dysfunction contributes to insulin resistance, vascular injury and consequent vascular disease. Several studies are investigating the possible role of adipokines as biomarkers for diabetic vascular complications. Evidences are still controversial about the negative, positive or independent association between the levels of different adipokines and cardiovascular risk factors.

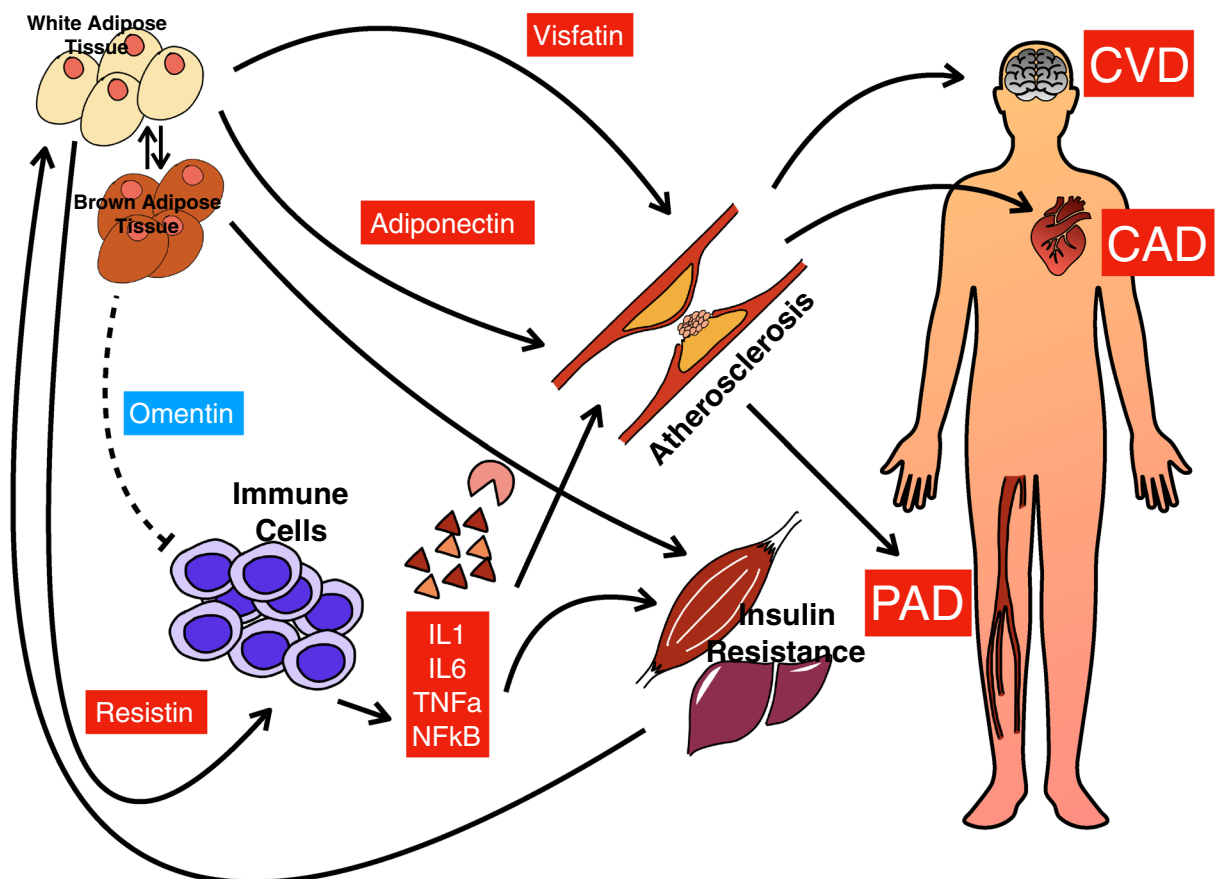


Fig. 1 Effect of different adipokines on inflammation and vascular complications. Different molecules, produced by different phenotypes of adipocytes, on inflammation, atherosclerotic plaque growth, glucose metabolism. Adipose-immune system cross talk influences the phenotype of adipose tissue and has an impact on the risk of coronary

artery disease (CAD), cerebrovascular disease (CVD) and peripheral artery disease (PAD). *IL1* interleukin 1, *IL6* interleukin 6, *TNF α* Tumor Necrosis Factor- α , *NF κ B* nuclear factor kappa-light-chain-enhancer of activated B cells

Adiponectin is a protein composed of 244 amino acids, encoded by a gene located on chromosome 3q27, which is related to T2DM and cardiovascular disease [43]. It is mainly synthesized in white adipose tissue. Several studies suggest that hypo adiponectinemia is associated with CAD indicating a protective role of adiponectin against the development of atherosclerosis process [44, 45]. Low serum adiponectin level may be considered an additional cardiovascular risk factor also for diabetic patients [46]. Al-Daghri et al. have studied this association at the genetic level. They found a link between a gene variant of adiponectin and CAD [47]. Other studies, performed in a population at intermediate-high CAD risk, documented that high plasma levels of adiponectin in diabetic patients are associated with higher rates of MACE rates [48]. Thus, adiponectin levels might need to be differently interpreted in populations with different degree of vascular impairment.

Omentin is a protein composed of 313 amino acids, encoded by a gene present on the chromosomal region 1q22-q23, that is related to T2DM [49]. Omentin mRNA is mostly expressed in the fraction of the vascular stroma of visceral adipose tissue, rather than in subcutaneous adipose tissue and in mature adipocytes. It has also been identified in other tissues, such as endothelium, epicardial adipose tissue, thymus, small intestine, colon, ovary, lung and placenta. There are two homologous isoforms: omentin-1, the most common form in human plasma, and omentin-2, which shares 83% of amino acids with isoform 1. Basic and clinical research has shown an anti-inflammatory effect of omentin-1 and an inverse correlation between its plasma levels and insulin resistance, diabetes, obesity and metabolic syndrome [50]. In mice, Yamawaki et al. have shown that treatment with omentin prevents inflammation of the endothelium by blocking the activation of JNK and NF- κ B induced by TNF α and reducing the expression of adhesion molecules [51]. Several studies have investigated the relationship between omentin-1 and CAD, stroke, and PAD [52] in diabetic patients. These studies, mostly cross-sectional and conducted on small and heterogeneous populations, provide discordant data about the negative, positive or independent correlation between omentin-1 levels and cardiovascular risk. Menzel et al. suggest that the different metabolic conditions may variably influence the levels of omentin and explain these results [53]. Therefore, prospective data are necessary to define the role of this adipokines as biomarker of an earlier diagnosis and management of vascular disease in diabetic patients.

Resistin is a protein synthesized in white adipose tissue, which is able to induce insulin resistance in muscles and liver and to promote the formation of early atherosclerotic lesion by proinflammatory pathways [54]. Several studies have documented that resistin plasma levels are elevated in obese and diabetic patients [55]. In particular, On et al. reported that resistin levels were higher in diabetic patients with CAD than

in those without CAD, and the pre-procedural resistin levels were higher in diabetic patients that manifested restenosis after stenting [56]. Indeed, resistin seems to induce the proliferation of smooth muscle cells and may have a role in restenosis of coronary lesions in patients with diabetes. Therefore, resistin assay may be useful in diabetic patients for assessing the risk of cardiovascular disease as well as the risk of restenosis after stenting [57].

