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UV-spectrophotometry versus HPLC–PDA for dual-drug dissolution profiling: which technique provides a closer step towards green biowaiver concept? Novel application on the recent FDA-approved mixture Aleve pm

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Abstract

Dissolution testing is a very significant tool that adds in vivo relevance to in vitro analytical data, thus providing a realistic in vitro/in vivo correlation. Dissolution profiling serves as a predictor of biological performance since the rate limiting step in the absorption of any drug is the rate of release from its pharmaceutical formulation. Being a routine procedure in quality control laboratories for initial approval process and scaling-up, the development of more and more eco-friendly methods for dissolution monitoring is considered as a worldwide trendy goal. In the present contribution, a comparison was highlighted between the two analytical techniques of utmost importance in acquisition of dissolution profiles: UV-spectrophotometry and HPLC-PDA focusing on the greenness of each one for further consolidation of the biowaiver concept. Both techniques were applied on the recently FDA-approved combination naproxen sodium (NAPR) and diphenhydramine hydrochloride (DIPH) formulated as Aleve pm[®] tablets. For the first time, this binary mixture was analyzed by three UV-spectrophotometric methods. NAPR was directly determined by zero-order spectrophotometry at 330 nm, where the spectrum of DIPH shows zero contribution. Whereas DIPH was determined by three simple methods exploiting the ratio spectra calculated using the spectrum of 20 µg/ml NAPR as a divisor. The first proposed method was ratio difference (RD), the second was ratio subtraction, and the last one was derivative ratio method. Being the simplest method, RD was the spectrophotometric method of choice that was applied in monitoring the dissolution of DIPH from Aleve pm® tablets. Although RD method has been widely described in many publications dealing with pharmaceutical analysis, it is the first article that opens a new horizon for RD method in the way of being a real-life application rather than only a method for determination. The second technique was a previously published HPLC-PDA, in which dissolution was then monitored by calculating the peak area of each component drug over time. Methods' validation was performed agreeing with the ICH guidelines. The advantages and challenges of each technique are discussed in a side-by-side comparison giving a key to recognize which one can positively influence environmental well-being.

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Graphical abstract



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Introduction

The international regulatory authorities such as the United States Pharmacopeia (USP), Food and Drug Administration (FDA), and International Conference on Harmonisation (ICH) stated that drug absorption from a solid oral dosage form depends on its release from the drug product (drug dissolution) (ICH 2010; USP 2011; Administration 2015). In vitro dissolution testing has been utilized as a predictor of drug's in vivo performance in the way that limits the need to costly and time-consuming human studies, giving a key to the biowaiver notion (Anand et al. 2011; Administration 2015; Korany et al. 2018a). Based on this general consideration, in vitro dissolution studies for solid oral dosage forms, such as tablets and capsules, are performed routinely in quality control laboratories to judge the quality of a drug product, also to evaluate the newly developed pharmaceutical formulations (Administration 2015). Recently, the term "green analytical chemistry" (GAC) was raised and gained a growing acceptance originated from the continuous worldwide strive towards a safe and healthy environment (Guardia and Garrigues 2012). The well-known GAC principles were introduced by Anastas and Warner in 1998. They presented a guiding framework applying to all disciplines of chemistry nevertheless analytical chemistry to reach the compromise between increasing quality of the results and raising environmental friendliness (Anastas and Eghbali 2010). In general, GAC prefers instrumental methods; however, they need to be ameliorated via reducing the amounts of reagents consumed and waste generated for increasing operator safety (Gałuszka et al. 2013; Korany et al. 2017a, b, 2018b). Therefore, many pharmaceutical scientists have directed their efforts on the way of greening most of analytical practices particularly those having a routine manner, consequently greening-up the dissolution process would be highly desirable.

USP stated that the standard methods for determining the extent of drug release from tablets or capsules are UVspectroscopy and high-performance liquid chromatography (HPLC), in which dissolution process is monitored via sampling at definite time intervals (every 5 min for 30 or 45 min) followed by analysis of the active principle (2011). In the present research, the greenness of each technique was investigated using a mixture of NAPR and DIPH present as OTCtablets Aleve pm[®]. Naproxen is a significant member of the class of non-steroidal anti-inflammatory drugs, it is used to treat pain and minor body aches (Uysal and Tunçel 2004). Diphenhydramine belongs to first-generation antihistaminic drugs; it is mainly used as a night-time sleep aid due to its sedating effect (Ge et al. 2011). Reviewing the literature carefully revealed that one spectrophotometric method has been reported for the analysis of this mixture yet although it is a widely used OTC drug (Mabrouk et al. 2015). However, other methods such as electrochemical (Abd El-Rahman et al. 2017; Shehata et al. 2017), high-performance thin-layer chromatographic (HPTLC) (Bhole et al. 2015), and HPLC/PDA (Nalluri et al. 2015) have been recently reported for simultaneous determination of both drugs in their binary formulation.

