



Comparative effectiveness of organic and inorganic amendments on cadmium bioavailability and uptake by *Pelargonium hortorum*

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Abstract

Purpose The aim of this study was to compare organic and inorganic amendments namely citric acid, ammonium nitrate, compost, and titanium dioxide nanoparticles (TiO₂ NPs) with ethylenediaminetetraacetic acid (EDTA) in enhancing the bioavailability of cadmium (Cd) in assisted phytoextraction.

Materials and methods Uncontaminated soil was spiked with different levels of Cd (25–150 mg kg⁻¹) by using CdSO₄ salt. Different levels of five amendments used were as follows: EDTA (0–5 mmol kg⁻¹), citric acid (0–10 mmol kg⁻¹), ammonium nitrate (0–10 mmol kg⁻¹), TiO₂ NPs (0–100 mg kg⁻¹), and compost (0–10%). *Pelargonium hortorum* plants were grown on amended soils for a period of 6 months and different parameter were considered to evaluate bioavailability of Cd upon application of amendments.

Results and discussion The bioavailability of Cd in soil was increased by 1.2-, 0.8-, 0.4-, and 0.2-fold in EDTA, citric acid, ammonium nitrate, and TiO₂ NPs, respectively. However, Cd bioavailability was decreased by 0.5-fold in compost-amended soils. The efficiency of amendments for mobilizing Cd followed the order: EDTA > citric acid > ammonium nitrate > TiO₂ NPs > compost. The maximum accumulations of Cd in root (350 mg Cd kg⁻¹) and shoot (943 mg Cd kg⁻¹) were observed upon EDTA application and minimum concentrations in root (125 mg Cd kg⁻¹) and shoot (104 mg Cd kg⁻¹) were observed in compost-amended soils. The maximum (2.7 g) and minimum (1.5 g) plant biomass was observed upon compost and EDTA application, respectively. The maximum Cd uptake per plant (1.44 mg) and metal extraction ratio (4.4%) were observed in citric acid-amended soils due to better biomass produced.

Conclusions Among all amendments, citric acid can be recommended as an environmentally friendly and effective substitute to EDTA for assisted phytoextraction of Cd to decontaminate polluted soil as Cd uptake per plant and metal extraction ratio were the highest upon application of citric acid.

Keywords Ammonium nitrate · Bioavailability · Citric acid · Compost · EDTA · Metal extraction ratio · TiO₂ NPs

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1 Introduction

Cadmium (Cd) is naturally found trace element and is being introduced in environment through various natural and anthropogenic activities (Khan et al. 2015; Gul et al. 2018a). Only 10% of Cd contamination is through natural activities and the remaining 90% is contributed by human activities (Khan et al. 2017). Among anthropogenic activities, fossil fuel combustion, phosphate fertilizers, industrial and municipal waste, metal smelting, and zinc mining (Guo et al. 2017; Abbas et al. 2018) have been the main sources of Cd contamination. Unlike organics, Cd is persistent, toxic, and non-biodegradable (Chhajro et al. 2016; Guo et al. 2017; Gul et al. 2018b), and high concentrations affect soil microorganisms, plant, and ultimately human through food chain (Li et al.

2016). Keeping in view the adverse effects of Cd on environment and human health, the remediation of Cd contaminated soil is required.

Various soil remediation techniques have been developed, but phytoremediation is accepted by public due to its aesthetic values and is also a cost-effective, environment-friendly technique (Souza et al. 2017). The success and efficiency of phytoremediation mainly depend on (a) the ability of plant to uptake metals from soil and translocate to shoot and (b) availability of metals in soil for plant uptake (Dede and Ozdemir 2016; Gul et al. 2018a). Various *Pelargonium* cultivars have inherent ability to accumulate high amount of lead (Pb) (Arshad et al. 2008, 2016; Manzoor et al. 2018) and Cd (Gul et al. 2018a, b) in shoot. Among all cultivars, *Pelargonium hortorum* is considered as Pb and Cd hyperaccumulator (Gul et al. 2018b). *P. hortorum* is scented geranium belonging to family Geraniaceae. It normally grows in areas with low precipitation and humidity and requires well-drained soil. *P. hortorum* has the capability to uptake soluble Cd through roots and translocate to shoots (Gul et al. 2018a, b). Therefore, bioavailability of metals is a serious limitation of phytoremediation. Metals are in insoluble form and they are not readily available for plant uptake. Chelating agents such as ethylenediaminetetraacetic acid (EDTA), nitrilotriacetate (NTA), diethylenetriaminepentaacetic acid (DTPA), *N*-hydroxy ethylenediaminetriacetic acid (HEDTA), ethylenediaminedisuccinate (EDDS), oxalic acid, and citric acid have been used to enhance the mobility of metals (Sarkar et al. 2008; Freitas et al. 2014; Shahid et al. 2014; Pedron et al. 2014; Gul et al. 2018a, b).

Most studies on chelate-induced phytoextraction have focused on the use of EDTA as a chelating agent. EDTA and formed complexes have low biodegradability and high solubility in soil, resulting in high risk of adverse environmental effects due to metal mobilization (Alkorta et al. 2004). In this respect, the amount of Cd taken up by plants has been reported to be much smaller than the amount of Cd mobilized from the soil during EDTA-induced Cd phytoextraction (Chen et al. 2004). Different researchers have shown that addition of EDTA increases the accumulation of Cd and other heavy metals in plants but the adverse effects of EDTA to soil enzymatic activities, soil microbes, and plants have also been reported (Epelde et al. 2008; Muhlbachova 2011; Johnson et al. 2010). Chen et al. (2004) reported that 3.5 and 20.6% of soil Pb and Cd, respectively, were leached from the soil after the application of 5 mmol kg⁻¹ of EDTA.

EDTA degrades very slowly and remains in soil after 5 to 6 months of application (Neugschwandtner et al. 2012). Therefore, the substitute of EDTA in enhancing the phytoextraction of Cd is required. Various other soil amendments such as citric acid, compost, and ammonium nitrate are commonly used to enhance the crop growth and yield. It is also reported that these amendments are

helpful in the uptake of metals from the contaminated sites (Chen et al. 2017; Gul et al. 2018a). Recently, researchers focused on the use of TiO₂ NPs to enhance the nutrient uptake and improve crop yield (Zahra et al. 2017; Rizwan et al. 2019). But the impact of TiO₂ NPs on the mobility of heavy metals is not studied in comparison to different amendments.

Therefore, the present study was designed to compare different organic and inorganic amendments with the commonly used non-biodegradable chelating agent, i.e., EDTA, for increasing Cd bioavailability and uptake by *P. hortorum*. The specific objectives were to (a) compare the effectiveness of organic and inorganic amendments on the mobility of Cd in soil and (b) compare soil amendments for enhanced accumulation of Cd by *P. hortorum*.