Visfatin is an adipokine produced and secreted by visceral fat, whose plasma levels are correlated with obesity, insulin resistance and T2DM [58]. Several studies documented a role of visfatin as inflammatory mediator. It is involved in endothelial dysfunction and atherosclerotic plaque instability [59]. Mazaheroun et al. have found that visfatin plasma levels were high in patients with acute myocardial infarction (AMI) and found that these levels were highly sensitive and specific for this condition [60]. Visfatin seems to play a role in the pathogenesis of vascular damage in diabetic patients. Further investigations are needed to confirm these data and verify a potential diagnostic role of this adipokine.

Biomarkers related to lipid metabolism

Sortilin

Sortilin is a protein of 95 kDa encoded by the gene SORT1 located on chromosome 1. This protein is mainly expressed in hepatocytes and some studies have evidenced its role in apolipoprotein trafficking inside hepatocytes. Membrane sortilin promotes the uptake of LDL by hepatocytes via an LDL-receptor (R)-independent mechanism and also by macrophages, in the process of foam cell formation in atheroma [61]. Furthermore, there is evidence of a role of sortilin in intracellular cytokine traffic and in platelet activation [62]. Based on these data several researchers have investigated an association among sortilin, atherosclerosis, diabetes and cardiovascular complications [63]. Oh and colleagues [64] reported that circulating sortilin levels are increased in diabetic patients with CAD and in patients with CAD risk factors. Our group found that circulating sortilin levels are independently associated with PAD in a statin-free diabetic population [65]. Additionally, we found that sortilin levels correlate with PAD severity in diabetic patients, suggesting a dose-dependent relationship. The significance of these interesting findings needs to be strengthened by prospective studies.

Biomarkers related to calcium/phosphorus metabolism

Osteoprotegerin

OPG is a member of the Tumor Necrosis Factor (TNF) receptor family involved in bone turnover and vessel

calcification. Several studies have documented the role of OPG in the progression of atherosclerosis in diabetic patients [66]. OPG has also been described as an independent risk factor for disease progression in patients with CAD [67]. In PAD, there are contrasting data on the role of OPG plasma levels. Recent studies demonstrated higher OPG concentrations in T2D patients with PAD than in those without PAD [40]. However, no data are available on the possible coexistence of CAD in these relatively small patient populations. Thus, whether OPG levels might have a different behavior in diabetic vs non-diabetic or in CAD vs PAD subjects still needs to be demonstrated.

Klotho

The lower serum levels of phosphorus detected in diabetic patients have suggested an unbalance of phosphorus regulation in these patients [68]. The phosphate homeostasis is guaranteed by several hormones and factors such as vitamin D, growth hormone, thyroid hormone, calcitonin, glucocorticoids, atrial natriuretic factor, parathyroid hormone (PTH) and FGF23 [69]. The precipitation of calcium salts in arterial walls leads to vascular calcification. This has been shown to be an independent risk factor of cardiovascular disease and mortality [70]. Calcium precipitation is promoted by a dysfunction of mineral metabolism and both low and high bone-turnover are associated to a higher mineral deposition in blood vessels. T2DM is correlated to a state of low bone turnover [71] and, additionally, high PTH serum levels are associated with early vascular calcification in lower limbs of T2DM patients. In these subjects, tibial artery calcification is a predictor of PAD, diabetic ulcer foot and MALE [72]. Vitamin D deficiency is a main trigger of PTH elevation, thus suggesting that vitamin D supplementation may be beneficial in reducing vascular complications of diabetic patients [72].

FGF-23/ α -klotho axis is an adjunctive regulator of mineral metabolism. The perturbation of this recently described pathway seems to accelerate arterial calcification and progression of vascular complications [73]. Indeed, elevated serum concentrations of FGF23 and low levels of circulating Klotho are markers of cardiac and large vessel complications in pre-diabetes and diabetes [74].

A number of studies document a strong association between FGF23 and cardiovascular adverse events in T2DM. Elevated serum concentrations of this hormone are correlated with a high incidence of MALE and mortality in diabetic patients as well as with high incidence of diabetic foot [66].

Klotho also plays a significant role in glucose metabolism and regulation [75]. Despite a possible promotion of insulin resistance, due to a decreased glucose cell uptake [75], its interference with insulin/IGF-1 pathway causes an inhibition

of AKT kinase. This prevents the phosphorylation of FOXO transcription factor and inhibits the expression of catalase or mitochondrial manganese superoxide dismutase which cooperate in the reduction of reactive oxygen species [75].

The role of Klotho in diabetes is also suggested by the fact that Klotho is a protective factor against diabetic ulcer foot, it attenuates the progression of diabetic nephropathy and is an independent protective factor against macrovascular major adverse events (stroke and myocardial infarction) [76].

Atherosclerotic inflammation reduces the expression of Klotho in arterial wall cells probably by the expression of NF- κ B, specifically through cytokines of TNF- α superfamily [77]. The lack of protective role of Klotho causes an endothelial dysfunction leading to a degenerative process of vessels wall such as endothelial cells apoptosis, decreased expression of endothelial cadherin, smooth muscle cells calcification, increase of adhesion molecules as VCAM-1 and ICAM-1, oxidation, decrease of nitric oxide and loss of endothelium integrity and function [78]. To confirm the protective role of Klotho, the infusion of exogenous Klotho reverses these degenerative effects likely through the induction of nitric oxide production and release [79]. These evidences suggest a possible implication of Klotho as predictor of sub-clinical atherosclerosis, since a reduction of the hormone serum levels is a marker of coronary disease, heart failure, stroke and peripheral artery disease [80].

Exosomes

Exosomes (EXOs) constitute a subgroup of extracellular vesicles (EVs), rich in bioactive molecules such as DNA, messenger RNA, micro-RNA (miRNA) and other proteins. Several findings show their role as drivers of intracellular communications. Many researchers have investigated a possible correlation between EXOs and CVD [81–83]. Indeed, in response to stress or injury conditions, contents of EXOs may be up- or down-regulated, giving a signal and this may be used to drive the diagnosis and prognosis for CVD. However, methods of EXOs collection, isolation, and purification are still undergoing standardization [84].

Among the identified EV RNAs, miRNA has been closely associated with CVD. Different miRNAs have different roles and implications [15]. For example, miRNA-26 has likely a role in the modulation of inflammation and leukocyte adherence [85]. Micro RNA-100, according to intravascular ultrasound discoveries that correlate its levels with atherosclerotic plaque composition [86], reduces the expression of mammalian target of rapamycin (mTOR) signaling a pathway promoting more stability of the coronary plaque [15]. An elevation of miRNA-126 concentrations was observed in diabetic patients with a strict glycemic control through an optimization of anti-diabetic treatment. This molecule

downregulates vascular inflammation and reduces the tissue factor expression in vessels with an antithrombotic effect [87].

