RESEARCH ARTICLE

Ghrelin and Apolipoprotein AIV Levels Show Opposite Trends to Leptin Levels During Weight Loss in Morbidly Obese Patients

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Abstract

Background Although bariatric surgery is the most common procedure used to induce weight loss in morbidly obese patients, its effect on plasma satiety factors (leptin, ghrelin, and apolipoprotein (apo)-AIV) is controversial.

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J. M. Fort · J. A. Baena-Fustegueras Endocrinology Surgery Unit, Hospital Universitari Vall D'hebron, Institut De Recerca Vall D'hebron, Universitat Autónoma De Barcelona, Barcelona, Spain The aim of this work was to analyze these parameters before and at different times after surgery.

Methods Plasma was obtained from 34 patients before undergoing Roux-en-Y gastric bypass and during weight loss in the 12 months following surgery.

Results Morbidly obese patients had significantly higher values (147%) of leptin than normal-weight (NW) persons, while their ghrelin levels were 46% less than NW. Apo-AIV levels had approximately the same value in both groups (obese and NW). During weight loss, leptin decreased by 75% and ghrelin increased by 78%. Both parameters reached values less than or near NW, respectively, at 1 year after surgery. During the first month after surgery, apo-AIV plasma levels decreased (47%) but later increased and finally returned to preoperative values. Apo-AIV levels were correlated negatively with leptin and positively with ghrelin. High-density lipoprotein (HDL) levels were positively correlated with those of ghrelin and apo-AIV.

Conclusions During weight loss, plasma leptin and ghrelin could be good markers of total fat decrease. Ghrelin could also indicate gastric mucous improvement, whereas apo-AIV could indicate the recovery of intestinal function. Changes produced in the HDL levels of morbidly obese patients during weight loss suggest a decreased risk of coronary disease.

Keywords Bariatric surgery \cdot RYGBP \cdot HOMA-IR \cdot HDL \cdot Satiety

Abbreviations

HOMA-IR	homeostasis model assessment of insulin
	resistance
NEFA	nonesterified fatty acid
TC	total cholesterol

Introduction

Surgical weight loss approaches are based on intestinal malabsorption and gastric reduction. "Success" has been defined as a maintained weight loss of \geq 50% of excess body weight, which in turn is defined as the difference between body weight and ideal weight (for a given patient's gender and height) [1]. The Roux-en-Y gastric bypass (RYGBP) has become the gold standard of bariatric surgery. It represents a mixed technique, combining restriction derived from leaving a small stomach pouch near the esophagogastric junction by excluding the greater curvature, together with a small malabsorptive component derived from bypassing most of the stomach and duodenum [1].

Leptin and ghrelin are peripheral signals that contribute to the central regulation of food intake. Body mass index (BMI) is the best predictor of circulating leptin concentrations. Unlike total body fat, the amount of upper or lower body adiposity or visceral fat has been found not to influence basal leptin concentrations [2]. Despite the strong correlation between body fat and leptin concentrations, there is great heterogeneity in leptin concentrations for all body fat indices [3, 4].

Leptin, a hormone released by adipocytes, provides information about energy status to hypothalamic regulatory centers [5]. In humans, circulating leptin levels decrease or increase in response to acute caloric shortage or surplus, respectively [6]. There is some evidence linking leptin to direct regulation of adipose tissue metabolism through inhibition of lipogenesis and stimulation of lipolysis [7]. Circulating leptin concentrations are elevated in obesity and decrease after weight loss [8].

Ghrelin, a peptide produced predominantly by the stomach and particularly in the fundus [9], is involved in energy balance regulation. However, in contrast to the anorexigenic effects of leptin, ghrelin stimulates appetite [10]. In human obesity, ghrelin concentrations are low. This might be related to high caloric intake, as body weight reduction in obese patients increases the concentration of ghrelin [8].

