#### **RESEARCH ARTICLE**



# **Diferences in bacterial community composition, structure and function between sediments in waterways and non‑navigable channels in a plain river network area**

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## **Abstract**

Bacterial communities greatly help maintain the balance of river ecosystems and are highly sensitive to changes in environmental conditions. Plain river network areas (PRNs) are characterized by dense river networks, low-lying terrain, and slow water flow, where the bottom sediment is frequently disturbed by ship navigation due to the limited water depth and width of waterways, providing a unique ecological niche for bacterial growth. Hence, understanding how bacterial communities in PRNs respond to changes in hydrodynamic conditions, physicochemical parameters, and pollutants under ship navigation is essential to maintaining the stability of inland waterway ecosystems. The Taihu Lake Basin, a typical PRN, was selected to explore the diferences in bacterial community composition, structure and function between sediments in waterways (WS) and non-navigable channels (NS). The results indicate that the sediment from NS possessed more diverse and complex bacterial communities than WS. NMDS and ANOSIM analyses further verifed the signifcant diferences in bacterial community structure between WS and NS. Combined with LEfSe, we observed the highly diferential taxonomy between WS and NS from phylum to order. Moreover, a comparison of beta diversity dissimilarity indices revealed that although species replacement dominated both the WS and NS beta-diversity patterns, species loss caused the diferences in the overall beta diversity between them. Variance partitioning analysis revealed that physicochemical parameters (clay content, pH, ORP, and others) and ship traffic volume (STV) were the main driving factors for bacterial community distribution between WS and NS, while pollutants (heavy metals, perfuoroalkyl acids, and others) had a relatively minor infuence. PICRUSt2 analysis revealed that the changes in pH, ORP, and STV under ship navigation might inhibit the bacterial ability to metabolize carbohydrates. The results reveal the comprehensive efects of ship navigation disturbance on sediment bacterial communities in the PRN and contribute to further understanding of inland waterway ecosystems.

**Keywords** Bacterial community · Hydrodynamic action · Pollutants · Beta-diversity pattern · Function prediction · Metabolism

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# **Introduction**

Bacteria play a vital role in biogeochemical processes and nutrient cycling (Chen et al. [2022;](#page-10-0) Wang et al. [2016;](#page-12-0) Winter et al. [2007\)](#page-12-1) via assimilation and mineralization as a fundamental dynamics of river ecosystems (Paudel Adhikari et al. [2019;](#page-11-0) Zhang et al. [2020a](#page-13-0)). As an indispensable carrier and media in river systems, sediment provides diverse habitats for bacterial growth (Behera et al. [2020b](#page-10-1)). Meanwhile, surface sediment represents one of the most active regions of bacterial activity that can be used to refect the changes in functional characteristics of river water environments under the infuence of external conditions (Tšertova et al. [2013\)](#page-12-2). However, the composition and activity of bacterial communities are highly sensitive to changes in environmental conditions (Wang et al. [2021b](#page-12-3)), and as a result, the attributing factor to these changes can be confounded by the diversity and diferences in infuencing factors (Shade et al. [2012](#page-12-4); Steven et al. [2008](#page-12-5)). Human activity is reported to be one of the crucial factors attributed to changes in river sediment characteristics and composition (Behera et al. [2021b](#page-10-2); Su et al. [2018\)](#page-12-6), which would further affect the assembly of bacterial community structure in river ecosystems (Paudel Adhikari et al. [2019;](#page-11-0) Zhang et al. [2020a\)](#page-13-0). Effects of common human activities such as water conservancy projects and urban river construction on sediment bacterial community diversity and structure have been investigated (Gao et al. [2021](#page-11-1); Li et al. [2019b;](#page-11-2) Zhang et al. [2020b\)](#page-13-1), while the role of inland waterway transport has received little attention.

Inland waterway transport plays a vital role in cargo transportation because of its large capacity, low cost, and low energy consumption. Plain river network areas (PRNs) are characterized by dense river networks, low-lying terrain, and slow water flow, making the inland water transportation industry highly developed (Li and Hua 2021; Ma et al. [2021](#page-11-3)). However, due to the limited water depth and width of most waterways in PRNs, benthic sediment is frequently disturbed by ship navigation, providing a unique ecological niche for bacterial growth (Behera et al. [2021a\)](#page-10-3). Contrary to rivers without ship navigation, the propeller jets and ship-induced waves in the waterway provide complicated hydrodynamic conditions with axial, radial, and tangential velocities (Donal et al. [2013](#page-11-4); Lam et al. [2011,](#page-11-5) [2013](#page-11-6)), which directly afect the physical structure and composition of bacteria (Cheng et al. [2019b](#page-10-4)). Meanwhile, the turbulent fow caused by ship navigation disrupts the equilibrium state of the sediment–water interface, changing the physicochemical parameters of sediment and triggering pollutants released from bottom sediment to overlying water (Cheng et al.  $2019a$ ; Xu et al.  $2018$ ), and ultimately affecting the environmental conditions for the survival of bacterial communities. Previous studies have shown that the changes in sediment physicochemical parameters can signifcantly afect the diversity and composition of microbial communities. For example, Rath et al. [\(2019\)](#page-12-8) observed large bacterial community diferences along with the shifts in pH and salinity. Yao et al. [\(2022](#page-12-9)) studied the sediment of a eutrophic river and found that phosphorus and pH were the key factors dominating the bacterial community structure. In addition, studies on the efects of pollutants (e.g., toxic metals and organic pollutants) on microbial communities have also been widely reported. Due to the increasingly frequent human activities, multitudinous pollutants have been discharged into rivers and accumulated in sediment through complex effects (Behera et al. [2020a\)](#page-10-5), which inevitably have affected the microbial community in sediment (Rout et al. [2022](#page-12-10)). Cr (chromium) and Mn (manganese) signifcantly negatively

afect microbial alpha diversity, and their contributions have been quantifed (Li et al. [2020a](#page-11-8)). Furthermore, organic pollutants, such as perfuoroalkyl acids (PFAAs) and polycyclic aromatic hydrocarbons (PAHs), have exhibited non-negligible impacts on microbial communities due to their unique chemical properties by changing gene expression and microbial activity (Liu et al. [2021b](#page-11-9); Qiao et al. [2018;](#page-12-11) Wu et al. [2021b](#page-12-12)). However, whether ship navigation in PRNs afects bacterial community structure and the relative contributions of hydrodynamic conditions, physicochemical parameters and pollutants to the process remain unknown.

Identifying bacterial community patterns that may reveal diferent processes is crucial for understanding bacterial community structures and their causes (Baselga [2010](#page-10-6)). Beta diversity has been defned as the extent of change in community composition among sites (Whittaker [1960\)](#page-12-13), which is used to characterize bacterial community patterns (Simpson [1943\)](#page-12-14). Indeed, overall beta diversity stems from one or a combination of biological phenomena: nestedness and spatial turnover (Baselga [2010;](#page-10-6) Wang et al. [2021a](#page-12-15)). Nestedness refects a non-random process of species gain or loss, which occurs when the biota of sites with smaller numbers of species are subsets of the biota at richer sites (Baselga [2013](#page-10-7)). In contrast, spatial turnover implies the occurrence of the replacement of some species by others across geographic zones as a result of environmental fltering, competition, and historical constraints (Legendre [2014;](#page-11-10) Leprieur et al. [2011](#page-11-11)). Although the two biological phenomena can in combination contribute to overall beta diversity, their relative importance may vary across habitats and biological groups (Si et al. [2015;](#page-12-16) Wang et al. [2021a](#page-12-15)). In particular, inland waterways where bacterial habitats may become fragmented due to ship navigation disturbance leas to higher ecosystem vulnerability (Si et al. [2015](#page-12-16)). However, the relative contribution of species turnover and nestedness to bacterial community patterns under ship navigation disturbance remains unclear. Therefore, partitioning the overall beta diversity into separate components would further reveal the ecological processes underlying the biodiversity in inland waterways.

The present study focused on a PRN, and five pairs of sampling sites were set in adjacent waterways (WS) and nonnavigable channels (NS) with little diferences in depth and width. The main objectives were to explore the following questions: (1) What are the diferences in sediment bacterial community diversity, composition, and structure between WS and NS? (2) What are the biological phenomena implied by the bacterial community patterns in WS and NS? (3) What are the relative contributions of hydrodynamic conditions, physicochemical parameters, and pollutants to the bacterial community distribution? (4) Would ship navigation disturbance afect the function of bacterial communities in sediment? The results provide theoretical support for the management of inland waterways and advocate further attention to the sustainable development of inland waterway ecosystems.

#### **Materials and methods**

#### **Sampling design**

The Taihu Lake Basin is in the core area of the Yangtze River Delta in eastern China, which is densely covered with river networks corresponding to a river density of 3.2 km/ km $^2$  (Wu et al. [2018;](#page-12-17) Yu et al. [2022a](#page-12-18), [b](#page-12-19)). Due to the hydrological characteristics of low slope, shallow depth, and complicated connectivity (Li and Hua [2021](#page-11-12)), most of the waterways in the basin are Grade III and below (Jiangsu Province Waterway Atlas). As a typical PRN, the basin was selected to explore the efects of ship navigation disturbance on the sediment bacterial community. Furthermore, ship traffic would inevitably discharge pollutants such as toxic metals, petroleum hydrocarbons, PFAAs, and antifouling coatings to the waterway due to collision, explosion or breaking (Ali et al. [2022](#page-10-8); Prabowo and Bae [2019\)](#page-12-20). Therefore, heavy metals (HMs), PAHs, PFAAs, and organochlorine pesticides (OCPs) were selected as the target pollutants to evaluate their potential role in structuring bacterial communities.

