

Simple non-enzymatic electrochemical sensor for hydrogen peroxide based on nafion/platinum nanoparticles/reduced graphene oxide nanocomposite modified glassy carbon electrode

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Received: 9 August 2016 / Revised: 4 December 2016 / Accepted: 18 December 2016 / Published online: 3 January 2017
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Abstract A facile and effective strategy to fabricate non-enzymatic H₂O₂ sensor was developed based on Nafion/Platinum nanoparticles/reduced graphene oxide (Nafion/Pt NPs/RGO) nanocomposite modified glassy carbon (GC) electrode. The morphology of Nafion/Pt NPs/RGO nanocomposite was characterized by transmission electron microscopy (TEM), energy-dispersive X-ray spectroscopy (EDX) analyzer, Fourier transform infrared spectrum (FT-IR), and X-ray diffraction (XRD) spectrum respectively. The electrochemical properties of the prepared H₂O₂ sensor were evaluated by cyclic voltammetry and chronoamperometry. The prepared H₂O₂ sensor exhibited excellent electroreduction activity toward H₂O₂ with a wide linear range of 0.005–3 mM, a remarkable sensitivity of 132.8 μA mM⁻¹ cm⁻², and a low detection limit of 0.4 μM (S/N = 3). In addition, it showed good selectivity, reproducibility, and long-term stability. The excellent performance of the sensor might be attributed to the synergic effect of nanohybrids. These favorable results indicated that the prepared Nafion/Pt NPs/RGO nanocomposite is promising for fabricating non-enzymatic H₂O₂ sensor.

Keywords Graphene · Pt nanoparticles · Nafion · Non-enzymatic · Hydrogen peroxide sensor

Introduction

Hydrogen peroxide (H₂O₂) is an essential intermediate in environmental and biological reactions. It is very important to reliable and fast determination of H₂O₂ in many areas such as medicine, food control, and industrial and environmental analysis [1, 2]. Hence, the study of reliable and fast determination of H₂O₂ has attracted extensive attention. Many analytical techniques have been developed for the determination of H₂O₂ such as fluorescence [3], chemiluminescence [4], spectrophotometry [5], high-performance liquid chromatography [6], electrochemical approaches [7, 8], etc. Because of the redox behavior of H₂O₂, the electrochemical method has received a significant interest in the determination of H₂O₂ over other techniques due to its simplicity, high sensitivity, selectivity, and compatibility toward miniaturization [9]. Among the electrochemical H₂O₂ sensors, the enzyme-modified electrodes are frequently used to detect H₂O₂ with satisfactory sensitivity. However, the enzyme-modified sensors have some inevitable drawbacks such as the complex fabrication procedure, limited lifetime, stability problem, and high cost of the enzymes. Thus, many efforts have been devoted to the non-enzymatic H₂O₂ sensors based on functional nanocomposites [10–12]. The development of a high sensitivity and good selectivity catalyst for non-enzymatic H₂O₂ detection is still highly desirable in this field.

Graphene, a monolayer of carbon atoms closely packed into honeycomb two-dimensional carbon material, has received considerable attentions, owing to its large surface-to-volume ratio, high electrical conductivity, chemical stability, and excellent electronic properties [13–16]. In

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Electronic supplementary material The online version of this article (doi:10.1007/s11581-016-1944-2) contains supplementary material, which is available to authorized users.

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electrochemical sensing, graphene has been particularly used as an electrode material, owing to its excellent electrical conductivity, high electron transfer rate, and enormous electroactive area. Graphene is usually obtained by various techniques, such as mechanical exfoliation, chemical vapor deposition, epitaxial growth, and liquid phase exfoliation [17, 18]. Among these techniques, the chemical reduction of graphene oxide (GO) is an effective method for preparing graphene [19]. Moreover, it is also a facile and effective strategy to fabricate graphene-based materials because of simplicity, low cost, and easy scale up. However, graphene agglomerates easily due to Van der Waals force, which limits its application [20]. Thus, it is necessary to select suitable biopolymers to improve the dispersion of graphene in aqueous solution.

Nafion, composed of mainly hydrophobic backbone ($-\text{CF}_2$ groups) and hydrophilic chains ($-\text{SO}_3^-$ groups), has been comprehensively employed as an electrode modifier agent with excellent properties such as its antifouling capacity, chemical inertness, and high permeability to cations [21]. Furthermore, it is easy to functionalize carbon nanomaterials based on supramolecular assembly by adding Nafion into alcoholic solutions of carbon nanomaterials [22]. Nafion could effectively improve the hydrophilicity and solubility of graphene in aqueous solution [23]. The perfluorocarbon chains in Nafion could maintain the hydrophobicity of graphene and improve its dispersity. Also, the sulphonic groups could prevent the stacking of graphene layers [24].

