

Synthesis, spectroscopic characterization, X-ray structure, DFT calculations, and antimicrobial studies of diorganotin (IV) complexes of monotopic oxygen nitrogen donor Schiff base

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Abstract Seven new diorganotin(IV) complexes, [Me₂SnL] (**1**), [Et₂SnL] (**2**), [(*n*-Bu)₂SnL] (**3**), [Ph₂SnL] (**4**), [(*n*-Oct)₂SnL] (**5**), [tert-Bu₂SnL] (**6**), and [*n*-BuClSnL] (**7**) have been synthesized from the reaction of *N'*-(2-hydroxybenzylidene)-4-tert-butylbenzohydrazide (H₂L) with the corresponding diorganotin(IV) dichloride/oxide or organotin(IV) chloride dihydroxide. The synthesized compounds were structurally characterized by FT-IR, multinuclear NMR (¹H and ¹³C) spectroscopies, elemental analysis, mass spectrometry, DFT/(B3LYP) calculations, and, for ligand single crystal, X-ray diffraction analysis. Spectroscopic evidence affirms coordination of ligand to the dialkyltin(IV) moieties through oxygen nitrogen donor sites in iminol form. The ¹J(¹¹⁹Sn, ¹³C) coupling constants, 584–655 Hz and ²J(¹¹⁹Sn–¹H) coupling constant, 79 Hz for

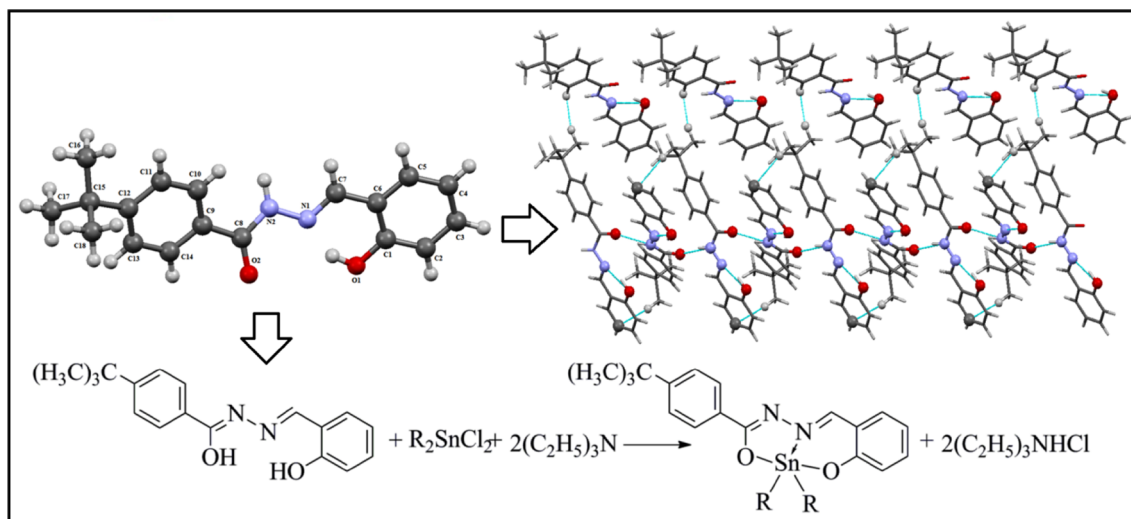
complex (**1**), suggest pentacoordination around Sn atom in solution. Single-crystal X-ray structure of ligand show its existence in amido form. Supramolecular architecture mediated by N(2)–H...O(2) and C–H...π interactions is formed in solid state. The DFT calculations have been performed to obtain various structure-based molecular properties as well as to support experimental results. The synthesized compounds were screened in vitro against various human pathogenic microbial strains. *Escherichia coli* and *Staphylococcus aureus* were most visibly inhibited by complexes (**4**) and (**7**). Highest antifungal activity was shown by compound (**6**) against *Fusarium solani*. Compound (**3**) displayed highest cytotoxicity among synthesized compounds with LD₅₀ 0.44 μg/mL.

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Graphical Abstract



Keywords Diorganotin(IV) complexes · DFT calculations · Antibacterial activity · Antifungal · Activity · Cytotoxicity

Introduction

Organotin(IV) compounds have been the center of attention of many research groups primarily due to diversity in their structures and wide range of applications. Substantial interest has been shown in their biological and pharmaceutical activities as anti-diabetic (Roy et al. 2016), antitumor (Fani et al. 2015), antituberculosis (Kovala-Demertzi et al. 2009), antileishmanial (Sirajuddin et al. 2014), and antimicrobial agents (Sharma et al. 2016). The biochemical behavior of these complexes is primarily influenced by the oxidation state, coordination number of metal atom, geometry, and thermodynamic and kinetic characteristics of complex. Furthermore, the groups directly bonded with tin atom not only confer special pharmaceutical characteristics to these compounds (Girasolo et al. 2014) by modifying their fat solubility but also play an essential role in the transportation of these compounds to specific sites (Gholivand et al. 2016).

Antimicrobial agents act on vital microbial functions like cell wall synthesis, nucleic acid synthesis, and folate metabolism, and may impede the role of cell membrane and ribosome. However, the microbes have developed ways of not being affected by these compounds. The emergence of such bacterial and fungal multidrug-resistant strains has increased the need to design and synthesize compounds which can be effectively used against such pathogens (Zhang et al. 2016).

Density functional theory (DFT) calculations of chemical species have become an integral part of research, where

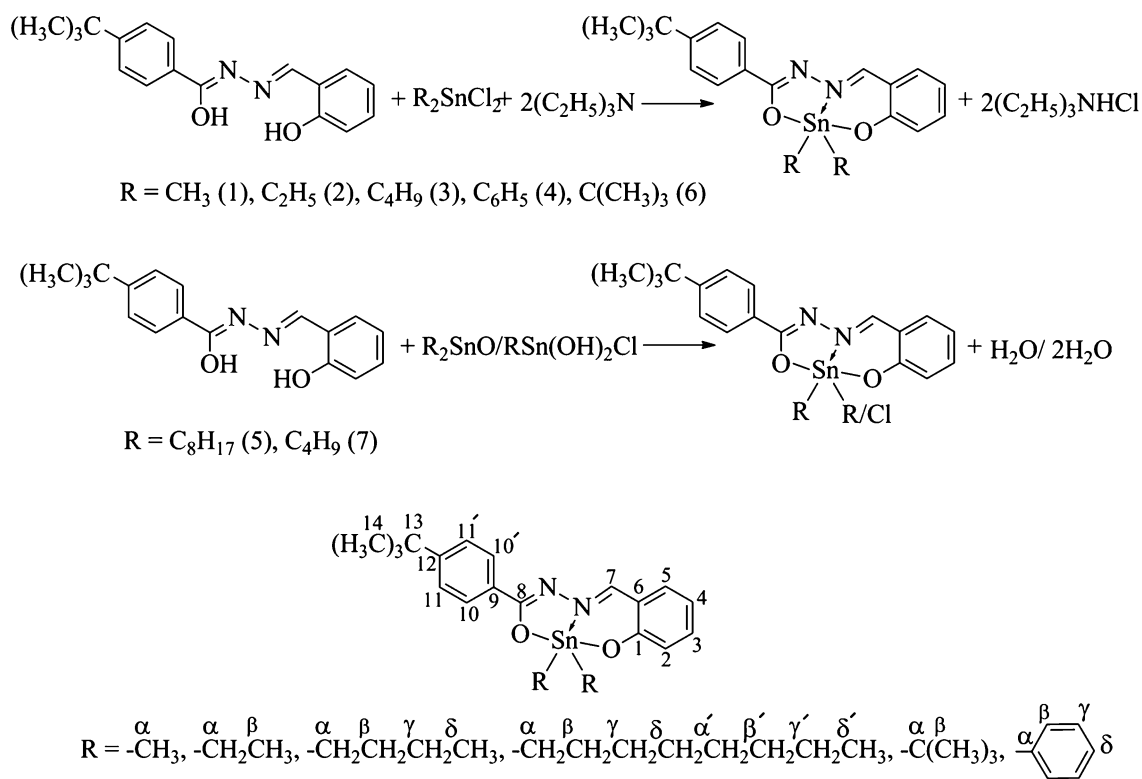
they play an important role in supplementing and supporting interpretation of experimental data. Such calculations well reproduce the experimental results and also provide various structure-based molecular properties. The B3LYP (Becke, 3-parameter, Lee–Yang–Parr) hybrid functional in DFT has been very popular, because it shows trade-off between accuracy and computational cost than the post Hartree–Fock methods and generally yields comparable results.

In continuation with our previous studies on diorganotin(IV) complexes and the rising demand for an efficient metal based compound that can inhibit the growth of pathogens, we report herein the synthesis of *N'*-(2-hydroxybenzylidene)-4-*tert*-butylbenzohydrazide (**H₂L**) and seven new diorganotin(IV) derivatives [R₂SnL] (R=–CH₃ (**1**), –C₂H₅ (**2**), *n*-C₄H₉ (**3**), –C₆H₅ (**4**), –C₈H₁₇ (**5**), –*tert*-C₄H₉ (**6**), *n*-C₄H₉Cl (**7**)) L=*N'*-(2-oxydobenzylidene)-4-*tert*-butylbenzohydrazide. All the compounds were characterized by FT-IR, multinuclear NMR (¹H and ¹³C), elemental analysis, mass spectroscopy, DFT/B3LYP calculations, and screened against selected pathogenic bacterial and fungal strains. Cytotoxicity assessed by in vivo lethality to brine shrimp nauplii is also reported.

Experimental

Materials and methods

Salicylaldehyde, 4-*tert*-butylbenzoic hydrazide, triethylamine, dimethyltin(IV) dichloride, diethyltin(IV) dichloride, dibutyltin(IV) dichloride, diphenyltin(IV) dichloride, dioctyltin(IV) oxide, and butyltin(IV) chloridedihydroxide were purchased from Sigma-Aldrich and used as received.



