ORIGINAL ARTICLE

Qualities and antioxidant activities of lotus leaf afected by diferent drying methods

Chunyan Guo¹ · Nan Zhang1 · Cunqi Liu1 · Jingjing Xue1 · Jianzhou Chu1 · Xiaoqin Yao1

Received: 18 September 2018 / Revised: 10 December 2019 / Accepted: 17 December 2019 / Published online: 10 January 2020 © Franciszek Górski Institute of Plant Physiology, Polish Academy of Sciences, Kraków 2020

Abstract

Drying treatments are an efective method for preserving original benefcial ingredients of postharvest plants. This work supplied the suitable drying conditions of three drying methods by investigating the changes of lotus leaves' quality and antioxidant activity. The results indicated that: (1) Oven drying (55–65 °C) was the most appropriate way to keep the higher contents of alkaloids and amino acids, and the higher activities in DPPH radical scavenging and hydroxyl radical scavenging in lotus leaf treated with three drying methods; (2) Steam kill-enzyme torrefaction (3–5 min) was the most appropriate method for keeping the higher contents of favone and the higher activities of superoxide radical scavenging in lotus leaf treated with three drying methods; (3) Microwave drying (680–850 W) was the most appropriate means to keep the higher contents of vitamin C, soluble sugar and protein in lotus leaf treated with three drying methods; (4) The color of lotus leaf treated with microwave treatments was more bright green than that treated with the other two methods; Therefore, the actual conditions of each drying means have to be taken into consideration when use the drying means.

Keywords Lotus leaf · Drying methods · Alkaloid · Radical scavenging activity

Introduction

Lotus (*Nelumbo nucifera* Gaertn.) is usually served as an economic ornamental, food, and pharmacy purposes, and generally grown in Eastern Asia, China, and India (Hu et al. [2012;](#page-6-0) Kato et al. [2015\)](#page-6-1). The cultivation and medicinal use of lotus were discovered in China 2000 years ago (Zhao et al. [2013\)](#page-7-0). Lotus leaves have been widely used as tonic medicine in Qin and Han dynasties. In November 1991, lotus leaves were included in the second batch of "Food and Drugs" list (Document No. 45 of the Health Supervision Bureau of the Ministry of Health of the People's Republic of China).

Tea is brewed with a portion of the organs of medicinal plants, which has been popular in the world for thousands of years (Mukhtar and Ahmad [2000\)](#page-7-1). Lotus leaves are rich

Communicated by P. Wojtaszek.

 \boxtimes Jianzhou Chu cjz01me@126.com

 \boxtimes Xiaoqin Yao yaoxiao301@126.com

¹ The College of Life Sciences, Hebei University, Baoding 071002, China

in many active and nutritional ingredients, such as favone, amino acids, vitamin C and so on (Huang and Zhang [2008](#page-6-2)). Additionally, many pharmacological and physiological activities have been confrmed in lotus leaf tea, such as antiobesity, anti-oxidant, antiviral, anti-hypertensive, and so on (Ono et al. [2006](#page-7-2); Liao and Lin [2013](#page-6-3); Kato et al. [2015](#page-6-1)). The lotus leaf resources are abundant, and the price is low (Jin et al. [2016\)](#page-6-4). Therefore, the lotus leaf tea has a broad development prospect. Currently, there are not many beverages related to lotus leaf on the market, and the processing methods reported and the product qualities obtained are diferent (Hao et al. [2012\)](#page-6-5). Therefore, this article systematically studied the diferent processing techniques of lotus leaf tea and optimized the technical parameters of each process, which were able to enhance active and nutritional ingredients of lotus leaf tea.

Drying can be used to quickly protect the medicinal qualities of the medicinal plants in a simple manner, which is considered the most common and basic step needed for postharvest plants (Zhang et al. [2015;](#page-7-3) Yuan et al. [2015](#page-7-4)). It can be seen from former work, most researches on drying methods have been mainly conducted on medicinal plants, fruits, and vegetables. The aims of this study were to probe the changing rules of qualities and antioxidant activities of

lotus leaf with three drying means and determine the most suitable drying method that simultaneously conserve the higher qualities and antioxidant activities in lotus leaves. Based on these, the results of this study will be helpful to develop lotus leaf tea as a healthy beverage.

Materials and methods

Equipment

Ultralow temperature refrigerated centrifuge (Thermo, Germany), Electronic balance (Sartorius, China), UV spectrophotometer (Unico, China), Electric blast drying oven (Teste, China), Microwave oven (Galanz, China), waterbath (Shengweili, China) and Ultrasonic cleaner (Kangjie, China).

Plant material and experimental design

Plant material

Lotus leaves were collected from Bai-yang Lake, Hebei province, China (38° 43′–39° 02′ N; 115° 38′-116° 07′ E), during September 2018. The lotus leaves were treated with three drying means.

