



Acute toxicity and teratogenicity of carbaryl (carbamates), tebufenpyrad (pyrazoles), cypermethrin and permethrin (pyrethroids) on the European sea bass (*Dicentrarchus labrax* L, 1758) early life stages

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Abstract

The toxicity of carbaryl, tebufenpyrad, cypermethrin and permethrin was evaluated in European sea bass *Dicentrarchus labrax* during the embryonic and larval development using six different concentrations per chemical. The order of the toxicity effectiveness was carbaryl > tebufenpyrad > cypermethrin > permethrin. The larvae were more sensitive to all tested chemicals than embryos. The LC50 of carbaryl, tebufenpyrad, cypermethrin and permethrin was determined as 13.88, 43.96, 92 and 142 ppm and 9.27, 25.67, 48.4 and 72.7 ppm in embryo and larvae, respectively. Furthermore, the tested pesticides exhibited teratogenic effects on *D. labrax* embryo-larval stages. The observed malformations were coagulation, no spherical egg, unhatched egg, pericardial oedemata, yolk oedemata, lordosis, kyphosis, scoliosis, no eye, cranial deformation and body atrophy. Malformations were induced with 0.5 ppm carbaryl, 10 ppm tebufenpyrad and 50 ppm cypermethrin and permethrin; the highest rates of malformation were noted with 16 ppm carbaryl, 160 ppm tebufenpyrad, 400 ppm cypermethrin and 400 ppm permethrin as 34.5%, 28%, 17.5% and 16%, respectively. A positive correlation between the incidence of malformation and the increase of pesticide concentration was established.

Keywords Carbaryl · Tebufenpyrad · Cypermethrin · Permethrin · *Dicentrarchus labrax* · Early life stages · Acute toxicity · Teratogenicity

Introduction

Pesticide usage has considerably increased during the last decade specially with the development of the agriculture; in fact, the worldwide use of those compounds was estimated to be 3 billion kilograms each year (Hayes et al. 2017). Definitely, the use of pesticides has become required in order to avoid losses of agricultural products, which were estimated

to exceed 70% for fruit, 50% for vegetable and 30% for cereal (Tudi et al. 2021). Pesticides were also used in urban and industrial activities, which contributes to the continual increase of pesticide occurrence in the aquatic ecosystems (Syafrudin et al. 2021). This is not without consequences on non-target species, particularly fish (Hernández-Moreno et al. 2011; Syafrudin et al. 2021).

Carbaryl or Sevin was commercialized for the first time in 1958. It is commonly used in agriculture for the control of pests, and as a molluscicide, its mechanism of action was documented to act as a reversible inhibitor of acetylcholinesterase (Hodgson 2012). Carbaryl toxicity was investigated in freshwater fish species as *Poecilia reticulata*, *Oncorhynchus mykiss*, *Catla calta*, *Mystus vittatus*, *M. cava-sius*, *Anabas testutus*, *Danio rerio*, *Cirrhinus mrigala*, *Labeo rohita* and *Channa punctatus* (Boran et al. 2007; Mahboob et al. 2015; Mustafa et al. 2014; Sambasiva Rao et al. 1985; Tilak et al. 1981; von Hellfeld et al. 2020; Zinkl et al. 1987). Comparative carbaryl toxicity with its degradation product

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(1-naphthol) was conducted for five freshwater fish species (Tilak et al. 1981). Carbaryl is among the most detected pesticides in streams and groundwater (Gilliom et al. 2006). It was also found in surface water and groundwater in Southern Malawi at concentrations 0.083–0.254 mg/l and 0.165–0.492 mg/l, respectively (Kanyika-Mbewe et al. 2020). In Tunisia, carbaryl was detected in drinking water and in Medjerda River, Mellègue River, Siliana River and Sidi Salem dam (Latrous El Atrache et al. 2016).

The second pesticide chosen is tebufenpyrad also known as Masai, Pyranica or Shirudo®, a pyrazole insecticide, and acaricide was mostly used for the treatment of citrus fruit against *Tetranychus urticae*, *Panonychus ulmi* and *P. citri* (Tomlin 1997). It was recorded as one of the most toxic pesticides for fish (EFSA 2008). Tebufenpyrad acts like a mitochondrial complex I electron transport inhibitor (Charli et al. 2016). Studies on the occurrence and ecotoxicology of this pesticide for fish are very limited; according to Kapsi et al. (2019), tebufenpyrad was detected in the Louros River (north-western Greece) at concentrations ranging from 0.127 to 0.337 ppm. In Tunisia, it was found in chili pepper at a concentration of 0.186 mg/kg (Toumi et al. 2018).

The third and fourth insecticides selected are two pyrethroids cypermethrin and permethrin; both were reported as toxic to non-target species, mainly to fish (Başer et al. 2003). Cypermethrin and permethrin are both synthetic pyrethroids type II and I, respectively (WHO 1989). Cypermethrin was synthesized in 1974 and commercialized for the first time in 1977; it is mostly used for the control of many pests especially Lepidoptera (WHO 1989). Cypermethrin affects the nervous system by disrupting the transport of sodium ion through the cell membrane (WHO 1989). Its toxicity was reported in various fish species like the Indian major carp *Labeo rohita* (Marigoudar et al. 2013), the common carp *Cyprinus carpio* (Suvetha et al. 2010), guppy fish *Poecilia reticulata* (Salako et al. 2020), the Nile tilapia *Oreochromis niloticus* (Majumder and Kaviraj 2021), the freshwater carp *Catla catla* (Sharma and Jindal 2020) and zebrafish *Danio rerio* (Guo et al. 2021).

