



Pollution resistance assessment of plants around chromite mine based on anticipated performance index, dust capturing capacity and metal accumulation index

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Abstract

Plant species sustaining under a polluted environment for a long time are considered as potentially resistant species. Those plant species can be considered as an eco-sustainable tool used to bio-monitor and mitigate pollution. This study was carried out on a total of ten commonly available plant species to assess their anticipated performance index (API), dust capturing capacity (DCC), and metal accumulation index (MAI) in chromite mine and control areas. According to the anticipated performance index (API), *Macaranga peltata* (Roxb.) Müll.Arg., *Holarrhena pubescens* Wall. ex G.Don and *Ficus hispida* Roxb. ex Wall. are highly tolerant species while *Terminalia arjuna* (Roxb. ex DC.) Wight & Arn. and *Trema orientalis* (L.) Blume are intermediate tolerant species. *F. hispida* was also shown to have the highest dust capturing capacity (5.94 ± 0.43 mg/cm²) whereas that of *Woodfordia fruticosa* Kurz (1.03 ± 0.11 mg/cm²) was found to be lowest. The metal accumulation index ranged from 17.29 to 4.5 and 6.38 to 1.94 at the mine and control areas, respectively. Two-way ANOVA analysis revealed area-wise significant differences between biochemical and physiological parameters. Also, results showed that the pollution level and heavy metal affected different biochemical and physiological parameters of plant species at the mining area. The plant species with the highest API, DCC, and MAI value could be recommended for greenbelt development in different polluted areas.

Keywords Bio-monitor · Anticipated performance index · Dust capturing capacity · Metal accumulation index · Chromite mine · Greenbelt development

Introduction

The economic growth and development of a nation is dependent on its mining sector to a great extent which is associated with the exploitation of the mineral resources (Lèbre et al. 2017), whereas mining plays a significant role in economic growth and at the same time its adverse impact on the environment. Worldwide mining activity is one of the

serious contributors to the environmental pollution and considered one of the significant sources of air, soil and water pollution (Oluwoye et al. 2017; Golui et al. 2021; Sahu and Basti 2020). From open-cast mining, a massive amount of gaseous pollutants, toxic substances, fine ore particles and dust are released to the environment (Das et al. 2020). These released pollutants mix with the atmosphere and are transported at a long distance through the air from the mine area and lead to pollution in the surrounding undisturbed area (Ni et al. 2018; Khazini et al. 2021).

Green vegetation around the mining area can act as an eco-friendly and cost-effective tool to reduce pollution. In a polluted environment, green plants play a crucial function in improving the environmental quality by accumulating toxic pollutants (Gheorghe and Ion 2011; Kaur and Nagpal 2017). They have great potential to the absorption, adsorption and accumulation of pollutants from both soil and air (Manara 2012; Remon et al. 2013). They transport toxic substances from soil and air to biotic environments. Green plants play

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a role as pollution sink as they provide a broad surface area for absorption and accumulation of pollutants; hence, plantation of highly tolerant plant species can help in effective green belt development to clean up polluted areas, further improving the environmental quality (Remon et al. 2013; Selmi et al. 2016; Karmakar and Padhy 2019;). Selection of suitable plants for greenbelt development differs from region to region.

The air pollution tolerance index (APTI) and anticipated performance index (API) are considered best and reliable indices to select appropriate tolerant and sensitive plant species. The highly tolerant plant species are chosen to perform against the environmental pollution, and the sensitive species might be used as a bioindicator (Rai 2016; Javanmard et al. 2020; Roy et al. 2020; Molnár et al. 2020). Four biochemical parameters, namely total chlorophyll content (TCC), leaf extract pH (pH), ascorbic acid (AA) and relative water content (RWC), were used to develop the APTI (Singh and Rao, 1991). API is an upgradation of APTI, which is a more appropriate index for the selection of tolerant and sensitive species in a particular region. API is developed with the combination of the APTI value of each individual species along with some morphological (plant habit, canopy structure, types of plant, laminar structure) and socio-economic characters (Mondal et al. 2011). Dust, particulate matter and heavy metal are the most prevalent environmental pollutants, causing critical problems to all living organisms (Xiu et al. 2020; Kong et al. 2021). Plants provide the most efficient and natural ways to remediate pollution by capturing dust and accumulating particulate matter. In order to combat such pollutants, dust capturing capacity (DCC) and metal accumulation index (MAI) are also considered to select the suitable plant species.

The Sukinda chromite mine area is one of the most populated areas in the world, having a number of open-cast chromite mines. This mining area holds 183 million tons of raw chromite reserves, which is a total of 97% of total reserves of the country and approximately 3.8 million tons of chromite produced per year. Due to various mining activities, a massive amount of ore particles and dust is produced and get blown away through air to surrounding areas, subsequently causing chromium (Cr) contamination by atmospheric deposition (Das et al. 2020). According to a report of Black Smith Institute Report (2007), extensive pollution of the area due to excessive chromite ore mining has made it the fourth most polluted place in the world.

Through this study, we provide an integrated approach to the selection of both pollution-tolerant and -sensitive plant species. To know the physiological and biochemical tolerance levels of plants, we studied the air pollution tolerance index (APTI) and anticipated performance index (API). Heavy metal accumulation and dust capturing capacities of a total of ten plants were also examined. This study is

confined only on commonly found native plant species in this area. So, our study will contribute convenient knowledge/information to find out the suitability of plant species for phytoremediation of toxic heavy metals in the vicinity of the chromite mine area as well as different other polluted areas to improve the levels of soil and air pollution.

