RESEARCH ARTICLE

Promising co‑inoculation strategies to reduce arsenic toxicity in soybean

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Abstract

Arsenic (As) is the cause for concern worldwide due to its high toxicity. Its presence in agricultural soils and groundwater adversely afects soybean (*Glycine max* L.) growth and yield and also endangers food safety. Plant growth-promoting rhizobacteria (PGPR) could be used as part of cost-efective and eco-friendly strategies to mitigate As phytotoxicity. However, simple inoculation of soybean with PGPR *Bradyrhizobium japonicum* E109 (E109), a common practice in Argentina, is not efective in counteracting the efects of As exposure. Our aim was to assess whether the response of soybean to arsenate (AsV) and arsenite (AsIII) could be helpfully modulated by co-inoculating E109 with the free-living PGPRs *Azospirillum brasilense* Cd (Cd) or *Bacillus pumilus* SF5 (SF5). Co-inoculation with E109+SF5 alleviated As-induced depletion of chlorophyll a and b, and carotenoid content, reaching an increase of 26, 28 y 31%, respectively. It also enhanced nodulation $(15–19%)$ under As exposure. E109 + Cd and E109 + SF5 induced changes in the antioxidant system, which could be related to the maintenance of redox homeostasis. Moreover, As accumulation was reduced by 53% in aerial parts of plants inoculated with E109+Cd, and by 16% in the roots of those inoculated with E109+SF5. The strains selected show interesting potential for the development of biotechnological schemes to improve soybean yield while guaranteeing safer food production.

Keywords Glycine max · Arsenate · Arsenite · Inoculant · Bacillus · Azospirillum

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Introduction

Arsenic (As) is a toxic metalloid that occurs naturally in certain soils and in groundwater. It is found in both organic and inorganic forms, although the latter are prevalent (Singh and Srivastava [2020;](#page-11-0) Jia et al. [2021](#page-10-0)). The relative presence of diferent As species depends on the pH and redox potential of the surrounding environment. Microorganisms are also known to play a crucial role in the As geocycle (Ben Fekih et al. [2018\)](#page-10-1).

The most common inorganic forms of As that adversely afect plants are arsenate (AsV) and arsenite (AsIII). Given that both can become accumulated in plant tissues, their presence in cultivable lands is cause for concern. Aquaporins mediate the uptake of AsIII and phosphate transporters that of AsV. The metalloid is then transferred to the shoots by xylem loading (Bali and Sidhu [2021](#page-10-2)). Once inside the plant, it negatively impacts morphological, physiological, biochemical, and metabolic processes. More precisely, it can disturb root-shoot biomass, chlorophyll content, photosynthetic rate, gas exchange, water content, and sugar metabolism as well as trigger the production of reactive oxygen species (ROS) (Abbas et al. [2018;](#page-9-0) Bali and Sidhu [2021](#page-10-2)). Since ROS are able to engage in reactions with proteins, lipids, carbohydrates, and DNA, they can cause irreparable damage unless scavenged through activation of the antioxidant system (Banerjee and Roychoudhury [2019\)](#page-10-3). This is why enzymatic and non-enzymatic antioxidants constitute one of the main defense mechanisms that determine As tolerance (Zulfqar and Ashraf [2022\)](#page-11-1). Furthermore, As interferes with nodulation and nitrogen fxation in legume-*Rhizobium* interactions (Armendariz et al. [2019;](#page-9-1) Pajuelo et al. [2019](#page-11-2)) and can thus reduce crop yield and afect food safety (Bustingorri and Lavado [2014\)](#page-10-4). Several plant food sources, in fact, have been found to contain harmful levels of As (Upadhyay et al. [2019](#page-11-3)).

Soybean (*Glycine max L*.), a major crop worldwide, is mainly produced in the USA, Brazil, and Argentina (Goonde and Ayana [2021](#page-10-5)). Because of its ability to grow under a wide range of environmental conditions, it has been introduced into marginal lands where it is frequently exposed to As. Evidence of the adverse efects of As has therefore been found in this plant species, both in studies conducted by our lab (Talano et al. [2013;](#page-11-4) Armendariz et al. [2016](#page-9-2); Vezza et al. [2018](#page-11-5), [2021\)](#page-11-6) and by other authors (Bustingorri and Lavado [2014](#page-10-4); Chandrakar et al. [2017](#page-10-6); Fatima et al. [2020](#page-10-7); Gálusová et al. [2020](#page-10-8)).