The observed reduction of circulating ghrelin concentrations in obese patients at 6 months after RYGBP surgery is not determined by active weight loss or improved insulin sensitivity but rather depends on the surgical bypass of the ghrelin-producing cell population of the fundus [11]. Other investigators have reported that stomach bypass surgery decreases ghrelin concentrations, suggesting that the size of the stomach might correlate directly with ghrelin concentrations [12]. However, other reports have shown a significant increase of serum ghrelin 12 months after biliopancreatic diversion [13]. Some reports have found that insulin is a physiological and dynamic modulator of plasma ghrelin levels and that insulinemia may mediate the effects of nutritional status on ghrelin concentration [14], whereas other reports have shown no relationship between ghrelin levels and other parameters [15]. Following the hypothesis of Cummings et al. [16] that ghrelin has a role in the long-term regulation of body weight, multiple studies have demonstrated increased [12], decreased [17], or unchanged [18] ghrelin levels after weight loss. Recently, it was reported that an immobilized form of ghrelin could specifically bind to different types of HDL associated with plasma sterase, paraoxonase, and clusterin [19].

Human apolipoprotein (apo)-AIV is synthesized and secreted exclusively by the small intestine into enterocytes. The jejunum is the major site of apo-AIV synthesis, but apo-AIV is also produced in the duodenum and ileum. Apo-AIV secretion is stimulated by lipid absorption [20]. Apo-AIV synthesis and secretion into the lymph are associated with chylomicron formation, as digested lipids are processed in mucosal cells. The majority of apo-AIV in the circulation exists as free protein with the remainder associated with circulating HDL [21]. Apo-AIV is also synthesized in the hypothalamus and has been proposed as an important signal for satiety in the central nervous system [22]. The effects of leptin and apo-AIV combine, perhaps synergistically, to reduce food intake [23]. Vergès et al. [24] have demonstrated a strong correlation between plasma apo-AIV and postprandial triacylglyceride (TAG) metabolism in both normoponderal and obese subjects, but these results could not be confirmed by other investigators [25]. Plasma apo-AIV decreases markedly in overweight adolescents undergoing short-term weight reduction, but this decrease is not related to the degree of weight loss, BMI, or plasma leptin. It has been demonstrated that common apo-AIV variants are associated with differences in BMI and body fat percentage [25, 26].

The main objective of this study was to investigate changes in hormone levels (leptin, ghrelin, and insulin) in morbidly obese patients before and after RYGBP and to study the relationship of plasma apo-AIV levels with other components of the lipid plasma profile and anthropometric parameters. We followed the patients' progress through 1 year of weight loss.

Methods

Patient Selection

A group of 34 morbidly obese patients (24 women and ten men) aged between 27 and 61 years and with a BMI> 40 kg/m^2 were recruited from the Vall d'Hebron Hospital in

Barcelona, Spain. The obesity criterion was in accordance with the Spanish consensus for the diagnosis of obesity [27]. Sixty-eight percent of patients had metabolic syndrome (Adult Treatment Panel III Criteria).

All subjects were free of inflammatory and infectious diseases and none were receiving anti-obesity or antiinflammatory drugs at the time of the study. Patients were excluded if they had neoplastic, renal, hepatic, or active systemic diseases, hypothyroidism or endocrine diseases other than diabetes, or if they had been on a restrictive diet during the week previous to the study. All patients reported that their weight was stable during the previous 3 months. None of the diabetic patients were being treated with insulin. The study protocol was reviewed and accepted by the hospital ethics committee and all subjects gave their written informed consent to participate.

For 2 days before the study, all subjects were placed on an isocaloric diet calculated on the basis of individual requirements. The diet was made up of 50% carbohydrates, 20% protein, and 30% fat. Blood samples were taken under fasting conditions at 8:00 and 10:00 AM 1 month before surgery (in tables and graphs: OB) and 1, 3, 6, 9, and 12 months after surgery (in tables and graphs: 1M, 3M, 6M, 9M, 12M). Plasma was separated immediately by centrifugation (2,000×g, 30 min, 4°C) and aliquots were frozen at -80° C for subsequent analysis.

After surgery and during the study, the patients' diet was: during the first month, a liquid diet of 810 kcal/day (45% carbohydrates, 33% protein, and 22% fat); during the second month, a triturated diet of 839 kcal/day (44% carbohydrates, 38% protein, and 18% fat); during the third month, a solid diet of 844.5 kcal/day (41% carbohydrates, 31% protein, and 28% fat); during the rest of the study, the patients were placed on a maintenance diet of 825 kcal/day (44% carbohydrates, 34% protein, and 22% fat).

