REVIEW

Progress in tumor vascular normalization for anticancer therapy: challenges and perspectives

Bingxue Shang, Zhifei Cao, Quansheng Zhou (🖂)

Cyrus Tang Hematology Center, Jiangsu Institute of Hematology, the First Affiliated Hospital of Soochow University, Suzhou 215123, China; Key Laboratory of Thrombosis and Hemostasis, Ministry of Health, Soochow University, Suzhou 215123, China

© Higher Education Press and Springer-Verlag Berlin Heidelberg 2012

Abstract Antitumor angiogenic therapy has been shown promising in the treatment of several advanced cancers since the approval of the first antiangiogenic drug Avastin in 2004. Although the current antiangiogenic drugs reduce the density of tumor blood vessels and result in tumor shrinkage at the early stage of treatment, recent studies have shown that antiangiogenic therapy has transient and insufficient efficacy, resulting in tumor recurrence in patients after several months of treatment. Blockage of blood and oxygen supplies creates a hypoxic and acidic microenvironment in the tumor tissues, which fosters tumor cells to become more aggressive and metastatic. In 2001, Jain proposed tumor vascular normalization as an alternative approach to treating cancers based on the pioneering work on tumor blood vessels by several other researchers. At present, normalizing the disorganized tumor vasculature, rather than disrupting or blocking them, has emerged as a new option for anticancer therapy. Preclinical and clinical data have shown that tumor vascular normalization using monoclonal antibodies, proteins, peptides, small molecules, and pericytes resulted in decreased tumor size and reduced metastasis. However, current tumor vascular normalizing drugs display moderate anticancer efficacy. Accumulated data have shown that a variety of vasculogenic/angiogenic tumor cells and genes play important roles in tumor neovascularization, growth, and metastasis. Therefore, multiple-targeting of vasculogenic tumor cells and genes may improve the efficacy of tumor vascular normalization. To this end, the combination of antiangiogenic drugs with tumor vascular normalizing therapeutics, as well as the integration of Western medicine with traditional Chinese medicine, may provide a good opportunity for discovering novel tumor vascular normalizing drugs for an effective anticancer therapy.

Keywords angiogenesis; vasculogenesis; neovascularization; tumor; vasculature; normalization; traditional Chinese medicine

Introduction

The link between the expansion, invasion, and metastasis of malignant tumors and the ability of the tumor to acquire an adequate vascular supply has now been well established [1]. Since Folkman [2] introduced a new concept of angiogenesis and proposed a novel antiangiogenic strategy for anticancer therapy in 1971, five antiangiogenic drugs have been used in a clinical setting to treat colon cancer, non-small cell lung cancer, and several other cancers. Avastin, one of the best antiangiogenic therapeutics, prolonged cancer patient life for 4 to 6 months [3–5]. While people were encouraged by this promising anti-antigenic therapy for a few cancers, recent

Received October 9, 2011; accepted November 16, 2011 Correspondence: quanshengzhou@yahoo.com clinical data have shown that these drugs exhibit transient and insufficient anticancer efficacy, resulting in tumor recurrence in patients after several months on the antiangiogenic drug [6,7]. One of the fundamental reasons for the moderate efficacy of antiangiogenic therapy is that current antiangiogenic drugs reduce blood and oxygen supplies, creating a hypoxic and acidic tumor microenvironment in tumor tissues that induces tumor cells to become more aggressive and metastatic, resulting in relapse in the cancer patients [8,9]. Hence, new strategies and approaches for antitumor neovascularization are highly desired.

Tumor vasculature is disorganized. In 2001, Jain [10] proposed a new concept of normalization of disorganized tumor vasculature, rather than blockage or disruption of tumor blood vessels for anticancer therapy, and the concept was re-enforced in 2005 [11]. Preclinical and clinical data have shown that normalization of the disorganized tumor

vasculature using monoclonal antibodies, proteins, peptides, small molecules, and pericytes reduced the tumor size and metastasis [12,13]. Tumor vascular normalization has recently emerged as an alternative approach to antitumor neovascularization and anticancer therapy [14].

In the current paper, recent progress in tumor vascular normalization will first be reviewed, the challenges in current tumor vascular normalization therapy will be discussed, and new strategies and approaches for novel tumor vascular normalizing drug discovery will be addressed. Finally, the perspectives on tumor vascular normalization for anticancer therapy will be discussed.

Morphological and molecular bases of tumor vascular normalization

When a tumor grows bigger than 2 mm, the cells in the middle of the tumor tissue live in a hypoxic microenvironment because oxygen diffusion in the tissue is only 1 mm in diameter. Hypoxia in the tumor profoundly changes the gene expression profile of tumor cells and initiates tumor angiogenesis and vasculogenesis [8,9]. As a consequence, the tumor tissue forms disorganized tumor vasculature with morphological and molecular characteristics that are distinct from normal blood vessels.

Morphological characteristics of tumor blood vessels

Tumor blood vessels are heterogeneous and exhibit unique morphological and functional characteristics compared with normal vasculature. Normal blood vessels are organized in a hierarchy of evenly distributed arteries, capillaries, and veins. The vessels are covered by pericytes to maintain vascular integrity. By contrast, tumor blood vessels consist of irregular branches with arteriovenous shunts, which are absent in pericyte coverage, and are highly permeable to plasma and plasma proteins (Fig. 1). Consequently, the tumor tissue usually suffers from local edema, bleeding, poor blood



Fig. 1 Comparison of normal vasculature with tumor vasculature. (A) Normal blood vessels are organized in a hierarchy of evenly distributed arteries, capillaries, and veins. The vessels are covered by pericytes to maintain the integrity of the vessels. (B) Tumor blood vessels are heterogeneous and consist of irregular branches with arteriovenous shunts.

circulation, and insufficient clearance of carbon dioxide and other metabolites. Indeed, tumor cells live in a hypoxic and acidic microenvironment, which induces the overexpression of genes responsible for tumor angiogenesis, vasculogenesis, growth, and metastasis [8,9]. In addition, tumor blood vessels lack functional lymphatic vessels inside the tumor, resulting in the elevation of interstitial fluid pressure in the tumor and the formation of a physiologic barrier to the delivery of therapeutic agents to the tumor [13]. These unique characteristics of the disorganized tumor vasculature provide the rationale for tumor vascular normalization [15].

Cell types and resources of tumor blood vessels

Tumor atavism has been recognized as a common phenomenon in tumor development for quite some time, and aggressive tumor cells usually exhibit the properties of embryonic stem/progenitor cells. Accumulated data have indicated that malignant tumors undergo embryonic vasculogenesis and angiogenesis to generate unique tumor vasculature in six distinct ways, namely, angiogenesis, vasculogenesis, intussusception, vessel co-option, vascular mimicry, and through cancer stem cell-differentiated tumor endothelial cells [16–19]. Increasing evidence has shown that aside from the well-known vascular endothelial growth factor (VEGF) and endothelial cell-mediated tumor angiogenesis, aggressive tumor cells and many other cells also play an important role in tumor neovascularization by undergoing vasculogenesis and angiogenesis. These vasculogenic cells include precancerous stem cells [20-22], cancer stem cells [23–29], bone marrow-derived endothelial progenitors (EPCs), blood monocytes, mast cells [30-35], mesenchymal stem cells (MSCs) [36,37], and tumor-associated macrophages (TAMs) [38,39]. In particular, a variety of vasculogenic tumor cells can form tumor blood vessels through either vasculogenic mimicry [40-48] or trans-differentiation into tumor endothelial cells [20-29]. Tumor endothelial cells exhibit tumor cell genetic, morphologic, and functional characteristics that are different from normal endothelial cells. Notably, recent studies revealed that more than 60% of the tumor blood vessels in glioma patients consisted of tumor endothelial cells trans-differentiated from glioma stem cells [25,26].

