

Effects of soil flooding and changes in light intensity on photosynthesis of *Eugenia uniflora* L. seedlings

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Abstract The increased frequency of heavy rains as a result of global climate change can lead to flooding and changes in light availability caused by the presence of thick clouds. To test the hypothesis that reduction in light availability can alleviate the harmful effects of soil flooding on photosynthesis, the authors studied the effects of soil flooding and acclimation from high to low light on the photosynthetic performance of *Eugenia uniflora*. Seedlings acclimated to full sunlight (about $35 \text{ mol m}^{-2} \text{ d}^{-1}$) for 5 months were transferred to partial sunlight (about $10 \text{ mol m}^{-2} \text{ d}^{-1}$) and were either subjected to soil flooding or not flooded. Chlorophyll fluorescence was measured throughout the flooding period and leaf gas exchange was measured 16 days after flooding was initiated. Minimal fluorescence yield (F_0) was significantly higher and the quantum efficiency of open PSII centres (F_v/F_m) was significantly lower in flooded than in non-flooded plants in full sunlight. Sixteen days after flooding was initiated, stomatal conductance ($g_{s_{\text{sat}}}$) and net photosyntheses expressed on a leaf area ($A_{\text{sat-area}}$), weight ($A_{\text{sat-wt}}$) and chlorophyll ($A_{\text{sat-Chl}}$) basis decreased in response to soil flooding. Flooding decreased stomatal conductance by similar amounts in full and partial sunlight, but $A_{\text{sat-area}}$ in partial and full sunlight was 3.4 and 16.8 times lower,

respectively, in flooded than in non-flooded plants. These results indicate that changes from full to partial sunlight during soil flooding can alleviate the effects of flooding stress on photosynthesis in *E. uniflora* seedlings acclimated to full sunlight. The responses of photosynthesis in trees to flooding stress may be dependent on changes in light environment during heavy rains.

Keywords Chlorophyll fluorescence · Global change · Leaf gas exchange · Stomatal conductance

Introduction

Plants have different strategies to survive and grow in high or low light environments (Smith 1982; Valladares and Niinemets 2008). In general, leaves have great phenotypic plasticity in relation to the light environment and high capacity to acclimate to changes in light availability (Walters 2005; Valladares and Niinemets 2008). Leaves acclimated to high light are very different in terms of their physiology, anatomy and morphology than shade-acclimated leaves. In general, sun leaves have greater leaf weight per area and chlorophyll a/b ratio, lower chlorophyll content, less nitrogen allocated to the light-harvesting complexes and more nitrogen allocated to the enzymes of the Calvin cycle than shade leaves (Evans and Poorter 2001; Valladares and Niinemets 2008). Photosynthetic acclimation of shaded leaves to full sun, or vice versa, may involve physiological adjustments within hours, days or weeks, or structural changes within weeks or months (Bongers and Popma 1990; Walters 2005; Mielke and Schaffer 2010a).

Flooding is a major environmental stress to terrestrial plants in ecosystems prone to high rainfall or poor soil

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drainage, as well as in ecosystems susceptible to high water table fluctuations (Schaffer 1998; Pezeshki 2001; Kozłowski 2002; Kreuzwieser et al. 2004). Soil flooding induces oxygen deficiency and low soil redox potential which affect different aspects of plant physiology and development, such as premature leaf senescence and abscission, changes in leaf formation and expansion, suppression of root metabolism and growth and absorption of macronutrients (Pezeshki 2001; Kozłowski 2002; Kreuzwieser et al. 2004). Changes in CO₂ assimilation is one of the first measurable responses of plants to soil flooding, which can be attributed to stomatal and non-stomatal limitations to photosynthesis (Mielke et al. 2003; Herrera et al. 2008). Studies have shown that the photosynthetic performance and growth of different plant species to soil flooding may be influenced by the interaction of flooding with light intensity (Wagner and Dreyer 1997; Gardiner and Krauss 2001; Lavinsky et al. 2007; Mielke and Schaffer 2010a, b).

