INVITED REVIEW

# HIV Neuropathogenesis: a Tight Rope Walk of Innate Immunity

Honghong Yao . Crystal Bethel-Brown . Cicy Zidong Li · Shilpa J. Buch

Received: 6 November 2009 / Accepted: 11 March 2010 / Published online: 31 March 2010  $©$  Springer Science+Business Media, LLC 2010

Abstract During the course of HIV-1 disease, virus neuroinvasion occurs as an early event, within weeks following infection. Intriguingly, subsequent central nervous system (CNS) complications manifest only decades after the initial virus exposure. Although CNS is commonly regarded as an immune-privileged site, emerging evidence indicates that innate immunity elicited by the CNS glial cells is a critical determinant for the establishment of protective immunity. Sustained expression of these protective immune responses, however, can be a double-edged sword. As protective immune mediators, cytokines have the ability to function in networks and co-operate with other host/viral mediators to tip the balance from a protective to toxic state in the CNS. Herein, we present an overview of some of the essential elements of the cerebral innate immunity in HIV neuropathogenesis including the key immune cell types of the CNS with their respective soluble immune mediators: (1) cooperative interaction of IFN- $\gamma$  with the host/virus factor (platelet-derived host factor (PDGF)/viral Tat) in the induction of neurotoxic chemokine CXCL10 by macrophages, (2) response of astrocytes to viral infection, and (3) protective role of PDGF and MCP-1 in neuronal survival against HIV Tat toxicity. These components of the cerebral innate immunity do not act separately from each other but form a functional immunity network. The ultimate outcome of HIV infection in the CNS will thus be dependent on the regulation of the net balance of cell-specific protective versus detrimental responses.

H. Yao  $\cdot$  C. Bethel-Brown  $\cdot$  C. Z. Li $\cdot$  S. J. Buch ( $\boxtimes$ ) Department of Pharmacology and Experimental Neuroscience, 985880 Nebraska Medical Center (DRC 8011), University of Nebraska Medical Center, Omaha, NE 68198-5880, USA e-mail: sbuch@unmc.edu

Keywords HIV neuropathogenesis. MCP-1 . CXCL10

## Introduction

It is estimated that almost 25% of untreated HIV-1-infected individuals and ∼7% of HIV-1-infected patients treated with combined antiretroviral therapy develop HIV-associated dementia (HAD) (Budka [1991;](#page-4-0) Spencer and Price [1992;](#page-5-0) McArthur et al. [1993](#page-4-0); Sacktor et al. [2001](#page-5-0)), a neurodegenerative syndrome that is clinically characterized by progressive cognitive, motor, and behavioral abnormalities (Gendelman et al. [1994](#page-4-0); Lipton and Gendelman [1995](#page-4-0)). The pathological manifestation of the syndrome, HIV encephalitis (HIVE), is accompanied by prominent microglial activation, formation of microglial nodules, perivascular accumulations of mononuclear cells, presence of multinucleated giant cells, and neuronal damage and loss (Gendelman et al. [1994;](#page-4-0) Bell [1998](#page-4-0); Nath [1999\)](#page-5-0). The primary cells infected by HIV-1 in the brain are macrophages/microglia and, to a lesser extent, astrocytes, but not neurons (Kaul et al. [2001](#page-4-0)). One broad explanation frequently advocated to explain the loss of neurons in this disease is that cellular and/or viral proteins released from the infected cells have a direct toxic effect on the neurons (Brenneman et al. [1988](#page-4-0); Dreyer et al. [1990](#page-4-0); Adamson et al. [1996](#page-4-0); Hof et al. [1998](#page-4-0); Kruman et al. [1998;](#page-4-0) Chen et al. [2000](#page-4-0); Patel et al. [2000](#page-5-0)).

HIV-1 neuroinvasion occurs during the early stages of disease; however, pathological phenotypes, like microglial activation, astrogliosis, and neuronal loss, are not manifested until late in the disease and, again, not in all infected individuals. This leads to the speculation that the ultimate disease outcome is a result of the balance of neuroprotective versus neurotoxic responses. Additionally, the

same pathways that mediate a protective response have also been shown to play critical roles in mediating toxicity. For example, antiviral cytokine such as IFN-γ can, under certain conditions, synergize with other inflammatory mediators such as TNF- $\alpha$ , resulting in induction of inflammatory mediators, such as the chemokine CXCL10 (Bokhari et al. [2009](#page-4-0); Williams et al. [2009\)](#page-6-0), thereby tipping the balance from a protective physiological response to a pathological one. Such cooperation and interplay of host and viral factors often culminates in end-organ HIV pathology observed in later stages of the disease. It is this dysregulation of host responses and the dual roles of these mediators in neuroAIDS that are the focus of this review. Herein, we will present an overview of some of the essential elements of the cerebral innate immunity in HIV-1 neuropathogenesis, specifically in the key immune cell types of the central nervous system (CNS).

