**ORIGINAL PAPER**



# **Determination of stoichiometry and stability constants of iron complexes of phenanthroline, Tris(2‑pyridyl)‑s‑triazine, and salicylate using a digital camera**

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#### **Abstract**

The determination of stoichiometric ratios and stability constants of metal–ligand complexes is of great importance for physical, chemical, biochemical, and environmental studies and often require relatively expensive and sophisticated instruments. Herein, we introduce a simple, accurate, and low-cost method for determining these parameters by a conventional digital camera. The stoichiometric ratios and stability constants of iron complexes of 1,10-phenanthroline, 2,4,6-Tris(2-pyridyl) s-triazine, and salicylate were determined, as model examples, using the molar ratio and the continuous variation methods. Digital images of solutions with various metal–ligand ratios were captured and analyzed, and the  $Y_{xy}$  color absorbance and the Δ*E*<sub>LUV</sub> color difference parameters were used as convenient analytical signals that favorably compete with conventional spectrophotometric signals. For the three studied iron complexes, the results of stoichiometric ratios and stability constants obtained from digital images were in excellent agreement with the spectrophotometric and the previously reported literature's data.

**Keywords** Digital imaging ·  $Y_{xy}$ color absorbance ·  $\Delta E_{LUV}$  color difference · Stability constant · Metal complexes · Absorption spectra

# **Introduction**

The stoichiometry and stability constants of metal complexes are important parameters for physical, chemical, biochemical, and environmental studies. Diferent techniques were used for the determination of these parameters including UV–VIS spectrophotometry (Carmody [1964;](#page-5-0) Chattopadhyaya and Singh [1974](#page-5-1); Dömötör et al. [2018;](#page-6-0) Gamov et al. [2019](#page-6-1); Kluska et al. [2018;](#page-6-2) Kocyla et al. [2017](#page-6-3); Ravichandran et al. [2014\)](#page-6-4), spectrofuorimetry (Chen et al. [2015;](#page-6-5) Cuprys et al. [2018](#page-6-6)), potentiometry (Dömötör et al. [2018](#page-6-0); Friend and Wall [2019;](#page-6-7) Jakusch [2018](#page-6-8); JANRAO et al. [2014](#page-6-9); Kluska et al. [2018](#page-6-2); Kocyla et al. [2017;](#page-6-3) S. Al–Farhan [2018\)](#page-5-2), voltammetry

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 $\boxtimes$  Ashraf A. Mohamed aamohamd@sci.asu.edu.eg (Geiger et al. [1991](#page-6-10); Kamyabi et al. [2016\)](#page-6-11), conductometry (Kazemi et al. [2016](#page-6-12); Al–Farhan [2018\)](#page-5-2), gel chromatography (Yoza [1977](#page-6-13)), ion exchange chromatography (Wacker and Seubert [2014\)](#page-6-14), liquid–liquid extraction (Omoto and Wall [2017\)](#page-6-15), electrophoresis (Ansorge et al. [2018](#page-5-3); Holm et al. [2013;](#page-6-16) TEWARI' et al. [1993](#page-6-17)), and isothermal calorimetry (Holm et al. [2013;](#page-6-16) Kluska et al. [2018](#page-6-2)). These techniques often require sophisticated and relatively expensive instruments, where the very popular UV–VIS spectrophotometry is most commonly used. Stoichiometry and stability constants can be determined spectrophotometrically by several methods including the Job's continuous variation method (Carmody [1964](#page-5-0)), the molar ratio method, and the Bjerrum method. (JANRAO et al. [2014](#page-6-9)).