A total of 62 human tumor cell lines were recently examined using an *in vitro* tube formation system; 25 tumor cell lines from various tumors were found to form capillary tubes on Matrigel, with most of these tumor cells actively participating in tumor angiogenesis and vasculogenesis *in vivo* (unpublished observation). On the other hand, the cellular and molecular mechanisms of tumor cell-predominant neovascularization remain to be elucidated. Collectively, vasculogenic tumor cells play a critical role in tumor neovascularization. These emerging data provide new insight into the disorganized tumor vasculature, broadening the prospects on tumor vascular normalization.

Molecular characteristics of tumor blood vessels

Malignant tumors can form new blood vessels via a complex process involving neovascularization, which is now believed to result from two distinct mechanisms: (1) induced *angiogenesis*, by which neovascularization is achieved through tumor cell-produced angiogenic factor induced ingrowth of the tumor vasculature supply from preexisting blood vessels [2,18,19], and (2) *vasculogenesis*, by which the tumor cells and several types of bone marrow cells directly develop into the blood vessels required for neovascularization [40–48]. However, the molecular mechanisms underlying the formation of tumor blood vessels remain to be determined.

Initial tumor neovascularization is driven by both tumor cell intrinsic factors and hypoxia. Tumor cell intrinsic factors play basic roles in the initiation and development of tumor blood vessels, whereas hypoxia triggers tumor vascularity. Genomic instability and epigenetic abnormality may cause chromosome translocations, gene deletions, insertions, and mutations, resulting in the formation of various oncogenes or inactivation of tumor suppressive genes that are closely associated with tumor neovascularization [49,50]. Several oncogenes and tumor suppressors have been implicated in tumor neovascularization, such as Von Hippel/Linda (VHL) [51], Piwi-like protein 2 (Piwil2) [52], and fibronectin extra domain B (ED-B) [53]. In brief, mutant VHL diminishes the degradation of the hypoxia-inducible factor (HIF) and promotes HIF-mediated gene transcription, resulting in tumor vascularization [53]. A splicing error causes intron sequence retention in fibronectin mRNA to produce fibronectin ED-B in a variety of tumors, and fibronectin ED-B functions as an extracellular matrix protein and contributes to tumor vasculogenesis [53]. Our previous study [52] revealed that overexpression of a 60 kDa Piwil2 in tumor cells promoted tumor vasculogenesis and tumorigenesis [54]. In short, these data suggest that tumor intrinsic factors are critical to tumor neovascularization. Of note, current understanding of tumor intrinsic factors in tumor neovascularization are very shallow; hence, identification of additional intrinsic factors responsible for tumor angiogenesis and vasculogenesis will not only help in understanding the mechanisms of tumor vasculature generation, but will also provide new targets for novel tumor vascular normalizing drug discovery.

Hypoxia induces the overexpression of HIF-1, HIF-2, and many other vasculogenic and angiogenic genes and is an indication of tumor neovascularization. HIF-1 and HIF-2 consist of oxygen-dependent α subunits and constitutive β subunits and play important roles in the initiation and development of tumor vasculature [8,9]. Under normoxia, the active HIF heterodimer is not abundantly present in the tumor cells because its formation is tightly regulated by a master HIF regulatory protein, prolyl hydroxylase domaincontaining protein 2 (PHD2). PHD2 promotes hydroxylation and proteasomal degradation of HIF α subunits in an oxygen-dependent manner, leading to the inactivation of HIF activity. However, under hypoxial circumstances, PHD2 is in an inactivated state, and the HIF-1 α subunits are stabilized to form an active heterodimer with the HIF-1 β subunits. The active HIFs go to the cell nucleus, bind to the gene promoter region containing hypoxia-response elements, and activate the transcription of hundreds of hypoxia-response genes, including a variety of key vasculogenic and angiogenic genes such as VEGF, VEGF receptor-2, EphrinB2, Eph receptor B2 (EphB2), EphB4, vascular endothelial cadherin (VE-cadherin), cluster of differentiation 31 (CD31), semaphorin 4D (Sema4D) and its receptor plexin-B1, integrins, extra cellular matrix proteins (ECMs), and metal matrix proteases (MMPs) [8,9].

Several hypoxia- and HIF-induced genes play an important role in tumor neovascularization. For example, hypoxia switches tumor cells on to express embryonic vasculogenic genes such as EphrinB2 and its receptor EphB4, and both of the proteins locate at the cell membrane surface and contribute to tumor vasculogenesis [55,56]. Hypoxia also actuates the overexpression of MMPs, which degrade ECMs and promote the migration of tumor cells to the surrounding normal tissue and enhance tumor cell invasion and metastasis [57]. HIF-induced VE-cadherin and EphB2 in various tumor cells function as important adhesive molecules that connect tumor cells to one another to form capillary-like structures [58]. HIF induces the overexpression of Sema4D and its receptor plexin-B1 in tumors and promotes tumor neovascularization and enhances tumor invasive growth and metastasis [59].

Taken together, tumor cell intrinsic factors are critical to tumor neovascularization. A hypoxial and acidic environment triggers tumor vascularity by inducing tumor cells to express a variety of vasculogenic and angiogenic genes, which enable the tumor cells to form blood vessels. On the other hand, tumor cell-released growth factors, cytokines, and chemokines recruit bone marrow and blood cells into the tumor tissue to form a tumor microenvironment that fosters tumor neovascularization. Apparently, tumor cells play a pivotal role in tumor vascular initiation and development. Hitherto, targeting vasculogenic tumor cells and impeding the expression of vasculogenic and angiogenic genes in tumors has become a reasonable strategy for disorganized tumor vascular normalization.

Tumor vascular normalization for anticancer therapy

Malignant tumors require blood vessels for growth and metastasis. Based on the tumor angiogenesis theory of Folkman [2], a tumor produces its new blood vessel branches from preexisting blood vessels. Antitumor angiogenic therapy has mainly targeted VEGF and vascular endothelial cell-mediated angiogenesis during the past three decades, and the widely held view is that antiangiogenic therapy should inhibit or block tumor blood vessels, thereby depriving the supply of oxygen and nutrients in the tumors [2,18,19]. Traditional antiangiogenic strategies attempt to reduce the tumor vascular supply, but their success at an early stage is hampered by insufficient efficacy and the subsequent development of resistance [6,7]. Based on pioneering research studies [60-63], Jain [10] first proposed the normalization of tumor blood vessels as a new paradigm for anticancer therapy in 2001. The new concept of tumor vascular normalization encompasses an increase in tumor vascular cell tight connection and pericyte coverage, as well as vascular integrity to remedy the disorganized tumor vasculature and improve the blood circulation in tumor tissue. Emerging preclinical and initial clinical evidence has shown that normalization of disorganized tumor vasculature using therapeutics, rather than the blockage or disruption of tumor blood vessels, reduces tumor hypoxia, interstitial fluid pressure, and hyperpermeability and facilitates the delivery of exogenous therapeutics, the efficacy of radiotherapy, and the immunity of immune cells. Meanwhile, the coverage of tumor blood vessels by pericytes reduces the migration of metastatic tumor cells into circulation, curtailing tumor metastasis. Tumor vascular normalization has become a complementary therapeutic paradigm for cancer and other vascular disorders [10–15]. Several types of tumor vascular normalizing agents have shown anticancer potential, including anti-VEGF signaling agents, PHD2 agonists, VE-cadherin functional inhibitory proteins and antibodies, various small molecules such as traditional Chinese herbal drugs, and potential therapeutics to increase pericyte vascular coverage.