### Macrophages

Cooperative interaction of IFN- $\gamma$  with the host/virus factor (PDGF/viral Tat) in the induction of neurotoxic chemokine CXCL10

Chemokines in the brain have been recognized as essential elements in neurodegenerative disease and related neuroinflammation through their regulation of inflammatory responses (Luster and Ravetch [1987;](#page-4-0) Conant et al. [1998](#page-4-0); Kelder et al. [1998](#page-4-0)), thereby contributing to injury and eventual loss of neurons (Asensio et al. [2001\)](#page-4-0). Cerebral expression of various chemokines, including CXCL10 (interferon  $\gamma$ -inducible peptide or IP-10), and its receptor is increased in HIVE (McArthur et al. [1993;](#page-4-0) Kolson and Pomerantz [1996;](#page-4-0) Sanders et al. [1998](#page-5-0); Kolb et al. [1999](#page-4-0)). Increased levels of CXCL10 have been detected in the cerebrospinal fluid and plasma of individuals with HIV-1 infection (Kolb et al. [1999\)](#page-4-0) and in the brains of individuals with HAD (McArthur et al. [1993](#page-4-0); Kolson and Pomerantz [1996;](#page-4-0) Sanders et al. [1998\)](#page-5-0). Importantly, the CXCL10 levels in the CNS of HIV-1-infected individuals correlated positively with disease progression. There is also evidence that CXCL10 participates in the neuropathogenesis in simian HIV (SHIV)-infected macaques (Sasseville et al. [1996;](#page-5-0) Westmoreland et al. [1998](#page-5-0)) by contributing to the degeneration of neurons, possibly through the activation of a calcium-dependent apoptotic pathway (Potula et al. [2004](#page-5-0); Sui et al. [2006](#page-5-0)). Increased CXCL10 levels are critical for the increased migration of inflammatory cells into the CNS, a hallmark feature of HAD (Navia et al. [1986](#page-5-0); Nath [1999](#page-5-0)).

In the brain, CXCL10 can be induced by a variety of factors, including viral gp120, Tat, and Nef, and cellular host factors such as IFN-γ (Kutsch et al. [2000;](#page-4-0) Asensio et

al. [2001;](#page-4-0) van Marle et al. [2004](#page-5-0)). Although both the cellular (IFN- $\gamma$  and TNF- $\alpha$ ) (Ohmori and Hamilton [1993;](#page-5-0) Majumder et al. [1998b\)](#page-4-0) and viral factors (Tat and gp120) (Kutsch et al. [2000;](#page-4-0) Asensio et al. [2001\)](#page-4-0) are known to cooperatively induce CXCL10, it remains unclear how the interplay of host factors and virus modulates chemokine expression. In fact, it has been shown that interactions of soluble host factors can synergistically induce the expression of CXCL10 (Ohmori and Hamilton [1993,](#page-5-0) [1995;](#page-5-0) Majumder et al. [1998a,](#page-4-0) [b\)](#page-4-0). In addition to the known inflammatory mediators, over-expression of the growth factor such as the platelet-derived growth factor (PDGF)-B chain has been demonstrated in the brains of macaques with SHIV-E (Sui et al. [2003](#page-5-0); Potula et al. [2004\)](#page-5-0). PDGF has been implicated in the pathogenesis of various inflammatory diseases (Ross et al. [1984](#page-5-0), [1985,](#page-5-0) [1986](#page-5-0); Ross and Raines [1988;](#page-5-0) Antoniades et al. [1990\)](#page-4-0). In addition to enhanced expression of IFN- $\gamma$ , there is also an increased expression of PDGF-B chain in the brains of macaques with SHIV-E (Lane et al. [1996;](#page-4-0) Orandle et al. [2002](#page-5-0); Shapshak et al. [2004\)](#page-5-0).

There is an increasing cumulative evidence that activated mononuclear phagocytes (macrophages/microglia) releasing inflammatory mediators in the CNS are a better correlate of HIV-associated neurocognitive disorders (HAND) than the actual viral load in the brain. The complex interplay of inflammatory mediators released by macrophages often leads to the induction of neurotoxins in HAD. Dissection of these complex interplays has led to the finding that, while PDGF alone had no effect on the induction of CXCL10 in human macrophages, in conjunction with IFN- $\gamma$ , it significantly augmented the expression of CXCL10 RNA and protein through transcriptional and posttranscriptional mechanisms. Signaling molecules, such as JAK and STATs, PI3K, mitogen-activated protein kinase (MAPK), and NF-κB, were found to play a role in the synergistic induction of CXCL10 (Dhillon et al. [2007\)](#page-4-0). Furthermore, PDGF via its activation of p38 MAPK was also able to increase the stability of IFN-γ-induced CXCL10 mRNA (Dhillon et al. [2007\)](#page-4-0).