Potential therapeutic implications

The occurrence of T2DM and related complications seems to be directly associated to inflammation which worsens the hyperglycemic state and insulin resistance. Moreover, the growing fat mass in obese patients is one important source of chronic inflammation in diabetic patients. The unbalance between the fast growth of adipose tissue and the ineffective vascular support leads to tissue hypoxia that generates and sustains subclinical inflammation [8]. Other assumed causes of this low grade pro-inflammatory process, in diabetic patients, are periodontitis [88] and diet-related gut microbiota unbalance [8]. In addition, several molecules, such as CCL2 (MPC-1), CCL5 (RANTES), CXCL8 (IL-8), TNF- α , IL-6, IL-1, CRP, are implied in this pro-oxidant chronic phenomenon which increases the incidence of vascular adverse events [8].

Clinical evaluation of inflammatory status: in which patients and when

Patients with PAD and diabetic PAD can be divided in two different groups: the asymptomatic and the symptomatic patients such as those with claudication [89].

It has been shown that, in patients with asymptomatic PAD, plasma levels of inflammatory markers are similar to those we find in control cases, while patients with intermittent claudication have higher levels of CRP and IL-6, and a lower flow mediated dilatation (FMD) than asymptomatic ones [90]. These data seem to indicate that the inflammatory status and the endothelial function are related to the severity of PAD [90]. This hypothesis is supported by the fact that ankle-brachial pressure index (ABPI), which represents the degree of circulatory impairment in the affected limb is positively correlated with plasma levels of CRP, soluble intercellular adhesion molecule-1 (sICAM-1) and Soluble vascular cell adhesion molecule-1 (sVCAM-1), and positively correlated with flow-mediated dilatation (FMD) [90].

The degree of systemic inflammation also has an inter-individual variability depending on the severity of the inflammatory status that is influenced by the quantity of atherosclerotic plaques and the quality of the composition of these lesions [90]. In fact, symptomatic claudicant patients are characterized by a systemic increased activation of neutrophil, augmented concentrations of adhesion molecules (soluble I-CAM and V-CAM), thromboxane, endothelin-1, and IL-8 with an associated endothelial dysfunction [90]. In particular, these patients present a chronic

sustained inflammation with transient recidivate burdens that promote the maintenance and the amplification of this pro-inflammatory substrate increasing the cardiovascular risk [90]. Moreover, these recurrent acute inflammation loads and impairment of endothelial function are markedly reduced after a procedure of revascularization. Or after an adequate aerobic rehabilitation program which decreases the ischemic insults occurring during walk [90]. Revascularization is an effective therapeutic procedure to restore a valid blood flow to ischemic tissues of lower limbs in diabetic PAD. However, the stressful stimulus on arterial wall provoked by ballooning and stent placement (particularly in long extended stenosis) promotes local inflammation with an increase of inflammatory proteins [10]. We propose to test the hypothesis that inflammatory markers may be useful to personalize the optimal timing of PAD revascularization procedures. Patients who after the optimization of pharmacological and rehabilitation treatment have persistently high hs-CRP, IL6 and TNF α should have an earlier revascularization and should be treated with stenting, whenever possible. Furthermore, they should receive a narrower follow-up than the others, given the high risk of MALE and MACE. In this view the monitorization of inflammatory markers might also help deciding in which patients the use of anti-inflammatory agents might be most helpful [91]. Also, antidiabetic agents have different anti-inflammatory properties.

Rizza et al. have proved that treatment with the thiazolidinedione, a class of insulin-sensitizing drugs, increases the expression and secretion of adiponectin [92]. Other studies have documented that pioglitazone, a thiazolidinedione, improves cardiovascular outcomes in diabetic patients [93] an effect that may be related to adiponectin levels. In the same way, fenofibrate seems to increase adiponectin levels. Studies show a reduction of cardiovascular events and an improvement of angiogenic repair in ischemic limbs in patients with T2DM treated with this drug [94].

A number of novel approaches were made recently available such as the biologic therapies targeting molecules [95], lipid mediators (phospholipase inhibitors and antileukotrienes [95] or intracellular signaling pathways (NADPH oxidase, p38 mitogen-activated protein kinase, phosphodiesterase [96], MAP kinase [95]. This scientific effort indicates the role attributed to inflammation in atherosclerosis and the search by the scientific community of ways to slow down the development and the progression of this degenerative process [96].

Conclusion

The assay of several cytokines, such as sortilin, omentin, OPG, klotho, IL-6, TNF- α and especially hs-CRP can be used to identify diabetic patients having an inflammatory

Table 1 Effect of different cytokines on diabetes vascular complications

	Predictive	Prognostic	Therapeutic
Adiponectin	↓Serum levels are associated with CAD	↑Serum levels are related to the severity of CAD	Pioglitazone ↑adiponectin levels and is associated improves cardiovascular outcomes in diabetic patients Fenofibrate ↑adiponectin levels and is associated a reduction of cardiovascular events and an improvement of angiogenic repair in ischemic limbs in patients with T2DM treated with this drug In mice, treatment with omentin prevents inflammation of the endothelium by blocking the activation of JNK and NF-κB induced by TNFα and reducing the expression of adhesion molecules
Omentin	↑/↓Serum levels are associated with CAD and PAD	↑Serum levels are related to the severity of PAD	
Resistin	↑Serum levels in CAD	↑Serum levels preprocedural in patients with restenosis after stenting	–
Visfatin	↑Serum levels in AMI	–	–
Ctrps	↑/↓Serum levels are associated with CAD and PAD	–	Some evidences in mice
Sortilin	↑Serum levels in CAD and in PAD	↑Serum levels are related to the severity of PAD	–
HMGB-1	↑Serum levels in CAD and in PAD	↑Serum levels are related to the severity of PAD	–
OPG	↑Serum levels in PAD	–	–
CRP	CRP promotes a major accuracy of risk estimation if used as complementary tool of Framingham score	↑CRP is related to worse vascular outcomes in diabetic PAD CRP is an independent predictor of amputation in diabetic patients	–
Pentraxin 3	↑Concentrations are detected in human carotid plaques and coronary atherosclerotic lesions during acute myocardial infarction and instable angina	Pentraxin 3 has a supposed role in vascular complications of diabetic patients	–
Interleukins	↑IL-18 is correlated with major incidence of diabetes mellitus and the vascular complications related to this disease ↑IL-6 correlates to a higher incidence of endovascular revascularization procedure failure	–	The administration of IL-1Ra improves glycemic control, reduces levels of CRP and decreases the incidence of cardiovascular disease Canakinumab allows a reduction in plasma concentrations of hs-CRP, IL-6 and fibrinogen, with possible benefits for patients who present a residual inflammatory risk; IL-1 inhibition treatment decreases the risks of overall MACE, unstable angina, and breakthrough or recurrence of heart failure;
FGF23	↑FGF23 and ↓circulating Klotho are markers of MALE, diabetic ulcer foot, mortality and cardiac and large vessels complications in pre-diabetes and diabetes	–	
Klotho	↓Klotho is related to coronary disease, heart failure, stroke and peripheral artery disease ↓Klotho is associated to a more severe grade of the coronary stenosis ↓Klotho is connected to a more intense severity of disease	–	Klotho is a protective factor against diabetic ulcer foot, it attenuates the progression of diabetic nephropathy, and it protects from macrovascular major adverse events (stroke and myocardial infarction) in patients affected by type 2 diabetes mellitus

