

# Contrasting arthropod communities associated with dwarf mistletoes *Arceuthobium globosum* and *A. vaginatum* and their host *Pinus hartwegii*

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**Abstract** Arthropod communities and epiphytic plants associated with tree canopies have been widely studied and have revealed a great diversity of organisms; however, the community hosted by parasitic plants, such as dwarf mistletoes, remains poorly known. In the coniferous forests of North America, dwarf mistletoe infection (*Arceuthobium* spp.) significantly damages the health of the forest, causing large financial losses for the forest industry, but it also positively affects diversity, especially of mammals and birds. This study examined the attributes of the arthropod communities associated with two species of dwarf

mistletoe [*Arceuthobium globosum* Hawksw. & Wiens and *A. vaginatum* (Humb. & Bonpl. ex Willd.) J. Presl] and their host *Pinus hartwegii* Lindl. In 2010, in five sites located in Zoquiapan (Central Mexico), we collected plant tissue from the three species bimonthly. Arthropods were separated from the plant tissue and identified to the finest level possible. We collected 32,059 individuals, for which 51 morphospecies were identified, belonging to 15 taxonomic orders; the most abundant orders for the three plants were Prostigmata, Thysanoptera and Homoptera. The community associated with *P. hartwegii* had the highest value of diversity ( $H' = 1.47$ ; *A. globosum*,  $H' = 0.64$ ; *A. vaginatum*,  $H' = 0.68$ ) and species richness ( $S = 40$ ; *A. globosum*,  $S = 30$ ; *A. vaginatum*,  $S = 35$ ); while abundance was significantly higher for the mistletoes (*A. globosum*,  $n = 407$  individuals/sample; *A. vaginatum*,  $n = 536$  individuals/sample; *P. hartwegii*,  $n = 134$  individuals/sample). Species richness, abundance and diversity were significantly different for the three studied plants, as well for sampling month and the interaction of these two factors (except for diversity). The results suggest that the canopy of *P. hartwegii* is an important element in the ecosystem, providing a mosaic of resources and conditions to the associated fauna. We also propose that mistletoes are key species within the forest canopy, as they greatly influence the establishment of diverse organisms, particularly arthropods.

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## Introduction

Insect–plant interaction is one of the most studied biotic relationships because it includes two major biological groups: plants, which represent the largest biomass source on Earth (Schoonhoven et al. 2005); and arthropods, which comprise most of the living taxa (Borror et al. 1989). Arthropods play important roles in plant community dynamics, acting as herbivores, pollinators and seed dispersers (Samways 1994), and plants offer food and shelter for these organisms (Schoonhoven et al. 2005). The attributes of the arthropod community within a host plant are influenced by the plant's features such as size, structural complexity, phenology (Lawton 1983; Langellotto and Denno 2004; López-Gómez and Cano-Santana 2010; Schowalter 2011), food availability, primary productivity, presence of prey (Polis et al. 1997; Mittelbach et al. 2001), environmental conditions (Schowalter 2011) and presence of key species (Power et al. 1996; Letourneau and Dyer 1998).

Forest canopies are characterized by a high diversity and abundance of organisms, including a great variety of arthropods (Southwood et al. 1982; Stork 1991; Basset and Arthington 1992; Tovar-Sánchez et al. 2003) because of the great variety of exploitable resources and conditions (Morecroft et al. 1985; Fitzjarrald and Moore 1995), including a vast number of epiphytic and parasitic plants that increase the environmental complexity and provide sites for colonization (Shaw 2004). Some studies have reported that epiphytic plants contain a large variety of microarthropods, mainly mites (Acari) and springtails (Collembola) (Yanoviak and Nadkarni 2001; Yanoviak et al. 2003). Nevertheless, little attention has been paid to an important component on forest canopies and its interaction with invertebrate organisms: mistletoes.

Mistletoes have long been treated as pests on forestry lands because of their negative effects on hosts performance, such as decreased growth and fecundity (Mathiasen et al. 2008), but are now known to play key role in their biological community by improving ecosystem processes such as decomposition and nutrient cycling (March and Watson 2007; Ndagurwa et al. 2014) and enhancing vertebrate and invertebrates diversity (Watson 2001; Watson and Herring 2012).

Dwarf mistletoe infection (*Arceuthobium* spp., Viscaceae) of coniferous trees is worrying worldwide because of their negative impacts on economic and forestry activities (Stevens and Hawksworth 1984; Hawksworth and Wiens 1996). In Mexico, dwarf mistletoe infection is considered the second leading cause of forest damage, after fires (Hawksworth 1983). The biology, physiology and systematic of dwarf mistletoes are well documented

(Hawksworth and Wiens 1996; Mathiasen et al. 2008); however, other important ecological aspects, such as the relationship with arthropods, have received less attention.

Mistletoes can be tightly associated with arthropods, since many species can act as pollinators (Coleoptera, Diptera, Hymenoptera, and Lepidoptera; Penfield et al. 1976; Whittaker 1984; Hawksworth and Wiens 1996), and others can feed on its tissues (Hawksworth and Wiens 1996). Nevertheless, few studies have focused on the arthropod community held within mistletoes (Whittaker 1984). Burns et al. (2011) found 10 arthropod orders inhabiting the box mistletoe (*Amyema miquelii*) and its host (*Eucalyptus* spp.), which had very similar arthropod community compositions, but eucalyptus had greater densities of arthropods. The most representative orders were, in order of abundance, Hemiptera, Thysanoptera, Coleoptera, Araneae, Hymenoptera and Psocoptera. Results from other studies of arthropods communities on mistletoes, which did not include any host comparisons, are very similar (Room 1972; Whittaker 1984; Tassone and Majer 1997; Anderson and Braby 2009) and include the presence of other orders such as Collembola, Coleoptera and Lepidoptera (Anderson and Braby 2009).

However, to our knowledge, no study has been published for the community associated with dwarf mistletoes (*Arceuthobium* spp.), although various studies report single species associations. Several mite species (Prostigmata and Mesostigmata) have been described as tightly associated with different *Arceuthobium* species (Stevens and Hawksworth 1970; Kennett 1963; Pritchard and Baker 1958; Mathiasen et al. 2008), and larvae of a phytophagous lepidopteran as a specific consumers (Mooney 2001; Larsen et al. 1995).

The goal of this study was to compare the arthropod community attributes (species richness, diversity, abundance and composition) among three plant species: two dwarf mistletoes [*Arceuthobium globosum* Hawksw. & Wiens and *A. vaginatum* (Humb. & Bonpl. ex Willd.) J. Presl] and its host plant (*Pinus hartwegii* Lindl.) in the Zoquiapan region of Central Mexico.

Increasing our knowledge about the arthropod community associated with dwarf mistletoes and their pine host will contribute to a better understanding of arthropod–plant interactions and the role of dwarf mistletoes in these communities, other than the traditional concept of these organisms as harmful pests. Even more, the understanding of the pine-dwarf mistletoe system may be useful to plan managing strategies when needed by detecting phytophagous insects associated with dwarf mistletoes and using them for biological control.

## Materials and methods

### Study area

The study was conducted in the Zoquiapan subregion within the Iztaccíhuatl-Popocatepetl National Park in Central Mexico (19°15–29'N, 98°37–45'W). The area occupies 19,400 ha and has a subhumid temperate climate with summer rains; the mean annual temperature is 9.7 °C, and the annual precipitation is 941 mm (Arriaga et al. 2002; SMN 2013). The vegetation types found in this area are *Pinus*, *Abies* and *Quercus* forests; nonetheless, the flora is mainly composed of *Pinus hartwegii* (Obieta and Sarukhán 1981). Other tree species, such as *P. montezumae*, *P. ayacahuite*, *Cupressus lusitanica* and *Alnus jorullensis*, occur at low frequency. Forest understory is represented by tussocky grasses such as *Muhlenbergia macroura*, *M. quadridentata* and *Festuca toluensis*. The rainy season is from May to October, and the dry season is from November to April (Obieta and Sarukhán 1981).