Another mechanism of increased CXCL10 expression in the CNS occurs via the interplay of HIV-1 protein Tat and IFN- $\gamma$  in macrophages (Dhillon et al. [2008\)](#page-4-0). Synergistic induction of CXCL10 by both Tat and IFN- $\gamma$  was susceptible to inhibition by the MEK1/2 inhibitor U0126 and the p38 MAPK inhibitor SB203580. In addition, JAK/ STAT pathway was shown to play a major role in Tat/IFNγ-mediated CXCL10 induction in macrophages (Dhillon et al. [2008\)](#page-4-0). Thus, the cooperative interaction of Tat and IFN- $\gamma$  resulted in enhanced chemokine expression, which in turn can manifest as an amplified inflammatory immune response within the CNS of patients with HAD, by a mechanism involving an increased recruitment of lymphocytes/monocytes in the brain.

#### **Astrocytes**

#### Response of astrocytes to viral infection

While abundant studies support the role of macrophages and microglia in HIV-1-induced CNS damage, other glial cells such as astrocytes are also important contributors to neuronal pathology and inflammation. In addition to macrophages, astrocytes, the most numerous cell type within the brain, provide a very important reservoir for the generation CXCL10 (Dong and Benveniste [2001](#page-4-0); Thompson et al. [2001;](#page-5-0) Minagar et al. [2002](#page-4-0)). In fact, it is becoming increasingly appreciated that the severity of HAD/HIVE correlates more closely with the presence of activated glial cells rather than with the presence and amount of HIV-infected cells in the brain. It is known that proinflammatory cytokines, IFN- $\gamma$  and TNF- $\alpha$ , are markedly increased in CNS tissues during HIV-1 infection in the brain and are implicated in the pathophysiology of HAD (Wesselingh et al. [1997;](#page-5-0) Shapshak et al. [2004](#page-5-0)). One mechanism for the induced expression of CXCL10 in astrocytes has been via the synergistic induction of both IFN-γ and TNF-α (Majumder et al. [1998b\)](#page-4-0). Additionally, HIV-1/HIV-1 proteins in the CNS can further interact with these proinflammatory cytokines to further dramatically induce the expression of CXCL10 in the brain (Kutsch et al. [2000;](#page-4-0) Asensio et al. [2001](#page-4-0)). In fact, recent findings have demonstrated a synergistic induction of CXCL10 mRNA and protein in human astrocytes exposed to HIV-1/HIV proteins Tat and the proinflammatory cytokines. Signaling molecules, including JAK, STATs, MAPK (via activation of ERK1/ 2 and p38), and NF- $\kappa$ B, were identified as key players in the synergistic induction of CXCL10.

Furthermore, dissection of mechanisms involved in the synergistic induction of CXCL10 clearly points to the role of oxidative stress as a major player in this process. HIV-1 induced oxidative stress in astrocytes has been shown to be a critical determinant of the NF-κB-targeted genes, specifically CXCL10 (Song et al. [2007\)](#page-5-0). One mechanism by which oxidative stress is able to impact signaling pathways and their corresponding transcription factors is through a respiratory burst orchestrated by the activation of NADPH oxidase (Sundaresan et al. [1995;](#page-5-0) Wang et al. [1998;](#page-5-0) Adler et al. [1999](#page-4-0); Park et al. [2004](#page-5-0); Turchan-Cholewo et al. [2009](#page-5-0)). NADPH oxidase, a multi-subunit membrane associated enzyme, is capable of producing superoxide (Chanock et al. [1994](#page-4-0); Babior [1999;](#page-4-0) El-Benna et al. [2005](#page-4-0); Raad et al. [2008\)](#page-5-0). It has recently been demonstrated that treatment of astrocytes with a mixture of Tat and the inflammatory cytokines resulted in a respiratory burst, an effect that was abrogated by apocynin, an NADPH oxidase inhibitor (Williams et al. [2010](#page-6-0)). Treatment with apocynin also decreased CXCL10 expression in Tat-, IFN- $\gamma$ -, and TNF- $\alpha$ -stimulated astrocytes. This synergistic

induction involved the activation of the ERK1/2, JNK, and Akt pathways with downstream translocation of NF-κB.