Drying methods

(1) Oven drying was conducted in an electric blast drying oven at three different temperatures (55 \degree C, 65 \degree C, and 75 °C). Dry time was about 24 h; (2) Steam kill-enzyme torrefaction was carried out in a pot flled with boiling water in three different kill-enzyme times of $1 (Z1)$, $3 (Z2)$ and 5 min (Z3). The samples after kill-enzyme torrefaction were dried in electric blast drying oven at 65 °C. Drying time was about 24 h; and (3) Microwave drying was conducted in a microwave oven in three different power levels of 50% (450 W1), 80% (650 W2) and 100% (850 W3). Drying time was about 24 min, 6 min, and 3 min at W1, W2 and W3 treatments, separately. The eventual water contents were approximately $15 \pm 0.73\%$ dry basis in each of drying methods (Jafari et al. [2016](#page-6-6)).

After the above processes, the samples were ground into a powder and conserved in polyethylene bags.

Measurement methods

Flavone and alkaloids content

Flavone content was determined according to the method described by He and Liu (2007) . Sample (0.2 g) was extracted with ethanol solution (20 mL, 50%) in an ultrasonic bath. The reaction mixture involved extract solution (5 mL), $AICl₃$ (8 mL, 1.5%), acetic acid-sodium acetate (4 mL). The absorption values were measured at 420 nm.

Alkaloids content was determined according to the method described by Wang et al. ([2005](#page-7-5)). Sample (0.2 g) was extracted with trichloromethane solution (5 mL) and ammonium hydroxide (1 mL). The reaction mixture contained extract solution (2 mL), trichloromethane solution (8 mL), citric acid-sodium citrate (10 mL) and bromothymol blue (1 mL). The absorption values were detected at 416 nm.

Soluble sugar, free amino acids, vitamin C and protein content

Soluble sugar was measured according to the method described by Machado et al. (2013) (2013) . Sample (0.2 g) was extracted with distilled water (5 mL). The reaction mixture included supernatant (1 mL), distilled water (1 mL), and anthrone- H_2SO_4 (5 mL). The absorption values were detected at 625 nm.

Free amino acids content was measured with minor modifcation according to the method described by Li et al. ([2004\)](#page-6-9). Sample (0.2 g) was extracted with acetic acid solution (8 mL, 10%). The reaction mixture was mixed with supernatant (2 mL), and ninhydrin (3 mL), ascorbic acid (0.1 mL). The absorption values were detected at 570 nm.

Vitamin C was determined according to State Standard of the People's Republic of China ([1990\)](#page-7-6) (GB 12392-90). Sample (0.2 g) was extracted with oxalic acid solution (10 mL, 1%) and fltered with active carbon (0.2 g). The reaction mixture was mixed with supernatant (5 mL), thiourea solution (5 mL, 2%), 2, 4-dinitrophenylhydrazine solution (1 mL, 2%) and were cultivated in the water bath (37 °C) for 3 h. The absorption values were detected at 500 nm.

Protein content was measured according to the Coomassie brilliant blue staining created by Bradford ([1976](#page-6-10)). Sample (0.2 g) was extracted with 5 mL phosphate bufer (50 mmol L⁻¹) involving EDTA (1 mmol L⁻¹), 2% (W/V) polyvinyl pyrrolidone. The reaction mixture involved extract solution (0.1 mL) and Coomassie brilliant blue (5 mL). The absorption values were detected at 595 nm.

Determination of antioxidant capacity

The DPPH free radical scavenging activity was measured according to the previously described method (Mohsen and Ammar [2009](#page-7-7)). Sample extract solution (2 mL) was mixed with DPPH solution (2 mL, 0.04 mg mL⁻¹, prepared in 100% ethanol) for 0.5 h. The absorption values were detected at 517 nm, and the calculation formula as follows:

Scavenging activity (%) = $(A_0 - A_1 + A_2)/A_0 \times 100\%$

 A_0 is the absorption value of the control (the ethanol replaced the sample solution); A_1 is the absorption value of

the sample; A_2 is the absorption value of the sample under similar conditions as A_1 with the ethanol replaced the DPPH solution.

The superoxide radical scavenging activity was assayed using the modifed pyrogallol autoxidation method (Chen et al. [2012\)](#page-6-11). Sample extract solution (1 mL) was mixed with Tris–HCl (4 mL, 0.05 mol L^{-1} , pH 8.2) and pyrogallol $(1 \text{ mL}, 0.2 \text{ mmol L}^{-1})$ in a water bath (25 °C) for 5 min. The reaction was stopped immediately with two drops of concentrated HCl. The absorption values were detected at 325 nm, and the calculation formula was as follows:

Scavenging activity $(\%) = (A_0 - A_1)/A_0 \times 100\%$

 A_0 is the absorption value of the control; A_1 is the absorption value of the sample.