Seven patients with normal weight were used as controls (in tables and graphs, NW). These patients were euthyroid (12 h fasted), normolipidemic, without digestive system disease, and underwent a laparotomy at the same time as blood was drawn.

Anthropometric and Body Composition Measurements

Body weight, excess weight, height, and waist and hip circumferences were measured according to standard procedures [28]. Weight and height were determined for all subjects, and the BMI was then calculated. Waist circumference was taken midway between the lower rib margin and the iliac crest. Hip circumference was determined as the widest circumference measured on the great trochanter. The waist-to-hip ratio (WHR) was then calculated. The percentage of body fat was calculated from the equation proposed by Deurenberg et al. [29], and the quantity of total subcutaneous and visceral fat was calculated from equations proposed by Bonora et al. [30].

Blood Assays

Insulin levels were determined using an IMMULITE 2500 autoanalyzer (Siemens Medical Diagnostics) in an immunometric chemiluminescent test with two binding sites (noncompetitive) on solid phase. Ghrelin (total: acylated and deacylated) was determined by competitive radioimmunoassay (catalog no H-4864, Peninsula Laboratories, Inc., Bachem, San Carlos, CA, USA) using ¹²⁵I and a double antibody as markers. This process requires the previous purification and concentration of the sample by chromatography on a Sep-Pak C18 (Waters) column. Leptin was determined by direct competitive radioimmunoassay with a double antibody and ¹²⁵I as the radioisotope (DRG Instrument GmbH, Germany).

Plasma protein was determined using the method developed by Bradford [31]. Fasting plasma glucose, TAG, nonesterified fatty acid (NEFA), total cholesterol (TC), and high- (cHDL) and low-density (cLDL) lipoprotein cholesterol were measured enzymatically in the hospital's routine chemistry laboratory. Total HDL quantification was done with a commercial kit for agarose gel electrophoresis (MIDIGEL LIPO Kit made by BIOMIDI, Toulouse, France). The homeostasis model assessment of insulin resistance (HOMA-IR) was calculated as previously described [32].

Apo-AIV Western Blot

We ran a single gel for each patient that was loaded with samples from before (obese) and after surgery (during weight loss) and plasma sample from NW as a control. Western blot (WB) assays to detect apo-AIV were run using 10% sodium dodecyl sulfate gel electrophoresis and transferred to Immobilon membranes of Millipore®. IgG anti-human apo-AIV obtained from rabbits in our laboratory was used as the primary antibody at 1/500 dilution (with blocking solution). Several years ago, our group was issued a patent on the use of human apo-AIV antibodies for clinical diagnosis [33]. Horseradish peroxidase (HRP)conjugated pig anti-rabbit IgG at 1/15,000 dilution (with blocking solution) was used as the secondary antibody (Dako, Glostrup, Denmark). Non-specific binding was prevented using a blocking solution (phosphate-buffered saline (PBS) with bovine serum albumin (BSA)) at 2% (w/v, BSA/PBS). WBs were developed using the Super-Signal WestPico Chemiluminescent substrate for HRP detection (Pierce[®], Rockford, IL, USA). Relative quantities were estimated by densitometric scanning (Phoretix 1D gel Analysis Software Non-linear Dynamics, Newcastle, UK).

Statistical Analysis

Results are given as mean±SEM. Statistical differences between mean values for obese and NW (control) at 1, 3, 6, 9, or 12 months after surgery (weight loss) were assessed using a one-way analysis of variance (ANOVA). Individual comparisons were made using Tukey's multiple-comparison test. Statistical differences between women and men at different months after surgery (weight loss) were assessed by a two-way ANOVA and Bonferroni posttests. Correlations between independent variables were determined by Spearman's correlation coefficient (rho). Statistical comparisons were considered significant at p<0.05.

All statistical analyses were computed using GraphPad Prism version 4.00 software for Windows (GraphPad Software, San Diego CA, USA, www.graphpad.com).

Results

Clinical Characteristics of Patients

The characteristics of the studied patients are shown in Table 1. The loss of body weight was already significant at 1 month after surgery and weight loss continued until it reached 37% after 1 year. The observed decrease represented a loss of 71% of the excess weight (calculated as the difference between the theoretical ideal weight and the patient's actual weight), a 27% decrease in the circumference of the waist, a 19% decrease in hip circumference (see Table 1), and a significant decrease in WHR.