## Neurons

Paradoxical roles of chemokines in HAND

In the brains of individuals with HAD, upregulation of chemokines in the CNS is often considered a correlate of neuroinflammation. However, recent increasing evidence raises the possibility that, in addition to their role as chemoattractants, chemokines might also act as neurotransmitters or neuromodulators (Rostene et al. [2007\)](#page-5-0). A classic example of this is the chemokine fractalkine, which is not only a neuroimmune modulator recruiting peripheral macrophages into the brain but can also function as a neuroprotective factor (Tong et al. [2000\)](#page-5-0). Similar to fractalkine, CCL2 has also been demonstrated to protect cardiac myocytes from hypoxia-induced apoptosis (Tarzami et al. [2005\)](#page-5-0). CCL2 is a constitutively expressed chemokine in the normal brain that is produced by a variety of CNS cells. It has been demonstrated that CCL2 can function not only to recruit monocytes into the CNS of HIV-1 patients but also to play a critical role in the protection of fetal neurons against HIV-1 Tat toxicity (Eugenin et al. [2003\)](#page-4-0). A recent study also identified an additional role for CCL2, that of neuroprotection against HIV-1 Tat toxicity in rat primary midbrain neurons involving transient receptor potential canonical (TRPC) (Yao et al. [2009b\)](#page-6-0).

Similarly, newer pleiotrophic roles of the chemokine fractalkine are also becoming increasingly appreciated in the literature. Specifically, although fractalkine is a known chemoattractant, it can also function to regulate neuronal survival via its antiapoptotic effects (Meucci et al., [1998;](#page-4-0) Meucci et al., [2000\)](#page-4-0).

In addition to the ability of the chemokines to exert diverse functions in varying cell types, there are other intrinsic mechanisms within the host that can manifest the pleiotropic nature of these chemokines. For example, it is well recognized that the activity of many neuroactive peptides is modulated by bioactive fragments, which are formed by the action of a variety of peptidases. This phenomenon appears to represent an important regulatory mechanism that modulates many neuropeptide systems. Similar processing of chemokines resulting in the alteration of their functions has recently been reported for the chemokine SDF-1α. These studies showed that, in addition to its direct neurotoxic effect, cleaved chemokine SDF (5-67) could actually amplify the HIVinduced ''bystander'' neurotoxicity. Herein, the authors elegantly demonstrated that the proteolytic cleavage of SDF resulted in a peptide that was highly immunogenic and that contributed to HIV neurodegeneration through engagement of

a chemokine receptor, CXCR3 (Vergote et al. [2006](#page-5-0)). Furthermore, this indirect effect occurred at a concentration range that was 100 times lower than the concentration required to induce a direct neuronal death. This is a classic example demonstrating yet another mechanism by which proteins can acquire neuropathogenic properties following proteolytic processing.

In response to cellular damage, the host is also capable of producing trophic growth factors (PDGF, fibroblast growth factor—FGF, or nerve growth factor—NGF) that may protect the neuronal, glial, and other resident cells of the brain. These neurotropic growth factors also play somewhat paradoxically diverse roles during the progression of CNS infection by promoting increased viral replication or cooperating with viral and/or host factors that promote neuronal degeneration. Many of these trophic factors also provide protection of neurons against the neurotoxic factors (cytokines, chemokines, and viral products). We previously demonstrated that, while PDGF expression in activated mononuclear phagocytes was closely associated in macaques with SHIV-E (Potula et al. [2004\)](#page-5-0), its expression in the neurons correlated with neuronal fitness (Peng et al. [2008b](#page-5-0)). Thus, depending on the cell type within the tissue, the same host factor can manifest diverse activation responses. Figure 1 illustrates the complex interactions that exist among trophic growth factors and cytokines/chemokines/viral proteins in the brains of HIV-infected patients. The ultimate outcome of infection in the CNS (neuronal survival versus damage) is a result of the ensuing shift in balance between the neurotrophic versus neurotoxic products manifested over time

492 J Neuroimmune Pharmacol (2010) 5:489–495

following infection. Recent findings demonstrated that PDGF-BB protected the neurons against gp120-mediated apoptosis as measured by lactate dehydrogenase and deoxynucleotidyltransferasemediated dUTP nick end labeling assay (Peng et al. [2008b](#page-5-0)). Antiapoptotic effects of PDGF-BB were also confirmed by regulation of caspase-3 activation and by examining levels of anti- and proapoptotic genes, Bcl-xL and Bax, respectively (Peng et al. [2008b](#page-5-0)). Further studies indicated the roles for both PI3K⁄ Akt and Bcl family pathways in PDGF-mediated neuroprotection in SH-SY5Y cells (Peng et al. [2008a\)](#page-5-0). In addition, it was also demonstrated that PDGF was able to rescue dopaminergic neurons from HIV Tat neurotoxicity in vivo (Yao et al. [2009a\)](#page-6-0). The confirmation of the neuroprotective role of PDGF against Tat toxicity was also corroborated in vitro in primary cultures of rat midbrain neurons. H. Yao and colleagues demonstrated a novel role of the  $Ca^{2+}$ -permeable channel TRPC in PDGF-mediated neuroprotection in rat neurons that indicated an exogenous PDGF-activated TRPC results in the amplification of downstream ERK signaling via the Pyk2 pathway, followed by nuclear translocation of CREB and ultimately culminating in neuronal survival (Yao et al. [2009a\)](#page-6-0). These studies implicate the robustness of PDGF neuroprotective ability against a range of diverse HIV-1 proteins.