Field sampling was conducted in August 2020. As shown in Fig. S1 and Table S1, the ten sampling sites were divided into five pairs. The water depth and width of WS and NS in each pair were similar (Table S2), and the sampling sites were in close proximity. Specifically, five sampling sites (WS1–WS5) were from Grand Canal, Wangyu River, Wutai Canal, and the Taipu River. The other fve sampling sites (NS1– NS5) were in NS, free from the disturbance of ship navigation.

## **Analysis of sediment physicochemical parameters and pollutants**

Three surface sediment samples (approximately 5-cm deep) were collected at each sampling site using a stainless-steel grab sampler and then thoroughly mixed and stored in 50 mL polypropylene centrifuge tubes. All the sediment samples were immediately transported to the laboratory in an ice-cooled box. Samples used for DNA extraction and other analyses were stored at−80 °C and−4 °C in the dark, respectively, until analysis. Physicochemical parameters of sediment samples included ORP, pH, salinity, total dissolved solids (TDS), conductivity, *foc*, D<sub>50</sub>, clay content, silt content, sand content, total nitrogen (TN), and total phosphorus (TP). Detailed measurement methods and results of the parameters are provided in Text S1 and Table S1, respectively.

The sediment samples were freeze-dried for 48 h and ground into a fne powder. Then, 1 g dw and 0.5 g dw of the sediment powder were used for HMs and PFAAs extraction (triplicate), respectively. HPLC grade solvents and deionized water were used for sample pretreatment and extraction. Detailed extraction methods of HMs and PFAAs can be found in Text S2. Specifcally, eight HMs (Cr, Cd, Ni, Cu, Zn, As, Pb, and Hg) and 13 PFAAs (C4–C14, PFBS, PFHxS, PFOS, PFBA, PFPeA, PFHxA, PFHpA, PFOA, PFNA, PFDA, PFUndA, PFDoA, and PFTeDA) were analyzed in the present study. The abbreviations, full names, molecular structure, and commercial sources of all PFAA and internal standards are summarized in Table S3. Detailed instrumental analyses are provided in Text S3 and Table S4.

The data on PAHs and OCPs were obtained from Zhao et al. ([2017\)](#page-13-2), which investigated the residues of PAHs and OCPs from 81 river sediment samples from the Taihu Lake Basin. ArcGIS (ESRI, Cal., USA) and Get-Data Graph Digitizer software were used to extract data. ArcGIS services map matching (geographic registration module) and Get-Data Graph Digitizer extracts data from fgures were based on a numerical algorithm (Yu et al. [2022](#page-12-18)). The extracted data were compared with the existing data in the paper, and the relative standard deviation ranged from 0.14% to 5.76%, indicating that the extracted data were reliable. Finally, the concentrations of PAHs and OCPs in sediment near the desired sampling sites were obtained after careful comparison with the Jiangsu Province Waterway Atlas.

#### **Quality assurance and quality control**

To avoid contamination, neither fuorinated substances nor glass was contacted during the feld sampling and pretreated processes. The feld blanks and transport blanks were treated, and a procedure blank and solvent blank (methanol) was added to every batch of 12 samples. No target pollutant was detected above the quantification limits in the blank samples. Internal standards were used for the calibration and standardization of target PFAAs. Native compounds were quantifed by a standard series of  $1-200 \mu g/L$ , and the regression coefficients of each PFAA standard curve were greater than 0.99. The limits of detection (LODs) and the limits of quantifcation (LOQs) were set as signal-to-noise ratios (S/N) of 3:1 and 10:1, respectively (Table S5). Based on the LODs and LOQs, the method detection limits (MDLs) and the method quantifcation (MQLs) were obtained by a batch of standard addition tests. The matrix-spiked recoveries of PFAAs (10 ng/g dw) ranged from 82.55% to 127.04% (Table S5).

#### **DNA extraction and sequencing analysis**

DNA of each sediment sample was extracted by the ALFA-SEQ Advanced Soil DNA Kit (Guangzhou mCHIP Biotechnology Co., Ltd, China). The regions of the bacterial 16S rRNA gene (V3–V4) were amplifed using the 338F (ACT CCTACGGGAGGCAGCA) and 806R (GGACTACHVGGG TWTCTAAT) primers. PCR amplifcation and Illumina Hiseq sequencing were conducted at Guangdong Magigene Biotechnology Co. Ltd. (Guangzhou, China). The detailed sequencing processes are described in Text S4.

#### **Statistical analyses**

To compare the diferences in physicochemical parameters, hydrodynamic conditions and pollutant concentrations between WS and NS sediments, the bacterial community diversity, structure, and function in response to the diferences were determined. First, the alpha diversity of each sample was calculated in QIIME 2 using the observed species diversity indices based on the OTU analysis. In addition, Spearman correlations were calculated between environmental factors and the alpha diversity of bacteria. Based on the Bray–Curtis distance, non-metric multidimensional scaling (NMDS) analysis was performed in R software ("vegan" package) to determine the dissimilarity of bacterial community structure between WS and NS (Hu et al. [2021](#page-11-13)). Next, an analysis of similarity (ANOSIM) was conducted to determine whether the diferences were signifcant. The linear discriminant analysis of efect sizes (LEfSe) was used to identify a highly diferential taxonomy between WS and NS (Zhang et al. [2019a](#page-12-19), [2021b\)](#page-13-3). In this analysis, the nonparametric Kruskal–Wallis rank-sum test was frst performed to fnd the species with signifcant diferences in abundance between the WS and NS. Next, the Wilcoxon rank-sum was used to test the diference consistency among the subgroups of the species obtained in the previous step, followed by linear regression analysis (LDA) to estimate the efect size of each species at log (10) values and visualize taxa in the cladogram.

Furthermore, to reveal the biological phenomena implied by the bacterial community patterns in WS and NS, the Sørensen dissimilarity index ( $β<sub>SOR</sub>$ ), Simpson dissimilarity index  $(\beta_{SIM})$ , and the nestedness-resultant dissimilarity index  $(\beta_{\text{NES}})$  of WS and NS were calculated in R-studio ("betapart" package) (Baselga [2010](#page-10-6), [2013\)](#page-10-7), respectively. Redundancy analysis (RDA) was conducted in Canoco 5 to elucidate the correlations between bacterial community structure and environmental factors. Variance infation factor analysis (VIF) was performed to select vital factors before RDA analysis. Variance partitioning canonical correspondence analysis (VPA) was used to quantify the relative contributions of physicochemical parameters, ship traffic volume (STV), and pollutants to the bacterial communities. A KEGG (Kyoto Encyclopedia of Genes and Genomes) pathway analysis was performed using PICRUSt 2 (phylogenetic investigation of communities by reconstruction of unobserved states) to investigate whether the changes in hydrodynamic conditions might lead to functional differences (Wang et al. [2018;](#page-12-21) Xu et al. [2020](#page-12-22)). Finally, the Student's *t*-test was conducted in IBM SPSS 22 software to test the bacterial metabolic pathways with signifcant diferences characterized at various levels between the WS and NS (Yin and Wang [2021](#page-12-23)). The Spearman correlations were calculated between the environmental factors and the top 15 bacterial classes.

## **Results**

## **Physicochemical parameters and pollutants concentrations**

The primary river characteristic parameters of WS and NS are provided in Table S2. Five WS, including Grade III and V, correspond to single-line straight waterway widths of 30–55 m and 22–35 m (Inland Waterway Navigation Standards), respectively. The average width of the five NSs is 39.60 m. Additionally, a few diferences in water depth between WS  $(4.22 \pm 1.22 \text{ m})$  and NS  $(3.97 \pm 0.46 \text{ m})$ were observed (Taihu Basin Authority of Ministry of Water Resources, Water Information of Jiangsu Province) (Table S2). Furthermore, the physicochemical parameters of the sediment samples collected from each sampling site are listed in Table S1. pH ranged from 6.90 to 7.86, indicating sediment in the study was neutral to slightly alkaline (Chen et al. [2017\)](#page-10-9). Salinity ranged from 0.12 to 0.72 psu (0.44 $\pm$ 0.15 psu). In addition, the ORP values of the investigated sediment were between 50.20 and 396.70 mv, with signifcantly higher ORP values observed in WS  $(249.48 \pm 116.60 \text{ mv})$  ( $p < 0.05$ ), which showed stronger oxidizability than NS  $(103.90 \pm 51.34 \text{ mv})$ . The organic carbon content (*f*oc) of all sediment ranged from 3.92% to 7.04%  $(5.21\% \pm 0.91\%)$ , with a small range in variation. Sediment from all sampling sites was mainly composed (accounting for more than 70% on average) of silt  $(2-63 \mu m)$ , followed by sand (63–2000 μm), and clay ( $\lt 2$  μm), suggesting that the sediment was mainly composed of grains larger than 2 μm in size. Meanwhile, the sediment grain size distribution between WS and NS exhibited certain diferences. The  $D_{50}$ , silt content, and sand content of WS sediment were higher than that of NS, whereas the clay content was significantly lower than NS  $(p < 0.05)$ . The TN and

TP concentrations in WS [TN  $(1671.33 \pm 160.36 \text{ mg/kg})$ dw) and TP (667.23  $\pm$  116.16 mg/kg dw)] were relatively lower than NS [TN  $(1727.92 \pm 181.30 \text{ mg/kg} \text{ dw}$  and TP  $(729.38 \pm 149.77 \text{ mg/kg} \text{ dw})$ ].