Permethrin was synthesized in 1973, and was then classified as “Restricted use pesticides” by the US Environmental Protection Agency (EPA) based on its high toxicity to aquatic organisms (Başer et al. 2003); among the studied fish species, we cite the narrow-clawed crayfish *Astacus leptodactylus* (Günel et al. 2021).

The European sea bass *Dicentrarchus labrax* was chosen for many reasons; it has been found effective for the biomonitoring of the marine ecosystem, a suitable model of laboratory bioassays (Hernández-Moreno et al. 2011; Mhadhbi et al. 2020; Varo et al. 2003), a highly consumed species with a broad range of distribution (FAO 2021). Indeed, the global farming production of this species has increased from 60,000 t in 2003 to 235,537 t in 2018, with

Mediterranean countries being the most important producer (FAO 2021). Besides this, *D. labrax* is relatively easy in handling, and is characterized by high relative fecundity ranging from 350,000 to 542,000 eggs/kg (Forniés et al. 2001). *D. labrax* is a euryhaline species with an interesting cycle of life, including an embryonic and a larval stage whose development occurs in the marine environment; juvenile development may occur in marine and/or freshwater environments (Jennings and Pawson 1992). This capacity to tolerate strong variation in salinity allowed for this fish species the colonization of various habitats, which make it an interesting model for ecotoxicological studies in both marine and freshwater ecosystems. As stated above, data reporting the toxicity of carbaryl, cypermethrin and permethrin are available for some fish species; however, there is no report of their effects on *Dicentrarchus labrax*. Also, investigations of tebufenpyrad toxicity on fish are very rare.

This study focuses on the assessment of carbaryl, tebufenpyrad, cypermethrin and permethrin toxicity on the European sea bass (*Dicentrarchus labrax*) early life stages. The lethal and sublethal effects of the selected pesticides were determined using six increasing concentrations. The results provided here could serve as useful baseline information, in the future, for the monitoring of contaminant in the aquatic system using *Dicentrarchus labrax* early life stages as a model.

Materials and methods

Sampling and experimental conditions

Eggs of the European sea bass *Dicentrarchus labrax* (diameter ~ 1.5 mm) were obtained from a broodstock maintained in captivity at the company SUD AQUACULTURE TUNISIE (SAT), located in the Djorf region of the governorate of Medenine, Southern Tunisia. An egg collector was used for the collection of the floating one, then they were transported to the laboratory in plastic bags put in portable ice boxes. At the laboratory, embryos were acclimated to laboratory conditions during 24 h; the temperature was maintained around 14.0 ± 1.0 °C, pH = 7.80 ± 0.10 , and a photoperiod of 12 h light: 12 h dark, and the salinity was measured to around 34‰.

Carbaryl (purity $\geq 98\%$), tebufenpyrad (purity $\geq 98\%$), cypermethrin (purity $\geq 98\%$) and permethrin (purity $\geq 98\%$) were purchased from Sigma-Aldrich, Co. (St. Louis, MO, USA). The physicochemical properties are summarized in Table S1. Stock solutions were made in 1% dimethyl sulfoxide (DMSO, Sigma-Aldrich), and were maintained at 4 °C until use. Effects of the selected pesticides were tested using six increasing concentrations chosen based on results from previous studies: 0.5, 1, 2, 4, 8 and 16 ppm for carbaryl

(Toumi et al. 2016); 10, 20, 40, 80, 120 and 160 ppm for tebufenpyrad (Kapsi et al. 2019); and 12.5, 25, 50, 100, 200 and 400 ppm for cypermethrin and permethrin (Ali et al. 2018; Günal et al. 2021).

Acute toxicity test

The acute toxicity test was realized according to OECD Guidelines for the testing of chemicals No. 203 (OECD 2019) and No. 236 (OCDE 2013). The bioassays were conducted on floating fertilized eggs being examined using a dissection microscope. The non-floating eggs and those exhibiting an abnormal development at the stages blastula were eliminated. Two hundred eggs were considered of each concentration, and 50 eggs ($\times 4$ replicates) were carefully distributed into 500-ml glass beakers filled with filtered seawater. Two controls were considered, one without pesticide and the other with DMSO added in volumes equal to that of the tested pesticide. The percentage of mortality was calculated as the following: number of dead embryos / total number for each concentration * 100.

In the control groups, the percentages of mortality were all below 10%, which is required for the validity of the test. Food and oxygen were not provided during the test period. Pesticides' effects on *D. labrax* embryos and larvae were monitored daily during 6 days; the first 2 days correspond to the embryo stage after which the larval stage occurs, which corresponds to 96 h of exposure to the tested compounds. The number of dead eggs and embryos was noted. The survival and malformations were daily recorded after hatching; successful hatching was marked by the rupture of the egg membrane. Mortality was recognized by a coagulation of the embryos, a missing of heartbeat, a failure in the development of somites and a non-detached tail.

Teratogenic assessment

Examinations were made every 24 h immediately after hatching; malformed embryo was considered when a coagulated fertilized egg, an abnormal egg shape and an unhatched egg were observed. Larval malformations were reported when a larva with yolk sac and pericardial oedemata is observed; a lordosis, kyphosis, scoliosis, absence of eye, cranial deformity and general atrophy of the body were detected (von Hellfeld et al. 2020). The observations were made under a binocular dissection microscope using a thick slide with a concave chamber filled with seawater (Table 1).