### Methods

#### Study system

Arthropod communities in association with two dwarf mistletoe species (*A. globosum* and *A. vaginatum*) and their host (*Pinus hartwegii*) were described. *Arceuthobium globosum* subsp. *grandicaule* (Viscaceae) is a highly branched parasitic shrub, 20–70 cm tall, with yellow or greenish stems; its staminate flowers are 5 mm wide. Anthesis was recorded between January and May, with a peak from March to April. Fruits ripen from July to October, and seeds are dispersed from July to November, with a peak in August and September. This plant parasitizes 12 species of Mexican pines; it is distributed from central Mexico to Guatemala and Honduras, within an altitudinal range of 2450–4000 m a.s.l. (Hawksworth and Wiens 1996).

*Arceuthobium vaginatum* subsp. *vaginatum* is a dark brown or black (sometimes reddish), hemiparasitic shrub 20–30 cm tall. The dioecious plants have a remarkable sexual dimorphism, with the staminate individuals larger than the pistillate. Anthesis usually occurs in March and April, and fruits reach maturity in August of the next year. *Arceuthobium vaginatum* parasitizes 13 species of Mexican pines; it is distributed from northern to central Mexico, in an altitudinal range of 2100–3900 m a.s.l. (Hawksworth and Wiens 1996).

The host for these two dwarf mistletoes in the study site is *Pinus hartwegii* (Pinaceae), which is an evergreen coniferous tree 15–30 m tall, distributed from Mexico to Honduras in temperate subhumid zones at 2850–4200 m a.

s.l. It has thick, rough, scaly, dark brown to gray bark; needles 7–14 cm long are in fascicles of 2–6 (de Rzedowski et al. 2001).

#### Sampling method

We selected five *Pinus hartwegii* stands of approximately 100 m<sup>2</sup> and at least 500 m from each other; the two dwarf mistletoe species were present in each stand. The sites were located between 19°17'0.5" and 19°18'19"N and 98°42'26.4" and 98°40'30.7"W at 3270–3430 m a.s.l. In each stand, we collected stems from both mistletoe species and terminal branches with needles of *P. hartwegii* that were infected by any of the two mistletoe species. Approximately, 725 cm<sup>3</sup> of tissue were collected for each species, from at least five different plants. This procedure was repeated every 2 months for 1 year, from March 2010 to January 2011 for a total of 90 samples: 5 sites × 3 studied species × 6 samplings.

Each sample was processed in a Berlese–Tullgren funnel for 3 days without heat and 3 days with a heat source, after which we collected the arthropods from the bottom of the funnel for posterior counting and identification to the finest taxonomic level possible. Different identification keys were used, according to the relevant taxonomic group.

#### Data analysis

To evaluate sampling efficiency, we built two species accumulation curves, one for all arthropod species pooled together and one separating the arthropods associated with each plant species (Soberón and Llorente 1993). For the two curves, we implemented the Vegan package of R (R Core Development Team 2015) using the random method with 100 permutations. We also computed species richness (*S*), abundance (*N*), diversity (*H'*), evenness (*J'*), Jackknife first order (*S'*, a species pool estimation) and the percentage of reached richness in the samples (based on the Jackknife estimator). The last two were also calculated for total arthropod community. Diversity was estimated with the Shannon–Wiener index (*H'*) using natural logarithm (Krebs 1999); for evenness, we used the Pielou Index (*J'*) (Krebs 1999).

Then, we compared species richness similarity between the three plant species with the Jaccard similarity index (*SI<sub>j</sub>*) (Krebs 1999). To estimate community similarities from species relative frequencies, we constructed a Bray–Curtis similarity dendrogram (Legendre and Legendre 1998). We also quantified the number of morphospecies present exclusively in one of the three plant species (i.e., two mistletoes and pine) and the number of morphospecies that were shared among the plant species. These results were illustrated in a Venn diagram.

Community composition to the order level was compared among species and sampling date using a contingency table to examine the frequency of each order with each plant species. The table was tested with a  $\chi^2$  test of independence (Zar 2010), and, if significant, a standardized residues test was performed (Zar 2010).

To determine the effect of the plant species and the sampling month on arthropod species richness, diversity and abundance, we conducted a repeated measure ANOVA for each attribute (Zar 2010). When the latter was significant ( $p < 0.05$ ), we performed the post hoc Tukey test to identify homogeneous groups. In cases where the Tukey test did not show any significantly different groups (even when the ANOVA test was significant), we used the Fisher test (Zar 2010).

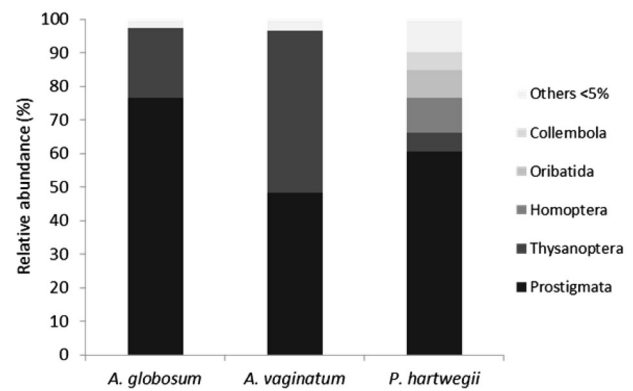
To determine differences among the arthropod community assemblage of three habitat types (pine, *A. vaginatum* and *A. globosum*), we used a principal coordinate analysis (PCoA). The Adonis test was used for finding significant differences in abundance of taxa comparing pine versus *A. vaginatum* versus *A. globosum* treatments of arthropods communities. When the Adonis test was significant ( $p < 0.05$ ), additional Adonis tests were done between all possible pair comparisons of treatments (pine vs. *A. vaginatum*, pine vs. *A. globosum*, *A. vaginatum* vs. *A. globosum*). All statistical analyses were done in the Vegan package of R (R Core Development Team 2015).

## Results

### General community composition

A total of 32,059 individuals were collected (both adult and immature stages) from 89 samples (*A. vaginatum* sample from March went missing). Of these, 51 morphospecies were recognized; 20,619 individual were collected during the rainy season (May, July and September) and 11,440 during the dry season (January, March and November). Of the 51 morphospecies, 15 were identified to order level, 26 to family, six to genera and only four to species (see Appendix for details).

Fifteen orders were determined, listed from most to least abundant: Prostigmata, Thysanoptera, Oribatida, Homoptera, Collembola, Psocoptera, Hymenoptera, Hemiptera, Coleoptera, Mesostigmata, Lepidoptera, Diptera, Araneae, Opiliones and Mecoptera (orders according Borror et al. 1989; Fig. 1). Subclass Acari was very abundant on the three species and was represented by three orders: Prostigmata, Mesostigmata, and Oribatida. Prostigmata was the dominant group in the three plants, and the most abundant genus was *Brevipalpus*. For Oribatida, the greatest abundance and diversity were associated with *P.*