The hydroxyl radical scavenging activity was measured according to the improved phenanthroline method (You et al. [2011](#page-7-8)). The mixtures were mixed with 1, 10-phenanthroline $(1 \text{ mL}, 0.75 \text{ mmol})$, phosphate buffer $(2 \text{ mL}, 0.2 \text{ mol } L^{-1})$, pH 7.4), sample extract solution (1 mL) , FeSO₄ (1 mL) , 0.75 mM) and H_2O_2 (1 mL, 0.01%) in a water bath (37 °C) for 1 h. The absorption values were detected at 536 nm, and the calculation formula was as follows:

Scavenging activity $(\%) = (A_0 - A_1)/(A_2 - A_1) \times 100\%$

 A_0 is the absorption value of the sample; A_1 is the absorption value of the control (the water replaced the sample); A_2 is the absorption value of the solvent.

Statistical analysis

Data were the mean \pm standard error. Differences among treatments were analyzed using one-way analysis of variance with Duncan's test $(p < 005)$. All analyses were performed by SPSS (version 13.0).

Results and discussion

The efects of drying methods on favone and alkaloids in lotus leaf

Flavone has various biological and medicinal properties, for example, anti-infammatory, anti-cancer (Manthey et al. [2001](#page-7-9)) and anti-obesity agent (Kim et al. [2010](#page-6-12)). The experimental study showed that favone content in lotus leaf was obviously raised as the enhance of stem kill-enzyme time (Fig. [1](#page-2-0)). The results were similar to Shi et al. ([2017\)](#page-7-10). In oven drying, favone content rose frst and then reduced as the enhance of the oven temperature. In the three drying methods, the lotus leaf processed with 5 min steam kill-enzyme exhibited the highest flavonoid content (2.537 mg g^{-1} DW), followed by the lotus leaf treated with the higher microwave power (2.337 mg g^{-1} DW) and lotus leaf dried by oven (65 °C) (1.836 mg g^{-1} DW).

Fig. 1 The favone content with three drying methods in lotus leaves. Oven drying (H1, 55 °C; H2, 65 °C; H3, 75 °C), steam kill-enzyme torrefaction (Z1, 1 min; Z2, 3 min; Z3, 5 min) and microwave drying (W1, 450 W; W2, 650 W; W3, 850 W). The bars with diferent letters are obviously diferent from each other at the same drying method (p <0.05). Values are means of four replicates \pm SE

Fig. 2 The alkaloids content with three drying methods in lotus leaves. Oven drying (H1, 55 °C; H2, 65 °C; H3, 75 °C), steam killenzyme torrefaction (Z1, 1 min; Z2, 3 min; Z3, 5 min) and microwave drying (W1, 450 W; W2, 650 W; W3, 850 W). The bars with diferent letters are obviously diferent from each other at the same drying method ($p < 0.05$). Values are means of four replicates \pm SE

Many studies have shown that the lotus leaf has a signifcant lipid-regulating efect, and its active site is mainly alkaloids (Du et al. [2000\)](#page-6-13). In the current research, alkaloids content signifcantly increased as the enhance of microwave power (Fig. [2\)](#page-2-1). Similar results have appeared in previous research (Zhang et al. [2011](#page-7-11)). In oven drying, alkaloids content in lotus leaf dropped frst and rose subsequently as the enhance of oven temperature, and achieved a maximum content when oven temperature was 75 °C. However, the alkaloids content in lotus leaf signifcantly reduced with the enhance of the steam kill-enzyme time. Oven drying was benefcial to preserving the content of alkaloids in lotus leaves among the three drying methods.

The efects of drying methods on the contents of soluble sugar, amino acids, vitamin C and protein in lotus leaf

In recent years, bio-polysaccharides have been proven to be anti-tumor, anti-viral, anti-aging, enhancing immunity, and hematopoietic properties (Khaskheli et al. [2015](#page-6-14); Hammi et al. [2016](#page-6-15)). At the same time, water-soluble sugar cannot only taste sweet alcohol in the tea feld but also can produce some aroma substances and colored substances through Maillard reaction and caramelization in the process, which afected the tea quality. Dehnad et al. [\(2016\)](#page-6-16) certifed that the drying method has a signifcant efect on the polysaccharide content. Figure [3](#page-3-0) indicated that the soluble sugar content in lotus leaf was afected by the actual conditions of each drying method. In the three drying means, higher microwave power (650–850 W) was beneficial for preserving soluble sugar content (1.550 and 1.630 mg g^{-1} DW) in lotus leaves, followed by lotus leaf treated by steam killenzyme (1.450 mg g^{-1} DW) and lotus leaf dried by oven $(65–75 °C)$ (1.253 and 1.258 mg g⁻¹ DW).

