Geography

June 2011 Vol.56 No.16: 1714–1720 doi: 10.1007/s11434-011-4418-y

Is the maximum carbon number of long-chain *n*-alkanes an indicator of grassland or forest? Evidence from surface soils and modern plants

RAO ZhiGuo^{1,2}, WU Yi^{1,3}, ZHU ZhaoYu^{1*}, JIA GuoDong¹ & HENDERSON Andrew⁴

Received November 10, 2010; accepted December 23, 2010

The molecular distribution of long-chain n-alkanes in 62 soil samples collected from diverse locations across eastern China was analyzed. The long-chain n-alkanes were mostly dominated by n- C_{29} or n- C_{31} , regardless of the overlying vegetation type at each site. The results were compared with those summarized from the literature, covering more than 100 soil samples within China and more than 300 genera of modern plants distributed worldwide. There were similar n-alkane distribution patterns for most genera, with no clear differences among grasses, shrubs, and trees. The evidence from analyses of surface soils and modern plants indicates that the relationship between the molecular distribution of long-chain n-alkanes of surface soils and source vegetation is highly complex, and is influenced by many factors. Further, it is suggested that source vegetation types should not be simply inferred from distribution patterns of long-chain n-alkanes in sediments.

surface soils, modern plants, long-chain n-alkanes, molecular distribution, vegetation type

Citation:

Rao Z G, Wu Y, Zhu Z Y, et al. Is the maximum carbon number of long-chain n-alkanes an indicator of grassland or forest? Evidence from surface soils and modern plants. Chinese Sci Bull, 2011, 56: 1714–1720, doi: 10.1007/s11434-011-4418-y

The evolution of terrestrial landscapes over geological time and modern history, including alternations between grassland and forest, has been driven by changes in the global climate. Efforts to reconstruct the history of terrestrial vegetation have mostly relied on the use of pollen [1,2] and phytolith [3] assemblages. Recently, the characteristic profiles of n-alkanes extracted from loess [4–6] and red clay [7,8] sediments have been used to reconstruct changes in vegetation over geological timescales. In some studies, the maximum carbon number (MCN) of long-chain n-alkanes, i.e., the carbon number of the most abundant homolog, has been used to indicate vegetation type. More specifically, it was suggested that long-chain n-alkanes dominated by n-C₂₇ or n-C₂₉ indicate forest, while those dominated by n-C₃₁

indicate grassland. Therefore, the ratio of n- C_{27} to n- C_{31} (or n- C_{27} +n- C_{29} /n- C_{31} +n- C_{33}) has been used to represent the input of woody versus herbaceous plants.

Leaf waxes of higher terrestrial plants contain abundant long-chain *n*-alkanes with significant odd-to-even carbon number preference [9,10]. There are demonstrable links between the molecular distribution of long-chain *n*-alkanes of modern plants and different seasons, study sites, plant age, and plant organs [9]. Thus, there should be a complex relationship between the molecular distribution of sedimentary long-chain *n*-alkanes from multiple source plants and the source vegetation.

Even all the long-chain n-alkanes produced by woody plants are dominated by n- C_{27} or n- C_{29} , while those of herbaceous plants are dominated by n- C_{31} , it is unclear whether the MCN of material from multiple plant sources can be

¹ Key Laboratory of Marginal Sea Geology, Guangzhou Institute of Geochemistry, Chinese Academy of Sciences, Guangzhou 510640, China;

² Key Laboratory of Western China's Environmental Systems (Ministry of Education), Lanzhou University, Lanzhou 730000, China;

³ Graduate University of Chinese Academy of Sciences, Beijing 100049, China;

⁴ School of Geographical & Earth Sciences, University of Glasgow, Glasgow G12 8QQ, UK

^{*}Corresponding author (email: zhuzy@gig.ac.cm)

used to reconstruct the source vegetation type. It is possible that long-chain n-alkanes mixed from sources dominated by n-C₂₇ (woody plants) and n-C₃₁ (herbaceous plants) could be dominated by the n-C₂₉, because it occurs at relatively high concentrations in both of the source plant types. For example, if a woody plant with a ratio of 10:7:3 for $n-C_{27}$, n-C₂₉, and n-C₃₁ and a herbaceous plant with a ratio of 1:7:10 for n- C_{27} , n- C_{29} , and n- C_{31} contributed equally to a certain sediment, the n-alkanes extracted from that sediment would show a ratio of 11:14:13 for $n-C_{27}$, $n-C_{29}$, and $n-C_{31}$. This would erroneously indicate that n-C29 was the dominant homolog in the source vegetation. Therefore, the MCN may not be a robust indicator of vegetation type, and the relative abundance of n- C_{27} to n- C_{31} does not indicate equal inputs from woody and herbaceous vegetation. Moreover, the production of long-chain n-alkanes may differ between different plants, which further complicate reconstructions of past vegetation.