Materials and methods

Study area

The present study was done at two areas, namely Sukinda chromite mine valley and Tomka Forest, located in Jajpur district, Odisha, India. Sukinda chromite mine valley is the largest chromite producer in India; almost 99% of the country's chromium is produced from this valley. Stretching between 21° 00' N to 21° 05' N latitude and 85° 44' E to 85° 53' E longitude occupies an area of 200 km² which holds an estimated 183 million tons of raw chromium deposits (Nayak and Kale 2020) and consists of an extensive plateau in the interior with a foreground of a wide coastal plain underlain by Precambrian rock. Currently, 20 open-cast and 2 underground mines are functional in the valley producing 160 million tons of overburden. It also releases 11.73 t of hexavalent chromium (Cr(VI)) to the environment every year, making it the fourth-worst polluted place in the world. (Dhal et al. 2011; Das et al. 2020). Tomka Forest is situated 21° 6'12" N to 21° 8' 11" N latitude and 85° 49' 12" E to 85°55' 58" E longitude, considered as a control area due to the pollution-free atmosphere, located about 40 km away from the mining area.

Plant species sampling

In total, leaf samples of ten plant species, namely *Ailanthus excelsa* Roxb., *Bixa Orellana* L., *Ficus hispida* Roxb. Ex Wall., *Macaranga peltata* (Roxb.) Müll.Arg., *Woodfordia fruticosa* Kurz, *Trema orientalis* (L.) Blume, *Terminalia arjuna* (Roxb. ex DC.) Wight & Arn., *Holarrhena pubescens* Wall. ex G.Don, *Callicarpa tomentosa* (L.) Murr. and *Combretum roxburghii* Spreng. plant species were collected during winter session 2020. These plant species were considered on the basis of dominance, closest to the mining site, visible dust deposition on foliage and ecological importance of the specific plant species. Three replicates of healthy, mature and fully developed plant leaf samples were collected for each species. The replicate plant species were chosen as those individual species having similar height and breast diameter. After being sampled, leaves were placed into zipper-locked bags and kept in a portable ice box, then transferred to the laboratory. Leaves were collected and transferred very carefully from plant to zipper-locked bag,

for ensuring least disturbance to their surface dust. For further biochemical analysis, leaves were washed with tap water followed by distilled water, and stored in $-20\text{ }^{\circ}\text{C}$.

Analysis of the biochemical parameters of the plants

Relative water content

After taking the fresh weight (FW), leaves were submerged in deionized water for 24 h. Then, turgid weight (TW) was recorded by soaking up the surface water of leaves. Overnight dried leaves in the oven at $80\text{ }^{\circ}\text{C}$ were used for taking dry weight (DW) (Liu and Ding 2008).

$$\text{RWC}(\%) = \left[\frac{\text{FW} - \text{DW}}{\text{TW} - \text{DW}} \right] \times 100$$

Total chlorophyll content

The total chlorophyll content of leaves was calculated following the procedure mentioned by Arnon (1949). One gram of fresh leaves was crushed by using 10 ml of 80% acetone and centrifuged at 5000 rpm for 5 min. After collection, the supernatant volume was made up to 30 ml by 80% acetone and absorbance was taken at 645 and 663 nm by using a spectrophotometer (Analytical UV–Vis 3090 V). The calculation of total chlorophyll content was done using the following formula:

$$T \left(\frac{\text{mg}}{\text{g}} \right) = \frac{(20.2 * A_{645} + 8.02 * A_{663}) * V}{1000 * W(\text{g})}$$

where A_{645} = absorbance at 645 nm, A_{663} = absorbance at 663 nm, V = total volume of extract (ml), and W = weight of leaf in grams.

Ascorbic acid

Ascorbic acid estimation was done by using the modified colorimetric 2,6-dichlorophenol indophenol technique. Fresh leaves (0.5 g) were extracted in 4% oxalic acid and made up to 30 ml volume and centrifuged at 6000 rpm for 10 min. Ten millilitres of 4% oxalic acid was added with 5 ml of the supernatant and titrated against the dye. Titration was done till a pink colour appeared which was present for a few seconds.

$$A \left(\frac{\text{mg}}{\text{g}} \right) = \frac{0.5(\text{mg}) * V_2(\text{ml}) * 30(\text{ml})}{V_1(\text{ml}) * 5(\text{ml}) * \text{weight of sample}(\text{g})}$$

where

- V_1 volume of the dye titrated against the working standard
- V_2 volume of dye titrated against the sample

Leaf extract pH

The pH of the leaves was determined with the help of a pH meter by crushing 1 g of fresh leaves and homogenizing it in 40 ml of deionized water.

Air pollution tolerance index

APTI was calculated following the equation described by Sing and Rao (1991).

$$\text{APTI} = [A(T + P) + R]/10$$

where A = ascorbic acid (mg/g), T = total chlorophyll content (mg/g), P = pH of leaf extract and R = relative water content (%).

Anticipated performance index

Combining some morphological and socio-economic characteristics (plant habit, type of plant, lamina characters, canopy type and economic value) with APTI value, API is estimated. The API score is calculated according to the following formula:

$$\text{API}(\%) = (\text{Total positives}) \times 100/10$$

The maximum positive allotted for a plant species is 16. According to API, score plant species were categorized into eight different groups (e.g. not recommended, very poor, poor, moderate, good, very good, excellent and best).

