

# Extracted magnetic resonance texture features discriminate between phenotypes and are associated with overall survival in glioblastoma multiforme patients

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**Abstract** GBM is a markedly heterogeneous brain tumor consisting of three main volumetric phenotypes identifiable on magnetic resonance imaging: necrosis (vN), active tumor (vAT), and edema/invasion (vE). The goal of this study is to identify the three glioblastoma multiforme (GBM) phenotypes using a texture-based gray-level co-occurrence matrix (GLCM) approach and determine whether the texture features of phenotypes are related to patient survival. MR imaging data in 40 GBM patients were analyzed. Phenotypes vN, vAT, and vE were segmented in a preprocessing step using 3D Slicer for rigid registration by T1-weighted imaging and corresponding fluid attenuation inversion recovery images. The GBM phenotypes were segmented using 3D Slicer tools. Texture features were extracted from GLCM of GBM phenotypes. Thereafter, Kruskal–Wallis test was employed to select the significant features. Robust predictive GBM features were identified and underwent numerous classifier analyses to distinguish phenotypes. Kaplan–Meier analysis was also performed to determine the relationship, if any, between phenotype texture features and survival rate. The simulation results showed that the 22 texture features were significant with  $p$  value  $<0.05$ . GBM phenotype discrimination based on texture features showed the best accuracy, sensitivity, and specificity of 79.31, 91.67, and 98.75 %, respectively. Three texture features derived from active tumor

parts: difference entropy, information measure of correlation, and inverse difference were statistically significant in the prediction of survival, with log-rank  $p$  values of 0.001, 0.001, and 0.008, respectively. Among 22 features examined, three texture features have the ability to predict overall survival for GBM patients demonstrating the utility of GLCM analyses in both the diagnosis and prognosis of this patient population.

**Keywords** GLCM · Glioblastoma · Survival · Texture · Feature

## 1 Introduction

Glioblastoma multiforme (GBM) is the most common and most aggressive primary brain malignancy in adults [39]. The inability to perform complete surgical tumor resection and poor drug delivery to the brain contributes notably to the lack of effective treatment and poor prognosis. Generally, tumors such as GBM are heterogeneous with intratumoral spatial variation in both cellularity and areas of necrosis [4]. Tumors with high intratumoral heterogeneity have been shown to have a poorer prognosis likely secondary to their intrinsically aggressive biology [38]. Tumors are classified into four grades of glioma, namely grade 1 (juvenile pilocytic astrocytoma; best prognosis), grade 2 (low-grade glioma), grade 3 (anaplastic astrocytoma), and grade 4 (glioblastoma: the most aggressive type). It is mainly located in the cerebral hemispheres, and the average patient survival with GBM is around 14.6 months [30, 39].

Given the poor patient survival in GBM, computer-aided detection techniques may play a role in early, accurate diagnosis. To this end, brain tumor diagnosis using the image processing and analysis has been an area of increased

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research interest in recent years allowing for partial automation of GBM identification [45]. This identification is based on the tumor heterogeneity which is a pattern feature of malignancy that represents areas of high cell density or GBM phenotypes such as: active tumor and edema parts, with low cell density of necrotic parts [6, 40, 43]. These phenotypes can be segmented manually or semiautomatically by image processing tools applied to MR images. Moreover, they can specifically be identified as active tumor (vAT) and necrosis (vN) using T1-weighted imaging (T1–WI) images and edema (vE) by fluid attenuation inversion recovery (FLAIR) sequences [12]. However, not all the parts of phenotypes can be clearly identified even though this identification is clinically and prognostically useful. Therefore, it is important to assess the GBM heterogeneity by quantifying its phenotypes and this is represented by the employment of many feature types including Gaussian mixture model (GMM) features [8], histogram-based features [17], wavelet-based features [28], and texture-based features [3, 5, 7, 28].

GMM-based features have been successfully employed to classify the normal from abnormal brain heterogeneity [13]. Similarly, histogram-based features have shown that GBM has two Gaussian distributions in FLAIR sequence which are represented by vAT and vE phenotypes. Among nine statistical features for classifying vAT and vE, kurtosis and skewness have shown a highest range of 58.33–75.00 % accuracy classifier [9]. Wavelet-based features have been used to extract space–frequency textures in order to predict MGMT gene methylation status in GBM [28]. Texture features based on gray-level co-occurrence matrix (GLCM) have been employed to differentiate between pathological and healthy tissue in different organs. Thus, texture features based on the GLCM have been extracted and can evaluate the relationships of gray-level intensity in the image by second-order statistics. In this context, GLCM-based textures can compute the gray-level intensity within an image and provide additional descriptors of tumor heterogeneity [7]. These feature types can enhance standard reporting techniques and help to more accurately characterize tumor heterogeneity [15].

In addition to their diagnostic utility, imaging features have also been associated with survival in GBM. For example, contrast-enhanced MR image features provided novel prognostic information and were accurate in predicting survival times in patients with advanced gliomas [34]. Additionally, two newer studies have been considered a standardized lexicon of imaging features derived from MRI [known Visually Accessible Rembrandt Images (VASARI)] which have been demonstrated to be feasible predictors of survival [21, 29]. New texture analysis of GBM phenotypes with texture-based survival prediction could be useful in clinical practice. Recently, it showed that the texture feature

ratios from contrast-enhancing, non-enhancing lesions and kinetic texture analysis obtained from perfusion parametric maps provide useful information for predicting survival in patients with GBM [27].

This paper focuses on GBM phenotype analysis using texture feature extraction based on GLCM from MR images. This approach further show the effect of radiomics analysis for GBM tumors using descriptors (features) that may be subsequently employed in automated glioma diagnosis with phenotype characterization, and demonstrate the feasibility of texture features to be associated with survival in GBM. They may also provide a more accurate assessment of the patient prognosis and underlying genomic composition.