Currently, there is no evidence that long-chain n-alkanes from woody plants are dominated by n- C_{27} and n- C_{29} , or that those of herbaceous plants are dominated by n- C_{31} . Similarly, there is little work showing how the MCN and molecular distribution of sedimentary long-chain n-alkanes from higher plants can be used to reconstruct paleovegetation. In this paper, we explore the relationship between the distribution of long chain n-alkanes and their respective vegetation sources based on the results from 100 surface soil samples collected from diverse areas across China and more than 300 globally distributed genera of modern plants.

1 Material and methods

For this study, a total of 62 surface soil samples were collected from diverse areas across China, extending from Hainan Province in southern China to Heilongjiang Province in northeastern China, spanning latitudes of 18°N to 50°N (Figure 1; Table S1). Vegetation across the study area ranges from tropical rainforest in southern China to temperate coniferous-deciduous broad-leaved mixed forest in northeastern China. The prevailing climate is monsoonal with concurrent rainy and hot seasons, with mean annual precipitation ranging from 500–2500 mm and a mean annual temperature of 0–26°C. The monsoonal climate trends from south to north with a strong dependence on latitude.

Surface soil samples were collected from the top 2–4 cm of weathering crust of basalt or other visible bedrocks in the field, thus avoiding contamination from underground water and allochthonous organic matter. The vegetation at all sampling sites was natural, with little human disturbance. Two to three surface samples were collected from different physiographic locations of each sampling site. A small grassland located in Damaping (114.5°E, 40.9°N, Figure 1), was sampled more intensively (12 samples). All samples

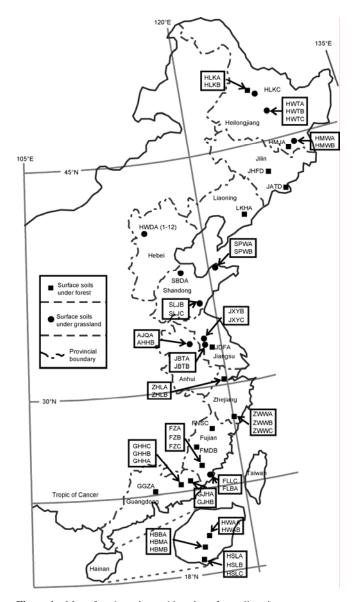


Figure 1 Map of study region and location of sampling sites.

were collected between September and November in 2005.

Rootlets and gravel were removed from the samples, and then they were ground and sieved through an 80- μ m mesh sieve. After immersion in dichloromethane for ~2 h, approximately 10 g powdered soil was ultrasonically extracted for 10 min. This process was repeated three times and the combined solvents were then concentrated by rotary evaporation. The total lipid extract was separated using silica gel flash-column chromatography. The sample was eluted with hexane to obtain the saturated hydrocarbon fraction containing long-chain n-alkanes.

The long-chain n-alkanes were analyzed using an HP 6890 gas chromatograph equipped with an HP-5 MS silica capillary column (30 m \times 0.32 mm \times 0.25 μ m). The oven temperature program was as follows: 80°C for 3 min, increasing to 200°C at a rate of 10°C/min, then increasing at

3°C/min to 290°C, and hold for 30 min. A reference material-Indiana STD (a set of n-alkanes consisting of 10 homologs; n-C₁₂, n-C₁₄, n-C₁₆, n-C₁₈, n-C₂₀, n-C₂₂, n-C₂₅, n-C₂₈, n-C₃₀, and n-C₃₂; Figure 2) was analyzed under exactly the same conditions. The carbon numbers of n-alkanes extracted from surface soil samples were determined by comparing their retention times with those of known alkanes in the reference material (Figure 2). The areas underneath n-alkane peaks were used to determine relative abundance.