2 Materials and methods

2.1 Soil preparation and analysis

Uncontaminated surface soil (0–15 cm) of clay loam texture having pH 7.39, electrical conductivity 0.33 mS/cm, and organic matter 0.38% was collected from the nursery area of National University of Sciences and Technology (NUST; 33.6426° N, 72.9929° E), Islamabad, Pakistan. Detailed characteristics are presented in Table S1 (ESM). The soil did not contain Cd. Area falls in semiarid climatic zone with moderate winters and summer. Soil of the area is derived from wind deposits and sedimentary rocks. Soil samples were air-dried, crushed, and sieved to remove undecomposed organic materials and rocks and stored in containers. All containers were properly labeled and soil was spiked with four levels of Cd (25, 50, 100, 150 mg kg⁻¹) by using cadmium sulfate (CdSO₄) solution. Soil was mixed thoroughly at regular intervals for homogenization and after metal stabilization; soil was used for further experiments.

2.2 Soil amendments

Different organic and inorganic soil amendments such as citric acid, compost, ammonium nitrate, and TiO₂ NPs were tested and compared with EDTA to increase the bioavailability of Cd. For the experimental work, different doses of EDTA (0–5 mmol kg⁻¹), citric acid (0–10 mmol kg⁻¹), ammonium nitrate (0–10 mmol kg⁻¹), TiO₂ NPs (0–100 mg kg⁻¹), and compost (0–10%) were used and different controls (without Cd, without Cd with addition of amendments, with Cd without addition of any amendments) were maintained for the comparisons.

2.3 Titania nanoparticle preparation and characterization

Titania nanoparticles were prepared by using titanium isopropoxide ($C_{12}H_{28}O_4Ti$), ethanol (C_2H_6O), distilled water, and hydrochloric acid (HCl) in a ratio 1:15:60:0.2 (Mehrizad et al. 2009). All the chemicals were added in water and stirred for 48 h at room temperature on hot plate. After stirring, solutions were kept in an oven at 90 °C for 48 h and golden yellow crystals were obtained. The crystals were powdered and calcinated at 400 °C for 2 h in muffle furnace (NEYO M-525 SERIES II). The crystallite phase and size of the prepared TiO_2 NPs were identified using X-ray diffractometer (XRD). The TiO_2 NPs were in anatase phase having average crystalline size of 13 nm. Scanning electron microscopy (SEM; JSM 6490 A, Jeol Ltd., Japan) was used to determine the particle size and surface morphology. Average particle size of TiO_2 NPs was 26.5 nm and the particles were spherical in shape (Fig. S1, Electronic Supplementary Material (ESM)).

2.4 Incubation experiments and soil extraction

Incubation experiments were carried out to evaluate changes in solubility of Cd in soil by the addition of different organic and inorganic amendments. Briefly, 10 g of spiked soil (60% WHC) was taken in acid-cleaned petri dishes, and specified amounts of EDTA, citric acid, ammonium nitrate, TiO_2 NP solution, and solid compost were added separately. For the control, 3 mL of distilled water was added and all treatments were incubated for 7 days at 28 °C. After incubation, samples were dried overnight in the oven at 65 °C (Shazia et al. 2014). The soluble fraction of Cd was extracted by taking 10 g of sample in a flask and 50 mL of 0.01 M $CaCl_2$ solution (1:5) and shaken for 2 h at 200 rpm. After shaking, the samples were filtered through Whatman filter paper no. 42 and filtrate was analyzed for Cd concentration by atomic absorption spectrophotometer (AAS, PerkinElmer 900T).

2.5 Culture experiments for phyto-accumulation of Cd in amended soils

Pot experiments were carried out to investigate the impact of amendments on Cd accumulation in *P. hortorum*. Pots were filled with 1 kg of Cd spiked and control (without Cd) soils. One-month-old seedlings of *P. hortorum* of approximately equal size were transferred to each pot. After 1 month of seedling transplantation, EDTA, citric acid, ammonia nitrate, and TiO_2 NPs and compost were applied. The experiments were carried out in greenhouse, illuminated with natural light, and pots were watered regularly to maintain ambient moisture level. The average temperature and humidity for the experimental period were 22.6 °C and 63%, respectively. The

maximum and minimum temperature was observed for the months of April and January, respectively. Maximum and minimum humidity was observed in the month of February and November, respectively. Plants were grown for the period of 6 months, and after this exposure time, they were carefully harvested.

2.6 Plant harvesting and analysis

Plants were removed, washed with 0.01 M HCl solution, and rinsed several times with distilled water to remove any external Cd. For fresh biomass, plants were divided in root and shoot, and fresh biomass was measured. After that, plants were dried in the oven at 65 °C for 48 h and dry biomass was measured. For the analysis of Cd, the root and shoot were digested in a mixture of HNO_3 and $HClO_4$ (3:1) and Cd content was determined through an AAS (PerkinElmer 900T).

2.7 Cd uptake, metal extraction ratio, and translocation factor

Uptake of Cd by plant is the total amount of Cd accumulated in whole plant, and Cd metal extraction ratio (MER) is the accumulation of Cd in shoot to that of soil. Translocation factor (TF) is the transfer of Cd from the root to shoot. These are very important parameters in the phytoextraction. Both parameters were calculated by using following formulae:

$$Cd \text{ uptake (mg plant}^{-1}) = [Cd_{shoot} \times SDW] + [Cd_{root} \times RDW] \quad (1)$$

where Cd_{shoot} and Cd_{root} are the concentrations of Cd in shoot and root, respectively, and SDW and RDW are the dry weights of shoot and root, respectively.

$$MER (\%) = [Cd_{shoot} \times W_{shoot}] / [Cd_{soil} \times W_{soil}] \times 100 \quad (2)$$

where Cd_{shoot} and Cd_{soil} are the concentrations of Cd in shoot and soil and W_{shoot} and W_{soil} are the dry weights of shoot and soil, respectively (Mertens et al. 2005).

$$TF = Cd_{shoot} / Cd_{root} \quad (3)$$

where Cd_{shoot} and Cd_{root} are the Cd concentrations in shoot and root, respectively (Zahedifar et al. 2016).

2.8 Statistical analysis

Completely randomized design (CRD) with three replicates for each treatment was used for experimental setup. Analysis of variance (ANOVA) was performed by Statistix 10.0 software followed by Tukey's post hoc test between the means of treatments for significance difference ($P < 0.05$).