In this work and for the first time, three spectrophotometric methods were developed on this mixture based on ratio spectra manipulation; they are RD, RS, and DD¹. Upon application of the proposed methods, RD was thought to be the most simple and reliable method. Plenty of publications have exploited the recently developed RD method in drug assay purposes (Lotfy and Abdel-Monem Hagazy 2012; Darwish et al. 2013; Saad 2015; Abdel-Halim et al. 2016; Abd El-Rahman et al. 2017; Ragab et al. 2018), yet our goal was to expand it beyond determination towards a real application. Thus, RD method was utilized in further dissolution investigations in parallel to a reported HPLC method (Shehata et al. 2017). This study enlightens the opportunities and limitations of the two different methodologies for both determination and dissolution tracking of NAPR and DIPH. The novelty of the current study arises from performing a side-by-side comparison with the aim of evaluating the sustainability of the analytical technique.

Experimental

Instrument

UV-visible spectrophotometer was SHIMADZU dual beam (Kyoto, Japan), model UV-1650 PC. The utilized software was UV-Probe personal spectroscopy software version 2.21 (SHIMADZU). The spectral band was 2 nm and scanning speed was 2800 nm/min with 0.1 nm interval.

Drugs' dissolution was tested using a VanKel (VK 7000 USA) equipped with standard USP type-I basket.

Samples

Reference samples

Diphenhydramine hydrochloride and naproxen sodium pure standards were purchased from Bayer Co.; the purity of each was certified to be more than 99.99%.

Pharmaceutical formulation

Aleve pm[®] tablets were formulated by Bayer Pharmaceutical Company, Germany. Batch No. NAA5FPO. Each tablet is labeled to contain 220 mg and 25 mg of NAPR and DIPH, respectively.

Solutions

Naproxen sodium and diphenhydramine hydrochloride stock standard solutions with a concentration of 500 and 100 μ g/ml, in distilled water, were prepared in two separate 100-ml volumetric flasks by dissolving 0.05 and 0.01 g of pure NAPR and DIPH, respectively.

Procedure

UV-spectrophotometric methods' development and calibration

Aliquots (0.4–5.2 ml) of NAPR and (0.4–4 ml) of DIPH standard solutions (500 and 100 μ g/ml, respectively), were accurately transferred into two separate series of 10-ml volumetric flasks then completed to volume with distilled water. The prepared series of standard solutions were scanned from 200 to 400 nm and the zero-order (D^0) absorption spectrum of each solution was recorded using distilled water as a blank.

NAPR was determined directly by plotting its absorbance in the zero-order spectra (D^0) at $\lambda_{max} = 330$ nm (where DIPH has no contribution) against its corresponding concentrations then the regression equation was computed. However, for quantitative measurement of DIPH in presence of NAPR, zero-order absorption spectra of DIPH were divided by the spectrum of 20 µg/ml NAPR, subsequently three different spectrophotometric methods were applied on the recorded DIPH ratio spectra.

Ratio difference (RD) spectrophotometric method The difference in peak amplitudes of the ratio spectra (DIPH/ NAPR) at 246 and 256 nm ($\Delta P_{256-246}$) was plotted against the corresponding concentrations of DIPH (µg/ml) then the regression equation was computed.

Ratio subtraction (RS) spectrophotometric method From the previously obtained ratio spectra, the absorbance of NAPR in the extended region (a constant value) at 275– 350 nm was subtracted. The obtained spectra were then multiplied by the spectrum of the divisor (20 µg/ml NAPR). The re-produced spectra (the same as DIPH zero-order spectra) can be directly used for estimation of DIPH at its $\lambda_{max} = 220$ nm. Calibration was obtained by plotting the absorbance values versus the corresponding concentrations then the regression equation was computed.