2 Results and discussion

2.1 Long-chain *n*-alkanes extracted from surface soils under different vegetation types

The carbon numbers of *n*-alkanes extracted from most surface soil samples ranged from 14 to 35 with significant bimodal distribution (Figure 2). The short chain components without strong odd-to-even carbon number preference were dominated by either n- C_{17} or n- C_{19} . All samples showed relatively high concentrations of n- C_{23} to n- C_{34} alkanes with strong odd-to-even preference, with n- C_{27} , n- C_{29} , and n- C_{31} being the most dominant. The CPI (carbon preference index) of the long-chain n-alkanes extracted from all 62 surface soil samples ranged from 3 to 12.8 with an average value of 6.3. As reported in previous studies, n- C_{15} to n- C_{20} alkanes without significant odd-to-even preference and dominated by n- C_{17} or n- C_{19} alkanes are mainly derived from lower organisms [11], while terrestrial higher plants produce abundant long-chain n-alkanes with significant odd-to-even preference (such as n- C_{27} , n- C_{29} , and n- C_{31}) and with a CPI value exceeding 5 [9,12]. According to these classifications, the molecular distributions of n-alkanes extracted from our surface soil samples are characteristic of both higher plants and lower organisms, while the long-chain *n*-alkane components are derived from terrestrial higher plants.

We analyzed the MCN and relative abundance of the three main homologs (n- C_{27} , n- C_{29} , and n- C_{31}) of long-chain n-alkanes in each sample. Half of the samples (31) were collected from forested locations. The MCNs of n-alkanes in these samples were 27, 29, or 31, reflecting dominance of n-C₂₇, n-C₂₉, or n-C₃₁ alkanes, respectively (Table S1; Figure 3). Of these 31 samples, 22 were dominated by $n-C_{29}$, 14 of which showed a relative abundance of $n-C_{29}>n-C_{31}>$ n-C₂₇, and 8 of which showed a relative abundance of $n-C_{29}>n-C_{27}>n-C_{31}$. Of the remaining 9 samples, 5 were dominated by n- C_{27} with a relative abundance of n- C_{27} > $n-C_{29}>n-C_{31}$, and 4 were dominated by $n-C_{31}$ with a relative abundance of n-C₃₁>n-C₂₉>n-C₂₇. Half of the samples (31) were collected from grassland. These had an MCN of 29 or 31, reflecting dominance of n-C₂₉ or n-C₃₁ alkanes, respectively. Of these 31 samples, 14 were dominated by $n-C_{31}$

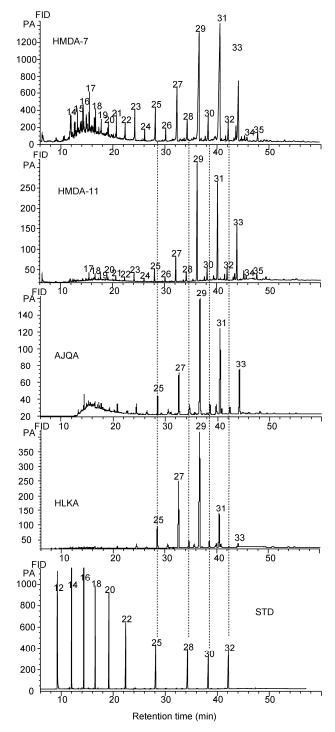


Figure 2 GC chromatogram of reference material and four typical surface soil samples (refer to Figure 1 and Table S1 for detailed information on soil samples).

with a relative abundance of n- C_{31} >n- C_{29} >n- C_{27} . The other 17 samples were dominated by n- C_{29} ; 13 samples showed a relative abundance of n- C_{29} >n- C_{31} >n- C_{27} and 4 samples showed a relative abundance of n- C_{29} >n- C_{27} >n- C_{31} .