## 2 Methods

### 2.1 Patients population and data acquisition

In this study, data were collected from The Cancer Imaging Archive (TCIA) (<http://cancerimagingarchive.net/>), and 40 GBM patients were used to validate the proposed method. The GBM data were acquired prior to any treatment from patients with brain tumors that were subsequently diagnosed as GBM. The GBM diagnosis was based on histological examination. These patients were visually assessed as having sufficient quality and as containing the phenotypes [necrosis parts (vN), contrast enhancement/active tumor (vAT), and edema/invasion (vE)]. For visual assessment, 3D Slicer software was used for illustrating the GBM phenotypes and testing that the patient images can be correctly registered using T1–WI and FLAIR sequence. Table 1 shows the characteristics of GBM patients by representing the average, median, minimum, and maximum of the phenotypes (vN, vAT, and vE), age, and overall survival.

The imaging protocol used whole-brain T1–WI and FLAIR scanning using a 3T MRI scanner (GE Healthcare). T1–WI scans were acquired based on the following parameters: slice thickness (ST) = 5 mm, spatial resolution (SR) = 1.04 mm, pixel spacing (PS) = 0.78 mm, repetition time (TR) = 650 ms, echo time (TE) = 9 ms, and flip angle (FA) = 90°. And, FLAIR scans were acquired using

**Table 1** Patient and tumor characteristics

	Average	Median	Minimum	Maximum
vE (mm <sup>3</sup> )	73,748	63,366	5568	199,900
vAT (mm <sup>3</sup> )	28,133	27,311	3282	57,631
vN (mm <sup>3</sup> )	12,585	11,939	587	44,737
Age (years)	57.15	56	34	81
Overall survival (days)	412.12	394.00	67.00	828.00

the following parameters: ST = 5 mm, SR = 1.24 mm, PS = 0.78 mm, TR = 10,002 ms, TE = 147 ms, FA = 90°, and acquisition time 10:24 min.

## 2.2 Data preprocessing

The entire sequences available for the patient MRI set were obtained, yet only post-contrast T1–WI and FLAIR were used for texture analyses. Moreover, from the available database 40 patients' data were randomly chosen to obtain full GBM tumor imaging MRI sets. All of the images had  $512 \times 512$  pixel acquisition matrices and were converted into grayscale before further processing. Note that the standard imaging parameters were used for each of the sequences as noted in the TCIA database. Standardization was employed by the linear normalization of each image. MRI raw data were filtered by an average filter (spatial filter) with size window of  $3 \times 3$  before further processing to minimize the effects of noise in images and other external factors. Thereafter, registration, segmentation, and texture feature extraction were employed on GBM phenotypes for data collection.

## 2.3 Registration and segmentation based on 3D Slicer

Registration obtains more information from different scan angles by using different slices and voxel thickness. It registers two corresponding scans or images and rigidly aligns them to each other to make an accurate registration. This operation affects the computation time which is a necessary obligation as we finish the registration step. The computation time changes depending on the image size and angle rotations. It can be more sophisticated with the three-dimensional data (3D), and the percentage accuracy may be decreased [18, 42]. Eventually, transformation of locations in one image to new locations in another image requires regulator parameters. The step of determining the correct transformation parameters is required during the image registration process. Since the FLAIR scans and T1–WI post-contrast scans obtained with different slice parameters, angles, and voxel thickness, we rigidly aligned and registered the scans to each other. Moreover, most of the voxel size of the FLAIR and T1–WI images was similar and was simply registered. However, in case that the voxel size was dissimilar, we resampled the FLAIR volume to the matrix of T1–WI voxel size. The patient's images which have complex rotation modifications and registrations were not considered in order to achieve an error less than 2 mm. The average of computational time necessary to complete each volume registration is 40–50 s.

Registration for each patient's data was done by using T1–WI and its corresponding FLAIR sequence using 3D Slicer (Fig. 1) [1]. Phenotypes were segmented based on

T1–WI, FLAIR, and its corresponding registration image by using the 3D Slicer (example in Fig. 1d). Histogram analysis clearly shows the difference in shape of phenotype data fitting. For example, distribution shape in the pixel intensity axis for each phenotype was not similar (Fig. 1). Moreover, phenotype was segmented manually slice-by-slice and organized in order to extract the texture features. Then, the texture features of necrosis (vN) and active tumor parts (vAT) were computed from T1–WI, while texture feature of edema/invasion (vE) was extracted from FLAIR sequence. Texture features extracted for each phenotype were employed on each axial slice using the GLCM technique.

## 2.4 Gray-level co-occurrence matrix computation

Texture feature extraction based on GLCM is a second-order texture that is based on the joint co-occurrence of gray values for pairs of pixels at a given distance  $d$  and direction  $\theta$  [10, 23]. Traditionally, the co-occurrence matrix  $P_{d,\theta}(i, j)$  of a given two-dimensional (2D) image  $I$  of size  $N \times N$  can be defined as

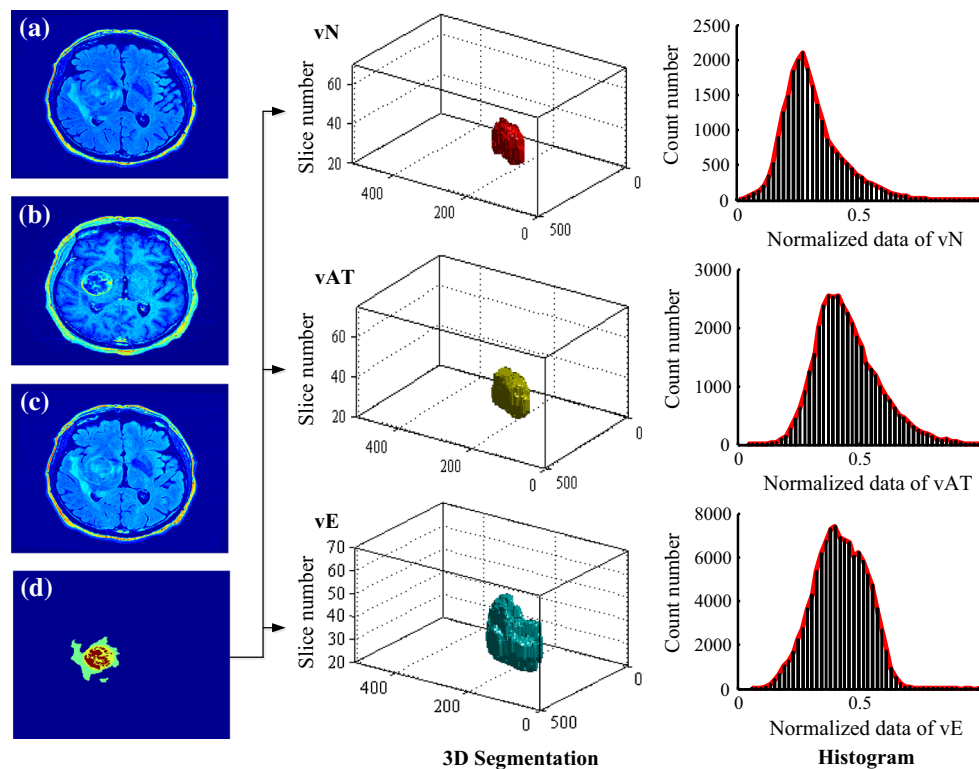
$$P_{d,\theta}(i, j) = \sum_{x=1}^N \sum_{y=1}^N \begin{cases} 1, & \text{if } I(x, y) = i \wedge I(x + dx, y + dy) = j \\ 0 & \text{otherwise} \end{cases} \quad (1)$$