Recently, graphene-based materials have been deeply investigated as novel electrode modification materials for the improvement of sensing sensitivity because of π - π stacking and synergetic effects with other materials. Recently, platinum nanoparticles (Pt NPs) have been attached an enormous amount of interests for their high electron transfer rate and excellent electrocatalytic activities toward H_2O_2 [25–27]. What's more, Pt NPs exhibit the property of validly lessening the oxidation/reduction overvoltage in the electrochemical detection of H_2O_2 , which can effectively avoid the interference from other co-existing substances [28]. Hence, it has become very popular in using Pt nanomaterials to fabricate H_2O_2 sensors. Furthermore, the introduction of metal nanoparticles into the dispersion of graphene sheets can inhibit the aggregation of graphene sheets, result in mechanically jammed, and exfoliate graphene agglomerate with very high surface area [29].

The aim of this work is to develop a facile and effective strategy to fabricate non-enzymatic H_2O_2 electrochemical sensor. Nafion/Pt NPs/RGO nanocomposite was prepared by a facile, eco-friendly, and controllable route. Since Nafion is composed of mainly hydrophobic and hydrophilic chains, it was used to effectively disperse RGO/Pt NPs hybrids in aqueous solution and to enhance the stability of the modified electrodes. A novel non-enzymatic electrochemical sensor for direct analytical detection of H_2O_2 was successfully papered

based on Nafion/Pt NPs/RGO nanocomposite modified glassy carbon electrode. The electrochemical properties were evaluated by cyclic voltammetry and chronoamperometry. The resulting electrode showed excellent electrocatalytic activity toward H_2O_2 .

Experimental

Reagents and materials

Graphene oxide (GO) was purchased from Nanjing XFNANO Materials Tech Co. Potassium Hexachloroplatinate (K_2PtCl_6), and Nafion was obtained from Sigma-Aldrich Co. Sodium borohydride (NaBH_4) and hydrogen peroxide (H_2O_2 , 30%, *v/v* aqueous solution) were purchased from Tianjin Chemical Factory (China). Freshly prepared 0.1 M phosphate buffer solution, consisting of Na_2HPO_4 and NaH_2PO_4 , was used as the supporting electrolyte. Na_2HPO_4 and NaH_2PO_4 were purchased from Tianjin Damao Chemical Reagent Co. (China). All aqueous solutions were prepared with doubly distilled water.

Apparatus

Electrochemical experiments were performed on a 283 Potentiostat-Galvanostat electrochemical workstation (EG&G PARC with M 270 software) with a conventional three-electrode system with the modified electrode as the working electrode, an Ag/AgCl electrode (saturated with KCl) as the reference electrode, and a platinum wire (1 mm diameter) as the counter electrode.

Nanocomposites were characterized by transmission electron microscopy (TEM, Tecnai G^2 F 20 instrument, Philips Holland), energy-dispersive X-ray spectroscopy analyzer (EDX, which was equipped on the Tecnai G^2 F 20 instrument), Fourier transform infrared spectrum (FT-IR, TENSOR 37, Bruker, German), and X-ray diffraction analysis (XRD, Rigaku, Japan).

Preparation of Nafion/Pt NPs/RGO

A total of 20 mg of GO and 20 mL of doubly distilled water were mixed through sonication for 2 h to obtain GO suspension solution (1 mg/mL). Subsequently, 10 μL of K_2PtCl_6 solution (6 mM) was added to GO suspension with sonication for 30 min. Afterwards, fresh NaBH_4 solution was added dropwise into the above suspension solution and kept stirring at room temperature for 24 h. Finally, Pt NPs/RGO was obtained through centrifugation and washed several times. Afterwards, the products were dried and resuspended in doubly distilled water at 1 mg/mL. Finally, 1 mL Nafion alcohol

solution (5 wt%) was added with sonication for 1 h to obtain Nafion/Pt NPs/RGO suspension solution.

Preparation of modified electrodes

Prior to experiment, the GC electrode was polished with 0.3 and 0.05 μm α -alumina powder sequentially, followed by ultrasonically cleaning in doubly distilled water and ethanol for 5 min, respectively. Afterwards, 6 μL of Nafion/Pt NPs/RGO suspension solution was immobilized on the surface of the GC electrode and dried naturally. The Nafion/Pt NPs/RGO modified GC electrode was directly used as a non-enzymatic H_2O_2 sensor for the determination of H_2O_2 .