Recently, digital imaging devices, e.g., digital cameras (Mohamed and Shalaby [2019;](#page-6-18) Mohamed et al. [2018](#page-6-19)), smartphone cameras (Dutta [2019;](#page-6-20) Mohamed and Shalaby [2019](#page-6-18); Mohamed et al. [2018\)](#page-6-19), and scanners (Mohamed and Shalaby [2019](#page-6-18); Mohamed et al. [2018](#page-6-19); Shokrollahi et al. [2015](#page-6-21)), served as sensors for colorimetric analysis. Captured images were analyzed to yield the Red, Green, and Blue (RGB) channel intensities (de Morais et al. [2016](#page-6-22); Huangfu

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et al. [2019;](#page-6-23) Mohamed and Shalaby [2019;](#page-6-18) Mohamed et al. [2018\)](#page-6-19). Recently, Morais et al. (de Morais et al. [2016\)](#page-6-22) applied the red color channel intensity as a signaling tool for the determination of the Iron-Phenanthroline binding constant. However, the channel intensities of the non-uniform RGB color space were characterized by poor calibration linearity (Mohamed and Shalaby [2019](#page-6-18); Mohamed et al. [2018\)](#page-6-19) and therefore were mathematically converted into the  $Y_{xy}$ and  $\Delta E_{\text{LUV}}$  color space signals that gave excellently linear calibration responses that favorably competed with signals of sophisticated spectrophotometers (Mohamed and Shalaby [2019](#page-6-18); Mohamed et al. [2018\)](#page-6-19).

Here we describe a low-cost, simple, sensitive, and accurate method for the determination of the stoichiometry and stability constants of metal–ligand complexes using digital image-based analysis (DIBA). The stoichiometric ratios and stability constants of the Iron complexes with 1,10-Phenanthroline (Phen), 2,4,6-Tris(2-pyridyl)-s-triazine (TPTZ), and Salicylate (SAL) were determined using the molar ratio and the continuous variation methods. A conventional digital camera was used to capture images of cuvettes containing reacting solutions of the iron complexes with various Fe/L ratios or mole fraction. Captured images were analyzed to obtain the non-uniform RGB color space parameters that were converted to the corresponding  $Y_{xy}$ and  $\Delta E_{LUV}$  color space analytical signals.(Mohamed and Shalaby [2019](#page-6-18); Mohamed et al. [2018\)](#page-6-19) The  $Y_{xy}$  color absorbance and  $\Delta E_{LUV}$ color diference values were plotted against the molar ratio or the mole fraction and the stoichiometric ratio and stability constant values were calculated.

## **Theoretical**

The formation of mononuclear complexes can be represented by the following chemical equilibrium  $M + nL \leftrightarrow$ MLn.

The stability constant  $\beta_n$  is given by Eq. [\(1](#page-1-0)). In most cases one of the reaction constituents takes part in other equilibrium reaction, e.g., the ligand can be protonated or competed by other metal ions present, so the real concentration of the free ligand may difer widely in solutions with the same total ligand concentration. In this case, the apparent stability constant is called a conditional stability constant " $\beta_n''$ " and the overall stability constant can be calculated using Eq. ([2\)](#page-1-1) (Chattopadhyaya and Singh [1974;](#page-5-1) Inczédy [1976\)](#page-6-24). Where  $\alpha_{L(H)}$  is the side reaction function of ligand protonation and can be calculated using Eq. ([3\)](#page-1-2) (Chattopadhyaya and Singh [1974](#page-5-1)).

$$
\beta_n = \frac{[ML_n]}{[M][L]^n} \tag{1}
$$

<span id="page-1-1"></span>
$$
\beta_n = \beta'_n (\alpha_{L(H)})^n \tag{2}
$$

<span id="page-1-2"></span>
$$
\alpha_{L(H)} = 1 + \frac{\left[H^+\right]}{K_1} + \frac{\left[H^+\right]^2}{K_1 K_2} + \dots
$$
\n(3)

The conditional and overall stability constants can be determined from the plots of the molar ratio and continuous variation methods, where at the point of stoichiometry, the ratio between the true observed absorbance (or color absorbance, or color difference)  $(A_t)$  to that extrapolated by the tangents  $(A_{ex})$  equal to the mole fraction of the metal ion or the ligand in the complex, Eq. ([4\)](#page-1-3). Thus, the concentration of the complex species can be expressed by Eq. ([5\)](#page-1-4), and the concentration of the free metal ion and the free ligand can be expressed by Eqs.  $(6)$  $(6)$  and  $(7)$  $(7)$ , respectively. Thus, by substitution in Eq.  $(1)$  $(1)$ , the conditional stability constant can be calculated from Eq. [\(8](#page-1-7)).(Inczédy [1976](#page-6-24))