Dust capturing capacity

Dust capturing capacity was quantified with the help of a Petri dish. A Petri dish was oven dried, and the initial weight (W_1) was weighed. The upper and lower surface dust of a leaf was washed with deionized water and transferred to the Petri dish. Then the Petri dish was completely dried and weighed to record the final weight (W_2). The surface area (A) of the leaf was measured with the help of graph paper. Dust capturing capacity was calculated by the following formula:

$$W = W_2 - W_1/A$$

Analysis of heavy metal concentration in soil

The metal concentration in soil was done following the procedure mentioned by Roy et al. (2020). A sieved soil sample (0.2 g) was digested using HCl and H_2SO_4 in the ratio of 1:3. The digested sample was filtered and made up to 50 ml with 2% HNO_3 . Then the digested solutions were analysed for

six heavy metals by using PerkinElmer ICP-OES (Model: Optima 2100 DV).

Analysis of heavy metal concentration in leaves

The washed leaves were completely dried at 60 °C in a hot air oven. One gram of leaf material was digested using 2 acids, i.e. HClO₄ and HNO₃, in the ratio of 1:2 (Samecka-Cymerman and Kempers 1999). Digested solutions were diluted with double-distilled water and the final volume made up to 50 ml. Then, the digested solutions were analysed for Al, Cr, Fe, Ni, Pb and Zn using PerkinElmer ICP-OES (Model: Optima 2100 DV).

Metal accumulation index

The metal accumulation capability of plant species is calculated by the using the metal accumulation index (MAI) formula (Hu et al. 2014).

$$MAI = \left(\frac{1}{N}\right) \sum_{J=1}^n IJ$$

where N = number of metals and IJ = sub-index of J gained by dividing the metal concentration by its standard deviation. It depends on the collected sample number.

Results and discussions

Ascorbic acid

Ascorbic acid or vitamin C is an important low molecular weight non-enzymatic antioxidant present in plant chloroplasts. This antioxidant plays a crucial function in the light reaction of photosynthesis, regulates various metabolic biosynthesis pathways and protects plants from biotic and abiotic stresses (Singh and Verma 2007). Throughout photosynthetic electron transfer, ascorbic acid reacts and scavenges reactive oxygen species (ROS). Therefore, the biosynthesis and concentration of this antioxidant are directly proportionate to the tolerance level of plants (Smrinoff and Wheeler, 2000; Wang et al. 2018). In the mining area, the highest ascorbic acid content (mg/g in fresh leaves) was found in *H. pubescence* (26.83 ± 1.01) followed by *M. peltata* (24.66 ± 0.99), *T. orientalis* (22.81 ± 0.8) and *F. hispida* (19.63 ± 2.77), and the lowest content was noticed in *C. roxburghii* (8.76 ± 0.95), followed by *C. tomentosa* (9.46 ± 1.93) and *W. fruticosa* (9.86 ± 1.22). In comparison to the control area, a considerable difference was found in ascorbic acid contents in all plant species (Fig. 1). In the control area, the highest amount was found in *M. peltata* (14.28 ± 0.93) followed by *T. arjuna* (12.85 ± 0.57) and *H. pubescence* (11.36 ± 2.22) and the lowest amount was

found in *C. tomentosa* (4.11 ± 0.7) followed by *C. roxburghii* (4.72 ± 1.03) and *A. excelsa* (5.56 ± 0.91).

Ascorbic acid reacts with various ROS (i.e. superoxide radical, singlet oxygen and hydroxyl radical) and protects pigments and nucleic acids. The elevated amount of ascorbic acid increases the tolerance level of plants under pollution environment as well as other stress conditions (like, drought, salt, temperature) (Wang et al. 2018). Hence, plants synthesized with a higher level of ascorbic acid in polluted environments are evaluated as tolerant species. In our study, the ascorbic acid content is higher at the mining area and lower at the control area for all examined plant species. The enhancement of ascorbic acid in all species from the mining area might be due to the elevated production of ROS, through accumulation of toxic metals in the plant body.

Relative water content

The relative water content signifies the balance between plant water uptake and release. The water content maintains turgor pressure and cell wall permeability in the plant body. During pollution conditions, different pollutants enhance cell permeability, which leads to deficiency in water and nutrients, leading to prior senescence of leaves and bark (Achakzai et al. 2017; Safari et al 2019). So, the relative water content of plants is a suitable physiological parameter to measure water condition in the plant body. All the species had a more relative water content in the control area than in the mining area (Fig. 1). The highest relative water content in the mining area was found in *M. peltata* (93.71 ± 2.69) and the lowest in *A. excelsa* (64.81 ± 7.76). In the control area, the relative water content of leaves of the plant species differed from a maximum of 98.22 ± 0.33 in *F. hispida* to a minimum of 69.31 ± 1.37 in *A. excelsa* (Table 1).

The relative water content of plant leaves in the mining area is decreased which might be due to high pollutants and heavy metal contamination; it also specifies the unbalanced physiological condition of the plant. Heavy metal contamination in the plant body reduces the water transport from root to leaves (Barceló and Poschenrieder 1990). Under any stress conditions, a high water content helps to sustain physiological and biochemical balance in a plant body. A higher level of relative water content in plants increases the tolerance strength of plant towards pollution stress (Gupta et al. 2016; Karmakar and Padhy 2019).