In many tropical areas, high rainfall is accompanied by the presence of thick clouds, causing a large reduction in light available for CO₂ assimilation and growth (Zotz and Winter 1994; Graham et al. 2003). In addition, in tropical and subtropical regions the increased frequency of heavy rains as a result of global climate change (Vera et al. 2006; Marengo et al. 2009) can lead to flooding (Michener et al. 1997; Milly et al. 2002; Hirabayashi et al. 2008) and changes in light availability caused by the presence of thick clouds. Thus, in ecosystems prone to soil flooding, it is probable that reductions in light caused by the presence of clouds can reduce the harmful effects of flooding on the photosynthetic performance of plants. Although there are many references on photosynthetic acclimation of plants transferred from shade to full sun (i.e., Bongers and Popma 1990; Evans and Poorter 2001; Krause et al. 2001; Houter and Pons 2005; Guo et al. 2006; Naramoto et al. 2006), and interactive effects of light environment and soil flooding (Wagner and Dreyer 1997; Gardiner and Krauss 2001; Lavinsky et al. 2007; Mielke and Schaffer 2010a, b), the authors are aware of no published studies on the effects of changes from high to low light on the physiological responses of plants subjected to soil flooding.

Eugenia uniflora L. (Myrtaceae) is a typical shrub or small tree species native to the Restingas (Margis et al. 2002), a marginal ecosystem of the Brazilian Atlantic Rainforest (Scarano 2002). Restingas occur along the coastline, including marshes, dry and swamp forests, and open clumped vegetation, where the occurrence of drought and soil flooding are often limiting factors for plant growth (Henriques et al. 1986). Studies have shown that *E. uniflora* is a high light demanding species and moderately sensitive to soil flooding (Mielke and Schaffer 2010a, b). Mielke and Schaffer (2010a) observed that the light availability during flooding and the pre-acclimation to different light

intensities affected the physiological performance and growth of seedlings subjected to soil flooding. In an experiment in which *E. uniflora* plants cultivated in a shade house were transferred to full or partial sunlight and subjected to soil flooding, the photosynthetic apparatus of leaves developed in shade had a relatively limited capacity to acclimate to changes in light intensity (Mielke and Schaffer 2010b). In that experiment, both stomatal and non-stomatal limitations to photosynthesis were related to the low capacity of photosynthetic acclimation of flooded seedlings after transference from shade to full sun.

To test the hypothesis that changes in light availability, that can be caused by the presence of thick clouds, can alleviate the harmful effects of soil flooding on photosynthesis of trees during periods of heavy rains, we conducted an experiment aimed at analyzing the effects of soil flooding and acclimation from high to low light on chlorophyll fluorescence, leaf chlorophyll content and leaf gas exchange of *E. uniflora* seedlings.

Materials and methods

The experiment was conducted at the Tropical Research and Education Center, University of Florida (TREC/UF), Homestead, Florida, USA (25.5°N 80.5°W). In March 2008, *E. uniflora* L. seedlings were obtained from a commercial nursery located in Homestead, Florida, USA. Plants were cultivated in 10-l plastic containers (ten to twelve plants per container) with a standard nursery substrate of 65 pine bark, 25 Florida peat and 10% coarse sand by volume. Plants were carefully selected to obtain a uniform sample. The seedlings were about 1 year old at the time of transference to TREC/UF. At this stage, the average height of the plants per container was 0.5–0.8 m and the average stem diameter 0.10 m above the soil surface was 3–10 mm. In an open field at TREC/UF, 3 × 2 m (an area sufficient for ten containers) blocks were selected perpendicular to the daily track of the sun, permitting plants in full sunlight to receive almost all solar radiation during the day. Shade cages (3 × 2 × 1.7 m) were constructed with PVC tubes covered with one layer of a neutral shade netting (25–30% of full sunlight) and placed on four blocks. The remaining four blocks were left open (no cages). Cages were spaced far enough apart so that they did not shade the open blocks.