where  $dx$  and  $dy$  specify the distance between the pixel of interest and its neighbor, along the  $x$  and  $y$  axis of an image, respectively. Note that the GLCM is a square matrix of size  $N_g \times N_g$ , where  $N_g$  represents the number of gray levels of the image.

For 2D images, typical values used for “ $d$ ” are {1, 2, 3, 4} and those for “ $\theta$ ” equal {0°, 45°, 90°, 135°}. The GLCMs corresponding to the additional directions {180°, 225°, 270°, 315°} added to the specification of the texture associated with combinations of the aforementioned four offsets and four directions. These additional directions were considered to cover the whole spatial relationship of offset  $d$ . In fact, 4 offsets with 4 symmetric directions may provide 16 GLCMs.

For each phenotype slice, 16 GLCMs are quantified by 22 descriptors with each slice being represented by a feature vector of 22 features. Each of these feature values is the average of 16 descriptor values.

For whole GBM tumor, a feature vector of 22 features represents the average of each descriptor in whole slice in each GBM patient. Note that these descriptors (features) have been proposed by Haralick [23] and an additional feature type based on GLCM originally described for sea ice texture analysis [37]. Table 2 shows the texture feature names and explains the mathematical formula representing the texture descriptors. All the 22 texture functions are analyzed and discussed in the result section.



**Fig. 1** Preprocessing of texturized MRI image from GBM patients. **a** T1–WI axial image, **b** corresponding axial FLAIR image, **c** corresponding axial registration, and **d** 2D segmentation (label map) of GBM phenotypes. Phenotypes segmentation: illustration of 3D seg-

mentation of phenotype vN, vAT, and vE. And, two-dimensional histogram distribution of normalized phenotypes: associated pixel histogram distribution of vN, vAT, and vE

## 2.5 Processing and analysis of features

The Kruskal–Wallis test was applied to assess the statistical significance between texture features and GBM phenotypes [31]. For this statistical test, a  $p$  value  $< 0.05$  was considered to be significant. Thus,  $z$  score normalization was employed on each of the feature vectors which converted to zero mean and unit variance. Note that the mean and standard deviation ( $\sigma$ ) of the feature vector were calculated as follows:

$$r_n = \frac{r - \text{mean}}{\sigma} \quad (2)$$

where  $r$  is the original value,  $r_n$  is the new value, and the mean and  $\sigma$  are the average and standard deviation of the original data, respectively.

## 2.6 Classifier setting

We implemented four classifier techniques, namely discriminant analysis (DA) [20], naïve Bayes (NB) [2], decision trees (DT) [35], and support vector machine (SVM) [24] using the *Statistics and Machine Learning Toolbox* in MATLAB software. The implementation of DA was performed using pseudo-inverse which is equivalent to approximating the

solution using a least-squares solution method. The implementation of NB considered a kernel estimation method which approximated the complex distributions of data.

For training based on DT classifier, two conceptual phases is considered: a “growing” phase where training examples are recursively split based on their attributes and a “pruning” phase which simplifies the tree by removing low discriminating branches of the tree. One of the most important components of decision trees is the split criterion, which selects for each node of the tree an attribute to separate examples along the branches of this node. The choice for best attribute splitting can be based on several techniques. For this work, we considered the Gini index ( $I_{G(t)}$ ) for splitting data and identify the feature subset leading to the highest accuracy.  $I_{G(t)}$  is an impurity-based criterion that measures the divergence between the probability distributions of the attribute’s values

$$I_{G(t)} = \sum_i p_i(1 - p_i) \quad (3)$$

where  $p_i$  is the relative frequency of class  $i$  at node  $t$  and node  $t$  represents any node at which a given split is performed.  $p_i$  is determined by dividing the total number of observations in the class by the total number of observations.