Of the total body weight (Table 1), 61% corresponded to fat, and 73% of this percentage corresponded to subcutaneous fat and 27% to visceral fat. The percentage of fat loss during the first 6 months after bariatric surgery was similar

for total fat, subcutaneous fat, and visceral fat (39%, 40%, and 38%, respectively). Over the next 6 months, the total fat loss was decreased: 15% of total or visceral fat and 11% of subcutaneous fat. In all cases, the changes were highly significant (Table 1).

Biochemical Plasma Parameters

TC in morbidly obese patients was significantly higher than in NW subjects, but during the first month after surgery this parameter completely recovered to normal levels (Table 2). During weight loss, cHDL remained at similar levels and was even higher than in NW subjects, whereas cLDL levels diminished throughout the months after surgery, reaching even lower values than in control subjects (Table 2). The largest difference was observed in plasma NEFA concentrations, which were twofold higher in morbidly obese patients as compared to NW subjects. NEFA levels even increased during the first month and remained elevated to a similar degree as the initial obese state throughout the study period (Table 2).

Glucose and the insulin levels in the morbidly obese patients were significantly above those of NW subjects (Table 2), but during the first month after surgery the glycemic level diminished to 24% and then remained at that level during the rest of the studied period (Table 2). Insulin diminished by 43% during the first month but after 3 months remained stable for the rest of the study period (Table 2). HOMA-IR reference values considered to represent insulin resistance are around 3.5–3.8 [33]. In the present study, morbidly obese patients had HOMA-IR values more than 3.5-fold higher than those of NW subjects. However, from the first month after bariatric surgery, HOMA-IR decreased by 57% to a point where we considered there was no insulin resistance (Table 2).

 Table 1 Clinical characteristics of obese patients before and after bariatric surgery

	OB	Period of weight loss								
		1M	3M	6M	9M	12M	p (ANOVA)			
Weight (kg)	130.3±3.4	115.2±3.2 o	103.5±2.8 o	93.4±2.8 o,1	86.1±3.0 o,1,3	82.1±2.9 o,1,3	< 0.0001			
Excess weight (kg)	65.4±2.6	49.9±1.3 o	38.4±2.2 o,1	28.5±2.2 o,1,3	21.8±2.3 o,1,3,6	19.2±2.3 o,1,3,6	< 0.0001			
BMI (kg/m ²)	$48.8 {\pm} 0.9$	42.3±1.6 o	38.7±0.8 o	35.0±0.8 o,1	32.1±0.9 o,1,3	30.9±0.9 o,1,3	< 0.0001			
Waist (cm)	136.1 ± 2.3	120.2 ± 5.4	112.0±3.7 o	105.1±4.5 o	91.4±3.0 o,1,3	98.5±2.9 0,1	< 0.0001			
Hip (cm)	145.0 ± 2.3	132.6±4.9	126.2±4.2 o	123.8±4.5 o	115.5±9.5 o	117.8±3.1 o	< 0.0001			
TAT (kg)	78.7±3.1	66.2±2.5 o	53.5±2.0 o,1	44.4±2.1 o,1	37.8±2.1 o,1,3	35.2±2.1 o,1,3	< 0.0001			
SAT (kg)	58.5 ± 2.7	46.8±5.7	38.1±3.2 o	35.4±3.3 o	24.3±5.0 o	30.5±2.4 o	< 0.0001			
VAT (kg)	20.9 ± 1.3	13.2 ± 2.6	14.3 ± 2.5	12.9±2.0 o	10.3±1.9 o	10.6±2.1 o	0.0001			

Results are expressed as mean±SEM. Statistics were computed using one-way ANOVA and individual comparisons were made with Tukey's multiple-comparison test between groups. Symbols denote differences between obese and 1, 3, 6, 9, or 12 months (M) after surgery classified by (o), 1M vs. rest of months (1), 3M vs. rest of months (3), 6M vs. rest of months (6). One symbol: p<0.01

BMI body mass index, TAT total adipose tissue, SAT subcutaneous adipose tissue, VAT visceral adipose tissue, OB obese