The experiment was arranged in a completely randomized design with a 2 × 2 factorial arrangement consisting of two light treatments (partial and full sunlight), two flood treatments (flooded and non-flooded) and four replications (blocks) per treatment. The containers were kept in full sunlight from March 2008 to October 2008. On 16 October 2008, half of the containers were transferred to the shade

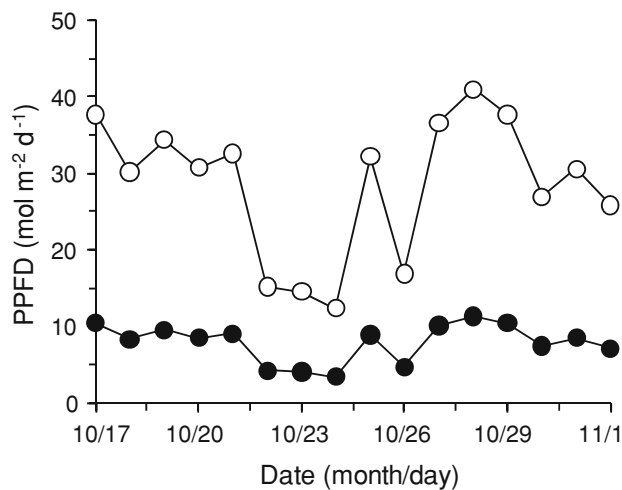


Fig. 1 Total daily photosynthetic photon flux density (PPFD) during the period of soil flooding (*open circles* represent plants in full sun and *closed circles* represent plants in partial sunlight) in Homestead, Florida, USA. Data from 17 October 2008 to 1 November 2008

cages and half remained in full sunlight. Half of the plants in each light treatment were flooded and the remaining plants were maintained with soil water content near field capacity using an automatic irrigation system. Plants in the flooded treatment were flooded by placing each container in a 19-l plastic bucket filled with tap water to 50–100 mm above the soil surface to ensure complete inundation of the root system. To ensure that the irrigation system was sufficient to maintain the soil water content of non-flooded plants at field capacity, tensiometers (Soil Moisture Equipment Corp., Santa Barbara, California, USA) were installed in three containers in the full sun treatment.

During the period of acclimation to full sunlight and throughout the experiment, the photosynthetic photon flux density (PPFD) was measured in each light treatment with model LI-190SA quantum sensors connected to LI-1000 data loggers (Li-Cor, Inc., Lincoln, Nebraska, USA). In full and partial sunlight treatments, air temperature (T_a) and relative humidity (RH) were monitored and recorded with Hobo H8 Pro Series dataloggers (Onset Computer, Bourne, Massachusetts, USA), and the vapour pressure deficit (VPD) calculated as described by Landsberg (1986). Total daily rainfall during the experiment was obtained from a weather station of the Florida Automated Weather Network

(<http://fawn.ifas.ufl.edu/>) located 50 m from the experiment. The same averages of T_a and VPD for the partial and full sunlight treatments were reached during the flooding period, i.e., 22.1°C and 0.5 kPa, respectively. Total daily PPFD during plant acclimation was between 4.7 mol m⁻² d⁻¹ (18 August 2008) and 56.5 mol m⁻² d⁻¹ (13 May 2008). The average value of rainfall during the period of soil flooding was 0.1 mm. The maximum, minimum and average values of total daily PPFD were 11.4, 3.4 and 7.9 mol m⁻² d⁻¹ in partial sunlight and 41.0, 12.3 and 28.4 mol m⁻² d⁻¹ in full sunlight (Fig. 1). In this experiment the authors did not measure the soil redox potential (Eh), but in a parallel experiment (Mielke and Schaffer 2010b) the values of soil Eh decreased to -80 mV and remained close to this value in both partial and full sunlight treatments 1 week after the soil was initially flooded.

One, five and eleven days after seedlings were flooded, chlorophyll fluorescence was measured for two recently matured, fully-expanded leaves per container (one container per replication) with a portable fluorescence system (model OS-30, Opti-Sciences Inc., Hudson, New Hampshire, USA) between 12 and 13:00 h. Total daily and maximum instantaneous values of PPFD during days in which chlorophyll fluorescence was measured are presented in Table 1. Leaves were acclimated in the dark for 30 min prior to each chlorophyll fluorescence measurement. Sixteen days after seedlings were flooded chlorophyll fluorescence, leaf chlorophyll content, leaf gas exchange and leaf weight per area (LWA) were measured on one recently matured, fully-expanded leaf per container (one container per replication). Chlorophyll fluorescence was measured as previously described between 8 and 9:00 am. The minimal (F_o), maximum (F_m) and variable (F_v) fluorescence yields and the quantum efficiency of open PSII centres (F_v/F_m) were measured and calculated as described by Maxwell and Johnson (2000). After chlorophyll fluorescence measurements, leaf gas exchange variables were measured with a portable photosynthesis system (CIRAS-2, PP Systems, Amesbury, Massachusetts, USA). Prior to each measurement, leaves were acclimated to the environmental conditions inside the leaf chamber for about 20 min. The air flow rate, T_a , VPD, PPFD inside the leaf chamber and the reference CO_2 concentration (C_a) during measurements were 200 ml min⁻¹, 25°C, 1 kPa, 1000 μ mol photons m⁻² s⁻¹