The MCN and relative abundance of the three main homologs of long-chain *n*-alkanes varied among surface soil

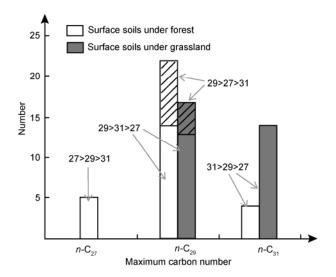


Figure 3 Maximum carbon number and relative abundance of three main homologs $(n\text{-}C_{27}, n\text{-}C_{29}, \text{ and } n\text{-}C_{31})$ of long-chain n-alkanes extracted from 62 surface soil samples collected from across eastern China.

samples from the same location under the same vegetation. At the Damaping site (Figure 1), the bedrock is basalt and the top soil layer is too thin to support the growth of woody plants. Indeed, no woody plants were observed in the study area. Therefore, this location may have been influenced by grassland for a relatively long time. Of the 12 samples collected from this sampling site, 10 were dominated by $n\text{-C}_{31}$ with a relative abundance of $n\text{-C}_{31}$ > $n\text{-C}_{29}$ > $n\text{-C}_{27}$ (HMDA-7 in Figure 2). The remaining two samples were dominated by $n\text{-C}_{29}$ with a relative abundance of $n\text{-C}_{29}$ > $n\text{-C}_{31}$ > $n\text{-C}_{27}$ (HMDA-11 in Figure 2).

Most of the soil samples collected from across eastern China were dominated by n- C_{29} and n- C_{31} (only 5 out of 62 samples were dominated by n- C_{27}). This trend has also been reported in other studies. In a similar smaller study of 26 surface soil samples from China and Mongolia, most samples were dominated by $n-C_{29}$ and $n-C_{31}$ with $n-C_{31}$ as the most frequent MCN, and only few samples were dominated by n-C₂₇ [13]. In another study, n-alkanes of 10 surface soil samples from different climatic and vegetation zones were analyzed. Four from forested locations (three of which were plantation forests) were dominated by n-C₂₉, which were suggested to reflect a mixture of woody and herbaceous source plants, and 6 samples from grassy locations were dominated by n- C_{31} [14]. Long-chain n-alkanes extracted from surface soil in the Heshang Cave, Qingjiang, Hebei Province, were dominated by $n-C_{31}$ [15]. Long-chain n-alkanes extracted from 9 surface soil samples collected from across the Loess Plateau were mainly dominated by n-C₂₉ and n-C₃₁, while only few samples were dominated by n-C₂₇ [16]. All of these surface soil samples were obtained from diverse locations with various overlying vegetation types and plant assemblages across a huge geographical region; however, the long-chain *n*-alkanes extracted from these samples were mainly and consistently dominated by n-C₂₉ and n-C₃₁.

2.2 Long-chain *n*-alkanes extracted from different modern plants

We conducted a systematic evaluation of long-chain nalkane distribution from 334 genera of modern plants reported in published studies [2,5,17-41] and unpublished data (Rao et al. personal unpublished data; Table S2). Most plants are dominated by n-C29 and n-C31, even when they are divided into different groups such as trees, shrubs, and grasses (Table S2). Of the 207 modern grasses, 124 were dominated by n- C_{31} and 39 by n- C_{29} . Of the 101 modern trees, 35 genera showed an MCN reflecting n- C_{31} and 31 showed an MCN reflecting n-C₂₉. Of the 26 modern shrubs, 6 showed an MCN reflecting n-C₃₁ and 15 showed an MCN reflecting n-C₂₉ (Figure 4). Overall, 80% of grasses, 65% of trees, and 80% of shrubs were dominated by either n-C₃₁ or n-C₂₉. Although this is a small sample size compared with the number of plant species worldwide, the data highlights that plants show common characteristics in the molecular distribution of long-chain *n*-alkanes.

2.3 Discussion

The relationship between the composition of long-chain n-alkanes in surface soils and the corresponding vegetation is complex, and there are many sources of uncertainty. It is possible that soils located under grasslands are affected by surrounding forests, and/or by deposition of atmospheric aerosols that act as a source of long-chain n-alkanes [42,43]. Considering the consistent odd-to-even preference and molecular distribution of long-chain n-alkanes from surface soils, it is likely that their variability results from overlying vegetation. For example, the n-alkane distribution in soils mainly dominated by n-C₂₉ and n-C₃₁ is similar to that identified in modern plants, regardless of the type of vegetation and plant.