Table 1 (continued)

	Predictive	Prognostic	Therapeutic
TNF- α	<p>↑TNF-α determines more incidence of failure of lower extremity endovascular revascularization procedures and a major occurrence of MACE and MALE</p> <p>Among diabetic patients, those who are affected by diabetic ulcer foot present a higher concentration of TNF-α</p>	–	Indeed, anti-TNF strategies have been thought to be beneficial in several inflammatory disease as atherosclerosis

pattern that might be directly dependent from diabetic vasculopathy and contribute to worsen its prognosis (Fig. 1; Table 1). These assays may be helpful in subjects with advanced disease to better assess their risk and possibly to guide therapeutic decisions. Their combined use with the assay of other molecules such as adipokines, klotho and FGF23 may help defining better risk prediction models and possibly personalizing the use of innovative strategies.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

Statement of human and animal rights The procedures employed to draft this manuscript respect the ethical standards in the Helsinki Declaration of 1975, as revised in 2000, as well as the national law.

Informed consent For this type of article, formal consent is not required.

Consent for publication All authors have read the paper and agree that it can be published.

References

1. Sarwar N, Gao P, Seshasai SR, Gobin R, Kaptoge S, Di Angelantonio E, Ingelsson E, Lawlor DA, Selvin E, Stampfer M, Stehouwer CD, Lewington S, Pennells L, Thompson A, Sattar N, White IR, Ray KK, Danesh J, Collaboration ERF (2010) Diabetes mellitus, fasting blood glucose concentration, and risk of vascular disease: a collaborative meta-analysis of 102 prospective studies. *Lancet* 375(9733):2215–2222. [https://doi.org/10.1016/S0140-6736\(10\)60484-9](https://doi.org/10.1016/S0140-6736(10)60484-9)
2. Sasso FC, Chiodini P, Carbonara O, De Nicola L, Conte G, Salvatore T, Nasti R, Marfella R, Gallo C, Signoriello S, Torella R, Minutolo R, Group NITDS (2012) High cardiovascular risk in patients with Type 2 diabetic nephropathy: the predictive role of albuminuria and glomerular filtration rate. *The NID-2 Prospective Cohort Study. Nephrol Dial Transplant* 27(6):2269–2274. <https://doi.org/10.1093/ndt/gfr644>
3. Minutolo R, Sasso FC, Chiodini P, Cianciaruso B, Carbonara O, Zamboli P, Tirino G, Pota A, Torella R, Conte G, De Nicola L (2006) Management of cardiovascular risk factors in advanced type 2 diabetic nephropathy: a comparative analysis in nephrology, diabetology and primary care settings. *J Hypertens* 24(8):1655–1661. <https://doi.org/10.1097/01.hjh.0000239303.93872.31>
4. Norris KC, Smoyer KE, Rolland C, Van der Vaart J, Grubb EB (2018) Albuminuria, serum creatinine, and estimated glomerular filtration rate as predictors of cardio-renal outcomes in patients with type 2 diabetes mellitus and kidney disease: a systematic literature review. *BMC Nephrol* 19(1):36. <https://doi.org/10.1186/s12882-018-0821-9>
5. Selvin E, Erlinger TP (2004) Prevalence of and risk factors for peripheral arterial disease in the United States: results from the National Health and Nutrition Examination Survey, 1999–2000.