Results and discussion

Characterization of Nafion/Pt NPs/RGO

Figure 1 shows the typical TEM images of the Nafion/RGO and Nafion/Pt NPs/RGO. As shown in Fig. 1a and Fig. S1, a typical wrinkled RGO sheet (Fig. S1) can be seen, and RGO sheets were compactly coated by polymer and exfoliated completely by Nafion (Fig. 1a). As shown in Fig. 1b, Pt NPs were well scattered on the surface of RGO sheet with very few aggregation, indicating that RGO made an outstanding contribution to promoting the uniform distribution of Pt NPs. Highly dispersed Pt NPs on RGO supports with larger surface

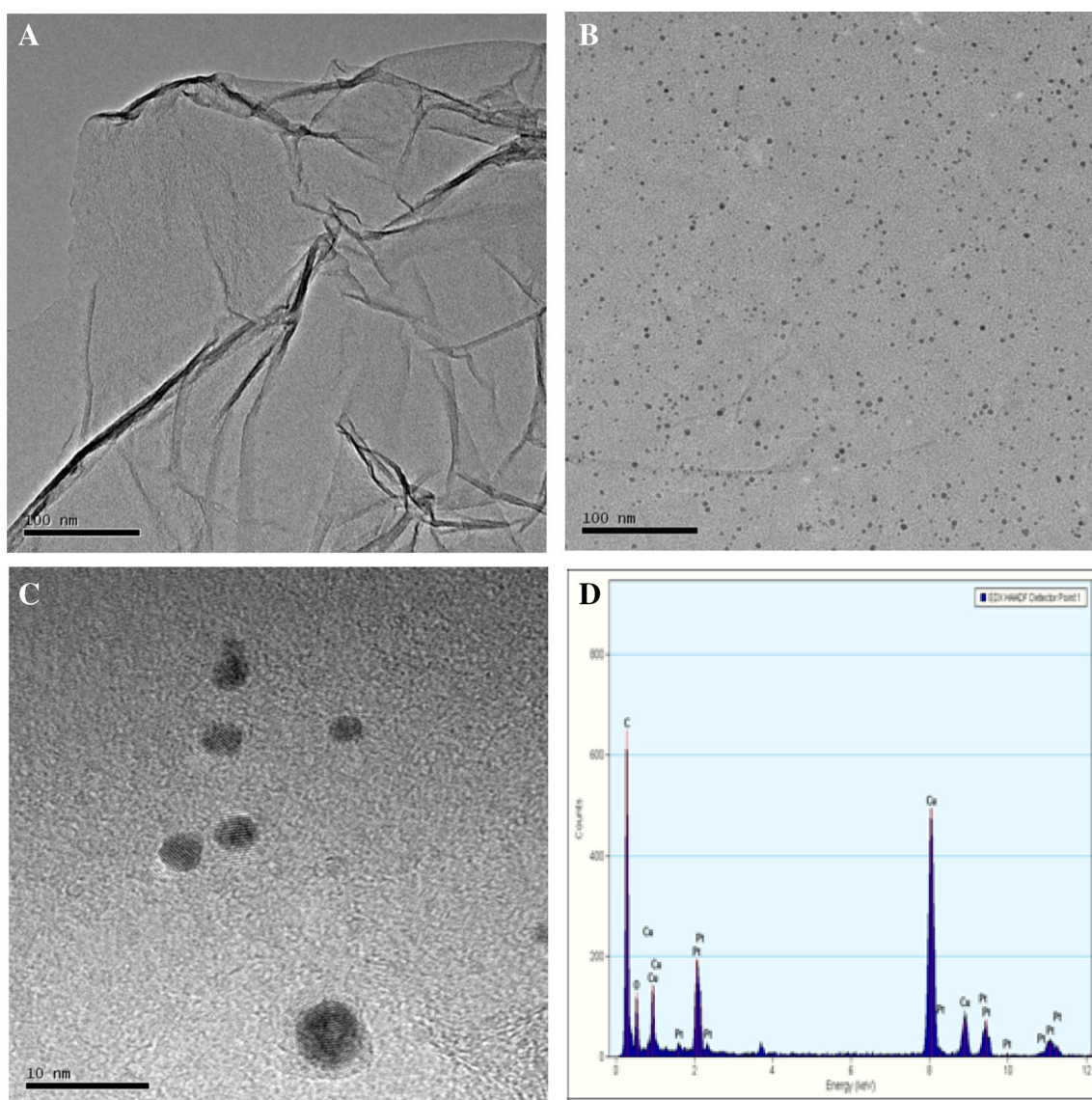


Fig. 1 TEM images of the Nafion/RGO (a) and Nafion/Pt NPs/RGO under different magnification (b and c); EDX spectrum of Pt NPs/RGO nanocomposite (d)