Several studies on *n*-alkanes in soils have been carried out on red earth in southern China, including 12 Pleistocene samples from Changxing, Zhejiang Province [8], 52 Pleistocene samples from Xuancheng, Anhui Province [7] and

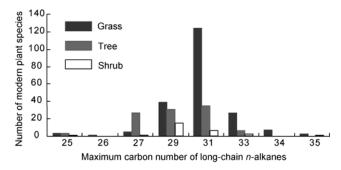


Figure 4 Maximum carbon number of long-chain *n*-alkanes extracted from modern plants, summarized from data in Table S2.

some Pleistocene samples from Xiushui, Jiangxi Province [44]. All of those samples were dominated by $n-C_{29}$ or n-C₃₁. Analyses of samples from the Chinese Loess Plateau in northern China, including the Yuanbao profile in Linxia [45], the Dadiwan profile in Qin'an [5], the Caoxian profile [46], the Xifeng profile [47], and the Luochuan profile [4,6], showed a predominance of n- C_{31} , with only few samples dominated by n- C_{29} . Thus, in the red earth areas in southern China and loess areas in northern China, almost all samples were dominated by n-C₂₉ and n-C₃₁, although it is unclear whether red earth or loess directly affects the n-alkane distribution in surface soils and modern plants in those areas. Together, these data indicate that there is a complex relationship between the molecular distribution of long-chain n-alkanes in surface soil and sediment derived from multiple plant sources and the type of source vegetation. There are several factors that control the molecular distribution of sedimentary n-alkanes, and therefore, affect their relationship with modern vegetation. These factors can be summarized as follows:

(1) Production of *n*-alkanes in plants: This can strongly influence the *n*-alkane profile, because a plant species that produces greater amounts of n-alkanes will bias the distribution of *n*-alkanes in sediments. Thus, the *n*-alkane profile of the sediment will not reflect the dominant vegetation. Differences in n-alkane production among plants in China have been reported [20]. In one study, the concentration of total *n*-alkanes from 93 gramineous plants ranged from 6.9 to 1860 µg/g with an average value of 164.1 µg/g [18]. Therefore, it is possible that the distribution of n-alkanes in sedimentary sequences reflects the plant source(s) with greatest n-alkane production rates, rather than those contributing the most biomass. Indeed, there is no evidence to suggest that the dominant plant shows the highest rate of n-alkanes production. On the contrary, there were no detectable n-alkanes in fresh pine needles of Lodgepole Pine (Pinus contorta), the dominant plant in a Canadian coniferous forest [19]. Likewise, the concentrations of n-alkanes extracted from some coniferous needles from Scandinavian regions were too low to permit isotopic measurement [17].

(2) Mixing effect of different sources: as mentioned above, a mixture of long-chain n-alkanes dominated by n- C_{27} and n- C_{31} may produce a signal indicating dominance of n- C_{29} , as all of the vegetation sources may contain relatively high concentrations of n- C_{29} . This is one possible reason why n- C_{29} is frequently the dominant component in surface soils and sediments. A recent study highlights this, as the molecular distribution of terrestrial n-alkanes in lake sediments was not characteristic of the dominant vegetation around the lake [17]. In that study, the authors analyzed diverse vegetation types and surface lake sediments collected from a wide geographical range (northern Finland to southern Italy). Their results may reflect, at least partly, the mixing process of n-alkanes.

(3) Variation of long-chain *n*-alkanes from the same

plant under different environmental conditions, and postdepositional alteration: Some studies have argued there is no significant variation in the MCN and the molecular distribution of long-chain n-alkanes during the decomposition process [16] or in different growth seasons [15]. However, there is a large body of evidence that variation and alterations do occur [17,19,35,38,39]. Results from Europe indicate that the molecular distributions of *n*-alkanes within the same species vary among different study sites, and the average chain length (ACL) of n-alkanes from deciduous tree leaves increases from northern to southern Europe [17]. Some studies have reported varied molecular distribution of *n*-alkanes from the same species across different study sites (Table S2). Together, these findings indicate that the molecular distribution of *n*-alkanes in modern plants varies significantly depending on the prevailing climatic and environmental conditions.

All of these factors result in a very complex relationship between the molecular distribution of sedimentary long-chain *n*-alkanes and their respective source vegetation. At present, it is uncertain whether it is valid to use molecular distributions of long-chain *n*-alkanes, and particularly the MCN of long-chain *n*-alkanes, as indicators of source vegetation.