Table 1 Total daily and maximum instantaneous values of PPFD during the days in which chlorophyll fluorescence data was collected

Day	Total PPFD (mol m ⁻² d ⁻¹)		Maximum PPFD (μ mol m ⁻² s ⁻¹)	
	Partial sunlight	Full sunlight	Partial sunlight	Full sunlight
10/17/08	10.4	37.7	506	1493
10/21/08	9.0	32.6	502	1489
10/27/08	10.1	36.6	472	1370
11/01/08	7.2	25.8	476	1426

Table 2 Minimal fluorescence yield (Fo) and the quantum efficiency of open photosystem II centres (Fv/Fm) of *E. uniflora* seedlings after one, five, eleven (between 12 and 13:00 h) and 16 days (between 8

and 9:00 h) of transference from full to partial sunlight and exposure to soil flooding

Variable	Days	Partial sunlight		Full sunlight		ANOVA		
		Flooded	Non-flooded	Flooded	Non-flooded	L	F	L × F
Fo (rel. units)	1	229 ± 40 a	238 ± 42 a	336 ± 132 a	327 ± 79 a	ns	ns	ns
	5	222 ± 40 a	224 ± 38 a	389 ± 240 a	288 ± 171 a	ns	ns	ns
	11	186 ± 41 aB	209 ± 11 aA	679 ± 107 aA	368 ± 190 bA	**	*	**
	16	244 ± 34 a	214 ± 41 a	285 ± 43 a	212 ± 5 b	ns	*	ns
Fv/Fm (rel. units)	1	0.71 ± 0.06 a	0.71 ± 0.06 a	0.68 ± 0.06 a	0.67 ± 0.03 a	ns	ns	ns
	5	0.71 ± 0.06 a	0.70 ± 0.06 a	0.67 ± 0.15 a	0.73 ± 0.11 a	ns	ns	ns
	11	0.76 ± 0.05 aA	0.71 ± 0.04 aA	0.49 ± 0.10 bB	0.64 ± 0.13 aA	**	ns	*
	16	0.69 ± 0.05 a	0.74 ± 0.03 a	0.55 ± 0.09 b	0.71 ± 0.02 a	**	**	ns

Data are the means ± standard deviations of four replications within each factor (light environment or soil flooding)

Interactions between light and flood treatments were analyzed by a two-way ANOVA: $P > 0.05$ (ns), $P \leq 0.05$ (*), $P \leq 0.01$ (**). Means followed by the *same letter* are not significantly different according to a non-paired *T*-test ($P < 0.05$). Capital letters represent comparisons of light effects within flood treatments and *lower case letters* represent comparisons of flood effects within light environments

and $375 \mu\text{mol mol}^{-1}$, respectively. The value of PPFD used in this experiment was above the light saturation point for *E. uniflora* (Mielke and Schaffer 2010b). The light saturated net photosynthetic rate on a leaf area basis ($A_{\text{sat-area}}$) and stomatal conductance to water vapour (g_{sat}) were calculated using the values of CO_2 and humidity variation inside the cuvette. Intrinsic water use efficiency (A/g_s) was calculated by dividing $A_{\text{sat-area}}$ by g_{sat} .

After leaf gas exchange measurements, total leaf chlorophyll content (Chl) was estimated with a SPAD meter (Model 502, Minolta Inc., Osaka, Japan). SPAD values were converted to total chlorophyll (Chl a + b) content using the equation derived by Mielke et al. (2010) for *E. uniflora*. Immediately after SPAD measurements, the leaves were collected and total leaf area and leaf dry weight were determined for each plant. Leaf area was determined with a leaf area meter (model LI-3000, Li-Cor Inc., Lincoln, Nebraska, USA) and leaf dry weight was obtained after oven-drying leaves at 75°C until a constant weight was reached. Leaf weight per area (LWA) was calculated by dividing the leaf dry weight by the leaf area. Light saturated net photosynthetic rates based on leaf weight ($A_{\text{sat-wt}}$) and leaf chlorophyll ($A_{\text{sat-Chl}}$) were calculated by dividing $A_{\text{sat-area}}$ by LWA and Chl a + b, respectively.