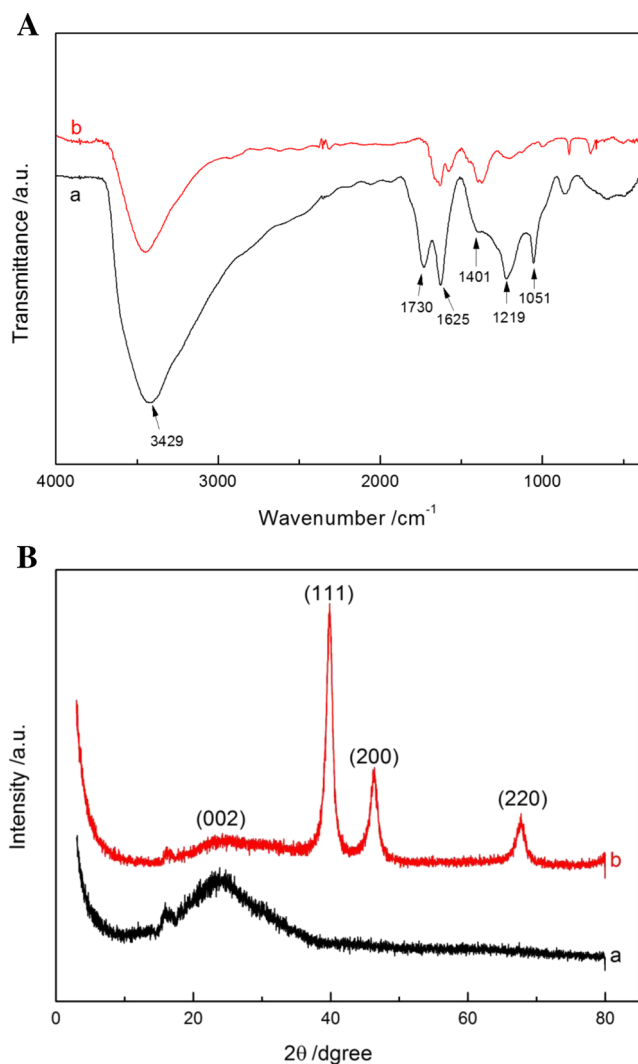


Fig. 2 **a** FT-IR spectra of GO (*a*) and RGO/Pt NPs (*b*). **b** XRD patterns of Nafion/RGO (*a*) and Nafion/Pt NPs/RGO/GC (*b*)

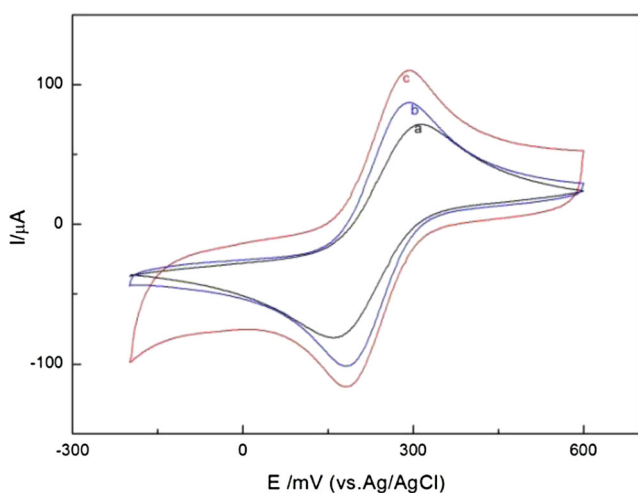


Fig. 3 CVs of bare GCE (*a*), Nafion/RGO/GC electrode (*b*), and Nafion/Pt NPs/RGO/GC electrode (*c*) recorded in 0.1 M KCl solution containing 10 mM $[\text{Fe}(\text{CN})_6]^{3-}$. Scan rate: 50 mV s^{-1}