3 Conclusions

In this study, analyses of long-chain *n*-alkanes in 62 surface soil samples highlighted that the molecular distribution of long-chain *n*-alkanes of surface soils is highly variable, even among different sites with the same type of vegetation (forest or grassland). However, there were similar molecular distributions of long-chain n-alkanes in surface soils under different vegetation types (forest and grassland). Of the 62 samples, most (57) were dominated by $n-C_{29}$ or $n-C_{31}$ (only 5 were dominated by n- C_{27}), regardless of the type of overlying vegetation. This is consistent with the findings of other surface soil studies [13-16]. More importantly, the patterns of *n*-alkane distribution in more than 100 surface soils are consistent with those reported for more than 300 modern plant species [2,5,17-41], because most modern plants, whether they are trees, grasses, or shrubs, are also dominated by n- C_{29} and n- C_{31} . These results confirm that long-chain n-alkanes extracted from soils or sediments are derived from terrestrial higher plants. However, they cannot accurately predict the source vegetation because of the complex relationship between the molecular distribution of sedimentary long-chain n-alkanes (including both surface soils and sediments) and the type of source vegetation. That is, there is no simple model of interpretation. Therefore, for a given and specific set of long-chain n-alkanes that are dominated by n- C_{29} or n- C_{31} , it is almost impossible to determine whether they are derived from herbaceous or woody plants, or whether they represent forest or grassland vegetation.

This work was supported by the National Natural Science Foundation of China (40901055), the Key Project of the Chinese Ministry of Education (109151), the Fund for Creative Research Groups (41021091), the Cross-discipline Innovative Study Fund for Youth Talent of Lanzhou University (LZUJC2007011) and the National Basic Research Program of China (2010CB950202 and 2010CB833405).

- 1 Sun X J, Song C Q, Wang F Y, et al. Vegetation history of the Loess Plateau of China during the last 100000 years based on pollen data. Quat Int, 1997, 37: 25–36
- 2 Schwark L, Zink K, Lechterbeck L. Reconstruction of postglacial to early Holocene vegetation history in terrestrial Central Europe via cuticular lipid biomarkers and pollen records from lake sediments. Geology, 2002, 30: 463–466
- 3 Lü H Y, Liu D S, Wu N Q, et al. Phytolith record of vegetation succession in the southern Loess Plateau since late Pleistocene (in Chinese). Quat Sci, 1999, 4: 336–349
- 4 Yang M S, Zhang H C, Lei G L, et al. Biomarkers in weakly developed paleosol (L1SS1) in the Luochuan loess section and reconstructed paleovegetation-environment during the interstade of the Last Glaciation (in Chinese). Quat Sci, 2006, 26: 976–984
- 5 Zhong Y X, Chen F H, An C B, et al. Holocene vegetation cover in Qin'an area of western Chinese Loess Plateau revealed by *n*-alkane. Chinese Sci Bull, 2007, 52: 1692–1698
- 6 Zhang H C, Yang M S, Zhang W X, et al. Molecular fossil and pale-ovegetation records of paleosol S₄ and adjacent loess layers in the Luochuan loess section, NW China. Sci China Earth Sci, 2008, 51: 321–330
- 7 Liang B, Xie S C, Gu Y S, et al. Distribution of *n*-alkanes as indicative of paleovegetation change in Pleistocene red earth in Xuancheng, Anhui (in Chinese). J Earth Sci, 2005, 30: 129–132
- 8 Wang Z Y, Yu J H, Gu Y S, et al. Molecular fossils as indicators for paleoenvironment from quaternary red earth in Changxing, Zhejiang (in Chinese). Mar Geol Quat Geol, 2002, 22: 97–102
- 9 Eglinton G, Hamilton R J. Leaf epicuticular waxes. Science, 1967, 156: 1322–1334
- Freeman K H, Hayes J M, Trendel J M, et al. Evidence from carbon isotope measurements for diverse origins of sedimentary hydrocarbons. Nature, 1990, 343: 254–256
- 11 Cranwell P A. Lipid geochemistry of sediments from Upton Broad, a small productive lake. Org Geochem, 1984, 7: 25–37
- Rieley G, Collier R J, Jones D M, et al. The biogeochemistry of Ellesmere Lake, U.K. I: Source correlation of leaf wax inputs to the sedimentary lipid record. Org Geochem, 1991, 17: 901–912
- 13 Wang Y L, Fang X M, Bai Y, et al. Distribution of lipids in modern soils from various regions with continuous climate (moisture-heat) change in China and their climate significance. Sci China Earth Sci, 2007, 50: 600-612
- 14 Wang Z Y, Liu Z H, Yi Y, et al. Features of lipids and their significance in modern soils from various climate-vegetation regions (in Chinese). Acta Pedol Sin, 2003, 40: 967–970
- Cui J W, Huang J H, Pu Y, et al. Comparison of lipid compositions between plant leaves and overlying soil in Heshang Cave, Qingjiang, Hubei Province and its significance (in Chinese). Quat Sci, 2007, 28: 35–42
- 16 Zhong Y X, Xue Q, Chen F H. n-alkane distributions in modern vegetation and surface soil from western Loess Plateau (in Chinese). Quat Sci, 2009, 29: 767–773
- 17 Sachse D, Radke J, Gleixner G. δD values of individual n-alkanes from terrestrial plants along a climatic gradient—Implications for the sedimentary biomarker record. Org Geochem, 2006, 37: 469–483
- 18 Maffei M. Chemotaxonomic significance of leaf wax alkanes in the Gramineae. Biochem Syst Ecol, 1996, 24: 53–64
- Otto A, Simpson M J. Degradation and preservation of vascular plant-derived biomarkers in grassland and forest soils from Western Canada. Biogeochemistry, 2005, 74: 377–409
- 20 Cui J W, Huang J H, Xie S C. Characteristics of seasonal variations