Scheme 2 Synthesis of diorganotin(IV) complexes (1–7) and numbering scheme of alkyl groups bonded to tin atom

3J ($^{119}\text{Sn}-^1\text{H}$) = 46 Hz), 8.02 (d, 2H, PhC10,10'-H, $^3J_{\text{H-H}} = 8.4$ Hz), 7.46 (d, 2H, PhC11,11'-H, $^3J_{\text{H-H}} = 8.4$ Hz), 1.37 (s, 9H, $\text{C}(\text{CH}_3)_3$), 0.85 (s, 6H, SnMe, 2J ($^{119/117}\text{Sn}-^1\text{H}$) = 79, 76 Hz). ^{13}C NMR (CDCl_3 ppm): 166.3, 135.1, 134.1, 121.7, 117.2, 116.8 (PhC1-C6), 161.1 (HC=N), 169.3 (OCNN), 154.6, 130.4, 127.4, 125.2 (PhC9-C12), 34.9, 31.2 ($\text{C}(\text{CH}_3)_3$), 1.4 (SnMe-C, 1J ($^{119/117}\text{Sn}-^{13}\text{C}$) = 655 Hz).

Diethyltin(IV) [N'-(2-oxidobenzylidene)-N-(oxido-(4-tert-butylphenyl)methylene) hydrazine] (2)

Yield: 1.2 g, 88%; m.p. paste; elemental analysis (%), calcd. for $\text{C}_{22}\text{H}_{28}\text{N}_2\text{O}_2\text{Sn}$: C, 56.08; H, 5.99; N, 5.95. Found: C, 56.11; H, 6.03; N, 5.93. FT-IR (KBr) $\nu_{\text{max}}/\text{cm}^{-1}$: 1606 (m, C=N), 1077 (w, N-N), 569 (w, Sn-O), 458 (w, Sn-N). EI-MS, m/z (%): $[\text{M}]^+$ 472(25.7); $[\text{M}-\text{Et}]^+$ 443(20.7); $[\text{M}-2\text{Et}]^+$ 414(44.0); $[\text{SnEt}]^+$ 149(2.7); $[\text{tert-BuPhCO}]^+$ 161(100.0); $[\text{Sn}]^+$ 120(4.6). ^1H NMR (CDCl_3 ppm): 6.81 (d, 1H, PhC2-H, $^3J_{\text{H-H}} = 8.4$ Hz), 7.34 (t, 1H, PhC3-H, $^3J_{\text{H-H}} = 7.8$ Hz), 6.73 (t, 1H, PhC4-H, $^3J_{\text{H-H}} = 7.4$ Hz), 7.18 (d, 1H, PhC5-H, $^3J_{\text{H-H}} = 7.8$ Hz), 8.79 (s, 1H, CH = N, 3J ($^{119}\text{Sn}-^1\text{H}$) = 43 Hz), 8.03 (d, 2H, PhC10,10'-H, $^3J = 8.4$ Hz), 7.46 (d, 2H, PhC11,11'-H, $^3J_{\text{H-H}} = 8.7$ Hz), 1.36 (s, 9H, $\text{C}(\text{CH}_3)_3$), 1.40–1.51 (m, 4H, SnEtC α -H), 1.32 (t, 6H, SnEtC β -H, $^3J_{\text{H-H}} = 7.2$ Hz) ^{13}C NMR (CDCl_3 ppm): 167.1, 135.0, 134.1, 121.6, 116.9, 116.8 (PhC1-C6), 161.2

(HC = N), 169.7 (OCNN), 154.5, 130.5, 127.4, 125.2 (PhC9-C12), 34.9, 31.2 ($\text{C}(\text{CH}_3)_3$), 14.3 (SnEt-C α , 1J ($^{119/117}\text{Sn}-^{13}\text{C}$) = 618 Hz), 9.3 (SnEt-C β , 2J ($^{119}\text{Sn}-^{13}\text{C}$) = 43 Hz).

Di-n-butyltin(IV) [N'-(2-oxidobenzylidene)-N-(oxido-(4-tert-butylphenyl)methylene) hydrazine] (3)

Yield: 1.2 g, 78%; m.p. 72–75 °C; elemental analysis (%), calcd. for $\text{C}_{26}\text{H}_{36}\text{N}_2\text{O}_2\text{Sn}$: C, 59.22; H, 6.88; N, 5.31. Found: C, 59.26; H, 6.84; N, 5.35. FT-IR (KBr) $\nu_{\text{max}}/\text{cm}^{-1}$: 1610 (m, C = N), 1082 (w, N-N), 570 (w, Sn-O), 465 (w, Sn-N). EI-MS, m/z (%): $[\text{M}]^+$ 528(29.7); $[\text{M}-n\text{-Bu}]^+$ 471 (20.1); $[\text{M}-2n\text{-Bu}]^+$ 414(31.6); $[\text{Sn}(\text{Bu})_2]^+$ 234(6.0); $[\text{tert-BuPhCO}]^+$ 161(100.0); $[\text{Sn}]^+$ 120(7.5). ^1H NMR (CDCl_3 ppm): 6.80 (d, 1H, PhC2-H, $^3J_{\text{H-H}} = 8.4$ Hz), 7.34 (t, 1H, PhC3-H, $^3J_{\text{H-H}} = 7.8$ Hz), 6.74 (t, 1H, PhC4-H, $^3J_{\text{H-H}} = 7.5$ Hz), 7.19 (d, 1H, PhC5-H, $^3J_{\text{H-H}} = 7.8$ Hz), 8.78 (s, 1H, CH = N, 3J ($^{119}\text{Sn}-^1\text{H}$) = 43 Hz), 8.05 (d, 2H, PhC10,10'-H, $^3J_{\text{H-H}} = 8.7$ Hz), 7.48 (d, 2H, PhC11,11'-H, $J = 8.4$), 1.38 (s, 9H, $\text{C}(\text{CH}_3)_3$), 1.64–1.72 (m, 4H, SnBuC α -H), 1.52–1.59 (m, 4H, SnBuC β -H), 1.35–1.42 (m, 4H, SnBuC γ -H), 0.90 (t, 6H, SnBuC δ -H, $^3J_{\text{H-H}} = 7.2$ Hz) ^{13}C NMR (CDCl_3 ppm): 166.9, 134.9, 134.1, 121.6, 117.1, 116.8 (PhC1-C6), 160.9 (HC = N), 169.6 (OCNN), 154.4, 130.6, 127.4, 125.2 (PhC9-C12), 34.9, 31.3 ($\text{C}(\text{CH}_3)_3$), 22.2 (SnBuC α ,

$^1J[^{119}\text{Sn}-^{13}\text{C}] = 604, 578 \text{ Hz}$, 26.9 (SnBuC β , $^2J[^{119}\text{Sn}-^{13}\text{C}] = 35 \text{ Hz}$), 26.5 (SnBuC γ , $^3J[^{119}\text{Sn}-^{13}\text{C}] = 88 \text{ Hz}$), 13.6 (SnBuC δ).

Diphenyltin(IV) [N'-(2-oxidobenzylidene)-N-(oxido-(4-tert-butylphenyl)methylene)hydrazine] (4)

Yield: 1.4 g, 82%; m.p. 151–153 °C; elemental analysis (%), calcd. for $\text{C}_{30}\text{H}_{28}\text{N}_2\text{O}_2\text{Sn}$: C, 63.52; H, 4.98; N, 4.94. Found: C, 63.49; H, 5.01; N, 4.91. FT-IR (KBr) $\nu_{\text{max}}/\text{cm}^{-1}$: 1608 (m, C=N), 1071 (w, N–N), 565 (w, Sn–O), 462 (w, Sn–N). EI-MS, m/z (%): $[\text{M}]^+$ 568(60.8); $[\text{M}-2\text{Ph}]^+$ 414(2.2); $[\text{SnPh}]^+$ 197(32.4); $[\text{tert-BuPhCO}]^+$ 161(100.0); $[\text{Sn}]^+$ 120(8.9). $^1\text{H NMR}$ (CDCl_3 ppm): 6.80 (d, 1H, PhC2-H, $^3J_{\text{H-H}} = 8.4 \text{ Hz}$), 7.36 (t, 1H, PhC3-H, $^3J_{\text{H-H}} = 7.8 \text{ Hz}$), 6.76 (t, 1H, PhC4-H, $J = 7.4 \text{ Hz}$), 7.15 (d, 1H, PhC5-H, $^3J_{\text{H-H}} = 8.1 \text{ Hz}$), 8.79 (s, 1H, CH = N, $^3J(^{119}\text{Sn}-^1\text{H}) = 52 \text{ Hz}$), 8.23 (d, 2H, PhC10,10'-H, $^3J_{\text{H-H}} = 8.7 \text{ Hz}$), 7.55 (d, 2H, PhC11,11'-H, $^3J_{\text{H-H}} = 8.7 \text{ Hz}$), 1.42 (s, 9H, $\text{C}(\text{CH}_3)_3$), 7.92–7.96 (m, 4H, SnPhC β -H), 7.40–7.50 (m, 6H, SnPhC γ , δ -H), $^{13}\text{C NMR}$ (CDCl_3 ppm): 167.2, 135.3, 134.3, 122.1, 117.5, 116.9 (PhC1–C6), 161.2 (HC=N), 169.2 (OCNN), 154.8, 130.5, 127.7, 125.3 (PhC9–C12), 35.0, 31.2 ($\text{C}(\text{CH}_3)_3$), 139.2 (SnPhC α), 136.3 (SnPhC β , $^2J[^{119}\text{Sn}-^{13}\text{C}] = 54 \text{ Hz}$), 128.9 (SnPhC γ , $^3J[^{119/117}\text{Sn}-^{13}\text{C}] = 83 \text{ Hz}$), 130.4 (SnPhC δ , $^4J[^{119}\text{Sn}-^{13}\text{C}] = 21 \text{ Hz}$).