Current anti-VEGF signaling agents in tumor vascular normalization

Under the guidance of classic angiogenesis theory, several antiangiogenic drugs that mainly target VEGF signaling and endothelial cell-mediated angiogenesis have been used in clinical or preclinical assessments for the anticancer therapy, including the humanized VEGF monoclonal antibody bevacizumab (Avastin), the anti-VEGFR2 antibody IMC-1121b, the small molecule VEGFR2 inhibitor vatalanib, the VEGF receptor chimeric protein afliberecept, the VEGFR2 inhibitor peptide CT-322, and a VEGFR downstream protein kinase inhibitor Sutent [3-5]. VEGF is undoubtedly a critical growth factor for vascular endothelial cell-mediated angiogenesis, and anti-VEGF therapy has become promising in the treatment of advanced colon and non-small cell lung cancers. The angiostatic effects of current anti-VEGF signaling drugs have been attributed to the inhibition of endothelial cellmediated angiogenesis [64,65] because VEGF receptors have traditionally been thought to be limited to vascular endothelial cells. However, increasing evidence has shown that VEGFR2 is also expressed in perivascular cells, monocytes/ macrophages, dendritic cells, hematopoietic stem cells, and

bone marrow-derived circulating cells [9], suggesting that these cells may be affected by VEGF-VEGFR signaling inhibitors. The lack of a close association between the levels of VEGF in cancer patients and the antiangiogenic efficacy of the anti-VEGF signaling drugs has been a puzzle for researchers and clinical doctors for quite some time. VEGF can not be regarded as a biomarker of anti-VEGF and antiangiogenic therapies [66], suggesting that other mechanisms may be contributing to the angiostatic function of Avastin and other VEGF inhibitors.

The production and release of VEGF by tumor cells has been well established. On the one hand, VEGF induces endothelial cell migration to the tumor tissue and the formation of tumor blood vessels [1,2]. On the other hand, it reduces pericyte responsiveness to the related growth factor, platelet-derived growth factor (PDGF), resulting in small pericyte coverage in tumor vasculature [67]. Increasing evidence has indicated that anti-VEGF signaling drugs have dramatic effects on mural and perivascular cells in the tumors that increase pericyte coverage of the tumor blood vessels, resulting in tumor vascular normalization [68,69]. Thus, normalization of disorganized tumor vasculature through an increase in pericyte coverage is one of the important mechanisms of the anti-VEGF drug activity.

Although several current antiangiogenic drugs inhibit VEGF signaling and produce tumor vascular normalization effects, these drugs are notably still insufficient in curtailing tumor growth and metastasis [6–9], indicating that more potent tumor vascular normalizing agents are needed to achieve high efficacy of anticancer therapies.

PHD2 and its agonists in tumor vascular normalization

PHD enzymes belong to the iron(II)-2-oxoglutaratedependent dioxygenase families and have potential in regulating the HIF level and activity in cells. Among the PHD enzymes, PHD2 is the most important in controlling the levels of HIF-1 α and HIF activity under normoxial conditions [70]. Increasing data have shown that PHD2 functions as a tumor suppressor gene through the degradation of the HIF-1 α unit and the decline in HIF activity. Convincingly, mutation of the PHD2 gene or loss of PHD2 activity under hypoxial conditions affects tumor angiogenesis and tumor development [70–72]. Accordingly, PHD2 has been targeted for tumor vascular normalizing drug discovery.

Choi *et al.* [73] screened a molecular library consisting of more than 600 small molecules using PHD2 as the target and found a potent PHD2 activator called KRH102053 [2-amino-4-methylsulphanyl-butylic acid-4-methoxy-6-(4-methoxy-benzylamino)-2,2-dimethyl-chroman-3-yl ester], which acted as a PHD2 agonist to decrease the levels of HIF-1 α and HIF activity. As a consequence, the expression of HIF-regulated downstream vasculogenic target genes decreased. KRH102053 also inhibited human umbilical vein endothelial cell-mediated tube formation and HIF-related migration and

invasion of human osteosarcoma cells [73]. Moreover, another PHD2 activator, KRH102140, effectively inhibited HIF-1 α activity and decreased angiogenesis [74]. *In vivo* studies of these PHD2 activators in tumor vascular normalization, growth, and metastasis are needed to evaluate the antitumor efficacy and toxicity of these compounds.

By contrast, several studies have revealed that a haplodeficiency of PHD2 in mice resulted in tumor vascular normalization [71,72]. Carmeliet *et al.* [71] found the vascular normalizing effect of endothelial PHD2 haplodeficiency. In mice, the haplodeficiency of PHD2 promotes vessel maturation and normalization, resulting in improved tumor perfusion and oxygenation and the inhibition of tumor cell invasion, migration, and metastasis [71,72]. In view of the discrepancy between PHD2 agonists and PHD2 haplodeficiency in tumor vascular normalization, the neat effect of PHD2 on blood vessel normalization needs to be verified.

Targeting VE-cadherin for tumor vascular normalization

VE-cadherin is located at the surface of vascular endothelial cells and functions as a bridge that connects vascular endothelial cells [61]. Knockout of the VE-cadherin gene in mice resulted in early embryonic death of mice because of failure to generate primitive blood vessels, indicating that VE-cadherin plays a pivotal role during early embryonic vasculogenesis and development. In a normal human body after birth, VE-cadherin only expresses in vascular endothelial cells and not in other normal tissues and cells; it is a major player in the maintenance of vascular integrity and function [58,75-77]. VE-cadherin was also found in the osteosarcoma cell line MG63, which could form a tube-like capillary structure in vitro, convincingly silencing VE-cadherin expression in MG63 cells via SiRNA-hindered angiogenic cell function [78]. Increasing evidence has shown that the expression of VE-cadherin in tumor cells is implicated in tumor vasculogenesis and progression [79,80]. More recently, VE-cadherin was found overexpressed in the human ovarian cancer cell line Hey1B and 22 other tumor cell lines. Interestingly, most of these tumor cells gained the ability to form tube-like capillary structures in vitro in Matrigel (unpublished observation), suggesting that VE-cadherin plays a key role in tumor cell-mediated vasculogenesis. Hence, tumor cell VEcadherin may be a good target for novel antitumor vasculogenic drug discovery. The mechanisms of tumor cell-dominant vasculogenesis are still not clear; vasculogenic genes and biomarkers in these tumor cells are currently being actively identified to come up with new strategies and approaches to normalize disorganized tumor vasculature.

For that reason, several VE-cadherin-functional inhibitory proteins and monoclonal antibodies have been tested to suppress tumor angiogenesis. T2-tryptophany-tRNA synthetase (T2-TrpRS) effectively inhibited retinal angiogenesis in a mouse model, but it did not affect or disrupt normal vasculature. Interestingly, T2-TrpRS promoted the regeneration of normal blood vessels in retinopathy but thwarted pathological retinol vasculature [81,82], suggesting that T2-TrpRS especially normalizes angiogenic blood vessels.

T2-TrpRS has recently been found to block tumor VEcadherin, and particularly suppress tumor endothelial cellmediated angiogenesis, although it did not affect normal endothelial cell-mediated angiogenesis [83]. More recently, our mechanism study revealed that T2-TrpRS bound to the two critical amino acids Trp2 and Trp4 in VE-cadherin extra cellular domain 1 and prohibited VE-cadherin to form dimers on the tumor cell surface. As a result, tumor endothelial cells with unengaged VE-cadherin failed to connect to each other to form capillary tubes [84], suggesting that T2-TrpRS can be a specific tumor vascular normalizing agent.

In addition, monoclonal antibodies against VE-cadherin have also exhibited specific antitumor effects. Actinium-225, an alpha-emitting antibody construct (E4G10), specifically bound to the unengaged form of VE-cadherin and showed potent and selective killing of tumor neovascular endothelium and EPCs in the bone marrow; however, no specific normaltissue uptake of E4G10 was seen via imaging or post-mortem biodistribution studies in mice [85].

Taken together, the unengaged form of VE-cadherin in tumor cells, as well as the tumor neovascular endothelium and bone marrow-derived EPCs can be targeted for special tumor vascular normalizing drug discovery.