**Table 2** Summary of texture features for GBM phenotypes and corresponding  $p$  values

Texture parameters	Functions	Average (normalized)			$p$ value
		vN	vE	vAT	
$f_1$ : autocorrelation	$f_1 = \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} (ij) P_{d,\theta}(i, j)$	0.31	0.90	0.43	$1.21 \times 10^{-67}$
$f_2$ : contrast	$f_2 = \sum_{n=0}^{N_g-1} n^2 \left\{ \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} P_{d,\theta}(i, j) \mid i - j = n \right\}$	0.30	0.91	0.42	$1.54 \times 10^{-66}$
$f_3$ : correlation <sub>1</sub>	$f_3 = \frac{\sum_{i=1}^{N_g} \sum_{j=1}^{N_g} (i - \mu_x)(j - \mu_y) P_{d,\theta}(i, j)}{\sigma_x \sigma_y}$	0.31	0.96	0.46	$9.49 \times 10^{-69}$
$f_4$ : correlation <sub>2</sub>	$f_4 = \frac{\sum_{i=1}^{N_g} \sum_{j=1}^{N_g} (ij) P_{d,\theta}(i, j) - \mu_x \mu_y}{\sigma_x \sigma_y}$	0.29	0.92	0.42	$6.77 \times 10^{-67}$
$f_5$ : cluster prominence	$f_5 = \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} (i + j - \mu_x - \mu_y)^4 P_{d,\theta}(i, j)$	0.33	0.95	0.55	$3.5 \times 10^{-104}$
$f_6$ : cluster shade	$f_6 = \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} (i + j - \mu_x - \mu_y)^3 P_{d,\theta}(i, j)$	0.33	0.92	0.56	$4 \times 10^{-115}$
$f_7$ : dissimilarity	$f_7 = \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} \mid i - j \mid P_{d,\theta}(i, j)$	0.34	0.91	0.55	$3.54 \times 10^{-98}$
$f_8$ : energy	$f_8 = \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} (ij) P_{d,\theta}(i, j)^2$	0.32	0.93	0.56	$2 \times 10^{-115}$
$f_9$ : entropy	$f_9 = - \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} P_{d,\theta}(i, j) \log(P_{d,\theta}(i, j))$	0.50	0.90	0.64	$1 \times 10^{-51}$
$f_{10}$ : homogeneity	$f_{10} = \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} P_{d,\theta}(i, j) / (1 + \mid i - j \mid)$	0.49	0.94	0.60	$1.4 \times 10^{-52}$
$f_{11}$ : local homogeneity	$f_{11} = \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} P_{d,\theta}(i, j) / (1 + \mid i - j \mid)^2$	0.48	0.92	0.63	$2.54 \times 10^{-61}$
$f_{12}$ : maximum probability	$f_{12} = \max_{i,j} (P_{d,\theta}(i, j))$	0.47	0.97	0.63	$3.29 \times 10^{-61}$
$f_{13}$ : sum of squares: variance	$f_{13} = \sum_{i=1}^{N_g} (i - \mu_x)^2 P_x(i)$	0.3	0.98	0.46	$3.02 \times 10^{-68}$
$f_{14}$ : sum average	$f_{14} = \sum_{n=2}^{2N_g} n \left( \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} P_{d,\theta}(i, j) \right)$	0.30	0.97	0.45	$6.75 \times 10^{-68}$
$f_{15}$ : sum variance	$f_{15} = \sum_{n=2}^{2N_g} (n - f_{14})^2 \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} P_{d,\theta}(i, j)$	0.31	0.95	0.46	$1.3 \times 10^{-68}$
$f_{16}$ : sum entropy	$f_{16} = - \sum_{n=2}^{2N_g} \left( \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} P_{d,\theta}(i, j) \right) \log \left( \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} P_{d,\theta}(i, j) \right)$	0.32	0.97	0.45	$8.24 \times 10^{-68}$
$f_{17}$ : difference variance	$f_{17} = \sum_{n=0}^{N_g-1} (n - \mu_{i-j})^2 \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} P_{d,\theta}(i, j)$ where $\mu_{i-j}$ is the mean of $\sum_{i=1}^{N_g} \sum_{j=1}^{N_g} P_{d,\theta}(i, j)$	0.31	0.90	0.40	$2.4 \times 10^{-67}$
$f_{18}$ : difference entropy	$f_{18} = - \sum_{n=0}^{N_g-1} \left( \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} P_{d,\theta}(i, j) \log \left( \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} P_{d,\theta}(i, j) \right) \right)$	0.33	0.91	0.39	$8.74 \times 10^{-67}$
$f_{19}$ : information measure of correlation <sub>1</sub>	$f_{19} = \frac{f_9 + \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} \log(P_x(i) P_y(j))}{- \sum_{i=1}^{N_g} P_x(i) \log(P_x(i))}$	0.03	0.98	0.40	$6.79 \times 10^{-68}$
$f_{20}$ : information measure of correlation <sub>2</sub>	$f_{20} = \sqrt{1 - e^{\frac{2(f_9 + \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} \log(P_x(i) P_y(j))}{- \sum_{i=1}^{N_g} P_x(i) \log(P_x(i))})}}$	0.30	0.95	0.39	$9.33 \times 10^{-67}$
$f_{21}$ : inverse difference	$f_{21} = \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} \frac{P_{d,\theta}(i, j)}{1 + \mid i - j \mid}$	0.31	0.97	0.43	$4.94 \times 10^{-67}$
$f_{22}$ : inverse difference moment	$f_{22} = \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} \frac{P_{d,\theta}(i, j)}{1 + (i - j)^2}$	0.31	0.96	0.42	$1.66 \times 10^{-66}$

$\mu_x, \mu_y, \sigma_x$  and  $\sigma_y$  are the means and standard deviation of  $P_x$  and  $P_y$ , the partial probability density functions  
 $\mu_x = \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} i \cdot P_{d,\theta}(i, j)$ ;  $\mu_y = \sum_{i=1}^{N_g} \sum_{j=1}^{N_g} j \cdot P_{d,\theta}(i, j)$ ;  $\sigma_x = \sqrt{\sum_{i=1}^{N_g} \sum_{j=1}^{N_g} (1 - \mu_x)^2 \cdot P_{d,\theta}(i, j)^2 \cdot P_{d,\theta}(i, j)}$   
 $P_x(i)$  and  $P_y(j)$  are the  $i$ th entry in the marginal-probability matrix obtained by summing the rows and columns of  $P_{d,\theta}(i, j)$ , respectively



SVM function ( $c$ ) uses a model to identify support vectors ( $s_i$ ), weighted ( $\alpha_i$ ), and bias ( $b$ ) that are used to classify vectors ( $x$ ) according to the following equation

$$c \approx \sum_i \alpha_i K(s_i, x) + b \quad (4)$$

$$K = e^{(-x_1 - x_2^2)} \quad (5)$$

where  $K$  is a kernel function (radial basis function).