As for the pollutants, the average concentration of HMs in the studied area followed the decreasing order of: Zn  $(158.43 \pm 46.11 \text{ mg/kg} \text{ dw})$  > Cr  $(104.71 \pm 20.03 \text{ mg/kg}$ dw) > Cu (44.70  $\pm$  18.24 mg/kg dw) > Ni (43.93  $\pm$  12.58 mg/ kg dw) > Pb  $(39.94 \pm 12.19 \text{ mg/kg} \text{ dw})$  > As  $(16.90 \pm 5.03 \text{ mg/kg} \text{ dw}) > Cd (0.51 \pm 0.16 \text{ mg/kg} \text{ dw}) > Hg$  $(0.15 \pm 0.07 \text{ mg/kg} \text{ dw})$  (Table S6). Moreover, the total concentrations of HMs (ΣHMs) in WS ranged from 249.44 to 517.16 mg/kg dw  $(384.67 \pm 88.26$  mg/kg dw), and 277.23 to 505.94 mg/kg dw  $(433.87 \pm 80.48 \text{ mg/kg} \text{ dw})$  in NS. Furthermore, seven PFAAs (PFBA, PFHpA, PFOA, PFDA, PFDoA, PFHxS, and PFOS) were 100% detected among the 13 PFAAs. PFBA, PFOS, PFHpA, and PFOA were identifed as the dominant PFAAs, with a proportion of 31.81%, 28.44%, 13.23%, and 12.48%, respectively (Table S7 and Fig. S2). The total concentrations of PFAAs (ΣPFAAs) in all samples ranged from 29.95 to 67.15 ng/g dw, with the median ΣPFAAs in NS being 32.88% higher than that of WS. In addition, the mean concentrations of PAHs and OCPs in WS were  $296.38 \pm 119.44$  ng/g dw and  $32.48 \pm 6.77$  ng/g dw, and those in NS were  $325.99 \pm 137.96$  ng/g dw and  $39.98 \pm 11.85$  ng/g dw, respectively.

#### **Bacterial alpha diversity**

After quality fltering, 383,028 high-quality sequences of the bacterial 16S rRNA gene were obtained and assigned to 39,984 OTUs. Diversity information of bacterial communities, including the number of OTUs, Chao1, Shannon, and Simpson indices, are provided in Table S8. The alpha diversity values of bacterial communities between WS and NS were different, with the mean Chao1 ( $4478 \pm 174$ ) and Shannon  $(10.00 \pm 0.20)$  indices of NS sediment being 1.27 and 1.34 times higher than WS [Chao1  $(3520 \pm 876)$  and Shannon indices  $(7.50 \pm 2.10)$ ], respectively. In addition, the mean Simpson index of WS sediment was 20.32 times that of NS.

To further investigate the relationships between bacterial richness, diversity, and environmental factors, Spearman correlation coefficients were calculated (Fig.  $1$ ). The results indicate that the STV, Pb, PFBA, clay content, PFHxS, and Cd signifcantly afected the bacterial alpha diversity in sediment  $(p < 0.05)$ . Specifically, STV, Pb, PFHxS, and Cd were signifcantly negatively correlated with the Chao1 and Shannon indice, and positively correlated with the Simpson index. The results revealed that ship navigation disturbance was unfavorable for bacterial alpha diversity and the richness and diversity of bacterial communities decreased with the increase of STV. Meanwhile, the exposure of the bacterial community to Pb, Cd, PFBA, and PFHxS could result in a reduction of alpha diversity. Conversely, clay content was signifcantly positively correlated with the Chao1 and Shannon indices and negatively correlated with the Simpson index, indicating that the increase in clay content was benefcial to bacterial richness and diversity.

#### **Bacterial community composition and beta diversity**

The composition and relative abundance across bacterial communities of diferent samples at the phylum and order levels are displayed in Fig. S3a–b. Proteobacteria (15.56–87.56%, mean 43.03%) were the dominant phylum in most all sediment samples. The other predominant bacterial phyla in all samples included Chlorofexi (1.08–25.46%, mean 12.38%), Bacteroidetes (1.41–20.77%, mean 11.85%), and Epsilonbacteraeota (0.09–49.71%, mean 7.86%), according to their overall abundance (Fig. S3a). At the order level, the dominant orders included Betaproteobacteriales (6.08–31.05%, mean 17.91%), Campylobacterales (0.09–49.71%, mean 7.86%), and Anaerolineales (0.30–10.44%, mean 6.60%) (Fig. S3b).

The NMDS analysis based on the Bray–Curtis distance was performed to characterize the beta diversity of bacterial communities and indicated that marked diferences in bacterial community structure between WS and NS were observed (Fig. [2](#page-5-0)). The ANOSIM confrmed the consistency of the results, which further verifed that the bacterial community structure in NS sediment was signifcantly diferent from that in WS ( $R = 0.276$ ,  $p = 0.006$ ). NMDS analysis suggested that the bacterial community structure in NS showed a tendency to cluster, whereas the bacterial community structure in WS was scattered, indicating that the bacterial community structure in NS sediment was more stable than WS (Fig. [2](#page-5-0) and Fig. S3).

LEfSe was then performed to assess the highly diferential taxonomy (from phylum to order levels) between the



<span id="page-4-0"></span>**Fig. 1** Heat map illustrates the correlations between environmental factors and bacterial alpha diversity indices (\*0.01≤*p*<0.05, \*\**p*<0.01)



<span id="page-5-0"></span>**Fig. 2** Non-metric multidimensional scaling (NMDS) ordination of the bacterial community (Bray−Curtis distance). Points close together represent samples with similar bacterial community composition, and the ovals represent the 95% confdence intervals of the grouped samples

samples from WS and NS (Zhang et al. [2019a\)](#page-12-19). The phylogenetic distribution of bacterial lineages is displayed in Fig. [3a,](#page-5-1) and the LDA values for each lineage are shown in Fig. [3b.](#page-5-1) LDA values greater than two indicated that species were signifcantly associated with sediment from the WS (green) and NS (red). The results indicate that 63 diferentially abundant taxa were detected between WS and NS, of which 62 were enriched in NS. At the class level, the bacterial taxa signifcantly enriched in the NS were identifed as Gammaproteobacteria, Bacteroidia, Deltaproteobacteria, Lentisphaeria, Hydrogenedentia, Fimbriimonadia, and others, among which Gammaproteobacteria, Bacteroidia, and Deltaproteobacteria belonged to the dominant class. Furthermore, Streptomycetales, belonging to Actinobacteria, exhibited the only signifcant presence in WS.

## **Ecological process underlying biodiversity**

The causality underlying the beta diversity is described by one or combinations of two biological phenomena of spatial



<span id="page-5-1"></span>**Fig. 3 a** The bacterial lineages with signifcant diferences between WS (green) and NS (red) and **b** LDA values for each lineage

turnover and nestedness, which are evaluated based on the presence and absence of species (Baselga [2010,](#page-10-6) [2013](#page-10-7)). To further determine the biological phenomena implied by the bacterial community patterns in WS and NS, the beta diversity dissimilarity indices were calculated, respectively, with results listed in Table S9. For WS, the estimated overall beta diversity  $β<sub>SOR</sub>$  value of the bacterial community among the sampling sites was 0.69, with a substantial contribution from spatial turnover ( $\beta_{SIM} = 0.61$ ) and a small contribution from nestedness ( $\beta_{\text{NES}}$ =0.08). Similarly, a considerable contribution from spatial turnover  $(\beta_{SIM} = 0.59)$  and a small contribution from nestedness  $(\beta_{\text{NES}}=0.01)$  was observed in NS, with the  $\beta_{\text{SOR}}$  of the bacterial community at 0.60.

Comparing the dissimilarity indices between WS and NS showed that the overall beta diversity of WS ( $\beta_{\text{SOR}}$ =0.69) was higher than for NS ( $\beta_{\text{SOR}}$ =0.60), also indicating the extent of variation in community composition among WS sampling sites was more drastic than NS. Furthermore, analysis of their spatial turnover and nestedness components found that spatial turnover was almost equally important to the WS ( $\beta_{SIM} = 0.61$ ) and NS ( $\beta_{SIM} = 0.59$ ), while nestedness contributed eight times more to the WS ( $\beta_{\text{NES}}$ =0.08) bacterial community structure than NS ( $\beta_{NFS}=0.01$ ). In comparison, beta-diversity patterns in WS resulted from both species replacements and species loss, while the betadiversity patterns in NS almost entirely attributed to species replacement only.

#### **Environmental factors afecting bacterial community structure**

The RDA was performed to explore the correlations between environmental factors and bacterial community structures (Fig. [4](#page-6-0)). Based on VIF analysis (Zhang et al. [2019b](#page-13-4)), STV, clay content, pH, ORP, HMs, PFAAs, OCPs, and TN were selected as the primary explanatory factors affecting the bacterial community in sediment, which could explain 78.24% of the total variation. As shown in Fig. [4](#page-6-0), STV  $(R^2 = 0.608)$ , pH ( $R^2$  = 0.596), and ORP ( $R^2$  = 0.486) had the greatest influences on the bacterial community structure, and other factors—namely, PFAAs, HMs, clay content, and TN—were also strong predictors for bacterial community distribution.

The VPA was next used to determine whether the STV, pollutants (HMs, PFAAs, and OCPs), or physicochemical parameters (clay content, pH, ORP, and TN) contributed more to the bacterial community distribution. The results suggest that the combination of STV, pollutants and physicochemical parameters contributed exceeded 75% to the bacterial community distribution (Fig. [5](#page-6-1)). Among them, physicochemical parameters and STV were the most important factors, which explained 38.2% and 32.7% of the variation in bacterial communities, respectively, much greater than the role of pollutants (12.2%).



