Letter to the Editor CULTURE OF MOSQUITO CELLS IN EAGLE'S MEDIUM

Dear Editor:

Aedes albopictus mosquito cells derived from the ATC-15 line of Singh (1967) have been extensively used in production of somatic cell mutants (Gerenday et al., 1997), expression of transfected genes (Fallon, 1989; Shotkoski and Fallon, 1994), and most recently, isolation and characterization of factors involved in the inducible immune response (Hernandez et al., 1994; Yoshiga et al., 1997). Although *Ae. albopictus* cells are often maintained in "insect" tissue culture media, such as the formulation of Mitsuhashi and Maramorosch (1964) which contains lactalbumin hydrolysate and yeastolate, the ability of mosquito cells to thrive in vertebrate tissue culture media under a standard 5% CO₂ atmosphere was noted in the original description of ATC-15 cells (Singh, 1967) and has facilitated comparative studies between mammalian and mosquito cells.

Apart from the serum component, vertebrate tissue culture media such as Eagle's medium are completely defined, a feature that facilitates development of selective conditions for recovery of somatic cell mutants and use of radioisotopes as metabolic precursors in biochemical studies. In particular, the C7-10 Ae. albopictus cell line used in current analyses was derived from cells that had been gradually adapted from the Mitsuhashi and Maramorosch (1964) medium to Eagle's (1959) vertebrate tissue culture medium supplemented with L-glutamine, nonessential amino acids, antibiotics, and 5% fetal bovine serum (Sarver and Stollar, 1977; reviewed in Fallon and Stollar, 1987). Preparation of this medium by combining Earle's salts, essential and nonessential amino acids, vitamins, glutamine, antibiotics, sodium bicarbonate, and fetal bovine serum has been detailed previously (Fallon, 1989). In this letter, we describe a simplified preparation protocol based on the commercially available powdered or liquid formulations shown in Table 1.

In a first application, we used a powdered Eagle's minimal essential medium (MEM) from Sigma Chemical Co. (St. Louis, MO) con-

TABLE 1

COMMERCIAL FORMULATIONS AND CATALOG DESCRIPTIONS OF CULTURE MEDIA

Formulation	Description	
Powder	Sigma M-0643: Eagle's Minimum Essential Medium (MEM); with Earle's salts, L-glutamine, and nonessential amino acids; without sodium bicarbonate	
Powder	GIBCO BRL #61100 (MEM with Earle's salts and L- glutamine; without sodium bicarbonate	
Liquid	GIBCO BRL #11095 (1 \times MEM with Earle's salts and L-glutamine)	
Liquid	GIBCO BRL #51200 (1 \times MEM with Earle's salts; without phenol red, without L-glutamine)	

taining Earle's salts, L-glutamine, and nonessential amino acids (Table 1) as a base. Briefly, 10 g of the powdered Sigma medium was dissolved in 875 ml of distilled water with constant stirring. To arrive at equivalent concentrations of key components, relative to the earlier formulation (Fallon, 1989), additional nonessential amino acids and glutamine, vitamins, D-glucose, antibiotics, and sodium bicarbonate (Table 2) were added, as detailed in Table 3. After adjusting the pH to 6.8 with 1 N HCl or 1 N NaOH, the volume was adjusted to 1000 mL, and the medium was sterilized by filtration and stored as 95-ml aliquots at 4° C. Prior to use, heat-inactivated fetal bovine serum was added to a final concentration of 5%. When C7-10 cells were maintained in the Sigma-based medium (Table 3), cell growth measured with a Coulter electronic cell counter was indistinguishable from that in medium prepared from components as described previously (Fallon, 1989).

TABLE 2

SOURCES AND CATALOG NUMBERS OF SUPPLEMENTS USED IN PREPARING CULTURE MEDIA

Component	Supplier		
Distilled water	_		
Sodium bicarbonate	(7.5% wt/vol; GIBCO BRL #25080)		
MEM nonessential amino acids	(10 mM = 100X; GIBCO BRL #11140)		
MEM vitamin solution	(100X; GIBCO BRL #11120)		
L-glutamine	(200 mM = 100X; GIBCO BRL #25030)		
Penicillin/Streptomycin	(10,000 U/ml; 10,000 μg/ml; GIBCO BRL #15140)		
D(+)glucose (dextrose)	(10% wt/vol; GIBCO BRL #15023; Sigma G-7021)		

TABLE 3

ADDITIONS TO MEDIA TO MAKE THEM COMPARABLE TO THOSE USED IN FALLON (1989)