3 Results

3.1 Influence of amendments on bioavailability of Cd in soil

Efficiencies of all the soil amendments for Cd release were different (Table 1). The application of amendments enhanced the bioavailability of Cd in soil except compost which showed a decrease in the bioavailability of Cd as compared to the control. Without application of any amendment, maximum 11.22 mg Cd kg⁻¹ was observed in 150 mg Cd kg⁻¹ treatment. The application of amendments improved the bioavailability of Cd in soil. The maximum bioavailable portion of Cd was observed upon 5 mmol kg⁻¹ EDTA application. Bioavailability of Cd in 150 mg Cd kg⁻¹ treatment was significantly (*P* < 0.05) increased by 1.2-, 0.8-, 0.4-, and 0.2-fold at 5 mmol kg⁻¹ EDTA, 10 mmol kg⁻¹ citric acid, 10 mmol kg⁻¹ ammonium nitrate, and 100 mg kg⁻¹ TiO₂ NP application, respectively, as compared to treatment without application of amendments. However, the application of 10% compost significantly reduced Cd bioavailability by 0.5-fold. The efficiency of amendments in

increasing the mobility of Cd in soil followed the sequence EDTA > citric acid > ammonium nitrate > TiO₂ NPs > compost.

3.2 Phyto-accumulation of Cd upon soil amendment application

3.2.1 Cd concentration in shoot

P. hortorum accumulated 38.1% Cd in shoot at 150 mg Cd kg⁻¹ treatment as compared to 25 mg Cd kg⁻¹ without application of amendments (Fig. 1). The application of amendments increased Cd concentration in shoot except for compost which reduced the accumulation. At 150 mg Cd kg⁻¹ treatment, Cd accumulation in shoot was increased by 3.9-, 2.4-, 1.0-, and 0.2-fold, upon 4 mmol kg⁻¹ EDTA, 10 mmol kg⁻¹ citric acid, 10 mmol kg⁻¹ ammonium nitrate, and 100 mg kg⁻¹ TiO₂ NPs, respectively, as compared to 25 mg Cd kg⁻¹ treatment without the application of amendments. However, application of 10% compost significantly (*P* < 0.05) decreased Cd concentration in shoot by 0.3-fold (Fig. 1). The maximum accumulation of Cd in shoot of *P. hortorum* was observed at EDTA application

Table 1 Phytoavailability of Cd in spiked and amended soils

Phyto-available Cd in spiked soils (mg kg ⁻¹)		Cd Treatments (mg kg ⁻¹)			
Amendment levels		25	50	100	150
EDTA (mmol kg ⁻¹)	0	1.7 ± 1.1 ^{jk}	3.9 ± 0.6 ^{ij}	7.5 ± 2.6 ^h	11.2 ± 1.1 ^{fg}
	2	7.4 ± 1.0 ^{hi}	8.5 ± 1.1 ^{gh}	11.1 ± 3.2 ^{fg}	16.4 ± 0.8 ^{b-d}
	4	12.8 ± 0.6 ^{ef}	13.4 ± 2.5 ^{d-f}	15.0 ± 4.4 ^{de}	18.1 ± 0.5 ^{bc}
	5	14.6 ± 0.5 ^{de}	15.3 ± 2.8 ^{c-e}	18.5 ± 2.3 ^b	25.7 ± 0.7 ^a
	10	17.1 ± 1.1 ^k	19.9 ± 0.9 ^{ij}	27.5 ± 2.6 ^{gh}	38.1 ± 1.1 ^{de}
Citric acid (mmol kg ⁻¹)	0	1.7 ± 1.1 ^{jk}	3.9 ± 0.9 ^{ij}	7.5 ± 2.6 ^{gh}	11.2 ± 1.1 ^{de}
	2	3.8 ± 0.7 ^{ij}	4.5 ± .5 ⁱ	9.2 ± 0.6 ^{e-g}	14.3 ± 1.7 ^c
	6	6.8 ± 1.7 ^h	8.6 ± 2.2 ^{f-h}	13.9 ± 1.2 ^c	17.3 ± 1.2 ^b
	10	9.9 ± 1.4 ^{ef}	13.5 ± .7 ^{cd}	17.6 ± 2.1 ^b	20.3 ± 1.6 ^a
	100	1.7 ± 1.2 ^{hi}	3.9 ± 0.9 ^{f-h}	7.5 ± 2.7 ^{de}	11.2 ± 1.1 ^{a-c}
Titanium nanoparticles (mg kg ⁻¹)	0	1.7 ± 1.2 ^{hi}	3.9 ± 0.9 ^{f-h}	7.5 ± 2.7 ^{de}	11.2 ± 1.1 ^{a-c}
	20	2.1 ± 1.2 ^{g-i}	3.1 ± 1.3 ^{f-h}	8.1 ± 1.6 ^d	11.5 ± 2.7 ^{a-c}
	60	2.8 ± 0.4 ^{f-h}	3.9 ± 0.9 ^{f-h}	8.9 ± 1.5 ^{cd}	12.2 ± 1.8 ^{ab}
	100	4.6 ± 1.4 ^{fg}	5.2 ± 2.1 ^{ef}	9.5 ± 3.9 ^{b-d}	13.6 ± 2.5 ^a
	Compost (%)	0	1.7 ± 1.2 ^{h-j}	3.9 ± 1.0 ^{e-g}	7.5 ± 2.7 ^{bc}
2		1.4 ± 0.6 ^{ij}	3.4 ± 1.0 ^{f-h}	6.9 ± 0.5 ^{bc}	10.5 ± 3.6 ^a
6		1.2 ± 0.6 ^{ij}	3.0 ± 0.3 ^{f-i}	5.9 ± 0.3 ^{cd}	8.4 ± 1.0 ^b
10		1.1 ± 0.5 ^{ij}	1.9 ± 0.3 ^{g-j}	4.6 ± 0.7 ^{d-f}	5.7 ± 1.1 ^{c-e}
Ammonium nitrate (mmol kg ⁻¹)		0	1.7 ± 1.2 ^{hi}	3.9 ± 1.0 ^{gh}	7.5 ± 2.7 ^{de}
	2	3.1 ± 1.4 ^{g-i}	4.3 ± 1.0 ^{f-h}	8.4 ± 2.2 ^{c-e}	12.8 ± 2.9 ^{ab}
	6	5.7 ± 2.0 ^{e-g}	7.4 ± 1.9 ^{d-f}	9.7 ± 3.3 ^{cd}	15.0 ± 4.5 ^a
	10	7.6 ± 2.5 ^{de}	9.5 ± 1.8 ^{cd}	13.4 ± 1.8 ^{ab}	16.3 ± 2.3 ^a

Data is the mean (±SD) of three replicates. Superscript letters a–k show the significant difference (*P* < 0.05, Tukey's Honestly Significant Difference Test) among all groups

followed by citric acid, ammonium nitrate, and TiO₂ NPs. In comparison with EDTA, Cd accumulation was significantly reduced in citric acid (31.6%), ammonium nitrate (59.7%), TiO₂ NPs (70.3%), and compost (86.7%) -amended soils.