- Circulation 110(6):738–743. <https://doi.org/10.1161/01.CIR.0000137913.26087.F0>
6. Selvin E, Marinopoulos S, Berkenblit G, Rami T, Brancati FL, Powe NR, Golden SH (2004) Meta-analysis: glycosylated hemoglobin and cardiovascular disease in diabetes mellitus. *Ann Intern Med* 141(6):421–431. <https://doi.org/10.7326/0003-4819-141-6-200409210-00007>
 7. Hirsch IB (2015) Glycemic variability and diabetes complications: does it matter? Of course it does! *Diabetes Care* 38(8):1610–1614. <https://doi.org/10.2337/dc14-2898>
 8. Lontchi-Yimagou E, Sobngwi E, Matsha TE, Kengne AP (2013) Diabetes mellitus and inflammation. *Curr Diab Rep* 13(3):435–444. <https://doi.org/10.1007/s11892-013-0375-y>
 9. Lu J, Marnell LL, Marjon KD, Mold C, Du Clos TW, Sun PD (2008) Structural recognition and functional activation of FcγR by innate pentraxins. *Nature* 456(7224):989–992. <https://doi.org/10.1038/nature07468>
 10. Biscetti F, Ferraro PM, Hiatt WR, Angelini F, Nardella E, Cecchini AL, Santoliquido A, Pitocco D, Landolfi R, Flex A (2019) Inflammatory cytokines associated with failure of lower extremity endovascular revascularization (LER): A Prospective Study of a Population with diabetes. *Diabetes Care*. <https://doi.org/10.2337/dc19-0408>
 11. Libby P (2017) Interleukin-1 beta as a target for atherosclerosis therapy: biological basis of CANTOS and beyond. *J Am Coll Cardiol* 70(18):2278–2289. <https://doi.org/10.1016/j.jacc.2017.09.028>
 12. Chen CH, Jiang T, Yang JH, Jiang W, Lu J, Marathe GK, Pownall HJ, Ballantyne CM, McIntyre TM, Henry PD, Yang CY (2003) Low-density lipoprotein in hypercholesterolemic human plasma induces vascular endothelial cell apoptosis by inhibiting fibroblast growth factor 2 transcription. *Circulation* 107(16):2102–2108. <https://doi.org/10.1161/01.CIR.0000065220.70220.F7>
 13. Stancel N, Chen CC, Ke LY, Chu CS, Lu J, Sawamura T, Chen CH (2016) Interplay between CRP, atherogenic LDL, and LOX-1 and Its potential role in the pathogenesis of atherosclerosis. *Clin Chem* 62(2):320–327. <https://doi.org/10.1373/clinchem.2015.243923>
 14. Balamir I, Ates I, Topcuoglu C, Turhan T (2018) Association of endocan, ischemia-modified albumin, and hscrp levels with endothelial dysfunction in type 2 diabetes mellitus. *Angiology* 69(7):609–616. <https://doi.org/10.1177/0003319717740781>
 15. Soeki T, Sata M (2016) Inflammatory biomarkers and atherosclerosis. *Int Heart J* 57(2):134–139. <https://doi.org/10.1536/ihj.15-346>
 16. Xu B, Yang CZ, Wu SB, Zhang D, Wang LN, Xiao L, Chen Y, Wang CR, Tong A, Zhou XF, Li XH, Guan XH (2017) Risk factors for lower extremity amputation in patients with diabetic foot. *Zhonghua Nei Ke Za Zhi* 56(1):24–28. <https://doi.org/10.3760/cma.j.issn.0578-1426.2017.01.007>
 17. Inoue K, Sugiyama A, Reid PC, Ito Y, Miyauchi K, Mukai S, Sagara M, Miyamoto K, Satoh H, Kohno I, Kurata T, Ota H, Mantovani A, Hamakubo T, Daida H, Kodama T (2007) Establishment of a high sensitivity plasma assay for human pentraxin3 as a marker for unstable angina pectoris. *Arterioscler Thromb Vasc Biol* 27(1):161–167. <https://doi.org/10.1161/01.ATV.0000052126.48375.d5>
 18. Mutlu M, Yuksel N, Takmaz T, Dincel AS, Bilgihan A, Altinkaynak H (2017) Aqueous humor pentraxin-3 levels in patients with diabetes mellitus. *Eye (Lond)* 31(10):1463–1467. <https://doi.org/10.1038/eye.2017.87>
 19. Ahmad J, Zubair M, Malik A, Siddiqui MA, Wangnoo SK (2012) Cathepsin-D, adiponectin, TNF-α, IL-6 and hsCRP plasma levels in subjects with diabetic foot and possible correlation with clinical variables: a multicentric study. *Foot (Edinb)* 22(3):194–199. <https://doi.org/10.1016/j.foot.2012.03.015>
 20. Silva LC, Ortigosa LC, Benard G (2010) Anti-TNF-α agents in the treatment of immune-mediated inflammatory diseases: mechanisms of action and pitfalls. *Immunotherapy* 2(6):817–833. <https://doi.org/10.2217/imt.10.67>
 21. Nidorf SM, Eikelboom JW, Budgeon CA, Thompson PL (2013) Low-dose colchicine for secondary prevention of cardiovascular disease. *J Am Coll Cardiol* 61(4):404–410. <https://doi.org/10.1016/j.jacc.2012.10.027>
 22. Banerjee M, Saxena M (2012) Interleukin-1 (IL-1) family of cytokines: role in type 2 diabetes. *Clin Chim Acta* 413(15–16):1163–1170. <https://doi.org/10.1016/j.cca.2012.03.021>
 23. Herder C, Dalmas E, Böni-Schnetzler M, Donath MY (2015) The IL-1 pathway in type 2 diabetes and cardiovascular complications. *Trends Endocrinol Metab* 26(10):551–563. <https://doi.org/10.1016/j.tem.2015.08.001>
 24. Dinarello CA, Ikejima T, Warner SJ, Orencole SF, Lonnemann G, Cannon JG, Libby P (1987) Interleukin 1 induces interleukin 1. I. Induction of circulating interleukin 1 in rabbits in vivo and in human mononuclear cells in vitro. *J Immunol* 139(6):1902–1910
 25. Suzuki K, Murtuza B, Smolenski RT, Sammut IA, Suzuki N, Kaneda Y, Yacoub MH (2001) Overexpression of interleukin-1 receptor antagonist provides cardioprotection against ischemia-reperfusion injury associated with reduction in apoptosis. *Circulation* 104(12 Suppl 1):I303–I308. <https://doi.org/10.1161/hc371.094871>
 26. Libby P, Warner SJ, Friedman GB (1988) Interleukin 1: a mitogen for human vascular smooth muscle cells that induces the release of growth-inhibitory prostanoids. *J Clin Invest* 81(2):487–498. <https://doi.org/10.1172/JCI113346>
 27. Schlesinger N, Alten RE, Bardin T, Schumacher HR, Bloch M, Gimona A, Krammer G, Murphy V, Richard D, So AK (2012) Canakinumab for acute gouty arthritis in patients with limited treatment options: results from two randomised, multicentre, active-controlled, double-blind trials and their initial extensions. *Ann Rheum Dis* 71(11):1839–1848. <https://doi.org/10.1136/annrheumdis-2011-200908>
 28. Zheng ZH, Zeng X, Nie XY, Cheng YJ, Liu J, Lin XX, Yao H, Ji CC, Chen XM, Jun F, Wu SH (2019) Interleukin-1 blockade treatment decreasing cardiovascular risk. *Clin Cardiol*. <https://doi.org/10.1002/clc.23246>
 29. Gosling J, Slaymaker S, Gu L, Tseng S, Zlot CH, Young SG, Rollins BJ, Charo IF (1999) MCP-1 deficiency reduces susceptibility to atherosclerosis in mice that overexpress human apolipoprotein B. *J Clin Invest* 103(6):773–778. <https://doi.org/10.1172/JCI5624>
 30. Panee J (2012) Monocyte chemoattractant protein 1 (MCP-1) in obesity and diabetes. *Cytokine* 60(1):1–12. <https://doi.org/10.1016/j.cyto.2012.06.018>
 31. Arakelyan A, Petrakova J, Hermanova Z, Boyajyan A, Lukl J, Petrek M (2005) Serum levels of the MCP-1 chemokine in patients with ischemic stroke and myocardial infarction. *Mediat Inflamm* 3:175–179. <https://doi.org/10.1155/MI.2005.175>
 32. Czemplik M, Kulma A, Wang YF, Szopa J (2017) Therapeutic strategies of plant-derived compounds for diabetes via regulation of monocyte chemoattractant protein-1. *Curr Med Chem* 24(14):1453–1468. <https://doi.org/10.2174/0929867324666170303162935>
 33. Ismail NA, Abd El Baky AN, Ragab S, Hamed M, Hashish MA, Shehata A (2016) Monocyte chemoattractant protein 1 and macrophage migration inhibitory factor in children with type 1 diabetes. *J Pediatr Endocrinol Metab* 29(6):641–645. <https://doi.org/10.1515/jpem-2015-0340>
 34. Hartman J, Frishman WH (2014) Inflammation and atherosclerosis: a review of the role of interleukin-6 in the development of atherosclerosis and the potential for targeted drug therapy. *Cardiol Rev* 22(3):147–151. <https://doi.org/10.1097/CRD.0000000000000021>