Total chlorophyll content

The photosynthetic pigment chlorophyll, found in the chloroplasts of green plants, is an index of productivity and is

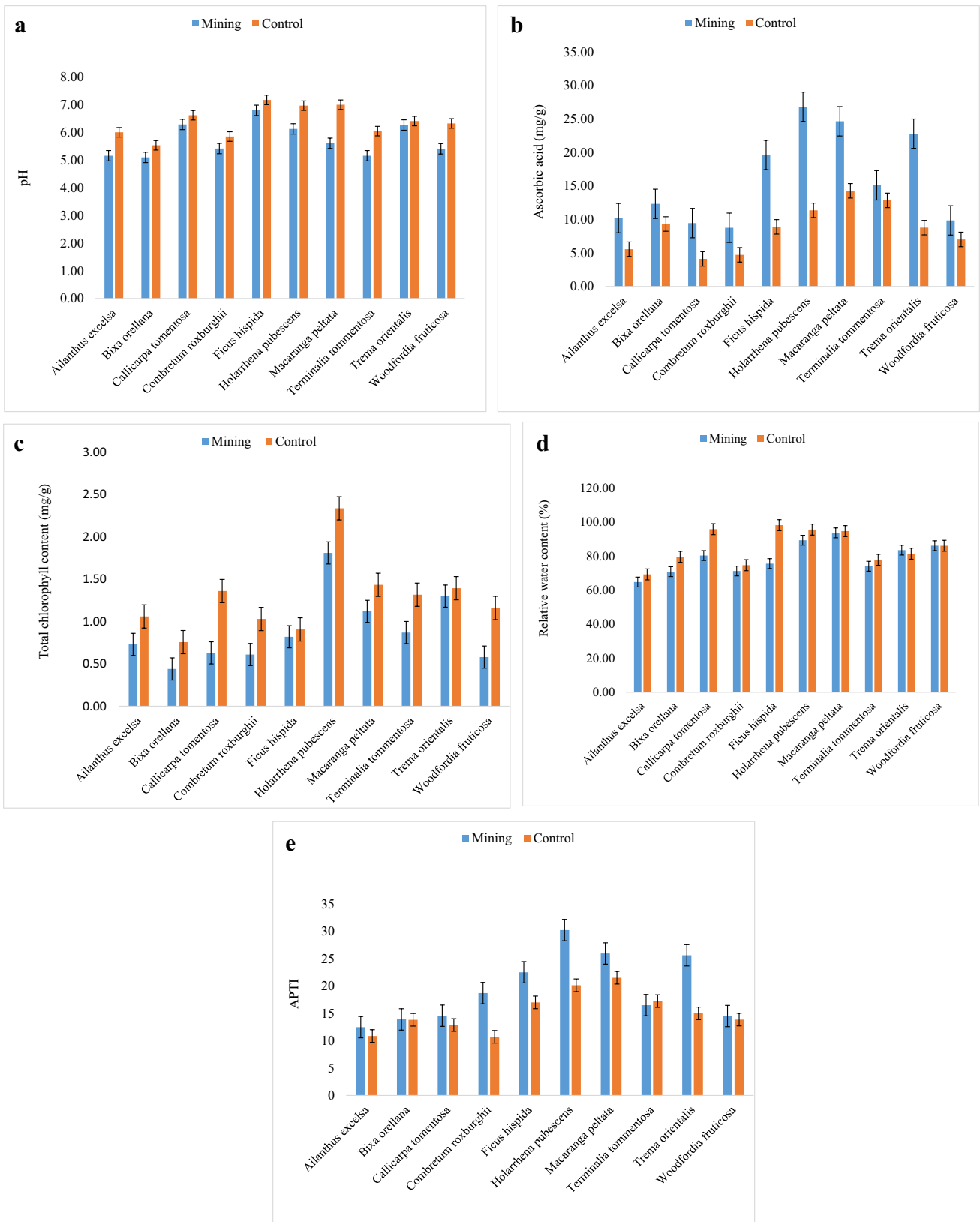


Fig. 1 Area-wise difference in **a** pH, **b** ascorbic acid, **c** total chlorophyll content, **d** relative water content, and **e** APTI of selected plant. Error bars refer to standard error

Table 1 Assessment of APTI of selected plant species from mining and control area