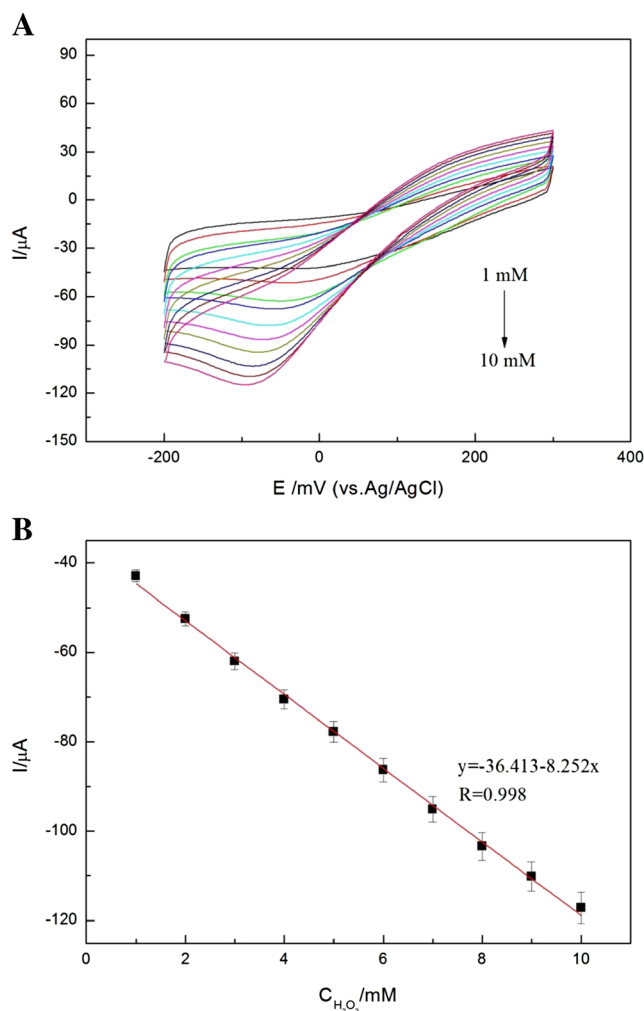


Fig. 4 **a** CVs of Nafion/Pt NPs/RGO/GC electrode in 0.1 M PBS (pH 7.0) at the scan rate 50 mV s^{-1} with different concentrations of H_2O_2 (from 1 to 10 mM). **b** The plot of the cathodic peak current values against the concentration

areas have many advantages in catalytic activity and sensor sensitivity [30]. The lattice structure of Pt NPs could be observed clearly under the high magnification. As shown in Fig. 1d, the EDX analysis further revealed that the nanocomposite was consisted of C, O, and Pt elements, and the Cu element in the spectrum should be from the substrate, which confirmed that Pt NPs had been coated on the RGO sheets.

FT-IR spectra further clarify the successful synthesis of Pt NPs/RGO. As shown in Fig. 2a (curve a), some peaks at 3429, 1730, 1625, 1401, 1219, and 1051 cm^{-1} can be seen, owing to O–H stretching vibration, carbonyl C=O stretching, aromatic C=C stretching, C–OH stretching, C–O–C stretching, and alkoxy C–O stretching [31]. However, in curve b of Pt NPs/RGO, the peak intensities at 3429, 1219, and 1055 cm^{-1}

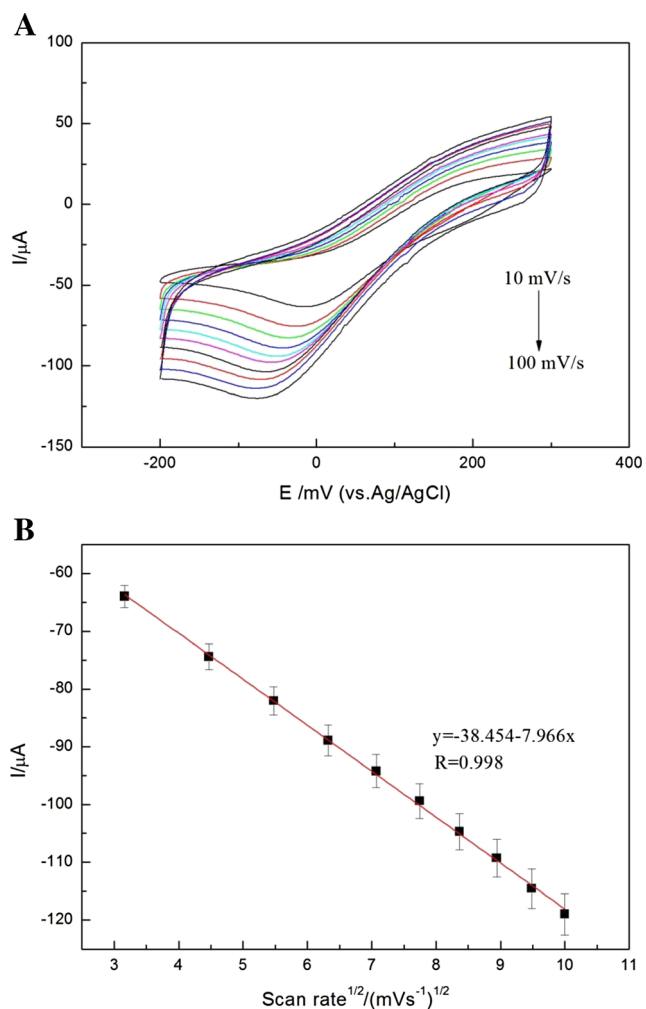


Fig. 5 **a** CVs of the Nafion/Pt NPs/RGO/GC electrode 0.1 M PBS (pH 7.0) with 5 mM H₂O₂ at different scan rate: 10, 20, 30, 40, 50, 60, 70, 80, 90, and 100 mV s⁻¹. **b** The plot of the cathodic peak current values against the scan rate

dramatically decreased after the reduction of GO. These intensities markedly decreased or even disappeared, confirming the successful reduction of GO to RGO [32]. The typical C=O absorption bands at 1730 cm⁻¹ almost disappeared, indicating that the carbonyl groups on the surface of GO sheets were modified by Pt NPs. This result further confirmed that Pt NPs/RGO nanocomposite had been synthesized successfully.