Di-n-octyltin(IV) [N'-(2-oxidobenzylidene)-N-(oxido-(4-tert-butylphenyl)methylene)hydrazine] (5)

Yield: 1.4 g, 75%; m.p. viscous liquid; elemental analysis (%), calcd. for $\text{C}_{34}\text{H}_{52}\text{N}_2\text{O}_2\text{Sn}$: C, 63.86; H, 8.20; N, 4.38. Found: C, 63.89; H, 8.18; N, 4.41. FT-IR (KBr) $\nu_{\text{max}}/\text{cm}^{-1}$: 1606 (m, C = N), 1070 (w, N–N), 566 (w, Sn–O), 463 (w, Sn–N). EI-MS, m/z (%): $[\text{M}]^+$ 640(40.6); $[\text{M-Oct}]^+$ 527(28.3); $[\text{M}-2\text{Oct}]^+$ 414(34.9); $[\text{HSnOct}]^+$ 234(5.2); $[\text{tert-BuPhCO}]^+$ 161(87.8); $[\text{Sn}]^+$ 120(3.3); $[\text{Bu}]^+$ 57(100.0). $^1\text{H NMR}$ (CDCl_3 ppm): 6.81 (d, 1H, PhC2-H, $^3J_{\text{H-H}} = 8.4 \text{ Hz}$), 7.34 (t, 1H, PhC3-H, $^3J_{\text{H-H}} = 7.5 \text{ Hz}$), 6.74 (t, 1H, PhC4-H, $^3J_{\text{H-H}} = 7.5 \text{ Hz}$), 7.18 (d, 1H, PhC5-H, $^3J_{\text{H-H}} = 8.1 \text{ Hz}$), 8.77 (s, 1H, CH = N, $^3J(^{119}\text{Sn}-^1\text{H}) = 42 \text{ Hz}$), 8.04 (d, 2H, PhC10,10'-H, $^3J_{\text{H-H}} = 8.7 \text{ Hz}$), 7.47 (d, 2H, PhC11,11'-H, $^3J_{\text{H-H}} = 8.4 \text{ Hz}$), 1.38 (s, 9H, $\text{C}(\text{CH}_3)_3$), $\text{H}\alpha$: 1.65–1.77 (m, 4H, SnOctC α -H), 1.53–1.58 (m, 4H, SnOctC β -H), 1.23–1.29 (br, 16H, SnOctC γ - γ' -H), 0.88 (t, 6H, SnOct δ' , $^3J_{\text{H-H}} = 6.6 \text{ Hz}$), $^{13}\text{C NMR}$ (CDCl_3 ppm): 166.9, 134.9, 134.1, 121.6, 117.0, 116.8 (PhC1–C6), 160.8 (HC=N); 169.6 (OCNN); 154.4, 130.6, 127.8, 125.1 (PhC9–C12), 34.9, 31.2, ($\text{C}(\text{CH}_3)_3$), 22.7 (SnOctC α , $^1J[^{119}\text{Sn}-^{13}\text{C}] = 592 \text{ Hz}$) 24.7 (SnOctC β , $^2J[^{119}\text{Sn}-^{13}\text{C}] = 36 \text{ Hz}$), 33.4 (SnOctC γ , $^3J[^{119}\text{Sn}-^{13}\text{C}] = 76 \text{ Hz}$), 29.2 (SnOctC δ), 29.1 (SnOctC α'), 31.2 (SnOctC β'), 22.5 (SnOctC γ'), 14.1 (SnOctC δ').

Di-tert-butyltin(IV) [N'-(2-oxidobenzylidene)-N-(oxido-(4-tert-butylphenyl)methylene)hydrazine] (6)

Yield: 1.3 g, 80%; m.p. 137–138 °C; elemental analysis (%), calcd. for $\text{C}_{26}\text{H}_{36}\text{N}_2\text{O}_2\text{Sn}$: C, 59.22; H, 6.88; N, 5.31. Found: C, 59.19; H, 6.91; N, 5.28. FT-IR (KBr) $\nu_{\text{max}}/\text{cm}^{-1}$: 1606 (m, C=N), 1075 (w, N–N), 566 (w, Sn–O), 458 (w, Sn–N). EI-MS, m/z (%): $[\text{M}]^+$ 528(5.4); $[\text{M}-2\text{tert-Bu}]^+$ 414(19.9); $[\text{Sn}(\text{tert-Bu})_2]^+$ 234(3.3); $[\text{tert-BuPhCO}]^+$ 161(21.5); $[\text{Sn}]^+$ 120(2.2); $[\text{Bu}]^+$ 57(100.0). $^1\text{H NMR}$ (CDCl_3 ppm): 6.86 (d, 1H, PhC2-H, $^3J_{\text{H-H}} = 8.1 \text{ Hz}$), 7.34 (t, 1H, PhC3-H, $^3J_{\text{H-H}} = 7.8 \text{ Hz}$), 6.72 (t, 1H, PhC4-H, $^3J_{\text{H-H}} = 7.5 \text{ Hz}$), 7.18 (d, 1H, PhC5-H, $^3J_{\text{H-H}} = 7.8 \text{ Hz}$), 8.80 (s, 1H, CH = N, $^3J(^{119}\text{Sn}-^1\text{H}) = 39 \text{ Hz}$), 8.09 (d, 2H, PhC10,10'-H, $^3J_{\text{H-H}} = 8.7 \text{ Hz}$), 7.49 (d, 2H, PhC11,11'-H, $^3J_{\text{H-H}} = 8.4 \text{ Hz}$), 1.38 (s, 9H, $\text{C}(\text{CH}_3)_3$), 1.36 (s, 18H, Sn-*tert*-BuC β -H, $^3J(^{119/117}\text{Sn}-^1\text{H}) = 109, 104 \text{ Hz}$). $^{13}\text{C NMR}$ (CDCl_3 ppm): 168.0, 134.7, 133.9, 121.7, 117.0, 116.5 (PhC1–C6), 160.7 (HC=N); 169.4 (OCNN); 154.3, 130.8, 127.4, 125.2 (PhC9–C12), 34.9, 31.3, ($\text{C}(\text{CH}_3)_3$), 40.4 (Sn-*tert*-BuC α , $^1J[^{119/117}\text{Sn}-^{13}\text{C}] = 584 \text{ Hz}$), 29.7 (Sn-*tert*-BuC β).

n-Butylchloridetin(IV) [N'-(2-oxidobenzylidene)-N-(oxido-(4-tert-butylphenyl)methylene)hydrazine] (7)

Yield: 1.2 g, 77%; m.p. 143–144 °C; elemental analysis (%), calcd. for $\text{C}_{22}\text{H}_{27}\text{ClN}_2\text{O}_2\text{Sn}$: C, 52.26; H, 5.38; N, 5.54. Found: C, 52.23; H, 5.34; N, 5.53. FT-IR (KBr) $\nu_{\text{max}}/\text{cm}^{-1}$: 1609 (m, C=N), 1072 (w, N–N), 570 (w, Sn–O), 461 (w, Sn–N). EI-MS, m/z (%): $[\text{M}]^+$ 506(13.9); $[\text{M}-\text{BuCl}]^+$ 414(7.6); $[\text{C}_7\text{H}_4\text{NOSn}]^+$ 238(2.5); $[\text{tert-BuPhCO}]^+$ 161(100.0); $[\text{Sn}]^+$ 120(5.1); $[\text{Bu}]^+$ 57(31.8). $^1\text{H NMR}$ (CDCl_3 ppm): 6.84 (d, 1H, PhC2-H, $^3J_{\text{H-H}} = 8.1 \text{ Hz}$), 7.36 (t, 1H, PhC3-H, $^3J_{\text{H-H}} = 8.1 \text{ Hz}$), 6.82 (t, 1H, PhC4-H, $^3J_{\text{H-H}} = 7.2 \text{ Hz}$), 7.20 (d, 1H, PhC5-H, $^3J_{\text{H-H}} = 7.8 \text{ Hz}$), 8.86 (s, 1H, CH = N), 8.01 (d, 2H, PhC10,10'-H, $^3J_{\text{H-H}} = 8.4 \text{ Hz}$), 7.52 (d, 2H, PhC11,11'-H, $^3J_{\text{H-H}} = 8.4 \text{ Hz}$), 1.32 (s, 9H, $\text{C}(\text{CH}_3)_3$), 1.76–1.84 (m, 4H, SnBuC α -H), 1.61–1.67 (m, 4H, SnBuC β -H), 1.45–1.57 (m, 4H, SnBuC γ -H), 0.97 (t, 6H, SnBuC δ -H, $J = 7.2 \text{ Hz}$) $^{13}\text{C NMR}$ (CDCl_3 ppm): 165.9, 135.0, 134.6, 122.1, 118.0, 117.1 (PhC1–C6), 158.1 (HC=N); 166.6 (OCNN); 154.7, 130.6, 127.5, 125.7 (PhC9–C12), 35.2, 31.4 ($\text{C}(\text{CH}_3)_3$), 25.6 (SnBuC α), 28.8 (SnBuC β), 27.4 (SnBuC γ), 14.1 (SnBuC δ).

Antibacterial activity

The antibacterial action of synthesized compounds was assessed against *Escherichia coli* ATCC 11229, *Bacillus subtilis* ATCC 11774, *Shigella flexneri* ATCC 10782, *Staphylococcus aureus* ATCC 25923, *Pseudomonas aeruginosa* ATCC 10245, and *Salmonella typhi* ATCC 10749, by agar well diffusion method. Imipenem was used as standard drug (Rehman et al. 2008). Six mm diameter wells were dug in the media by means of sterile metallic borer. Eight-hour-old

bacterial inoculums containing approximately 10^4 – 10^6 CFU/mL were spread over the surface of nutrient agar with the help of sterile cotton swab. The test samples with a concentration of 2 mg/mL in DMSO were introduced into the wells. Reference antibacterial drug and DMSO served as positive and negative controls, respectively. After incubation at 37 °C for 20 h, the zones of inhibition (mm) were measured and compared with the standard drug.