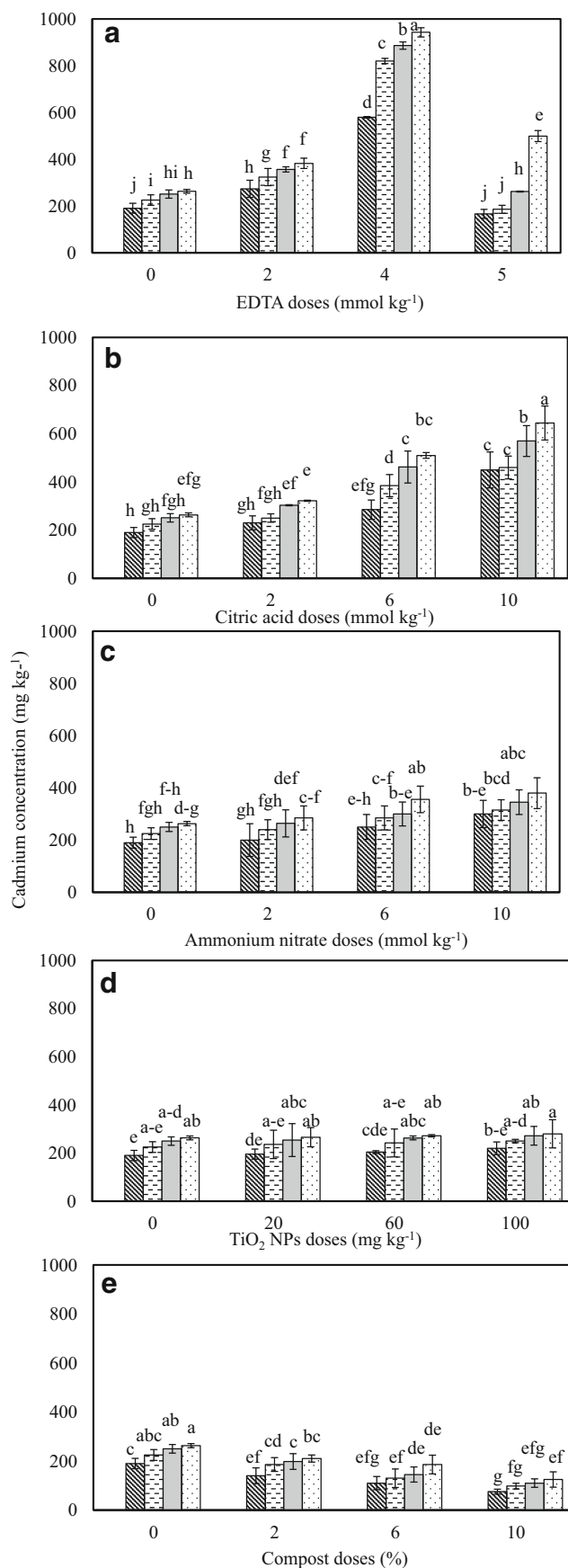
3.2.2 Cd concentration in root

Figure 2 illustrates the concentration of Cd in roots. *P. hortorum* has the ability to accumulate Cd in roots without addition of any amendment, and concentration of Cd in roots increased with increasing levels of Cd. *P. hortorum* accumulated 67.6% more Cd in roots at 150 mg Cd kg⁻¹ treatment as compared to 25 mg Cd kg⁻¹ without application of amendments. The increase in Cd accumulation in roots of *P. hortorum* was observed upon amendment application. *P. hortorum* showed 2.3-, 1.5-, 1.0-, and 0.9-fold increase in Cd accumulation in root upon application of 4 mmol kg⁻¹ EDTA, 10 mmol kg⁻¹ citric acid, 10 mmol kg⁻¹ ammonium nitrate, and 100 mg kg⁻¹ TiO₂ NPs, respectively, at 150 mg Cd kg⁻¹ treatment as compared to plant grown in 25 mg Cd kg⁻¹ treatment without the application of amendments. On the other hand, addition of 10% compost decreased Cd accumulation by 0.92-fold. The efficiency of all amendments for Cd accumulation in roots follows the sequence: EDTA > citric acid > ammonium nitrate > TiO₂ NPs > compost.

3.2.3 Plant biomass

Dry plant biomass of *P. hortorum* decreased with increasing levels of Cd without application of amendments (Table 2). The maximum dry plant biomass was observed in control group. At 150 mg Cd kg⁻¹ treatment, the biomass was decreased by 22.8%. Upon application of soil amendments, the dry plant biomass of *P. hortorum* decreased with increasing levels of EDTA. However, for the other amendments, the plant dry biomass increased as the levels of amendments increased in control soil (without Cd). Addition of 5 mmol kg⁻¹ EDTA in control group decreased the dry plant biomass by 62%. However, the application of maximum level of citric acid, ammonium nitrate, TiO₂ NPs, and compost increased the dry plant biomass by 10, 11, 12, and 14%, respectively. By comparing combined effects of Cd treatments and amendments, the dry plant biomass was significantly decreased by 46.1, 15.1, 11.9, 5.6, and 4.9% upon the application of maximum level of EDTA, citric acid, ammonium nitrate, TiO₂ NPs, and compost, respectively, at 150 mg Cd kg⁻¹ as compared to control (without Cd and amendments).

Fig. 1 Cadmium accumulation in shoots of *P. hortorum* in Cd-spiked and amended soils. **a**) EDTA. **b**) Citric acid. **c**) Ammonium nitrate. **d**) TiO₂ NPs. **e**) Compost (▨ = 25 mg Cd kg⁻¹; ▤ = 50 mg Cd kg⁻¹; ▥ = 100 mg Cd kg⁻¹; ▦ = 150 mg Cd kg⁻¹)