Several physicochemical methods have been developed to remediate As, but most of them have practical limitations. In addition to being costly, they may generate toxic sludge, be inefficient under natural conditions, and pose operational problems. By contrast, bioremediation is sustainable, costefective, and reliable (Irshad et al. [2021\)](#page-10-9). The potential of As-tolerant plant growth-promoting rhizobacteria (PGPRs) as biofertilizers, phytostimulants, biopesticides, and bioremediators may be harnessed to improve plant performance and contribute to the mitigation of As (Molina et al. [2021](#page-10-10); Kumar et al. [2022\)](#page-10-11). Well-chosen strains could induce the antioxidant defense system, repair photosynthetic activity, improve nutritional status, and help to re-establish symbiotic interaction (Mondal et al. [2021\)](#page-10-12). Microbial activity can also strongly infuence the bioavailability, adsorption/absorption, and translocation of As within the soil–plant system (Srivastava and Shukla [2019](#page-11-7); Khan et al. [2021](#page-10-13)). In short, appropriately selected PGPRs could potentially be used alone or as part of consortia to regulate stress caused by As in host plants.

In Argentina, soybean is frequently inoculated with formulations based on *Bradyrhizobium japonicum* E109 (E109), designed to boost yield. However, previous studies by our laboratory have shown that E109 is not a suffcient strategy to assist plant growth in environments with moderate-to-high As concentrations. Certain benefts have been reported for the co-inoculation of rhizobium with other

strains in leguminous plants subjected to regular and stressful conditions. For instance, the application of *Azospirillum brasilense* strains together with rhizobium was observed to increase biomass, nitrogen content, favonoid signals, nodgene transcription, and grain yield (Dardanelli et al. [2008](#page-10-14); Galindo et al. [2018;](#page-10-15) Rondina et al. [2020;](#page-11-8) Filipini et al. [2021](#page-10-16)). Similar results were obtained after co-inoculating rhizobium and *Bacillus pumilus* strains (Stefan et al. [2010](#page-11-9); Hasibuan et al. [2021](#page-10-17)). Specifcally, *Azospirillum brasilense* Cd (Cd) is a collection strain used for commercial inoculant formulation, while *Bacillus pumilus* SF5 is an endophytic strain isolated from the rhizosphere under drought stress and exhibits interesting PGPR properties and tolerance to a wide range of As concentrations (Forchetti et al. [2007;](#page-10-18) Vezza et al. [2020](#page-11-10)). However, nothing is known about the mechanisms underlying these properties or the strains' ability to contribute to plant success in actual As-contaminated felds.

Here, we explored the effects of co-inoculating soybean with $E109 + Cd$ and $E109 + SF5$ on the plant's response to As during the early vegetative stage. Exposure to AsV and AsIII was considered, since they are the most frequent chemical forms in the environment and they difer in phytotoxicity. Parameters associated with As stress were analyzed, such as those related to growth, nodulation, photosynthetic pigments, the antioxidant system, and As accumulation in plant tissues. Our working hypothesis was that co-inoculation with $E109 + Cd$ and $E109 + SF5$ could be a better strategy than E109 simple inoculation on soybean early development, nodulation, and limitation of As uptake in metalloid-contaminated environments. The fndings are encouraging enough to consider the selected strains for future research on a larger and deeper scale.

Materials and methods

Biological material

Soybean (*Glycine max* L.) cv. Don Mario 4670 seeds were surface disinfected as described by Vezza et al. [\(2018](#page-11-5)). The bacterial strains used for seed inoculation were *Bradyrhizobium japonicum* E109 (E109), *Azospirillum brasilense* Cd (Cd), and *Bacillus pumilus* SF5 (SF5). E109 and Cd are collection strains used to produce commercial inoculant formulations worldwide. SF5 is an endophytic bacteria isolated from sunfower (*Helianthus annuus* L.) roots grown under drought (Forchetti et al. [2007](#page-10-18)), which was gently provided by Dr. S. Alemano.

Experimental design and growth conditions

Experiments were carried out in a completely randomized factorial design with three seed inoculation conditions: E109 simple inoculation and co-inoculations with $E109 + Cd$ or E109+SF5, and three As treatments: without As (control), and exposure to AsV or AsIII as described below.

Soybean inoculation with E109 is a very widespread practice in Argentina, and this strain remains in the soil from one season to the next (Narożna et al. [2015](#page-11-11)), so it constituted the control condition for the comparison to co-inoculation strategies. Non-inoculated condition compared with E109 simple inoculation has already been evaluated in our previous works (Talano et al. [2013;](#page-11-4) Armendariz et al. [2019](#page-9-1)).