Interactions between light and flood treatments were analyzed by a two-way ANOVA. When interactive effects of light × flood treatments were observed, a non-paired *T*-test was used for comparisons of light effects within flood treatments and flood effects within light environments. When interactive effects of light × flood treatments were not observed, only comparisons between flood treatments within light environments were analyzed by a non-paired *T*-test.

Results

From the first to the fifth day after flooding was initiated there were non-significant differences in Fo and Fv/Fm among treatments (Table 2). Eleven days after flooding, the average values of Fo were significantly higher and the average Fv/Fm lower ($P \leq 0.05$) in flooded plants in full sunlight compared with flooded plants in partial sunlight. Also, from the first to the eleventh day after flooding there was an increase of 102% in the average values of Fo and 39% decrease of Fv/Fm in the leaves of flooded plants in full sun. Sixteen days after transferring plants from full to partial sunlight and initiating flooding treatments (Table 1), there were significant differences between full and partial sunlight treatments for Fv/Fm ($P \leq 0.01$) and significant effects of flooding on Fv/Fm ($P \leq 0.01$) and Fo ($P \leq 0.05$). The average Fv/Fm was significantly higher in non-flooded than in flooded plants only in full sunlight. No significant difference ($P > 0.05$) was observed between Fo in flooded and non-flooded plants in partial sunlight, but a significant difference ($P \leq 0.05$) for this variable was observed between flooded and non-flooded plants in full sunlight. In the full sunlight treatment, the average value of Fo was 35% higher in flooded than in non-flooded plants. The average values of Fv/Fm were, respectively, 1.1 and 1.3 times higher in non-flooded than in flooded plants in partial and in full sunlight.

Sixteen days after transferring plants from full to the partial sunlight and flooding (Table 3), there were significant differences between full and partial sunlight treatments for Chl a + b, g_{sat} , $A_{\text{sat-area}}$, $A_{\text{sat-wt}}$ ($P \leq 0.01$) and A/g_s ($P \leq 0.05$) and significant effects of flooding on g_{sat} , $A_{\text{sat-area}}$, $A_{\text{sat-wt}}$, $A_{\text{sat-Chl}}$ ($P \leq 0.01$) and A/g_s ($P \leq 0.05$).

Table 3 Leaf weight per area (LWA) and leaf-level photosynthetic characteristics of *E. uniflora* seedlings after 16 days of transference from full to partial sunlight and exposure to soil flooding

Variable	Partial sunlight		Full sunlight		ANOVA		
	Flooded	Non-flooded	Flooded	Non-flooded	L	F	L × F
LWA (g m ⁻²)	109.6 ± 8.7 a	113.1 ± 6.5 a	123.4 ± 16.3 a	114.5 ± 5.1 a	ns	ns	ns
Chl a + b (μmol m ⁻²)	367.3 ± 99.0 a	401.7 ± 111.9 a	207.9 ± 21.2 b	305.3 ± 51.7 a	**	ns	ns
g _{Sat} (mmol m ⁻² s ⁻¹)	18.8 ± 3.5 b	72.8 ± 17.7 a	11.3 ± 4.4 b	49.0 ± 5.0 a	**	**	ns
A _{Sat-area} (μmol m ⁻² s ⁻¹)	2.6 ± 0.9 b	8.7 ± 1.9 a	0.4 ± 0.5 b	6.7 ± 1.4 a	**	**	ns
A _{Sat-wt} (μmol kg ⁻¹ s ⁻¹)	24.5 ± 9.9 b	77.4 ± 19.6 a	2.8 ± 3.7 b	58.8 ± 14.8 a	**	**	ns
A _{Sat-Chl} (μmol mmol ⁻¹ s ⁻¹)	7.1 ± 1.9 b	22.0 ± 2.9 a	1.7 ± 2.2 b	23.8 ± 5.5 a	ns	**	ns
A/g _S (μmol mol ⁻¹)	141.2 ± 53.1 aA	120.4 ± 12.9 aA	32.1 ± 37.6 bB	135.1 ± 20.2 aA	*	*	**