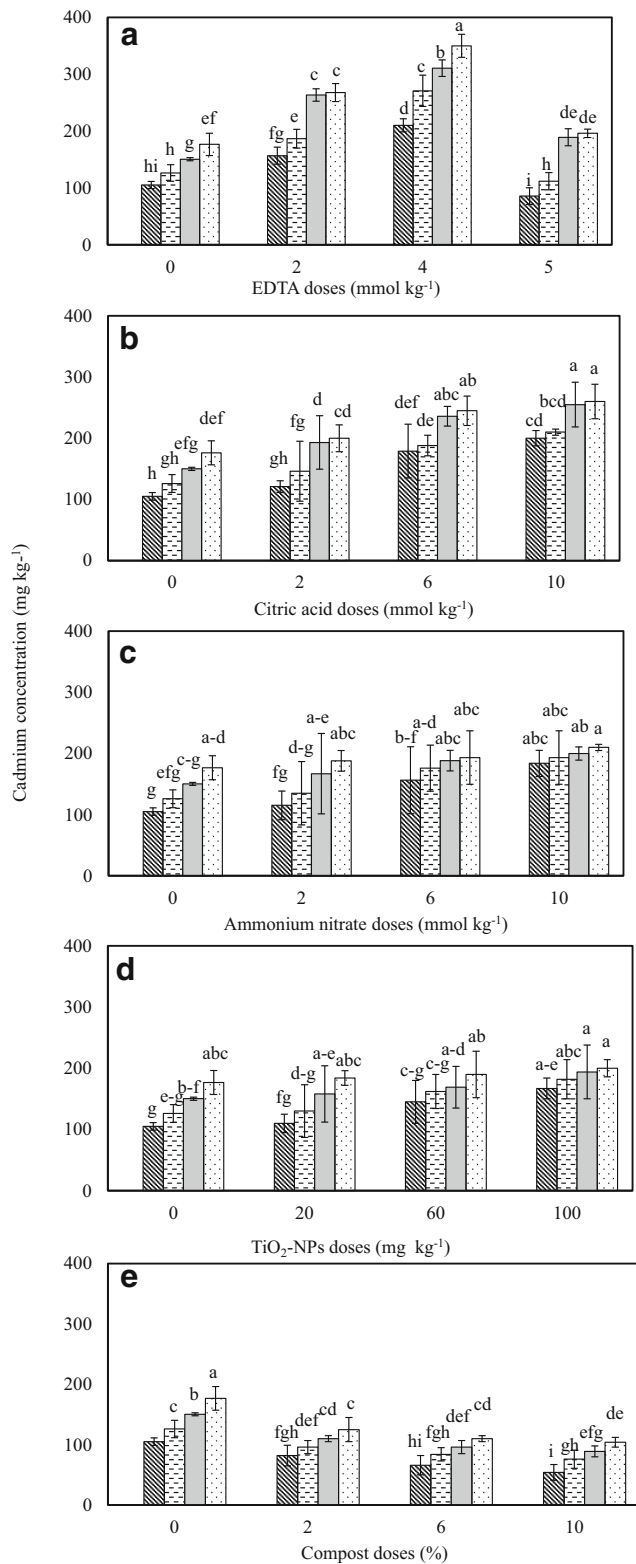


Fig. 2 Concentration of Cd in roots of *P. hortorum* in Cd spiked and organic and inorganic amended soil. **a)** EDTA. **b)** Citric acid. **c)** Ammonium nitrate. **d)** TiO₂ NPs. **e)** Compost (▨ = 25 mg Cd kg⁻¹; ▩ = 50 mg Cd kg⁻¹; ▧ = 100 mg Cd kg⁻¹; ▦ = 150 mg Cd kg⁻¹)

3.2.4 Cadmium uptake per plant

Cd uptake by *P. hortorum* in amendment-mediated soil is shown in Fig. 3. Without the application of soil amendments, the maximum (0.57 mg) and minimum (0.48 mg) Cd uptake was observed in 100 and 25 mg Cd kg⁻¹ treatment, respectively. The application of amendments increased Cd uptake with increasing levels except compost which showed decreasing trend. Among all amendments, the maximum Cd uptake was observed in citric acid-amended soil followed by EDTA, ammonium nitrate, and TiO₂ NPs. The Cd uptake in citric acid-, EDTA-, ammonium nitrate-, and TiO₂ NP-amended soils was increased by 2-, 1.9-, 0.9-, and 0.5-fold, respectively, at the 150-mg Cd kg⁻¹ treatment as compared to 25 mg Cd kg⁻¹ level without application of amendments. On the other hand, the addition of compost reduced Cd uptake by 0.3-fold.

3.2.5 Metal extraction ratio

The metal extraction ratio of *P. hortorum* grown in Cd treatment without the application of amendments decreased with increasing levels of Cd (Fig. 4). The maximum MER of 1.8% was observed in 25 mg Cd kg⁻¹ treatment without the application of amendments. The application of EDTA, citric acid, ammonium nitrate, and TiO₂ NPs increased MER as the dose of amendments increased at same Cd treatment level. The maximum MER of 4.4% was observed upon 10 mmol kg⁻¹ citric acid application in 25 mg Cd kg⁻¹ treatment followed by EDTA (4.3%), ammonium nitrate (3.1%), TiO₂ NPs (2.2%), and compost (0.8%). The MER of *P. hortorum* decreased with increasing levels of Cd and amendments. The MER of *P. hortorum* was decreased by 46, 50, 72, 75, and 88% at 150 mg Cd kg⁻¹ upon application of higher levels of EDTA, citric acid, ammonium nitrate, TiO₂ NPs, and compost, respectively.

3.2.6 Translocation factor

Table 3 illustrates the TF of *P. hortorum* upon amendment application in Cd-spiked soils. TF of *P. hortorum* was > 1 in Cd-spiked soils without amendment application. In the present study, with increasing levels of Cd (from 25 to 150 mg kg⁻¹), TF decreased, but at all levels, it was > 1. The application of soil amendments increased the TF, and with increasing levels of amendments and Cd, transfer of Cd from root to shoot increased further. The maximum TF (3.05) was observed at 50 mg Cd kg⁻¹ in combination with 4 mmol kg⁻¹ EDTA and minimum TF (1.19) was observed at 150 mg Cd kg⁻¹ in combination with 10% compost. TF values greater than 1 highlighted the translocation of Cd from roots to shoots.

Table 2 Dry plant biomass of *P. hortorum* in Cd spiked and amendments mediated soil