Species	Area	Total chlorophyll (mg/g)	pH	Relative water content (%)	Ascorbic acid (mg/g)	APTI
<i>A. excelsa</i>	Mining	0.73 ± 0.01	5.16 ± 0.04	64.81 ± 7.76	10.2 ± 0.81	12.48
	Control	1.06 ± 0.13	6 ± 0.29	69.31 ± 1.32	5.56 ± 0.91	10.85
<i>B. orellana</i>	Mining	0.44 ± 0.01	5.1 ± 0.06	70.90 ± 1.44	12.33 ± 2.86	13.92
	Control	0.75 ± 0.09	5.53 ± 0.12	79.62 ± 1.37	9.32 ± 0.92	11.82
<i>C. tomentosa</i>	Mining	0.63 ± 0.006	6.29 ± 0.11	80.37 ± 1.64	9.46 ± 1.93	14.58
	Control	1.36 ± 0.19	6.62 ± 0.16	95.87 ± 0.75	4.11 ± 0.7	12.86
<i>C. roxburghii</i>	Mining	0.61 ± 0.009	5.42 ± 0.05	71.3 ± 0.95	8.76 ± 0.95	18.71
	Control	1.03 ± 0.12	5.85 ± 0.24	74.71 ± 1.3	4.72 ± 1.03	10.71
<i>F. hispida</i>	Mining	0.82 ± 0.002	6.8 ± 0.13	75.63 ± 1.15	19.53 ± 2.69	22.52
	Control	0.9 ± 0.07	7.17 ± 0.14	98.22 ± 0.33	8.89 ± 0.78	16.99
<i>H. pubescens</i>	Mining	1.81 ± 0.01	6.13 ± 0.02	89.38 ± 2.63	26.83 ± 1.01	30.24
	Control	2.33 ± 0.22	6.97 ± 0.14	95.63 ± 1.6	11.36 ± 2.22	20.12
<i>M. peltata</i>	Mining	1.12 ± 0.05	5.61 ± 0.03	93.71 ± 2.69	24.66 ± 0.1	25.96
	Control	1.43 ± 0.24	7 ± 0.09	94.72 ± 0.88	14.28 ± 0.93	21.51
<i>T. arjuna</i>	Mining	0.87 ± 0.006	5.16 ± 0.09	74.1 ± 2.13	15.1 ± 1.61	16.51
	Control	1.31 ± 0.2	6.05 ± 0.17	77.87 ± 3.03	12.85 ± 0.57	17.24
<i>T. orientalis</i>	Mining	1.31 ± 0.11	6.27 ± 0.03	83.56 ± 5.06	22.81 ± 0.8	25.64
	Control	1.39 ± 0.19	6.41 ± 0.04	81.47 ± 1.36	8.78 ± 1.07	15
<i>W. fruticosa</i>	Mining	0.59 ± 0.007	5.41 ± 0.02	86.16 ± 2.88	9.86 ± 1.22	14.52
	Control	1.16 ± 0.19	6.32 ± 0.12	86.12 ± 4.74	7.01 ± 0.33	13.85

called a photoreceptor. Chlorophyll plays an important role in plant photosynthesis, so measurement of total chlorophyll content is a significant measure to assess the effect of pollution on the plant. The decreased level of the total chlorophyll content of all species was noticed in the mining area as compared to the control area (Fig. 1). At the mining area, the highest chlorophyll content (mg/g of fresh weight) was found in *H. pubescens* (1.81 ± 0.01) followed by *T. orientalis* (1.3 ± 0.11) and *M. peltata* (1.12 ± 0.05). The lowest content was found in *B. orellana* (0.44 ± 0.01) followed by *W. fruticosa* (0.58 ± 0.007) and *C. roxburghii* (0.61 ± 0.009). At the control area, the highest content was found in *H. pubescens* (2.33 ± 0.22) followed by *M. peltata* (1.43 ± 0.24) and *T. orientalis* (1.39 ± 0.19). The lowest content was found in *B. orellana* (0.75 ± 0.09) followed by *F. hispida* (0.9 ± 0.07) and *A. excelsa* (1.06 ± 0.13). Chlorophyll has a highly organized state that may go through various photochemical reactions like oxidation, reduction and reversible bleaching in different pollutant conditions (Karmakar et al. 2021). The decreased level of chlorophyll at the mining area might be due to dust deposition on the leaf surface and heavy metal accumulation. Protochlorophyllide is the precursor of chlorophyll; during chlorophyll biosynthesis, this precursor is reduced by NADPH to chlorophyllide in the presence of light, but due to the adherence of dust on the leaf surface hindering the pathway of light, thus interfering in the process of chlorophyll formation (Roy et al.

2020). After accumulation of heavy metals, Mg^{++} ion of chlorophyll is replaced by another metal ion and breakdown into phaeophytin (Karakoti et al 2014). High pollution stress reduces the total chlorophyll content at the polluted site. But some plant species produce tolerance mechanisms for detoxification of metal toxicity. Production of a high level of phytochelatin, metallothioneins, organic acids and thiol-reactive peptides which are bound with toxic metals and compartmentalized into vacuoles is performed. Those species are able to maintain a high level of chlorophyll known as tolerant species.

Leaf extract pH

Leaf extract pH is a biochemical parameter of plants that signifies as an indication of stress. The pH of the leaf extracts ranges from 6.8 ± 0.13 to 5.1 ± 0.06 and 7.17 ± 0.14 to 5.53 ± 0.12 , at the polluted and control areas, respectively, with *F. hispida* and *B. orellana* having the highest and lowest values. The pH of plants has shown a good relationship with the susceptibility to pollution. Low pH reduced the photosynthetic activity by altering the stomatal activity while a higher level of pH in plants can increase tolerance towards pollution by enhancing the synthesis rate of ascorbic acid from hexose sugar (Escobedo et al. 2008). The presence of acidic pollutants in the environment increases the acidic nature of the leaf by decreasing the pH. Hence, those

plant species have a higher leaf extract pH which are considered as pollution-tolerant species. It is evidenced that all the plant species collected from the mining area showed an acidic pH. In comparison with the control area, all plants showed an acidic nature of leaf extract which may be due to the accumulation of acidic nature heavy metals in apoplast. Mostly heavy metals like Cr, Ni and Zn form acidic radicals in the leaf matrix by reacting with cellular water. Karmakar and Padhy (2019) and Nadgórska-Socha et al. (2017) have reported that leaf pH is influenced by the presence of heavy metals like Cd, Cr, Ni and Zn.