Free amino acids are a crucial biologically active compound that has been shown specifc functions in maintaining metabolism, growth, reproduction, and immune function (Cylwik et al. [2005;](#page-6-17) Morris et al. [2005](#page-7-12); Mateo et al. [2007](#page-7-13)). Both the drying methods and the actual drying conditions will affect the content of free amino acids in the lotus leaf (Fig. [4\)](#page-3-1). The present study showed amino acids in lotus leaf was obviously dropped while the enhance of the oven temperature (55–75 °C) and achieved a maximum content (1019 μg g^{-1} DW) while oven temperature was 55 °C. However, amino acids in lotus leaf increased frstly and then declined while the enhance of steam kill-enzyme time and microwave power, and achieved the maximum content (596

Fig. 3 The soluble sugar content with three drying methods in lotus leaves. Oven drying (H1, 55 °C; H2, 65 °C; H3, 75 °C), steam killenzyme torrefaction (Z1, 1 min; Z2, 3 min; Z3, 5 min) and microwave drying (W1, 450 W; W2, 650 W; W3, 850 W). The bars with diferent letters are obviously diferent from each other at the same drying method ($p < 0.05$). Values are means of four replicates \pm SE

Fig. 4 The amino acids content with three drying methods in lotus leaves. Oven drying (H1, 55 °C; H2, 65 °C; H3, 75 °C), steam killenzyme torrefaction (Z1, 1 min; Z2, 3 min; Z3, 5 min) and microwave drying (W1, 450 W; W2, 650 W; W3, 850 W). The bars with diferent letters are obviously diferent from each other at the same drying method ($p < 0.05$). Values are means of four replicates \pm SE

and 144 μ g g⁻¹ DW) as the steam kill-enzyme torrefaction and microwave drying was 3 min and 650 W, separately. The research showed that the decomposition of amino acids might be caused by the higher microwave power, the higher oven temperature, and the longer kill-enzyme time. In the three drying methods, oven drying was benefcial to preserving the amino acid content in lotus leaves. Therefore, the actual conditions of each processing methods should be considered.

In humans, there are many important biosynthetic processes need vitamin C to take part, for example, to reduce the oxidative stress of ascorbate peroxidase substrates. Yang and He [\(2009\)](#page-7-14) put forward that microwave-dried red dates had the highest vitamin C content, which was higher than that of natural drying and electrothermic drying. Lv et al. ([2011\)](#page-6-18) also reported that microwave drying could not only efectively shorten the drying time and increase the drying rate, but also could greatly improve the preservation rate of vitamin C in dry hawthorn slices compared with the traditional natural drying and hot air drying. In the current research, vitamin C in lotus leaf increased frst and then reduced while the enhance of microwave power, and achieved the maximum content (22.344 mg g^{-1} DW) when the microwave power was 650 W (Fig. [5\)](#page-4-0). Vitamin C content in lotus leaf signifcantly rose while the enhance of the steam kill-enzyme time. But, vitamin C content dropped frst and rose subsequently with the increased of the oven temperature. Microwave drying was benefcial to preserving the content of vitamin C in lotus leaves among the three drying methods.

Protein is the material basis of life and the main undertaker of life activity. Wang et al. [\(2008\)](#page-7-15) reported that the protein content of alfalfa hay could be reserved better with the high-temperature drying Wang et al. ([2010](#page-7-16)). also reported

Fig. 5 The vitamin C content with three drying methods in lotus leaves. Oven drying (H1, 55 °C; H2, 65 °C; H3, 75 °C), steam killenzyme torrefaction (Z1, 1 min; Z2, 3 min; Z3, 5 min) and microwave drying (W1, 450 W; W2, 650 W; W3, 850 W). The bars with diferent letters are obviously diferent from each other at the same drying method ($p < 0.05$). Values are means of four replicates \pm SE

Fig. 6 The protein content with three drying methods in lotus leaves. Oven drying (H1, 55 °C; H2, 65 °C; H3, 75 °C), steam kill-enzyme torrefaction (Z1, 1 min; Z2, 3 min; Z3, 5 min) and microwave drying (W1, 450 W; W2, 650 W; W3, 850 W). The bars with diferent letters are obviously diferent from each other at the same drying method $(p<0.05)$. Values are means of four replicates \pm SE

that microwave-treated lotus leaves are more conducive to preserve protein content than traditional drying method. The study further showed that protein content in lotus leaf was also afected by microwave power. In the experimental condition, lotus leaf treated by 850 W microwave had the highest protein content (24.461 mg $g^{-1}DW$), followed by lotus leaf treated by the oven (55 °C) (22.645 mg g^{-1} DW) and lotus leaf processed with 3 min steam kill-enzyme $(21.720 \text{ mg g}^{-1} \text{DW})$ (Fig. [6\)](#page-4-1).