Note that the SVM is a binary classifier which can be extended by fusing several of its kind into a multiclass classifier [16]. For 3 classes as we have the phenotypes (vN, vAT, and vE), 3 classifiers are necessary: one SVM classifies vN from vAT and vE, a second SVM classifies vAT from vN and vE, and a third SVM classifies vE from vN and vAT. The decision then is a combination of targets of all the separate SVMs. For example, vectors from classes vN, vAT, and vE have codes (1, -1, -1), (-1, 1, -1), and (-1, -1, 1), respectively. If each of the separate SVMs classifies a phenotype correctly, it means that no error for this phenotype classifier. However, if at least one of the SVMs misclassifies the phenotype, the class selected for this phenotype is the one its target code closest in the Hamming distance sense to the actual output code and this may be an erroneous decision. The reason to use these specific classifier methods is to achieve the trade-off performance metrics.

The low number of patient samples was a limitation to prove the technical method and the medical analysis. In this case, leave-one-out cross-validation can be a solution to evaluate the performance metric by the swapping test and training sample data [36]. In this way, all of the GBM phenotypes were used for both training and testing. Thus, we computed the classifier accuracy, sensitivity, and specificity to evaluate the classifier feature. In addition, receiver operating characteristic (ROC) curve was employed which provides the true positive versus (vs) false positive rates. ROC curves are widely employed to evaluate the performance of a medical test or model and are associated with area under the curve (AUC). An AUC value close to 1 shows better classification performance [22, 33].

Moreover, survival curves were plotted by Kaplan–Meier method and compared by the log-rank test [25]. We considered the median of each feature to grouping the patients in two groups based on cutoff with the threshold being the median feature value. For survival analysis, we considered  $p$  value  $<0.01$  to be statistically significant.

### 3 Results

Proposed approaches were implemented in 3D Slicer [1] and MATLAB 2013b (version 8.2 [44]) and performed on

**Table 3** Summary of the classification results

Classifier	Accuracy	Sensitivity	Specificity
DA	79.31	91.67	98.75
DT	70.69	75.00	90.00
NB	75.00	91.67	96.25
SVM	77.59	91.67	95.00

GBM brain tumor MR images. The GBM images were registered, and phenotypes were segmented and identified by our considered techniques.

#### 3.1 GLCM-based texture features

The 22 texture features exhibited  $p$  values  $<0.05$  which formed the basis for their use in further analysis (Table 2). Each of the 22 texture functions provided a unique value for each GBM phenotype. Texture functions  $f_{17}$ ,  $f_{18}$ ,  $f_{19}$ , and  $f_{20}$  show high values among the 22 features, and across the three GBM phenotypes, with maximum values of texture function seen for  $f_{19}$ . Note that the values for each feature take into consideration the average of the values derived from different phases and offsets for that particular feature and also the average derived from multiple samples. This correlation of texture feature values with the three GBM heterogeneity phenotypes allows these phenotypes to be distinguished using their distinct texture features.

#### 3.2 Classification

The Kruskal–Wallis test showed that whole 22 texture feature extracted from GLCMs were significant with  $p$  value  $<0.05$  (Table 2). Performance metrics of the texture feature classifier showed a maximum accuracy, sensitivity, and specificity of 79.31, 91.67, and 98.75 %, respectively, using DA technique, while the classifier accuracy value was decreased with SVM (77.59 %), NB (75.00 %), and DT (70.69 %), respectively (Table 3).

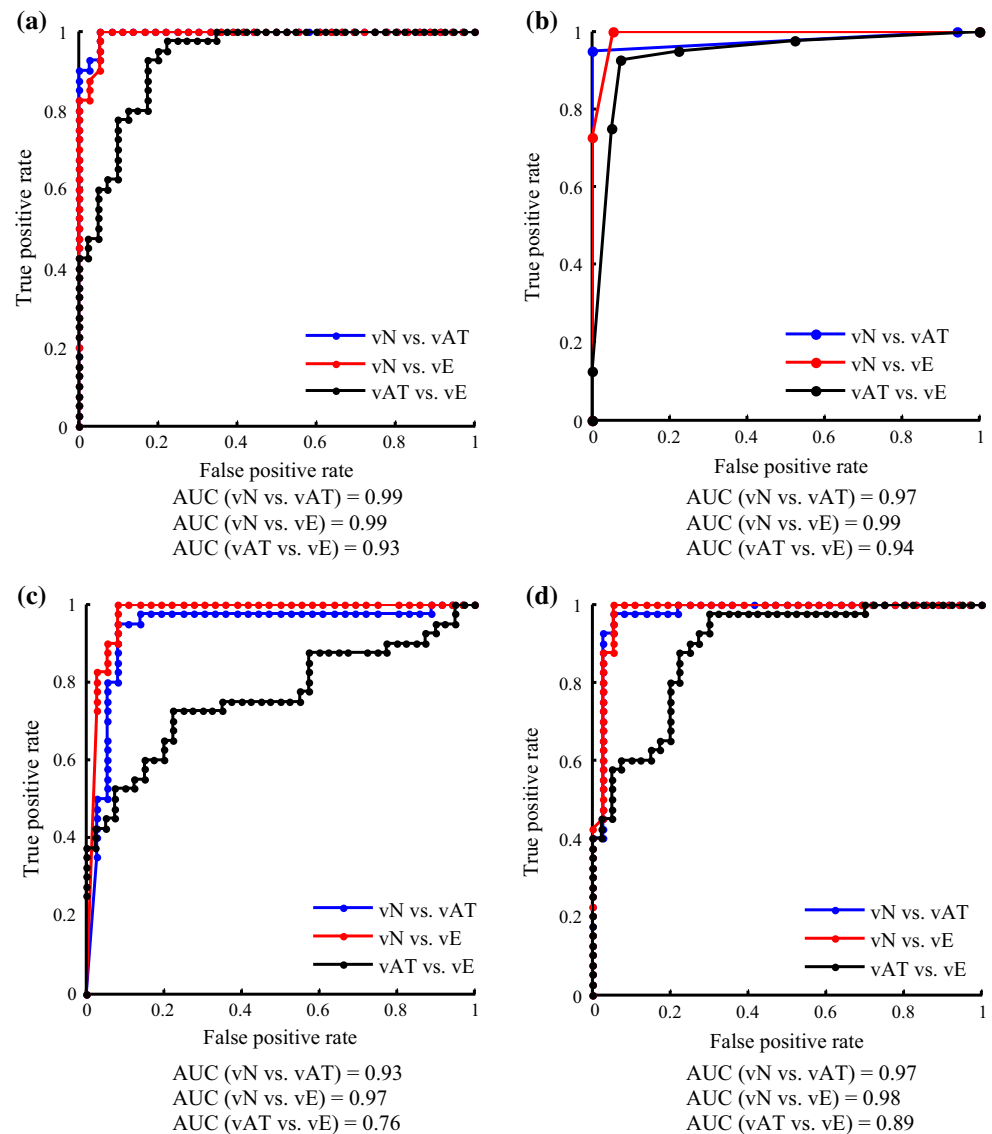
Moreover, from 36 vN, 40 vAT, and 40 vE phenotypes, confusion matrix showed that 33 vN and 32 vE were correctly classified using DA, while the highest vAT samples (29 vAT) were correctly classified using SVM classifier (Table 4).