Data are the means ± standard deviations of four replications within each factor (light environment or soil flooding)

Interactions between light and flood treatments were analyzed by a two-way ANOVA: $P > 0.05$ (ns), $P \leq 0.05$ (*), $P \leq 0.01$ (**). Means followed by the *same letter* are not significantly different according to a non-paired *T*-test ($P < 0.05$). *Capital letters* represent comparisons of light effects within flood treatments and *lower case letters* represent comparisons of flood effects within light environments. (*Chl a + b*) total chlorophyll, (*g_{Sat}*) stomatal conductance of water vapour at light saturation, (*A_{Sat-area}*) net photosyntheses expressed on an area, (*A_{Sat-wt}*) weight and (*A_{Sat-Chl}*) chlorophyll basis and (*A/g_S*) intrinsic water use efficiency

There were significant interactions ($P \leq 0.01$) between light and flooding treatments for *A/g_S*. The average values of *Chl a + b*, *g_{Sat}*, *A_{Sat-area}*, *A_{Sat-wt}* and *A_{Sat-Chl}* were significantly higher ($P \leq 0.05$) in non-flooded than in flooded plants in both full and partial sunlight; whereas *A/g_S* was significantly higher in non-flooded than in flooded plants only in full sunlight. In the full sunlight treatment, the average *A/g_S* was 72% higher in non-flooded than in flooded plants. In partial sunlight, the average *Chl a + b*, *g_{Sat}*, *A_{Sat-area}*, *A_{Sat-wt}* and *A_{Sat-Chl}* in non-flooded plants were, respectively, 1.1, 4.0, 3.4, 3.2 and 3.1 times higher than in flooded plants, whereas full sunlight the same differences were, respectively, 1.5, 4.3, 16.8, 21.0 and 14.0 times higher.

Discussion

In this experiment, transferring plants acclimated to full sun to partial sunlight was aimed at simulating changes in light availability due to the appearance of thick clouds during tropical storms or heavy rains. Thus, plants acclimated to full sunlight were subjected to continuous flooding and full and partial (30%) sunlight treatments for 16 days in an open field experiment. There are few references reporting changes in light availability caused by the presence of clouds on photosynthesis of tropical trees. In a seasonal rainforest in Panama, Graham et al. (2003) reported that monthly average values of PPFD during the wet season were about 70% of the values measured during the dry season and that changes in light availability was an important factor limiting CO₂ assimilation and growth of trees during rainy season. However, considering the

variations between clear and cloudy days, these differences can be much larger. On heavily clouded days, the total daily solar radiation can reach values at or lower than 30% of full sun. For example, during the acclimation of plants to full sunlight (data not shown), it was calculated a maximum total daily PPFD of 56.5 mol m⁻² d⁻¹ on 13 May 2008, when the total daily rainfall was 0 mm, and a minimum value of 4.7 mol m⁻² d⁻¹ on 18 August 2008, when the total daily rainfall was 68 mm; a percentage difference of about 92% (or 8% in relation to full sunlight).

Despite variations in the total daily values of PPFD in full sun during the experiment, on the first 3 days of chlorophyll fluorescence measurements PPFD values were within the range found in two previous studies conducted by Mielke and Schaffer (2010a, b). Moreover, the maximum daily values of PPFD in full and partial sunlight were, respectively, higher and lower than the values of the light saturation point for *E. uniflora* (Mielke and Schaffer 2010b).

Significant differences observed between flooded and non-flooded plants in full sunlight for *F_o* and *F_v/F_m* indicate that the leaves of flooded plants in full sunlight were more susceptible to the photoinhibition of photosynthesis (Baker 2008). Photoinhibition occurs when the rate of absorption of light energy by photosynthetic pigments exceeds the utilization rate in chloroplasts and several studies have shown the occurrence of synergistic effects between high light and other stress factors on photoinhibition of photosynthesis (Murata et al. 2007). Changes in *F_v/F_m* in plants subjected to soil flooding and high light have been reported by others (Lavinsky et al. 2007; Mielke and Schaffer 2010a). In this experiment, 11 and 16 days after transferring plants from full to the partial sunlight and

the onset of flooding, significant differences in Fv/Fm and Fo between flooded and non-flooded plants in full sun were observed. On day 5, although differences were not significant between flooded and non-flooded treatments for Fv/Fm and Fo, those variables were, respectively, 31% lower and 35% higher in flooded than in non-flooded plants. Even though other complementary methods should be used to access the extension of photoinhibition in leaves (Logan et al. 2007), the changes in Fv/Fm and Fo indicated that flooding plants in full sunlight affected the flow of energy through the photosynthetic processes, probably making these plants susceptible to photoinhibition (Baker 2008).