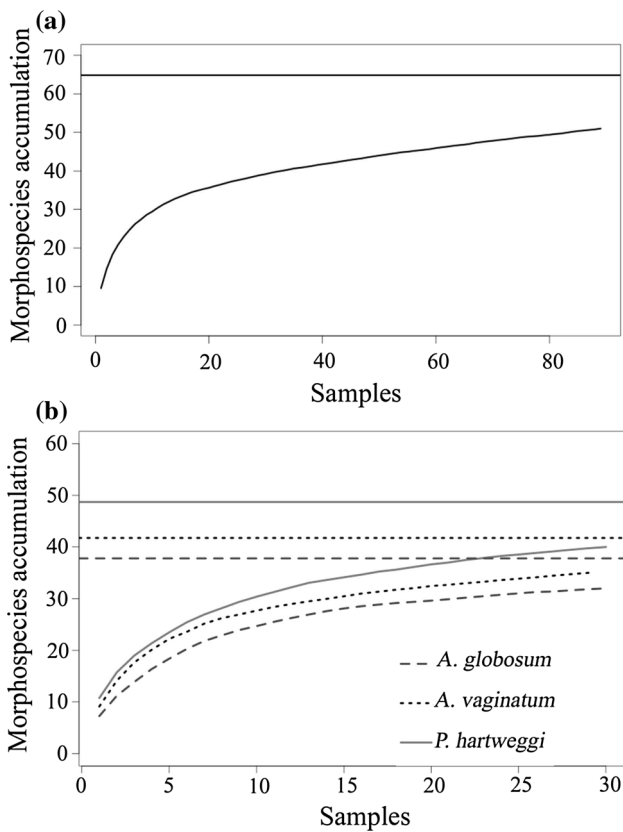
**Fig. 1** Relative abundance (%) of the arthropod orders associated with *P. hartwegii* ( $N = 4294$ ), *A. globosum* ( $N = 12,224$ ) and *A. vaginatum* ( $N = 15,541$ ). “Others” are orders with < 5% of abundance (Psocoptera, Hymenoptera, Hemiptera, Coleoptera, Mesostigmata, Lepidoptera, Diptera, Araneae, Opiliones and Mecoptera)

*hartwegii*, on which 8% of the specimens were found. Mesostigmata, however, was very infrequent (< 2%), and most individuals belonged to the family Phytoseiidae.

Thysanoptera, the second most abundant order, represented 21, 48 and 5% of the arthropods associated with *A. globosum*, *A. vaginatum* and *P. hartwegii*, respectively (Fig. 1). At least three species were found: *Frankliniella toluensis*, *F. fallaciosus* and *F. minuta*, plus a possible new species for the genus. Although Homoptera had a relative low abundance, it represented 10% of the arthropods associated with *P. hartwegii*. Collembola order, represented by 21 species of suborder Entomobryomorpha and 22 morphospecies of suborder Poduromorpha, represented 5% of the abundance on pine needles, while on both mistletoes it contributed < 1% of the abundance. The remaining nine orders (Psocoptera, Hymenoptera, Hemiptera, Coleoptera, Lepidoptera, Diptera, Araneae, Opiliones and Mecoptera) represented < 5% of the abundance for each plant species.

Sampling efficiency was high because the arthropod richnesses were very close to the species pool estimations ( $S'$ ) for the total community (Fig. 2a) and for each plant (Fig. 2b), and due to the elevated percentages of the richnesses (> 78%; Table 1).

The Bray–Curtis dendrogram and Jaccard similarity index ( $SI_j$ ) showed that similarity between communities is greater between both dwarf mistletoe species ( $SI_j = 0.625$ ), than any of the mistletoes with *P. hartwegii* (*A. vaginatum*  $SI_j = 0.53$ ; *A. globosum*  $SI_j = 0.59$ ; Fig. 3). Likewise, most of morphospecies (47.05%) were present in the three plants, and a few were exclusive for one of the plants (*A. vaginatum* = 11.76%, *A. globosum* = 5.88%, *P. hartwegii* = 19.60%; Fig. 4).

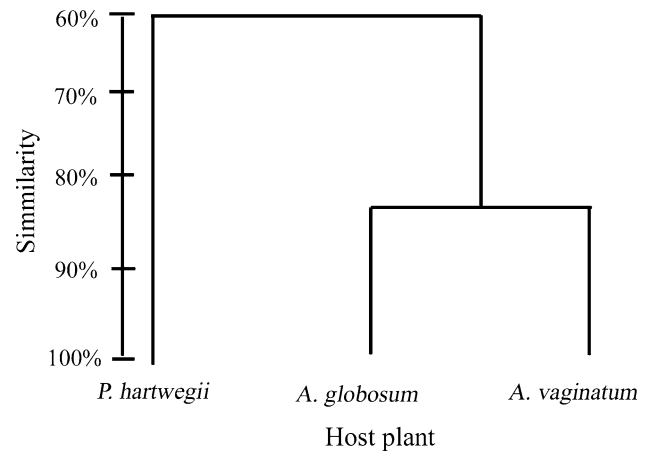


**Fig. 2** Accumulation curve of **a** pooled morphospecies and **b** morphospecies for each plant species. Horizontal lines represent species pool estimation ( $S'$ ) for each plant

**Community composition per plant species**

*Arceuthobium vaginatum* and *A. globosum* had the greatest values of abundance and density of arthropods, but *P. hartwegii* had the greatest values of species richness, diversity and evenness (Table 1).

The abundance of the most abundant arthropods (Prostigmata and Thysanoptera) depended on the type of plant ( $\chi^2 = 9539$ ,  $df = 28$ ,  $p < 0.001$ ; Fig. 5a). Prostigmata species were significantly more abundant in *A. globosum* than expected from random, and in the same way, significantly lower in *P. hartwegii* (Fig. 5a). Meanwhile, individuals of Thysanoptera were significantly more abundant



**Fig. 3** Bray-Curtis dendrogram comparing arthropod communities associated with *A. globosum*, *A. vaginatum* and *P. hartwegii*

in *A. vaginatum* and significantly lower in *A. globosum* (Fig. 5a).

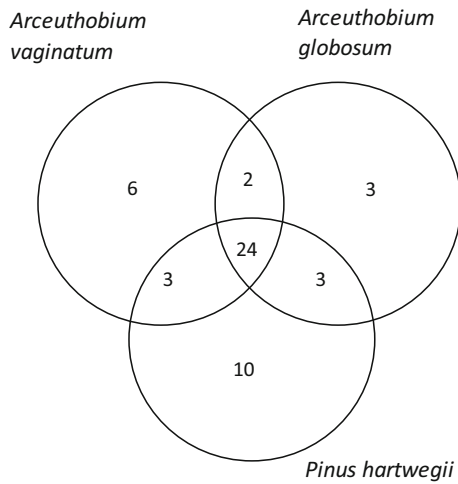
The abundance of each order was also dependent on sampling month ( $\chi^2 = 14,625$ ,  $df = 55$ ,  $p < 0.001$ ; Fig. 5b). The most notable trends were for Prostigmata, which was significantly more abundant in July, September, November and January (than expected for random), and for Thysanoptera, which was significant more abundant in March and May (Fig. 5b); on the contrary, the frequencies of Prostigmata and Thysanoptera were significant lower in the remaining months of sampling, for each case (i.e., March and May for Prostigmata, and July, September, November and January for Thysanoptera; Fig. 5b).