<span id="page-6-0"></span>**Fig. 4** Redundancy analysis (RDA) shows the relationships between the most signifcant environmental factors and bacterial community structure in sediment



<span id="page-6-1"></span>**Fig. 5** Variance partitioning canonical correspondence analysis (VPA) illustrates the efects of STV, pollutants (HMs, PFAAs, and OCPs), and physicochemical parameters (clay content, pH, ORP, and TN) on the bacterial communities

## **Metabolism functional prediction of bacterial community**

Among the six categories of biological metabolic pathways in level 1, the relative abundance of genes related to metabolism was the highest (78.80–79.71%), followed by genetic information processing (12.60–12.79%), cellular processes (4.84–5.59%), and environmental information processing (2.09–2.37%), while the abundances of genes related to organismal systems (0.34–0.35%) and human diseases  $(0.21-0.28\%)$  were much lower (Fig. [6a\)](#page-7-0).

In comparison, the relative abundance of genes related to metabolism in WS was significantly lower than NS  $(p<0.05)$  among the six categories of biological metabolic pathways (Fig. S4), indicating that ship navigation would constrain the metabolic function of bacteria in WS sediment. Subsequently, the distribution of bacterial metabolic function in level 2 was further analyzed (Fig.  $6a$ ). The results suggested that the relative abundances of most sub-functions in WS were lower than in NS, such as the metabolism of cofactors and vitamins, carbohydrate metabolism, and amino acid metabolism that occupied relatively high abundances  $(>10\%)$ . In particular, the relative abundances of genes related to carbohydrate metabolism, amino acid metabolism, biosynthesis of other secondary metabolites, and transcription in WS were significantly lower than in NS ( $p < 0.05$ , Fig. [6a\)](#page-7-0). Then, carbohydrate metabolism (12.53%–12.73%) and amino acid metabolism (12.39–12.53%) with relatively higher abundance were selected to take a closer look, and the diferences of pathways belonging to these two functions in level 3 between WS and NS were tested. For carbohydrate





- Fructose and mannose metabolism
	- Galactose metabolism
- Ascorbate and aldarate metabolism
- 8. Starch and sucrose metabolism
	- Amino sugar and nucleotide sugar metabolism
		- Pyruvate metabolism
	- Glyoxylate and dicarboxylate metabolism
	- Propanoate metabolism

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- Butanoate metabolism
- C5-Branched dibasic acid metabolism
- Inositol phosphate metabolism

<span id="page-7-0"></span>**Fig. 6 a** Relative abundance of metabolic pathways on KEGG categories in level 2; **b** relative abundance of carbohydrate metabolism on KEGG categories in level 3 (\*0.01 ≤ *p* < 0.05, \*\**p* < 0.01)

metabolism, the relative abundance of genes related to the pentose phosphate pathway, pentose and glucuronate interconversions, and galactose metabolism in NS sediment were significantly higher than WS ( $p < 0.05$ , Fig.  $6b$ ), indicating that the ability of bacteria in WS sediment to degrade various sugars might be inhibited (Yin and Wang [2021\)](#page-12-23). Spearman correlation analysis showed that the pentose phosphate pathway and galactose metabolism were signifcantly stimulated by pH ( $rho = 0.652$  and 0.673,  $p < 0.05$ ), while the pentose and glucuronate interconversions were inhibited by ORP and STV (*rho*=– 0.624 and – 0.661, *p*<0.05). Furthermore, no signifcant diference was found between WS and NS concerning amino acid metabolism in level 3.

## **Discussion**

To investigate the dynamic changes of bacterial communities under ship navigation disturbance in the PRN, we compared the diferences in diversity, composition and function of bacterial communities between WS and NS sediments. WS sediment is frequently disturbed by ship navigation, whereas NS sediment is free from the disturbance of ship navigation.

The alpha diversity analysis showed there was an uneven distribution in bacterial communities between WS and NS. The Chao1 index refects the bacterial richness, while the Shannon and Simpson indices represent the bacterial diversity. The Chao1 and Shannon indices of WS were lower than those of NS, while the Simpson index was higher than that of NS (Table S8), indicating that the sediment from NS possessed more diverse and complex bacterial communities than WS. In addition, the bacterial community compositions in NS varied slightly among the diferent sampling sites, whereas the bacterial community structure in WS showed considerable spatial heterogeneity (Fig. [2\)](#page-5-0). A similar phenomenon was observed in the bacterial community structure of sediment between the infowing estuary and the central area of Dongting Lake (Zhang et al. [2019a\)](#page-12-19), and they attributed this diference to the combined efects of sediment transport and deposition under diferent hydrodynamic conditions. Then, LefSe analysis further showed that WS sediment had a unique bacterial community pattern compared with NS. Ship navigation disturbance appears to have an apparent environmental filtering effect on selecting adaptive species in WS sediment. Zhang et al. ([2021a\)](#page-13-5) also reported a similar fltering efect of frequent water level fuctuations on the bacteria in riparian zones.

A comparison of beta diversity dissimilarity indices suggested that spatial turnover and nestedness contributed differently to the bacterial community structures, with species replacement dominating the formation of beta-diversity patterns in WS and NS rather than species loss, illustrating the high proportion of endemic bacterial species present in the study area (Baselga [2010\)](#page-10-6). Wu et al. [\(2021c](#page-12-24)) also reported that spatial turnover explained more variation in planktonic bacterial community structure than nestedness. However, although species loss contributed less than species replacement to the WS and NS beta-diversity patterns, it caused a diference in the overall beta diversity between WS and NS, which should also deserve attention. It is generally accepted that both geographic and environmental factors would afect the patterns of bacterial communities (Baselga [2010](#page-10-6); Liu et al. [2018](#page-11-14)). For example, research on the beta-diversity patterns of European longhorn beetles reported an orderly increase in species loss with an increase in geographic distance from southern to northern Europe (Baselga [2010](#page-10-6)). Liu et al. ([2018](#page-11-14)) also reported the increase in the diferences between bacterial communities from the upper reach of the Yangtze River to the river mouth (4300 km). However, the geographic distance between sampling sites in NS and WS was not likely a factor since they were close to each other in each pair (Fig. S1 and Table S1), which was far less distant than the geographical distances that led to the dissimilarities of the river bacterial community composition. Therefore, the effect of geographic distance on the differences in bacterial community structure between WS and NS can almost be ignored. Meanwhile, environmental factors were most likely the dominant contributing factors to the diferences observed and appear to be important factors afecting bacterial communities (Wang et al. [2021a;](#page-12-15) Zhang et al. [2019b](#page-13-4)). As previously discussed, the hydrodynamic conditions, physicochemical characteristics, and pollutant concentrations in WS sediment difered from NS. Compared to NS, the higher contribution of species loss to the bacterial community structure observed in WS suggested that the local environmental conditions provided by WS sediment might not be conducive to bacterial growth.

The RDA and VPA analyses further revealed that the physicochemical parameters and STV contributed more to the variation in bacterial communities between WS and NS than pollutants (Figs. [4](#page-6-0) and [5\)](#page-6-1). In general, hydrodynamic disturbances are the primary physical force afecting bacterial living environments (Cheng and Hua [2016;](#page-10-10) Jalil et al. [2019](#page-11-15)). For example, Besemer et al. [\(2009](#page-10-11)) reported that the spatial variation of hydrodynamic forces largely explained the change of community composition along with bedforms. In the present study, STV represents the intensity of ship navigation disturbance to a certain extent, providing additional hydrodynamic forces for WS sediment. The riverbed of a waterway is proven to undergo signifcant changes when subjected to a strong swirling jet flow induced by a rotating ship propeller (Hong et al. [2016](#page-11-16); Nadia et al. [2019](#page-11-17)). Strong shear stress would break the stability of the riverbed and resuspend bottom sediment (Lam et al. [2010](#page-11-18)), thus forcing the bacterial communities to strip away from the colonized substrate and random difusion from sediment to overlying water (Lv et al. [2021](#page-11-19); Shen et al. [2015](#page-12-25)) and leading to the species loss observed in WS sediment. Sediment particle size distribution, which was reported as a driving factor causing the diferent structures of the microbial community (Zhang et al. [2020b\)](#page-13-1), may further verify the speculation. Generally, smaller particles favor bacterial attachment since they have larger specifc surface areas (Gao et al. [2021;](#page-11-1) Wu et al. [2021a](#page-12-26)). The signifcant lower clay content in WS than NS indicated that the erosion caused by ship navigation on WS sediment could carry away fne grained sediment, resulting in the coarse-graining of WS sediment, which was consistent with Grant et al. ([2018](#page-11-20)) and Wei et al. ([2016](#page-12-27)), who found that coarser-grained sediment is deposited in areas with high current velocity. As a result, the bacterial communities originally colonized on clay may be lost in the process, which also explained why the higher contribution of species loss to bacterial community structure was observed in WS than in NS. Meanwhile, the frequent disturbance could alter the degree of bacterial aggregation in sediment (Li et al. [2020b;](#page-11-21) Shen et al. [2015](#page-12-25)), subsequently increasing the possibility of bacterial community migration and leading to the distribution of bacterial communities in sediment exhibiting considerable spatial variability. Therefore, the bacterial community structure in WS sediment could be altered under the direct physical disturbance of ship navigation. A similar phenomenon was observed in the study of water diversion-induced changes in sediment bacterial communities (Lv et al. [2021](#page-11-19)), which reported that water diversion totally altered the natural seasonal changes in bacterial communities.