E109 was grown in yeast extract mannitol (YEM) medium (Vincent [1970](#page-11-12)), while Luria–Bertani (LB) (Bertani [1951](#page-10-19)) medium was used for the growth of Cd and SF5, at 28 °C and 150 rpm. Cultures in the exponential growth phase were adjusted to 1×10^{-9} , 1×10^{-6} , and 1×10^{-8} CFU mL⁻¹ (colony forming units per milliliter), respectively. Inoculation doses were adjusted to a fnal volume of 250 μL for each 10 g of seeds, using a mixture of equal volumes of each strain for co-inoculations. Inoculated seeds were placed into pots containing sterile perlite as an inert substrate to avoid interferences with As treatment. They were irrigated by capillarity with 1/4 Hoagland solution without nitrogen and kept in a growth chamber (model E15, Conviron, Winnepeg, Canada) under controlled conditions: 14-h light/10-h dark cycle, 200 µmol m⁻² s⁻¹ light intensity, 28 °C, and 80% relative humidity. Seven days after inoculation, plants were irrigated with water without As (control), supplemented with 25-μM sodium arsenate $(Na_2HAsO_4.7H_2O)$ (SIGMA) (AsV) or 25 -μM sodium arsenite (NaAsO₂) (SIGMA) (AsIII). The As concentration used was selected because it is found in many rural areas around the world (Mariño et al. [2020;](#page-10-20) Singh and Srivastava [2020](#page-11-0)) and causes alterations in soybean growth, according to our frst report (Talano et al. [2013\)](#page-11-4). The plants were evaluated at 35 days of growth. Harvested plant tissues were homogenized with liquid $N₂$ and immediately kept at−80 °C until their further use.

Plant growth and nodulation

Dry (DW) and fresh weight (FW) of total aerial part, leaves, and roots were registered. In addition, the number and FW of total nodules and those located in the main root were evaluated.

Photosynthetic pigments

The determination of photosynthetic pigments was performed according to Dere et al. [\(1998](#page-10-21)). For this, 100 mg of fresh leaf tissues were homogenized with 1 mL of 96% methanol. After centrifugation at 5000 rpm for 7 min, supernatant absorbance was measured at 662 nm, 645 nm, and 470 nm. The content of chlorophyll a (Ca), chlorophyll b (Cb), and carotenoids (C_{x+c}) was calculated using

the following formulas: Ca = $15.65 \times A_{666} - 7.34 \times A_{653}$; $Cb = 27.05 \times A_{653} - 11.21 \times A_{666}$; $C_{x+c} = 1000 \times A_{470}$ $-2.86 \times Ca - 129.2 \times Ch/245$. The results were expressed as mg g⁻¹ FW.

Antioxidant system

Antioxidant enzyme activity

Enzyme extraction was performed by homogenizing plant tissues (leaves and roots separately) with bufer solution $(1:10 \text{ w/v})$ [50 mM KH₂PO₄/K₂HPO₄ buffer pH 7.8, 0.5 mM EDTA, and polyvinylpolypyrrolidone (PVPP)] and subsequent centrifugation at 10,000 rpm and 4 °C for 30 min. After that, the supernatant was used for assays of total peroxidase (Px), ascorbate peroxidase (APx), and superoxide dismutase (SOD) activities. Total Px activity was determined spectrophotometrically with o-dianisidine and H_2O_2 as substrates, following the appearance of a colored product [bis (3,3-dimethoxy-4amino) azo-biphenyl] (ε 11.3 mM⁻¹ cm⁻¹) at 470 nm (Agostini et al. [1997](#page-9-3)). One Px unit was defned as the amount of enzyme that generates 1 μmol of the colored product after 1 min of reaction. APx activity was determined following the oxidation of L-ascorbic acid (ε 2.8 M⁻¹ cm⁻¹) at 290 nm (Hossain and Asada [1984\)](#page-10-22). One APx unit was defned as the amount of enzyme that produced the oxidation of 1 μmol of ascorbic acid after 1 min of reaction. SOD activity was determined according to Beauchamp and Fridovich ([1973\)](#page-10-23) with some modifcations. An SOD unit was defned as the amount of enzyme required to cause 50% inhibition of the rate of photochemical reduction of nitroblue tetrazolium (NBT) at 560 nm (Armendariz et al. [2016](#page-9-2)).