**Effect of plant species and sampling month on the community attributes**

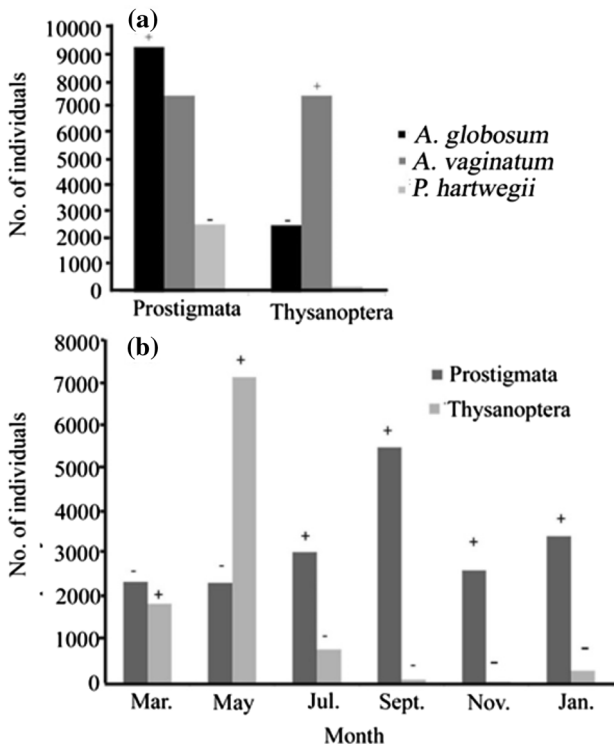
There were significant effects of the plant ( $F_{2, 14} = 14.91$ ,  $p < 0.001$ ), the sampling month ( $F_{5, 14} = 4.02$ ,  $p = 0.003$ ), and their statistical interaction (plant  $\times$  sampling month;  $F_{10, 14} = 2.90$ ,  $p = 0.046$ ) on the arthropod morphospecies richness. Arthropod richness was higher on *P. hartwegii* and *A. globosum* communities than on *A. vaginatum* (Fig. 6a). Significant differences in sampling month were only found in March and May; *P. hartwegii* had a higher

**Table 1** Abundance ( $N$ ), density, morphospecies richness ( $S$ ), evenness ( $J'$ ), diversity ( $H'$ ), Jackknife first order ( $S'$ ) and percentage of reached richness (using Jackknife species pool estimator) of arthropods associated with three plant species

Plant species	$N$	Density $\pm$ SE (individuals/sample)	$S$	$J'$	$H'$	$S' \pm$ SE	Percentage of richness reached (%)
<i>A. globosum</i> ( $n = 30$ )	12,224	407.5 $\pm$ 79.4	30	0.330	0.642	37.8 $\pm$ 2.74	79.36
<i>A. vaginatum</i> ( $n = 29$ )	15,541	545.9 $\pm$ 134.8	35	0.298	0.645	41.75 $\pm$ 2.55	83.83
<i>P. hartwegii</i> ( $n = 30$ )	4294	143.1 $\pm$ 22.6	40	0.625	1.468	48.7 $\pm$ 3.50	82.13
Total arthropod community			51			64.84 $\pm$ 3.95	78.65



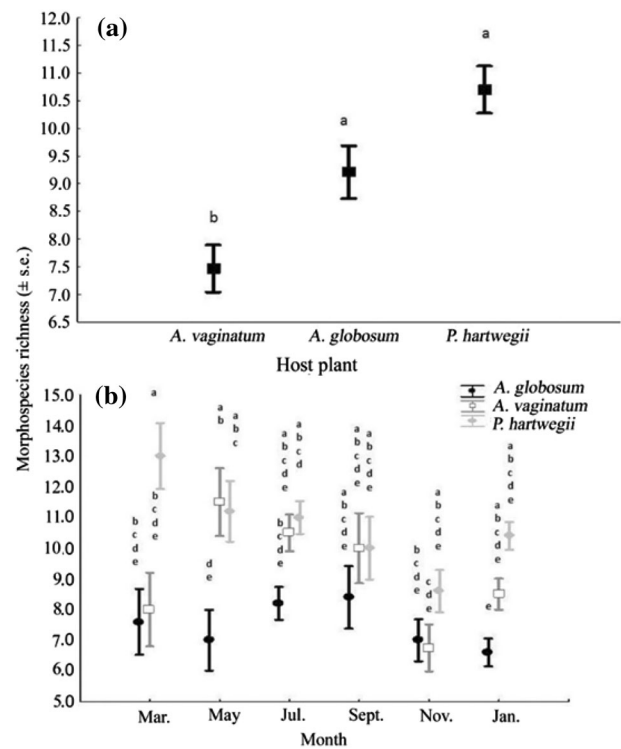
**Fig. 4** Number of arthropod morphospecies exclusive to and shared by the three plant species studied in this work (*A. vaginatum*, *A. globosum* or *P. hartwegii*)



**Fig. 5** Absolute frequencies of the most abundant orders of arthropods **a** on each plant and **b** for each sampling month. + higher frequency than expected; - lower frequency than expected

richness in March than the other two plants did, whereas *A. globosum* had a lower richness on May (Fig. 6b).

We also found a significant effect of plant species ( $F_{2, 14} = 14.9, p < 0.001$ ), sampling month ( $F_{5, 14} = 4.5, p = 0.001$ ), and their interaction (plant  $\times$  sampling month;  $F_{10, 14} = 3.6, p = 0.001$ ) on arthropod mean abundance. Arthropod abundance was significantly higher on the two

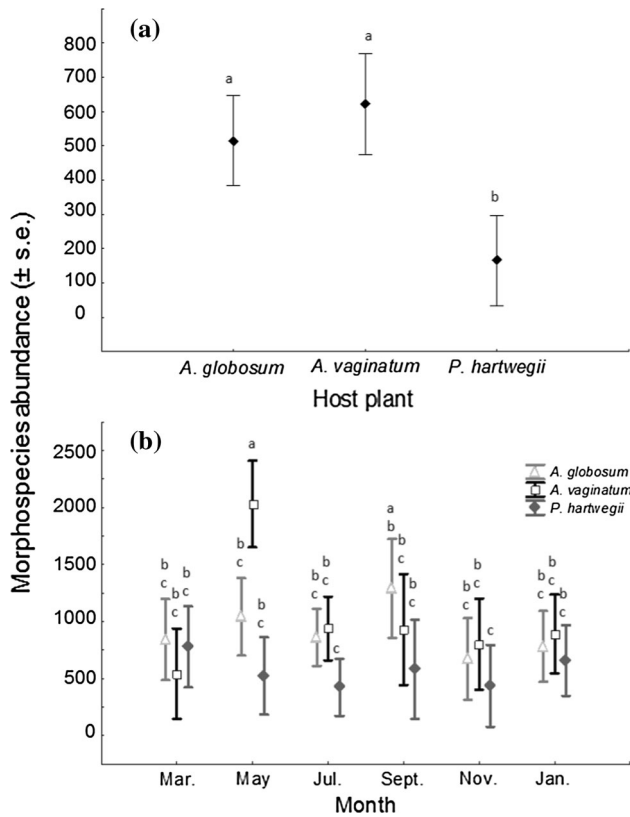


**Fig. 6** **a** Morphospecies richness ( $\pm$  SE) on each plant species. **b** Morphospecies richness ( $\pm$  SE) on each plant species by sampling month. Letters represent significant differences among groups, according to Tukey's post hoc test ( $p < 0.05$ )

mistletoe species, than on the pine host (Fig. 7a). On the other hand, mean abundance on *A. vaginatum* was significantly higher only in May than the other two plant species (Fig. 7b).

For diversity ( $H'$ ), there was a significant effect of plant species ( $F_{2, 14} = 47.47, p < 0.001$ ) and sampling month ( $F_{5, 14} = 2.57, p = 0.037$ ), but not for their interaction ( $F_{10, 14} = 1.22, p = 0.299$ ). The community associated with *P. hartwegii* was more diverse than the communities associated with the mistletoes (Fig. 8a). Moreover, diversity was higher in March, declined over time, and diversity in March was higher than in November and January (Fig. 8b).

According to the PCoA, the arthropod communities differed significantly among the three habitat types (Adonis test,  $p < 0.001$ ; Fig. 9), besides there were significant differences between arthropod communities of pine versus *A. vaginatum* (Adonis test,  $p < 0.001$ ) and pine versus *A. globosum* (Adonis test,  $p < 0.001$ ); nevertheless, *A. vaginatum* versus *A. globosum* comparison did not show any significant difference (Adonis test,  $p = 0.30$ ; Fig. 9).