The texture feature classifier of phenotypes was then evaluated based on ROC which showed that the AUC value based on DT classifier is better than the DA, SVM, and NB classifier (Fig. 2). AUC of (vN vs. vE) is greater than (vN vs. vAT) and (vAT vs. vE) of 99.23, 97.63, and 94.12 %, respectively (Fig. 2b). We choose the AUC of (vN vs. vE), (vN vs. vAT), and (vAT vs. vE) because the ROC must be operated between two classes.

**Table 4** Confusion matrix of the GBM phenotypes

Samples	DA			DT			NB			SVM		
	vN	vAT	vE	vN	vAT	vE	vN	vAT	vE	vN	vAT	vE
36 vN	33	0	3	27	3	6	33	0	3	33	2	1
40 vAT	0	27	13	4	28	8	0	25	15	2	29	9
40 vE	1	7	32	4	9	27	3	8	29	2	10	28

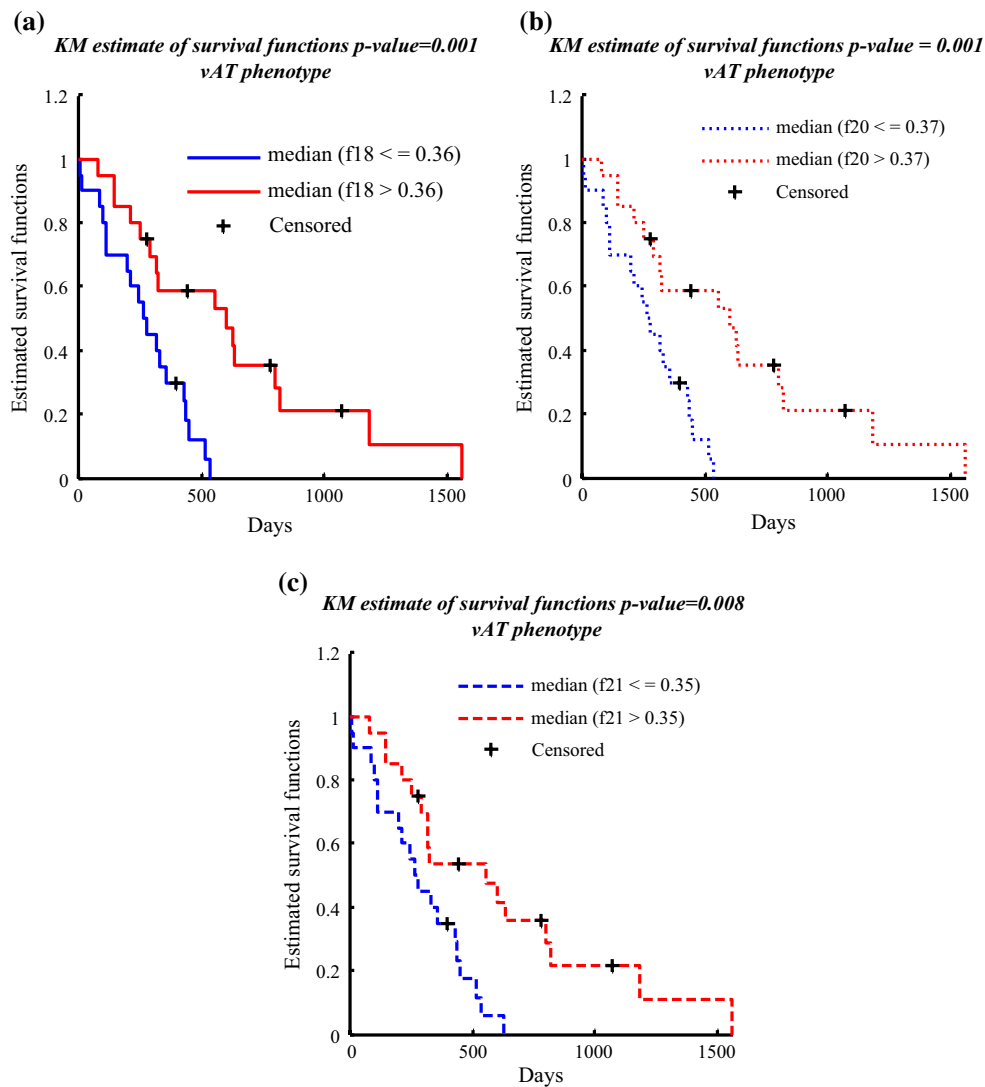
**Fig. 2** A ROC curve plotting true positive rate against false positive rate for vN versus vAT, vN versus vE, and vAT versus vE in four classifier techniques: **a** DA, **b** DT, **c** NB, and **d** SVM



**3.3 Survival analysis**

Kaplan–Meier curves were significantly different for three texture features: difference entropy  $f_{18}$  ( $p$  value = 0.001), information measure of correlation  $f_{20}$  ( $p$  value = 0.001), and inverse difference  $f_{21}$  ( $p$  value = 0.008) (Fig. 3; Table 5). Kaplan–Meier analysis confirmed that three features based on active tumor phenotype were predictors of overall survival. For edema and necrosis parts, the texture

features were not statistically significant for predicting overall survival with considered the  $p$  value <0.01. We observed that the most texture feature statistically significant to predict the survival time of GBM patient was in the active tumor parts (contrast enhancement, vAT), and this is confirmed if we considered the  $p$  value = 0.01. In these terms, we can also consider the features ( $f_1, f_2, f_3, f_4, f_{17}, f_{19},$  and  $f_{22}$ ) to predict the survival time of GBM patient (Table 5).