35. Mai J, Virtue A, Shen J, Wang H, Yang XF (2013) An evolving new paradigm: endothelial cells—conditional innate immune cells. *J Hematol Oncol* 6:61. <https://doi.org/10.1186/1756-8722-6-61>
36. Erlandsson Harris H, Andersson U (2004) Mini-review: the nuclear protein HMGB1 as a proinflammatory mediator. *Eur J Immunol* 34(6):1503–1512. <https://doi.org/10.1002/eji.200424916>
37. Kalinina N, Agrotis A, Antropova Y, DiVitto G, Kanellakis P, Kostolias G, Ilyinskaya O, Tararak E, Bobik A (2004) Increased expression of the DNA-binding cytokine HMGB1 in human atherosclerotic lesions: role of activated macrophages and cytokines. *Arterioscler Thromb Vasc Biol* 24(12):2320–2325. <https://doi.org/10.1161/01.ATV.0000145573.36113.8a>
38. Biscetti F, Straface G, De Cristofaro R, Lancellotti S, Rizzo P, Arena V, Stigliano E, Pecorini G, Egashira K, De Angelis G, Ghirlanda G, Flex A (2010) High-mobility group box-1 protein promotes angiogenesis after peripheral ischemia in diabetic mice through a VEGF-dependent mechanism. *Diabetes* 59(6):1496–1505. <https://doi.org/10.2337/db09-1507>
39. Oozawa S, Sano S, Nishibori M (2014) Usefulness of high mobility group box 1 protein as a plasma biomarker in patient with peripheral artery disease. *Acta Med Okayama* 68(3):157–162. <https://doi.org/10.18926/AMO/52656>
40. Biscetti F, Flex A, Alivernini S, Toluoso B, Gremese E, Ferraccioli G (2017) The role of high-mobility group box-1 and its cross-talk with microbiome in rheumatoid arthritis. *Mediat Inflamm* 2017:5230374. <https://doi.org/10.1155/2017/5230374>
41. Yang J, Shah R, Robling AG, Templeton E, Yang H, Tracey KJ, Bidwell JP (2008) HMGB1 is a bone-active cytokine. *J Cell Physiol* 214(3):730–739. <https://doi.org/10.1002/jcp.21268>
42. Hajer GR, van Haeften TW, Visseren FL (2008) Adipose tissue dysfunction in obesity, diabetes, and vascular diseases. *Eur Heart J* 29(24):2959–2971. <https://doi.org/10.1093/eurheartj/ehn387>
43. Vionnet N, Hani EH, Dupont S, Gallina S, Francke S, Dotte S, De Matos F, Durand E, Leprêtre F, Lecoœur C, Gallina P, Zekiri L, Dina C, Froguel P (2000) Genomewide search for type 2 diabetes-susceptibility genes in French whites: evidence for a novel susceptibility locus for early-onset diabetes on chromosome 3q27-qter and independent replication of a type 2-diabetes locus on chromosome 1q21-q24. *Am J Hum Genet* 67(6):1470–1480. <https://doi.org/10.1086/316887>
44. Pilz S, Horejsi R, Möller R, Almer G, Scharnagl H, Stojakovic T, Dimitrova R, Weihrauch G, Borkenstein M, Maerz W, Schauenstein K, Mangge H (2005) Early atherosclerosis in obese juveniles is associated with low serum levels of adiponectin. *J Clin Endocrinol Metab* 90(8):4792–4796. <https://doi.org/10.1210/jc.2005-0167>
45. Sasso FC, Pafundi PC, Marfella R, Calabrò P, Piscione F, Furbatto F, Esposito G, Galiero R, Gragnano F, Rinaldi L, Salvatore T, D'Amico M, Adinolfi LE, Sardu C (2019) Adiponectin and insulin resistance are related to restenosis and overall new PCI in subjects with normal glucose tolerance: the prospective AIRE Study. *Cardiovasc Diabetol* 18(1):24. <https://doi.org/10.1186/s12933-019-0826-0>
46. Ezenwaka CE, Kalloo R (2005) Caribbean female patients with type 2 diabetes mellitus have lower serum levels of adiponectin than nondiabetic subjects. *Neth J Med* 63(2):64–69
47. Al-Daghri NM, Al-Attas OS, Alokail MS, Alkharfy KM, Husain T (2011) Adiponectin gene variants and the risk of coronary artery disease in patients with type 2 diabetes. *Mol Biol Rep* 38(6):3703–3708. <https://doi.org/10.1007/s11033-010-0484-5>
48. Hung WC, Wang CP, Lu LF, Yu TH, Chiu CA, Chung FM, Chen HJ, Houg JY, Shin SJ, Lee YJ (2010) Circulating adiponectin level is associated with major adverse cardiovascular events in type 2 diabetic patients with coronary artery disease. *Endocr J* 57(9):793–802
49. Xiang K, Wang Y, Zheng T, Jia W, Li J, Chen L, Shen K, Wu S, Lin X, Zhang G, Wang C, Wang S, Lu H, Fang Q, Shi Y, Zhang R, Xu J, Weng Q (2004) Genome-wide search for type 2 diabetes/ impaired glucose homeostasis susceptibility genes in the Chinese: significant linkage to chromosome 6q21-q23 and chromosome 1q21-q24. *Diabetes* 53(1):228–234
50. Watanabe K, Watanabe R, Konii H, Shirai R, Sato K, Matsuyama TA, Ishibashi-Ueda H, Koba S, Kobayashi Y, Hirano T, Watanabe T (2016) Counteractive effects of omentin-1 against atherogenesis. *Cardiovasc Res* 110(1):118–128. <https://doi.org/10.1093/cvr/cvw016>
51. Yamawaki H (2011) Vascular effects of novel adipocytokines: focus on vascular contractility and inflammatory responses. *Biol Pharm Bull* 34(3):307–310
52. Biscetti F, Nardella E, Bonadia N, Angelini F, Pitocco D, Santoliquido A, Filipponi M, Landolfi R, Flex A (2019) Association between plasma omentin-1 levels in type 2 diabetic patients and peripheral artery disease. *Cardiovasc Diabetol* 18(1):74. <https://doi.org/10.1186/s12933-019-0880-7>
53. Menzel J, di Giuseppe R, Biemann R, Wittenbecher C, Aleksandrova K, Pischon T, Fritsche A, Schulze MB, Boeing H, Isermann B, Weikert C (2016) Omentin-1 and risk of myocardial infarction and stroke: Results from the EPIC-Potsdam cohort study. *Atherosclerosis* 251:415–421. <https://doi.org/10.1016/j.atherosclerosis.2016.06.003>
54. Verma S, Li SH, Wang CH, Fedak PW, Li RK, Weisel RD, Mickle DA (2003) Resistin promotes endothelial cell activation: further evidence of adipokine-endothelial interaction. *Circulation* 108(6):736–740. <https://doi.org/10.1161/01.CIR.0000084503.91330.49>
55. Degawa-Yamauchi M, Bovenkerk JE, Juliar BE, Watson W, Kerr K, Jones R, Zhu Q, Considine RV (2003) Serum resistin (FIZZ3) protein is increased in obese humans. *J Clin Endocrinol Metab* 88(11):5452–5455. <https://doi.org/10.1210/jc.2002-021808>
56. On YK, Park HK, Hyon MS, Jeon ES (2007) Serum resistin as a biological marker for coronary artery disease and restenosis in type 2 diabetic patients. *Circ J* 71(6):868–873. <https://doi.org/10.1253/circj.71.868>
57. Calabro P, Samudio I, Willerson JT, Yeh ET (2004) Resistin promotes smooth muscle cell proliferation through activation of extracellular signal-regulated kinase 1/2 and phosphatidylinositol 3-kinase pathways. *Circulation* 110(21):3335–3340. <https://doi.org/10.1161/01.CIR.0000147825.97879.E7>
58. Chen MP, Chung FM, Chang DM, Tsai JC, Huang HF, Shin SJ, Lee YJ (2006) Elevated plasma level of visfatin/pre-B cell colony-enhancing factor in patients with type 2 diabetes mellitus. *J Clin Endocrinol Metab* 91(1):295–299. <https://doi.org/10.1210/jc.2005-1475>
59. Adya R, Tan BK, Chen J, Randeve HS (2008) Nuclear factor-kappaB induction by visfatin in human vascular endothelial cells: its role in MMP-2/9 production and activation. *Diabetes Care* 31(4):758–760. <https://doi.org/10.2337/dc07-1544>
60. Mazaherioun M, Hosseinzadeh-Attar MJ, Janani L, Vasheghani Farahani A, Rezvan N, Karbaschian Z, Hossein-Nezhad A (2012) Elevated serum visfatin levels in patients with acute myocardial infarction. *Arch Iran Med* 15(11):688–692
61. Patel KM, Strong A, Tohyama J, Jin X, Morales CR, Billheimer J, Millar J, Kruth H, Rader DJ (2015) Macrophage sortilin promotes LDL uptake, foam cell formation, and atherosclerosis. *Circ Res* 116(5):789–796. <https://doi.org/10.1161/CIRCRESAHA.116.305811>
62. Ogawa K, Ueno T, Iwasaki T, Kujiraoka T, Ishihara M, Kunitomo S, Takayama T, Kanai T, Hirayama A, Hattori H (2016) Soluble sortilin is released by activated platelets and its circulating levels are associated with cardiovascular risk factors.