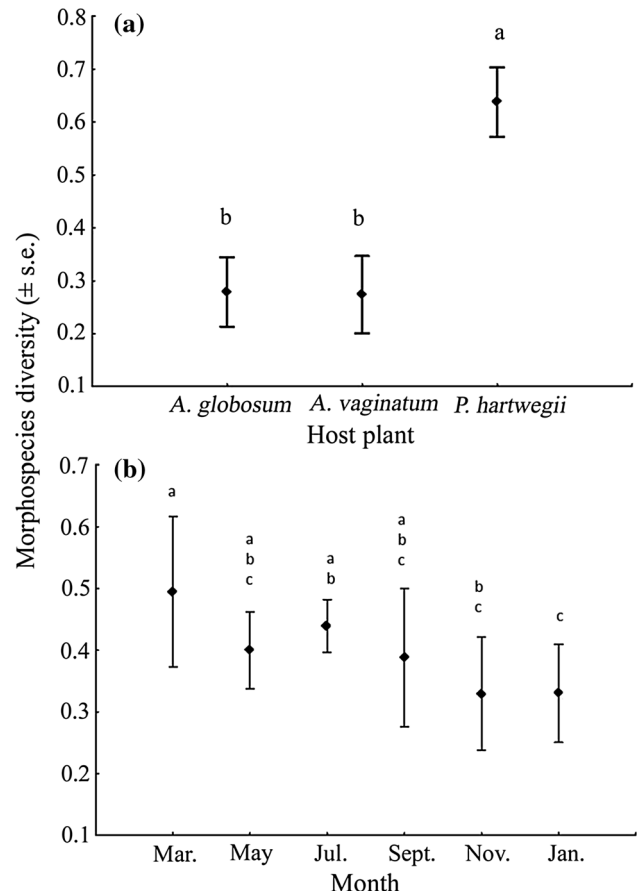
**Fig. 7** **a** Morphospecies abundance (± SE) on each species. **b** Morphospecies abundance (± SE) on each species by sampling month. Letters represent significant differences among groups, according to Tukey's post hoc test ( $p < 0.05$ )

**Discussion**

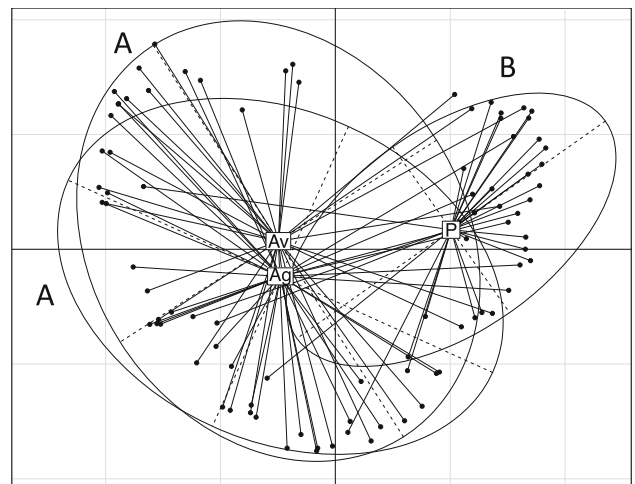
**Community structure and temporal variation**

Results clearly showed that *P. hartwegii* has greater morphospecies richness, diversity and evenness, and a lower abundance of arthropods compared with dwarf mistletoes' aerial tissues. Communities associated with the two mistletoes have similar abundances of certain groups, such as Thysanoptera and Prostigmata, which can be attributed to their phylogenetic closeness and great phenotypic resemblance (Hawksworth and Wiens 1996), thus the provision of similar resources and conditions.

The great diversity of arthropods found on *P. hartwegii* needles can be attributed to two things: (1) this species provides a larger colonization island than the mistletoes do (Schoener 1976; Connor and McCoy 1979; Lomolino 2000; Williamson et al. 2001; López-Gómez and Cano-Santana 2010), and (2) needles on terminal branches have a rosette-like shape, sheltering arthropods from climatic variations and predators (López-Gómez and Cano-Santana 2010). The scale-like leaves of the dwarf mistletoes provide a smaller colonization island and are ineffective



**Fig. 8** **a** Diversity ( $H'$ ) on each species and **b** in each sampling month. Letters represent significant differences among groups, according to Tukey's post hoc test ( $p < 0.05$ )



**Fig. 9** PCoA analysis comparing species abundance in the arthropod community samples from the three habitats: pine (P), *A. vaginatum* (Av), *A. globosum* (Ag). Different letters denote significant differences in an Adonis test ( $p < 0.05$ )

shelters for many arthropods, especially those of medium to large size. In addition, the higher diversity of arthropods

on pine needles is likely favored by tourist species (i.e., a nonpredatory species that does not have a lasting association with a host plant; Moran and Southwood 1982) looking for the more heterogeneous resources that pines provide. Individuals belonging to Homoptera, Coleoptera, Diptera, Hemiptera, Homoptera, Araneae and Hymenoptera were found living inside pine fascicles, which also tend to accumulate debris (our personal observations) that can be used as feeding sites by saprophagous arthropods such as springtails, oribatid mites, Opiliones, and Psocoptera barkflies (Borrer et al. 1989; Schowalter 2011).

The greater variety of habitats and resources of the pine foliage may also increase the number of trophic levels that favor top predators such as spiders, mites and parasitic wasps Mesostigmata (Schoener 1989; Post et al. 2000). Winged insects, however, may be underestimated since they can fly to avoid capture.

Although *A. globosum* (up to 70 cm tall) and *A. vaginatum* (maximum height 55 cm) clearly differ in size (Hawksworth and Wiens 1996), our results showed that the smaller *A. vaginatum* had a greater abundance, density and richness of arthropods than did *A. globosum*. However, we collected the same volume of tissue for both species, regardless of their size, so we cannot draw precise conclusions about any size effect. Moreover, in a study in 2009 (a year before this study) in the same area, *A. vaginatum* was more prevalent than *A. globosum* (27.6 and 6.7% of infected trees, respectively), as well more abundant (mean dwarf mistletoe rating system: 2.05 and 1.18, respectively) (Queijeiro-Bolaños et al. 2014). Thus, despite its size *A. vaginatum* apparently provides more available habitat for the arthropod community. Further studies are clearly necessary to clarify contributions of size and abundance to the arthropod community composition.

Arthropod communities on dwarf mistletoes had a lower species richness compared with that reported for other mistletoe species such as *Decaisnina signata* (113 arthropod species and 12 orders; Anderson and Braby 2009) and *Amyema miquelli* (10 orders; Burns et al. 2011). Loranthaceous mistletoes are generally larger (including branches, leaves, flowers and fruits) than dwarf mistletoes and may thus offer greater colonization space and feeding resources. In contrast, dwarf mistletoes have small scale-like leaves that may be a more suitable refuge for microarthropods (body length < 5 mm) such as thrips and mites. Because of their smaller flowers, dwarf mistletoes may depend on insects such as encyrtid wasps (Hymenoptera: Encyrtidae) for pollination (when they are not wind-pollinated) (Penfield et al. 1976).

*Arceuthobium vaginatum* presents a marked sexual dimorphism, with staminate plants usually larger than the pistillate (Hawksworth et al. 2002), but *A. globosum* does not. We did not differentiate between male and female

plants, and as far as we know, no biotic associations have been correlated with the sexual dimorphism of mistletoes. However, differences in species richness and composition can be expected based on variations in size derived from sexual dimorphism; larger mistletoes can host greater species richness (Anderson and Braby 2009). It has been reported for plants in general that, depending on the gender, nutrients can vary, and frequently arthropods prefer male plants (Agren et al. 1999). Nevertheless, studies focused on sexual dimorphism in mistletoes are still needed.

Temporal variation of arthropod communities showed subtle changes in richness and composition; the greatest abundance of organisms was present in the rainy season (May–October), with a notable increase in abundance of individuals belonging to orders Thysanoptera, Collembola, Psocoptera, Hymenoptera, Coleoptera and Oribatida. Throughout this study, we did not record humidity or leaf production or herbivory rates because the rainy season is known to be strongly related to increased presence of herbivores as a direct consequence of greater resource availability (Basset 1996; Stork et al. 1997).