Targeting tumor-associated macrophages for tumor vascular normalization

During tumor development, several types of bone marrowderived cells, including EPCs, TAMs, and MSCs, are recruited from the blood to the tumor tissues [38]. Increasing evidence has shown that TAMs contribute to tumor angiogenesis and vasculogenesis, and TAMs with proangiogenic and immune-suppressive (M2-like) phenotypes are hallmarks of malignancy [38,39]. Rolny et al. [86] reported that histidine-rich glycoprotein (HRG) inhibited tumor growth and metastasis through the enhancement of vessel normalization and antitumor immune responses. Mechanism studies indicated that HRG suppressed the expression of placental growth factor and induced the polarization of tumorassociated macrophages, resulting in tumor vascular normalization and enhancement of immunity [86]. This finding suggests that the modulation of TAMs may be another way to curb tumor neovascularization.

Traditional Chinese herbal drugs in tumor vascular normalization

Historically, traditional Chinese medicine has been widely used in China for thousands of years and has shown promising results in the prevention and treatment of cancers. For example, the Realgar-*Indigo naturalis* formula has been proven very effective in the treatment of human acute promyelocytic leukemia [87]. One type of traditional Chinese medicinal drug named the "activating blood circulation to dissipate blood stasis" (活血化瘀) has been used in China to prevent and treat both cardiac vascular diseases and tumors for many years [88-98]. Accumulated data have shown that "activating blood circulation to dissipate blood stasis" drugs can increase blood microcirculation, facilitate oxygen and nutrition supply, and promote the removal of waste materials from tissues, resulting in an improvement of tissue blood circulation and vascular function [88]. So far, various "activating blood circulation to dissipate blood stasis" herbal extracts and purified active compounds, including celastrol [89], morelloflavone [90], gambogic acid [91], acetyl-11keto-β-boswellic acid [92,93], 1'-acetoxychavicol acetate [94], cudratricusxanthone G [95], Salvia miltiorrhiza Bunge [96], Erigeron breviscapus Hand-Mazz [97], and Hedvotis diffusea Willd [98], have been used to improve tissue blood circulation and treat malignant tumors. In particular, the combination of the "activating blood circulation to dissipate blood stasis" drugs with current anticancer chemotherapeutics exerted a better effect as a collective anti-tumor treatment [99].

Manipulation of the pericytes in tumor tissues for vascular normalization

Tumor vasculature is characterized by dilated and tortuous vessels, thickened basement membranes, small pericyte coverage, immaturity, and high permeability. The mechanisms of the generation of the disorganized tumor vasculature are under investigation. One possibility is that tumor blood vessels can be lined by tumor cells through vasculogenesis. Although tumor cells gain vasculogenic capability, the cells clearly have not acquired the entire function of normal vasculature, as previously mentioned [16–30,41–48,100]. Another possibility is that tumor tissues have high VEGF levels, which impede pericyte vascular coverage.

The inhibition of VEGF and its receptor can have dramatic and unexpected effects on mural and perivascular cell functions in the tumor tissue, resulting in vascular smooth muscle cell and pericyte activation as well as blood vessel normalization/maturation [63-66,101,102]. Angiopoietin-1 (Ang1) has been shown to act as an angiogenic promoter in embryonic angiogenesis by promoting vascular branching, pericyte recruitment, and endothelial survival. Increasing experimental data suggest that Ang1-stimulated association of mural cells with endothelial cells leads to the stabilization of newly formed blood vessels. Ang1 can be used, alone or in combination with VEGF, to promote therapeutic angiogenesis [101,102]. In addition, inhibition of angiogenesis with an antibody DC101 that blocks VEGFR2 significantly promotes pericyte recruitment and increases the number of pericytecovered vessels [71,72,103]. Increasing data have shown that

normalization of tumor blood vessels can lead to an improvement in the metabolic profile of the tumor microenvironment, as well as in the delivery and efficacy of exogenously administered therapeutics, efficacy of radiotherapy and of effective immune cells, and a reduction in the number of metastatic cells shed by tumors into circulation [10–14,71,72,101–104]. Therefore, manipulation of pericytes in tumor tissues has opened another option to normalize the disorganized tumor vasculature.

Challenges and perspectives

Since the new concept of the normalization of disorganized tumor vasculature for anticancer therapy was raised by Jain in 2001, sound progress has been made in preclinical and initial clinical trials. Normalization of disorganized tumor blood vessels results in an increase in tumor tissue blood and oxygen supply, improvement of the distribution and efficacy of other anticancer drugs, and reduction in tumor growth and metastasis [10–14]. However, moderate antitumor effects of current tumor vascular normalizing drugs have shadowed the prospect of vascular normalization strategies in anticancer therapy. In the subsequent section, with the development of new strategies and approaches to uncover potent tumor vascular normalizing agents and correct the disorganized tumor vasculature for an effective anticancer therapy will be discussed.

Targeting vasculogenic cells in tumors for tumor vascular normalization

Current antitumor angiogenesis and vascular normalization therapies have mainly focused on anti-VEGF and endothelial cell-predominant angiogenesis, overlooking tumor vasculogenesis mediated by tumor cells and BM-derived cells. As previously mentioned, aside from endothelial cells, various other cells, including tumor cells, EPCs, MSCs, TAMs, and pericytes, play an important role in tumor neovascularization through vasculogenesis [1,18,19]. In particular, a variety of tumor cells, including pre-cancer stem cells, cancer stem cells, tumor endothelial cells, and various other aggressive tumor cells, participate in the formation of tumor blood vessels through at least three ways [18]: (1) blood vessel co-option between tumor cells and vascular endothelial cells, (2) tumor cell-predominant vascular mimicry (independent of vascular endothelial cells), and (3) cancer stem cell transdifferentiation into tumor endothelial cells (Fig. 2). Tumor cell-consisting blood vessels have unique morphological characteristics that differ from normal vasculature, as previously stated [8,9,13,15], and account for poor blood circulation in the tumor tissues. Accumulated data have shown that vasculogenic tumor cells play critical roles in tumor neovascularization, growth, and metastasis [16-30,41-48,100]. Hence, vasculogenic tumor cells should be targeted for the normalization of disorganized tumor vasculature.



Fig. 2 Normalization of disorganized tumor blood vessels for anticancer therapy. Malignant tumors may make their own blood vessels via a complex process of neovascularization through (1) VEGF and endothelial cell-mediated angiogenesis, (2) vessel co-option between endothelial cells and tumor cells, and (3) tumor cell-predominant vasculogenesis. The disorganized tumor blood vessels could be normalized using several options, including (1) anti-VEGF therapeutics, (2) PHD2 agonists, (3) VE-cadherin blocking agents, (4) targeting tumor-associated macrophages, (5) traditional Chinese medicinal herbal drugs, and (6) manipulation of pericytes.

Vasculogenic and angiogenic tumor cells must be targeted for effective normalization of the disorganized tumor vasculature because of two reasons. First, tumor cells can directly generate blood vessels via vasculogenesis. Second, tumor cells secrete a variety of angiogenic factors that initiate and promote tumor angiogenesis. Incredibly, most of the aggressive tumor cells have vasculogenic and/or angiogenic capabilities; therefore, targeting vasculogenic and angiogenic tumor cells may be a "one stone hits two birds" approach to impeding tumor neovascularization and hindering tumor cellmediated tumorigenesis. In addition, targeting vasculogenic tumor cells and genes may provide a new opportunity for novel antitumor vasculogenic drug discovery. Moreover, the combination of antitumor vasculogenic agents with antiangiogenic drugs may exhibit synergistic effects in anticancer therapy.