On the other hand, hydrodynamic disturbances can induce sediment–water interface material exchange (Cheng et al. [2019b](#page-10-4); Wang et al. [2015](#page-12-28)), causing changes in the physicochemical characteristics of sediment and indirectly afecting the bacterial community. ORP and pH have been considered two of the critical factors that signifcantly afect the structure and composition of microbial communities (Chang et al. [2014](#page-10-12); Meng et al. [2021;](#page-11-22) Zhang et al. [2021b\)](#page-13-3). In the present study, the mean value of ORP in WS sediment was significantly higher than NS ( $p < 0.05$ ), and the pH was correspondingly lower than NS (Table S1). ORP values can laterally refect the oxygen content in sediment (Li et al. [2019a](#page-11-23)), and the signifcant higher ORP values in WS suggested that ship navigation disturbance might directly accelerate the oxygen exchange rate at the sediment–water interface (Ma et al. [2021\)](#page-11-3), which oxygenates the sediment. This condition was conducive to the decomposition of organic matter, which in turn releases acidic substances (Fang et al. [2021](#page-11-24); Mastný et al. [2018](#page-11-25)) and reduces the pH of WS sediment. Lower pH may be stressful for bacterial growth (Kim et al. [2016\)](#page-11-26); the relatively higher ORP and lower pH conditions in WS sediment might also be responsible for the diferences in bacterial communities between WS and NS.

Additionally, HMs and organic pollutants are also important factors afecting the diversity and composition of microbial communities. For example, Yao et al. ([2022\)](#page-12-9) reported that HMs dominated the variation of microbial communities in eutrophic rivers and exhibited different effects on bacterial and fungal communities. Several studies indicated that PFAAs would lead to the evolution of bacterial communities with characteristics that resisted toxicity, which had the dual efects of stimulating and inhibiting the growth of the bacterial community (Lu et al. [2020;](#page-11-27) Qiao et al. [2018](#page-12-11)). Furthermore, the effects of PAHs and OCPs on microbial communities were also confrmed (Egbe et al. [2021;](#page-11-28) Liu et al. [2021b](#page-11-9); Yan et al. [2019\)](#page-12-29). However, although the concentrations of studied pollutants in WS sediment were relatively lower than those in NS, no signifcant diferences were observed in the mean concentrations of pollutants in sediment between WS and NS ( $p > 0.05$ ). The results may be a function of surface sediment being an important sink for pollutants (Hua et al. [2021\)](#page-11-29), and in turn refecting the results of the long-term deposition of pollutants in the sediment. Second, Taihu Lake Basin is a typical annular river network with an uncertain flow direction. The sampling sites from WS and NS in each pair were close and afected by approximate external pollution sources. Thus, ship navigation disturbance may trigger the release of these pollutants from sediment in a short time (Ma et al. [2021\)](#page-11-3), but the concentration of pollutants in sediment is generally stable. Therefore, although the pollutants had varying degrees of impact on the alpha diversity of bacterial communities (Fig. [1](#page-4-0)), they were not the main factor causing the diferences in bacterial community structure between WS and NS (Fig. [5](#page-6-1)).

Predictions of metabolic pathways are essential for understanding the metabolic activity of bacterial communities (Liu et al. [2021a](#page-11-30); Wu et al. [2021c](#page-12-24)). Although the functional predictions of bacterial communities in the present study are putative, this can still provide valuable predictive information for exploring the functional diferences of bacteria under ship navigation disturbance. The results of the present study were consistent with previous studies that found that metabolism and genetic information processing are the two dominant functions of bacteria in sediment (Abia et al. [2018;](#page-10-13) Rajeev et al. [2021](#page-12-30)). However, the bacterial communities in sediment exhibited considerable variations in functional composition under diferent hydrodynamic conditions. Notably, ship navigation disturbance might inhibit the carbohydrate metabolism function of bacterial communities in WS by reducing pH and increasing ORP and STV. Therefore, attention should be paid to the possible inhibitory efects of ship navigation disturbance on bacterial metabolism function, especially in plain river networks with dense WS, where the adverse efects of ship navigation disturbance on the bacterial metabolism activities could be greater.

# **Conclusions**

The present study contributes to a better understanding of the dynamic changes of sediment bacterial communities under complex hydrodynamic environments. We found that the sediment bacterial alpha diversity decreased gradually from NS to WS and that the bacterial community structure in WS was less stable than that in NS. Moreover, the underlying factor of the diferences in beta diversity between WS and NS was species loss caused by the changes in hydrodynamic conditions and physicochemical parameters in WS sediments. In addition, ship navigation disturbance may inhibit the carbohydrate metabolic function of bacterial communities in WS by reducing pH and increasing ORP and STV. Our study provides a reference for understanding the response of bacterial communities in sediment to ship navigation disturbance in the PRN, although further studies are needed to elucidate how this response changes in diferent seasons. While inland transport brings great economic benefts, it is important to pay attention to the ecosystem health of inland waterways.

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**Data availability** The sequenced data can be found in the NCBI Sequence Read Archive (accession number: PRJNA785136).

## **Declarations**

**Ethics approval and consent to participate** Not applicable.

**Consent for publication** Not applicable.

**Competing interests** The authors declare no competing interests.

## **References**

<span id="page-10-13"></span>Abia ALK, Alisoltani A, Keshri J, Ubomba-Jaswa E (2018) Metagenomic analysis of the bacterial communities and their functional profles in water and sediments of the Apies River, South Africa, as a function of land use. Sci Total Environ 616– 617:326–334.<https://doi.org/10.1016/j.scitotenv.2017.10.322>