### Functional role of arthropods

We found 15 orders of arthropods, which include phytophagous and predatory organisms, on the three plant species. Stevens and Hawksworth (1970, 1984) described generalist and specialist insects feeding or living within dwarf mistletoes; the most frequent belonged to the orders Lepidoptera, Hemiptera, Coleoptera, Thysanoptera, Prostigmata, Mesostigmata, and Araneae. Even so, there is very little evidence that phytophagous insects on dwarf mistletoes in Mexico can function as agents to control these parasitic plants.

Prostigmata was the most abundant order of arthropods associated with the three plant species; the species within these order are reported as phytophagous (Borrer et al. 1989), and some have been described as specialist shoot predators for dwarf mistletoes (such as *Typhlodromus arceuthobius* [Phytoseiidae], *T. pusillum*, *Paraphytopus arceuthobii* [Eriophyidae] and *Brevipalpus porca* [Tenuipalpidae]; Keifer 1952; Pritchard and Baker 1958; Kennett 1963). Although we do not know the identity of all the morphospecies, the genus *Brevipalpus* and members of the families Phytoseiidae were present, indicating that they could act as predators for dwarf mistletoes' shoots. Mites are also a major group of saprophagous insects (Maraun and Scheu 2000) that can carry out the facilitation process, decomposition and nutrient cycling (Visser 1985).

On the other hand, the presence of Collembola in forest canopies has been reported as largely due to the association they have with epiphytic plants (Prinzing 1997). Although



springtails are considered general consumers, they can feed on fungal hyphae and spores, bacteria, detritus, pollen and mineral particles, and some have predatory habits (Castaño-Meneses et al. 2004). The rosette-like structure of pine needle fascicles may increase the retention of debris, favoring the presence of saprophagous arthropods such as springtails.

Thysanoptera was the second most abundant order. Thrips are commonly associated with dwarf mistletoes and have been described as shoot feeders (Stevens and Hawksworth 1970, 1984). The highest abundance of thrips (especially *Frankliniella toluensis*) was reported in the May sample of *A. vaginatum*, when it was flowering (Hawksworth and Wiens 1996). Thus, thrips could be pollinivores, although this possibility needs confirmation. These thrips are also an important prey for Phytoseiidae mites and egg feeding wasps (Morse and Hoddle 2006).

Although the homopteran members could not be identified to a more specific taxonomic level, they probably belong to the Mitiidae family, which are shoot feeders of *A. vaginatum* subsp. *cryptopodum* in the southern United States and northern Mexico (Kuijt 1955).

Adult Curculionidae beetles (Coleoptera) often oviposit on pine branches, and the hatching larvae feed on pine buds (Hawksworth and Wiens 1996). So beetle larvae might also exploit mistletoe shoots as a feeding resource.

It is feasible that wasps (Hymenoptera) are closely associated with the three plant species. Some wasp species feed on nectar and pollen of dwarf mistletoe flowers (Goulet and Huber 1993), but they can parasitize other arthropods, such as members of Araneae, Coleoptera, Heteroptera, Homoptera and Lepidoptera, which were all present in the three plant species.

Members of the order Lepidoptera, such as the phytophagous larvae of *Mitoura* sp. and *Neophasia* sp. found on *A. vaginatum* subsp. *cryptopodum* (Grimble and Beckworth 1993) may be mistletoes specialist consumers. We therefore need to determine the morphospecies of the lepidopterans to the finest taxonomic level (which we were not able to do, since we found only larvae) because these specialist organisms may have potential to control dwarf mistletoe.

Spiders are also predators and likely to be able to feed on all the other arthropods present on the mistletoes and the pine; it also can be parasitized by wasps.

## Conclusions

Each of the three plant species maintain different arthropod communities, which are very similar to each other, possibly because of the phylogenetic proximity of the three species. The pine species, however, is able to host a greater variety of organisms by providing a wider range of resources and refuge sites.

The role that parasitic plants such as mistletoes play in their communities can be diverse and complex, as was evident in our study system. Although dwarf mistletoes have been traditionally viewed as harmful parasites, they can also be a positive component of the canopy by supporting a vast arthropod community and providing suitable microhabitats and resources for a great variety of organisms. The great abundance of microarthropods found within these parasitic plants is supportive evidence for the theory that mistletoes act as a key group within their biological communities (Watson 2001; Watson and Herring 2012).

Deeper studies on the phytophagous groups (Thysanoptera, Prostigmata and Lepidoptera) should focus on their potential as biocontrol agents for dwarf mistletoe.

The study of such complex systems (like pine-mistletoe-arthropods) in forests should lead to more integrative strategies for forest management.

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## Appendix

List and abundances of arthropods species associated with the dwarf mistletoes *Arceuthobium globosum* and *A. vaginatum*, and their host *Pinus hartwegii*. Organisms were collected in the Iztaccíhuatl-Popocatepétl National Park, Mexico. The taxonomic classification was based on Borror et al. (1989).

Morphospecies	Taxonomic classification	Host plant		
		<i>A. globosum</i>	<i>A. vaginatum</i>	<i>P. hartwegii</i>
	Phylum Arthropoda			
	Class Arachnida			
	Order Araneae			
	Suborder Araneomorphae			
	Superfamily Araneoidea			
	Family Araneidae			
1	Araneidae sp. 1	1	0	0
2	Araneidae sp. 2	0	1	0
	Family Theridiidae			
3	Theridiidae sp. 1	7	5	2
4	Theridiidae sp. 2	0	1	0
5	Theridiidae sp. 3	0	0	1
	Family Linyphiidae			
6	Linyphiidae sp. 1	0	3	0
7	Linyphiidae sp. 2	0	0	1
	Family Tetragnathide			
8	Tetragnathide sp. 1	0	0	3
	Order Opiliones			
9	Opilione sp. 1	0	1	1
	Class Acari			
	Order Mesostigmata			
	Suborder Monogynapsida			
	Cohort Gamasina			
	Subcohort Dermanyssia			
	Super Family Phytoseioidea			
	Family Phytoseiidae			
10	Phytoseiidae sp. 1	5	16	10
11	Phytoseiidae sp. 2	5	5	48
12	Phytoseiidae sp. 3	5	0	3
	Suborder Prostigmata			
	Cohort Raphignathina			
	Superfamily Tetranychoidae			
	Family Tenuipalpidae			
	Genera <i>Brevipalpus</i>			
13	<i>Brevipalpus</i> sp. 1	9304	7427	2400
	Superfamily Tetranychoidae			
	Family Tetranychidae			
14	Tetranychidae sp. 1	15	45	153
	Superfamily Raphignathoidea			
	Family Camerobiidae			
	Genera <i>Camerobia</i>			
15	<i>Camerobia</i> sp. 1	18	8	12
	Cohort Parasitengomina			
	Superfamily Erythraeoidea			
	Family Erythraeidae			
16	Erythraeidae sp. 1	8	12	36
	Suborder Oribatida			
	Cohort Brachypylina			