Antifungal activity

The agar tube dilution test was used to investigate the in vitro antifungal activity of synthesized compounds against six fungal strains [*Trichophyton longifusus* ATCC 22397, *Candida albicans* ATCC 2192, *Aspergillus flavus* ATCC 1030, *Microsporum canis* ATCC 9865, *Fusarium solani* ATCC 11712, and *Candida glabrata* ATCC 90030 (Atta-ur-Rahman and Thomsen 2001)]. Reference drugs Miconazole and Amphotericin B were used for comparison. DMSO solutions of synthesized compounds (200 mg/mL) were prepared. The Sabouraud dextrose agar media (4 mL) prepared by mixing Sabouraud (32.5 g), glucose agar (4%), and agar–agar (20 g) in 500 mL of distilled water was dispensed into screw-capped tubes and autoclaved for 15 min at 121 °C. The prepared test compound solutions were added to non-solidified Sabouraud agar media (50 °C). This was then solidified at room temperature and inoculated with 4 mm diameter portion of inoculums, derived from a 7-day-old respective fungal culture. An agar surface streak was employed to ensure non-mycelial growth. The tubes were incubated at 27–29 °C for 7–10 days and the growth inhibition was measured with reference to the control.

Cytotoxicity

The cytotoxicity of the synthesized compounds was investigated by the Brine shrimp lethality test (Atta-ur-Rahman and Thomsen 2001)]. Brine shrimps (*Artemia salina*) eggs were hatched in a vessel containing sterile simulated seawater, prepared using 38 g.L⁻¹ sea salt; pH was adjusted to 8.5 using 1 M NaOH at room under constant aeration, for 2 days. Thirty active nauplii were placed in a vial containing brine solution (4.5 mL) and a drop of yeast suspension. In each experiment, test solution (0.5 mL) was added to the vial and surviving larvae were counted at ambient temperature after 24 h. Experiments were conducted in triplicate for each concentration (1, 10, and 100 mg/mL) of test substances and compared with the control. Data were analyzed with Finney's probit analysis to determine the LD₅₀ (Finney 1971)]. Etoposide was used as the standard drug.

Computational details

The present quantum chemical calculations have been performed using the DFT method employing B3LYP hybrid functional as implemented in the Gaussian 09 software (Becke 1988; Frisch et al. 2009; Lee et al. 1988). A graphical user interface, GaussView 5, was used to prepare input files as well as to visualize/plot the theoretical results (Dennington et al. 2009). Geometry optimization and harmonic frequency calculations were carried out for isolated gas phase H₂L compound and seven new diorganotin (IV) complexes in the ground electronic state at B3LYP/6-311 ++G(d,p) level and B3LYP/6-311 ++G(d,p)/LANL2DZ level of theory, respectively. The basis set LANL2DZ with effective core potential was chosen for Sn atom while for other atoms (C, H, N, O, and Cl) 6-311 ++G(d,p) basis set was used. The lack of imaginary values in the calculated wavenumbers revealed the optimized geometry corresponding to an energy minimum on potential energy surface. The harmonic frequencies generally overestimate the experimental one mainly by ignoring anharmonic terms in potential energy expression. This overestimation appears systematically. Therefore, various types of scaling factors have been employed to match theoretical IR band position with experimental one. In this work, a uniform scaling factor, 0.960 in the high wavenumbers region and by 0.988 in the low wavenumbers region (below 1800 cm⁻¹), was used to scale down the harmonic wavenumbers (Borba et al. 2010). The IR spectra of the compounds were simulated using Lorentzian band shape with FWHM of 5 cm⁻¹. The NMR spectra (¹H and ¹³C) of the present compounds were simulated in the solvent phase (chloroform) using gauge independent atomic orbital (GIAO) method within DFT/B3LYP framework. The integral equation formalism (IEF) version of the polarizable continuum model (IEF-PCM) was employed for the NMR spectra calculations in the solvent phase. Each solvent-phase optimized structure of the present ligand as well as complexes was subjected to single point GIAO-NMR calculations in a polarized continuum of chloroform. Relative chemical shift values (in ppm) were obtained by subtracting the corresponding tetramethylsilane (TMS) shielding for ligand at GIAO-B3LYP/6-311 + G(2d,p) and complexes at GIAO-B3LYP/6-311 + G(2d,p)/LANL2DZ level of theory. Furthermore, various other molecular properties such as atomic charges, dipole moment, chemical reactivity descriptors based on highest occupied molecular orbital (HOMO) and lowest unoccupied molecular orbital (LUMO), molecular electrostatic potential (MEP), thermodynamic properties, and HOMO–LUMO analysis of the ligand, as well as complexes were also reported.

Results and discussion

The condensation of salicylaldehyde with 4-*tert*-butylbenzoic hydrazide in ethanol afforded the ligand *N'*-(2-hydroxybenzylidene)-4-*tert*-butylbenzohydrazide (**H₂L**), which, on reaction with diorganotin(IV) compounds in stoichiometric ratio, yielded R₂SnL (**1–6**) (R=–CH₃, –C₂H₅, –*n*-C₄H₉, –C₆H₅, –C₈H₁₇, –*tert*-C₄H₉) and BuClSnL (**7**) in good yield (75–88%, Scheme 1, 2). All the compounds are stable in air at room temperature, soluble in common organic solvents and solids under normal conditions except compound (**5**). The elemental analysis data provided in the experimental section support the formation of complexes and suggest a 1:1 diorganotin(IV) and ligand ratio.

Infrared spectroscopy

Comparison of IR spectra of complexes with the free ligand helps in the identification of characteristic vibrational frequencies and gives an insight into the binding modes of ligand and structure of complexes formed. In FT-IR spectrum of ligand, the bands observed at 3455 and 3178 cm⁻¹ are assigned to ν OH and ν NH stretching vibrations respectively, whereas the corresponding theoretical wavenumbers are 3292 and 3388 cm⁻¹ respectively. Theoretically, O–H stretching wavenumber is predicted at lower region than N–H stretching wavenumber due to consideration of intramolecular interaction (O–H–N) only in calculation. The **H₂L** compound is appeared with its dimer structure in crystal having interactions, O–H–N (intramolecular) and C=O–H–N (intermolecular). Therefore, the frequencies of O–H, N–H, and C=O stretching vibrations are shifted considerably towards lower region. A sharp band at 1669 cm⁻¹ indicates the presence of C=O group and existence of ligand in the amido form. The corresponding theoretical IR frequency is found to be 1729 cm⁻¹. The absence of all these bands in the IR spectra of complexes suggests that during complex formation, the ligand undergoes tautomeric shift from amido to enolic form followed by deprotonation and coordination to diorganotin(IV) moieties. The absorption band due to ν C=N in ligand observed at 1616 cm⁻¹ shifts to lower values in the spectra of all diorganotin (IV) complexes indicating shifting of electron density from the nitrogen of C=N group to tin atom or coordination of azomethine nitrogen with the tin center (Pettinari et al. 2001). Furthermore, this causes a decrease in inter lone-pair repulsion on the nitrogen atoms and shifts the ν (N–N) frequencies to higher values at 1070–1080 cm⁻¹ in the spectra of organotin(IV) complexes. These IR frequencies are also found in accordance with theoretical values. The band due to C=N stretching vibration in ligand is predicted at 1648 cm⁻¹, while in all tin complexes, this vibration is appeared comparatively at lower value around 1613 cm⁻¹. The N–N stretching

vibration is predicted within the region 1030–1050 cm⁻¹ for the tin complexes, whereas this vibration is found 1158 cm⁻¹ for **H₂L** compound. In all the synthesized complexes, new IR bands also emerge in the regions of 559–570 cm⁻¹ and 451–458 cm⁻¹ due to the formation of Sn–O and Sn–N bonds, respectively (Hong et al. 2011; Muhammad et al. 2012). The corresponding theoretical IR bands due to Sn–O and Sn–N stretching vibrations are occurred in the regions 615–605 cm⁻¹ and 575–560 cm⁻¹, respectively. The appearance of these absorption bands supports the formation of complexes. Even after scaling the IR frequencies, the values are found larger than those of experimental one due to the fact that calculations have been carried out for the isolated molecule in the gaseous phase, while the experiments belong to solid phase.

NMR spectroscopy

The comparison of ¹H NMR spectrum of ligand and diorganotin (IV) complexes indicates some explicit structural changes in the ligand and also provides useful information regarding the binding mode of ligand. The disappearance of resonance signals due to OH and NH groups in the ¹H NMR spectra of all diorganotin(IV) complexes **1–7** indicates that the ligand undergoes enolization and deprotonation before forming the organotin(IV) complexes (Shujah et al. 2013). The proton and carbon (¹³C) NMR spectra of ligand as well as all complexes obtained by GIAO-DFT/B3LYP method have been compared with the experimental one and are found in good agreement.

The aromatic protons of the ligand and methyl protons of *tert*-butyl group give signal in the region δ 6.92–7.30 and 1.32 ppm, respectively, whereas theoretically, the proton NMR signals occurred around 7–8 ppm and 1–2 ppm, respectively. Since these groups are not involved in bonding, so all these signals remain unaffected in the NMR spectra of diorganotin(IV) complexes.