- of leaf *n*-alkanes and *n*-alkenes in modern higher plants in Qingjiang, Hubei Province, China. Chinese Sci Bull, 2008, 53: 2659–2664
- 21 Bi X H, Sheng G Y, Liu X H, et al. Molecular and carbon and hydrogen isotopic composition of *n*-alkanes in plant leaf waxes. Org Geochem, 2005, 36: 1405–1417
- 22 Chikaraishi Y, Naraoka H. Compound-specific δ D- δ ¹³C analyses of n-alkanes extracted from terrestrial and aquatic plants. Phytochemistry, 2003, 63: 361–371
- 23 Chikaraishi Y, Naraoka H, Poulson S R. Carbon and hydrogen isotopic fractionation during lipid biosynthesis in a higher plant (*Cryptomeria japonica*). Phytochemistry, 2004, 65: 323–330
- 24 Conte M H, Weber J C, Carlson P J, et al. Molecular and carbon isotopic composition of leaf wax in vegetation and aerosols in a northern prairie ecosystem. Oecologia, 2003, 135: 67–77
- 25 Gormann R, Schreiber L, Kolodziej H. Cuticular wax profiles of leaves of some traditionally used African Bignoniaceae. Z Naturforsch, 2004, 59: 631–635
- 26 Huang Y, Eglinton G, Ineson P, et al. Absence of carbon isotope fractionation of individual *n*-alkanes in a 23-year field decomposition experiment with *Calluna valgaris*. Org Geochem, 1997, 26: 497–501
- 27 Jansen B, Nierop K, Hageman J, et al. The straight-chain lipid biomarker composition of plant species responsible for the dominant biomass production along two altitudinal transects in the Ecuadorian Andes. Org Geochem, 2006, 37: 1514–1536
- 28 Kawamura K, Ishimura Y, Yamazaki K. Four years' observations of terrestrial lipid class compounds in marine aerosols from the western North Pacific. Glob Biogeochem Cycle, 2003, 17: 1003, doi: 10.1029 /2001GB001810
- 29 Krull E, Sachse D, Mügler I, et al. Compound-specific δ^{13} C and δ^{2} H analyses of plant and soil organic matter: A preliminary assessment of the effects of vegetation change on ecosystem hydrology. Soil Biol Biochem, 2006, 38: 3211–3221
- 30 Li B C, Dong Y L, Li C, et al. Distribution and compound specific carbon isotope of individual long chain alkanes from leaves of *Kan-delia Candel* and *Ficus Microcarpa* and their photosynthesis (in Chinese). J Trop Oceanol, 2003, 22: 62–69
- 31 Liu W G, Huang Y S. Compound-specific D/H ratios and molecular distributions of higher plant leaf waxes as novel paleoenvironmental indicators in the Chinese Loess Plateau. Org Geochem, 2005, 36: 851–860
- 32 Lockheart M J, Van Bergen P F, Evershed R P. Variations in the stable carbon isotope compositions of individual lipids from the leaves of modern angiosperms: Implications for the study of higher land plant-derived sedimentary organic matter. Org Geochem, 1997, 26: 137–153
- 33 Lockheart M J, Van Bergen P F, Evershed R P. Chemotaxonomic classification of fossil leaves from the Miocene Clarkia lake deposit, Idaho, USA based on *n*-alkyl lipid distributions and principal component analyses. Org Geochem, 2000, 31: 1223–1246
- 34 Marseille F, Disnar J, Guillet B, et al. n-Alkanes and free fatty acids in humus and A1 horizons of soils under beech, spruce and grass in the Massif-Central (Mont-Lozère), France. Eur J Soil Sci, 1999, 50: 433–441
- 35 Mügler I, Sachse D, Werner M, et al. Effect of lake evaporation on δD values of lacustrine n-alkanes: A comparison of Nam Co (Tibetan Plateau) and Holzmaar (Germany). Org Geochem, 2008, 39: 711–729
- 36 Nichols J E, Booth R K, Jackson S T, et al. Paleohydrologic reconstruction based on *n*-alkane distributions in ombrotrophic peat. Org Geochem, 2006, 37: 1505–1513
- 37 Nott C J, Xie S C, Avsejs L A, et al. n-alkane distributions in ombrotrophic mires as indicators of vegetation change related to climatic variation. Org Geochem, 2000, 31: 231–235
- Rommerskirchen F, Plader A, Eglinton G, et al. Chemotaxonomic significance of distribution and stable carbon isotopic composition of long-chain alkanes and alkan-1-ols in C₄ grass waxes. Org Geochem, 2006, 37: 1303–1332
- 39 Sessions A L. Seasonal changes in D/H fractionation accompanying lipid biosynthesis in Spartina alterniflora. Geochim Cosmochim Acta,