- <span id="page-10-8"></span>Ali MM, Islam MS, Islam AR, Bhuyan MS, Ahmed AS, Rahman MZ, Rahman MM et al (2022) Toxic metal pollution and ecological risk assessment in water and sediment at ship breaking sites in the Bay of Bengal Coast. Bangladesh. Mar Pollut Bull. 175:113274. <https://doi.org/10.1016/j.marpolbul.2021.113274>
- <span id="page-10-6"></span>Baselga A (2010) Partitioning the turnover and nestedness components of beta diversity. Global Ecol Biogeogr 19(1):134–143. <https://doi.org/10.1111/j.1466-8238.2009.00490.x>
- <span id="page-10-7"></span>Baselga A (2013) Separating the two components of abundancebased dissimilarity: balanced changes in abundance vs. abundance gradients. Methods Ecol Evol. 4(6):552-7. [https://doi.](https://doi.org/10.1111/2041-210X.12029) [org/10.1111/2041-210X.12029](https://doi.org/10.1111/2041-210X.12029)
- <span id="page-10-5"></span>Behera BK, Chakraborty HJ, Patra B, Rout AK, Dehury B, Das BK, Sarkar DJ, Parida PK, Raman RK, Rao AR, Rai A et al (2020) Metagenomic analysis reveals bacterial and fungal diversity and their bioremediation potential from sediments of River Ganga and Yamuna in India. Front Microbiol. 11:556136. [https://doi.](https://doi.org/10.3389/fmicb.2020.556136) [org/10.3389/fmicb.2020.556136](https://doi.org/10.3389/fmicb.2020.556136)
- <span id="page-10-1"></span>Behera BK, Patra B, Chakraborty HJ, Sahu P, Rout AK, Sarkar DJ, Parida PK, Raman RK, Rao AR, Rai A, Das BK et al (2020) Metagenome analysis from the sediment of river Ganga and Yamuna: In search of beneficial microbiome. PLoS One. 15(10):e0239594.<https://doi.org/10.1371/journal.pone.0239594>
- <span id="page-10-3"></span>Behera BK, Dehury B, Rout AK, Patra B, Mantri N, Chakraborty HJ, Sarkar DJ, Kaushik NK, Bansal V, Singh I, Das BK et al (2021) Metagenomics study in aquatic resource management: recent trends, applied methodologies and future needs. Gene Rep 25:101372.<https://doi.org/10.1016/j.genrep.2021a.101372>
- <span id="page-10-2"></span>Behera BK, Sahu P, Rout AK, Parida PK, Sarkar DJ, Kaushik NK, Rao AR, Rai A, Das BK, Mohapatra T et al (2021) Exploring microbiome from sediments of River Ganga using a metagenomic approach. Aquat Ecosyst Health Manage 24(4):12–22. <https://doi.org/10.14321/aehm.024.04.04>
- <span id="page-10-11"></span>Besemer K, Singer G, Hödl I, Battin TJ (2009) Bacterial community composition of stream bioflms in spatially variable-fow environments. Appl Environ Microbiol 75(22):7189–7195. [https://](https://doi.org/10.1128/AEM.01284-09) [doi.org/10.1128/AEM.01284-09](https://doi.org/10.1128/AEM.01284-09)
- <span id="page-10-12"></span>Chang Y-J, Chang Y-T, Hung C-H, Lee J-W, Liao H-M, Chou H-L (2014) Microbial community analysis of anaerobic bio-corrosion in diferent ORP profles. Int Biodeterior Biodegrad 95:93– 101. <https://doi.org/10.1016/j.ibiod.2014.04.008>
- <span id="page-10-9"></span>Chen Y, Liu Y, Wang X (2017) Spatiotemporal variation of bacterial and archaeal communities in sediments of a drinking reservoir, Beijing, China. Appl Microbiol Biotechnol 101(8):3379–3391. <https://doi.org/10.1007/s00253-016-8019-1>
- <span id="page-10-0"></span>Chen X, Wang Juncai, You Yimin, Wang Renyuan, Chu Shaohua, Chi Yaowei et al (2022) When nanoparticle and microbes meet: the effect of multi-walled carbon nanotubes on microbial community and nutrient cycling in hyperaccumulator system. J. Hazard. Mater. 423(PT A):126947. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.jhazmat.2021.126947) [jhazmat.2021.126947](https://doi.org/10.1016/j.jhazmat.2021.126947)
- <span id="page-10-10"></span>Cheng H, Hua Z (2016) Efects of hydrodynamic disturbances and resuspension characteristics on the release of tetrabromobisphenol A from sediment. Environ Pollut 219:785–793. [https://doi.](https://doi.org/10.1016/j.envpol.2016.07.055) [org/10.1016/j.envpol.2016.07.055](https://doi.org/10.1016/j.envpol.2016.07.055)
- <span id="page-10-4"></span>Cheng H, Wang Y, Zhu T, Wang L, Xie Z, Hua Z et al (2019b) Efects of hydrodynamic disturbances on biodegradation of tetrabromobisphenol A in water-sediment systems. Environ Sci Pollut Res Int 26(30):31392–31400. [https://doi.org/10.1007/](https://doi.org/10.1007/s11356-019-06291-8) [s11356-019-06291-8](https://doi.org/10.1007/s11356-019-06291-8)
- Cheng H, Hua Z (2018) Distribution, release and removal behaviors of tetrabromobisphenol A in water-sediment systems under prolonged hydrodynamic disturbances. Sci Total Environ 402–10. [https://doi.org/10.1016/j.scitotenv.2018.04.276.](https://doi.org/10.1016/j.scitotenv.2018.04.276)
- <span id="page-11-7"></span>Cheng H, Wang Y, Zhu T, Wang L, Xie Z, Hua Z, Jiang X et al. (2019a). Effects of hydrodynamic disturbances on biodegradation of tetrabromobisphenol a in water-sediment systems. Environ Sci Pollut Res Int 31392–400. [https://doi.org/10.1007/](https://doi.org/10.1007/s11356-019-06291-8) [s11356-019-06291-8](https://doi.org/10.1007/s11356-019-06291-8)
- <span id="page-11-4"></span>Donal R, Gerard A, Hamill; Harold T. Johnston, (2013) Determining propeller induced erosion alongside quay walls in harbours using Artifcial Neural Networks. Ocean Eng 59(1):142–151. [https://](https://doi.org/10.1016/j.oceaneng.2012.10.018) [doi.org/10.1016/j.oceaneng.2012.10.018](https://doi.org/10.1016/j.oceaneng.2012.10.018)
- <span id="page-11-28"></span>Egbe CC, Oyetibo G, Oyetibo G, Ilori MO (2021) Ecological impact of organochlorine pesticides consortium on autochthonous microbial community in agricultural soil. Ecotoxicol Environ Saf 207:111319.<https://doi.org/10.1016/j.ecoenv.2020.111319>
- <span id="page-11-24"></span>Fang J, Dong Junyu, Li Changchao, Chen Hao, Wang Lifei, Lyu Tianshu et al (2021) Response of microbial community composition and function to emergent plant rhizosphere of a constructed wetland in northern China. Appl Soil Ecol 168(61):104141. [https://](https://doi.org/10.1016/j.apsoil.2021.104141) [doi.org/10.1016/j.apsoil.2021.104141](https://doi.org/10.1016/j.apsoil.2021.104141)
- <span id="page-11-1"></span>Gao Y, Zhang W, Li Y, Wu H, Yang N, Hui C (2021) Dams shift microbial community assembly and imprint nitrogen transformation along the Yangtze River. Water Res 189:116579. [https://doi.](https://doi.org/10.1016/j.watres.2020.116579) [org/10.1016/j.watres.2020.116579](https://doi.org/10.1016/j.watres.2020.116579)
- <span id="page-11-20"></span>Grant SB, Azizian M, Cook P, Boano F, Rippy MA (2018) Factoring stream turbulence into global assessments of nitrogen pollution. Science 359(6381):1266–1269. [https://doi.org/10.1126/science.](https://doi.org/10.1126/science.aap8074) [aap8074](https://doi.org/10.1126/science.aap8074)
- <span id="page-11-16"></span>Hong J-H, Chiew Y-M, Hsieh S-C, Cheng N-S, Yeh P-H (2016) Propeller jet–induced suspended-sediment concentration. J Hydraul Eng 142(4):4015064. [https://doi.org/10.1061/\(ASCE\)HY.1943-7900.](https://doi.org/10.1061/(ASCE)HY.1943-7900.0001103) [0001103](https://doi.org/10.1061/(ASCE)HY.1943-7900.0001103)
- <span id="page-11-13"></span>Hu D, Hong H, Rong B, Wei Y, Zeng J, Zhu J, Bai L, Guo F, Yu X et al (2021) A comprehensive investigation of the microbial risk of secondary water supply systems in residential neighborhoods in a large city. Water Res. 205:117690. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.watres.2021.117690) [watres.2021.117690](https://doi.org/10.1016/j.watres.2021.117690)