	Source, catalog number, and amount			
Characteristic or ingredient	Sigma M-0643	GIBCO 61100	GIBCO 11095	GIBCO 51200
Weight or volume (prepackaged)	10 g	9.5 g	1 L	1 L
Distilled water	875 ml	875 ml		
Glucose	10 ml	10 ml	10 ml	10 ml
Glutamine	10 ml	10 ml	10 ml	20 ml
Vitamins	10 ml	10 ml	10 ml	10 ml
Nonessential amino acids	10 ml	20 ml	20 ml	20 ml
Antibiotics	10 ml	10 ml	10 mł	10 ml
Sodium bicarbonate	29.3 ml	29.3 ml	_	

As expected, comparable formulations of powdered or liquid media from GIBCO BRL (Grand Island, NY; Table 1), supplemented as detailed in Table 3, also supported identical growth rates of C7-10 cells. Finally, for studies with the steroid hormone 20-hydroxyecdysone, we have begun to maintain C7-10 cells in a medium formulated without phenol red (Berthois et al., 1986) (GIBCO #51200; Table 1), and containing fetal bovine serum (heat-inactivated at 56° C for 30 min) either untreated or made steroid-free by dextran-charcoal treatment. Particularly with dextran-charcoal-treated serum, growth rates in the phenol red-free medium are reduced and frequent refeeding is required. Because this phenol red-free preparation also lacks glutamine, additional glutamine is added as described in Table 3.

Commercial preparations of culture media are cost-effective and simple to use, save time used in preparation of medium from components, and reduce the possibility of batch-to-batch variation. Because comparison of the many available formulations of vertebrate media is tedious, we anticipate that the updated information provided here will supersede our earlier publication (Fallon, 1989) and facilitate use of mosquito cells by other investigators.

ACKNOWLEDGMENTS

This work was supported by grants AI20385 and HD24869 from the National Institutes of Health and by the University of Minnesota Agricultural Experiment Station (publication # 981170005), St. Paul, MN.

References

Berthois, Y.; Katzenellenbogen, J. A.; Katzenellenbogen, B. S. Phenol red in tissue culture media is a weak estrogen: implications concerning the study of estrogen-responsive cells in culture. Proc. Natl. Acad. Sci. USA 83:2496–2500; 1986.

- Eagle, H. Amino acid metabolism in mammalian cell cultures. Science 130:432–437; 1959.
- Fallon, A. M. Optimization of gene transfer in cultured insect cells. J. Tissue Cult. Methods 12:1-6; 1989.
- Fallon, A. M.; Stollar, V. The biochemistry and genetics of mosquito cells in culture. In: Maramorosch, K., ed. Advances in cell culture. Vol. 5. New York: Academic Press; 1987:97-137.
- Gerenday, A.; Shih, K. M.; Herman, C. C., et al. Increased ribonucleotide reductase activity in hydroxyurea-resistant mosquito cells. Arch. Insect Biochem. Physiol. 34:31-41; 1997.
- Hernandez, V. P.; Gerenday, A.; Fallon, A. M. Secretion of an inducible cecropin-like activity by cultured mosquito cells. Am. J. Trop. Med. Hyg. 50:440-447; 1994.
- Mitsuhashi, J.; Maramorosch, K. Leafhopper tissue culture: embryonic, nymphal and imaginal tissues from aseptic insects. Contrib. Boyce Thompson Inst. 22:435-460; 1964.
- Sarver, N.; Stollar, V. Sindbis virus-induced cytopathic effect in clones of Aedes albopictus (Singh) cells. Virology 80:390-400; 1977.
- Shotkoski, F. A.; Fallon, A. M. Expression of an antisense dihydrofolate reductase transcript in transfected mosquito cells: effects on growth and plating efficiency. Am. J. Trop. Med. Hyg. 50:433-439; 1994.
- Singh, K. R. P. Cell cultures derived from Aedes albopictus (Skuse) and Aedes aegypti (L.). Curr. Sci. 36:506-508; 1967.
- Yoshiga, T.; Hernandez, V. P.; Fallon, A. M., et al. Mosquito transferrin, an acute phase protein that is up-regulated upon infection. Proc. Natl. Acad. Sci. USA 94:12337–12342; 1997.

Ann M. Fallon¹

Karen M. Shih

Anna Gerenday

Department of Entomology University of Minnesota 1980 Folwell Avenue St. Paul, Minnesota 55108

(Received 16 March 1998)

^{&#}x27;To whom correspondence should be addressed.