Total antioxidant activity

Total antioxidant activity (TAA) was determined as the capacity of radical scavenging against stable DPPH (2,2-diphenyl-1-picrylhydrazyl) according to Brand-Williams et al. [\(1995](#page-10-24)) with some modifcations. Leaf and root tissues (200 mg) were homogenized separately with 1 mL of 50% (v/v) methanol, incubated in ice for 2 h (vortexed every 30 min) and centrifuged at 10,000 rpm and 4 °C for 15 min. The supernatants (15 μL and 80 μL from leaf and root extracts, respectively) were mixed with a necessary amount of 60 μM DPPH/methanol solution to complete a fnal volume of 1 mL and then incubated in the dark for 30 min at room temperature. The absorbance was measured at 515 nm before and after incubation. Finally, TAA was calculated and expressed as follows: DPPH inhibition $(\%) = [(IA-FA)/$ IA \times 100, where IA is the initial absorbance (before incubation) and FA is the fnal absorbance (after incubation).

Total As concentration

Total As concentration in soybean leaves and roots was measured using homogenized oven-dried material. For this, an optimized method comprising fow injection hydride generation atomic absorption spectroscopy (FI-HGAAS) was used, according to Sigrist et al. [\(2016](#page-11-13)). Data were expressed as μ g As per g⁻¹ DW, and translocation factor (TF) was calculated as follows: TF $(\%) =$ (As concentration in leaves/As concentration in roots) \times 100.

Statistical analysis

The data were submitted to normality and homogeneity of variance assumptions using Shapiro–Wilk and Levene tests, respectively. When these assumptions were confrmed, the comparisons between the diferent treatments were carried out using an analysis of variance (ANOVA), followed by a comparison of multiple treatment levels using Tuckey's test. Differences were considered to be significant for $p < 0.05$. InfoStat software (v. 2012e; InfoStat, National University of Cordoba, Argentina) was used for this.

Results

Plant growth and nodulation

Growth-related parameters were evaluated in soybean plants during the early vegetative stage (V5). As a result of exposure to As, biomass production was around 25–39% lower in aerial parts and 16–20% lower in roots compared to the control (without As) (Table [1\)](#page-3-0). Co-inoculation with $E109 + Cd$ and $E109 + SF5$ did not disrupt plant growth: The values obtained were the same as those for simple inoculation with E109 (Table [1](#page-3-0)).

Nodules on the main root (Fig. $1a$) and total nodules (Fig. [1b](#page-4-0)) were also registered. The amount of nodules on the lateral roots was very few, and therefore, it was not reported here. The adverse efects of As were more pronounced on the nodules' FW than on their number. In plants inoculated only with E109, the FW of the main root nodules was reduced by 20 and 15% in the presence of AsV and AsIII, respectively (Fig. [1a](#page-4-0)). Co-inoculation led instead to increased nodule FW $(17\%$ for E109 + Cd and 14% for E109 + SF5), in comparison to simple inoculation under AsV treatment. Still, co-inoculation with E109+SF5 in plants exposed to AsIII, specifcally, caused a more relevant increase of 20% with respect to simple inoculation (Fig. [1a](#page-4-0)).

Photosynthetic pigments

The presence of As considerably reduced the content of chlorophyll a (Fig. $2a$) and b (Fig. $2b$) in plants inoculated only with E109. The effects were more severe with AsV (28% reduction in type a and 25% reduction in type b) than with AsIII (19 and 16% less, respectively). There was also a decrease in carotenoid content (of around 25%) under AsV treatment (Fig. $2c$). In plants exposed to AsV, small increases in the photosynthetic pigment content were obtained after co-inoculation with $E109 + Cd$, compared to E109 on its own. More signifcant advantages were achieved with $E109 + SF5$, since these plants had a higher content of chlorophyll a (26 and 13%), b (28 and 14%), and carotenoids (31 and 9%) compared to those inoculated with E109 under AsV and AsIII stress, respectively (Fig. [2\)](#page-5-0).