Air pollution tolerance index

The APTI is an established method for the evaluation of plants with regard to their sensitivity to pollutants. Depending on the APTI value, plant species are categorized into four different groups, i.e. ≤ 15 = sensitive, $15-19$ = intermediate, $20-24$ = moderately tolerant and > 24 = tolerant (Singh et al. 1991). The APTI values of each plant species were calculated using the mean values of T, P, R and A values, so the standard deviation of the APTI values is not mentioned (Table 1). At the mining area, it is shown that the APTI value ranges from 30.24 to 12.48. The highest value was recorded in *H. pubescens* (30.24) followed by *M. peltata* (25.96), *T. orientalis* (25.62) and *F. hispida* (22.52). According to the APTI range, *H. pubescens*, *M. peltata* and *T. orientalis* are tolerant species as their APTI value ≥ 24 . *F. hispida* (22.52) is an only moderately tolerant species. *C. roxburghii* (18.71) and *T. arjuna* (16.51) are intermediate species, whereas *A. excelsa*, *B. orellana* and *C. tomentosa* are sensitive species (APTI ≤ 15). At the control area, the APTI showed a declining (Fig. 1) value ranging from 21.51 to 11.41, with *M. peltata* and *C. roxburghii* having the highest and lowest values, respectively. All species are grouped into three categories, i.e. sensitive, intermediate and moderately tolerant; no species could qualify as tolerant at the control areas. A strong correlation between ascorbic acid and APTI at both mining ($R^2 = 0.86$) and control area ($R^2 = 0.83$) was noticed, whereas the remaining three parameters (pH, total chlorophyll and relative water content) have an insignificant low correlation with APTI. This indicates that, as the pollution level increases ascorbic acid in plants also increases to combat the pollution stress. Some previous studies have also found a similar kind of correlation between ascorbic acid and APTI value (Rai and Panda 2014; Bharti et al. 2018; Roy et al. 2020), since the concentration of heavy metal is higher in the mining area (Table 4) and the MAI of the plants is also higher, which leads to an increase in the antioxidant level to combat such high stress. In the control area, such stressors are below the permissible level, plant species not increasing their antioxidant level. Depending on the stressor level, same species showed different APTI indices in both areas.

Plantation of tolerant plant species, to mitigate environmental pollution, is a sustainable prospective to meet industrial and commercial growth.

Anticipated performance index

Different species act differently to different pollution stresses. Therefore, grouping of species on the basis of the APTI value is only useful to evaluate the tolerance status of the plant. Hence, only the APTI value is not enough for the selection of appropriate tolerant species to develop greenbelts in polluted areas. By incorporating the APTI value with economic value and morphological characters like plant habit, canopy structure, laminar character and plant types, the API grade of different species is calculated. In the mining area, *M. peltata* scored the highest API value followed by *H. pubescens* and *F. hispida* which were qualified as excellent, very good and good species, respectively, for greenbelt development (Table 2). Among the rest of the species, two (*T. arjuna* and *T. orientalis*) were qualified as moderate. *C. tomentosa*, *C. roxburghii*, *A. excelsa*, *Bixa orellana* and *W. fruticosa* are poor, very poor and not recommended for greenbelt development, because of their low API value (50–25). In control areas, a similar result was also found. *M. peltata* scored the highest API value followed by *H. pubescens* and *F. hispida*. But all three species were qualified as good species for greenbelt development. Among the rest of the plant species, *T. arjuna* is moderate, *C. tomentosa* is poor, three species (*A. excelsa*, *B. orellana* and *C. roxburghii*) are very poor and *W. fruticosa* is not recommended for greenbelt development. Screening all the species by using only the APTI value, *H. pubescens*, *M. peltata* and *T. orientalis* have been assessed as tolerant species and *F. hispida* was a moderately tolerant species. But after combining the APTI value with the morphological and economic value, then *F. hispida* qualified as a tolerant species along with *H. pubescens* and *M. peltata* and *T. orientalis* qualified as moderate along with *T. arjuna*.

Dust capturing capacity

The variation of dust capturing capacity of collected species at the mining and control areas are depicted in Table 3. *F. hispida* was found to have the highest dust capturing capacity (5.94 mg/cm²) in the mining area followed by *C. tomentosa* (4.26 mg/cm²), *H. pubescens* (3.6 mg/cm²), *M. peltata* (3.51 mg/cm²) and *T. orientalis* (3.5 mg/cm²). The remaining species (*A. excelsa*, *B. orellana*, *C. roxburghii*, *T. arjuna*, *W. fruticosa*) showed the lower dust capturing capacities in the mining area. In control areas, *M. peltata* (0.39 mg/cm²) showed the highest dust capturing capacities followed by *C. tomentosa* (0.37 mg/cm²) and *F. hispida* (0.36 mg/cm²) and the lowest dust capturing capacities shown by *W. fruticosa*