Dry plant biomass of <i>P. hortorum</i> in Cd spiked soils (g)						
Cd treatments (mg kg ⁻¹)	▶	0	25	50	100	150
Amendment levels	▼					
EDTA (mmol kg ⁻¹)	0	2.84 ± 0.5 ^a	2.66 ± 0.3 ^{a-c}	2.50 ± 0.7 ^{a-d}	2.36 ± 0.6 ^{a-d}	2.19 ± 0.2 ^{a-d}
	2	2.75 ± 0.6 ^{ab}	2.43 ± 1.2 ^{a-d}	2.13 ± 0.2 ^{a-d}	1.88 ± 0.5 ^{a-d}	1.63 ± 0.2 ^{cd}
	4	2.31 ± 0.8 ^{a-d}	2.06 ± 0.5 ^{a-d}	1.80 ± 0.6 ^{b-d}	1.69 ± 0.4 ^{cd}	1.60 ± 0.1 ^d
	5	2.22 ± 1.2 ^{a-d}	1.90 ± 0.6 ^{a-d}	1.78 ± 0.5 ^{b-d}	1.63 ± 0.7 ^{cd}	1.53 ± 0.1 ^d
Citric acid (mmol kg ⁻¹)	0	2.84 ± 0.45 ^{a-c}	2.66 ± 0.3 ^{a-c}	2.50 ± 0.7 ^{a-c}	2.36 ± 0.6 ^{a-c}	2.19 ± 0.2 ^c
	2	2.86 ± 0.4 ^{ab}	2.73 ± 0.3 ^{a-c}	2.58 ± 0.2 ^{a-c}	2.42 ± 0.2 ^{a-c}	2.24 ± 0.4 ^{bc}
	6	2.90 ± 0.5 ^a	2.78 ± 0.3 ^{a-c}	2.71 ± 0.2 ^{a-c}	2.58 ± 0.3 ^{a-c}	2.35 ± 0.1 ^{a-c}
	10	2.94 ± 0.5 ^a	2.83 ± 0.4 ^{a-c}	2.75 ± 0.4 ^{a-c}	2.64 ± 0.3 ^{a-c}	2.41 ± 0.3 ^{a-c}
Titania nanoparticles (mg kg ⁻¹)	0	2.84 ± 0.5 ^{ab}	2.66 ± 0.3 ^{ab}	2.50 ± 0.7 ^{ab}	2.36 ± 0.6 ^{ab}	2.19 ± 0.2 ^{ab}
	20	2.91 ± 0.6 ^{ab}	2.83 ± 0.6 ^{ab}	2.76 ± 0.4 ^{ab}	2.62 ± 0.1 ^{ab}	2.49 ± 0.1 ^{ab}
	60	2.94 ± 0.7 ^{ab}	2.90 ± 0.7 ^{ab}	2.84 ± 0.5 ^{ab}	2.71 ± 0.3 ^{ab}	2.62 ± 0.1 ^{ab}
	100	2.96 ± 0.7 ^a	2.92 ± 0.6 ^{ab}	2.87 ± 0.9 ^{ab}	2.80 ± 0.5 ^{ab}	2.68 ± 0.2 ^b
Compost (%)	0	2.84 ± 0.5 ^{ab}	2.66 ± 0.3 ^{ab}	2.50 ± 0.7 ^{ab}	2.36 ± 0.6 ^{ab}	2.19 ± 0.2 ^{ab}
	2	2.92 ± 0.5 ^{ab}	2.85 ± 0.6 ^{ab}	2.80 ± 0.3 ^{ab}	2.64 ± 0.4 ^{ab}	2.51 ± 0.1 ^{ab}
	6	2.96 ± 0.5 ^a	2.92 ± 0.5 ^{ab}	2.86 ± 0.6 ^{ab}	2.75 ± 0.3 ^{ab}	2.63 ± 0.1 ^{ab}
	10	2.98 ± 0.5 ^a	2.94 ± 0.5 ^a	2.90 ± 0.5 ^{ab}	2.83 ± 0.5 ^{ab}	2.70 ± 0.1 ^b
Ammonium nitrate (mmol kg ⁻¹)	0	2.84 ± 0.5 ^a	2.66 ± 0.3 ^a	2.50 ± 0.7 ^a	2.36 ± 0.6 ^a	2.19 ± 0.2 ^a
	2	2.88 ± 0.5 ^a	2.79 ± 0.4 ^a	2.64 ± 0.9 ^a	2.58 ± 0.2 ^a	2.37 ± 0.2 ^a
	6	2.92 ± 0.6 ^a	2.83 ± 0.6 ^a	2.74 ± 0.4 ^a	2.64 ± 0.1 ^a	2.42 ± 0.2 ^a
	10	2.95 ± 0.4 ^a	2.89 ± 0.6 ^a	2.70 ± 0.6 ^a	2.73 ± 0.4 ^a	2.50 ± 0.4 ^a

Data is the mean (±SD) of three replicates; superscript letter shows the significant difference ($P < 0.05$, Tukey's Honestly Significant Difference Test) among all groups

4 Discussion

The current study showed that amendments play an important role in increasing the bioavailability of Cd in soil except compost which reduced the bioavailable portion in soil. Most of the chelate-assisted phytoextraction studies focused on the use of EDTA and considered it as the best chelating agent in enhancing the mobility of Cd (Xie et al. 2012). The present study also showed that EDTA was more efficient in enhancing the bioavailability and accumulation of Cd in shoots of *P. hortorum*. The Cd solubility in EDTA-amended soil after 7 days of incubation time was significantly increased by 1.2-fold as compared to control. This increase in Cd solubility may involve different mechanisms, such as the adsorption of Cd on free EDTA and then the detachment of Cd ions from minerals, resulting in increasing Cd solubility in soils (Shahid et al. 2014). The addition of citric acid, ammonium nitrate, and TiO₂ NPs increased the solubility of Cd in soil by 0.8-, 0.4-, and 0.2-fold, respectively, but this increase was lower than that with EDTA. In contrast to this, the addition of compost significantly decreased the solubility of Cd by 50%. The stabilizing role of Cd in soil has been reported. High organic

matter content increased the retention of Cd and decreased the solubility (Khan et al. 2017).

For the successful phytoextraction of Cd, bioavailable portion must be taken by the plants. *Pelargonium* species have ability to accumulate heavy metals in shoot and are considered as hyperaccumulator of Pb and Cd (Arshad et al. 2008; Manshadi et al. 2013; Arshad et al. 2016; Manzoor et al. 2018). In the present study, *P. hortorum* accumulated 262.8 and 176.6 mg Cd kg⁻¹ in shoot and root, respectively, at 150 mg Cd kg⁻¹ without application of amendments. Mahdih et al. (2013) reported that scented geranium, *Pelargonium roseum*, accumulated 1957 mg Cd kg⁻¹ of shoot dry weight in 14 days of exposure. *Pelargonium* species have inherent ability to uptake heavy metals by acidifying the rhizosphere soil which is the main factor in the metal availability (Arshad et al. 2016). In this study, EDTA application significantly increased the accumulation of Cd in shoot and root by 3.9- and 2.3-fold, respectively. Our findings were coherent with another study which reported that the application of 4 g EDTA kg⁻¹ increased Cd accumulation in shoot of marigold (*Tagetes* sp.) by 3.4-fold as compared to the control (Ali et al. 2016). The application of citric acid, ammonium nitrate, and