Table 2	Plasma	parameters	of o	obese	patients	before	and	after	bariatric	surgery
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	NW	OB	Period of weight loss							
			1M	3M	6M	9M	12M	p (ANOVA)		
TC (mg/dl)	164.3±2.9	204.6±9.2 c	149.4±7.8 o	151.2±7.9 o	168.5±9.2 o	171.9±8.3	162.2±4.5 o	0.0001		
cHDL (mg/dl)	$42.7 {\pm} 0.9$	47.2 ± 2.1	33.5±1.2 c,o	38.4±1.2 o,1	44.5±1.2 1	49.2±2.0 c,1,3	51.0±1.8 c,1,3	< 0.0001		
cLDL (mg/dl)	133.3 ± 3.0	128.1 ± 6.3	91.3±6.8 c,o	97.0±5.5 c	103.2±7.3 c	99.5±5.8 c	92.2±3.9 c,o	< 0.0001		
NEFA (mg/dl)	8.7±0.4	16.3±3.2 c	29.2±4.2 c,o	18.7±1.9 c,1	14.5±2.0 c,1	14.4±3.4 c,1	15.6±2.8 c,1	< 0.0001		
TAG (mg/dl)	57.8 ± 2.1	146.9±12.3 c	147.1±10.6 c	130.2±8.9 c	115.4±8.8 c	104.6±9.9 c,o,1	95.1±5.7 c,o,1,3	< 0.0001		
Gluc (mg/dl)	72.0±2.1	126.3±9.6 c	96.2±3.4 c,o	94.0±2.7 c,o	91.7±2.3 c,o	90.5±2.0 c,o	91.1±3.0 c,o	< 0.0001		
Insulin (UI/L)	$11.0 {\pm} 0.8$	24.3±2.9 c	13.9±1.6 o	11.7±1.1 o	9.4±0.9 o,1	8.3±0.9 c,o,1,3	8.0±0.8 c,o,1,3	< 0.0001		
HOMA-IR	$2.0{\pm}0.1$	7.7±1.1 c	3.3±0.5 c,o	2.7±0.3 c,o	2.2±0.2 o,1	1.9±0.3 o,1	1.9±0.3 o,1	< 0.0001		

Data are expressed as mean±SEM. Statistics were computed using one-way ANOVA and Tukey posttest between groups. Symbols denote differences between normal weight (c) and obese or 1, 3, 6, 9, or 12 months (M) after surgery classified by (o), 1M vs. remaining months (1) and 3M vs. remaining months (3). Symbol: p < 0.01.

TC total cholesterol, cHDL cholesterol in HDL, cLDL cholesterol in LDL, NEFA nonesterified fatty acid, TAG triacylglycerides, Gluc glucose, HOMA-IR homeostasis model assessment-insulin resistant, NW normal weight, OB obese

Leptin and Ghrelin Levels in Obese Patients Before and After RYGBP

Fig. 1 (top panel) shows that leptin levels in obese patients before surgery was 60% higher than in NW subjects (dotted line in the middle of the shaded zone showing mean \pm SEM; 15.0 \pm 0.9 ng/ml plasma). At 6 months after surgery, leptin levels had already recovered to NW levels and continued to decrease to approximately 50%.

The ghrelin profile (Fig. 1, middle panel) was nearly the opposite of that observed for leptin. The dotted line in the middle of the shaded zone indicates the mean±SEM for NW values: 132.0 ± 0.3 pg/ml plasma. In obese patients, ghrelin levels were 46% lower than in NW subjects but began to increase only 1 month after surgery. After 1 year, ghrelin levels had recovered completely to NW levels; the difference was significant only at this point due to high variability. The correlation between leptin and ghrelin was negative (rho=-0.16) and significant (p < 0.05).

Leptin levels in obese patients were positively and significantly correlated with the BMI (rho=0.49, p<0.01), percentage of body fat (rho=0.59, p<0.001) and, to a lesser extent, with subcutaneous fat (rho=0.44, p<0.05). Ghrelin levels in the morbidly obese were not correlated with any of these parameters. Thus, where leptin levels were positively and significantly correlated with weight loss in all cases (p<0.001; Table 3), ghrelin levels were negatively and significantly correlated with weight loss (p<0.01) only in several cases (Table 3).