Derivative ratio (DD¹) spectrophotometric method The first derivative of the previously stored ratio spectra was obtained with $\Delta\lambda$ of 8 nm and a scaling factor equals to 100. The peak amplitudes of the first derivative of the ratio spectra were measured at 228 and 268 nm. Calibration graphs relating the peak amplitudes of the DD¹ curves at 228 and 268 nm to the corresponding concentrations of DIPH were constructed, and the corresponding regression equations were computed.

To prepare laboratory-prepared mixtures containing different ratios of NAPR and DIPH, aliquots from their respective standard solutions (500 and 100 µg/ml) were transferred accurately into a series of 10-ml volumetric flasks and completed to the mark with distilled water. NAPR was determined directly at zero-order by measuring the absorbance of each laboratory-prepared mixture at 330 nm, and then the concentrations of the drug were computed from its corresponding regression equation. For determination of DIPH, zero-order absorption spectrum of each laboratory-prepared mixture was divided by the spectrum of 20 µg/ml of NAPR and then the three proposed spectrophotometric methods were applied on the recorded ratio spectra. The concentration of DIPH in each mixture was obtained from the corresponding regression equation using the same procedure mentioned in the previous section (UV-spectrophotometric methods' development and calibration).

Assay of NAPR and DIPH content in Aleve pm[®] tablets

Ten Aleve pm[®] tablets were separately weighed, powdered, and well mixed. An amount of powder equivalent to one tablet was accurately weighed and transferred into a 100ml beaker, sonicated for 15 min in 50 ml distilled water. The solution was filtered into 100-ml volumetric flask, and the residue was washed two times each using 5 ml distilled water and the volume was completed to the mark with the same solvent. Afterwards; 1 ml of the prepared solution was transferred into a 10-ml volumetric flask, completed to the mark with water, and analyzed using the proposed spectrophotometric methods. The concentration of each drug was calculated from the corresponding regression equation. The validity of the adopted methods was assessed by applying standard addition technique by spiking the pharmaceutical formulation with known amounts of standard drug powders. The recoveries of the added standards were then calculated after applying the proposed methods.

Dissolution testing

was used, the tablet was introduced into the basket, and the vessel content was agitated using a rate of 75 rpm.

Samples were withdrawn from the dissolution medium at 5-min time intervals, filtered, and the volume was completed with distilled water. NAPR's concentration was calculated directly by measuring absorbance values of the zero-order spectrum at 330 nm, while RD method was applied to get the concentration of DIPH. Percentage dissolution was then calculated for both drugs, and dissolution curves were plotted.

Results and discussion

Dissolution monitoring is a mandatory routine step for all solid oral dosage forms to ascertain that the active drug(s) is released at the desired rate to exert the required therapeutic effect (Costa and Lobo 2001). Moreover; dissolution testing has been extensively discussed by pharmaceutical scientists as a surrogate for in vivo bioequivalence studies which undoubtedly affect human's safety and health (Maggio et al. 2013). In this context, dissolution testing has a crucial role in regulatory decision-making and it may be used to waive in vivo bioequivalence study requirements, serving the purpose of biowaiver concept (Karam 2015). Officially stated methods for dissolution monitoring depend mainly on one of the well-established analytical techniques: UV-spectrophotometry and HPLC (2011). Recently, the analytical society has paid considerable attention to the usage of "Sustainable Analytical Procedures'' that meet the requirements of GAC (Welch et al. 2010; Gałuszka et al. 2013; Abd El-Rahman et al. 2016). Since the two methodologies are feasible in most of the quality control laboratories worldwide, this research addresses their implementation in the analysis and dissolution testing of Aleve pm[®] tablets. Additionally, a fair comparison was held between both techniques to get to the greenly recommended one that adheres more closely to the GAC principles.

UV-spectrophotometric methods development and optimization

The zero-order absorption spectra of NAPR and DIPH shows overlapping that hinders direct determination of DIPH; however, NAPR spectrum is more extended in the way that allows its direct determination in presence of DIPH at $\lambda_{max} = 330$ nm, where DIPH shows no interference (Fig. 1). Accordingly, three spectrophotometric methods manipulating ratio spectra are adopted in this work for the simultaneous estimation of DIPH in presence of NAPR in their laboratory-prepared mixtures and pharmaceutical formulation. For estimation of NAPR, a linear relationship was plotted between the absorbance at 330 nm and the corresponding **Fig. 1** Zero-order absorption spectra of NAPR 20–260 μ g/ml (red dotted line) and DIPH 4–40 μ g/ml (green solid line), showing calibration of NAPR at 330 nm