Statistical analyses

Lethal concentrations (LC_{10-50}) and their 95% confidence intervals were calculated according to the Probit method (Finney 1971) using SPSS software, version 19. The no

observed effect concentration (NOEC) and the lowest observed effect concentration (LOEC) were calculated through ANOVA with Dunnett's post hoc test using the SPSS application, version 19.0. Data that did not meet the assumptions for ANOVA were tested using non-parametric tests (Kruskal–Wallis and Mann–Whitney *U*). Significant differences were considered for ($p < 0.05$).

Results

Acute toxicity

Results from the exposure of *Dicentrarchus labrax* embryos and larvae to six increasing concentrations of carbaryl, tebufenpyrad, cypermethrin and permethrin showed an increase lethally associated with the received dose of pesticides; this was expressed by a clear dose–effect relationship (Fig. 1). In embryos and larvae, mortality begins to occur after 24 h of exposure to 2 ppm of carbaryl, 10 ppm of tebufenpyrad, 25 ppm of cypermethrin and 25 ppm of permethrin (Table 2). Embryo and larval mortality was time and dose dependent (Table 2). The percentage of hatching success decreased with the increase in pesticide concentration, and a significant difference was detected for a concentration greater than 8 ppm carbaryl, 20 ppm tebufenpyrad, 100 ppm cypermethrin and 100 ppm permethrin (Table 2). LC_{50} were calculated as 13.88, 43.96, 92 and 142 ppm and 9.27, 25.67, 48.4 and 72.7 ppm for embryos and larvae treated with carbaryl, tebufenpyrad, cypermethrin and permethrin, respectively (Table 3). Also, NOEC and LOEC were higher in embryos than in larvae (Table 3). *D. labrax* larvae were found more sensitive than embryos to chemicals. Carbaryl was the most toxic pesticide to *D. labrax* early life stages followed by tebufenpyrad, cypermethrin and permethrin.

Teratogenic effects

The assessment of the effect of carbaryl, tebufenpyrad, cypermethrin and permethrin during the embryonic development of the European sea bass revealed the occurrence of several malformations (Figs. 2 and 3). Coagulation, unhatched egg, pericardial oedemata, yolk oedemata, lordosis, kyphosis and scoliosis were induced with carbaryl and tebufenpyrad (Fig. 4). One specimen was found without eye following the exposure to 16 ppm carbaryl; also for the same concentration, a cranial deformity was detected (Figs. 3 and 4). A no spherical egg was observed with 120 and 160 ppm (Figs. 2 and 4). A delay in egg hatching was noted in the groups exposed to carbaryl and tebufenpyrad; the longer period (~ 5 h) was recorded in specimens exposed to 8 and 16 ppm carbaryl against (~ 3 h) in those exposed to 120 and 160 ppm tebufenpyrad.

Table 1 Sublethal concentration of carbaryl, tebufenpyrad, cypermethrin, and permethrin reported in the literature for several fish species

Chemical	Fish species	Life stage	Time of exposure	LC ₅₀	References	
Carbaryl	<i>Poecilia reticulata</i>	Juvenile	96 h	1.38 mg/L	Boran et al. (2007)	
	<i>Oncorhynchus mykiss</i>	Juvenile	48 h	0.79 mg/l	Zinkl et al. (1987)	
		Adult	96 h	0.52 mg/L		
				24 h	1.41 mg/L	
	<i>Catla catla</i>	Larvae	96 h	6.4 mg/L	Tilak et al. (1981)	
	<i>Mystus vittatus</i>	Adult	96 h	2.4 mg/L		
	<i>M. cavasius</i>	Adult	96 h	4.6 mg/L		
	<i>Anabas testutus</i>	Adult	96 h	4.7 mg/L		
	<i>Danio rerio</i>	Embryos	96 h	12.2 mg/L	von Hellfeld et al. (2020)	
	<i>Cirrhinus mrigala</i>	fingerling	96 h	4.75 mg/L	Mahboob et al. (2015)	
	<i>Labeo rohita</i>	fingerling	96 h	8.24 mg/L	Mustafa et al. (2014)	
	<i>Channa punctatus</i>	-	48 h	8.71 mg/L	Sambasiva Rao et al. (1985)	
	<i>Dicentrarchus labrax</i>	Embryos	48 h	13.88 ppm	Present study	
Larvae		96 h	9.27 ppm			
Tebufenpyrad	<i>O. mykiss</i>	-	-	23–30 ppm	EFSA (2008)	
	<i>Dicentrarchus labrax</i>	Embryos	48 h	51 ppm	Present study	
		Larvae	96 h	25.67 ppm		
Cypermethrin	<i>Cnesterodon decemmaculatus</i>	Juvenile	96 h	0.43 ppm	Carriquiriborde et al. (2007)	
	<i>Heteropneustes fossilis</i>	Adult	96 h	0.67–1.27 ppm	Saha and Kaviraj (2003)	
	<i>Poecilia reticulata</i>	Adult	24 h	29.12 ppm	Salako et al. (2020)	
				72 h	27.16 ppm	
				96 h	27.07 ppm	
	<i>Mystus cavasius</i>	Embryos	48 h	8.25 ppm	Ali et al. (2018)	
		Larvae	72 h	6.12 ppm		
	<i>Oreochromis niloticus</i>	Adult	72 h	5.05 ppm	Majumder and Kaviraj (2021)	
	<i>Tilapia mossabica</i>	Adult	96 h	35.6 ppm	Prashanth et al. (2011)	
	<i>Heterobranchus bidorsalis</i>	fingerling	96 h	0.036 mg/L	Olufayo and Alade (2012)	
	<i>Cyprinus carpio</i>	Adult	96 h	2.6 ppm	Saha and Kaviraj (2008)	
	<i>Labeo rohita</i>	Early life stage	-	8.43 ppm	Dawar et al. (2016)	
	<i>Dicentrarchus labrax</i>	Embryos	48 h	92 ppm	Present study	
Larvae		96 h	48.4 ppm			
Permethrin	<i>Channa striatus</i>	-	24 h	2 ppm	Singh and Agarwal (1994)	
	<i>Oryzias latipes</i>	Juvenile	48 h	0.024 mg/L	Rice et al. (1997)	
					0.011 mg/L	
	<i>Sciaenops ocellatus</i>	Juvenile	96 h	23 ppm	Parent et al. (2011)	
	<i>Fundulus heteroclitus</i>	Adult	96 h	8 ppm		
	<i>Poecilia reticulata</i>	Adult	48 h	245.73 ppm	Başer et al. (2003)	
	<i>Dicentrarchus labrax</i>	Embryos	48 h	142 ppm	Present study	
		Larvae	96 h	72.7 ppm		