<span id="page-1-3"></span>
$$
\frac{A_t}{A_{ex}} = \frac{[ML_n]}{C_M} = \frac{n[ML_n]}{C_L} \tag{4}
$$

<span id="page-1-4"></span>
$$
\left[ML_n\right] = \frac{A_t}{A_{ex}} C_M \tag{5}
$$

<span id="page-1-5"></span>
$$
[M] = C_M - \frac{A_t}{A_{ex}} C_M \tag{6}
$$

<span id="page-1-6"></span>
$$
[L] = C_L - n \frac{A_t}{A_{ex}} C_M \tag{7}
$$

<span id="page-1-7"></span>
$$
\beta'_{n} = \frac{[ML_{n}]}{[M][L]^{n}} = \frac{\frac{A_{i}}{A_{ex}}C_{M}}{[C_{M} - \frac{A_{i}}{A_{ex}}C_{M}][C_{L} - n\frac{A_{i}}{A_{ex}}C_{M}]^{n}}
$$
(8)

where  $C_M$  and  $C_L$  are the analytical concentrations of the metal and ligand, whereas [*M*], [*L*], and [*MLn*] are the equilibrium concentrations of the metal, ligand, and complex, respectively.

## **Experimental**

#### **Apparatus and software**

<span id="page-1-0"></span>Digital Image-Based Analysis (DIBA) measurements were made using a simple homemade platform. (Mohamed and Shalaby [2019](#page-6-18); Mohamed et al. [2018\)](#page-6-19) In brief, the platform consisted of (1) two matched 10 mm glass cells (2) the digital camera and (3) a white cartoon paper as background diffuser. The difuser, the 3D printed cell-holder and the camera

were fixed on a  $20 \times 20$  cm wood plate; each of them was 5-cm distance apart. Digital images were captured, on the bench of our laboratory, using a Canon PowerShot A810 digital camera that is equipped with a 16.0 Mega Pixel CCD sensor, where the conventional fuorescent daylight-lamp fxed to the ceiling served as the light source. (Mohamed and Shalaby [2019](#page-6-18); Mohamed et al. [2018\)](#page-6-19) A conventional HP-EliteBook 2540P notebook running under windows 10 was used for treatment of data and analysis. Photoshop CC 2017 and imageJ 1.52c software were used for digital image cropping and RGB channel intensities calculations, respectively. The obtained RGB intensities were converted into the corresponding coordinates of the  $Y_{xy}$  and LUV color spaces using the free colormine online converter (Colormine color converter. [2019\)](#page-6-25).

Spectrophotometric measurements, for comparison only, were made on a Shimadzu 1650 UV/VIS spectrophotometer controlled by an UVProbe-2.5 software (Kyoto, Japan).

Eppendorf 10-100 and 100-1000 μL vary-pipettes (Westbury, NY, USA) and a calibrated EDT pH-mV meter (Dover Kent, UK) were used.

#### **Materials and chemicals**

All reagents were of ACS grade and were used as received from Sigma-Aldrich (St. Louis, MO, USA), Merck (Darmstadt, Germany) or BDH (Poole, UK). Unless otherwise stated, de-ionized water and aqueous solutions were used throughout. *1,10*-phenanthroline, *2,4,6*-Tris(2-pyridyl) s-triazine, sodium salicylate, ferrous ammonium sulfate hexahydrate, ferric ammonium sulfate dodecahydrate, and L-ascorbic acid, sodium acetate, sodium nitrate, and concentrated  $H_2SO_4$ , concentrated  $HNO_3$  were also used.