Identification of biomarkers for tumor vascular normalization

VEGF, which is an important angiogenic factor, is overexpressed in a variety of tumors and has been targeted for antiangiogenic therapy. However, clinical investigation has indicated that VEGF levels in cancer patients are generally poorly associated with the outcome of antiangiogenic therapy, and that VEGF is not a good biomarker for antiangiogenesis and anticancer therapies [69]. Although five antiangiogeneic drugs have been utilized for anticancer therapy in clinical settings, no validated biomarkers for monitoring and evaluating antitumor angiogenic therapy has been found at present. Ironically, hypertension, a toxic side effect of antiangiogenic therapy, has been reported to be closely associated with a better outcome of antiangiogenic therapy [6,7,69]. Therefore, identifying tumor angiogenic and vasculogenic biomarkers beyond VEGF signaling is highly desirable.

Sorensen *et al.* [105] have recently reported a "vascular normalization index," which measured the changes in the Ktrans, microvessel volume, and circulating collagen IV in recurrent glioblastoma patients, as a biomarker for tumor vascular normalization. They showed that the vascular normalization index was closely correlated to the duration of overall survival and/or to the progression-free survival (P < 0.05) of the patients. Determining whether the vascular

Tumor vascular normalization for anticancer therapy

normalization index is a universal biomarker for tumor vascular normalization in other types of tumor would be worthwhile.

The interstitial fluid pressure is usually more than 10-fold higher in a variety of tumor tissues compared with those of normal tissues. Tumor vasculature is highly permeable, and intravascular fluids and plasma proteins are accumulated in the tumor tissues [13]. Hence, interstitial fluid pressure and intravascular plasma proteins in the tumor may be considered as biomarker candidates for tumor vascular normalization. Accordingly, appropriate techniques will be required to measure these biomarkers. In this context, new, noninvasive imaging technologies, including noninvasive variable-magnification in vivo-fluorescence imaging and fluorescence tomography [106], magnetic resonance imaging [107], and acoustic radiation force-induced optical spectroscopy, would be very useful. Further clinical validation of these biomarkers and new technologies is needed to evaluate the anticancer efficacy of tumor vascular normalization drugs and improve anticancer therapy.

Multiple-targeting of tumor blood vessels for tumor vascular normalization

Tumor tissues consist of cancer cells and non-malignant cells, which include endothelial cells, pericytes, fibroblasts, macrophages, lymphocytes, dendritic cells, mast cells, and many other types of cells. Tumor cells orchestrate with other cells to form a disorganized tumor vasculature and a hostile tumor microenvironment that promotes tumor growth, invasion, metastasis, and resistance to various therapies. In addition, various vasculogenic and angiogenic genes are overexpressed in tumor vascular cells and contribute to tumor neovascularization. Thus, the multiple-targeting of vasculogenic tumor cells, bone marrow-derived cells, and other vasculogenic cells in a tumor microenvironment, as well as the retardation of vasculogenic and angiogenic gene overexpression, may improve the efficacy of tumor vascular normalization.

In addition, the combination of tumor vascular normalization agents with current antiangiogenic and cytotoxic drugs may achieve a synergistic effect during tumor vascular normalization. Furthermore, many traditional Chinese herbal drugs display antiangiogenic effects and enhance tumor vascular normalization. Therefore, the integration of Western medicine with traditional Chinese medicine may provide a good opportunity for achieving a high efficacy of tumor vascular normalization.

Although tumor vascular normalization has only been studied for 10 years, the sound progress in tumor vascular normalization indicates a bright prospect for this approach and warrants further investigation. Success in the identification of specific genes, cells, and biomarkers in disorganized tumor vasculature is predicted, and multiple-targeting of key vasculogenic cells and genes in tumor blood vessels, as well as the integration of Western medicine with traditional Chinese medicine, will bring a breakthrough in tumor vascular normalization for an effective anticancer therapy.

Acknowledgements

This study was supported by grants from the National Natural Science Foundation of China (Grant No. 30971138), the Science Foundation of Suzhou City (No. SWG0904 and No. SS201004), a project funded by the Priority Academic Program Development of Jiangsu Higher Education Institutions (PAPD), and a Special National Strategic Leader Project of China (No. XDA01040200).

References

- Ebos JM, Kerbel RS. Antiangiogenic therapy: impact on invasion, disease progression, and metastasis. Nat Rev Clin Oncol 2011; 8 (4): 210–221
- Folkman J. Tumor angiogenesis: therapeutic implications. N Engl J Med 1971; 285(21): 1182–1186
- Heath VL, Bicknell R. Anticancer strategies involving the vasculature. Nat Rev Clin Oncol 2009; 6(7): 395–404
- 4. Ribatti D. Endogenous inhibitors of angiogenesis: a historical review. Leuk Res 2009; 33(5): 638–644
- Ribatti D. The discovery of antiangiogenic molecules: a historical review. Curr Pharm Des 2009; 15(4): 345–352
- Van Cutsem E, Lambrechts D, Prenen H, Jain RK, Carmeliet P. Lessons from the adjuvant bevacizumab trial on colon cancer: what next? J Clin Oncol 2011; 29(1): 1–4
- Miles D, Harbeck N, Escudier B, Hurwitz H, Saltz L, Van Cutsem E, Cassidy J, Mueller B, Sirzén F. Disease course patterns after discontinuation of bevacizumab: pooled analysis of randomized phase III trials. J Clin Oncol 2011; 29(1): 83–88
- Otrock ZK, Hatoum HA, Awada AH, Ishak RS, Shamseddine AI. Hypoxia-inducible factor in cancer angiogenesis: structure, regulation and clinical perspectives. Crit Rev Oncol Hematol 2009; 70(2): 93–102
- Osinsky S, Zavelevich M, Vaupel P. Tumor hypoxia and malignant progression. Exp Oncol 2009; 31(2): 80–86
- Jain RK. Normalizing tumor vasculature with anti-angiogenic therapy: a new paradigm for combination therapy. Nat Med 2001; 7 (9): 987–989
- Jain RK. Normalization of tumor vasculature: an emerging concept in antiangiogenic therapy. Science 2005; 307(5706): 58–62
- Carmeliet P, Jain RK. Principles and mechanisms of vessel normalization for cancer and other angiogenic diseases. Nat Rev Drug Discov 2011; 10(6): 417–427
- Goel S, Duda DG, Xu L, Munn LL, Boucher Y, Fukumura D, Jain RK. Normalization of the vasculature for treatment of cancer and other diseases. Physiol Rev 2011; 91(3): 1071–1121
- Sato Y. Persistent vascular normalization as an alternative goal of anti-angiogenic cancer therapy. Cancer Sci 2011; 102(7): 1253– 1256
- Fukumura D, Jain RK. Tumor microvasculature and microenvironment: targets for anti-angiogenesis and normalization. Microvasc Res 2007; 74(2–3): 72–84
- 16. Hess AR, Margaryan NV, Seftor EA, Hendrix MJ. Deciphering the

signaling events that promote melanoma tumor cell vasculogenic mimicry and their link to embryonic vasculogenesis: role of the Eph receptors. Dev Dyn 2007; 236(12): 3283–3296