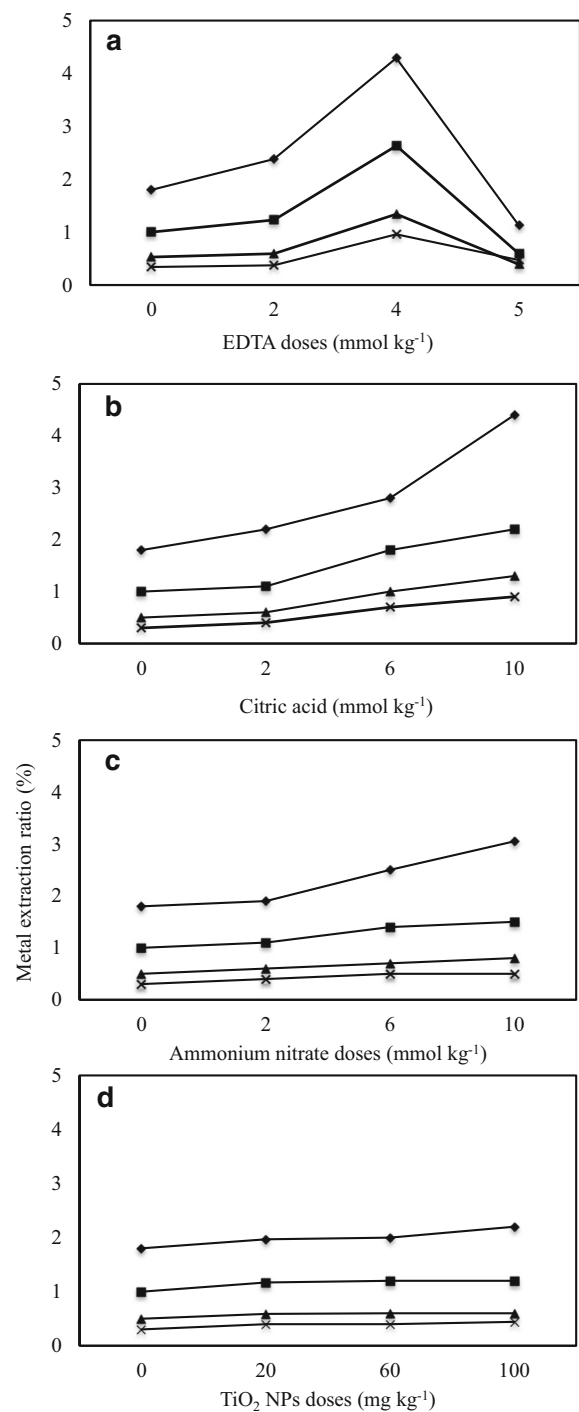
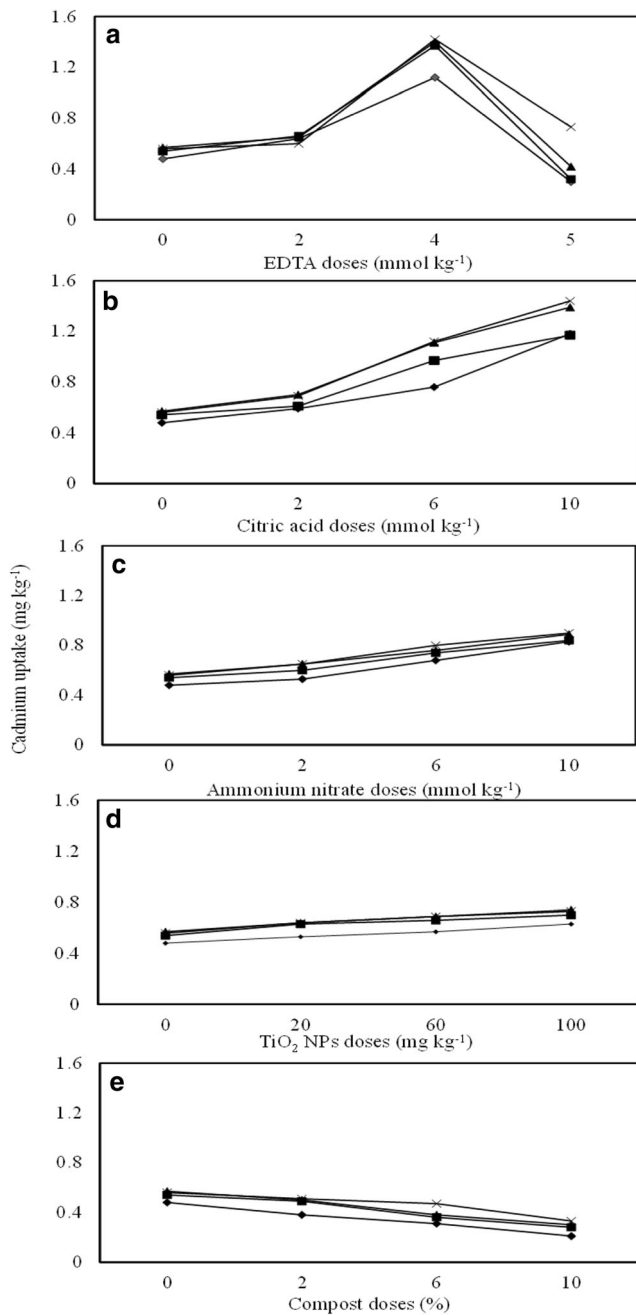


Fig. 3 Cd uptakes per plant by *P. hortorum*. **a**) EDTA. **b**) Citric acid. **c**) Ammonium nitrate. **d**) TiO₂ NPs. **e**) Compost (◆ = 25 mg Cd kg⁻¹; ■ = 50 mg Cd kg⁻¹; ▲ = 100 mg Cd kg⁻¹; × = 150 mg Cd kg⁻¹)

TiO₂ NPs increased the accumulation of Cd in shoot and root by 2.4-, 1.0-, and 0.2-fold and 1.5-, 1.0-, and 0.9-fold, respectively. In contrast, the application of compost showed decrease in Cd accumulation by shoots and root by 0.92- and 0.3-fold, respectively, and this decrease was due to possible stabilization of Cd in soil (Khan et al. 2017). The accumulation of Cd in shoot and root was significantly decreased by 31, 60, 70, 87, and 26, 40, 43, and 70%, respectively, upon the application of citric acid, ammonium nitrate, TiO₂ NPs, and compost

Fig. 4 Metal extraction ratio of *P. horticola*. **a)** EDTA. **b)** Citric acid. **c)** Ammonium nitrate. **d)** TiO₂ NPs. **e)** Compost (◆ = 25 mg Cd kg⁻¹; ■ = 50 mg Cd kg⁻¹; ▲ = 100 mg Cd kg⁻¹; × = 150 mg Cd kg⁻¹)

as compared to EDTA. By comparing all amendments in term of Cd accumulation, the EDTA was more efficient.

Another factor involved in phytoextraction is the ability of plant to survive and produce higher biomass. *P. horticola* has the ability to survive in heavy metal-contaminated soil but the biomass decreased at higher Cd levels (Gul et al. 2018b). The maximum plant biomass was observed in the control group, and with the increasing levels of Cd from 0 to 150 mg Cd kg⁻¹, the biomass of *P. horticola* was decreased by 22.8%. It is well reported that Cd causes phytotoxicity by decreasing the photosynthetic activity and ultimately reduces the plant biomass (Gul et al. 2018b). The addition of amendments showed various responses. Upon addition of EDTA in control groups, the biomass was decreased by 62%. Similar results were also reported by another study where the dry biomass of *Echinochloa crus galli* L. was decreased by 49.7% in the control group upon 5 mmol kg⁻¹ EDTA application. This decrease in the dry plant

biomass was due to the presence of high content of mobilized heavy metals and toxicity of free EDTA (Ebrahimi 2015). The dry plant biomass of *P. horticola* in citric acid-, ammonium nitrate-, TiO₂ NP-, and compost-amended soils was 2.94, 2.95, 2.96, and 2.98 g per plant, respectively. Among all amendments, 10% addition of compost in control group showed 14% increase in the dry plant biomass. Ahmad et al. (2015) showed 33.2% increase in the dry plant biomass of wheat upon addition of compost. The increase in dry plant biomass in compost-amended soil might be due to improved nutrient level, soil structure, and water holding capacity and reduction in the bioavailable fraction of Cd in soil (Oldari et al. 2011).