Table 1 Plant growth parameters in inoculated soybean plants exposed to AsV and AsIII

| | | Total aerial part | | Leaves | | Roots | |
|---------|------------------|-------------------|------------------|------------------|------------------|--------------------|--------------------|
| | | FW | DW | FW | DW | FW | DW |
| Control | E109 | $1.81 + 0.08 Ab$ | $0.45 + 0.02 Ab$ | $0.89 + 0.04 Ab$ | $0.23 + 0.01 Ab$ | $1.69 + 0.07 Ab$ | 0.18 ± 0.01 Ab |
| | $E109 + Cd$ | $1.91 + 0.09 Ab$ | $0.49 + 0.03$ Ab | $0.92 + 0.04 Ab$ | $0.24 + 0.01 Ab$ | $1.70 + 0.06 Ab$ | 0.18 ± 0.01 Ab |
| | $E109 + SF5$ | $1.85 + 0.07 Ab$ | $0.46 + 0.02 Ab$ | $0.92 + 0.04 Ab$ | $0.23 + 0.01$ Ac | 1.54 ± 0.05 Ab | 0.18 ± 0.01 Ab |
| AsV | E ₁₀₉ | $1.33 + 0.07$ Aa | $0.30 + 0.02$ Aa | $0.67 + 0.03$ Aa | $0.14 + 0.01$ Aa | $1.45 + 0.07$ Aa | $0.16 + 0.01$ Aa |
| | $E109 + Cd$ | $1.38 + 0.06$ Aa | $0.31 + 0.01$ Aa | $0.67 + 0.03$ Aa | $0.14 + 0.01$ Aa | $1.37 + 0.05$ Aa | $0.15 + 0.01$ Aa |
| | $E109 + SF5$ | $1.39 + 0.06$ Aa | $0.33 + 0.02$ Aa | $0.64 + 0.03$ Aa | $0.14 + 0.01$ Aa | 1.33 ± 0.05 Aa | 0.15 ± 0.01 Aa |
| AsIII | E ₁₀₉ | $1.40 + 0.08$ Aa | $0.33 + 0.02$ Aa | $0.70 + 0.04$ Aa | $0.16 + 0.01$ Aa | 1.34 ± 0.08 Aa | 0.16 ± 0.01 Aa |
| | $E109 + Cd$ | $1.38 + 0.05$ Aa | $0.33 + 0.01$ Aa | $0.66 + 0.02$ Aa | $0.15 + 0.01$ Aa | $1.31 + 0.04$ Aa | $0.15 + 0.01$ Aa |
| | $E109 + SF5$ | $1.43 + 0.06$ Aa | $0.36 + 0.02$ Aa | $0.69 + 0.03$ Aa | $0.17 + 0.01 Ab$ | $1.29 + 0.05$ Aa | $0.15 + 0.01$ Aa |

Results represent the mean \pm SE ($n=15-20$). Different capital letters indicate significant differences between inoculation conditions for the same As treatment, while diferent lowercase letters indicate signifcant diferences between As treatment for the same inoculation condition (Tukey test, *p* ≤ 0.05)

Fig. 1 Nodule FW and number from the main root (**a**) and all the roots (total) (**b**) in inoculated soybean plants exposed to AsV and AsIII. Results represent the mean \pm SE (*n* = 15–20). Diferent capital letters indicate signifcant diferences between inoculation conditions for the same As treatment, while different lowercase letters indicate signifcant diferences between As treatment for the same inoculation condition (Tukey test, $p \le 0.05$)

Nodule FW • Nodule number

Antioxidant system

The activity of antioxidant enzymes (APx, Px, and SOD) and TAA was estimated in leaves (Fi[g](#page-6-0). $3a$, [c,](#page-6-0) [e](#page-6-0) and g) and roots (Fig. [3b, d](#page-6-0), [f](#page-6-0) and [h\)](#page-6-0). Diferences were found between As-stressed and control plants, and in turn, the responses exhibited variations depending on the inoculation strategy applied.

In leaves, APx increased signifcantly in E109-inoculated plants exposed to either AsV or AsIII. When the combination E109+Cd was used, APx rose only under AsV treatment, and this increment was greater in magnitude than that observed after simple inoculation with E109 (Fig. [3a](#page-6-0)). Both chemical forms of As led to an increase in Px activity with respect to the control under all inoculation conditions, but similar to what happened with APx (Fig. [3a](#page-6-0)), the enhancement was greater with $E109 + Cd$ under AsV treatment (Fig. [3c\)](#page-6-0). In general, SOD did not vary in any relevant way from one condition to the other (Fig. [3e](#page-6-0)). Regarding TAA, higher values were obtained through inoculation with E109 in plants exposed to AsV, and with $E109 + Cd$ in plants exposed to AsIII. The combination $E109 + SF5$, for its part, rendered higher TAA levels under both As treatments (Fig. [3g](#page-6-0)).