- Kučera T, Lammert E. Ancestral vascular tube formation and its adoption by tumors. Biol Chem 2009; 390(10): 985–994
- Carmeliet P, Jain RK. Molecular mechanisms and clinical applications of angiogenesis. Nature 2011; 473(7347): 298–307
- Potente M, Gerhardt H, Carmeliet P. Basic and therapeutic aspects of angiogenesis. Cell 2011; 146(6): 873–887
- Shen R, Ye Y, Chen L, Yan Q, Barsky SH, Gao JX. Precancerous stem cells can serve as tumor vasculogenic progenitors. PLoS ONE 2008; 3(2): e1652
- Menakuru SR, Brown NJ, Staton CA, Reed MW. Angiogenesis in pre-malignant conditions. Br J Cancer 2008; 99(12): 1961– 1966
- Hong D, Gupta R, Ancliff P, Atzberger A, Brown J, Soneji S, Green J, Colman S, Piacibello W, Buckle V, Tsuzuki S, Greaves M, Enver T. Initiating and cancer-propagating cells in TEL-AML1-associated childhood leukemia. Science 2008; 319(5861): 336–339
- Hill RP, Marie-Egyptienne DT, Hedley DW. Cancer stem cells, hypoxia and metastasis. Semin Radiat Oncol 2009; 19(2): 106–111
- Zhao Y, Dong J, Huang Q, Lou M, Wang A, Lan Q. Endothelial cell transdifferentiation of human glioma stem progenitor cells *in vitro*. Brain Res Bull 2010; 82(5–6): 308–312
- Wang R, Chadalavada K, Wilshire J, Kowalik U, Hovinga KE, Geber A, Fligelman B, Leversha M, Brennan C, Tabar V. Glioblastoma stem-like cells give rise to tumour endothelium. Nature 2010; 468(7325): 829–833
- Ricci-Vitiani L, Pallini R, Biffoni M, Todaro M, Invernici G, Cenci T, Maira G, Parati EA, Stassi G, Larocca LM, De Maria R. Tumour vascularization via endothelial differentiation of glioblastoma stem-like cells. Nature 2010; 468(7325): 824–828
- Soda Y, Marumoto T, Friedmann-Morvinski D, Soda M, Liu F, Michiue H, Pastorino S, Yang M, Hoffman RM, Kesari S, Verma IM. Transdifferentiation of glioblastoma cells into vascular endothelial cells. Proc Natl Acad Sci USA 2011; 108(11): 4274– 4280
- Chiao MT, Yang YC, Cheng WY, Shen CC, Ko JL. CD133⁺ glioblastoma stem-like cells induce vascular mimicry *in vivo*. Curr Neurovasc Res 2011; 8(3): 210–219
- Ping YF, Bian XW. Consice review: contribution of cancer stem cells to neovascularization. Stem Cells 2011; 29(6): 888–894
- Ahn GO, Brown JM. Role of endothelial progenitors and other bone marrow-derived cells in the development of the tumor vasculature. Angiogenesis 2009; 12(2): 159–164
- Ria R, Piccoli C, Cirulli T, Falzetti F, Mangialardi G, Guidolin D, Tabilio A, Di Renzo N, Guarini A, Ribatti D, Dammacco F, Vacca A. Endothelial differentiation of hematopoietic stem and progenitor cells from patients with multiple myeloma. Clin Cancer Res 2008; 14(6): 1678–1685
- Vacca A, Ribatti D. Bone marrow angiogenesis in multiple myeloma. Leukemia 2006; 20(2): 193–199
- 33. Chen H, Campbell RA, Chang Y, Li M, Wang CS, Li J, Sanchez E, Share M, Steinberg J, Berenson A, Shalitin D, Zeng Z, Gui D, Perez-Pinera P, Berenson RJ, Said J, Bonavida B, Deuel TF, Berenson JR. Pleiotrophin produced by multiple myeloma induces

transdifferentiation of monocytes into vascular endothelial cells: a novel mechanism of tumor-induced vasculogenesis. Blood 2009; 113(9): 1992–2002

- 34. Scavelli C, Nico B, Cirulli T, Ria R, Di Pietro G, Mangieri D, Bacigalupo A, Mangialardi G, Coluccia AM, Caravita T, Molica S, Ribatti D, Dammacco F, Vacca A. Vasculogenic mimicry by bone marrow macrophages in patients with multiple myeloma. Oncogene 2008; 27(5): 663–674
- Maltby S, Khazaie K, McNagny KM. Mast cells in tumor growth: angiogenesis, tissue remodelling and immune-modulation. Biochim Biophys Acta 2009; 1796(1): 19–26
- Ball SG, Shuttleworth CA, Kielty CM. Mesenchymal stem cells and neovascularization: role of platelet-derived growth factor receptors. J Cell Mol Med 2007; 11(5): 1012–1030
- Chen MY, Lie PC, Li ZL, Wei X. Endothelial differentiation of Wharton's jelly-derived mesenchymal stem cells in comparison with bone marrow-derived mesenchymal stem cells. Exp Hematol 2009; 37(5): 629–640
- Siveen KS, Kuttan G. Role of macrophages in tumour progression. Immunol Lett 2009; 123(2): 97–102
- Coffelt SB, Hughes R, Lewis CE. Tumor-associated macrophages: effectors of angiogenesis and tumor progression. Biochim Biophys Acta 2009; 1796(1): 11–18
- Maniotis AJ, Folberg R, Hess A, Seftor EA, Gardner LM, Pe'er J, Trent JM, Meltzer PS, Hendrix MJ. Vascular channel formation by human melanoma cells *in vivo* and *in vitro*: vasculogenic mimicry. Am J Pathol 1999; 155(3): 739–752
- Folberg R, Hendrix MJ, Maniotis AJ. Vasculogenic mimicry and tumor angiogenesis. Am J Pathol 2000; 156(2): 361–381
- 42. Seftor RE, Seftor EA, Koshikawa N, Meltzer PS, Gardner LM, Bilban M, Stetler-Stevenson WG, Quaranta V, Hendrix MJ. Cooperative interactions of laminin 5 gamma2 chain, matrix metalloproteinase-2, and membrane type-1-matrix/metalloproteinase are required for mimicry of embryonic vasculogenesis by aggressive melanoma. Cancer Res 2001; 61(17): 6322–6327
- 43. Sood AK, Fletcher MS, Zahn CM, Gruman LM, Coffin JE, Seftor EA, Hendrix MJ. The clinical significance of tumor cell-lined vasculature in ovarian carcinoma: implications for anti-vasculogenic therapy. Cancer Biol Ther 2002; 1(6): 661–664
- Hendrix MJ, Seftor EA, Hess AR, Seftor RE. Vasculogenic mimicry and tumour-cell plasticity: lessons from melanoma. Nat Rev Cancer 2003; 3(6): 411–421
- Folberg R, Maniotis AJ. Vasculogenic mimicry. APMIS 2004; 112 (7–8): 508–525
- Zhang S, Zhang D, Sun B. Vasculogenic mimicry: current status and future prospects. Cancer Lett 2007; 254(2): 157–164
- Rak J, Milsom C, Yu J. Vascular determinants of cancer stem cell dormancy—do age and coagulation system play a role? APMIS 2008; 116(7–8): 660–676
- Begg AC, Stewart FA, Vens C. Strategies to improve radiotherapy with targeted drugs. Nat Rev Cancer 2011;11(4):239–253
- Chiarugi V, Magnelli L, Cinelli M, Ruggiero M. Oncogenes, p53, and tumor angiogenesis. J Cancer Res Clin Oncol 1998; 124(9): 523–525
- Giri D, Ittmann M. Inactivation of the PTEN tumor suppressor gene is associated with increased angiogenesis in clinically localized prostate carcinoma. Hum Pathol 1999; 30(4): 419–424