## **Determination of stoichiometry and stability constants of the iron complexes**

For the determination of Fe(II)-Phen stoichiometry using the molar ratio method, the following solutions were used: (1) 100 mL solutions of Fe(II)  $6.00 \times 10^{-4}$  mol L<sup>-1</sup>, and (2) 100 mL Phen solution  $6.00 \times 10^{-4}$  mol L<sup>-1</sup>. In a series of dry 10 mm path length glass spectrophotometric cells, 200 µL aliquots of Fe(II) were mixed with 300 µL of 1.0% *w/v* ascorbic acid [to reduce any Fe(III) traces present in the Fe(II)] and 300 µL of 1.0 mol L<sup>-1</sup> acetate buffer (pH=3.5). Increasing volumes of Phen (0 to 1200 µL) were then added to the reaction cuvettes to give increasing [Phen]/[Fe] ratios from 0.0 to 6.0. The reacting solutions were diluted with water to exactly 3000 µL and then mixed well. All measurements were made at 25 °C with the ionic strength adjusted to 0.1 mol  $L^{-1}$  using sodium nitrate solution.

For the determination of Fe(II)-Phen stoichiometry using the Job's method,  $3.00 \times 10^{-4}$  mol L<sup>-1</sup> solutions of Fe(II) and Phen were used. In another series of dry 10 mm path length glass spectrophotometric cells, various volumes of Fe(II) (100 to 1900  $\mu$ L) and Phen (1900 to 100  $\mu$ L) were mixed to give the desired mole fractions. A 300 µL of each of ascorbic acid and acetate buffer ( $pH = 3.5$ ) was added, and the final volume of each cuvette was then diluted to 3000 µL with water and the reacting solutions were mixed well. Similarly, all measurements were made at 25 °C with the ionic strength adjusted to 0.1 mol  $L^{-1}$  using sodium nitrate solution.

The molar ratio and the continuous variation methods were similarly followed for the Fe(II)-TPTZ and Fe(III)-SAL systems. However, the appropriate experimental conditions of the respective procedures were used. Namely, for the Fe(II)-TPTZ system,  $3.80 \times 10^{-4}$  and  $1.00 \times 10^{-4}$  mol L<sup>-1</sup> Fe(II) solutions were used for the molar ratio and the continuous variation methods, respectively. Moreover, acetate bufer of pH 4.5 was used. However, for the Fe(III)-SAL system,  $1.80 \times 10^{-3}$  mol L<sup>-1</sup> Fe(III) solution was used for both the molar ratio and the continuous variation methods. Further, 50 µL of 0.10 mol  $L^{-1}$  nitric acid was used to adjust the medium acidity, in the latter system.



<span id="page-2-0"></span>**Fig. 1** Digital images of Fe(II)-phenanthroline complex at diferent [phenanthroline]/[Fe(II)] molar ratio (**a**) and the cropped Area of interest (**b**). Except for the studied variable, other conditions were those given in th recommended procedure. Captured images were arbitrarily compressed to ft into the page margins; however, for image analysis, the original uncompressed images were used

# **Results and discussion**

#### **Spectra and Image acquisition**

In the molar ratio method, solutions having various [*L*]/[*M*] molar ratios were prepared for the Fe(II)-Phen, Fe(II)-TPTZ, and Fe(III)-SAL complexes. A portion from each solution was placed in the spectrophotometric cell and was placed in our homemade platform to capture respective digital images that were automatically outputted in JPEG format with 180 dpi resolution. From each image, a squared area  $(360 \times 360$  pixel) was cropped as an area of interest (AOI), Fig. [1](#page-2-0) and Figs. S1–S2. However, any other predefned AOI can be selected. Thereafter, the AOI was analyzed, using the freeware ImageJ, to obtain the RGB intensities that were converted to the corresponding  $Y_{xy}$  and  $\Delta E_{\text{LUV}}$  parameters, based on their excellent responses compared to the original RGB intensities (Mohamed and Shalaby [2019](#page-6-18); Mohamed et al. [2018](#page-6-19)). The color absorbance of x or y parameters and the color difference,  $\Delta E_{\text{LUV}}$ , were calculated from Eqs. ([9\)](#page-3-0) and ([10\)](#page-3-1), respectively.