**Fig. 3** Kaplan–Meier curves show a significant difference in survival for **a** difference entropy, **b** information measure of correlation<sub>2</sub>, and **c** inverse difference with log-rank  $p$  values of 0.001, 0.001, and 0.008, respectively

Considered  $p$  value  $<0.01$ , median survival time based on texture feature is 273.5 days ( $f_{18} \leq 0.36$ ;  $f_{20} \leq 0.37$ ;  $f_{21} \leq 0.35$ ), 443 days ( $f_{18} > 0.36$ ;  $f_{20} > 0.37$ ), and 325 days ( $f_{21} > 0.35$ ). These texture features were statistically significant based on active tumor phenotype, while the features of edema and necrosis parts were not significant using Kaplan–Meier and log-rank test (Fig. 3; Table 5).

#### 4 Discussion

The goal of this study was to provide high discrimination accuracy of GBM phenotypes using texture feature extraction from GLCM computation and find the texture features that predict the overall survival of GBM patients. The results of our study provide further evidence that texture image

features that describe tumor spatial variations are useful in describing the GBM phenotypes and in predicting survival.

Data fitting showed that the histogram shape was distinct in data distribution of GBM phenotypes (Fig. 1). Moreover, the average, median, minimum, and maximum function of GBM phenotypes showed maximum, medium, and minimum values in vE, vAT, and vN, respectively (Table 1). Similarly, texture features relating to GBM phenotypes showed maximum, medium, and minimum values in vE, vAT, and vN, respectively (Table 2). This suggests that each GBM phenotype has higher values of texture vE than vAT and vN. Test of significance using texture feature to discriminate between GBM phenotypes was obtained using Kruskal–Wallis test. Furthermore, we observed that the whole GLCM-based feature set showed high significance ( $p$  value  $<0.05$ ) which is represented by 22 texture features

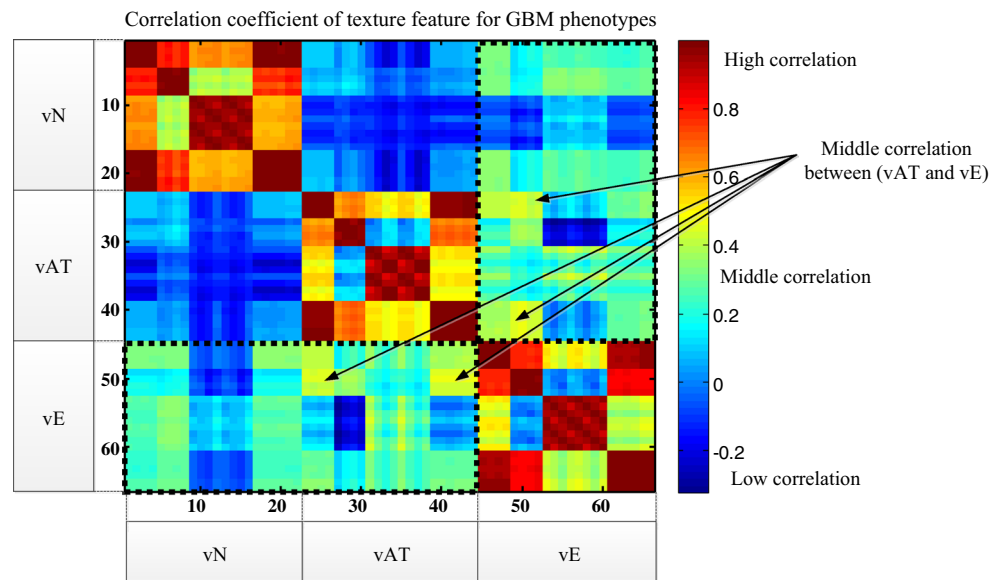


**Table 5** Summary of Kaplan–Meier analysis for GBM phenotypes texture features

Features	Cutoff (median)			Median survival (days)						<i>p</i> value		
				Above threshold			Below threshold					
	vN	vAT	vE	vN	vAT	vE	vN	vAT	vE	vN	vAT	vE
$f_1$	0.2	0.318	0.30	325.0	325.0	327.5	315.0	285.0	298.0	0.84	0.01	0.8
$f_2$	0.2	0.320	0.30	325.0	325.0	327.5	315.0	285.0	298.0	0.84	0.01	0.8
$f_3$	0.2	0.319	0.30	325.0	325.0	327.5	315.0	285.0	298.0	0.84	0.01	0.8
$f_4$	0.2	0.317	0.30	325.0	325.0	327.5	315.0	285.0	298.0	0.84	0.01	0.01
$f_5$	0.4	0.378	0.35	325.0	318.5	394.5	315.0	336.0	240.5	0.89	0.18	0.01
$f_6$	0.5	0.417	0.37	304.0	325.0	394.5	328.0	298.0	273.5	0.97	0.05	0.25
$f_7$	0.5	0.429	0.39	325.0	319.5	344.0	315.0	317.0	273.5	0.67	0.13	0.01
$f_8$	0.4	0.431	0.37	345.5	319.5	394.5	268.0	317.0	240.5	0.55	0.13	0.97
$f_9$	0.9	0.962	0.96	324.0	322.5	345.5	317.0	306.5	303.0	0.33	0.74	0.99
$f_{10}$	0.8	0.935	0.94	344.0	322.5	345.5	291.0	306.5	303.0	0.65	0.67	0.57
$f_{11}$	0.9	0.954	0.97	324.0	316.0	327.5	317.0	327.5	316.0	0.35	0.97	0.91
$f_{12}$	0.8	0.944	0.96	297.0	322.5	327.5	328.0	306.5	303.0	0.27	0.72	0.97
$f_{13}$	0.9	0.962	0.96	324.0	322.5	345.5	317.0	306.5	303.0	0.33	0.74	0.99
$f_{14}$	0.8	0.935	0.94	344.0	322.5	345.5	291.0	306.5	303.0	0.65	0.67	0.57
$f_{15}$	0.9	0.954	0.97	324.0	316.0	327.5	317.0	327.5	316.0	0.35	0.97	0.91
$f_{16}$	0.8	0.944	0.96	297.0	322.5	327.5	328.0	306.5	303.0	0.27	0.72	0.98
$f_{17}$	0.2	0.374	0.30	316.0	443.0	318.5	328.0	273.5	322.5	0.99	0.01	0.98
$f_{18}$	0.1	<b>0.36</b>	0.30	316.0	<b>443.0</b>	318.5	328.0	<b>273.5</b>	322.5	0.99	<b>0.001</b>	0.98
$f_{19}$	0.2	0.37	0.30	316.0	325.0	318.5	328.0	298.0	322.5	0.99	0.01	0.98
$f_{20}$	0.1	<b>0.37</b>	0.30	316.0	<b>443.0</b>	318.5	328.0	<b>273.5</b>	322.5	0.99	<b>0.001</b>	0.98
$f_{21}$	0.2	<b>0.35</b>	0.31	325.0	<b>325.0</b>	327.5	315.0	<b>273.5</b>	304.0	0.84	<b>0.008</b>	0.9
$f_{22}$	0.2	0.352	0.31	325.0	325.0	327.5	315.0	285.0	304.0	0.84	0.01	0.9