## Conclusion

During HIV-1 infection, cytokines/chemokines, elaborated initially as protective immune mediators, have the potential

Fig. 1 Imbalance of neurotrophic versus neurotoxic factors in the CNS leads to development of HAND. In the normal brain, neuronal homeostasis is maintained by an increased expression of neurotrophic factors compared with the neurotoxins. High levels of neurotrophic factors are neuroprotective. On the other hand, following injury or virus infection, an increased expression of neurotoxins in the CNS shifts the balance to a diseased state that is exemplified as neuronal loss

**Normal Brain** Neurotropic Factor Neurotoxins BDNF, NGF, GDNF, PDGF Cytokine, Chemokine, HIV proteins Neurotropic Factor Neurotoxins **HAND** BDNF, NGF, GDNF, PDGF Cytokine, Chemokine, HIV proteins <span id="page-4-0"></span>to function in networks and can co-operate with other host/ viral mediators to tip the balance from a protective to a toxic response in the CNS. A better understanding of the basic mechanisms of HAND is thus critical for the development of treatments for neurological compliant of AIDS.

Acknowledgement This work was supported by grants MH-068212, DA020392, DA023397, DA024442, and DA 027729 from the National Institutes of Health (SB).

## References

- Adamson DC, Dawson TM, Zink MC, Clements JE, Dawson VL (1996) Neurovirulent simian immunodeficiency virus infection induces neuronal, endothelial, and glial apoptosis. Mol Med 2:417–428
- Adler V, Yin Z, Tew KD, Ronai Z (1999) Role of redox potential and reactive oxygen species in stress signaling. Oncogene 18:6104–6111
- Antoniades HN, Bravo MA, Avila RE, Galanopoulos T, Neville-Golden J, Maxwell M, Selman M (1990) Platelet-derived growth factor in idiopathic pulmonary fibrosis. J Clin Invest 86:1055–1064
- Asensio VC, Maier J, Milner R, Boztug K, Kincaid C, Moulard M, Phillipson C, Lindsley K, Krucker T, Fox HS, Campbell IL (2001) Interferon-independent, human immunodeficiency virus type 1 gp120-mediated induction of CXCL10/IP-10 gene expression by astrocytes in vivo and in vitro. J Virol 75:7067–7077
- Babior BM (1999) NADPH oxidase: an update. Blood 93:1464–1476
- Bell JE (1998) The neuropathology of adult HIV infection. Rev Neurol (Paris) 154:816–829
- Bokhari SM, Yao H, Bethel-Brown C, Fuwang P, Williams R, Dhillon NK, Hegde R, Kumar A, Buch SJ (2009) Morphine enhances Tat-induced activation in murine microglia. J Neurovirol 22:1–10
- Brenneman DE, Westbrook GL, Fitzgerald SP, Ennist DL, Elkins KL, Ruff MR, Pert CB (1988) Neuronal cell killing by the envelope protein of HIV and its prevention by vasoactive intestinal peptide. Nature 335:639–642
- Budka H (1991) Neuropathology of human immunodeficiency virus infection. Brain Pathol 1:163–175
- Chanock SJ, el Benna J, Smith RM, Babior BM (1994) The respiratory burst oxidase. J Biol Chem 269:24519–24522
- Chen Z, Huang Y, Zhao X, Skulsky E, Lin D, Ip J, Gettie A, Ho DD (2000) Enhanced infectivity of an R5-tropic simian/human immunodeficiency virus carrying human immunodeficiency virus type 1 subtype C envelope after serial passages in pig-tailed macaques (Macaca nemestrina). J Virol 74:6501–6510
- Conant K, Garzino-Demo A, Nath A, McArthur JC, Halliday W, Power C, Gallo RC, Major EO (1998) Induction of monocyte chemoattractant protein-1 in HIV-1 Tat-stimulated astrocytes and elevation in AIDS dementia. Proc Natl Acad Sci U S A 95:3117–3121
- Dhillon NK, Peng F, Ransohoff RM, Buch S (2007) PDGF synergistically enhances IFN-gamma-induced expression of CXCL10 in blood-derived macrophages: implications for HIV dementia. J Immunol 179:2722–2730
- Dhillon N, Zhu X, Peng F, Yao H, Williams R, Qiu J, Callen S, Ladner AO, Buch S (2008) Molecular mechanism(s) involved in the synergistic induction of CXCL10 by human immunodeficiency virus type 1 Tat and interferon-gamma in macrophages. J Neurovirol 14:196–204