Apolipoprotein AIV and HDL Levels in Obese Patients and After RYGBP

The levels of plasma apo-AIV in obese patients (Fig. 1, bottom panel) were similar to those of NW individuals

(dotted line in the middle of the shaded zone indicates mean±SEM; 2.02±0.3 arbitrary units). One month after surgery, however, these apo-AIV levels in obese patients had decreased by 47% and then began to recover gradually until they had returned to NW levels after a year. In obese patients, correlations with other parameters were not observed; however, during weight loss, apo-AIV levels were correlated negatively with those of leptin (rho=-0.33, p<0.05) and positively with those of ghrelin (rho=0.89, p<0.001). Additionally, other correlations were observed for anthropometric or biochemical parameters (Table 3).

In order to verify that apo-AIV and HDL levels were related in obese individuals as described in other studies [20], we used agarose gels (0.5%) dyed with Sudan black (specific for lipids) to detect the total amount of plasma lipoprotein. Fig. 2 shows that the total amounts of HDL and apo-AIV (obtained by Western blotting) were nearly identical. The correlation between both parameters was rho=0.82 (p<0.01).

Discussion

This study presents, for the first time, changes in the plasma profile of several satiety factors as well as lipid parameters for morbidly obese patients receiving RYGBP surgery. The evolution of these factors and parameters was studied for 1 year after surgery, and we observed a recovery towards NW individual levels that was concomitant with the normalization of anthropometric measurements (body weight, body fat, etc.).

It should be noted that bariatric surgery is one of the most effective techniques for weight loss in morbidly obese patients and also makes it possible to quickly recover insulin sensitivity and correct dyslipidemia [34]. In the



Fig. 1 Plasma leptin, ghrelin, and apo-AIV. Results are expressed as mean±SEM. Statistics were computed using one-way ANOVA and individual comparisons were made using Tukey's multiple-comparison test between groups. The *numbers* in the *left* and *right corner* of each *panel* depict the ANOVA result. The results of the apo-AIV graph represent the mean±SEM densitometry values for all gels (one for each patient) in arbitrary units. The NW value for each graph is indicated by a *dotted line* in the middle of a shadow zone that indicates mean±SEM. N=34 patients for each graph. *Symbols* denote differences between NW (control) and different groups (*c*); between obese (*OB*) and 1, 3, 6, 9, or 12 months (*M*) after surgery classified by (*o*), 1M vs. remaining months (*I*), 3M vs. remaining of months (*3*). *One symbol:* p<0.05; *two symbols:* p<0.01; *three symbols:* p<0.001

present study, subjects lost approximately 37% of their body weight in the year after surgery, as well as 47% of their subcutaneous fat and the same amount of visceral fat; correspondingly, HOMA-IR was reduced by 48% in the first month. The use of other techniques, such as individualized diets with simultaneous physical exercise [35] or liquid diets to maintain a "baseline weight" [36], does not improve other co-morbidities. Phillips et al. [37] reported that, 3 months after adjustable laparoscopic gastric banding surgery in obese patients, no improvement in insulin sensitivity was seen despite 20% and 15% reductions of subcutaneous and visceral fat, respectively. Reinehr et al. [38] observed decreased leptin and improved insulin resistance, which we also observed in the present study.

During weight loss, we observed a great improvement in plasma lipid profile (TAG, TC, and cHDL) similar to that described by other investigators [39] for the same type of surgery. In contrast to other studies that did not observe any relationship with decreased BMI, body fat, glucose, triacylglycerides, cholesterol, insulin, or leptin in patients who received RYGBP [40], we observed that all these parameters had changed significantly by 1 month after surgery and continued to change throughout at least the first year after surgery.

Leptin and Ghrelin

Total plasma leptin usually reflects the amount of fat present in the body, and lower levels have been consistently reported in patients losing weight [41]. Our study showed that significant reduction of leptin levels occurred early: 1 month after surgery, when BMI and total adipose tissue were still high but significantly different from the presurgical (obese) state. A similar result was reported previously in a 9-month study [34], but we additionally observed that leptin levels continued to decrease up to at least 1 year after surgery. Contrary to other reports [42], leptin levels were correlated with BMI, the total fat percentage, subcutaneous fat, and visceral fat, both in the obese state and after surgery. Moreover, the progressive decrease in plasma leptin levels did not appear to depend on the type of diet or on the number of ingested calories. According to the literature, changes in plasma leptin during weight loss mainly reflect changes in metabolic rate, decreased adipose tissue, decreased size of ingested meals, etc. [43]. Liou et al. [44] recently reported that leptin levels in morbidly obese patients were significantly reduced at 1, 3, 6, and 12 months after laparoscopic minigastric bypass surgery, similar to the present results; however, ghrelin concentrations were not significantly changed after surgery in contrast to our results.