Leaf weight per area is a very important morphological indicator of acclimation to changes in light availability and is often associated with plasticity in relation to light acclimation (Evans and Poorter 2001; Aranda et al. 2004; Valladares and Niinemets 2008). In general, sun leaves have higher LWA than shade leaves which is related to thicker leaves and/or reduced leaf area of sun leaves compared to shade leaves. The increased thickness of leaves is usually associated with anatomical changes, such as more photosynthetic cells per unit of leaf area (Evans and Poorter 2001) causing a higher photosynthetic capacity in sun than in shade leaves (Meir et al. 2008). In this experiment all photosynthetic measurements were done on leaves that had been grown in full sunlight and LWA did not vary substantially after transferring plants from full to partial sunlight. The average values of LWA varied between 109.6 and 123.4 g m⁻² and were similar to values observed for leaves of *E. uniflora* acclimated to full sunlight and higher than values for leaves acclimated to partial sunlight in previous experiments conducted under similar conditions (Mielke and Schaffer 2010a, b). Thus, the differences among treatments cannot be explained by changes in leaf structure.

Leaf chlorophyll content is known to be higher in shade than in sun leaves (Valladares and Niinemets 2008). The estimated average Chl values in *E. uniflora* leaves acclimated to full sunlight were lower than those of leaves acclimated to partial sunlight (Mielke and Schaffer 2010b). The average leaf Chl a + b was approximately 32% higher in partial than in full sunlight. These results indicate that short-term acclimation to partial sunlight in *E. uniflora* leaves is related to an increase in chlorophyll content. Losses of chlorophyll and leaf chlorosis, in contrast, are stress symptoms observed in tropical or temperate tree species subjected to soil flooding (Gravatt and Kirby 1998; Gardiner and Krauss 2001; Oliveira and Joly 2010). Sixteen days after flooding, despite the 9% difference in leaf Chl a + b content between flooded and non-flooded plants in partial sunlight, a 47% difference was observed between flooded and non-flooded plants in full sunlight. In addition, the average Chl a + b value was 20% higher in flooded

plants in partial sunlight than in non-flooded plants in full sunlight, indicating that changes from high to low light may alleviate the chlorophyll loss in plants subjected to soil flooding. Also, such differences may be related to the effects of soil flooding on the ability of leaf acclimation after transference from full to partial sunlight.

The values of $A_{\text{sat-area}}$, $A_{\text{sat-wt}}$ and g_{sat} in leaves of non-flooded plants in partial sunlight were similar to values obtained in two previous experiments in which *E. uniflora* plants were grown in the same shade cages used in this experiment (Mielke and Schaffer 2010a, b). In general, sun-acclimated leaves have higher $A_{\text{sat-area}}$ and g_{sat} , and lower $A_{\text{sat-wt}}$ than shade-acclimated leaves (Valladares and Niinemets 2008). Interestingly, in this study there was an increase in $A_{\text{sat-area}}$ after the transference of plants from full to partial sunlight. It is possible that the increase in $A_{\text{sat-area}}$ may be related to increases in Chl and g_{sat} , since Chl and g_{sat} increased after plants were transferred from full to partial sunlight. The values of $A_{\text{sat-Chl}}$ are often higher in sun than in shade acclimated leaves (Pons and Anten 2004), which is related to increased activity of the photochemistry of photosynthesis at the expense of activity of the Calvin cycle (Percy 2000). The $A_{\text{sat-area}}$, $A_{\text{sat-wt}}$ and $A_{\text{sat-Chl}}$ declined as a result of flooding treatments in both full and partial sunlight. Reductions in net CO₂ assimilation and g_{sat} are common responses of tree species to soil flooding (Gravatt and Kirby 1998; Pezeshki and DeLaune 1998; Nuñez-Elisea et al. 1999; Gardiner and Krauss 2001; Mielke et al. 2003; Lavinsky et al. 2007). In many flood-tolerant plant species, increases in A/gs due to stomatal limitation of photosynthesis are related to decreases in stomatal conductance associated with maintenance of high photosynthetic rates (Mielke et al. 2003; Lavinsky et al. 2007). The dramatic decrease in A/gs in flooded plants in full sunlight and the 17% increase in A/gs observed in flooded plants when compared with non flooded plants in partial sunlight 16 days after flooding indicate that stomatal limitation of photosynthesis occurred only in the plants transferred to the partial sunlight.