The azomethine proton (CH = N) gives a signal at 8.64 ppm, while theoretically, it appeared at 8.41 ppm (in **H₂L**). In all the diorganotin(IV) complexes, a downfield shift to 8.76–8.80 ppm and appearance of tin satellite with ³*J*(¹¹⁹Sn–¹H) spin–spin coupling constant of 39–52 Hz indicate the shifting of electron density from azomethine nitrogen to Sn atom and presence of Sn–N coordinated bond in solution. The chemical shifts and coupling constants are consistent with reported diorganotin(IV) complexes of ONO donor type ligands (Shujah et al. 2011). The diorganotin(IV) complexes show additional signals owing to the protons of alkyltin(IV) moieties. The CH₃ protons in complex (**1**) resonate as a singlet at 0.85 ppm (around 0.5 ppm in theoretical ¹H-NMR spectrum) with Sn satellites corresponding to ²*J*(¹¹⁹Sn–¹H) coupling constants of 79 Hz. The ²*J*(¹¹⁹Sn, ¹H) coupling constant is a valuable parameter used to estimate

Table 1 (C–Sn–C) angles ($^\circ$) based on NMR parameter of selected organotin(IV) complexes

Comp. no.	$^1J(^{119}\text{Sn}, ^{13}\text{C})$ (Hz)	$^2J(^{119}\text{Sn}, ^1\text{H})$ (Hz)	Angle ($^\circ$)	
			$\theta(^1J)$	$\theta(^2J)$
1	655	79	134.2	129.6
2	618	–	131.0	–
3	604	–	135.3	–
5	592	–	134.1	–
6	584	–	133.4	–

Me–Sn–Me bond angle in solution state. In the present study, only the dimethyltin(IV) derivative exhibits $^2J(^{119}\text{Sn}, ^1\text{H})$ coupling in solution. On substitution the value of 2J in Lockhart's equation $\theta = 0.0105 [^2J]^2 - 0.799[^2J] + 122.4$, we get θ , the Me–Sn–Me angle, equal to 129.6° indicating a pentacoordinated tin atom in solution (Lockhart and Manders 1986). The terminal methyl of diethyl, dibutyl, dioctyl, and

butylchlorotin(IV) complexes (**2**), (**3**), (**5**), and (**7**) appear as triplet at 1.32, 0.90, 0.88, and 0.97 ppm with a $^3J(^1\text{H}-^1\text{H})$ coupling constant 7.2, 7.2, 6.6, and 7.2 Hz, respectively. Rest of the protons resonate in the expected range. In complex (**6**), the *tert*-butyl protons resonate at 1.36 ppm as singlet with a $^3J(^{117/119}\text{Sn}-^1\text{H})$ coupling constant of 104 and 109 Hz. The protons of diphenyltin moiety in complex (**4**) appear as multiplets in the regions 7.92–7.96 and 7.40–7.50 ppm. The ^{13}C NMR spectra data of complexes show a downfield shift of all carbon resonances, compared with that of the ligand, because coordination leads to the shifting of electron density from the ligand to diorganotin(IV) moieties. For organotin(IV) compounds, the $^1J[^{119}\text{Sn}, ^{13}\text{C}]$ coupling constants are important parameters used to evaluate the molecular structure and establish the coordination around tin in solution. Among the synthesized complexes $^1J[^{119}\text{Sn}, ^{13}\text{C}]$ coupling satellites were observed only for compounds 1, 2, 3, 5, and 6. The C–Sn–C bond angles in solution based on $^1J[^{119}\text{Sn}, ^{13}\text{C}]$ coupling constants have been calculated

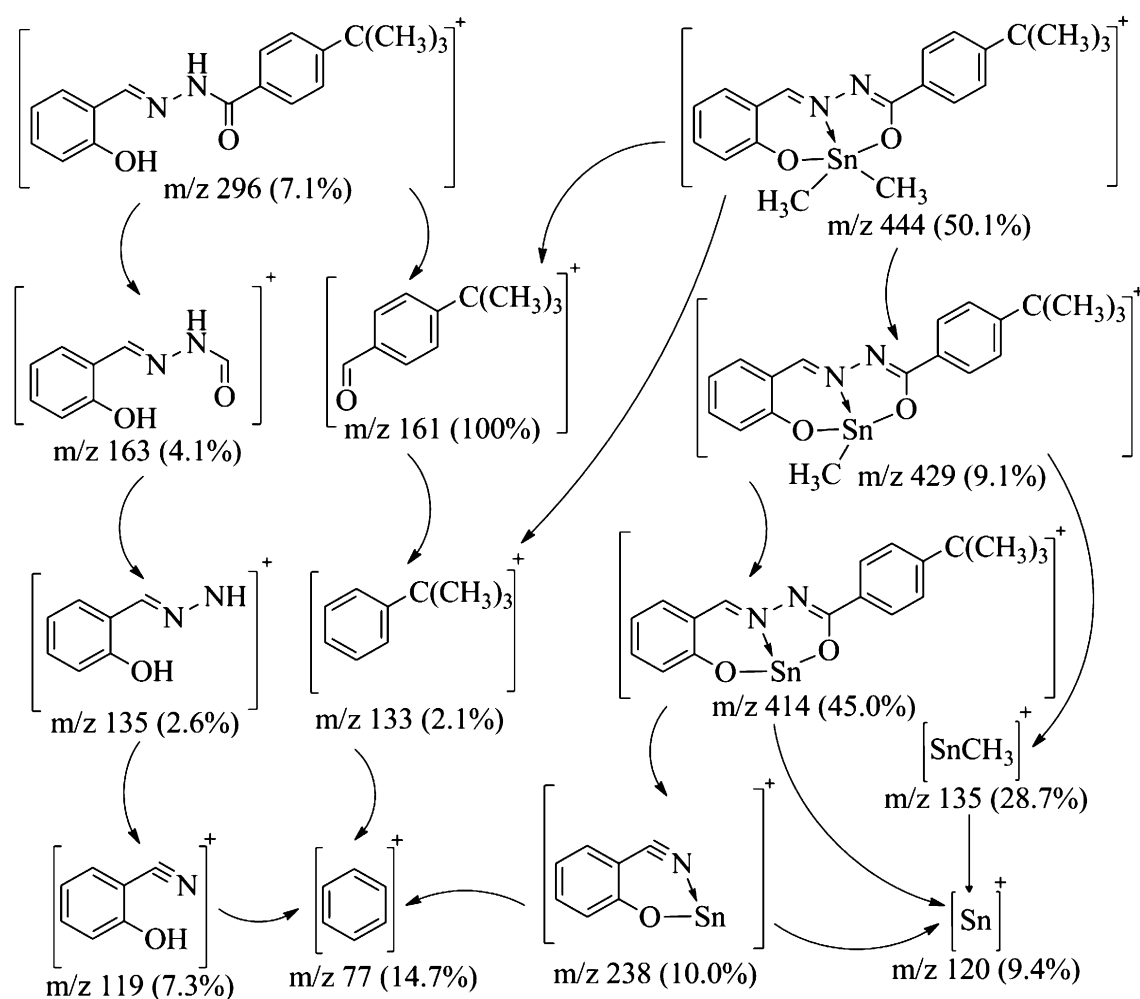
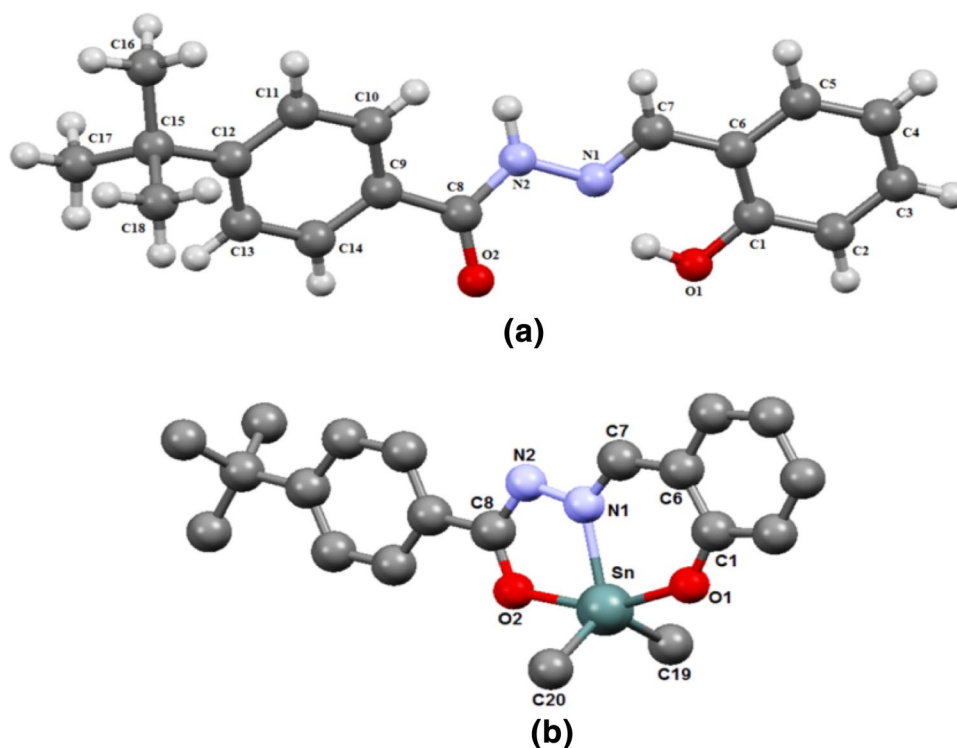
**Scheme 3** Proposed mass fragmentation pattern of ligand (H_2L) and dimethyltin(IV) complex (**1**)

Fig. 1 Molecular structure of ligand (a) and optimized structure of dimethyltin(IV) complex (b) with the atomic numbering scheme



and results reported in Table 1. The coupling constants for compounds (1), (2), (3), (5), and (6) were found to be in the range 584–655 Hz, which suggest pentacoordination around Sn atom in solution (Holeček and Lyčka 1986).

Mass spectrometry

The EI-MS spectral data of ligand (H_2L) and diorganotin(IV) complexes (R_2SnL , 1–7) are provided in the experimental section. The molecular ion peak and fragments containing tin were recognized on the basis of unique peak pattern attributed to ten stable isotopes of tin. The molecular ion ($C_{18}N_{18}N_2O_2SnR_2$)⁺ is observed in all cases; however, the peak intensity is low in case of complex (6) and (7). The first fragmentation step involves the formation of ($C_{18}N_{18}N_2O_2SnR_2$)⁺ fragment due to the loss of R group (R = CH₃, C₂H₅, C₄H₉, C₈H₁₇ except for complex (4) and (7) where the loss of both R leads to the formation of ($C_{18}N_{18}N_2O_2Sn$)⁺. The fragments ((CH₃)₃CC₆H₄CO)⁺ and (C₄H₉)⁺ are base peaks for H_2L , (1), (2), (3), (4), (7), and 5, 6, respectively. In addition, RSn^+ and Sn^+ ions of significant peak intensity are also observed for all the complexes (1–7). The mass spectral data for the fragmented ions are in close agreement with the proposed structure of the diorganotin(IV) complexes. The proposed fragmentation pattern of ligand and dimethyltin(IV) complex is provided in Scheme 3.