Table 1 Validation and regression parameters of the proposed methods

Validation parameter	UV-spectrophotometric methods							
	NAPR	DIPH						
	D ₀ at 330 nm	RD	RS	DD1				
				228 nm	268 nm			
Linearity range	20–260 µg/ml	4–40 µg/ml						
Slope	0.0065	0.0188	0.0354	0.0827	0.0279			
Intercept	0.0024	0.0076	0.0083	0.0208	0.00004			
SE of slope	0.00001	0.00008	0.00008	0.00057	0.00017			
SE of intercept	0.00197	0.002	0.00058	0.01444	0.00432			
r	1	0.9999	0.9999	0.9998	0.9999			
LOD ^a	1.19	0.40	0.33	0.67	0.59			
LOQ ^a	3.62	1.22	1.00	2.02	1.79			
Accuracy (mean \pm SD)	100.40 ± 0.488	99.00 ± 0.476	99.70 ± 0.842	99.91±1.115	100.20 ± 0.697			
Specificity ^b (mean \pm SD)	100.14 ± 1.099	100.49 ± 1.574	100.25 ± 1.078	100.38 ± 1.214	100.50 ± 1.134			
Precision ^c (RSD%)								
Repeatability	0.908	0.621	0.719	0.420	0.457			
Intermediate precision	1.266	1.025	1.644	0.857	1.045			

^aThe ratio of standard deviation of residuals (Sy/x) to the slope was used for calculating LOD and LOQ

^bSpecificity was assessed by the analysis of different laboratory-prepared mixtures containing different ratios of each drug

^cPrecision was evaluated by measuring the response of three concentrations of each drug (30, 50 and 70 for NAPR; 15, 20, and 30 for DIPH) three separate times on the same day (repeatability) and on 3 different days (intermediate precision)

concentrations, linearity, regression, and validation parameters are illustrated in Table 1.

Ratio difference (RD) spectrophotometric method

Ratio difference method was newly proposed as a simple method for determination of mixture compounds with overlapping spectra, having the advantage of slight mathematical manipulation of spectral data (Elzanfaly et al. 2012; Eissa 2017). The method includes dividing the spectrum of the analyte by a divisor concentration of the interfering substance, a ratio spectrum will be obtained, and a linear relationship between the difference in amplitudes at any two wavelengths and the corresponding concentrations of the analyte will result. Meanwhile, the ratio spectrum of the interferent will have a constant amplitude parallel to the x-axis, so at any two wavelengths it shows zero difference in amplitudes (Riad et al. 2015) (Fig. 2).

The method includes two critical steps: the first is the selection of the divisor concentration as it should provide minimal noise and maximum sensitivity. Different divisor concentrations of NAPR were tried and 20 μ g/ml showed the best results. The second step is the choice of the wavelength pair at which measurements would be recorded. Any two values of wavelength with different amplitudes in the ratio spectrum can be used provided that good linearity is present at each of them. Many wavelength pairs were tried

and 246–256 nm showed the best results. Linear correlation was obtained between the differences in peak amplitudes at 246–256 nm and the corresponding DIPH concentrations.

Ratio subtraction (RS) spectrophotometric method

Ratio subtraction method is a successfully applied spectrophotometric method for resolving binary mixtures provided that the spectrum of one component is more extended than the other (Darwish et al. 2011). The method was used for the determination of DIPH in its binary mixture with NAPR that have a more extended spectrum. In this method, the ratio spectra of the mixture can be expressed as DIPH/ NAPR + a constant (NAPR/NAPR'). Subtraction of the values of these constants in the extended region (275–350 nm) is followed by multiplication of the obtained spectra by the divisor NAPR'. Finally, the original spectra of DIPH could be obtained as shown in Fig. 3, and DIPH can be directly measured at its $\lambda_{max} = 220$ nm. The obtained absorbance values were plotted against the corresponding DIPH concentrations then the regression parameters were computed.