The non-significant difference between light treatments on day 16 for Fo may be related to the fact that chlorophyll fluorescence was measured in the early morning whereas on days 1, 5 and 11 the chlorophyll fluorescence was measured around midday. Also, the non-significant difference between light treatments on day 16 for $A_{\text{sat-Chl}}$, at same time in which $A_{\text{sat-area}}$ and $A_{\text{sat-weight}}$ were significantly higher in partial than in full sunlight, could be related to the increase in chlorophyll content after the transference of the plants from full to partial sunlight. As discussed previously, shade leaves have higher leaf area, higher chlorophyll content and lower leaf weight per area than sun leaves (Valladares and Niinemets 2008). The rapid increase in chlorophyll content after the transference from full to partial sunlight can be

interpreted as a mechanism of acclimation of the photosynthetic apparatus since the time after transference was not sufficient for the plants to produce new leaves morphologically acclimated to shade.

In addition to reporting photosynthetic variables, other longer-term studies of the effects of flooding in tropical trees have included growth responses (Davanso et al. 2002; Mielke et al. 2003; Lavinsky et al. 2007; Medina et al. 2009). In this study, the authors did not analyze the effects of flooding on plant growth because the duration of the study (about 2 weeks) was too short to expect sufficient plant growth to allow for comparisons between treatments. However, in a previous study in which *E. uniflora* trees were pre-acclimated to full and partial sunlight and flooded for 36 days, soil flooding caused significant decreases in plant growth (Mielke and Schaffer 2010). Thus, the long-term effects of flooding and decreases in light availability caused by occasional heavy rains warrants further investigation.

There are many references predicting increases in the frequency of heavy rains in tropical and subtropical regions as a result of global climate change (Vera et al. 2006; Marengo et al. 2009). Field studies have shown that cloud cover and changes in light intensity during rainy season limits net CO₂ assimilation in sun leaves (Zotz and Winter 1994; Graham et al. 2003). Thus, the changes in light caused by the presence of thick clouds can be a disadvantage for CO₂ assimilation and growth of trees. On the other hand, the increased frequency of heavy rains may be followed by an increase in the frequency of floods (Michener et al. 1997; Milly et al. 2002; Hirabayashi et al. 2008). In addition, the existence of interactive effects of light environment and soil flooding on photosynthesis and growth of temperate and tropical tree species has been reported by several authors (Wagner and Dreyer 1997; Gardiner and Krauss 2001; Lavinsky et al. 2007; Mielke and Schaffer 2010a, b). Despite some limitations in this experimental procedure, especially in relation to the continuous exposure of flooded plants to low and high light and the absence of a precise control of light intensity during the experiment, the results of chlorophyll fluorescence, leaf chlorophyll content and leaf gas exchange were sufficient to support the hypothesis that changes in light availability can alleviate the harmful effects of soil flooding on photosynthesis of *E. uniflora* leaves acclimated to full sun. To explore the extensiveness of the hypothesis tested in this study, additional studies should analyze the effects of changes in light availability and soil flooding at the whole plant and ecosystems levels. Furthermore, similar studies with other tree species should be conducted to analyze the impacts of changes in rainfall on the ecophysiological responses of cultivated and native species in areas prone to soil flooding.

In summary, these results indicate that changes in light availability during soil flooding can alleviate the effects of flooding stress on photosynthesis in *E. uniflora* seedlings acclimated to full sunlight, demonstrating that the responses of trees to flooding stress may be dependent on changes in light environment during heavy rains. Interactions between flooding stress and sun/shade acclimation on photosynthesis and growth of trees should be considered in studies aimed at predicting changes in the plant production and native vegetation distribution as a function of changes in rainfall associated with global climate change.

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