X-ray crystal structure of *N'*-(2-hydroxybenzylidene)-4-tert-butylbenzohydrazide (H_2L)

The molecular structure along with the atomic numbering scheme for ligand (H_2L) is shown in Fig. 1. Crystal parameters, selected bond lengths, and bond angles are given in Tables 2 and 3 respectively. The asymmetric unit of title compound consists of one molecule. In the solid state, the compound exists in the amido form, carbonyl and phenolic –OH groups are retained in the *cis* position. The N(1)–C(7) and O(2)–C(8) bond lengths [1.288(2), 1.243(2) Å] indicate their double bond nature. However, the N(1)–N(2), N(2)–C(8), and O(1)–C(1) bond lengths [1.381(2), 1.354(3), and 1.362(3) Å] support their single bond nature. Rest of the bond lengths are in the normal range. In the solid state, supramolecular architecture is formed mediated by N(2)–H...O(2) and C–H...π interactions (Fig. 2). The data relating to hydrogen bonds are depicted in Table 4.

Optimized molecular geometry

The comparison of optimized structural parameters (bond lengths and bond angles) with the XRD data of H_2L compound has been presented in Table 3. The synthesis scheme of the H_2L compound has two possibility of product namely T1 and T2 (Scheme 1). The optimized geometries of isolated H_2L compound (T1 and T2) with numbering scheme are illustrated in Fig. 3. The self-consistent field (SCF) minimum energy on the potential energy surface (PES) of T1 and

Table 2 Crystal data and structure refinement parameters for ligand (H_2L)

Empirical formula	$\text{C}_{18}\text{H}_{20}\text{N}_2\text{O}_2$
Formula mass	296.37
Crystal system	Orthorhombic
Space group	<i>Pbca</i>
<i>a</i> (Å)	13.982(4)
<i>b</i> (Å)	9.269(3)
<i>c</i> (Å)	24.557(7)
α (°)	90.00
β (°)	90.00
γ (°)	90.00
<i>V</i> (Å ³)	3182.6 (16)
<i>Z</i>	8
Crystal habit	Needle
Size (mm)	0.61 × 0.20 × 0.14
<i>T</i> (K)	100 (1)
ρ (g·cm ⁻³)	1.237
μ (Mo <i>K</i> α) (cm ⁻¹)	0.81
<i>F</i> (000)	1264
Total reflections	14719
Independent reflections	3234
For ($F_o \geq 4.0 \sigma(F_o)$)	1946
$R(F) = \Sigma(F_o - F_c) / \Sigma F_o $	0.0486
For $F_o > 4.0 \sigma(F_o)$	
$wR(F^2) = [\Sigma[w(F_o^2 - F_c^2)^2] / \Sigma[w(F_o^2)^2]]^{1/2}$	0.1264
Goodness-of-fit	1.007
θ Range (deg)	3.12–26.72
Data/restrictions/params	3234/0/279
Largest diff. peak and hole (eÅ ⁻³)	–0.19 and 0.23(5)

Table 3 Comparison of optimized structural parameters [bond lengths (Å) and bond angles (°)] with the XRD data of H_2L compound

Bond lengths	Experimental	B3LYP/6-311 ++G(d,p)
O(1)–C(1)	1.362(3)	1.344
O(2)–C(8)	1.243(2)	1.214
N(1)–N(2)	1.381(2)	1.357
N(1)–C(7)	1.288(3)	1.286
N(2)–C(8)	1.354(3)	1.389
C(8)–C(9)	1.486(3)	1.498
Bond angles		
N(2)–N(1)–C(7)	116.96(16)	118.9
N(1)–N(2)–C(8)	118.04(16)	120.3
O(1)–C(1)–C(2)	117.46(18)	117.7
O(1)–C(1)–C(6)	122.16(18)	122.9
N(1)–C(7)–C(6)	120.12(17)	121.6
O(2)–C(8)–N(2)	122.33(18)	122.4
O(2)–C(8)–C(9)	120.95(17)	123.1
N(2)–C(8)–C(9)	116.70(16)	114.5

T2 is found to be –601262.145 and –601258.786 kcal/mol respectively at B3LYP/6-311 ++G(d,p) level of theory. The minimum molecular energy has been obtained for T1 geometry (with N–H); therefore, it is found more stable than T2 geometry. This has been also confirmed from geometry (T1) of H_2L molecule obtained by XRD. The further theoretical investigations have been carried out on optimized geometry of T1. The optimized geometries of other seven new complexes have been also presented in Fig. 4.

Calculated geometric parameters of the present diorganotin(IV) complexes are summarized in Table 5. The 6-311 ++G(d,p) basis set led calculations of bond lengths and bond angles are comparable to reported experimental data in one of our research articles (Shujah et al. 2011).

In diorganotin(IV) complexes, the tin atom is present in a strongly distorted square-pyramidal geometry. In complexes (1–7), the distortions in bond lengths and angles of optimized geometry obtained at DFT/B3LYP from a possible geometry (Shujah et al. 2011) are found respectively around 0.1 Å and 1–4°.

Other theoretical molecular parameters

Various important molecular parameters such as molecular energy, dipole moment, thermodynamic parameters, HOMO–LUMO energy eigen values, HOMO–LUMO energy gaps, and global reactivity descriptors of the ligand, as well as complexes have been depicted in Table 6. The dipole moment values for complexes 1–5 and 7 are found to be around zero as these molecules have center of symmetry. The HOMO and LUMO are the main molecular orbitals which are involved in charge transfer within the molecule. The HOMO shows the ability to donate an electron, while LUMO exhibits the capability to accept an electron. The energy eigen values of HOMO and LUMO are directly related to ionization potential and electron affinity, respectively. The energy gap between HOMO and LUMO determines the kinetic stability and chemical reactivity of molecule. The plots of frontier molecular orbitals (HOMO and LUMO) for H_2L and complexes (1–7) are shown in Fig. 5. These plots show the charge transfer through conjugated pi bonds as well as through Sn connected bonds. The low HOMO–LUMO gap estimated around 4 eV for H_2L compound and around 3.5 eV for the complexes (1–7) shows the significant degree of intramolecular charge transfer, high chemical reactivity, and low kinetic stability. The global reactivity descriptors related to HOMO–LUMO energy eigen values of the present molecules obtained at DFT level, such as hardness, softness, chemical potential, and electrophilicities, are reported to understand their chemical reactivities. The large HOMO–LUMO gap indicates a hard molecule with low chemical reactivity, while low HOMO–LUMO energy gap shows a soft molecule of

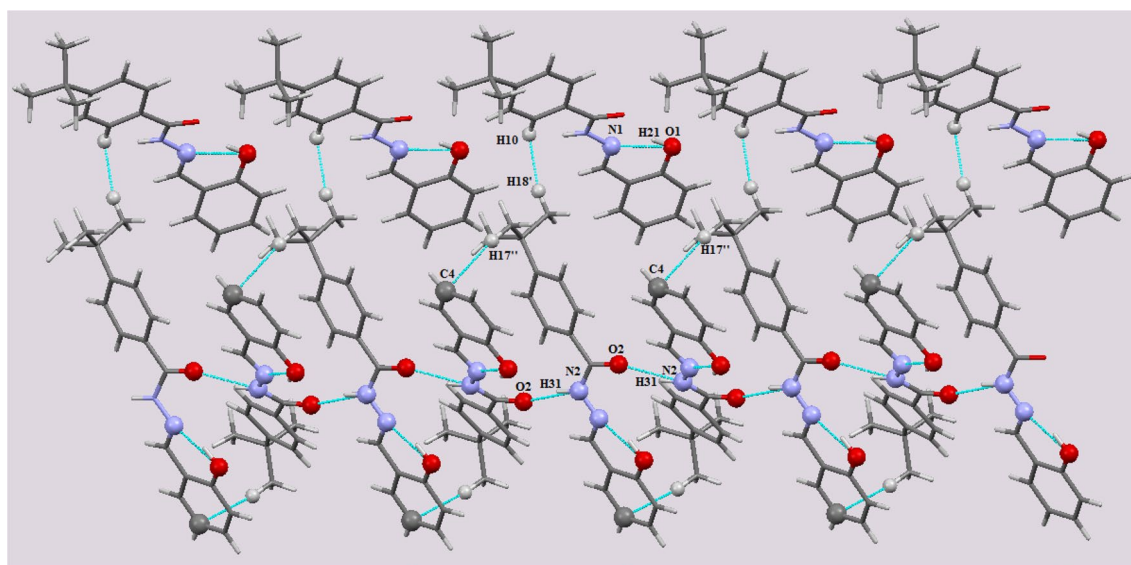


Fig. 2 Supramolecular architecture mediated by N(2)-H...O(2) and C-H... π interactions in the crystal lattice of H_2L

Table 4 Hydrogen-bond geometry (\AA , $^\circ$) for H_2L

D-H...A	D-H	H...A	D...A	D-H...A
O(1)-H(21)...N(1)	0.93	1.79(3)	2.633(2)	149.0(3)
N(2)-H(31)...O(2)	0.938(19)	1.895(19)	2.800(2)	161.4(19)
C(7)-H(7)...O(2)	0.989(19)	2.41(2)	3.198(3)	136.3(16)
C(10)-H(10)...O(2)	0.93(2)	2.57(2)	3.412(3)	150.2(16)

high chemical reactivity. The negative chemical potentials also define the stability of molecule. The electrophilicity index measures the degree of transfer of electrons from a donor to an acceptor system. The equations used to obtain these descriptors and their significance can be found in the literature (Ayers et al. 2005; Chermette 1999; Geerlings and De Proft 2008; Geerlings et al. 2003; Kohn et al. 1996; Parr and Yang 1989, 1995). The derived reactivity indices based on HOMO and LUMO energy eigen values given in this work are for qualitative comparison purposes.

The MEP surface plot of molecule is very useful in qualitative interpretation of the electrophilic and nucleophilic reactions as well as intermolecular hydrogen-bonding

interactions. MEP surface plots, generated for the present molecules by mapping of electrostatic potential on to the constant electron density surface, are shown in Fig. S10.