- Atherosclerosis 249:110–115. <https://doi.org/10.1016/j.atherosclerosis.2016.03.041>
63. Goettsch C, Kjolby M, Aikawa E (2018) Sortilin and its multiple roles in cardiovascular and metabolic diseases. *Arterioscler Thromb Vasc Biol* 38(1):19–25. <https://doi.org/10.1161/ATVBAHA.117.3.10292>
 64. Oh TJ, Ahn CH, Kim BR, Kim KM, Moon JH, Lim S, Park KS, Lim C, Jang H, Choi SH (2017) Circulating sortilin level as a potential biomarker for coronary atherosclerosis and diabetes mellitus. *Cardiovasc Diabetol* 16(1):92. <https://doi.org/10.1186/s12933-017-0568-9>
 65. Biscetti F, Bonadia N, Santini F, Angelini F, Nardella E, Pitocco D, Santoliquido A, Filippini M, Landolfi R, Flex A (2019) Sortilin levels are associated with peripheral arterial disease in type 2 diabetic subjects. *Cardiovasc Diabetol* 18(1):5. <https://doi.org/10.1186/s12933-019-0805-5>
 66. Biscetti F, Straface G, Pitocco D, Angelini F, Tinelli G, Landolfi R, Flex A (2016) Fibroblast growth factor 23 serum level in type 2 diabetic Italian subjects with peripheral arterial disease and critical limb ischemia. *Eur Rev Med Pharmacol Sci* 20(19):4048–4054
 67. Kiechl S, Schett G, Wenning G, Redlich K, Oberhollenzer M, Mayr A, Santer P, Smolen J, Poewe W, Willeit J (2004) Osteoprotegerin is a risk factor for progressive atherosclerosis and cardiovascular disease. *Circulation* 109(18):2175–2180. <https://doi.org/10.1161/01.CIR.0000127957.43874.BB>
 68. Fang L, Li X (2016) Level of serum phosphorus and adult type 2 diabetes mellitus. *Zhong Nan Da Xue Xue Bao Yi Xue Ban* 41(5):502–506. <https://doi.org/10.11817/j.issn.1672-7347.2016.05.009>
 69. Razzaque MS (2009) The FGF23-Klotho axis: endocrine regulation of phosphate homeostasis. *Nat Rev Endocrinol* 5(11):611–619. <https://doi.org/10.1038/nrendo.2009.196>
 70. Rennenberg RJ, Kessels AG, Schurgers LJ, van Engelshoven JM, de Leeuw PW, Kroon AA (2009) Vascular calcifications as a marker of increased cardiovascular risk: a meta-analysis. *Vasc Health Risk Manag* 5(1):185–197. <https://doi.org/10.2147/vhrm.s4822>
 71. Starup-Linde J, Vestergaard P (2016) Biochemical bone turnover markers in diabetes mellitus—a systematic review. *Bone* 82:69–78. <https://doi.org/10.1016/j.bone.2015.02.019>
 72. Mary A, Hartemann A, Brazier M, Aubert CE, Kemel S, Salem JE, Cluzel P, Liabeuf S, Massy Z, Mentaverri R, Bourron O, Kamel S (2018) Higher parathyroid hormone levels are associated with increased below-the-knee arterial calcification in type 2 diabetes. *Diabetes Metab* 44(3):305–308. <https://doi.org/10.1016/j.diabet.2017.04.008>
 73. Freedman BI, Divers J, Russell GB, Palmer ND, Bowden DW, Carr JJ, Wagenknecht LE, Hightower RC, Xu J, Smith SC, Langefeld CD, Hruska KA, Register TC (2015) Plasma FGF23 and calcified atherosclerotic plaque in African Americans with type 2 diabetes mellitus. *Am J Nephrol* 42(6):391–401. <https://doi.org/10.1159/000443241>
 74. Berezin AE, Berezin AA (2019) Impaired function of fibroblast growth factor 23/Klotho protein axis in prediabetes and diabetes mellitus: promising predictor of cardiovascular risk. *Diabetes Metab Syndr* 13(4):2549–2556. <https://doi.org/10.1016/j.dsx.2019.07.018>
 75. Flotyńska J, Uruska A, Araszkiwicz A, Zozulińska-Ziółkiewicz D (2018) Klotho protein function among patients with type 1 diabetes. *Endokrynol Pol* 69(6):696–704. <https://doi.org/10.5603/EP.a2018.0070>
 76. Pan HC, Chou KM, Lee CC, Yang NI, Sun CY (2018) Circulating Klotho levels can predict long-term macrovascular outcomes in type 2 diabetic patients. *Atherosclerosis* 276:83–90. <https://doi.org/10.1016/j.atherosclerosis.2018.07.006>
 77. Martín-Núñez E, Donate-Correa J, López-Castillo Á, Delgado-Molinos A, Ferri C, Rodríguez-Ramos S, Cerro P, Pérez-Delgado N, Castro V, Hernández-Carballo C, Mora-Fernández C, Navarro-González JF (2017) Soluble levels and endogenous vascular gene expression of. *Clin Sci (Lond)* 131(21):2601–2609. <https://doi.org/10.1042/CS20171242>
 78. Six I, Okazaki H, Gross P, Cagnard J, Boudot C, Maizel J, Druke TB, Massy ZA (2014) Direct, acute effects of Klotho and FGF23 on vascular smooth muscle and endothelium. *PLoS ONE* 9(4):e93423. <https://doi.org/10.1371/journal.pone.0093423>
 79. Olejnik A, Franczak A, Krzywonos-Zawadzka A, Kałużna-Oleksy M, Bil-Lula I (2018) The biological role of klotho protein in the development of cardiovascular diseases. *Biomed Res Int* 2018:5171945. <https://doi.org/10.1155/2018/5171945>
 80. Semba RD, Cappola AR, Sun K, Bandinelli S, Dalal M, Crasto C, Guralnik JM, Ferrucci L (2011) Plasma klotho and cardiovascular disease in adults. *J Am Geriatr Soc* 59(9):1596–1601. <https://doi.org/10.1111/j.1532-5415.2011.03558.x>
 81. Loyer X, Zlatanova I, Devue C, Yin M, Howangyin KY, Klaihmou P, Guerin CL, Kheloufi M, Vilar J, Zannis K, Fleischmann BK, Hwang DW, Park J, Lee H, Menasché P, Silvestre JS, Boulanger CM (2018) Intra-cardiac release of extracellular vesicles shapes inflammation following myocardial infarction. *Circ Res* 123(1):100–106. <https://doi.org/10.1161/CIRCRESAHA.117.311326>
 82. Sluijter JPG, Davidson SM, Boulanger CM, Buzás EI, de Kleijn DPV, Engel FB, Giricz Z, Hausenloy DJ, Kishore R, Lecour S, Leor J, Madonna R, Perrino C, Prunier F, Sahoo S, Schiffelers RM, Schulz R, Van Laake LW, Ytrehus K, Ferdinandy P (2018) Extracellular vesicles in diagnostics and therapy of the ischaemic heart: Position Paper from the Working Group on Cellular Biology of the Heart of the European Society of Cardiology. *Cardiovasc Res* 114(1):19–34. <https://doi.org/10.1093/cvr/cvx211>
 83. Boulanger CM, Loyer X, Rautou PE, Amabile N (2017) Extracellular vesicles in coronary artery disease. *Nat Rev Cardiol* 14(5):259–272. <https://doi.org/10.1038/nrcardio.2017.7>
 84. Bellin G, Gardin C, Ferroni L, Chachques JC, Rogante M, Mitrečić D, Ferrari R, Zavan B (2019) Exosome in cardiovascular diseases: a complex world full of hope. *Cells*. <https://doi.org/10.3390/cells8020166>
 85. Harris TA, Yamakuchi M, Ferlito M, Mendell JT, Lowenstein CJ (2008) MicroRNA-126 regulates endothelial expression of vascular cell adhesion molecule 1. *Proc Natl Acad Sci USA* 105(5):1516–1521. <https://doi.org/10.1073/pnas.0707493105>
 86. Soeki T, Yamaguchi K, Niki T, Uematsu E, Bando S, Matsuura T, Ise T, Kusunose K, Hotchi J, Tobiume T, Yagi S, Fukuda D, Taketani Y, Iwase T, Yamada H, Wakatsuki T, Shimabukuro M, Sata M (2015) Plasma microRNA-100 is associated with coronary plaque vulnerability. *Circ J* 79(2):413–418. <https://doi.org/10.1253/circj.CJ-14-0958>
 87. Witkowski M, Weithauser A, Tabaraie T, Steffens D, Kränkel N, Stratmann B, Tschoepe D, Landmesser U, Rauch-Kroehnert U (2016) Micro-RNA-126 reduces the blood thrombogenicity in diabetes mellitus via targeting of tissue factor. *Arterioscler Thromb Vasc Biol* 36(6):1263–1271. <https://doi.org/10.1161/ATVBAHA.115.306094>
 88. Lalla E, Papananou PN (2011) Diabetes mellitus and periodontitis: a tale of two common interrelated diseases. *Nat Rev Endocrinol* 7(12):738–748. <https://doi.org/10.1038/nrendo.2011.106>
 89. Mascarenhas JV, Albayati MA, Shearman CP, Jude EB (2014) Peripheral arterial disease. *Endocrinol Metab Clin North Am* 43(1):149–166. <https://doi.org/10.1016/j.ecl.2013.09.003>
 90. Silvestro A, Scopacasa F, Ruocco A, Oliva G, Schiano V, Zincarelli C, Brevetti G (2003) Inflammatory status and endothelial function in asymptomatic and symptomatic peripheral arterial