- <span id="page-11-29"></span>Hua Z, Yu L, Liu X, Zhang Y, Ma Y, Lu Y et al (2021) Perfuoroalkyl acids in surface sediments from the lower Yangtze River: Occurrence, distribution, sources, inventory, and risk assessment. Sci Total Environ. 798:149332. [https://doi.org/10.1016/j.scitotenv.](https://doi.org/10.1016/j.scitotenv.2021.149332) [2021.149332](https://doi.org/10.1016/j.scitotenv.2021.149332)
- <span id="page-11-15"></span>Jalil A, Li Y, Zhang Ke, Gao X, Wang W, Khan HO, Sarwar A et al (2019) Wind-induced hydrodynamic changes impact on sediment resuspension for large, shallow Lake Taihu. China Int J Sediment Res 34(3):205–215.<https://doi.org/10.1016/j.ijsrc.2018.11.003>
- <span id="page-11-26"></span>Kim JM, Roh A-S, Choi S-C, Kim E-J, Choi M-T, Ahn B-K et al (2016) Soil pH and electrical conductivity are key edaphic factors shaping bacterial communities of greenhouse soils in Korea. J Microbiol 54(12):838–845.<https://doi.org/10.1007/s12275-016-6526-5>
- <span id="page-11-18"></span>Lam W-H, Hamill G, Robinson D, Raghunathan S (2010) Observations of the initial 3D flow from a ship's propeller. Ocean Eng 37(14– 15):1380–1388. <https://doi.org/10.1016/j.oceaneng.2010.07.008>
- <span id="page-11-5"></span>Lam W, Hamil GA, Song YC, Robinson DJ, Raghunathan S (2011) A review of the equations used to predict the velocity distribution within a ship's propeller jet. Ocean Eng 38(1):1–10. [https://doi.](https://doi.org/10.1016/j.oceaneng.2010.10.016) [org/10.1016/j.oceaneng.2010.10.016](https://doi.org/10.1016/j.oceaneng.2010.10.016)
- <span id="page-11-6"></span>Lam WH, Hamill GA, Robinson DJ (2013) Initial wash profles from a ship propeller using CFD method. Ocean Eng 72:257–266. [https://](https://doi.org/10.1016/j.oceaneng.2013.07.010) [doi.org/10.1016/j.oceaneng.2013.07.010](https://doi.org/10.1016/j.oceaneng.2013.07.010)
- <span id="page-11-10"></span>Legendre P (2014) Interpreting the replacement and richness diference components of beta diversity. Global Ecol Biogeogr 23(11):1324– 1334.<https://doi.org/10.1111/geb.12207>
- <span id="page-11-11"></span>Leprieur F, Tedesco PA, Hugueny B, Beauchard O, Dürr HH, Brosse S, Oberdorff T et al (2011) Partitioning global patterns of freshwater fish beta diversity reveals contrasting signatures of past climate changes. Ecol Lett. 14(4):325–34. [https://doi.org/10.1111/j.1461-](https://doi.org/10.1111/j.1461-0248.2011.01589.x) [0248.2011.01589.x](https://doi.org/10.1111/j.1461-0248.2011.01589.x)
- <span id="page-11-12"></span>Li X-Q, Hua Z (2021) Multiphase distribution and spatial patterns of perfuoroalkyl acids (PFAAs) associated with catchment characteristics in a plain river network. Chemosphere. 263:128284. <https://doi.org/10.1016/j.chemosphere.2020.128284>
- <span id="page-11-23"></span>Li W, Zhang Shaokang, Zhang Lieyu, Li Xiaoguang, Wang Fan, Li Guowen et al (2019) In-situ remediation of sediment by calcium nitrate combined with composite microorganisms under low-DO regulation. Sci Total Environ. 697:134109. [https://doi.org/10.](https://doi.org/10.1016/j.scitotenv.2019.134109) [1016/j.scitotenv.2019.134109](https://doi.org/10.1016/j.scitotenv.2019.134109)
- <span id="page-11-2"></span>Li Y, Gao Yu, Zhang W, Wang C, Wang P, Niu L et al (2019b) Homogeneous selection dominates the microbial community assembly in the sediment of the Three Gorges Reservoir. Sci Total Environ 690:50–60.<https://doi.org/10.1016/j.scitotenv.2019.07.014>
- <span id="page-11-8"></span>Li C, Quan Quan, Gan Yandong, Dong Junyu, Fang Jiaohui, Wang Lifei et al (2020) Efects of heavy metals on microbial communities in sediments and establishment of bioindicators based on microbial taxa and function for environmental monitoring and management. Sci Total Environ. 749:141555. [https://doi.org/10.1016/j.scitotenv.](https://doi.org/10.1016/j.scitotenv.2020.141555) [2020.141555](https://doi.org/10.1016/j.scitotenv.2020.141555)
- <span id="page-11-21"></span>Li Y, Wan M, Du J, Lin L, Cai W, Wang L (2020) Microbial enhanced corrosion of hydraulic concrete structures under hydrodynamic conditions: microbial community composition and functional prediction. Constr. Build. Mater. 248:118609. [https://doi.org/10.](https://doi.org/10.1016/j.conbuildmat.2020.118609) [1016/j.conbuildmat.2020.118609](https://doi.org/10.1016/j.conbuildmat.2020.118609)
- <span id="page-11-14"></span>Liu T, Zhang AN, Wang J, Liu S, Jiang X, Dang C, Ma T, Liu S, Chen Q, Xie S, Zhang T et al (2018) Integrated biogeography of planktonic and sedimentary bacterial communities in the Yangtze River. Microbiome. 6(1):16.<https://doi.org/10.1186/s40168-017-0388-x>
- <span id="page-11-30"></span>Liu X, Hu Sihai, Sun Ran, Wu Yaoguo, Qiao Zixia, Wang Sichang et al (2021) Dissolved oxygen disturbs nitrate transformation by modifying microbial community, co-occurrence networks, and functional genes during aerobic-anoxic transition. Sci Total Environ. 790:148245.<https://doi.org/10.1016/j.scitotenv.2021a.148245>
- <span id="page-11-9"></span>Liu Y, Huang Yu-Hong, Lü Huixiong, Li Hui, Li Yan-Wen, Mo Ce-Hui et al (2021) Persistent contamination of polycyclic aromatic hydrocarbons (PAHs) and phthalates linked to the shift of microbial function in urban river sediments. J Haz Mat. 414:125416. <https://doi.org/10.1016/j.jhazmat.2021b.125416>
- <span id="page-11-27"></span>Lu B, Qian Jin, Wang Peifang, Wang Chao, Hu Jing, Li Kun et al (2020) Efect of perfuorooctanesulfonate (PFOS) on the rhizosphere soil nitrogen cycling of two riparian plants. Sci Total Environ. 741:140494.<https://doi.org/10.1016/j.scitotenv.2020.140494>
- <span id="page-11-19"></span>Lv J, Yuan Ruiqiang, Wang Shiqin (2021) Water diversion induces more changes in bacterial and archaeal communities of river sediments than seasonality. J Environ Manage. 293:112876. [https://](https://doi.org/10.1016/j.jenvman.2021.112876) [doi.org/10.1016/j.jenvman.2021.112876](https://doi.org/10.1016/j.jenvman.2021.112876)
- <span id="page-11-3"></span>Ma Y, Wang Peng, Hua Zulin, Lu Ying, Yang Yundong (2021) Ship navigation disturbance alters multiphase distribution of perfuoroalkyl acids and increases their ecological risk in waterways. Sci Total Environ. 793:148576. [https://doi.org/10.1016/j.scitotenv.](https://doi.org/10.1016/j.scitotenv.2021.148576) [2021.148576](https://doi.org/10.1016/j.scitotenv.2021.148576)
- <span id="page-11-25"></span>Mastný J, Kaštovská E, Bárta J, Chroňáková A, Borovec J, Šantrůčková H et al (2018) Quality of DOC produced during litter decomposition of peatland plant dominants. Soil Biol Biochem 121(1– 3):221–230.<https://doi.org/10.1016/j.soilbio.2018.03.018>
- <span id="page-11-22"></span>Meng L, Zuo Rui, Wang Jin-sheng, Li Qiao, Du Can, Liu Xin et al (2021) Response of the redox species and indigenous microbial community to seasonal groundwater fluctuation from a typical riverbank fltration site in Northeast China. Ecol. Eng. 159(3):106099.<https://doi.org/10.1016/j.ecoleng.2020.106099>
- <span id="page-11-17"></span>Nadia P, D'Alessandro F, Gaudio R, Tomasicchio GR (2019) Threedimensional analysis of local scouring induced by a rotating ship propeller. Ocean Eng 188:106294 1-106294 11. [https://doi.org/](https://doi.org/10.1016/j.oceaneng.2019.106294) [10.1016/j.oceaneng.2019.106294](https://doi.org/10.1016/j.oceaneng.2019.106294)
- <span id="page-11-0"></span>Paudel Adhikari N, Liu Y, Liu K, Zhang F, Adhikari S, Chen Y et al (2019) Bacterial community composition and diversity in Koshi