- Dong Y, Benveniste EN (2001) Immune function of astrocytes. Glia 36:180–190
- Dreyer EB, Kaiser PK, Offermann JT, Lipton SA (1990) HIV-1 coat protein neurotoxicity prevented by calcium channel antagonists. Science 248:364–367
- El-Benna J, Dang PM, Gougerot-Pocidalo MA, Elbim C (2005) Phagocyte NADPH oxidase: a multicomponent enzyme essential for host defenses. Arch Immunol Ther Exp (Warsz) 53:199–206
- Eugenin EA, D'Aversa TG, Lopez L, Calderon TM, Berman JW (2003) MCP-1 (CCL2) protects human neurons and astrocytes from NMDA or HIV-tat-induced apoptosis. J Neurochem 85:1299–1311
- Gendelman HE, Lipton SA, Tardieu M, Bukrinsky MI, Nottet HS (1994) The neuropathogenesis of HIV-1 infection. J Leukoc Biol 56:389–398
- Hof PR, Lee PY, Yeung G, Wang RF, Podos SM, Morrison JH (1998) Glutamate receptor subunit GluR2 and NMDAR1 immunoreactivity in the retina of macaque monkeys with experimental glaucoma does not identify vulnerable neurons. Exp Neurol 153:234–241
- Kaul M, Garden GA, Lipton SA (2001) Pathways to neuronal injury and apoptosis in HIV-associated dementia. Nature 410:988–994
- Kelder W, McArthur JC, Nance-Sproson T, McClernon D, Griffin DE (1998) Beta-chemokines MCP-1 and RANTES are selectively increased in cerebrospinal fluid of patients with human immunodeficiency virus-associated dementia. Ann Neurol 44:831–835
- Kolb SA, Sporer B, Lahrtz F, Koedel U, Pfister HW, Fontana A (1999) Identification of a T cell chemotactic factor in the cerebrospinal fluid of HIV-1-infected individuals as interferongamma inducible protein 10. J Neuroimmunol 93:172–181
- Kolson DL, Pomerantz RJ (1996) AIDS dementia and HIV-1-induced neurotoxicity: possible pathogenic associations and mechanisms. J Biomed Sci 3:389–414
- Kruman I, Guo Q, Mattson MP (1998) Calcium and reactive oxygen species mediate staurosporine-induced mitochondrial dysfunction and apoptosis in PC12 cells. J Neurosci Res 51:293–308
- Kutsch O, Oh J, Nath A, Benveniste EN (2000) Induction of the chemokines interleukin-8 and IP-10 by human immunodeficiency virus type 1 tat in astrocytes. J Virol 74:9214–9221
- Lane TE, Buchmeier MJ, Watry DD, Fox HS (1996) Expression of inflammatory cytokines and inducible nitric oxide synthase in brains of SIV-infected rhesus monkeys: applications to HIVinduced central nervous system disease. Mol Med 2:27–37
- Lipton SA, Gendelman HE (1995) Seminars in medicine of the Beth Israel Hospital, Boston. Dementia associated with the acquired immunodeficiency syndrome. N Engl J Med 332:934–940
- Luster AD, Ravetch JV (1987) Biochemical characterization of a gamma interferon-inducible cytokine (IP-10). J Exp Med 166:1084–1097
- Majumder S, Zhou LZ, Chaturvedi P, Babcock G, Aras S, Ransohoff RM (1998a) Regulation of human IP-10 gene expression in astrocytoma cells by inflammatory cytokines. J Neurosci Res 54:169–180
- Majumder S, Zhou LZ, Chaturvedi P, Babcock G, Aras S, Ransohoff RM (1998b) p48/STAT-1alpha-containing complexes play a predominant role in induction of IFN-gamma-inducible protein, 10 kDa (IP-10) by IFN-gamma alone or in synergy with TNFalpha. J Immunol 161:4736–4744
- McArthur JC, Hoover DR, Bacellar H, Miller EN, Cohen BA, Becker JT, Graham NM, McArthur JH, Selnes OA, Jacobson LP et al (1993) Dementia in AIDS patients: incidence and risk factors. Multicenter AIDS Cohort Study. Neurology 43:2245–2252
- Meucci O, Fatatis A, Simen AA, Bushell TJ, Gray PW, Miller RJ (1998) Chemokines regulate hippocampal neuronal signaling and gp120 neurotoxicity. Proc Natl Acad Sci U S A 95:14500–14505
- Meucci O, Fatatis A, Simen AA, Miller RJ (2000) Expression of CX3CR1 chemokine receptors on neurons and their role in neuronal survival. Proc Natl Acad Sci U S A 97:8075–8080
- Minagar A, Shapshak P, Fujimura R, Ownby R, Heyes M, Eisdorfer C (2002) The role of macrophage/microglia and astrocytes in the pathogenesis of three neurologic disorders: HIV-associated