- disease. *Vasc Med* 8(4):225–232. <https://doi.org/10.1191/1358863x03vm503oa>
91. Hirabara SM, Gorjão R, Vinolo MA, Rodrigues AC, Nachbar RT, Curi R (2012) Molecular targets related to inflammation and insulin resistance and potential interventions. *J Biomed Biotechnol* 2012:379024. <https://doi.org/10.1155/2012/379024>
92. Rizza S, Cardellini M, Porzio O, Pecchioli C, Savo A, Cardolini I, Senese N, Lauro D, Sbraccia P, Lauro R, Federici M (2011) Pioglitazone improves endothelial and adipose tissue dysfunction in pre-diabetic CAD subjects. *Atherosclerosis* 215(1):180–183. <https://doi.org/10.1016/j.atherosclerosis.2010.12.021>
93. Dormandy JA, Charbonnel B, Eckland DJ, Erdmann E, Massi-Benedetti M, Moules IK, Skene AM, Tan MH, Lefèbvre PJ, Murray GD, Standl E, Wilcox RG, Wilhelmsen L, Betteridge J, Birkeland K, Gøtzsche A, Heine RJ, Korányi L, Laakso M, Mokán M, Norkus A, Pirags V, Podar T, Scheen A, Scherbaum W, Schernthaner G, Schmitz O, Skrha J, Smith U, Taton J, Investigators P (2005) Secondary prevention of macrovascular events in patients with type 2 diabetes in the PROactive Study (PROspective pioglitazone Clinical Trial In macroVascular Events): a randomised controlled trial. *Lancet* 366(9493):1279–1289. [https://doi.org/10.1016/S0140-6736\(05\)67528-9](https://doi.org/10.1016/S0140-6736(05)67528-9)
94. Rajamani K, Colman PG, Li LP, Best JD, Voysey M, D’Emden MC, Laakso M, Baker JR, Keech AC, Investigators Fs (2009) Effect of fenofibrate on amputation events in people with type 2 diabetes mellitus (FIELD study): a prespecified analysis of a randomised controlled trial. *Lancet* 373(9677):1780–1788. [https://doi.org/10.1016/S0140-6736\(09\)60698-X](https://doi.org/10.1016/S0140-6736(09)60698-X)
95. Bertrand MJ, Tardif JC (2017) Inflammation and beyond: new directions and emerging drugs for treating atherosclerosis. *Expert Opin Emerg Drugs* 22(1):1–26. <https://doi.org/10.1080/14728214.2017.1269743>
96. Bäck M, Hansson GK (2015) Anti-inflammatory therapies for atherosclerosis. *Nat Rev Cardiol* 12(4):199–211. <https://doi.org/10.1038/nrcardio.2015.5>

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