- Bohonowych JE,Gopal U, Isaacs JS. Hsp90 as a gatekeeper of tumor angiogenesis: clinical promise and potential pitfalls. J Oncol 2010; 2010: 412985
- Gao JX. Cancer stem cells: the lessons from pre-cancerous stem cells. J Cell Mol Med 2008; 12(1): 67–96
- Midulla M, Verma R, Pignatelli M, Ritter MA, Courtenay-Luck NS, George AJ. Source of oncofetal ED-B-containing fibronectin: implications of production by both tumor and endothelial cells. Cancer Res 2000; 60(1): 164–169
- 54. Ye Y, Yin DT, Chen L, Zhou Q, Shen R, He G, Yan Q, Tong Z, Issekutz AC, Shapiro CL, Barsky SH, Lin H, Li JJ, Gao JX. Identification of Piwil2-like (PL2L) proteins that promote tumorigenesis. PLoS ONE 2010; 5(10): e13406
- 55. Oike Y, Ito Y, Hamada K, Zhang XQ, Miyata K, Arai F, Inada T, Araki K, Nakagata N, Takeya M, Kisanuki YY, Yanagisawa M, Gale NW, Suda T. Regulation of vasculogenesis and angiogenesis by EphB/ephrin-B2 signaling between endothelial cells and surrounding mesenchymal cells. Blood 2002; 100(4): 1326–1333
- 56. Djokovic D, Trindade A, Gigante J, Badenes M, Silva L, Liu R, Li X, Gong M, Krasnoperov V, Gill PS, Duarte A. Combination of Dll4/Notch and Ephrin-B2/EphB4 targeted therapy is highly effective in disrupting tumor angiogenesis. BMC Cancer 2010; 10(1): 641–652
- McColgan P, Sharma P. Polymorphisms of matrix metalloproteinases 1, 2, 3 and 9 and susceptibility to lung, breast and colorectal cancer in over 30,000 subjects. Int J Cancer 2009; 125(6): 1473–1478
- Taveau JC, Dubois M, Le Bihan O, Trépout S, Almagro S, Hewat E, Durmort C, Heyraud S, Gulino-Debrac D, Lambert O. Structure of artificial and natural VE-cadherin-based adherens junctions. Biochem Soc Trans 2008; 36(2): 189–193
- Sun Q, Zhou H, Binmadi NO, Basile JR. Hypoxia-inducible factor-1-mediated regulation of semaphorin 4D affects tumor growth and vascularity. J Biol Chem 2009; 284(46): 32066–32074
- Suri C, Jones PF, Patan S, Bartunkova S, Maisonpierre PC, Davis S, Sato TN, Yancopoulos GD. Requisite role of angiopoietin-1, a ligand for the TIE2 receptor, during embryonic angiogenesis. Cell 1996; 87(7): 1171–1180
- Suri C, McClain J, Thurston G, McDonald DM, Zhou H, Oldmixon EH, Sato TN, Yancopoulos GD. Increased vascularization in mice overexpressing angiopoietin-1. Science 1998; 282 (5388): 468–471
- Thurston G, Suri C, Smith K, McClain J, Sato TN, Yancopoulos GD, McDonald DM. Leakage-resistant blood vessels in mice transgenically overexpressing angiopoietin-1. Science 1999; 286 (5449): 2511–2514
- Hayes AJ, Huang WQ, Yu J, Maisonpierre PC, Liu A, Kern FG, Lippman ME, McLeskey SW, Li LY. Expression and function of angiopoietin-1 in breast cancer. Br J Cancer 2000; 83(9): 1154– 1160
- 64. Willett CG, Boucher Y, di Tomaso E, Duda DG, Munn LL, Tong RT, Chung DC, Sahani DV, Kalva SP, Kozin SV, Mino M, Cohen KS, Scadden DT, Hartford AC, Fischman AJ, Clark JW, Ryan DP, Zhu AX, Blaszkowsky LS, Chen HX, Shellito PC, Lauwers GY, Jain RK. Direct evidence that the VEGF-specific antibody bevacizumab has antivascular effects in human rectal cancer. Nat Med 2004; 10(2): 145–147

- Ferrara N, Hillan KJ, Novotny W. Bevacizumab (Avastin), a humanized anti-VEGF monoclonal antibody for cancer therapy. Biochem Biophys Res Commun 2005; 333(2): 328–335
- Jain RK, Duda DG, Willett CG, Sahani DV, Zhu AX, Loeffler JS, Batchelor TT, Sorensen AG. Biomarkers of response and resistance to antiangiogenic therapy. Nat Rev Clin Oncol 2009; 6 (6): 327–338
- Greenberg JI, Cheresh DA. VEGF as an inhibitor of tumor vessel maturation: implications for cancer therapy. Expert Opin Biol Ther 2009; 9(11): 1347–1356
- Tong RT, Boucher Y, Kozin SV, Winkler F, Hicklin DJ, Jain RK. Vascular normalization by vascular endothelial growth factor receptor 2 blockade induces a pressure gradient across the vasculature and improves drug penetration in tumors. Cancer Res 2004; 64(11): 3731–3736
- 69. Winkler F, Kozin SV, Tong RT, Chae SS, Booth MF, Garkavtsev I, Xu L, Hicklin DJ, Fukumura D, di Tomaso E, Munn LL, Jain RK. Kinetics of vascular normalization by VEGFR2 blockade governs brain tumor response to radiation: role of oxygenation, angiopoietin-1, and matrix metalloproteinases. Cancer Cell 2004; 6(6): 553– 563
- Ladroue C, Carcenac R, Leporrier M, Gad S, Le Hello C, Galateau-Salle F, Feunteun J, Pouysségur J, Richard S, Gardie B. PHD2 mutation and congenital erythrocytosis with paraganglioma. N Engl J Med 2008; 359(25): 2685–2692
- 71. Mazzone M, Dettori D, Leite de Oliveira R, Loges S, Schmidt T, Jonckx B, Tian YM, Lanahan AA, Pollard P, Ruiz de Almodovar C, De Smet F, Vinckier S, Aragonés J, Debackere K, Luttun A, Wyns S, Jordan B, Pisacane A, Gallez B, Lampugnani MG, Dejana E, Simons M, Ratcliffe P, Maxwell P, Carmeliet P. Heterozygous deficiency of PHD2 restores tumor oxygenation and inhibits metastasis via endothelial normalization. Cell 2009; 136(5): 839– 851
- Kim JW, Johnson RS. You don't need a PHD to grow a tumor. Dev Cell 2009; 16(6): 781–782
- Choi HJ, Song BJ, Gong YD, Gwak WJ, Soh Y. Rapid degradation of hypoxia-inducible factor-1alpha by KRH102053, a new activator of prolyl hydroxylase 2. Br J Pharmacol 2008; 154(1): 114–125
- 74. Nepal M, Gong YD, Park YR, Soh Y. An activator of PHD2, KRH102140, decreases angiogenesis via inhibition of HIF-1α. Cell Biochem Funct 2011; 29(2): 126–134
- Vestweber D. VE-cadherin: the major endothelial adhesion molecule controlling cellular junctions and blood vessel formation. Arterioscler Thromb Vasc Biol 2008; 28(2): 223–232
- Gavard J. Breaking the VE-cadherin bonds. FEBS Lett 2009; 583 (1): 1–6
- Dejana E, Orsenigo F, Lampugnani MG. The role of adherens junctions and VE-cadherin in the control of vascular permeability. J Cell Sci 2008; 121(13): 2115–2122
- Zhang LZ, Mei J, Qian ZK, Cai XS, Jiang Y, Huang WD. The role of VE-cadherin in osteosarcoma cells. Pathol Oncol Res 2010; 16 (1): 111–117
- Cavallaro U, Liebner S, Dejana E. Endothelial cadherins and tumor angiogenesis. Exp Cell Res 2006; 312(5): 659–667
- 80. Labelle M, Schnittler HJ, Aust DE, Friedrich K, Baretton G, Vestweber D, Breier G. Vascular endothelial cadherin promotes

breast cancer progression via transforming growth factor beta signaling. Cancer Res 2008; 68(5): 1388–1397