Plants having TF > 1 is considered as a hyperaccumulator. In Cd-treated groups without amendment application, TF > 1 (Table 3) indicated that *P. horticola* has the ability to transfer Cd from root to shoot. Similarly, Gul et al. (2018b) reported that *P. horticola* showed TF > 1 in Cd-spiked soils. The application of soil amendments further increased the TF. The maximum TF was observed upon EDTA application followed by citric acid, TiO₂ NPs, ammonium nitrate, and compost.

The maximum Cd uptake per plant (1.44 mg) was observed in citric acid-amended soil, followed by EDTA, ammonium

Table 3 Translocation factor of *P. horticola*

Translocation factor					
Cd treatments (mg kg ⁻¹)	▶	25	50	100	150
Amendment levels	▼				
EDTA (mmol kg ⁻¹)	0	1.81 ± 0.26 ^{bc}	1.79 ± 0.17 ^{bc}	1.67 ± 0.10 ^{bc}	1.50 ± 0.12 ^{bc}
	2	1.74 ± 0.07 ^{bc}	1.74 ± 0.22 ^{bc}	1.35 ± 0.04 ^c	1.43 ± 0.03 ^{bc}
	4	2.76 ± 0.16 ^a	3.05 ± 0.27 ^a	2.86 ± 0.10 ^a	2.70 ± 0.22 ^a
	5	1.96 ± 0.11 ^b	1.69 ± 0.33 ^{bc}	1.39 ± 0.12 ^c	2.55 ± 0.20 ^a
	10	1.81 ± 0.26 ^a	1.79 ± 0.17 ^a	1.67 ± 0.10 ^a	1.50 ± 0.12 ^a
Citric acid (mmol kg ⁻¹)	0	1.81 ± 0.26 ^a	1.79 ± 0.17 ^a	1.67 ± 0.10 ^a	1.50 ± 0.12 ^a
	2	1.92 ± 0.28 ^a	1.89 ± 0.80 ^a	1.62 ± 0.36 ^a	1.62 ± 0.16 ^a
	6	1.68 ± 0.52 ^a	2.04 ± 0.15 ^a	1.95 ± 0.20 ^a	2.10 ± 0.23 ^a
	10	2.27 ± 0.48 ^a	2.19 ± 0.20 ^a	2.29 ± 0.55 ^a	2.49 ± 0.20 ^a
	100	1.66 ± 0.45 ^a	1.73 ± 0.06 ^a	1.72 ± 0.15 ^a	1.81 ± 0.30 ^a
Titanium nanoparticles (mg kg ⁻¹)	0	1.81 ± 0.26 ^a	1.79 ± 0.17 ^a	1.67 ± 0.10 ^a	1.50 ± 0.12 ^a
	20	1.75 ± 0.53 ^a	1.93 ± 0.70 ^a	1.70 ± 0.60 ^a	1.51 ± 0.16 ^a
	60	1.77 ± 0.80 ^a	1.64 ± 0.16 ^a	1.59 ± 0.11 ^a	1.91 ± 0.55 ^a
	100	1.66 ± 0.45 ^a	1.73 ± 0.06 ^a	1.72 ± 0.15 ^a	1.81 ± 0.30 ^a
	100	1.66 ± 0.45 ^a	1.73 ± 0.06 ^a	1.72 ± 0.15 ^a	1.81 ± 0.30 ^a
Compost (%)	0	1.81 ± 0.26 ^a	1.79 ± 0.17 ^a	1.67 ± 0.10 ^a	1.50 ± 0.12 ^a
	2	1.81 ± 0.78 ^a	1.98 ± 0.52 ^a	1.79 ± 0.51 ^a	1.73 ± 0.42 ^a
	6	1.67 ± 0.07 ^a	1.58 ± 0.64 ^a	1.52 ± 0.35 ^a	1.70 ± 0.38 ^a
	10	1.46 ± 0.48 ^a	1.33 ± 0.34 ^a	1.23 ± 0.08 ^a	1.19 ± 0.21 ^a
	10	1.46 ± 0.48 ^a	1.33 ± 0.34 ^a	1.23 ± 0.08 ^a	1.19 ± 0.21 ^a
Ammonium nitrate (mmol kg ⁻¹)	0	1.81 ± 0.26 ^a	1.79 ± 0.17 ^a	1.67 ± 0.10 ^a	1.50 ± 0.12 ^a
	2	1.80 ± 0.24 ^a	1.96 ± 0.79 ^a	1.67 ± 0.56 ^a	1.45 ± 0.24 ^a
	6	1.46 ± 0.35 ^a	1.49 ± 0.20 ^a	1.61 ± 0.40 ^a	1.49 ± 0.43 ^a
	10	1.33 ± 0.21 ^a	1.40 ± 0.18 ^a	1.46 ± 0.46 ^a	1.39 ± 0.25 ^a
	10	1.33 ± 0.21 ^a	1.40 ± 0.18 ^a	1.46 ± 0.46 ^a	1.39 ± 0.25 ^a

Data is the mean (±SD) of three replicates; letter shows the significant difference ($P < 0.05$, Tukey's Honestly Significant Difference Test) among all groups

nitrate, and TiO₂ NPs and minimum (0.33 g) was observed in compost-amended soil. In the presence of citric acid, the accumulation of Cd in *P. hortorum* was 30% lower but the plant biomass was 50% higher than EDTA-amended soil. Therefore, Cd uptake by plant was higher in citric acid-amended soil as compared to EDTA-amended soil. The compost showed maximum plant biomass and growth, but the accumulation was reduced with increasing compost levels as it stabilized the Cd in soil instead of mobilization; therefore, the minimum Cd uptake was observed.

In assisted phytoextraction of Cd, the metal extraction ratios play an important role in identifying the efficiency of amendments in cleaning of soil. The citric acid showed maximum MER and minimum was observed for compost as compared to other amendments. By comparing all amendments, citric acid, ammonium nitrate, and TiO₂ NPs can be used as substitute to EDTA, as they showed minimum toxicity to plant in term of biomass and other physiological parameters.

5 Conclusions

EDTA, citric acid, ammonium nitrate, and TiO₂ NPs increased the mobility of Cd. Efficiency of amendments to increase mobility of Cd followed the sequence: EDTA > citric acid > ammonium nitrate > TiO₂ NPs > compost. The maximum accumulation of Cd in shoot and root was observed in EDTA-amended soil. But MER and Cd uptake were higher in citric acid-amended soil in comparison to other amendments. Overall, citric acid can be used as a substitute to EDTA in assisted phytoextraction of Cd.

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Compliance with ethical standards

Conflict of interest The authors have no conflict of interest to declare related to reported work here.

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