Table 2 Assessment of API of selected plant species

Species	APTI	Plant habit	Canopy structure	Type of plant	Laminar		Hardiness		Economic importance	Grade allotted		Assessment
					Size	Texture	Hardiness	% scoring				
Mining												
<i>A. excelsa</i>	+	+	+	-	+	+	-	+	+	6	37.5	Very poor
<i>B. orellana</i>	+	+	+	+	+	-	-	+	+	6	37.5	Very poor
<i>C. tomentosa</i>	+	+	+	-	+	+	+	+	+	8	50	Poor
<i>C. roxburghii</i>	++	+	+	+	+	-	+	+	+	8	50	Poor
<i>F. hispida</i>	+++	+	+	-	+	+	+	+	+	11	68.75	Good
<i>H. pubescens</i>	++++	+	+	+	+	-	-	+	+	12	75	Very good
<i>M. peltata</i>	++++	+	++	+	+	+	-	+	+	13	81.25	Excellent
<i>T. tomentosa</i>	++	++	++	-	+	-	-	+	+	9	56.25	Moderate
<i>T. orientalis</i>	+++	+	+	+	-	+	-	+	+	9	56.25	Moderate
<i>W. fruticosa</i>	+	-	-	+	-	+	-	+	+	4	25	Not recommended
Control												
<i>A. excelsa</i>	-	+	+	-	+	+	-	+	+	5	31.25	Very poor
<i>B. orellana</i>	+	+	+	+	+	-	-	+	+	6	37.5	Very poor
<i>C. tomentosa</i>	+	+	+	-	+	+	+	+	+	8	50	Poor
<i>C. roxburghii</i>	-	+	+	+	+	-	+	+	+	6	37.5	Very poor
<i>F. hispida</i>	++	+	+	-	+	+	+	+	+	10	62.5	Good
<i>H. pubescens</i>	+++	+	+	+	+	-	-	+	+	10	62.5	Good
<i>M. peltata</i>	++	++	++	+	+	+	-	+	+	11	68.75	Good
<i>T. arjuna</i>	++	++	++	-	+	-	-	+	+	9	56.25	Moderate
<i>T. orientalis</i>	+	+	+	+	+	+	-	+	+	6	37.5	Very poor
<i>W. fruticosa</i>	+	-	-	+	+	+	-	+	+	4	25	Not recommended

Table 3 DCC of plants (mg/cm²) in mining and control area

Species	Mining	Control
<i>A. excelsa</i>	1.4 ± 0.26	0.17 ± 0.02
<i>B. orellana</i>	1.07 ± 0.19	0.19 ± 0.1
<i>C. tomentosa</i>	4.26 ± 0.27	0.37 ± 0.07
<i>C. roxburghii</i>	1.25 ± 0.30	0.16 ± 0.14
<i>F. hispida</i>	5.94 ± 0.43	0.36 ± 0.2
<i>H. pubescens</i>	3.61 ± 0.43	0.23 ± 0.04
<i>M. peltata</i>	3.52 ± 0.32	0.39 ± 0.1
<i>T. arjuna</i>	2.54 ± 0.55	0.33 ± 0.2
<i>T. orientalis</i>	3.55 ± 0.36	0.28 ± 0.07
<i>W. fruticosa</i>	1.03 ± 0.11	0.14 ± 0.06

Table 4 Concentrations of heavy metals in soil (mg/kg)

Element	Mining	Control
Al	24,226.67 ± 2591.85	253.41 ± 35.34
Cr	78,013.33 ± 4978.29	108.3 ± 30.75
Fe	154,766.67 ± 10,150.92	1277.92 ± 354.92
Ni	2038.67 ± 181.75	40.77 ± 5.92
Pb	16.67 ± 2.96	2.6 ± 0.52
Zn	209.67 ± 5.33	46.4 ± 14.31

(0.14 mg/cm²) followed by *C. roxburghii* (0.16 mg/cm²) and *B. orellana* (0.17 mg/cm²). The dust capturing capacity of plants significantly depended on species to species. Several micro-characters, macro-characters and the surrounding environment affect the plant's dust capturing capacity. Micro-characters such as stomatal size and density, presence of trichome, pubescence, wax layer and macro-characters like height of plant, canopy structure, leaf arrangement on stem and petiole area significantly influence the dust capturing capacity of plant species (Mo et al. 2015; Leonard et al. 2016). *F. hispida*, *C. tomentosa*, *H. pubescens*, *M. peltata* and *T. orientalis* contribute a high level of dust capturing capacities due to the presence of trichome in both sides of the leaf surface. Similar findings were supported by some recent studies by Roy et al. (2020), and Chaudhary and Rathore (2019) found higher dust capturing capacities in genus *Ficus*. Roy et al. (2020) also found that *A. excelsa* had the lowest value due to its small and glabrous leaf petiole. Single-factor ANOVA showed significant differences in dust capturing capacities in both areas ($F = 24.11$, $F_{critical} = 4.41$), at the 0.05 significance level. Area-wise differences may be due to high atmospheric pollution and anthropogenic activity.

Assessment of heavy metal in soil

Mining area soils showed a relatively very high amount of different heavy metals in order Fe > Cr > Al > Ni > Zn > Pb. The concentration of Fe is higher than those of other metals which is 154,766.67 ± 10,150.92 mg/kg. The high Fe content in the mining soil is due to the presence of hematite and goethite. The concentrations of the other five heavy metals namely Cr, Al, Ni, Zn and Pb are depicted in Table 4. All heavy metal concentrations in the mining area soils surpass the ecotoxicological level as per WHO standard. On the other hand, all the heavy metal concentrations in the control forest area are within the limit of the WHO standard.