- Otani A, Slike BM, Dorrell MI, Hood J, Kinder K, Ewalt KL, Cheresh D, Schimmel P, Friedlander M. A fragment of human TrpRS as a potent antagonist of ocular angiogenesis. Proc Natl Acad Sci USA 2002; 99(1): 178–183
- 82. Banin E, Dorrell MI, Aguilar E, Ritter MR, Aderman CM, Smith AC, Friedlander J, Friedlander M. T2-TrpRS inhibits preretinal neovascularization and enhances physiological vascular regrowth in OIR as assessed by a new method of quantification. Invest Ophthalmol Vis Sci 2006; 47(5): 2125–2134
- Zhou Q, Kiosses WB, Liu J, Schimmel P. Tumor endothelial cell tube formation model for determining anti-angiogenic activity of a tRNA synthetase cytokine. Methods 2008; 44(2): 190–195
- 84. Zhou Q, Kapoor M, Guo M, Belani R, Xu X, Kiosses WB, Hanan M, Park C, Armour E, Do MH, Nangle LA, Schimmel P, Yang XL. Orthogonal use of a human tRNA synthetase active site to achieve multifunctionality. Nat Struct Mol Biol 2010; 17(1): 57–61
- 85. Jaggi JS, Henke E, Seshan SV, Kappel BJ, Chattopadhyay D, May C, McDevitt MR, Nolan D, Mittal V, Benezra R, Scheinberg DA. Selective alpha-particle mediated depletion of tumor vasculature with vascular normalization. PLoS ONE 2007; 2(3): e267
- 86. Rolny C, Mazzone M, Tugues S, Laoui D, Johansson I, Coulon C, Squadrito ML, Segura I, Li X, Knevels E, Costa S, Vinckier S, Dresselaer T, Åkerud P, De Mol M, Salomäki H, Phillipson M, Wyns S, Larsson E, Buysschaert I, Botling J, Himmelreich U, Van Ginderachter JA, De Palma M, Dewerchin M, Claesson-Welsh L, Carmeliet P. HRG inhibits tumor growth and metastasis by inducing macrophage polarization and vessel normalization through downregulation of PIGF. Cancer Cell 2011; 19(1): 31–44
- Wang L, Zhou GB, Liu P, Song JH, Liang Y, Yan XJ, Xu F, Wang BS, Mao JH, Shen ZX, Chen SJ, Chen Z. Dissection of mechanisms of Chinese medicinal formula Realgar-Indigo naturalis as an effective treatment for promyelocytic leukemia. Proc Natl Acad Sci USA 2008; 105(12): 4826–4831
- Xiong L, Tian SX. A concept of regulating tumor microenvironment immune and normalizing angiogenesis by Chinese medicine drug therapy for supporting zheng-qi to prop up root. Chin J Integr Traidt West Med (Zhongguo Zhong Xi Yi Jie He Za Zhi) 2010; 30 (2): 201–204 (in Chinese)
- Pang X, Yi Z, Zhang J, Lu B, Sung B, Qu W, Aggarwal BB, Liu M. Celastrol suppresses angiogenesis-mediated tumor growth through inhibition of AKT/mammalian target of rapamycin pathway. Cancer Res 2010; 70(5): 1951–1959
- 90. Pang X, Yi T, Yi Z, Cho SG, Qu W, Pinkaew D, Fujise K, Liu M. Morelloflavone, a biflavonoid, inhibits tumor angiogenesis by targeting rho GTPases and extracellular signal-regulated kinase signaling pathways. Cancer Res 2009; 69(2): 518–525
- 91. Qiang L, Yang Y, You QD, Ma YJ, Yang L, Nie FF, Gu HY, Zhao L, Lu N, Qi Q, Liu W, Wang XT, Guo QL. Inhibition of glioblastoma growth and angiogenesis by gambogic acid: an *in vitro* and *in vivo* study. Biochem Pharmacol 2008; 75(5): 1083–1092
- Pang X, Yi Z, Zhang X, Sung B, Qu W, Lian X, Aggarwal BB, Liu M. Acetyl-11-keto-beta-boswellic acid inhibits prostate tumor growth by suppressing vascular endothelial growth factor

receptor 2-mediated angiogenesis. Cancer Res 2009; 69(14): 5893-5900

- Park B, Sung B, Yadav VR, Cho SG, Liu M, Aggarwal BB. Acetyl-11-keto-β-boswellic acid suppresses invasion of pancreatic cancer cells through the downregulation of CXCR4 chemokine receptor expression. Int J Cancer 2011; 129(1): 23–33
- 94. Pang X, Zhang L, Lai L, Chen J, Wu Y, Yi Z, Zhang J, Qu W, Aggarwal BB, Liu M. 1'-Acetoxychavicol acetate suppresses angiogenesis-mediated human prostate tumor growth by targeting VEGF-mediated Src-FAK-Rho GTPase-signaling pathway. Carcinogenesis 2011; 32(6): 904–912
- Kuang L, Wang L, Wang Q, Zhao Q, Du B, Li D, Luo J, Liu M, Hou A, Qian M. Cudratricusxanthone G inhibits human colorectal carcinoma cell invasion by MMP-2 down-regulation through suppressing activator protein-1 activity. Biochem Pharmacol 2011; 81(10): 1192–1200
- 96. Liu XD, Fan RF, Zhang Y, Yang HZ, Fang ZG, Guan WB, Lin DJ, Xiao RZ, Huang RW, Huang HQ, Liu PQ, Liu JJ. Down-regulation of telomerase activity and activation of caspase-3 are responsible for tanshinone I-induced apoptosis in monocyte leukemia cells *in vitro*. Int J Mol Sci 2010; 11(6): 2267–2280
- Wu Y, Fan Q, Lu N, Tao L, Gao Y, Qi Q, Guo Q. Breviscapineinduced apoptosis of human hepatocellular carcinoma cell line HepG2 was involved in its antitumor activity. Phytother Res 2010; 24(8): 1188–1194
- Lin J, Wei L, Xu W, Hong Z, Liu X, Peng J. Effect of *Hedyotis diffusa* Willd extract on tumor angiogenesis. Mol Med Report 2011; 4(6): 1283–1288
- 99. You J. Study on the tumor microenvironment and tumor vascular normalization in integrative treatment of tumor by Chinese medicine and western medicine.Chin J Integr Traidt West Med (Zhongguo Zhong Xi Yi Jie He Za Zhi) 2011; 31(8): 1127–1131 (in Chinese)
- Hida K, Hida Y, Amin DN, Flint AF, Panigrahy D, Morton CC, Klagsbrun M. Tumor-associated endothelial cells with cytogenetic abnormalities. Cancer Res 2004; 64(22): 8249–8255
- 101. Tian S, Hayes AJ, Metheny-Barlow LJ, Li LY. Stabilization of breast cancer xenograft tumour neovasculature by angiopoietin-1. Br J Cancer 2002; 86(4): 645–651
- Metheny-Barlow LJ, Li LY. The enigmatic role of angiopoietin-1 in tumor angiogenesis. Cell Res 2003; 13(5): 309–317
- 103. Inai T, Mancuso M, Hashizume H, Baffert F, Haskell A, Baluk P, Hu-Lowe DD, Shalinsky DR, Thurston G, Yancopoulos GD, McDonald DM. Inhibition of vascular endothelial growth factor (VEGF) signaling in cancer causes loss of endothelial fenestrations, regression of tumor vessels, and appearance of basement membrane ghosts. Am J Pathol 2004; 165(1): 35–52
- 104. Coulon C, Georgiadou M, Roncal C, De Bock K, Langenberg T, Carmeliet P. From vessel sprouting to normalization: role of the prolyl hydroxylase domain protein/hypoxia-inducible factor oxygen-sensing machinery. Arterioscler Thromb Vasc Biol 2010; 30 (12): 2331–2336
- 105. Sorensen AG, Batchelor TT, Zhang WT, Chen PJ, Yeo P, Wang M, Jennings D, Wen PY, Lahdenranta J, Ancukiewicz M, di Tomaso E, Duda DG, Jain RK. A "vascular normalization index" as potential mechanistic biomarker to predict survival after a single dose of cediranib in recurrent glioblastoma patients. Cancer Res

2009; 69(13): 5296-5300

106. Zhang Q, Bindokas V, Shen J, Fan H, Hoffman RM, Xing HR. Time-course imaging of therapeutic functional tumor vascular normalization by antiangiogenic agents. Mol Cancer Ther 2011; 10(7): 1173–1184

107. Hormigo A, Gutin PH, Rafii S. Tracking normalization of brain tumor vasculature by magnetic imaging and proangiogenic biomarkers. Cancer Cell 2007; 11(1): 6–8