<span id="page-5-0"></span>dementia, Alzheimer disease, and multiple sclerosis. J Neurol Sci 202:13–23

- Nath A (1999) Pathobiology of human immunodeficiency virus dementia. Semin Neurol 19:113–127
- Navia BA, Cho ES, Petito CK, Price RW (1986) The AIDS dementia complex: II. Neuropathology. Ann Neurol 19:525–535
- Ohmori Y, Hamilton TA (1993) Cooperative interaction between interferon (IFN) stimulus response element and kappa B sequence motifs controls IFN gamma- and lipopolysaccharidestimulated transcription from the murine IP-10 promoter. J Biol Chem 268:6677–6688
- Ohmori Y, Hamilton TA (1995) The interferon-stimulated response element and a kappa B site mediate synergistic induction of murine IP-10 gene transcription by IFN-gamma and TNF-alpha. J Immunol 154:5235–5244
- Orandle MS, MacLean AG, Sasseville VG, Alvarez X, Lackner AA (2002) Enhanced expression of proinflammatory cytokines in the central nervous system is associated with neuroinvasion by simian immunodeficiency virus and the development of encephalitis. J Virol 76:5797–5802
- Park J, Choi K, Jeong E, Kwon D, Benveniste EN, Choi C (2004) Reactive oxygen species mediate chloroquine-induced expression of chemokines by human astroglial cells. Glia 47:9–20
- Patel CA, Mukhtar M, Pomerantz RJ (2000) Human immunodeficiency virus type 1 Vpr induces apoptosis in human neuronal cells. J Virol 74:9717–9726
- Peng F, Dhillon NK, Yao H, Zhu X, Williams R, Buch S (2008a) Mechanisms of platelet-derived growth factor-mediated neuroprotection—implications in HIV dementia. Eur J NeuroSci 28:1255–1264
- Peng F, Dhillon N, Callen S, Yao H, Bokhari S, Zhu X, Baydoun HH, Buch S (2008b) Platelet-derived growth factor protects neurons against gp120-mediated toxicity. J Neurovirol 14:62–72
- Potula R, Dhillion N, Sui Y, Zien CA, Funa K, Pinson D, Mayo MS, Singh DK, Narayan O, Buch S (2004) Association of platelet-derived growth factor-B chain with simian human immunodeficiency virus encephalitis. Am J Pathol 165:815– 824
- Raad H, Paclet MH, Boussetta T, Kroviarski Y, Morel F, Quinn MT, Gougerot-Pocidalo MA, Dang PM, El-Benna J (2008) Regulation of the phagocyte NADPH oxidase activity: phosphorylation of gp91phox/NOX2 by protein kinase C enhances its diaphorase activity and binding to Rac2, p67phox, and p47phox. Faseb J
- Ross R, Raines EW (1988) Platelet-derived growth factor—its role in health and disease. Adv Exp Med Biol 234:9–21
- Ross R, Faggiotto A, Bowen-Pope D, Raines E (1984) The role of endothelial injury and platelet and macrophage interactions in atherosclerosis. Circulation 70:III77–III82
- Ross R, Bowen-Pope DF, Raines EW (1985) Platelet-derived growth factor: its potential roles in wound healing, atherosclerosis, neoplasia, and growth and development. Ciba Found Symp 116:98–112
- Ross R, Raines EW, Bowen-Pope DF (1986) The biology of plateletderived growth factor. Cell 46:155–169
- Rostene W, Kitabgi P, Parsadaniantz SM (2007) Chemokines: a new class of neuromodulator? Nat Rev Neurosci 8:895–903
- Sacktor N, Lyles RH, Skolasky R, Kleeberger C, Selnes OA, Miller EN, Becker JT, Cohen B, McArthur JC (2001) HIV-associated neurologic disease incidence changes: multicenter AIDS cohort study, 1990–1998. Neurology 56:257–260
- Sanders VJ, Pittman CA, White MG, Wang G, Wiley CA, Achim CL (1998) Chemokines and receptors in HIV encephalitis. AIDS 12:1021–1026
- Sasseville VG, Smith MM, Mackay CR, Pauley DR, Mansfield KG, Ringler DJ, Lackner AA (1996) Chemokine expression in simian