Frübeck et al. [11] found that reduced circulating ghrelin concentrations 6 months after RYGBP surgery were not generated by active weight loss or improved insulin sensitivity but rather by the decreased size of secretory zone of the hormone (the fundus of the stomach) [11]. In our study, the disappearance of a large part of the stomach after RYGBP (the stomach was reduced to about 200 cm³, but a portion of the fundus might have remained active) was likely not enough to result in reduced plasma ghrelin. Additionally, ghrelin levels were lower in morbidly obese patients than in NW subjects but increased to NW levels during weight loss. We therefore consider that loss of the fundus likely did not cause the reduction of ghrelin levels because this effect was not seen at all times studied.

Table 3 Correlations between parameters during weight loss

	Leptin		Ghrelin		Apo-AIV	
	rho	р	rho	р	rho	р
BMI	0.73	< 0.001	-0.20	< 0.01	_	_
Weight	0.56	< 0.001	-0.26	< 0.01	-	_
Excess weight	0.73	< 0.001	-0.31	< 0.01	_	_
Hip circumference	0.58	< 0.001	-	-	-	_
Waist circumference	0.61	< 0.001	-0.31	< 0.05	_	_
Body fat (%)	0.59	< 0.001	-0.30	< 0.01	-0.44	< 0.01
Subcutaneous fat (kg)	0.63	< 0.001	-	_	_	_
Visceral fat (kg)	0.30	< 0.05	-	-	-	_
Insulin	0.37	< 0.001	-0.26	< 0.01	-	_
Glucose	0.31	< 0.001	-	_	-0.47	< 0.01
HOMA-IR	0.41	< 0.001	-	_	_	_
TC	0.35	< 0.001	-	_	_	_
HDL	_	-	0.69	< 0.001	0.82	< 0.01
TAG	0.47	< 0.001	-	_	_	_
Glycerol	0.53	< 0.001	_	_	-0.58	< 0.01

Correlations between independent anthropometric and body composition variables or biochemical parameters with leptin, ghrelin, or apo-AIV plasma levels during weight loss. Correlations were determined by Spearman's correlation coefficient (rho). Statistical comparisons were considered significant at P < 0.05

HOMA-IR homeostasis model assessment-insulin resistant, TC total cholesterol, HDL high-density lipoprotein, TAG triacylglycerides

Fig. 2 Plasma apo-AIV and HDL. Results represent the mean±SEM densitometry values for all gels (one for each patient) in arbitrary units. Statistics were computed using oneway ANOVA and individual comparisons were made using Tukey's multiple-comparison test between groups. The numbers in the top or bottom corner represent ANOVA results for apo-AIV (squares) and HDL (dots), respectively. Arrow shows the application line of plasma. N=34 patients for each graph. Symbols denote differences between NW (control) and different groups (c); between obese (OB) and 1, 3, 6, 9, or 12 months (M) after surgery classified by (o), 1M vs. remaining months (1), 3M vs. remaining months (3). One *symbol*: *p*<0.05; *two symbols*: *p* <0.01; *three symbols*: *p*<0.001



Nevertheless, we cannot discard that other intestinal cells could also secrete ghrelin, resulting in a net increase in plasma levels. In the present study, ghrelin levels were significantly correlated not only with weight loss but also with decreased insulin resistance; these results are in accord with previous studies [13, 45].

Although many studies [45, 46] have shown that ghrelin levels are downregulated in human obesity, the existing data on ghrelin levels during weight loss are contradictory. For example, after biliopancreatic diversion surgery, some investigators have observed increased ghrelin levels [13], whereas others observed diminished levels [46]. Still other investigators [47] have found that, during weight loss, ghrelin levels increase by 12% in the first 6 months, whereas in the present study we observed a larger increase (39%) in the same time interval. Other studies did not observe any substantial changes in ghrelin levels during weight loss in children and obese adolescents submitted to a low-fat high-carbohydrate diet, nor did they observe any correlation with insulin levels [39]. In the present study, we observed a negative correlation between ghrelin and leptin or insulin levels.