<span id="page-3-0"></span>
$$
A_{xy} = \left| \log \left( I_o / I \right) \right| \tag{9}
$$

<span id="page-3-1"></span>
$$
\Delta E_{\text{LUV}} = \left[ (\Delta L)^2 + (\Delta U)^2 + (\Delta V)^2 \right]^{\frac{1}{2}}
$$
\n(10)

where  $I_0$  and *I* are the *x* or *y* value of the blank and sample, respectively, whereas  $\Delta$  means the difference in the parameter value between the sample and blank, respectively. For validation, the spectrophotometric cell was placed in the spectrophotometer and spectral curves were recorded, Fig. [2](#page-3-2)a–c. The [L]/[M] ratio was plotted against the color absorbance of the x or y parameters, Fig. [2d](#page-3-2)–f, or against the color difference  $(\Delta E)$  calculated from the LUV color space parameters, Fig. [2g](#page-3-2)–i.



<span id="page-3-2"></span>**Fig. 2** Spectra obtained at diferent L:M ratio for the studied Fe complexes with Phenanthroline, TPTZ, and Salicylate (**a**)–(**c**), Molar ratio graphs plotted using spectrophotometric absorbance at the appro-

priate wavelength and color absorbance of *x* or *y* parameter  $(d)$ – $(f)$ , and color diference ΔELUV (**g**)–(**i**). Other conditions were those of Fig. [1](#page-2-0)



<span id="page-4-0"></span>**Fig. 3** Digital images of Fe(II)-phenanthroline complex at diferent Fe(II) mole fraction (**a**) and the cropped Area of interest (**b**). Other conditions were those of Fig. [1](#page-2-0)

For comparison and validation, the [*L*]/[*M*] ratio was simultaneously plotted against the spectrophotometric absorbance at the appropriate wavelength for each complex, Fig. [2d](#page-3-2)–i. The DIBA results showed 1:3, 1:2 and 1:1 (*M*:*L*) ratios for the Fe(II)-Phen, Fe(II)-TPTZ and Fe(III)- SAL systems, respectively, in excellent agreement with the spectrophotometric data.

Similarly, in the Job's method of continuous variation, solutions having various mole fractions were prepared for the Fe(II)-Phen, Fe(II)-TPTZ, and Fe(III)-SAL complexes and similarly treated as those of the molar ratio method. Images of these solutions were given in Fig. [3](#page-4-0) and Figs. S3–S4, for the Fe(II)-Phen, Fe(II)-TPTZ, and Fe(III)-SAL complexes, respectively. The Job's plots, Fig. [4a](#page-4-1)–f, of the studies systems exhibited maxima at 0.25, 0.33 and 0.5, for the Fe(II)-Phen, Fe(II)-TPTZ, and Fe(III)-SAL complexes, respectively, showing that iron reacts with phen, TPTZ, and SAL in molar ratios of 1:3, 1:2 and 1:1, respectively. These data refect the excellent agreement between the spectrophotometric and DIBA measurements.

It's worthy to mention that, in addition to the formed trisphenanthroline-Iron(II) complex ion, other monophenanthroline and diphenanthroline complex ions can be formed. However, compared to the very stable trisphenanthroline complex, the mono- and diphenanthroline complexes are relatively unstable and its existence can be neglected at high phanenthroline-to-Iron(II) ratios. For example, the monophenanthroline-Fe(II) complex is formed only at large



<span id="page-4-1"></span>**Fig. 4** Continuous variation graphs plotted using the spectrophotometric absorbance and the color absorbance of x or y coordinate of Yxy color (**a**)–(**c**) and color diference ΔELUV (**d**)–(**f**) for the studied

Fe complexes with Phenanthroline, TPTZ, and Salicylate. Conditions and symbols were those of Fig. [1](#page-2-0)