immunodeficiency virus-induced AIDS encephalitis. Am J Pathol 149:1459–1467

- Shapshak P, Duncan R, Minagar A, Rodriguez de la Vega P, Stewart RV, Goodkin K (2004) Elevated expression of IFN-gamma in the HIV-1 infected brain. Front Biosci 9:1073–1081
- Song HY, Ryu J, Ju SM, Park LJ, Lee JA, Choi SY, Park J (2007) Extracellular HIV-1 Tat enhances monocyte adhesion by upregulation of ICAM-1 and VCAM-1 gene expression via ROSdependent NF-kappaB activation in astrocytes. Exp Mol Med 39:27–37
- Spencer DC, Price RW (1992) Human immunodeficiency virus and the central nervous system. Annu Rev Microbiol 46:655–693
- Sui Y, Potula R, Pinson D, Adany I, Li Z, Day J, Buch E, Segebrecht J, Villinger F, Liu Z, Huang M, Narayan O, Buch S (2003) Microarray analysis of cytokine and chemokine genes in the brains of macaques with SHIV-encephalitis. J Med Primatol 32:229–239
- Sui Y, Stehno-Bittel L, Li S, Loganathan R, Dhillon NK, Pinson D, Nath A, Kolson D, Narayan O, Buch S (2006) CXCL10-induced cell death in neurons: role of calcium dysregulation. Eur J NeuroSci 23:957–964
- Sundaresan M, Yu ZX, Ferrans VJ, Irani K, Finkel T (1995) Requirement for generation of  $H_2O_2$  for platelet-derived growth factor signal transduction. Science 270:296–299
- Tarzami ST, Calderon TM, Deguzman A, Lopez L, Kitsis RN, Berman JW (2005) MCP-1/CCL2 protects cardiac myocytes from hypoxia-induced apoptosis by a G(alphai)-independent pathway. Biochem Biophys Res Commun 335:1008–1016
- Thompson KA, McArthur JC, Wesselingh SL (2001) Correlation between neurological progression and astrocyte apoptosis in HIV-associated dementia. Ann Neurol 49:745–752
- Tong N, Perry SW, Zhang Q, James HJ, Guo H, Brooks A, Bal H, Kinnear SA, Fine S, Epstein LG, Dairaghi D, Schall TJ, Gendelman HE, Dewhurst S, Sharer LR, Gelbard HA (2000) Neuronal fractalkine expression in HIV-1 encephalitis: roles for macrophage recruitment and neuroprotection in the central nervous system. J Immunol 164:1333–1339
- Turchan-Cholewo J, Dimayuga VM, Gupta S, Gorospe RM, Keller JN, Bruce-Keller AJ (2009) NADPH oxidase drives cytokine and neurotoxin release from microglia and macrophages in response to HIV-Tat. Antioxid Redox Signal 11:193–204
- van Marle G, Henry S, Todoruk T, Sullivan A, Silva C, Rourke SB, Holden J, McArthur JC, Gill MJ, Power C (2004) Human immunodeficiency virus type 1 Nef protein mediates neural cell death: a neurotoxic role for IP-10. Virology 329:302–318
- Vergote D, Butler GS, Ooms M, Cox JH, Silva C, Hollenberg MD, Jhamandas JH, Overall CM, Power C (2006) Proteolytic processing of SDF-1alpha reveals a change in receptor specificity mediating HIV-associated neurodegeneration. Proc Natl Acad Sci U S A 103:19182–19187
- Wang X, Martindale JL, Liu Y, Holbrook NJ (1998) The cellular response to oxidative stress: influences of mitogen-activated protein kinase signalling pathways on cell survival. Biochem J 333(Pt 2):291–300
- Wesselingh SL, Takahashi K, Glass JD, McArthur JC, Griffin JW, Griffin DE (1997) Cellular localization of tumor necrosis factor mRNA in neurological tissue from HIV-infected patients by combined reverse transcriptase/polymerase chain reaction in situ hybridization and immunohistochemistry. J Neuroimmunol 74:1–8
- Westmoreland SV, Rottman JB, Williams KC, Lackner AA, Sasseville VG (1998) Chemokine receptor expression on resident and inflammatory cells in the brain of macaques with simian immunodeficiency virus encephalitis. Am J Pathol 152:659–665
- <span id="page-6-0"></span>Williams R, Dhillon NK, Hegde ST, Yao H, Peng F, Callen S, Chebloune Y, Davis RL, Buch SJ (2009) Proinflammatory cytokines and HIV-1 synergistically enhance CXCL10 expression in human astrocytes. Glia 57:734–743
- Williams R, Yao H, Peng F, Yang Y, Bethel-Brown C, Buch S (2010) Cooperative induction of CXCL10 involves NADPH oxidase: implications for HIV dementia. Glia 58:611–621
- Yao H, Peng F, Fan Y, Zhu X, Hu G, Buch SJ (2009a) TRPC channelmediated neuroprotection by PDGF involves Pyk2/ERK/CREB pathway. Cell Death Differ 16:1681–1693
- Yao H, Peng F, Dhillon N, Callen S, Bokhari S, Stehno-Bittel L, Ahmad SO, Wang JQ, Buch S (2009b) Involvement of TRPC channels in CCL2-mediated neuroprotection against tat toxicity. J Neurosci 29:1657–1669