Derivative ratio (DD¹) method

The method was designed by Salinas (1990), based on derivatization of the ratio spectra for resolving binary or ternary mixtures. The main advantage of this method

Fig. 2 Ratio spectra of DIPH 4–40 µg/ml (green solid line), NAPR 20 µg/ml (red dotted line), and a lab-mixture of 20 µg/ml NAPR and 40 µg/ml DIPH (red broken line) using 20 µg/ml NAPR as a divisor





Fig.3 a Zero-order absorption spectra of different NAPR and DIPH lab-mixtures (prepared as mentioned in Table 3). **b** Ratio spectra of the lab-mixtures using 20 μ g/ml NAPR as a divisor. **c** Ratio spectra

of the lab-mixtures after subtraction of the constant. **d** Re-obtained DIPH spectra after multiplication by the divisor (20 μ g/ml NAPR)

is that the whole spectrum of interfering substance is canceled, so any wavelength can be for calibration; thus, easy measurements can be done at the wavelength of highest sensitivity either a maximum or a minimum (Abd El-Rahman et al. 2015).

Different $\Delta \lambda$ and scaling factors were tested, where $\Delta \lambda = 8$ and a scaling factor = 100 were suitable to enlarge the signals of DIPH and to diminish error in readings (Fig. 4). The linearity of the peak amplitudes of the DD¹ curve at both 228 and 268 nm was studied. Linear relationships were obtained for DIPH, and the regression parameters were computed.

Validation of the three proposed spectrophotometric methods has been performed according to ICH guidelines (Conference et al. 2005), and the calculated linearity and validation parameters are summarized in Table 1.

Determination of laboratory-prepared mixtures containing different ratios of NAPR and DIPH

The results obtained upon analysis of the laboratoryprepared mixtures containing different ratios of NAPR and DIPH revealed that the proposed spectrophotometric methods can be successfully used for selective determination of NAPR and DIPH in presence of each other (Table 2).

Determination of NAPR and DIPH in Aleve pm® tablets

The proposed spectrophotometric methods were successfully applied for simultaneous determination of NAPR and DIPH contents in their combined dosage form (Aleve pm[®] tablets),

Fig. 4 DD¹ spectra of DIPH 4–40 µg/ml using 20 µg/ml NAPR as a divisor



Table 2Determination ofNAPR and DIPH in laboratory-
prepared mixtures with the
proposed spectrophotometric
methods

Ratio (NAPR:DIPH)	NAPR conc. (µg/ml)	DIPH conc. (µg/ml)	Recovery % ^a					
			NAPR	DIPH				
			D_0 at 330 nm	RD	RS	DD1		
						228 nm	268 nm	
2:1	20	10	100.46	101.91	99.63	99.66	100.70	
1:1	20	20	98.15	98.83	100.24	100.44	101.78	
1:2	20	40	101.23	101.94	98.43	98.49	98.74	
3:1	60	20	99.90	99.10	100.81	101.83	100.71	
4:1	40	10	101.00	99.26	101.33	100.39	99.63	
8.8:1	88	10	100.10	101.91	101.05	101.48	101.42	
Mean			100.14	100.49	100.25	100.38	100.50	
SD			1.099	1.574	1.078	1.214	1.134	
RSD%			1.097	1.567	1.076	1.209	1.129	

^aAverage of three determinations

Table 3 Quantitative determination of NAPR and DIPH in Aleve pm[®] tablets by the proposed methods and the results of standard addition technique

Aleve pm [®] tablets (batch no. NAA4FPO)	UV-spectrophotometric methods						
	NAPR	DIPH					
	<i>D</i> ₀ at 330 nm	RD	RS	DD1			
				228 nm	268 nm		
R% of dosage form ^a ± RSD	101.53 ± 0.552	98.85 ± 0.938	100.19 ± 1.076	100.49 ± 0.674	98.87±0.604		
R% of standard added ^a ± RSD	99.88 ± 1.004	100.52 ± 0.780	101.24 ± 0.442	100.45 ± 1.261	99.01 ± 0.864		

^aAverage of three determinations

and standard addition technique was performed (Table 3). To examine the validity of the proposed methods, the obtained results were statistically compared to those of the reported HPLC method, and no significant differences were obtained (Table 4).