Antibacterial activity

All the compounds (H_2L , **1-7**) were tested against pathogenic bacterial strains using the agar well diffusion method (Table 7). The in vitro inhibitory activity was performed against *Escherichia coli*, *Bacillus subtilis*, *Sigella flexneri*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, and *Salmonella typhi*, using imipenem as standard drug. The ligand showed moderate activity only against *Shigella flexneri*, Gram-negative bacteria, mainly responsible for diarrhoeal diseases in humans. The results indicate that on complexation, the antibacterial activity increases which is also supported by earlier reports (Anwer et al. 2013).

Most of the antibacterial agents are more active against Gram-positive bacteria, because they do not possess outer membrane and only have a loosely packed porous polyglycane exterior layer through which the penetration of drug into the cell is easy. However, in the present study,

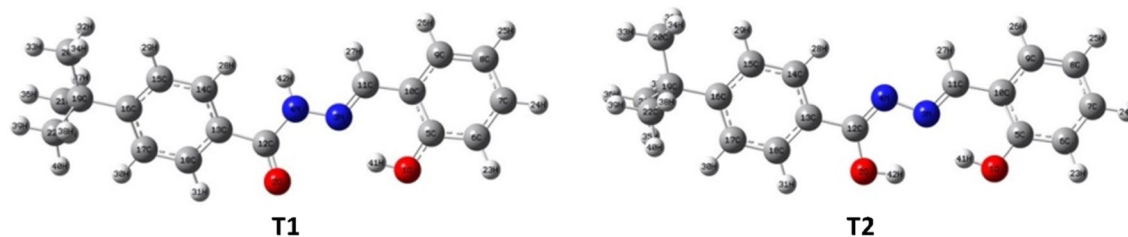


Fig. 3 Optimized geometry of two possible structure of Schiff base (H_2L) (T1 and T2) obtained at B3LYP/6-311++G(d,p) level of theory

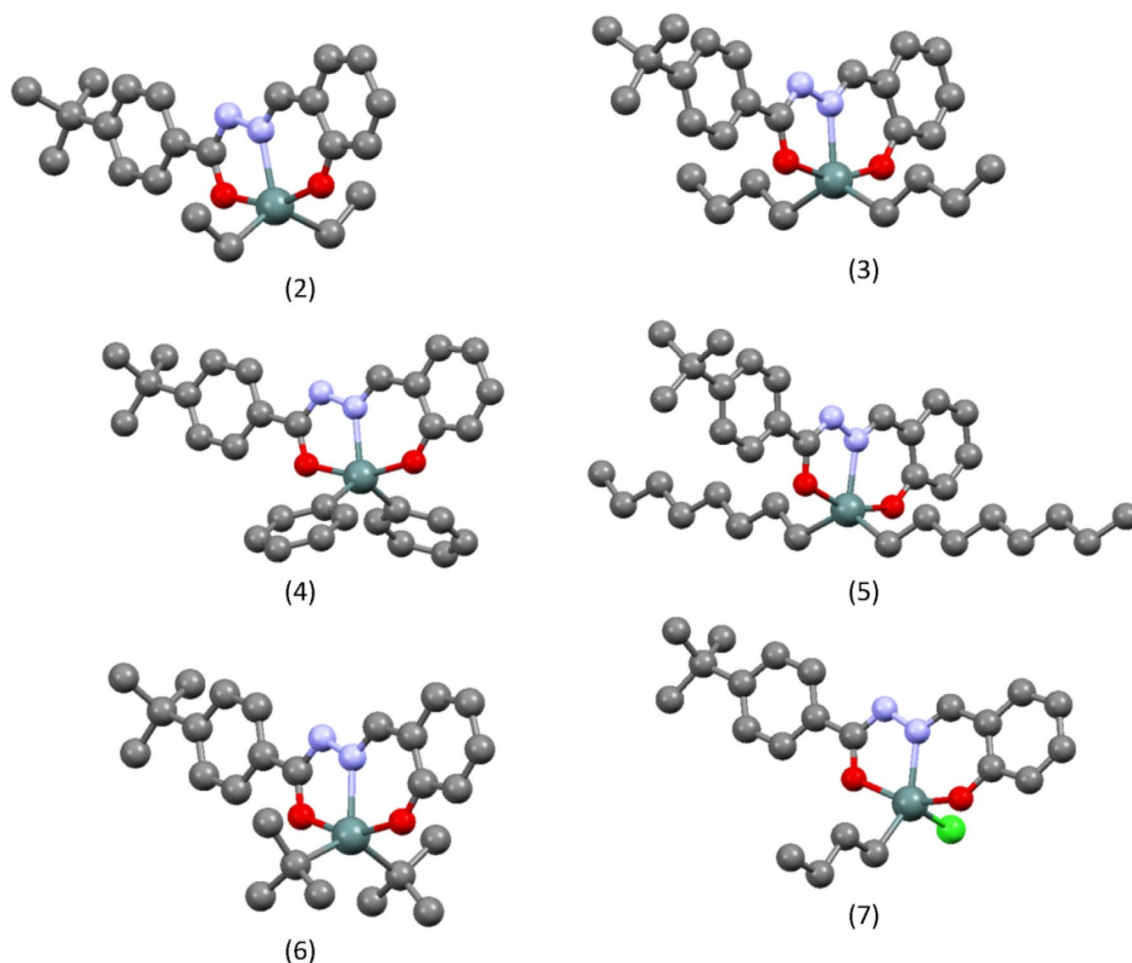


Fig. 4 Optimized structures of diorganotin(IV) complexes (2-7). Hydrogen atoms have been omitted for clarity

it was found that the synthesized compound show moderate activity against all the investigated bacterial strains, although some individual differences also existed among the tested bacteria; however, the diorganotin(IV) complexes (1-7) exhibited a better antibacterial activity than the ligand H_2L . No significant difference in the antibacterial activities of complexes against Gram-positive bacteria and Gram-negative bacteria has been observed. Among the tested bacteria, *Staphylococcus aureus* and *Escherichia coli* were most visibly inhibited by (4) and (7), respectively. The exact mechanism of action of synthesized compounds is yet to be investigated, however, Overtone's concept and Tweedy's chelation theory, which demands a better lipophilicity for antimicrobial action can be used to rationalized the enhanced activity of complexes (Fukushima et al. 2012; Zia ur et al. 2009). None of the synthesized compounds was more active than the standard drug.

Antifungal activity

The in vitro antifungal activities of ligand H_2L and diorganotin(IV) complexes (1-7) were tested against six human pathogenic fungal strains including yeasts (*Candida albicans* and *Candida glabrata*), dermatophytes (*Microsporum canis* and *Trichophyton longifusus*), and opportunistic molds (*Aspergillus flavus* and *Fusarium solani*) using the agar tube dilution protocol. Amphotericin B and Miconazole were used as standard drug. The results are presented in Table 8. Metallation usually boosts the antifungal activity; thus, the diorganotin(IV) complexes show a noticeably high antifungal activity than the ligand. Highest activity was shown by compound (6) against *F. solani*. Moderate activity against *A. falvis* and *M. canis* was shown by compounds (3) and (1), respectively. All the compounds showed insignificant activity against *Trichophyton longifusus* and *Candida glabrata*. The mechanistic details of inhibitory action against various fungal strains are yet to be explored; however, the

Table 5 Calculated geometric parameters [bond length (Å) and bond angles (°)] of diorganotin(IV) complexes (1–7)

	(1)	(2)	(3)	(4)	(5)	(6)	(7)	Exp.
Bond lengths (Å)								
O(1)–C(1)	1.312	1.311	1.311	1.316	1.311	1.312	1.321	1.324
O(2)–C(8)	1.301	1.301	1.300	1.305	1.300	1.301	1.307	1.299
N(1)–N(2)	1.384	1.384	1.384	1.382	1.382	1.384	1.379	1.412
N(1)–C(7)	1.305	1.305	1.305	1.303	1.305	1.305	1.306	1.298
N(2)–C(8)	1.316	1.316	1.316	1.314	1.315	1.315	1.314	1.296
C(8)–C(9)	1.480	1.480	1.480	1.479	1.480	1.481	1.475	–
Sn–N(1)	2.193	2.197	2.196	2.182	2.197	2.206	2.179	2.161
Sn–O(1)	2.075	2.083	2.083	2.066	2.080	2.089	2.037	2.065
Sn–O(2)	2.116	2.126	2.127	2.110	2.128	2.135	2.081	2.145
Sn–C(19)	2.122	2.138	2.137	2.126	2.135	2.187	–	2.119
Sn–C(20)	2.122	2.138	2.137	2.126	2.137	2.187	2.124	2.122
Bond angles (°)								
O(1)–Sn–O(2)	155.56	155.04	155.10	156.68	155.22	154.23	152.80	158.48
O(1)–Sn–N(1)	82.85	82.53	82.57	83.53	82.84	82.02	83.08	84.50
O(1)–Sn–C(19)	95.74	95.61	95.64	95.47	94.98	95.16	–	96.20
O(1)–Sn–C(20)	95.74	95.61	95.64	95.47	96.70	96.28	94.73	95.13
O(2)–Sn–N(1)	72.71	72.51	72.53	73.15	72.52	72.22	73.13	74.00
O(2)–Sn–C(19)	95.53	95.71	95.62	95.24	94.65	96.18	–	93.70
O(2)–Sn–C(20)	95.53	95.71	95.62	95.24	96.17	95.05	95.32	94.87
N(1)–Sn–C(19)	117.5	117.05	117.03	117.39	118.9	116.26	–	115.19