Signification features for predicting survival patients with GBM are shown in bold

**Fig. 4** Heat map of correlation coefficients between texture features of GBM phenotypes. vN, vAT, and vE are the necrosis, active tumor, and edema parts, respectively. *Black rectangle* is the part of correlation coefficient between (vE) and (vAT and vN). Correlation coefficient value >0.6, <0.2–0.6>, and <0.2 represents high, middle, and low correlation, respectively



(Table 2). This suggests that spatial textures based on the GLCM provide significant features to distinguish between phenotypes.

For classification of the GBM phenotypes, we observed that the trade-off between performance and accuracy was

achieved using DA classifier. The misclassification rate is resulted from some texture features that have common characteristics between GBM phenotypes which is represented by an AUC value between vAT versus vE in four classifier techniques (Fig. 2). This is further demonstrated

by higher coefficient of correlation values between phenotype features. The heat map of the phenotype feature correlation showed that edema parts have some correlation with necrosis and active tumor features (Fig. 4). Specifically, middle correlation coefficient range of 0.4–0.6 presented certain texture features between vAT and vE as illustrated in Fig. 4. The middle range of correlation coefficient values demonstrated a middle similarity between vAT and vE texture features.

In reality, overlap between GBM phenotypes can be represented by the misclassification ( $\approx 20\%$ ), and thus, it is not surprising that a certain texture feature shares a similar value. Note that the link between the texture feature and raw data is not directly correlated. For instance, a raw image with regular texture generates an irregular GLCM matrix which provides low value of the entropy ( $f_9$ ). Looking at the entropy in GBM phenotypes, vE has a maximum value followed by vAT and vN. It can be conditional that the edema phenotype is more irregular texture than vAT and vN, respectively (Table 2). This technique was successfully employed in abnormal colon cell discrimination [11], and the prediction of the clinical and pathological response to neoadjuvant chemotherapy (NAC) in patients with locally advanced breast cancer before NAC is started [41].

Additionally, survival analysis of GBM patients based on texture feature derived from GLCM demonstrated that the three features (difference entropy, information measure of correlation, and inverse difference) of vAT phenotype were statistically significant with median overall survival range of 273.5–443 days (Fig. 3). We observed that the texture in vAT phenotype may predict overall survival of GBM patients. Recently, a new study based on spatial habitat features has shown 14 spatial features associated with molecular subtype and 12-month survival GBM [26].

Our approach can be a potentially valuable tool in estimating characteristics invisible to the radiologist on inspection. To our knowledge, this is the first study looking into the discrimination of GBM phenotypes using texture feature extraction based on GLCM and to show the ability of the technique to predict overall survival of GBM patients based on their phenotype features. Many studies, however, have been carried out to discriminate between tumor types. For example, texture analysis proved to differentiate benign from malignant based on T1–WI and similarly to discriminate pleomorphic adenomas and warthin tumors [19]. Also, texture analysis has been utilized effectively in the characterization of posterior fossa tumors of children [32]. Moreover, a recent study using MRI showed that texture features derived from GLCM were significant features in differentiating true progression from pseudo-progression in GBM [14].

The influence of increased diagnostic accuracy is still limited to establish clinical use. However, one potential

advantage of the texture analysis is that the methods applied and their results are not limited to simple GBM phenotype discrimination. Our results, in addition to considering the optimal method of GBM phenotype discrimination, are also promising in their ability to characterize GBM heterogeneity, while the selection of texture features helps to accurately predict overall survival of GBM patients.

## 5 Conclusions

In this study, a novel approach based on texture feature extraction has been presented for GBM phenotype discrimination and predicting of overall patient survival. We have demonstrated the potential of texture features extracted from GLCM to characterize GBM phenotypes. The most important finding of this work is that the whole 22 texture feature set has been found to significantly classify GBM phenotypes, and three features were statistically significant for predicting overall survival. This study provides preliminary information of GBM phenotypes characterization and survival analysis of GBM patients. We note that a larger prospective trial is needed to fully evaluate the performance metrics of the proposed approach. Considering all the texture types extracted from GBM phenotypes that are under investigation, we posit that we are on the verge of a watershed moment in the identification and prediction of each phenotype by their texture features.

### Compliance with ethical standards

**Conflict of interest** The authors declare that there is no conflict of interest regarding the publication of this article.

**Ethical statement** The materials are in compliance with all applicable laws, regulations, and policies for the protection of medical data, and any necessary approvals, authorizations, and informed consent documents were obtained.

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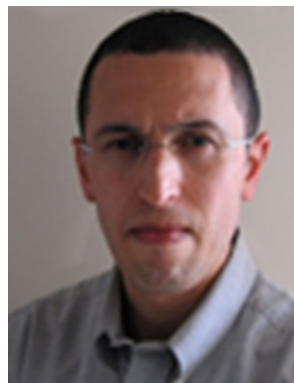
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