In roots, APx decreased signifcantly in plants inoculated with E109 and $E109 + Cd$ under AsV treatment, while activity values in those inoculated with E109+SF5 were similar to the control (Fig. [3b](#page-6-0)). No signifcant variations under the diferent conditions were observed for Px (Fig. [3d](#page-6-0)). Coinoculation with E109+SF5 improved SOD activity upon a

exposure to AsV and AsIII. The use of $E109 + Cd$ also enhanced the activity of this enzyme under AsIII treatment (Fig. [3f\)](#page-6-0). A similar trend was found for TAA, which was higher in plants co-inoculated with E109 + SF5 and exposed to AsV (Fig. [3h](#page-6-0)).

Total As concentration

Total As content was determined in aerial parts (Fig. [4a\)](#page-7-0) and roots (Fig. [4b\)](#page-7-0). The metalloid was more accumulated in the latter than in the former. In plants inoculated with E109 alone, the TF was 5.4% upon AsV exposure and 5.7% upon AsIII exposure (Fig. [4c](#page-7-0)). Co-inoculation with $E109 + Cd$ reduced the As content by 53% in aerial parts with respect to E109-inoculated plants under AsV treatment (Fig. [4a](#page-7-0)). Accordingly, TF was also lower (2.7%) under these conditions (Fig. [4c](#page-7-0)). Moreover, the roots of plants inoculated with E109+SF5 and treated with AsIII featured around 16% less total As than those otherwise inoculated (Fig. [4b](#page-7-0)).

Discussion

The presence of As in the environment poses a serious risk to the growth of crops such as soybean, and also in terms of food-chain contamination. In agricultural soils with high levels of As, solutions to this issue could take the form of inoculation strategies based on the use of PGPRs, such as those tested here: $E109$, $E109 + Cd$, and $E109 + SF5$.

In agreement with previous reports, we observed that soybean plants exposed to stress by AsV and AsIII were

impaired in their growth in comparison to the respective controls (Table [1\)](#page-3-0) (Talano et al. [2013](#page-11-4); Armendariz et al. [2016;](#page-9-2) Vezza et al. [2019](#page-11-14)). In addition, a signifcant reduction in nodulation was detected after inoculation with E109 (Fig. [1](#page-4-0)). Arsenic causes alterations in photosynthesis, nutrient uptake, phytohormone balance, and oxidative status, all of which contribute to defcient plant development (Bali and Sidhu [2021](#page-10-2); Khan et al. [2021\)](#page-10-13). The application of strains Cd and SF5 in combination with E109 did not afect plant growth (Table [1](#page-3-0)), but did slightly improve nodulation, mainly in plants exposed to AsIII and inoculated with E109+SF5 (Fig. [1](#page-4-0)). The impact of PGPRs on biometric parameters could largely depend on crop management practices, environmental conditions, and the plant's phenological stage. The scope of the present study was limited to the early vegetative stage, since the assay was conducted in a growth chamber. When it comes to other stages, Stefan et al. [\(2010](#page-11-9)) found the inoculation of soybean with *B. pumilus* Rs3 to have more marked efects during fowering and fruiting. Studies on the inoculation of several crops with *Azospirillum*, on the other hand, have come up with multi-causal variable results, as in the case of the soybean-*A. brasilense* association analyzed by Cassán et al. [\(2020\)](#page-10-25).

Assessing photosynthetic pigment content is a good method to indirectly estimate plant productivity. Chlorophyll metabolism is usually sensitive to changes in the environment and in the plant's physiological state, so pigment content could be an indicator of how the plant responds to stress (Sharma et al. [2020](#page-11-15)). The efects of metal(loid)s depend on the chemical species in question. For instance, soybean plants exposed to AsV experienced more noticeable

 $\mathsf a$ 3.5

Fig. 3 Antioxidant response in leaves (**a**, **c**, **e**, **g**) and roots (**b**, **d**, **f**, **h**) in soybean plants exposed to AsV and AsIII and under diferent inoculation conditions. Results represent the mean±SE ($n=6$). Different letters indicate significant differences (Tukey test, $p \le 0.05$)