Table a continued

Morphospecies	Taxonomic classification	Host plant		
		<i>A. globosum</i>	<i>A. vaginatum</i>	<i>P. hartwegii</i>
	Superfamily Cymbaeremaeoidea			
	Family Cymbaeremaeidae			
	Genera <i>Scapheremaeus</i>			
17	<i>Scapheremaeus</i> sp. 1	5	8	7
	Superfamily Ceratozetoidea			
	Family Ceratozetidae			
	Genera <i>Trichoribates</i>			
18	<i>Trichoribates</i> sp. 1	20	113	307
	Superfamily Oripodoidea			
	Family Oripodidae			
19	Oripodidae sp. 1	5	20	44
	Cohort Nothrina			
	Superfamily Crotonioidea			
	Family Camisiidae			
	Genera <i>Camisia</i>			
20	<i>Camisia</i> sp. 1	5	6	3
	Class Hexapoda			
	SubClass Collembola			
	Order Entomobryomorpha			
	Superfamily Entomobryoidea			
	Family Entomobryidae			
21	Entomobryidae sp. 1	28	45	227
	Order Poduromorpha			
	Superfamily Hypogastruroidea			
	Family Hypogastruridae			
22	Hypogastruridae sp. 1	0	4	1
	Class Insecta			
	Order Psocoptera			
	Family Lachesillidae			
	Subfamily Lachesillinae			
	Genera <i>Lachesilla</i>			
23	<i>Lachesilla pinicola</i>	24	44	161
	Order Thysanoptera			
	Family Thripidae			
	Genera <i>Frankiniella</i>			
24	<i>Frankiniella toluensis</i>	2548	7477	238
25	<i>Frankiniella fallaciosa</i>	6	14	0
26	<i>Frankiniella minuta</i>	0	2	0
27	<i>Frankiniella</i> sp. 1	0	1	0
	Order Hemiptera			
	Suborder Heteroptera			
28	Heteroptera Nymph sp. 1	23	84	24
29	Heteroptera sp. 1	9	13	13
30	Heteroptera sp. 2	0	0	5
	Suborder Homoptera			
31	Homoptera Nymph sp. 1	15	24	417
	Superfamily Membracoidea			

Table a continued

Morphospecies	Taxonomic classification	Host plant		
		<i>A. globosum</i>	<i>A. vaginatum</i>	<i>P. hartwegii</i>
	Family Membracidae			
32	Membracidae sp. 1	2	0	8
	Family Cicadellidae			
33	Cicadellidae sp. 1	0	0	22
	Order Hymenoptera			
	Suborder Apocrita			
34	Hymenoptera Larva sp. 1	64	58	83
	Superfamily Vespoidea			
	Family Vespidae			
35	Vespidae sp. 1	0	1	0
	Superfamily Ichneumonoidea			
	Family Ichneumonidae			
36	Ichneumonidae sp. 1	0	0	1
	Superfamily Platygastroidea			
	Family Platygastroidea			
37	Platygastroidea sp. 1	5	1	3
	Superfamily Ceraphonidea			
	Family Ceraphonidae			
38	Ceraphonidae sp. 1	0	1	0
	Superfamily Chilsidoidea			
	Family Chilsidoidea			
39	Chilsidoidea sp. 1	0	0	1
	Family Eulophidae			
40	Eulophidae sp. 1	1	0	0
	Order Lepidoptera			
41	Lepidoptera Larva sp. 1	23	30	5
42	Lepidoptera sp. 1	0	0	1
	Order Coleoptera			
43	Coleoptera Larva sp. 1	50	45	19
44	Coleoptera Larva sp. 2	13	5	16
	Family Curculionidae			
45	Curculionidae sp. 1	4	7	5
46	Curculionidae sp. 2	0	0	3
	Order Diptera			
47	Diptera Larva sp. 1	0	6	5
48	Diptera sp. 1	5	8	3
49	Diptera sp. 2	1	0	0
50	Diptera sp. 3	0	0	1
	Order Mecoptera			
51	Mecoptera sp. 1	0	0	1

## References

- Agren J, Danell K, Elmqvist T, Ericson L, Hjältén J (1999) Sexual dimorphism and biotic interactions. In: Geber MA, Dawson TE, Delph LF (eds) Gender and sexual dimorphism in flowering plants. Springer, New York, pp 217–246
- Anderson SJ, Braby MF (2009) Invertebrate diversity associated with tropical mistletoe in a suburban landscape from northern Australia. *North Territ Nat* 21:2–23
- Arriaga L, Espinoza JM, Aguilar C, Martínez E, Gómez L, Loa E (2002) Regiones terrestres prioritarias de México. Comisión Nacional para el Conocimiento y Uso de la Biodiversidad, Mexico City