A positive correlation between the incidence of malformation and the concentration of the tested pesticide was detected; r is > 0.92 for all the tested pesticides. In fact, the highest rates of malformation were observed with 16 ppm carbaryl (34.5%), 160 ppm tebufenpyrad (28%), 400 ppm cypermethrin (17.5%) and 400 ppm permethrin (16%) (Table 2). No malformations were observed in *D. labrax* embryos and larvae exposed to cypermethrin and permethrin concentrations ≤ 25 ppm (Fig. 3). Malformations observed in embryos were mainly coagulation (Fig. 2 and 4). In larvae, the most observed malformations were

yolk oedemata, pericardial oedemata, lordosis and kyphosis (Figs. 3 and 4).

Discussion

Decades of research on ecotoxicology have allowed the assessment of insecticide effects on non-target organisms in both terrestrial and aquatic environments (Antwi and Reddy 2015; Hill 1985). Toxicity of several insecticides like deltamethrin, carbofuran, fenitrothion, chlorpyrifos, dichlorvos,

Fig. 1 Dose–effect curves of lethal toxicity caused by carbaryl (A), tebufenpyrad (B), cypermethrin (C) and permethrin (D) on *Dicentrarchus labrax* embryos (open circles) 48 h and larvae (filled squares) 96 h (mean value ± standard error)

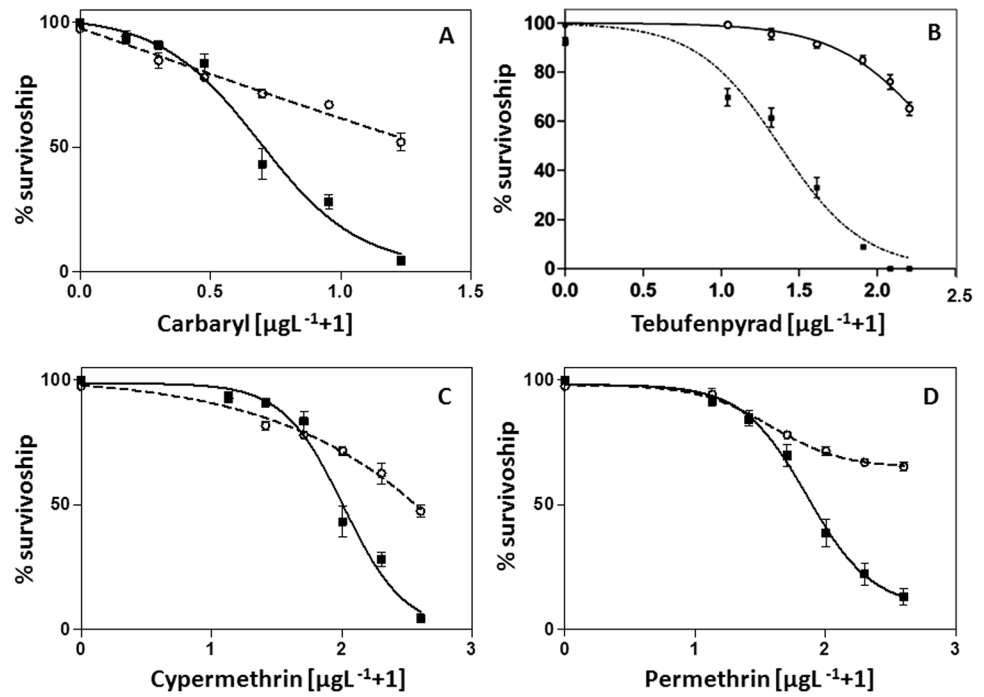


Table 2 Mortality, survival and percentage of malformation at embryo and larval stages of *Dicentrarchus labrax* exposed to carbaryl, tebufenpyrad, cypermethrin and permethrin