We also observed significant differences between women and men in plasma leptin at all times after surgery but not in plasma ghrelin. Although women had higher leptin levels than men (p<0.001), ghrelin levels were similar across genders. Gender explained 23% of the variation in leptin levels; this result was similar that observed by Saad et al. [47].

Apo-AIV and HDL

Few data in the literature describe the relationship between apo-AIV and obesity, and the majority of this work was done in rodents. It was only recently suggested that apo-AIV could act as a satiety factor [22], but current evidence supports an important role in the regulation of food intake [48]. In the present study, morbidly obese patients had apo-AIV levels similar to those of NW individuals, but, as with HDL, levels decreased by 50% within 1 month after surgery. This decrease was not only related to reduced duodenum size, as the duodenum is not the primary source of apo-AIV; rather the observed decrease in apo-AIV was likely the result of the general (not only lipids, which stimulate the apo-AIV secretion) decrease in caloric ingestion [49]. We observed 1 month after surgery that plasma apo-AIV levels began to increase until they reached NW levels. Similarly, HDL levels increased. However, during this period, patients did not increase their lipid ingestion. Therefore, the change in apo-AIV levels might have resulted from increased total intestinal absorption due to increased numbers of enterocytes or development of vellosities. Although not addressed in the present study, our

research group has previously described that, in human intestinal atrophy, plasma apo-AIV diminishes [31, 50] due to reduced activity or numbers of enterocytes. The present HDL and apo-AIV data further reinforce the concept that human plasma apo-AIV is synthesized exclusively by the intestine and is at least partly associated with HDL as previously described [51].

Some investigators [25] studying weight loss in children and obese adolescents enrolled in a low-fat diet did not observe any relationship between plasma levels of apo-AIV and TAG; this finding agrees with our data. However, the same study found no correlation between levels of apo-AIV and changes in BMI or plasma leptin; these findings are in contrast to ours.

In ob/ob mice and human small intestine epithelial CACO-2 cells, respectively, Shen et al. [23] and Morton et al. [52] observed decreased apo-AIV levels after leptin administration. We also found a relationship between leptin and apo-AIV levels; women had higher plasma leptin and lower apo-AIV levels, and men showed the opposite.

Thus, circulating leptin levels increase as individuals become more obese, whereas the intestinal apo-AIV response to lipids is attenuated [20]; during weight loss, this situation is inverted. It is possible, as suggested by other investigators, that the effects of leptin and apo-AIV combine, perhaps synergistically, to reduce food intake [23].

In the present study, plasma apo-AIV levels significantly varied between women and men at all times after surgery. Men had higher apo-AIV levels than women (p < 0.05), and gender explained 10% of the variance in plasma apo-AIV levels. Van Aalst-Cohen et al. [53] genotyped a cohort of 1,002 heterozygous familial hypercholesterolemia patients for polymorphisms in the gene encoding apo-AIV and other lipid genes. Multiple linear regressions showed that, together, polymorphisms could explain only 3.9% of the variation in HDL-C plasma levels. Application of a regression model that also incorporated sex, smoking, alcohol use, body mass index, and concomitant betablocker use as covariates increased the single-nucleotidepolymorphism-attributed variation of cHDL plasma levels to 32.5% [53]. Recently, a link was suggested between ghrelin and HDL subtypes; some investigators [19] have also proposed a limited number of binding sites for ghrelin in HDL. However, we only observed a good correlation between ghrelin and HDL levels over time. As only one study [19] has thus far examined relationships between these factors in detail, a future extensive study about apo-AIV and HDL levels and their relationship with obesity and cardiovascular diseases is important.

Conclusion The present study found that morbidly obese patients who underwent the RYGBP procedure to reduce

body weight had a greatly improved lipid profile at 1 year after surgery, likely resulting in decreased risk of coronary disease. In addition, they showed decreased resistance to insulin. The study of changes in plasma levels of leptin, ghrelin, and apo-AIV after surgery might allow better diagnosis of fat loss and recovery of gastro-intestinal functionality, which is important for clinical monitoring of patients.

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