River, the largest river of Nepal. Ecol Indic 104:501–511. [https://](https://doi.org/10.1016/j.ecolind.2019.05.009) [doi.org/10.1016/j.ecolind.2019.05.009](https://doi.org/10.1016/j.ecolind.2019.05.009)

- <span id="page-12-20"></span>Prabowo AR, Bae Dong Myung (2019) Environmental risk of maritime territory subjected to accidental phenomena: correlation of oil spill and ship grounding in the Exxon Valdez's case. Results Eng. 4(Supplemen 1):100035. [https://doi.org/10.1016/j.rineng.](https://doi.org/10.1016/j.rineng.2019.100035) [2019.100035](https://doi.org/10.1016/j.rineng.2019.100035)
- <span id="page-12-11"></span>Qiao W, Xie Z, Zhang Y, Liu X, Xie S, Huang J et al (2018) Perfuoroalkyl substances (PFASs) infuence the structure and function of soil bacterial community: Greenhouse experiment. Sci Total Environ 642:1118–1126. [https://doi.org/10.1016/j.scitotenv.2018.](https://doi.org/10.1016/j.scitotenv.2018.06.113) [06.113](https://doi.org/10.1016/j.scitotenv.2018.06.113)
- <span id="page-12-30"></span>Rajeev M, Sushmitha TJ, Aravindraja C, ToletiPandian SRSK (2021) Exploring the impacts of heavy metals on spatial variations of sediment-associated bacterial communities. Ecotoxicol Environ Saf. 209:111808.<https://doi.org/10.1016/j.ecoenv.2020.111808>
- <span id="page-12-8"></span>Rath KM, Fierer N, MurphyRousk DVJ (2019) Linking bacterial community composition to soil salinity along environmental gradients. ISME J 13(3):836–846. [https://doi.org/10.1038/](https://doi.org/10.1038/s41396-018-0313-8) [s41396-018-0313-8](https://doi.org/10.1038/s41396-018-0313-8)
- <span id="page-12-10"></span>Rout AK, Dehury B, ParidaSarkarDasRaiBehera PKDJBKABK et al (2022) Taxonomic profling and functional gene annotation of microbial communities in sediment of river Ganga at Kanpur, India: insights from whole-genome metagenomics study. Environ Sci Pollut Res Int 29(54):82309–82323. [https://doi.org/10.1007/](https://doi.org/10.1007/s11356-022-21644-6) [s11356-022-21644-6](https://doi.org/10.1007/s11356-022-21644-6)
- <span id="page-12-4"></span>Shade A, Peter H, Allison SD, Baho DL, Berga M, Bürgmann H, Huber DH et al (2012) Fundamentals of microbial community resistance and resilience. Front Microbiol 3:417. [https://doi.org/10.3389/](https://doi.org/10.3389/fmicb.2012.00417) [fmicb.2012.00417](https://doi.org/10.3389/fmicb.2012.00417)
- <span id="page-12-25"></span>Shen Y, Monroy GL, Derlon N, Janjaroen D, Huang C, Morgenroth E, Boppart SA, Ashbolt NJ, Liu WT, Nguyen TH (2015) Role of bioflm roughness and hydrodynamic conditions in Legionella pneumophila adhesion to and detachment from simulated drinking water bioflms. Environ Sci Technol. 49(7):4274–82. [https://doi.](https://doi.org/10.1021/es505842v) [org/10.1021/es505842v](https://doi.org/10.1021/es505842v)
- <span id="page-12-16"></span>Si X, Baselga Andrés, Ding Ping (2015) Revealing Beta-diversity patterns of breeding bird and lizard communities on inundated landbridge islands by separating the turnover and nestedness components. PLoS One. 10(5):e0127692. [https://doi.org/10.1371/journ](https://doi.org/10.1371/journal.pone.0127692) [al.pone.0127692](https://doi.org/10.1371/journal.pone.0127692)
- <span id="page-12-14"></span>Simpson GG (1943) Mammals and the nature of continents. Am J Sci 241:1–31. <https://doi.org/10.2475/ajs.241.1.1>
- <span id="page-12-5"></span>Steven DA, Jennifer BH, Martiny., (2008) Resistance, resilience, and redundancy in microbial communities. PNAS 105:11512–11519. <https://doi.org/10.1073/pnas.0801925105>
- <span id="page-12-6"></span>Su Z, Dai T, Tang Y, Tao Y, Huang B, Mu Q et al (2018) Sediment bacterial community structures and their predicted functions implied the impacts from natural processes and anthropogenic activities in coastal area. Mar Pollut Bull 131(Pt A):481–495. [https://doi.org/](https://doi.org/10.1016/j.marpolbul.2018.04.052) [10.1016/j.marpolbul.2018.04.052](https://doi.org/10.1016/j.marpolbul.2018.04.052)
- <span id="page-12-2"></span>Tšertova N, Kisand A, Baty F, Kisand V (2013) Homogeneous microbial diversity in the upper sediment layers of a shallow lake. Aquat Microb Ecol 70(1):77–85. <https://doi.org/10.3354/ame01647>
- <span id="page-12-28"></span>Wang P, Hua Z, Cai Y, Shen X, Li Q, Liu X (2015) Effects of hydrodynamic conditions on the sorption behaviors of aniline on sediment with coexistence of nitrobenzene. Environ Sci Pollut Res Int 22(15):11595–11605.<https://doi.org/10.1007/s11356-015-4399-5>
- <span id="page-12-0"></span>Wang J, Pan F, Soininen J, Heino J, Shen Ji (2016) Nutrient enrichment modifes temperature-biodiversity relationships in largescale feld experiments. Nat Commun 7:13960. [https://doi.org/](https://doi.org/10.1038/ncomms13960) [10.1038/ncomms13960](https://doi.org/10.1038/ncomms13960)
- <span id="page-12-21"></span>Wang L, Zhang J, Li H, Yang H, Peng C, Peng Z et al (2018) Shift in the microbial community composition of surface water and sediment along an urban river. Sci Total Environ 627:600–612. [https://](https://doi.org/10.1016/j.scitotenv.2018.01.203) [doi.org/10.1016/j.scitotenv.2018.01.203](https://doi.org/10.1016/j.scitotenv.2018.01.203)
- <span id="page-12-15"></span>Wang J, He Nianpeng, Wang Yin, Li Jingwen, Li Mingxu (2021) Divergent drivers determine soil bacterial β-diversity of forest and grassland ecosystems in Northwest China. Global Ecol Conserv. 28(19):e01622.<https://doi.org/10.1016/j.gecco.2021a.e01622>
- <span id="page-12-3"></span>Wang L, Li Yi, Zhao Zhe, Cordier Tristan, Worms Isabelle, Lihua AN et al (2021) Microbial community diversity and composition in river sediments contaminated with tetrabromobisphenol A and copper. Chemosphere. 272(2):129855. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.chemosphere.2021.129855) [chemosphere.2021.129855](https://doi.org/10.1016/j.chemosphere.2021.129855)
- <span id="page-12-27"></span>Wei M, Blanckaert K, Heyman J, Li D, Schleiss AJ (2016) A parametrical study on secondary fow in sharp open-channel bends: experiments and theoretical modelling. J Hydgro-Environ Res 13(12):1–13. <https://doi.org/10.1016/j.jher.2016.04.001>
- <span id="page-12-13"></span>Whittaker RH (1960) Vegetation of the Siskiyou Mountains, Oregon and California. Ecol Monogr 30:279–338. [https://doi.org/10.2307/](https://doi.org/10.2307/1948435) [1948435](https://doi.org/10.2307/1948435)
- <span id="page-12-1"></span>Winter C, Hein T, Kavka G, Mach R, L; Farnleitner, Andreas H. (2007) Longitudinal changes in the bacterial community composition of the Danube River: a whole-river approach. Appl Environ Microbiol 73(2):421–431. <https://doi.org/10.1128/AEM.01849-06>
- <span id="page-12-17"></span>Wu Z, Wang X, Chen Y, Cai Y, Deng J (2018) Assessing river water quality using water quality index in Lake Taihu Basin, China. Sci Total Environ 612:914–922. [https://doi.org/10.1016/j.scitotenv.](https://doi.org/10.1016/j.scitotenv.2017.08.293) [2017.08.293](https://doi.org/10.1016/j.scitotenv.2017.08.293)
- <span id="page-12-26"></span>Wu H, Li Yi, Zhang W, Niu L, Gao Yu, Hui C et al (2021a) Hydrostatic pressure infuence activity and assembly of bacterial communities in reservoir sediments. Freshw Biol 66(6):1049–1059. [https://doi.](https://doi.org/10.1111/fwb.13697) [org/10.1111/fwb.13697](https://doi.org/10.1111/fwb.13697)
- <span id="page-12-12"></span>Wu J-Y, Gu Li, Hua Zu-lin, Liang Zhong-Yan, Chu Ke-Jian, He Xin-Xin (2021) Per- poly-fuoroalkyl substances (PFASs) pollution in benthic riverine ecosystem: Integrating microbial community coalescence and biogeochemistry with sediment distribution. Chemosphere. 281:130977. [https://doi.org/10.1016/j.chemosphere.](https://doi.org/10.1016/j.chemosphere.2021.130977) [2021.130977](https://doi.org/10.1016/j.chemosphere.2021.130977)
- <span id="page-12-24"></span>Wu J-Y, Hua Zu-lin, Gu Li (2021) Planktonic microbial responses to perfuorinated compound (PFC) pollution: Integrating PFC distributions with community coalescence and metabolism. Sci Total Environ. 788:147743. [https://doi.org/10.1016/j.scitotenv.2021c.](https://doi.org/10.1016/j.scitotenv.2021c.147743) [147743](https://doi.org/10.1016/j.scitotenv.2021c.147743)
- <span id="page-12-7"></span>Xu G, Sun Z, Fang W, Liu J, Xu X, Lv C (2018) Release of phosphorus from sediments under wave-induced liquefaction. Water Res 144:503–511. <https://doi.org/10.1016/j.watres.2018.07.038>
- <span id="page-12-22"></span>Xu Y, Teng Ying, Wang Xiaomi, Li Ran, Christie Peter (2020) Exploring bacterial community structure and function associated with polychlorinated biphenyl biodegradation in two hydrogenamended soils. Sci Total Environ. 745:140839. [https://doi.org/](https://doi.org/10.1016/j.scitotenv.2020.140839) [10.1016/j.scitotenv.2020.140839](https://doi.org/10.1016/j.scitotenv.2020.140839)
- <span id="page-12-29"></span>Yan Z, Hao Z, Wu H, Jiang H, Yang M, Wang C (2019) Co-occurrence patterns of the microbial community in polycyclic aromatic hydrocarbon-contaminated riverine sediments. J Haz Mat 367:99–108. <https://doi.org/10.1016/j.jhazmat.2018.12.071>
- <span id="page-12-9"></span>Yao Y, Zhao Jiaqi, Miao Lingzhan, Hua J (2022) Efects of sediment physicochemical factors and heavy metals on the diversity, structure, and functions of bacterial and fungal communities from a eutrophic river. Environ Pollut. 303:119129. [https://doi.org/10.](https://doi.org/10.1016/j.envpol.2022.119129) [1016/j.envpol.2022.119129](https://doi.org/10.1016/j.envpol.2022.119129)
- <span id="page-12-23"></span>Yin Y, Wang J (2021) Predictive functional profling of microbial communities in fermentative hydrogen production system using PICRUSt. Int J Hydrog Energy 46(5):3716–3725. [https://doi.org/](https://doi.org/10.1016/j.ijhydene.2020.10.246) [10.1016/j.ijhydene.2020.10.246](https://doi.org/10.1016/j.ijhydene.2020.10.246)
- <span id="page-12-18"></span>Yu L, Liu Xiaodong, Hua Zulin, Zhang Yuan, Xue Hongqin (2022) Spatial and temporal trends of perfuoroalkyl acids in water bodies: a case study in Taihu Lake china (2009-2021). . Environ Pollut. 293:118575. <https://doi.org/10.1016/j.envpol.2021.118575>
- <span id="page-12-19"></span>Zhang W, Gu J, Li Yi, Lin Li, Wang P, Wang C et al (2019a) New insights into sediment transport in interconnected river-lake

systems through tracing microorganisms. Environ Sci Technol 53(8):4099–4108. <https://doi.org/10.1021/acs.est.8b07334>

- <span id="page-13-4"></span>Zhang W, Lei M, Li Yi, Wang P, Wang C, Gao Yu et al (2019b) Determination of vertical and horizontal assemblage drivers of bacterial community in a heavily polluted urban river. Water Res 161:98– 107.<https://doi.org/10.1016/j.watres.2019.05.107>
- <span id="page-13-0"></span>Zhang L, Fang Wangkai, Li Xingchen, Lu Wenxuan, Li Jing (2020) Strong linkages between dissolved organic matter and the aquatic bacterial community in an urban river. Water Res. 184:116089. <https://doi.org/10.1016/j.watres.2020a.116089>
- <span id="page-13-1"></span>Zhang W, Wang Haolan, Li Yi, Lin Li, Hui Cizhang, Gao Yu et al (2020) Bend-induced sediment redistribution regulates deterministic processes and stimulates microbial nitrogen removal in coarse sediment regions of river. Water Res. 170:115315. [https://](https://doi.org/10.1016/j.watres.2019.115315) [doi.org/10.1016/j.watres.2019.115315](https://doi.org/10.1016/j.watres.2019.115315)
- <span id="page-13-5"></span>Zhang H, Sun Liwei, Li Yi, Zhang Wenlong, Niu Lihua, Wang Longfei (2021a) The bacterial community structure and N-cycling gene abundance in response to dam construction in a riparian zone. Environ Res 194:110717. [https://doi.org/10.1016/j.envres.2021a.](https://doi.org/10.1016/j.envres.2021a.110717) [110717](https://doi.org/10.1016/j.envres.2021a.110717)
- <span id="page-13-3"></span>Zhang L, Delgado-Baquerizo Manuel, Shi Yu, Liu Xu, Yang Yunfeng, Chu Haiyan (2021b) Co-existing water and sediment bacteria are driven by contrasting environmental factors across glacierfed aquatic systems. Water Res. 198:117139. [https://doi.org/10.](https://doi.org/10.1016/j.watres.2021.117139) [1016/j.watres.2021.117139](https://doi.org/10.1016/j.watres.2021.117139)
- <span id="page-13-2"></span>Zhao Z, Jiang Yu, Li Q, Cai Y, Yin H, Zhang Lu et al (2017) Spatial correlation analysis of polycyclic aromatic hydrocarbons (PAHs) and organochlorine pesticides (OCPs) in sediments between Taihu Lake and its tributary rivers. Ecotoxicol Environ Saf 142:117– 128.<https://doi.org/10.1016/j.ecoenv.2017.03.039>

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