Chemical	$\mu\text{g L}^{-1}$	Embryos		Larvae				Percentage of malformation
		Number of dead embryos	Hatching success (%)	24 h	48 h	72 h	96 h	
Carbaryl	0.5	-	100	-	-	-	12	1
	1	-	100	-	4	8	25	1.5
	2	17	91.5	12	10	13	39	10.5
	4	27	86.5	18	15	18	42	19
	8	88	56*	19	27	20	46	26
	16	118	41*	12	21	19	30	34.5
Tebufenpyrad	10	-	100	-	5	6	15	0.5
	20	11	94.5*	8	9	20	40	1
	40	32	84*	10	18	22	47	6
	80	56	72*	15	19	36	52	12.5
	120	78	61*	16	22	20	35	23
	160	103	48.5*	19	20	19	39	28
Cypermethrin	12.5	-	100	-	-	1	2	0
	25	16	92	5	8	7	12	0
	50	17	91.5	6	13	16	26	1
	100	67	66.5*	7	15	18	30	5
	200	90	55*	8	18	22	33	7.5
	400	117	41.5	8	21	23	30	17.5
Permethrin	12.5	-	100	-	-	4	4	0
	25	15	92.5	4	4	8	8	0
	50	21	89.5	-	-	8	12	0.5
	100	33	83.5*	8	8	10	12	2.5
	200	90	55*	10	15	12	16	8
	400	101	49.5	15	19	17	29	16
Control without pesticide	-	4	98	0	0	4	4	0

Table 3 Toxicity thresholds (NOEC, LOEC and LC₁₀), and sublethal concentrations (LC₅₀) for carbaryl, tebufenpyrad, cypermethrin and permethrin in *Dicentrarchus labrax* early life stages

	Chemical	NOEC	LOEC	LC ₁₀	LC ₅₀
Embryos (48 h)	Carbaryl	1	2	5.68 (3.34–9.21)	13.88 (8.31–20.32)
	Tebufenpyrad	10	20	23.2 (22.78)	43.96 (40.25–46.12)
	Cypermethrin	-	12.5	23.45 (12.94–29.7)	92 (77.1–121.9)
	Permethrin	12.5	25	33 (31.77–36.96)	142 (118.12–160.4)
Larval (96 h)	Carbaryl	0.5	1	3.48 (1.89–6.81)	9.27 (6.4–13.19)
	Tebufenpyrad	-	10	10.3 (10.11–11.95)	25.67 (23.74–27.13)
	Cypermethrin	-	12.5	13.29 (9.23–19.77)	48.4(32.38–60.27)
	Permethrin	12.5	25	27.31 (26.22–29.47)	72.7 (13.8–21.19)