a

photosynthetic pigment depletion than those under AsIII treatment (Fig. [2\)](#page-5-0). This becomes more evident after prolonged periods of exposure and intensifes when As concentrations increase (Armendariz et al. [2016](#page-9-2)). Shahzad et al. ([2021](#page-11-16)) did not detect symptoms of chlorosis or necrosis in maize plants exposed to cadmium and inoculated with *Bacilus pumilus*, which suggests that appropriate inoculants can alleviate the damage wrought by exposure to metals. Likewise, soybean plants treated with AsV had markedly higher levels of chlorophyll (Fig. [2a](#page-5-0) and [b\)](#page-5-0) and carotenoids (Fig. [2c](#page-5-0)) when co-inoculated with $E109 + SF5$ than when receiving only E109. This co-inoculation strategy contributes to counteracting the harmful infuence of As, since pigment content did not signifcantly difer between the control (without As) and the plants treated with either form of As (Fig. [2\)](#page-5-0). The $E109 + Cd$ co-inoculation also led to improvements in pigment content over inoculation with E109, although the diferences were not statistically signifcant (Fig. [2](#page-5-0)). The dependence of photosynthetic pigment content on the presence and ratio of mineral elements was established for diferent crops (Bojović and Stojanović, [2005](#page-10-26)), and PGPRs could be responsible for increased mineral bioavailability. Phosphate solubilization and siderophore production have been reported for SF5 and Cd, respectively, and both strains have shown nitrogen fxation capacity (Forchetti et al. [2007](#page-10-18); Vezza et al. [2020\)](#page-11-10). These properties may

work synergistically with the biofertilizing abilities of E109, which have proven to be very sensitive to As (Talano et al. [2013](#page-11-4); Armendariz et al. [2019](#page-9-1)). Furthermore, a decrease in photosynthetic pigments has been posited as one of the injuries caused by metal(loid)-induced oxidative stress, so redox state regulation may play a vital role (Awan et al. [2020](#page-9-4)).

ROS are intermediates in several cellular cascades and play an important part in cell signaling. However, their production is exacerbated by the uptake of As, and this can cause oxidative damage to biologically relevant macromol-ecules (Chandrakar et al. [2017](#page-10-6)). The damage can be offset by non-enzymatic and enzymatic antioxidants (Armendariz et al. [2016](#page-9-2); Vezza et al. [2019\)](#page-11-14). Interestingly, our results suggest that the bacterial strains chosen for inoculation are able to modulate antioxidant defense in As-stressed soybean plants (Fig. [3](#page-6-0)). Co-inoculation with Cd and SF5 induced noticeable diferences in the plant's response compared to that observed with E109 alone. For example, APx and Px were up-regulated in the leaves of plants inoculated with $E109 + Cd$ (Fig. [3a](#page-6-0) and [c\)](#page-6-0), while APx and SOD activity was enhanced in the roots of those inoculated with $E109 + SF5$ under exposure to AsV (Fig. $3b$ and [f](#page-6-0)). Besides, the redox potential determined from the non-enzymatic components and estimated through TAA was improved in leaves and roots under treatment with AsIII and AsV, respectively, due to the application of $E109 + Cd$ and $E109 + SF5$ (Fig. [3g](#page-6-0) and

[h](#page-6-0)). This could be the result of transcriptional and translational regulation, since PGPRs have afected the expression of genes encoding diferent antioxidant enzymes, as was described for tomato plants exposed to cadmium (Khanna et al. [2019a\)](#page-10-27). The effects on antioxidant defense appear to be dependent on the strain, plant organ, chemical As form, and antioxidant components. The activity of catalase (CAT), SOD, and Px isoforms followed diferent trends when rice under stress by AsV was inoculated with *B. thuringiensis* and *P. glucanolytus* (Banerjee et al. [2020](#page-10-28)). In soybean exposed to cadmium, *B. cereus* induced an increase in SOD and CAT but down-regulated the activity of other enzymes such as Px (Sahile et al. [2021\)](#page-11-17). Other examples of fluctuations in the antioxidant defense machinery modulated by PGPRs were described by Qadir et al. ([2020](#page-11-18)) and Saleem et al. ([2018\)](#page-11-19) for sunfower plants treated with chromium and lead, respectively. What all the reports have in common, however, is the fact that they found a beneficial correlation between changes in the antioxidant system mediated by PGPRs, free radical scavenging, and, hence, the mitigation of oxidative stress brought about by metal(loid)s. Therefore, the inoculation of soybean with $E109 + Cd$ and $E109 + SF5$ may very well contribute to creating a state of adequate antioxidant activity that would maintain cellular redox homeostasis in the presence of As.