Table 6 Various theoretical molecular parameters

Parameters	H ₂ L	(1)	(2)	(3)	(4)	(5)	(6)	(7)
SCF Energy (kcal/mol)	– 601262.1450	– 652775.6812	– 702122.9925	– 800830.0047	– 893460.5399	– 998230.9442	– 990572.3344	– 800823.4798
Dipole moment (Debye)	6.349	0.417	0.261	0.256	0.803	0.206	3.328	0.381
ZPVE (kcal/mol)	216.531	248.016	284.322	355.389	316.020	498.178	280.057	354.077
Total thermal energy (kcal/mol)	229.425	264.566	302.469	376.960	336.216	526.537	298.613	375.646
C _v (cal/mol/K)	80.417	99.675	108.904	128.402	124.955	166.995	110.530	131.465
Total entropy, S (cal/mol/K)	152.105	182.150	194.835	223.802	211.254	280.351	199.433	215.815
E _{HOMO} (eV)	– 6.104	– 5.769	– 5.719	– 5.706	– 5.880	– 5.619	– 6.115	– 5.701
E _{LUMO} (eV)	– 1.976	– 2.232	– 2.198	– 2.186	– 2.305	– 2.078	– 2.506	– 2.194
E _{LUMO-HOMO} Gap (eV)	4.128	3.537	3.521	3.520	3.575	3.541	3.609	3.507
Global reactivity descriptors (eV)								
Ionization potential (IP), i.e., -HOMO	6.104	5.769	5.719	5.706	5.880	5.619	6.115	5.701
Electron affinity (EA), i.e., -LUMO	1.976	2.232	2.198	2.186	2.305	2.078	2.506	2.194
Electronegativity (χ)	4.040	4.001	3.959	3.946	4.093	3.849	4.311	3.948
Chemical potential (μ)	– 4.040	– 4.001	– 3.959	– 3.946	– 4.093	– 3.849	– 4.311	– 3.948
Electrophilicity (ω)	3.954	4.525	4.450	4.424	4.685	4.183	5.148	4.443
Hardness (η)	2.064	1.769	1.761	1.760	1.788	1.771	1.805	1.754
Softness (S)	0.242	0.283	0.284	0.284	0.280	0.282	0.277	0.285

Fig. 5 HOMO–LUMO plots of Schiff base ligand (H_2L) and diorganotin(IV) complexes (1–7)

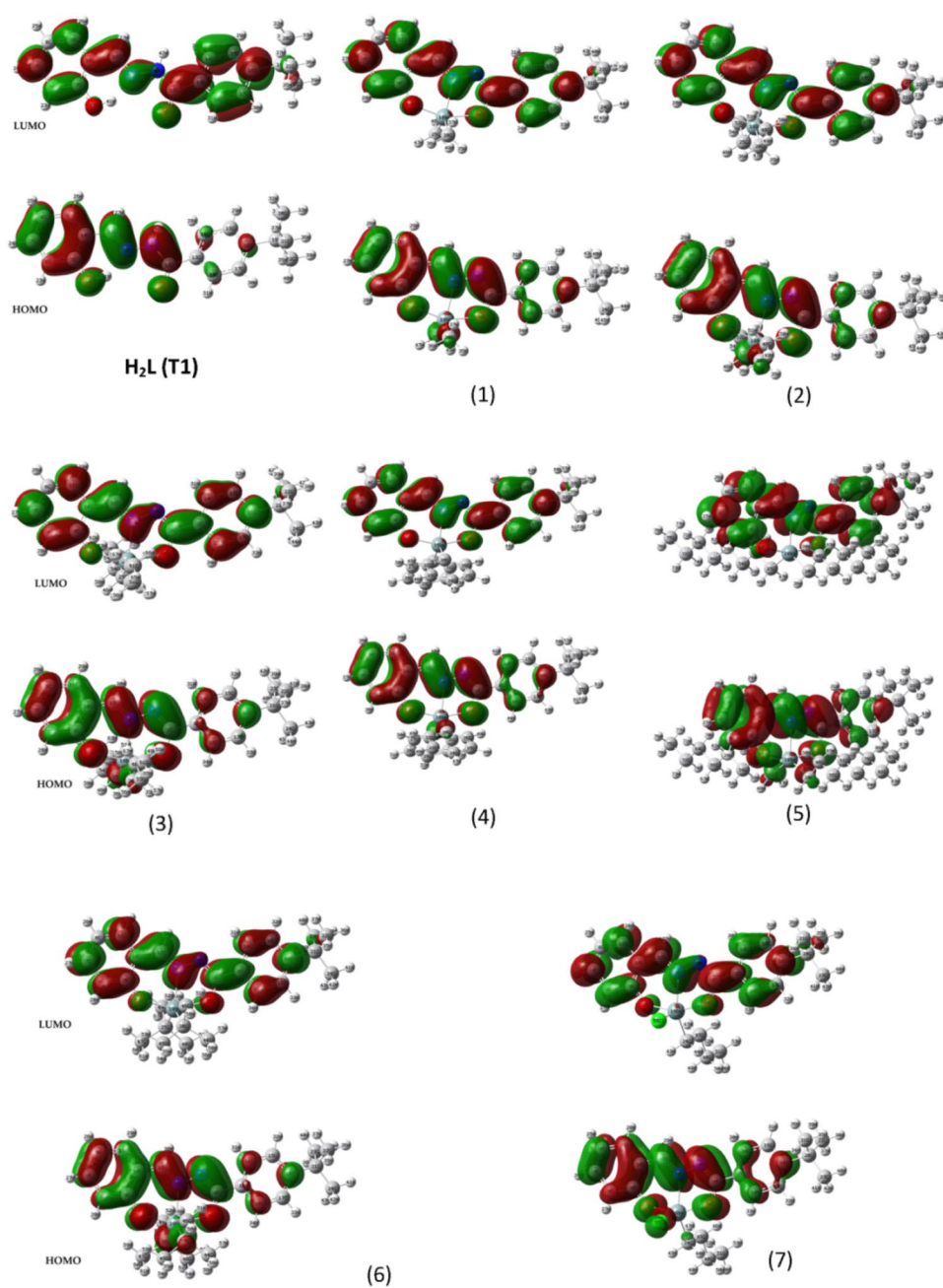


Table 7 Antibacterial activity of N' -(2-hydroxybenzylidene)-4-*tert*-butylbenzohydrazide (H_2L) and its organotin(IV) complexes

Bacterium	Inhibition zone diameter (mm)								Reference drug ^b
	H_2L	(1)	(2)	(3)	(4)	(5)	(6)	(7)	
<i>Escherichia coli</i>	–	14	–	15	13	–	12	16	30
<i>Bacillus subtilis</i>	–	–	12	15	–	–	10	11	37
<i>Shigella flexneri</i>	10	12	10	12	–	12	12	14	36
<i>Staphylococcus aureus</i>	–	11	–	10	15	–	13	–	26
<i>Pseudomonas aeruginosa</i>	–	10	–	8	–	10	–	–	32
<i>Salmonella typhi</i>	–	8	12	10	9	–	–	–	30

^aConc. 1 mg/mL of DMSO, ^bReference drug, Imipenem, – Insignificant activity

Table 8 Antifungal activity^{a-c} (% inhibition) of *N'*-(2-hydroxybenzylidene)-4-tert-butylbenzohydrazide (**H₂L**) and its organotin(IV) complexes

Fungus	Inhibition (%)							Standard drug ^c	MIC ^b (µg/mL)	
	H ₂ L	(1)	(2)	(3)	(4)	(5)	(6)			(7)
<i>Trichophyton longifusus</i>	–	–	–	–	–	–	–	–	Miconazole	70.0
<i>Candida albicans</i>	–	–	–	–	–	10	–	–	Miconazole	110.8
<i>Aspergillus flavus</i>	20	30	–	65	40	–	60	–	Amphotericin B	20.0
<i>Microsporium canis</i>	30	70	60	–	–	40	–	20	Miconazole	98.4
<i>Fusarium solani</i>	–	20	50	–	70	30	80	–	Miconazole	73.2
<i>Candida glabrata</i>	–	–	–	–	–	–	–	–	Miconazole	110.8

^aConcentration: 200 µg/mL of DMSO, ^bMIC: Minimum inhibitory concentration, ^cPercent inhibition (standard drug) = 100, – no inhibition

Table 9 Brine Shrimp (*Artemia salina*) lethality bioassay data of diorganotin(IV) derivatives of *N'*-(2-hydroxybenzylidene)-4-tert-butylbenzohydrazide

Sample code	(H ₂ L)	(1)	(2)	(3)	(4)	(5)	(6)	(7)
LD ₅₀ (µg/mL)	–	26.61	–	0.44	–	–	–	–

Standard drug: Etoposide, LD₅₀ 7.46 µg/mL

activity of synthesized compounds can be ascribed to their potential to interact with intracellular bioreceptors' triggering disruption in the movement of ribosome and obstructing nuclear protein and DNA synthesis with in the cell (Ayoko et al. 2003; Chaudhary et al. 2009).

Cytotoxicity

The synthesized ligand **H₂L** and diorganotin(IV) complexes (**1–7**) were assessed for cytotoxicity by in vivo lethality to brine shrimp nauplii (Table 4). The ability of organotin(IV) compounds to interact with the donor sites in DNA and proteins, their potential to stimulate oxidative damage and impede ATP synthesis and ability to cause apoptosis, necrosis or blockage of estrogen receptors are the primary factors responsible for their cytotoxic character (Shpakovsky et al. 2014; Tariq et al. 2014).

Among the synthesized compound, the dibutyltin(IV) derivative (**3**) displayed highest toxicity with LD₅₀ 0.44 µg/mL (Table 9), rest of the compounds does not show any significant toxicity against the Brine Shrimp (larvae).

Conclusions

Seven new diorganotin(IV) derivatives of *N'*-(2-hydroxybenzylidene)-4-tert-butylbenzohydrazide have been synthesized and characterized by elemental analysis, mass spectroscopy, FT-IR, multinuclear NMR (¹H, ¹³C and ¹¹⁹Sn), and DFT/B3LYP calculations. The ligand coordinates with the dialkyltin(IV) moieties in the iminol form through ONO donor sites. The ¹J(¹¹⁹Sn, ¹³C) coupling

constant values found were in the range 584–655 Hz and ²J(¹¹⁹Sn-¹H) coupling constants of 79 Hz for complex (**1**) suggest pentacoordination around Sn atom in solution. Single-crystal X-ray structure of ligand (**H₂L**) shows its existence in amido form. All the complexes exhibited a significant inhibitory activity against both Gram-positive and Gram-negative bacteria. Highest antifungal activity was shown by compound (**6**) against *Fusarium solani*. Compound (**3**) displayed highest cytotoxicity among the synthesized compounds with LD₅₀ 0.44 µg/mL.

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