Figure 2b exhibits the XRD pattern of Nafion/RGO (curve a) and Nafion/Pt NPs/RGO (curve b). A characteristic diffraction peak at 16.9° was assigned to the Nafion [23, 33], and the peak at 23.99° (002) was the typical diffraction peak of RGO, owing to the reduction of GO [34], which indicated that Nafion/RGO was successfully acquired. In curve b, the strong peaks at 39.8°, 46.3°, and 67.8° were attributed to the characteristic (111), (200), and (220) crystals of the Pt [35, 36]. Moreover, the characteristic diffraction peaks of Nafion

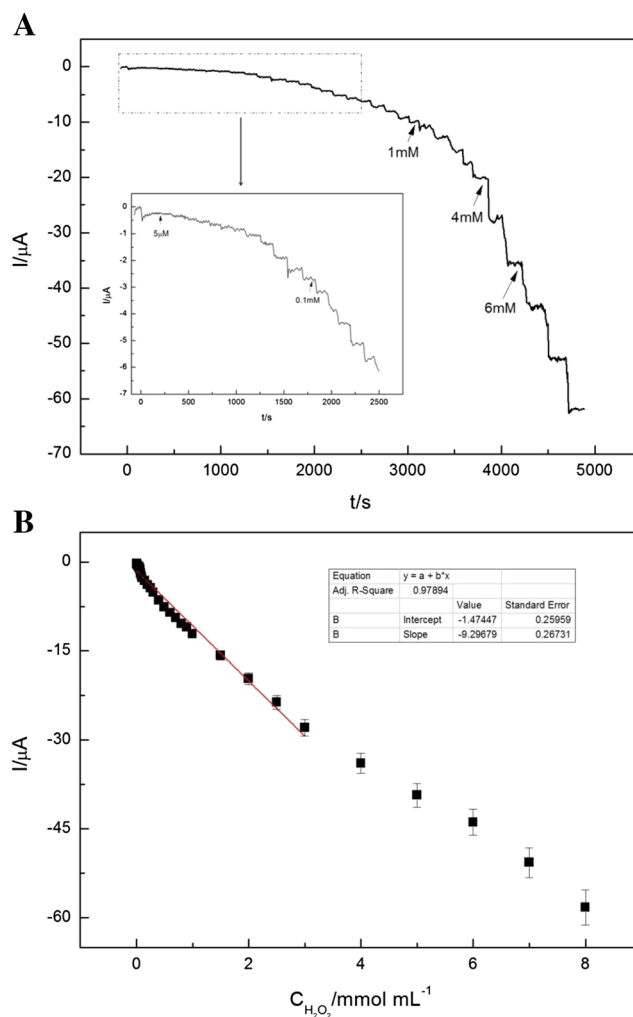


Fig. 6 **a** Amperometric response of Nafion/Pt NPs/RGO/GC electrode upon successive additions of H₂O₂ into 0.1 M PBS solution under stirring. The inset was the amplification of curve with the lower concentration region, Scan rate: 50 mV s⁻¹. **b** The calibration curve between amperometric response and H₂O₂ concentration

and RGO were also observed, indicating that Nafion/Pt NPs/RGO nanocomposite had been successfully prepared.

Electrochemical characterization of Nafion/Pt NPs/RGO/GC electrode

Cyclic voltammetry (CV) of bare GC electrode (a), Nafion/RGO/GC electrode (b), and Nafion/Pt NPs/RGO/GC electrode (c) recorded in 0.1 M KCl solution containing 10 mM [Fe(CN)₆]³⁻ are shown in Fig. 3. As shown in curve c, there are a couple of well-defined redox peaks of Nafion/Pt NPs/RGO/GC electrode at 290 and 190 mV, indicating Nafion/Pt NPs/RGO/GC electrode has a fast electron transfer rate. By the calculation, the electroactive surface area of Nafion/Pt NPs/RGO/GC electrode

Table 1 Comparison of Pt-based electrodes response to H₂O₂

Electrode	Sensitivity (μA mM ⁻¹ cm ⁻²)	LOD (μM)	linear range (μM)	Reference
PVA-MWCNTs-PtNPs	122.63	0.7	2–3800	[28]
Pt-PPy-GC	80.4	1.2	1000–8000	[38]
PtPNW@C nanoparticles	6.28	0.002	0.01–1.2	[39]
PtNP-TiO ₂	1.68	4	4–1250	[40]
Pt/rGO–CNT paper NPs	1.41	0.01	0.1~25	[41]
Graphene-Pt nanocomposite	–	0.5	2~710	[42]
Pt NP-PAni	96.1	0.7	10–8000	[43]
Pt-DENs/GOx/Pt-DENs/PAni/PSS	39.63	0.5	10–4500	[44]
Nafion/Pt NPs/RGO	132.8	0.4	5–3000	This work

RGO/GC electrode was about 1.26 and 1.55 times higher than those of Nafion/RGO/GC electrode and bare