- 2006, 70: 2153-2162
- Wiesenberg G L B, Schmidt M W I, Schwark L. Plant and soil lipid modifications under elevated atmospheric CO₂ conditions: I. Lipid distribution patterns. Org Geochem, 2008, 39: 91–102
- 41 Xie S C, Nott C J, Avsejs L A, et al. Molecular and isotopic stratigraphy in ombrotrophic mire for paleoclimate reconstruction. Geochim Cosmochim Acta, 2004, 68: 2849–2862
- 42 Guo F Q, Zhang C J, Zhang Y, et al. Distribution of *n*-alkane and environmental significance about the Lanzhou atmospheric dust (in Chinese). Environ Sci Technol, 2009, 32: 9–11
- 43 Xie M J, Wang G H, Hu S Y, et al. Aliphatic alkanes and polycyclic aromatic hydrocarbons in atmospheric PM₁₀ aerosols from Baoji, China: Implications for coal burning. Atmos Res, 2009, 93: 840–848
- 44 Xie S C, Yi Y, Liu Y Y, et al. The Pleistocene vermicular red earth in

- South China signaling the global climatic change: The molecular fossil record. Sci China Earth Sci, 2003, 46: 1114–1120
- 45 Wang Z Y, Xie S C, Chen F H. n-Alkane distribution as indicator for paleo-vegetation: An example from Yuanbao S₁ paleosol in Linxia, Gansu Province (in Chinese). Quat Sci, 2004, 24: 231–235
- 46 Zhong Y X. Environmental change in western Chinese Loess Plateau since the Last Glacial: Organic geochemical records in Loess (in Chinese). Dissertation for the Doctoral Degree. Lanzhou: Lanzhou University, 2008. 60–90
- 47 Liu W G, Huang Y S, An Z S, et al. Summer monsoon intensity controls C₄/C₃ plants abundanceduring the last 35 ka in the Chinese Loess Plateau: Carbon isotope evidence from bulk organic matter and individual leaf waxes. Palaeogeogr Palaeoclimatol Palaeoecol, 2005, 220: 243–254

Open Access This article is distributed under the terms of the Creative Commons Attribution License which permits any use, distribution, and reproduction in any medium, provided the original author(s) and source are credited.

Supporting Information

- Table S1 Detailed information on 62 surface soil samples collected from across eastern China, and composition of long-chain n-alkanes
- **Table S2** Maximum carbon number of long-chain *n*-alkanes extracted from modern plants and references

The supporting information is available online at csb.scichina.com and www.springerlink.com. The supporting materials are published as submitted, without typesetting or editing. The responsibility for scientific accuracy and content remains entirely with the authors.