The efects of drying methods on the antioxidant activity in lotus leaf

Scientifc researches have shown that cancer, aging, or other diseases were mostly related to the excess production

Fig. 7 The radical scavenging activity with three drying methods in lotus leaves (**a** DPPH radical scavenging activity; **b** superoxide radical scavenging activity; **c** hydroxyl radical scavenging activity). Oven drying (H1, 55 °C; H2, 65 °C; H3, 75 °C), steam kill-enzyme torrefaction (Z1, 1 min; Z2, 3 min; Z3, 5 min) and microwave drying (W1, 450 W; W2, 650 W; W3, 850 W). The bars with diferent letters are obviously diferent from each other at the same drying method $(p<0.05)$. Values are means of four replicates \pm SE

of free radicals. So, antioxidant activity is listed as one of the main research directions by food, health care products, and cosmetics companies. DPPH was mainly used in the detection of vitro antioxidant activity of antioxidant substances (Kurd and Samavatib [2015](#page-6-19)). Superoxide radical could disrupt human functions (Seedevi et al. [2017](#page-7-17)). Hydroxyl radical was one of the reactive oxygen species that could kill red blood cells, degrade DNA, and cell membranes (Bagchi and Kumar [2016](#page-6-20)). Zhang et al. **Plate 1** Drying treatment in lotus leaves (from the right to the left and from the top to the bottom: oven drying (H1, 55 °C; H2, 65 °C; H3, 75 °C), steam kill-enzyme torrefaction (Z1, 1 min; Z2, 3 min; Z3, 5 min) and microwave drying (W1, 450 W; W2, 650 W; W3, 850 W)

([2017\)](#page-7-18) reported that the lotus plumule could be served as natural antioxidants in functional foods because it was an extremely high antioxidant activity. Jin et al. ([2016\)](#page-6-4) also reported that lotus leaf tea has been popular in China, which is a wholesome drink, especially for antioxidant function.

The DPPH radical scavenging activity in lotus leaf dropped frst and rose subsequently with the increase of the oven temperature and microwave power, and reached a peak value at H1 (55 \degree C) and W1 (450 W), respectively. However, the DPPH radical scavenging activity was obviously declined as the enhance of steam kill-enzyme time (Fig. [7a](#page-4-2)). Among the three drying methods, the lotus leaf processed with oven drying (55 ℃) showed the higher DPPH radical scavenging activity, followed by the lotus leaf processed with the lower microwave power (450 W), and the lotus leaf processed with 1 min steam kill-enzyme.

In Fig. [7](#page-4-2)b demonstrated that the superoxide radical scavenging activity in lotus leaf rose frst and then declined as the enhance of the steam kill-enzyme time and microwave power. The superoxide radical scavenging activity in lotus leaf treated by the oven temperature (55 °C) reached the peak value. In the three drying methods, lotus leaf dried by the 3 min steam kill-enzyme exhibited the higher superoxide radical scavenging activity, while the lotus leaf treated by oven drying exhibited the lowest superoxide radical scavenging activity.

Figure. [7](#page-4-2)c indicated that the hydroxyl radical scavenging activity in lotus leaf signifcantly increased as the enhance of the steam kill-enzyme time and microwave power. However, the hydroxyl radical scavenging activity in lotus leaf rose frst and then declined as the enhance of the oven temperature. Among the three drying methods, the lotus leaf

processed with oven temperature (65 °C) and 5 min steam kill-enzyme drying both had the higher hydroxyl radical scavenging activity than that treated by the microwave drying.

The comprehensive analysis showed that the DPPH radical and the superoxide radical scavenging activity were the higher in lotus leaf treated with oven drying and steam kill-enzyme torrefaction, respectively. In addition, the lotus leaves treated by oven drying and steam kill-enzyme torrefaction both had the higher hydroxyl radical scavenging activity.

The color of lotus leaves in drying methods.

Color parameters, which was one of the important sensory properties of tea (Wu and Sun [2013](#page-7-19)). Jin et al. ([2016\)](#page-6-4) reported that there were many diverse colors in herbal teas. As is shown in Plate [1](#page-5-0), the color of lotus leaf treated with microwave treatments were more bright green than that treated with the other two methods. The reason might be that microwave treatments could be better to keep the higher chlorophyll contents in the lotus leaf. Next, experiments will be performed to prove the hypothesis and fnd the real reason.