GC electrode, respectively, according to the Randles-Sevcik eq. [37]:

$$I_p = 2.69 \times 10^5 AD^{1/2} n^{3/2} \nu^{1/2} C \quad (1)$$

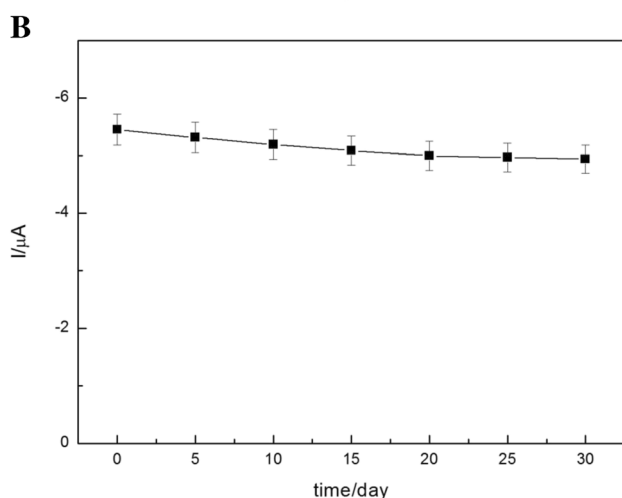
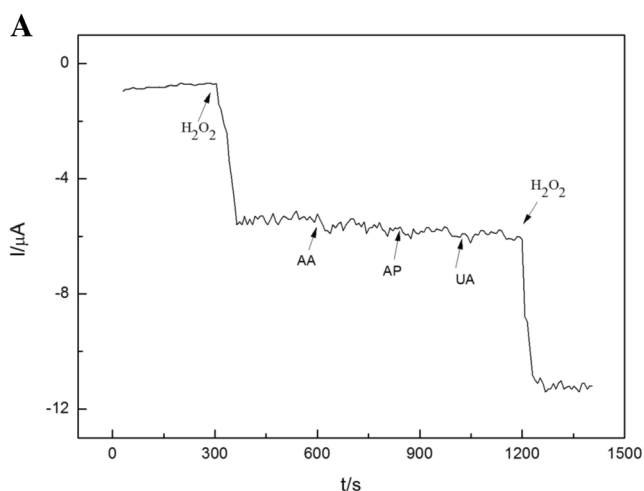


Fig. 7 a Amperometric response of Nafion/Pt NPs/RGO/GC electrode upon successive addition of 0.5 mM H₂O₂, 0.1 mM ascorbic acid (AA), 0.1 mM acetaminophen (AP) 0.1 mM uric acid (UA) and 0.5 mM H₂O₂. b The long-term stability of the non-enzymatic H₂O₂ sensor

Where I_p represents the redox peak current, A is the electrode's electroactive surface area, D is the diffusion coefficient of the molecule in solution which is $(6.70 \pm 0.02) \times 10^{-6} \text{ cm}^2 \text{ s}^{-1}$, n is the number of electron participating in the reaction which is equal to 1, ν is the scan rate (V s^{-1}), and C is the concentration of the probe molecule in the solution. These excellent results indicate that the prepared Nafion/Pt NPs/RGO nanocomposite is undeniably suitable as an electron transfer mediator between $[\text{Fe}(\text{CN})_6]^{3-}$ and the GC electrode, which should be attributed to the large edge plane/basal plane ration and high electrical conductivity of RGO and Pt NPs. Moreover, there may be a synergistic effect between RGO and Pt NPs, which can facilitate the electronic transfer.

Cyclic voltammetric behavior of Nafion/Pt NPs/RGO/GC electrode to H₂O₂

Figure 4a shows the CVs of Nafion/Pt NPs/RGO/GC electrode in 0.1 M PBS (pH 7.0) at the scan rate 50 mV s^{-1} with different concentrations of H₂O₂ (from 1 to 10 mM). It can be seen that the cathodic peak current gradually increased with the increase of H₂O₂ concentration. As shown in Fig. 4b, the cathodic peak current was linearly proportional to the concentration of H₂O₂ ($R^2 = 0.998$), indicating that it was possible to construct an electrochemical sensor which would behave well in the amperometric experiments.

CVs of Nafion/Pt NPs/RGO/GC electrode in 0.1 M PBS (pH 7.0) containing 5 mM H₂O₂ with varying the scan rates are shown in Fig. 5a. It can be found that the cathodic peak current increased as the scan rate increased from 10 to 100 mV s^{-1} , while the cathodic peak potential shifted to a

more negative region. As shown in Fig. 5b, there was a good linearity between the cathodic peak current and the scan rate ($R^2 = 0.998$), indicating the electrochemical process was diffusion controlled.