Values are in µg/l, with 95 % confidence intervals in parentheses

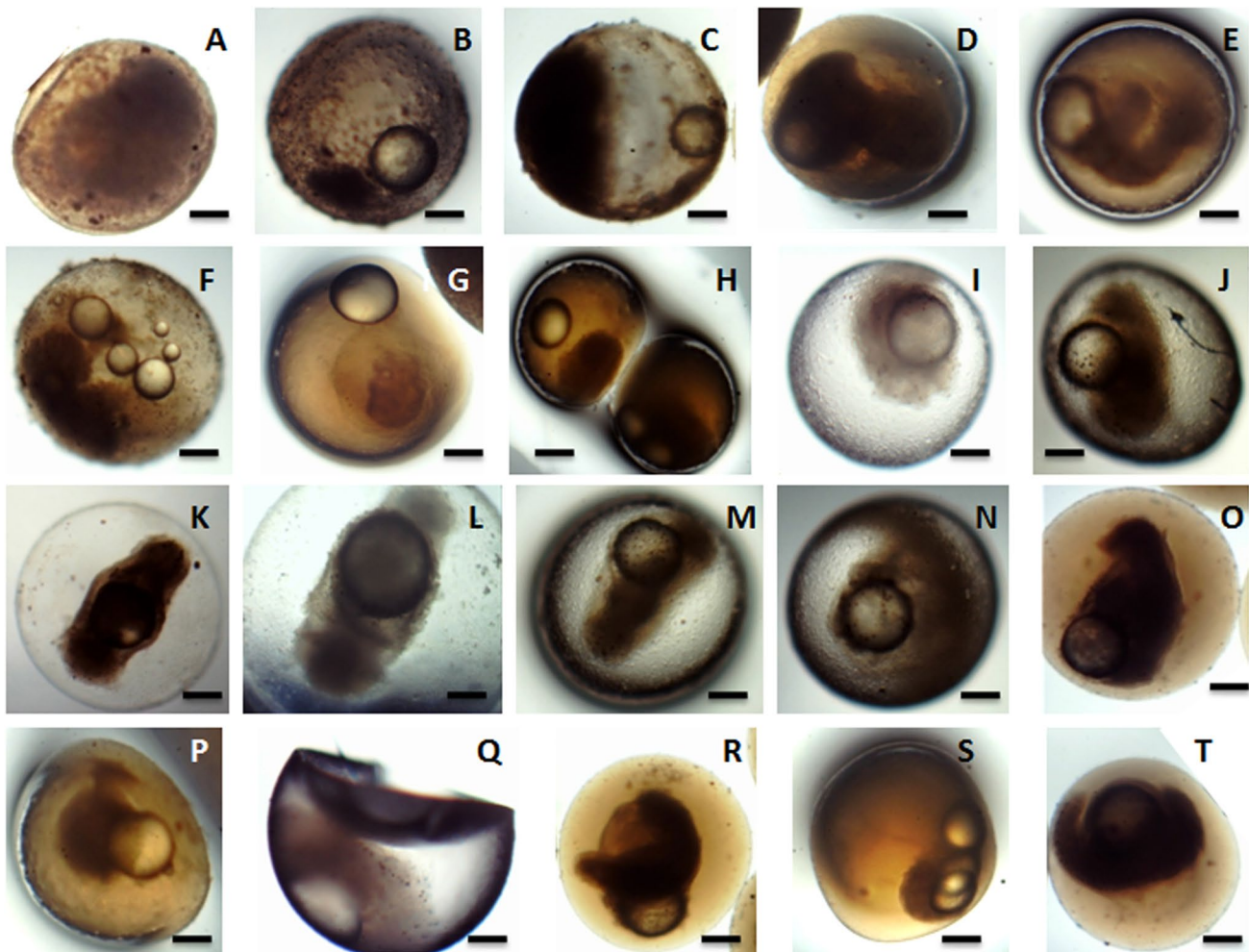


Fig. 2 Morphological abnormalities observed in *Dicentrarchus labrax* embryos exposed to carbaryl, tebufenpyrad, cypermethrin and permethrin. Coagulation caused by carbaryl (a–o), no spherical egg (p–q) and un-hatched egg (r, t). Malformation observed

with 8 ppm carbaryl (a–r), 16 ppm carbaryl (d), 80 ppm tebufenpyrad (b), 120 ppm tebufenpyrad (c, l, p), 160 ppm tebufenpyrad (s, q), 400 ppm cypermethrin (e–g, t), 100 ppm permethrin (h–k) and 400 ppm permethrin (m–o). Scale bars 300 µm

fipronil and trichlorfon was investigated on the European sea bass *Dicentrarchus labrax* (Almeida et al. 2010; Banni et al. 2011; Dallarés et al. 2020; Hernández-Moreno et al. 2011; Varo et al. 2003). However, most of the conducted studies

were focused on the assessment of pesticide toxicity on adult sea bass. Non-target organisms were frequently subjected to pesticide pollution through two ways, directly following spray deposition and drift and indirectly during surface

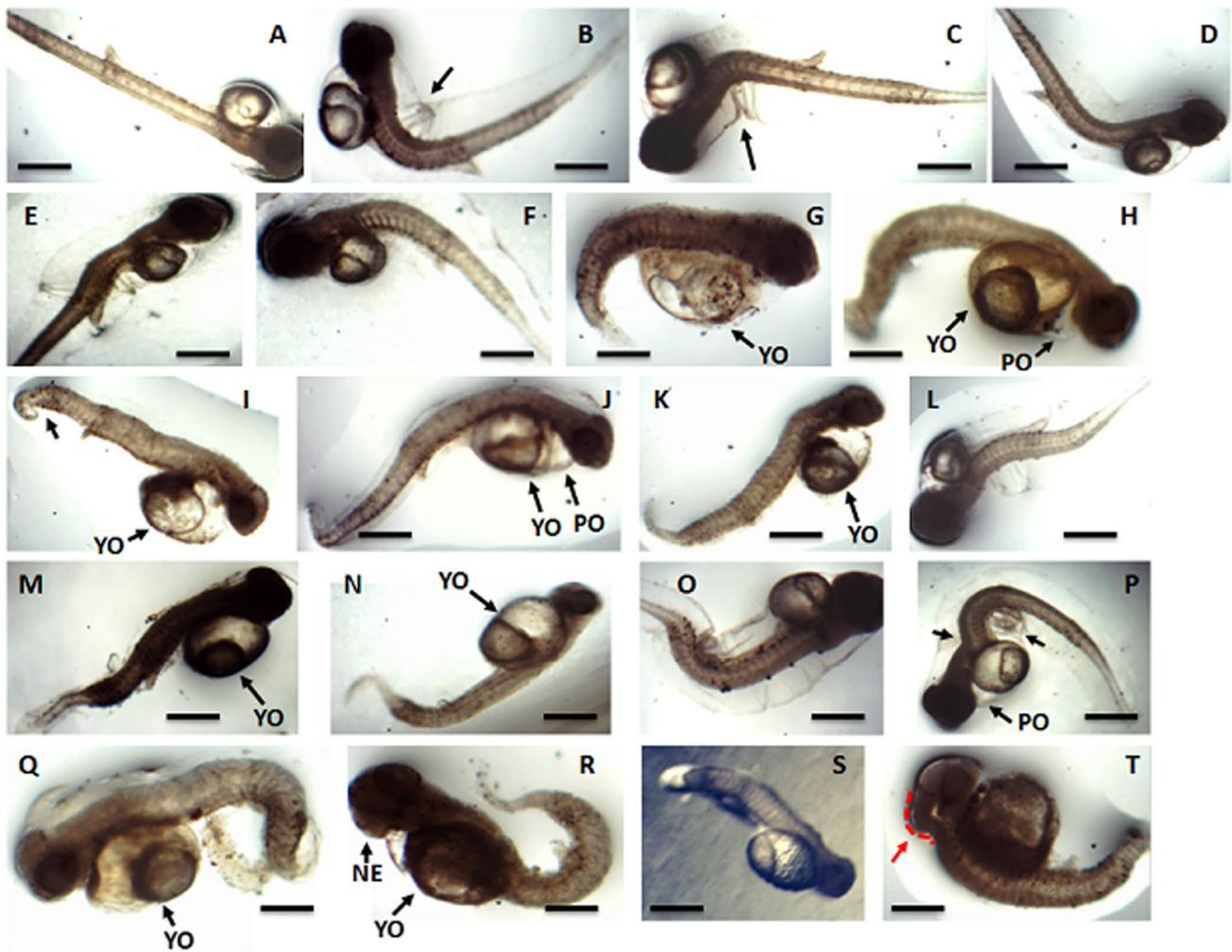


Fig. 3 Morphological abnormalities reported in *Dicentrarchus labrax* larvae exposed to carbaryl, tebufenpyrad, cypermethrin and permethrin. Normal larvae (a), lordosis (b–d), kyphosis (e–h), scoliosis (i–r), yolk oedemata (g, i, k, m, n, q, r), pericardial oedemata (p), pericardial and yolk oedemata (h, j), no eye (r), body atrophy (s) and