Conclusion

Both the drying methods and the specifc conditions of each processing methods will significantly affect the qualities and antioxidant activities of lotus leaves. The experimental study showed that: (1) Oven drying $(55-65 \degree C)$ was

benefcial to keeping the higher contents of alkaloids and amino acids, and the higher activities in DPPH radical scavenging and hydroxyl radical scavenging; (2) Steam kill-enzyme torrefaction (3–5 min) could gain the higher contents of favone and the higher activities of superoxide radical scavenging; (3) The appropriate microwave power and microwave time were about 680–850 W and 3–6 min, separately, which could gain the higher contents of vitamin C, soluble sugar and protein. (4) The color of lotus leaf treated with microwave treatments was more bright green than that treated with the other two methods. Therefore, the actual conditions of each processing methods should be considered when the drying method was selected. These results provided a theoretical basis for further research and improved the qualities of lotus leaf tea.

Author contribution statement Contribution of the authors of the paper was as follows: (1) Idea of the project: XY, JC, CL. (2) Experimentation: CG (3). Data interpretation: CG, NZ, JX. (4) Manuscript writing: CG.

Funding This work has been supported by the National Natural Science Foundation of China (No. 31300321), Natural Science Foundation of Hebei Province (No. C2012201080), and Open Foundation of Hebei Key Laboratory of Wetland Ecology and Conservation (No. hklk201904).

Compliance with ethical standards

Conflict of interest Chun-Yan Guo declares that she has no confict of interest. Nan Zhang declares that she has no confict of interest. Cun-Qi Liu declares that he has no confict of interest. Jing-Jing Xue declares that she has no confict of interest. Jian-Zhou Chu declares that he has no confict of interest. Xiao-Qin Yao declares that she has no confict of interest.

Ethical approval This article does not contain any studies involving human participants or animals performed by any of the authors.

Informed consent Not applicable as this study does not include any human participants.