- Basset Y (1996) Local communities of arboreal herbivores in Papua New Guinea predictors of insect variables. *Ecology* 77:1906–1919
- Basset Y, Arthington J (1992) The arthropod community of an Australian rainforest tree: abundance of component taxa. *Aust J Ecol* 17:89–98
- Borror DJ, Johnson NF, Triplehorn CA (1989) An introduction to the study of insects. Saunders College, Philadelphia
- Burns AE, Cunningham SA, Watson DM (2011) Arthropod assemblages in tree canopies: a comparison of orders on box mistletoe (*Amyema miquelii*) and its host eucalypts. *Aust J Entomol* 50:221–230
- Castaño-Meneses G, Palacios-Vargas JL, Cutz-Pool L (2004) Feeding habits of Collembola and their ecological niche. *Anales del Instituto de Biología, Serie Zoológica* 75:135–142
- Connor EF, McCoy E (1979) The statistics and biology of the species-area relationship. *Am Nat* 113:791–833
- de Rzedowski GC, Rzedowski J, and contributors (2001) Flora fanerogámica del Valle de México. Instituto de Ecología, A. C. and Comisión Nacional para el Conocimiento y Uso de la Biodiversidad. Pátzcuaro
- Fitzjarrald DR, Moore KE (1995) Physical mechanism of heat and mass exchange between forest and the atmosphere. In: Lowman MD, Nadkarni NM (eds) *Forest canopies*. Academic Press, San Diego, pp 45–72
- Goulet H, Huber J (1993) Hymenoptera of the world: an identification guide to families. Research Branch Agriculture, Ontario
- Grimble DG, Beckworth RC (1993) Temporal changes in presence of late instar *Mitoura spinetorum* (Lycaenidae) in eastern Oregon. *J Lepid Soc* 47:329–330
- Hawksworth FG (1983) Mistletoes as forest parasites. In: Calder M, Bernhardt P (eds) *The biology of mistletoes*. Academic Press, New York, pp 320–329
- Hawksworth FG, Wiens D (1996) Dwarf mistletoes: biology, pathology and systematics. *Agriculture Handbook* 709. U.S.D.A. Forest Service, Washington, DC
- Hawksworth FG, Wiens D, Geils BW (2002) *Arceuthobium* in North America. In: Geils BW, Cibrián-Tovar J, Moody B (eds) *Mistletoes of North American conifers*. U.S.D.A. Forest Service, Ogden, pp 29–56
- Keifer HH (1952) Eriophyid studies. XIX. Bulletin 41. State of California, Department of Agriculture, Sacramento, CA, pp 65–74
- Kennett CE (1963) Some species of *Typhlodromus* from dwarf mistletoes in North America (Acarina-Phytoseidae). *Pan Pac Entomol* 39:247–252
- Krebs CJ (1999) *Ecological methodology*. Addison-Wesley Educational Publication, Menlo Park
- Kuijt J (1955) Dwarf mistletoes. *Bot Rev* 10:569–619
- Langellotto GA, Denno RF (2004) Responses of invertebrate natural enemies to complex-structures habitats: a meta-analytical synthesis. *Oecologia* 139:1–10
- Larsen EM, Rodrick E, Milner R (eds) (1995) *Management recommendations for Washington's priority species, vol 1: invertebrates*. Washington Department of Fish and Wildlife, Olympia
- Lawton JH (1983) Plant architecture and the diversity of phytophagous insects. *Annu Rev Entomol* 28:23–39
- Legendre L, Legendre P (1998) *Numerical ecology*. Elsevier, Amsterdam
- Letourneau DK, Dyer LA (1998) Density patterns of *Piper* ant-plants and associated arthropods: top-predator trophic cascades in a terrestrial system. *Biotropica* 30:162–169
- Lomolino MV (2000) Ecology's most general, yet protean pattern: the species-area relationship. *J Biogeogr* 27:17–26
- López-Gómez V, Cano-Santana Z (2010) Best host-plant attribute for species-area relationship, and effects of the shade, conspecific distance and plant phenophase in an arthropod community within the grass *Muhlenbergia robusta*. *Entomol Sci* 13:174–182
- Maraun M, Scheu S (2000) The structure of oribatid mite communities (Acari, Oribatida): patterns, mechanisms and implications for future research. *Ecography* 23:374–383
- March WA, Watson D (2007) Parasites boost productivity: effects of mistletoe on litterfall dynamics in a temperate Australian forest. *Oecologia* 154:339–347
- Mathiasen RL, Nickrent DL, Shaw DC, Watson DM (2008) Mistletoes. Pathology, systematic, ecology and management. *Plant Dis* 92:988–1006
- Mittelbach GG, Steiner CF, Scheiner SM, Gross KL, Reynolds HL, Waide RB, Willig MR, Dodson SI, Gough L (2001) What is the observed relationship between species richness and productivity? *Ecology* 82:2381–2396
- Mooney KA (2001) The life history of *Dasyphyga alternosquamella* Ragonot (Pyralidae) feeding on the Southwestern dwarf mistletoe (*Arceuthobium vaginatum*) in Colorado. *J Lepid Soc* 55:140–143
- Moran VC, Southwood TRE (1982) The guild composition of arthropods communities in trees. *J Anim Ecol* 51:289–306
- Morecroft MT, Taylor P, Oliver HR (1985) Air and soil microclimates of deciduous woodland compared to an open. *Agric For Meteorol* 90:141–156
- Morse JG, Hoddle MS (2006) Invasion biology thrips. *Annu Rev Entomol* 51:67–89
- Ndagurwa GT, Dube JS, Mlambo D (2014) The influence of mistletoes on nutrient cycling in a semi-arid savanna, southwest Zimbabwe. *Plant Ecol* 215:15–26
- Obieta C, Sarukhán J (1981) Estructura y composición la vegetación herbácea de un bosque uniespecífico de *Pinus hartwegii*. I. Estructura y composición florística. *Boletín de la Sociedad Botánica de México* 41:75–125
- Penfield FB, Stevens RE, Hawksworth FG (1976) Pollination ecology of three Rocky Mountain dwarf mistletoe. *For Sci* 22:473–484
- Polis GA, Anderson WB, Holt RD (1997) Toward an integration of landscape and food web ecology: the dynamics of spatially subsidized food webs. *Annu Rev Ecol Syst* 28:289–316
- Post DM, Conners ME, Goldberg DS (2000) Prey preference by a top predator and the stability of linked food chains. *Ecology* 81:8–14
- Power ME, Tilman D, Estes JA, Menge B, Bond W, Mills LS, Daily G, Castilla JC, Lubchenco J, Paine RT (1996) Challenges in the quest for keystones. *Bioscience* 46:609–620
- Prinzling A (1997) Spatial and temporal use of microhabitats as a key strategy for the colonization of tree bark by *Entomobrya nivalis* L. (Collembola, Entomobryidae). In: Stork N, Adis J, Didham R (eds) *Canopy arthropods*. Chapman and Hall, London, pp 453–476
- Pritchard AE, Baker EW (1958) *The false spider mites (Acarina: Tenuipalpidae)*. University of California Publications in Entomology, Berkeley 14:207–208
- Queijeiro-Bolaños M, Cano-Santana Z, García-Guzmán G (2014) Incidence, severity and aggregation patterns of two sympatric dwarf mistletoe species (*Arceuthobium* spp.) in Central Mexico. *Eur J Forest Res* 113:297–306
- R Core Development Team (2015) R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. <http://www.R-project.org/>. Accessed 01 May 2016
- Room PM (1972) The constitution and natural history of the fauna of the mistletoe *Tapinanthus bangwensis* (Engl. & K. Krause) growing on cocoa in Ghana. *J Anim Ecol* 41:519–535
- Samways MJ (1994) *Insect conservation biology*. Chapman and Hall, London
- Schoener TW (1976) Species area relationship within archipelagoes: model and evidence from island birds. *Proc XVI Int Ornithol Congr* 6:629–642

- Schoener TW (1989) Food webs from the small to the large: the Robert H. MacArthur Award Lecture. *Ecology* 70:1559–1589
- Schoonhoven LM, van Loon JJA, Dicke M (2005) *Insect-plant biology*. Oxford University, New York
- Schowalter TD (2011) *Insect ecology. An ecosystem approach*. Elsevier Academic Press, Amsterdam
- SMN, Servicio Meteorológico Nacional (2013) Normales climatológicas de México. <http://smn.cna.gob.mx/>. Accessed 01 June 2013
- Shaw D (2004) Vertical organization of canopy biota. In: Lowman MD, Rinker HB (eds) *Forest canopies*. Elsevier Academic Press, Amsterdam, pp 73–101
- Soberón J, Llorente J (1993) The use of species accumulation functions for the prediction of species richness. *Conserv Biol* 7:480–488
- Southwood TRE, Moran VC, Kennedy CEJ (1982) The richness, abundance, and biomass of the arthropod communities in trees. *J Anim Ecol* 51:635–650
- Stevens RE, Hawksworth FG (1970) Insects and mites associated with dwarf mistletoes. Research Paper RM-59. US Department of Agriculture, Forest Service, Rocky Mountain Forest and Range Experiment Station, Fort Collins, CO
- Stevens RE, Hawksworth FG (1984) Insect-dwarf mistletoe associations: an update. In: Hawksworth FG, Scharpf RF (tech. coords). *Biology of dwarf mistletoes: proceedings of the symposium; General Technical Report RM-111*. US Department of Agriculture, Forest Service, Rocky Mountain Forest and Range Experiment Station, Fort Collins, CO, pp 94–101
- Stork NE (1991) The composition of the arthropod fauna of Bornean lowland rain forest trees. *J Trop Ecol* 7:161–180
- Stork NE, Adis J, Didham RK (eds) (1997) *Canopy arthropods*. Chapman and Hall, London
- Tassone RA, Majer JD (1997) Abundance of arthropods in tree canopies of *Banksia* woodland on the Swan Coastal Plain. *J R Soc West Aust* 80:281–286
- Tovar-Sánchez E, Cano-Santana Z, Oyama K (2003) Canopy arthropod communities on Mexican oaks at sites with different disturbance regimes. *Biol Conserv* 115:79–87
- Visser S (1985) Role of the soil invertebrates in determining the composition of soil microbial communities. In: Fitter AH (ed) *Ecological Interactions in soil. Plants, microbes and animals*. Blackwell, New York, pp 279–317
- Watson DM (2001) Mistletoe-A keystone resource in forest and woodland worldwide. *Annu Rev Ecol Syst* 32:219–249
- Watson DM, Herring M (2012) Mistletoe as a keystone resource: an experimental test. *Proc Biol Sci* 279:3853–3860
- Whittaker PL (1984) The insect fauna of mistletoe (*Phoradendron tomentosum*, Loranthaceae) in Southern Texas. *Southwest Nat* 29:435–444
- Williamson M, Gaston KJ, Lonsdale WM (2001) The species-area relationship does not have an asymptote. *J Biogeogr* 28:827–830
- Yanoviak SP, Nadkarni NM (2001) Arthropod diversity in epiphytic bryophytes of a neotropical cloud forest. In: Ganeshiah KN, Shaanker RU, Bawa KS (eds), *Tropical ecosystems: structure, diversity and human welfare. Proceedings of the international conference on tropical ecosystems, Bangalore, India, 15–18 July 2001*. Oxford-IBH, pp 416–419
- Yanoviak SP, Nadkarni NM, Gering JC (2003) Arthropods in epiphytes: a diversity component that is not effectively sampled by canopy fogging. *Biodivers Conserv* 12:731–741
- Zar JH (2010) *Biostatistical analysis*. Prentice-Hall, Upper Saddle River