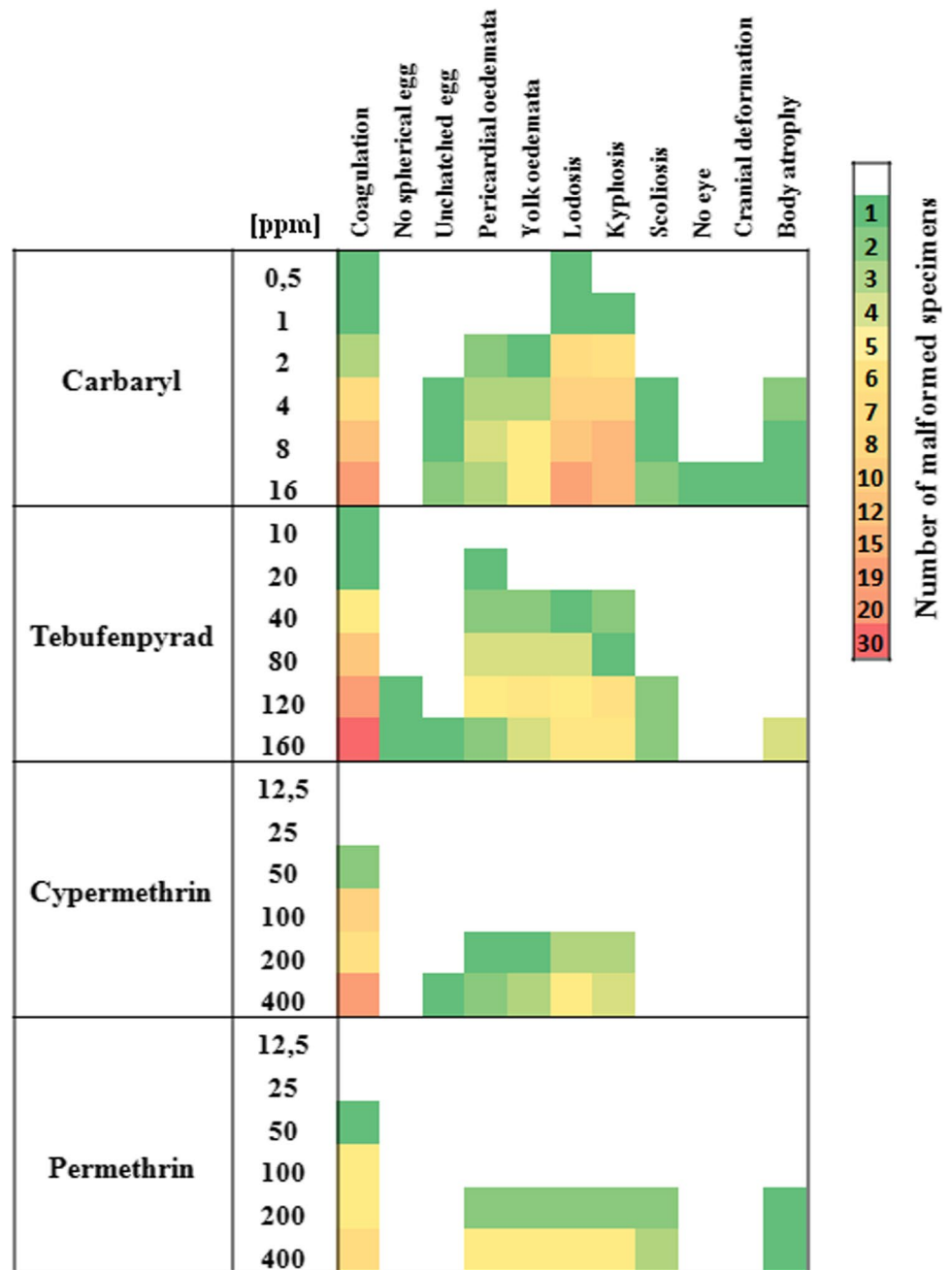
cranial deformity (t). YO yolk oedemata, PO pericardial oedemata, NE no eye. Malformation observed with 4 ppm carbaryl (i), 16 ppm carbaryl (b, d, h, j, r, t), 160 ppm tebufenpyrad (c, g, m, p, q, s), 400 ppm cypermethrin (e) and 400 ppm permethrin (f, k, l, n, o). Scale bars 500 μ m

runoff and erosion (Antwi and Reddy 2015). Pesticides were also used in aquaculture for the control of parasite in farmed *Dicentrarchus labrax*; trichlorfon and teflubenzuron were used for the treatment of *Diplectanum aequans* infestations (Tokşen et al. 2010, 2013). Moreover, deltamethrin is used for the treatment of *Ceratomyxa oestroides* infestations (Çolak et al. 2019).