References

- Bagchi S, Kumar KJ (2016) Studies on water-soluble polysaccharides from *Pithecellobium dulce* (Roxb.) Benth. Seeds Carbohyd Polym 138:215–221. [https://doi.org/10.1016/j.carbp](https://doi.org/10.1016/j.carbpol.2015.11.018) [ol.2015.11.018](https://doi.org/10.1016/j.carbpol.2015.11.018)
- Bradford MM (1976) A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. Anal Biochem 72(1–2):248–254. [https](https://doi.org/10.1016/0003-2697(76)90527-3) [://doi.org/10.1016/0003-2697\(76\)90527-3](https://doi.org/10.1016/0003-2697(76)90527-3)
- Chen R, Li Y, Dong H, Liu Z, Li S, Yang S, Li X (2012) Optimization of ultrasonic extraction process of polysaccharides from *Ornithogalum caudatum* ait and evaluation of its biological activities. Ultrason Sonochem 19(6):1160–1168. [https://doi.](https://doi.org/10.1016/j.ultsonch.2012.03.008) [org/10.1016/j.ultsonch.2012.03.008](https://doi.org/10.1016/j.ultsonch.2012.03.008)
- Cylwik D, Mogielnicki A, Buczko W (2005) l-Arginine and cardiovascular system. Pharmacol Rep 57:14–22
- Dehnad D, Jafari SM, Afrasiabi M (2016) Infuence of drying on functional properties of food biopolymers: from traditional to novel dehydration techniques. Trends Food Sci Tech 57:116– 131. <https://doi.org/10.1016/j.tifs.2016.09.002>
- Du LJ, Sun H, Li M, Jin W, Xu LZ (2000) Studies on the active components in the leaf of *Nelumbo nucifera* and Black Soybean and their mixture with blood lipid-regulating efect. Chin Tradit Herbal Drugs 31(7):526–527
- Hammi KM, Hammamib M, Rihouey C, Cerf DL, Ksouri R, Majdoub H (2016) Optimization extraction of polysaccharide from Tunisian *Zizyphus lotus* fruit by response surface methodology: composition and antioxidant activity. Food Chem 212:476–484. <https://doi.org/10.1016/j.foodchem.2016.06.004>
- Hao LL, Wang ZP, Xu Y, Li HC, Ma SF (2012) Study on the preparation of lotus leaf tea beverage. J Anhui Agri Sci 40:13613– 13615.<https://doi.org/10.13989/j.cnki.0517-6611.2012.27.032>
- He SM, Liu JL (2007) Study on the determination method of favone content in tea. Chin J Anal Chem 35:1365–1368
- Hu JH, Pan L, Liu HG, Wang SZ, Wu ZH, Ke WD, Ding Y (2012) Comparative analysis of genetic diversity in sacred lotus (Nelumbo nucifera Gaertn) using AFLP and SSR markers. Mol Biol Rep 39(4):3637–3647. [https://doi.org/10.1007/s1103](https://doi.org/10.1007/s11033-011-1138-y) [3-011-1138-y](https://doi.org/10.1007/s11033-011-1138-y)
- Huang KY, Zhang ZG (2008) The researchsituation of Lotus leaves. Chin Pharm 17:77–78
- Jafari SM, Azizi D, Mirzaei H, Dehnad D (2016) Comparing quality characteristics of oven-dried and refractance window-dried kiwifruits. J Food Process Pres 40(3):362–372. [https://doi.](https://doi.org/10.1111/jfpp.12613) [org/10.1111/jfpp.12613](https://doi.org/10.1111/jfpp.12613)
- Jin L, Li XB, Tian DQ, Fang XP, Yu YM, Zhu HQ, Ge YY, Ma GY, Wang WY, Xiao WF, Li M (2016) Antioxidant properties and color parameters of herbal teas in China. Ind Crops Prod 87:198– 209.<https://doi.org/10.1016/j.indcrop.2016.04.044>
- Kato E, Inagaki Y, Kawabata J (2015) Higenamine 4'-*O*-β-D-glucoside in the lotus plumule induces glucose uptake of L6 cells through theβ2-adrenergic receptor. Bioorgan Med Chem 23(13):3317– 3321. <https://doi.org/10.1016/j.bmc.2015.04.054>
- Khaskheli SG, Zheng W, Sheikh SA, Khaskheli AA, Liu Y, Soomro AH, Feng X, Sauer MB, Wang YF, Huang W (2015) Characterization of *Auricularia auricular* polysaccharides and its antioxidant properties in the fresh and pickled product. Int J Biol Macromol 81:387–395.<https://doi.org/10.1016/j.ijbiomac.2015.08.020>
- Kim J, Lee I, Seo J, Jung M, Kim Y, Yim N, Bae K (2010) Vitexin orientin and other favonoids from *Spirodela Polyrhiza* inhibit adipogenesis in 3T3-L1 cells. Phytother Res 24(10):1543–1548. <https://doi.org/10.1002/ptr.3186>
- Kurd F, Samavati V (2015) Water-soluble polysaccharides from *Spirulina platensis*: extraction and in vitro anti-cancer activity. Int J Biol Macromol 74:498–506. [https://doi.org/10.1016/j.ijbio](https://doi.org/10.1016/j.ijbiomac.2015.01.005) [mac.2015.01.005](https://doi.org/10.1016/j.ijbiomac.2015.01.005)
- Li HS, Sun Q, Zhao SJ, Zhang WH (2004) Plant physiological and biochemical principle and experimental technique. Higher Education Press, Beijing
- Liao CH, Lin JY (2013) Lotus (*Nelumbo nucifera* Gaertn.) plumule polysaccharide ameliorates pancreatic islets loss and serum lipid profles in non-obese diabetic mice. Food Chem Toxicol 58:416– 422.<https://doi.org/10.1016/j.fct.2013.05.018>
- Lv YZ, Liang ZH, Liu G, Shi MJ (2011) Efects of diferent drying methods on the stability of vitamin C in the dry process of *Cornus* Officinalis. Process Agric Prod 6:70-71
- Machado RAR, Ferrieri AP, Robert CAM, Glauser G, Kallenbach M, Baldwin IT, Erb M (2013) Leaf-herbivore attack reduces carbon reserves and regrowth from the roots via jasmonate and auxin

signaling. New Phytol 200(4):1234–1246. [https://doi.org/10.1111/](https://doi.org/10.1111/nph.12438) [nph.12438](https://doi.org/10.1111/nph.12438)