Table 4	Statistical analysis	of the results obt	ained by the propos	sed methods and t	the reported HPLC	C method for the de	etermination of NA	PR and
DIPH in	n pure powder form							

Parameter	NAPR		DIPH					
	D_0 at 330 nm	Reported method ^a	RD	RS	DD ₁		Reported method ^a	
					228 nm	268 nm		
Mean	100.40	99.66	99.00	99.70	99.91	100.20	100.12	
Variance	0.238	0.756	0.227	0.709	1.243	0.486	0.528	
n	7	3	7	7	7	7	3	
Student's t test ^b	1.769 (2.262)		2.152 (2.262)	0.747 (2.262)	0.295 (2.262)	0.165 (2.262)		
F value ^b	3.176 (5.14)		2.326 (5.14)	1.343 (19.33)	2.354 (19.33)	1.086 (5.14)		

^aRP-HPLC method using a mobile phase of 15 mM ammonium acetate buffer: acetonitrile (60:40 V/V) at a flow rate of 1.0 ml/min and detection at 220 nm

^bFigures between parentheses represent the corresponding tabulated values t and F values at P = 0.05

Dissolution monitoring

Monitoring of the dissolution of NAPR and DIPH was performed using RD-spectrophotometric method and compared to the reported HPLC method. Typically, dissolution curve is a plotted between the dissolved percentage of the drug and time (Kassem et al. 2014). Among the three proposed spectrophotometric methods, RD was preferably chosen over RS and DD¹ methods for further dissolution investigations because it exhibited simpler data manipulation steps and reliable results. To the best of our knowledge; RD method has been reported in many publications, yet this is the first article applying RD method to dissolution monitoring process. Samples were withdrawn from the dissolution medium at 5-min intervals, purified, then the concentrations of NAPR and DIPH by RD and HPLC methods were converted into %dissolution and plotted against time in minutes (Fig. 5). Dissolution profiles by the two methods revealed that NAPR took 35 min while DIPH took 40 min to dissolve completely, that complied successfully with the acceptance criterion stated by the USP which specified that the dissolution curves of immediate-release products typically reach 85–100% at about 30–45 min (2011). According to the Biopharmaceutics Classification System (BCS), NAPR and DIPH could be categorized as BCS class I drugs as they have high solubility and high permeability (Karam 2015). Thus, both drugs are eligible for the biowaiver procedure in which in vitro dissolution testing can be considered as an alternative to in vivo bioequivalence studies eliminating the need to harmful and costly procedure.

Although RD spectrophotometry and HPLC are in good agreements in accurate prediction of the dissolution profiles,



Fig. 5 Dissolution curves of NAPR and DIPH obtained by RD-spectrophotometric method and HPLC-PDA method

the two analytical methodologies differ in number of features particularly from a green analytical point of view. HPLC is the most applicable and versatile analytical technique in dissolution studies. Besides being a separation-based technique, photodiode array detection permits both qualitative and quantitative drug analysis. PDA's major advantage is related to the prediction of peak purity as the peak shape cannot assure that it corresponds to one or two (or even more) components. In such a case, absorbance at more than a single wavelength is very helpful in deciding whether the peak represents a single compound or not. However, in a case of Aleve pm[®] tablets, RD method excels as a greener alternative to HPLC. The proposed RD method depends on distilled water as solvent which is considered as green alternative for the organic solvents, since water is environmentally acceptable and recyclable. On the other side, HPLC utilized acetonitrile which is a costly solvent particularly after its prices have been skyrocketed after the so-called "Great acetonitrile crisis" provoked in late 2008. Moreover, acetonitrile is recently considered as a must-replaced solvent due to the debates which rose about its negative environmental impact. Also, the amount of generated waste is decreased dramatically in RD, compared to HPLC which consumes a lot of expensive pure solvents in column washing and conditioning steps. RD spectrophotometry can be considered a "greener" analytical protocol as it exhibits increasing safety of the operator through using environmentally friendly solvents, reduction of the analytical waste, in addition to lower energy consumption and reduced analysis time. To this end, RD spectrophotometry was found to be more closely adhering to the GAC principles, so it is strongly recommended for a greener and environmentally safer dissolution biowaiver studies.

Conclusion

Two different analytical methodologies, UV-spectrophotometry and HPLC, were developed for the simultaneous determination of NAPR and DIPH in their combined dosage form Aleve pm[®] tablets. The suggested spectrophotometric methods were RD, RS, and DD¹ that were used successfully for resolving the spectral overlap between NAPR and DIPH without prior separation based on ratio spectra manipulation. All the developed spectrophotometric and HPLC methods can be used for the determination of NAPR and DIPH in their pure bulk powder and in their combined dosage form with acceptable accuracy and sensitivity. Furthermore, RD method and HPLC were applied successfully to study the dissolution of both drugs from their tablet dosage form; then, the two techniques were compared from a green chemistry perspective. The proposed RD method provided a greener methodology as it utilized safer solvents, less energy

consumption, less generated waste, reduced analysis time, and enhanced operator safety.

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