Data from the studies of pesticide toxicity on fish species showed a difference in their effects among species and the same time, even in the same species between live stages (Hernández-Moreno et al. 2011). The results for this study showed that the four tested insecticides (carbaryl, tebufenpyrad, cypermethrin and permethrin) exhibited different effects on *D. labrax* early life stages. Carbaryl was found more toxic than tebufenpyrad, cypermethrin and permethrin. LC_{50} calculated for *D. labrax* exposed

to carbaryl were 13.88 ppm, 48 h, and 9.27 ppm, 96 h. Those values were lower than those of LC_{50} reported for other fish species (Table 1). Sublethal concentrations from the literature were in milligrams per litre level and ranged between 0.52 mg/l for *O. mykiss* (Boran et al. 2007) and 12.2 mg/l for zebrafish embryos (*Danio rerio*) (von Hellfeld et al. 2020). Carbaryl was found to induce several morphological malformations in zebrafish embryo at concentration < 6.6 mg/l (von Hellfeld et al. 2020). Malformations were induced in *D. labrax* early life stages with 0.5 ppm carbaryl, and the occurrence of malformed embryos and larvae increased with the increase in the concentration. The results of the bioassay conducted on *D. labrax* early life stages have proven the toxicity of tebufenpyrad; values of LC_{50} estimated for embryos 43.96 ppm and larvae 25.67 ppm were approximately similar to those

Fig. 4 Heatmap summarizing the occurrence of abnormalities in *Dicentrarchus labrax* embryos and larvae exposed to carbaryl, tebufenpyrad, cypermethrin and permethrin



calculated for *O. mykiss* 23 and 30 ppm (EFSA 2008), while they were higher than the LC₅₀ value, 10.98 ppm 48 h, reported for *Daphnia pulex* (Asselman et al. 2014). Tebufenpyrad teratogenicity was not reported previously; the results from this study revealed the occurrence of malformed embryos and larvae with 10 and 20 ppm, respectively. Teratogenic effects of pyrazole carboxamides as bixafen and isopyrazam were demonstrated on *D. rerio* and *Xenopus tropicalis* during embryonic development (Li et al. 2016; Xiao et al. 2021).

Despite the increased number of studies on the assessment of cypermethrin toxicity, data about its toxicity

on the European sea bass *D. labrax* especially early life stages lacked. Cypermethrin was reported as very toxic for fish based on results conducted on laboratory, with 96 h LC_{50s} being generally within the range of 0.4 and 2.8 ppm (WHO 1989). Sublethal concentrations of 92 ppm 48 h and 48.4 ppm 96 h measured for *D. labrax* early life stages were higher than values measured for *Labeo rohita*, *Mystus cavasius* and *Cnesterodon decemmaculatus* early life stages of 8.43, 6.12 and 0.48 ppm, respectively (Ali et al. 2018; Carriquirborde et al. 2007; Dawar et al. 2016). Malformation was observed in *D. labrax* embryos and larvae exposed to cypermethrin for concentrations ≥ 50 ppm, while it was reported

to induce malformations in *Mystus cavasius* embryos and larvae with 16 and 32 ppm (Ali et al. 2018).

Concerning permethrin, it was classified as toxic to fish since LC₅₀ values were in micrograms per litre level (Başer et al. 2003). Indeed, LC₅₀ ranged between 2 ppm, 24 h in *Channa striatus* and > 75 mg/l, 48 h in *Pimephales promelas* (Milam et al. 2000, Singh & Agarwal 1994). Sublethal concentrations measured for *D. labrax* were 142 ppm, 48 h and 72.7 ppm, 96 h; those values were higher compared to the LC₅₀ estimated for other Actinopterygii (Başer et al., 2003 and reference therein). The teratogenic effect of permethrin was reported for *Danio rerio* (Dach et al. 2019). Also, the results for this study showed that permethrin has teratogenic effects on *D. labrax* early life stages; the majority of the malformed specimens were detected for 100 and 200 ppm.

Dicentrarchus labrax larvae were found more sensitive than embryos; Mhadhbi et al. (2012) reported similar results in *Psetta maxima* exposed to the pesticides (alachlor, atrazine, diuron). The low sensitivity of fish embryos was attributed to the protective role of the external envelope, chorion (Groot and Alderdice 1985). However, it seems that pesticides affected the structure of this envelope by blocking the canal pore, which inhibits the egg permeability (Groot and Alderdice 1985). This could explain the increased rate of coagulated *D. labrax* embryos. In addition, it was found that pesticides induce a delay and even an inhibition in embryo hatching as a consequence to a disturbance in the morphology and function of the hatching gland cells; therefore, the secretion of the hatching enzyme is reduced Xiao et al. (2021).

Conclusion

Carbaryl, tebufenpyrad, cypermethrin and permethrin were found toxic to the European sea bass, *Dicentrarchus labrax*, early life stages. The toxicity followed the order carbaryl > tebufenpyrad > cypermethrin > permethrin. Embryo and larval mortality was time and dose dependent. Larvae were found more sensitive than embryos to the studied pesticides. Teratogenic effects of the studied pesticides were recognized of *D. labrax* early life stages. Results from the current survey provided baseline information that could be used in the future for the risk assessment of pesticides in the aquatic system using *Dicentrarchus labrax* early life stages as a model.

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Data availability All data are mentioned in the body of manuscript, tables, and figures.

Declarations

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