- Manthey JA, Guthrie N, Grohmann K (2001) Biological properties of citrus favonoids pertaining to cancer and infammation. Curr Med Chem 8(2):135–153. <https://doi.org/10.2174/0929867013373723>
- Mateo RD, Wu G, Bazer FW, Park JC, Shinzato I, Kim SW (2007) Dietary l-arginine supplementation enhances the reproductive performance of gilts. J Nutr 137(3):652–656. [https://doi.org/10.1093/](https://doi.org/10.1093/jn/137.3.652) [jn/137.3.652](https://doi.org/10.1093/jn/137.3.652)
- Mohsen SM, Ammar ASM (2009) Total phenolic contents and antioxidant activity of corn tassel extracts. Food Chem 112(3):595–598. <https://doi.org/10.1016/j.foodchem.2008.06.014>
- Morris CR, Kato GJ, Poljakovic M, Wang X, Blackwelder WC, Sachdev V, Hazen SL, Vichinsky EP, Morris JSM, Gladwin MT (2005) Dysregulated arginine metabolism, hemolysis-associated pulmonary hypertension and mortality in sickle cell disease. J Am Med Assoc 294(1):81–90.<https://doi.org/10.1001/jama.294.1.81>
- Mukhtar H, Ahmad N (2000) Tea polyphenols: prevention of cancer and optimizing health. Am J Clin Nutr 71(6):1698–1702. [https://](https://doi.org/10.1093/ajcn/71.6.1698s) doi.org/10.1093/ajcn/71.6.1698s
- Ono Y, Hattori E, Fukaya Y, Imai S, Ohizumi Y (2006) Anti-obesity efect of *Nelumbo nucifera* leaves extract in mice and rats. J Ethnopharmacol 106(2):238–244. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.jep.2005.12.036) [jep.2005.12.036](https://doi.org/10.1016/j.jep.2005.12.036)
- Seedevi P, Moovendhan M, Viramani S, Shanmugam A (2017) Bioactive potential and structural characterization of sulfated polysaccharide from seaweed *(Gracilaria cortical*). Carbohyd Polym 155:516–524. <https://doi.org/10.1016/j.carbpol.2016.09.011>
- Shi XF, Chu JJ, Zhang YF, Liu CQ, Yao XQ (2017) Nutritional and active ingredients of medicinal chrysanthemum fower heads afected by diferent drying methods. Ind Crops Prod 104:45–51. <https://doi.org/10.1016/j.indcrop.2017.04.021>
- State Standard of the People's Republic of China (1990) Method for determination of total ascorbic acid in fruits, vegetables and derived products (GB 12392–90)
- Wang L, Zhao YJ, Zhang YY, Wu JF, Zhang XN, Ji ZQ, Yue W (2005) Determination of alkaloids in pinellia tuber and its antiemetic study. Chin Pharmacol bull 21:864–867
- Wang XL, Yan XD, Xu YP, Zhao ZX (2008) Preliminary study on the factors afecting the protein contents of alfalfa hay. Pratacult Sci 3:47–50
- Wang DL, Xu WX, Li C, Huang Y (2010) Microwave drying of tea beverage quality. In: Transformation of economic development mode and independent innovation-The 12th China Science and Technology Association Annual Meeting (Volume II)
- Wu D, Sun DW (2013) Colour measurements by computer vision for food qualitycontrol—a review. Trends Food Sci Tech 29:5–20
- Yang YJ, He HS (2009) Determination of vitamin C content in red dates of diferent drying methods. Mod Prevent Med 36:42–43
- You L, Zhao M, Regenstein JM, Ren J (2011) In vitro antioxidant activity and in vivo anti-fatigue efect of loach (*Misgurnus anguillicaudatus*) peptides prepared by papain digestion. Food Chem 124(1):188–194.<https://doi.org/10.1016/j.foodchem.2010.06.007>
- Yuan J, Hao LJ, Wu G, Wang S, Duan JA, Xie GY, Qin MJ (2015) Efects of drying methods on the phytochemicals contents and antioxidant properties of chrysanthemum fower heads harvested at two developmental stages. J Funct Foods 19:786–795. [https://](https://doi.org/10.1016/j.jff.2015.10.008) [doi.org/10.1016/j.jf.2015.10.008](https://doi.org/10.1016/j.jff.2015.10.008)
- Zhang L, Wu HY, Deng DW, Huang GH (2011) Study on new technology of lotus leaf tea by microwave-assisted drying. Sci Tech Food Indus 32(04):295–298. [https://doi.org/10.13386/j.issn1](https://doi.org/10.13386/j.issn1002-0306.2011.04.070) [002-0306.2011.04.070](https://doi.org/10.13386/j.issn1002-0306.2011.04.070)
- Zhang L, Liu TF, Xue YJ, Liu C, Ru H, Dong MH, Yu ZF (2015) Efects of drying methods on the aroma components and quality of *Capsella Bursa-Pastoris* L. J Food Process Eng 39(2):107–120. <https://doi.org/10.1111/jfpe.12204>
- Zhang LJ, Cheng Z, Zhao Q, Wang MS (2017) Green and efficient PEG-based ultrasound-assisted extraction of polysaccharides from superfne ground lotus plumule to investigate their antioxidant activities. Ind Crops Prod 109:320–326. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.indcrop.2017.08.018) [indcrop.2017.08.018](https://doi.org/10.1016/j.indcrop.2017.08.018)
- Zhao J, Deng JW, Chen YW, Li SP (2013) Advanced phytochemical analysis of herbal tea in China. J Chromatogr A 1313:2–23. [https](https://doi.org/10.1016/j.chroma.2013.07.039) [://doi.org/10.1016/j.chroma.2013.07.039](https://doi.org/10.1016/j.chroma.2013.07.039)

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional afliations.