Amperometric response of Nafion/Pt NPs/RGO/GC electrode to H₂O₂

Amperometric responses of Nafion/Pt NPs/RGO/GC electrode upon successive addition of H₂O₂ into 0.1 M PBS under optimum conditions (Optimization of experimental parameter is presented in Supplementary material) are shown in Fig. 6a. It can be clearly seen that the Nafion/Pt NPs/RGO/GC electrode showed a rapid and stable response to the addition of H₂O₂, and the time of current response before it reached 95% of steady-state current was less than 6 s. The calibration curves are shown in Fig. 6b, and the regression equation is $y = -1.474 - 9.297x$ ($R = 0.979$). We found that the prepared non-enzymatic H₂O₂ sensor exhibited wide linear range from 0.005 to 3 mM with remarkable sensitivity of 132.8 $\mu\text{A mM}^{-1} \text{cm}^{-2}$, and the detection limit was calculated from $3 \times$ blank variance/slope found to be 0.4 μM at the signal-to-noise ratio of 3. In Table 1, we also compared the characteristics of the prepared non-enzymatic sensor with other relevant sensors collected from the literature. It can be seen that the performances of the Nafion/Pt NPs/RGO/GC electrode were obviously better than previously reported sensors. These excellent performances further revealed that Nafion/Pt NPs/RGO nanocomposites are promising modified electrode materials in H₂O₂ detection. These excellent performances of the Nafion/Pt NPs/RGO/GC electrode may be attributed to the synergistic effect between RGO and Pt NPs, which promotes large amount of H₂O₂ adsorbing on the electrode surface, accelerates the electron transfer rate between the surface of GC electrode and H₂O₂, and increases the electrocatalytic active area.

Selectivity, reproducibility, and stability of the modified electrode

The selectivity was studied by comparing the amperometric responses to successive addition of 0.5 mM H₂O₂, 0.1 mM ascorbic acid (AA), 0.1 mM acetaminophen (AP), 0.1 mM

uric acid (UA), and 0.5 mM H₂O₂ in 10 mL of 0.1 M PBS (pH 7.0) at the applied potential of -50 mV. As shown in Fig. 7a, the response current increased obviously after addition of H₂O₂, and no obvious changed after the addition of AA, AP, and UA respectively, while the current obviously increased again after H₂O₂ was second added, indicating Nafion/Pt NPs/RGO/GC electrode showed good selectivity toward H₂O₂ and the responses caused by AA, AP and UA could be negligible. The reproducibility was assessed by detecting the responses to 0.5 mM H₂O₂ at five electrodes independently, and the relative standard deviation was calculated to 3.2%. Furthermore, the long-term stability of the prepared H₂O₂ sensor was also evaluated by measuring the response current to 0.5 mM H₂O₂ solution every day for 1 month. The electrode remained 90.6% of its initial current response for H₂O₂ after 1 month (Fig. 7b). Thus, these results indicated the prepared H₂O₂ sensor exhibited an acceptable reproducibility and long-term stability.

Real sample analysis

In order to evaluate the practical applications of the prepared non-enzymatic H₂O₂ sensor, the Nafion/Pt NPs/RGO/GC electrode was investigated to detect H₂O₂ in disinfected fetal bovine serum (FBS). As shown in Table 2, the recovery was in the range of 95.00–105.00% and RSD ranged from 3.29 to 4.12%, indicating that the H₂O₂ sensor developed in this work showed potential applicability to real samples analysis.

Conclusion

In summary, we have developed a facile and effective route to prepare prepared non-enzymatic H₂O₂ sensor based on Nafion/Pt NPs/RGO nanocomposite via one-pot method. Since Nafion is composed of mainly hydrophobic and hydrophilic chains, it could be used as stabilizer to effectively disperse RGO/Pt NPs hybrids in aqueous solution. As a sensor, the data obtained from the electrochemical experiments showed that the performances of the fabricated sensor were suitable for quantitative detection of H₂O₂ with high sensitivity, wide linear range, and low detection limit, which were much better than many other H₂O₂ sensors previously

Table 2 Determination of H₂O₂ concentration in FBS samples ($n = 6$)

Sample	H ₂ O ₂ added (mM)	H ₂ O ₂ found (mM)	Recovery (%)	R.S.D. (%)
1	0.20	0.21	105.00	3.41
2	0.40	0.38	95.00	3.29
3	0.60	0.61	101.67	4.12
4	0.80	0.81	101.25	3.67
5	1.00	0.99	99.00	3.89

reported. Furthermore, the prepared non-enzymatic H_2O_2 sensor could be also used in the detection of H_2O_2 in disinfected fetal bovine serum with an acceptable RSD. With simple synthetic method, using of low-cost materials, and excellent electrocatalytic activity, there is no doubt that Nafion/Pt NPs/RGO nanocomposite has broad prospect in fabricating high performance sensors.

Acknowledgements This work is supported by the Natural Science Foundation of China (Grant Nos. 81127001 and 81273993).

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