Another key issue is the accumulation of As in plant tissues, not only due to its direct impact on metabolism but also because of the likelihood of edible parts becoming contaminated (Corguinha et al. [2015](#page-10-29)). Cd and SF5 could also be benefcial in this respect, since their application in combination with E109 noticeably impacted on the content and distribution of As in soybean plants. $E109 + Cd$ markedly reduced the amount of As in aboveground tissues and allowed for improved rhizostabilization under AsV treatment (Fig. [4a](#page-7-0)). A signifcant decrease in As accumulation was also observed in roots exposed to AsIII when they were inoculated with $E109 + SF5$ (Fig. [4b](#page-7-0)).

The plant responses described so far may be mediated by PGPRs modulating phytohormone levels. Indole acetic acid (IAA) produced by Cd could induce As translocation changes in soybean. Accordingly, inoculation of wheat with *A. brasilense* Sp 245 was found to decrease As accumulation (Lyubun et al. [2006\)](#page-10-30), while *A. lipoferum* 137 modifed cadmium content in barley roots (Belimov and Dietz [2000\)](#page-10-31) and *A. brasilense* Az39 reduced As translocation to soybean plant shoots (Armendariz et al. [2019\)](#page-9-1). Similar effects were obtained with IAA-producing bacteria belonging to other genera (Wang et al. [2017;](#page-11-20) Das and Sarkar [2018\)](#page-10-32), as well as with the exogenous application of IAA (Zhu et al. [2013](#page-11-21)). Several authors have suggested that IAA could improve the synthesis of phytochelatins (PCs) and, by this means, increase the capacity for As chelation in roots (Pál et al. [2018](#page-11-22); Singh et al. [2021\)](#page-11-23). PCs are known to chelate As and mediate its compartmentalization within the cell vacuole, thus immobilizing it. The mobility of As in apoplastic compartments and/or the surrounding soil and thus its entry into root cells could also be slowed down by other bacteriaassisted mechanisms, such as As bioaccumulation, precipitation, oxidation–reduction, and biosorption to surface components like bioflm (Mondal et al. [2021\)](#page-10-12). Microbial ability to convert AsIII into dimethyl AsV [DMA(V)], trimethylarsine [TMA(III)], and then TMA(III) oxide (TMAO), which is volatile, could also be harnessed to decrease As bioavailability in soils (Guo et al. [2020](#page-10-33)). Events such as these might be mediated by SF5 and explain the decrease in AsIII uptake by soybean roots when inoculated with this strain (Fig. [4b](#page-7-0)). In addition, PGPR-supplemented plants are thought to feature a down-regulation of metal transporter gene expression (Khanna et al. [2019b\)](#page-10-34).

Our main findings are summarized in the diagram in Fig. [5.](#page-9-5) We believe that several mechanisms acting synergistically could be responsible for the attenuation of the negative effects by As that was observed after coinoculating soybean with $E109 + Cd$ and $E109 + SF5$. Nevertheless, the abilities of Cd and SF5 to bioremediate and respond to As are still poorly understood. The present work was designed as an exploratory, laboratory-scale study to observe the benefits of Cd and SF5 on soybean stressed by AsV and AsIII. The results should encourage further studies aimed at elucidating the underlying mechanisms and testing the strains' potential under field and scaled-up conditions.

Conclusions

Our findings evidence that the response of soybean to As can be modulated by bacterial inoculants, in a manner dependent on the combination of strains used and the chemical As form to which the plants are exposed. In general terms, seed co-inoculation with $E109 + Cd$ and E109+SF5 was advantageous compared to simple inoculation with E109. The positive impact on nodulation, photosynthetic pigment content, and the activity of the antioxidant system, as well as the significant reduction in As accumulation in plant tissues exposed to AsV and AsIII, constitute novel findings attributable to the co-inoculation strategies tested here. Therefore, new research lines are opened to study the potential use of $E109 + Cd$ and E109 + SF5 for the formulation of inoculants to improve soybean performance under stress by As and reduce the metalloid's entry into the food chain. A better understanding of the modes of action that restrict As uptake and mobility, among others, will be necessary to exploit the strains' beneficial properties in polluted fields.

of As

Ref.: As: arsenic; APx: ascorbate peroxidase; Px: peroxidases; TAA: total antioxidant activity; SOD: superoxide dismutase

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Author contribution MEV, EA, and **MAT**: research design. **MEV**: methodology, investigation, writing – original draft. **RPP**: data collection. **ALWO**: writing – review and editing. **EA and MAT**: funding acquisition, supervision, writing – review and editing.

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Data Availability The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Declarations

Ethics approval Not applicable.

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