Complex	$\text{Log}\beta'^a$ (I = 0.1, 25 °C)			$\text{Log}\beta^b$ (I = 0.1, 25 °C)			Literature value
	<b>DIBA</b> $(A_{Yxy})$	<b>DIBA</b> $(\Delta E_{LUV})$	Spectropho- tometry	DIBA $(A_{Yxy})$	<b>DIBA</b> $(\Delta E_{LUV})$	Spectropho- tometry	$\text{Log }\beta$ $(I=0.1, 25^{\circ}C)$
$Fe(II)$ -phenanthroline	17.1	17.1	17.2	21.18	21.18	21.28	21.3 $Mckenzie$ 1955 <sup><math>\prime</math></sup> 21.2 (McBRYDE 1978)
$Fe(II)$ - Tris $(2$ -pyridyl)-s-triazine	12.30	12.03	12.19	12.39	12.12	12.28	12.3 Martell and Smith 1989 <sup>)</sup>
Fe(III)-Salicylate	5.42	5.52	5.43	16.31	16.41	16.32	16.3 Martell and Smith 1977 <sup>)</sup>

<span id="page-5-4"></span>**Table 1** Stability constant values of the studied complexes obtained using DIBA and spectrophotometric methods

a Conditional stability constant

b Overall stability constant

excess of ferrous (Kolthoff et al. [1950](#page-6-26)), so that, it can be formed only at iron mole fraction larger than 0.5 and [phenanthroline]/[Fe] molar ratio less than 1.0.

Further, the conditional and overall stability constants of the three studied complexes were determined from the molar ratio plots using Eqs. [\(8](#page-1-7)) and [\(2](#page-1-1)), respectively. The side reaction functions of each ligand protonation were calculated from Eq ([3](#page-1-2)) using the previously reported  $pK_a$  values for 1,10-phenanthroline (4.84) (CRC handbook of chemistry and physics [2017\)](#page-6-27), TPTZ (3.53 and 2.73) (Martell and Smith [1989](#page-6-28)), and Salicylic acid (13.6 and 2.98) (CRC handbook of chemistry and physics [2017\)](#page-6-27). The resulting conditional and overall stability constants of the Fe(II)-Phen, Fe(II)- TPTZ, and Fe(III)-SAL complexes, based on spectrophotometric and DIBA measurements are given in Table [1.](#page-5-4) The data refects the excellent agreement not only between the DIBA results and the spectrophotometric measurements but also with the previously reported literature values (Chattopadhyaya and Singh [1974;](#page-5-1) Martell and Smith [1977,](#page-6-29) [1989](#page-6-28); McKenzie [1955\)](#page-6-30). Thus, our simple camera-based platform can be conveniently applied to the determination of complex stoichiometry and stability constants in a manner competing well with the sophisticated spectrophotometers showing its feasibility for practical application in undergraduate students' laboratories.

# **Conclusions**

The stoichiometric ratio and binding constant of metal complexes are important physical and analytical chemistry concepts. Conventional methods for the determination of these parameters often require expensive and/or bulky instruments. Herein, we reported a low-cost and simple approach based on digital imaging. The stoichiometric ratio and binding constants of three iron complexes with 1,10-phenanthroline, 2,4,6-Tris(2-pyridyl)-s-triazine, and salicylate were determined using digital imaging and following the molar ratio and continuous variation methods. The color absorbance of non-uniform  $Y_{xy}$  color space and the color diference Δ*E*LUV derived from the uniform LUV color space were used as convenient analytical signaling tools. The  $Y_{xy}$  color absorbance and the  $\Delta E_{\text{LUV}}$  color difference signaling tools can be conveniently applied to investigate a wide range of various colored complexes including not only mononuclear complexes but also polynuclear complexes of various species. Moreover, the current results of composition and conditional and overall stability constants are in excellent agreement with those obtained using sophisticated spectrophotometers and specialized software packages and also in conformity to previously reported literature values. This is very advantageous under resource-limited settings.

## **Compliance with ethical standards**

**Conflict of interest** On behalf of all authors, the corresponding author states that there is no confict of interest.

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