Assessment of heavy metal within the leaf

Heavy metal accumulation in plants can be from both adsorption through stomata from areal dust deposition and root uptake from soil. So, it is impossible to distinguish whether the accumulated heavy metal came from the soil or from the air (Serbula et al. 2012; Norouzi et al. 2015). In mining areas, the analysed plant leaves revealed that they surpassed the permissible limit of the WHO (1996) standards. The Al, Cr and Fe contents were extremely higher within all investigated plant leaves. The average metal accumulation ranges of all investigated plants at both the areas are represented in Table 5. The Fe concentration was found to be higher in all plant leaves of the mining area ranging from 1028.87 ± 55.69 to 6793.93 ± 797.15 mg/kg and in the control area ranging between 245.89 ± 36.76 and 57.04 ± 15.18. The highest Fe accumulation was found in *F. hispida* leaves in the mining area (6793.93 ± 797.15 mg/kg) and in *C. tomentosa* leaves in the control area (245.89 ± 36.76 mg/kg). Al accumulation ranged between 159.43 ± and 1358.70 ± mg/kg at the mining area. The highest Al accumulation was detected in *F. hispida* followed by *M. peltata* (833.07 ± mg/kg), and the lowest was detected in *W. fruticosa* followed by *T. orientalis* (183.20 ± mg/kg). Compared with the control area, the highest accumulation was found in *F. hispida* (5.23 ± mg/kg) and the lowest was found in *T. orientalis* (0.62 ± mg/kg). The highest content of Cr accumulation was found in *F. hispida* (432.27 ± mg/kg) followed by *M. peltata* (328.03 ± mg/kg), and the lowest accumulation was found in *C. tomentosa* (48.97 ± mg/kg) followed by *B. orellana* (52.83 ± mg/kg). The maximum accumulation of Ni was found in *C. roxburghii* (64.57 ± mg/kg) followed by *M. peltata* (62.57 ± mg/kg).

Metal accumulation index

MAI is used to evaluate the heavy metal accumulation capacities in plants using a standard formula (Hu et al. 2014; Karmakar and Padhy 2019). The MAI values of the analysed

Table 5 Heavy metal in plant leaves (mg/kg) and their respective MAI

Species	Al	Cr	Fe	Ni	Pb	Zn	MAI
Mining							
<i>A. excelsa</i>	389.76 ± 20.06	195.7 ± 18.74	2570 ± 188.47	38.06 ± 2.8	11.17 ± 1.26	35.6 ± 2.95	7.46
<i>B. orellana</i>	241.16 ± 36.26	52.58 ± 2.8	3105.33 ± 374.78	37.83 ± 6.4	2.06 ± 0.69	18.86 ± 1.62	5.17
<i>C. tomentosa</i>	239.56 ± 28.35	48.96 ± 2.58	3072 ± 30.99	19.66 ± 2.43	1.8 ± 0.53	18.65 ± 0.44	17.29
<i>C. roxburghii</i>	430.76 ± 23.7	151.71 ± 24.56	3870.3 ± 695.79	64.56 ± 7.66	2.03 ± 0.2	24.23 ± 2.65	5.52
<i>F. hispida</i>	1358.76 ± 23.7	432.26 ± 24.16	6793.93 ± 797.15	32.06 ± 1.21	1.06 ± 0.07	15.86 ± 2.25	10.31
<i>H. pubescens</i>	404.42 ± 13.47	110.4 ± 5.57	1891.93 ± 67.55	45.13 ± 4.22	1.23 ± 0.26	19.13 ± 2.37	9.74
<i>M. peltata</i>	833.06 ± 25.15	328.03 ± 14.1	5239 ± 253.49	62.56 ± 4.33	2.76 ± 0.61	37.93 ± 3.92	10.16
<i>T. tomentosa</i>	326.16 ± 11.2	99.36 ± 12.26	1677.53 ± 170.98	33.06 ± 2.6	6.8 ± 0.79	30.06 ± 6.03	7.05
<i>T. orientalis</i>	183.2 ± 10.42	72.23 ± 6.37	1028.86 ± 55.69	19.26 ± 1.85	3.13 ± 0.9	20.93 ± 2.01	6.89
<i>W. fruticosa</i>	159.43 ± 14.29	162.13 ± 31.5	2138.16 ± 232.16	27.5 ± 3.01	2.18 ± 0.32	17.2 ± 1.62	4.99
Control							
<i>A. excelsa</i>	1.44 ± 0.33	1.85 ± 0.19	74 ± 21.41	1.82 ± 0.38	0.24 ± 0.07	4.23 ± 1.05	2.82
<i>B. orellana</i>	2.37 ± 0.67	1.29 ± 0.32	193.33 ± 54.07	2.11 ± 0.8	0.05 ± 0.01	3.58 ± 1.37	1.94
<i>C. tomentosa</i>	2.52 ± 0.48	1.69 ± 0.41	245.89 ± 36.75	1.09 ± 0.25	0.06 ± 0.01	4.95 ± 1.27	2.88
<i>C. roxburghii</i>	4.43 ± 0.72	2.56 ± 0.45	184 ± 43.18	1.3 ± 0.8	0.07 ± 0.006	0.94 ± 0.2	3.4
<i>F. hispida</i>	5.23 ± 1.36	3.1 ± 0.08	240.9 ± 26.78	1.88 ± 0.22	0.05 ± 0.009	2.07 ± 0.82	6.38
<i>H. pubescens</i>	0.89 ± 0.22	2.01 ± 0.46	61.55 ± 7.94	2.51 ± 0.43	0	2.01 ± 0.82	2.34
<i>M. peltata</i>	1.69 ± 0.53	1.49 ± 0.26	215.11 ± 31.46	2.03 ± 0.51	0.08 ± 0.009	5.2 ± 1.22	3.15
<i>T. tomentosa</i>	0.76 ± 0.17	1.76 ± 0.15	117.47 ± 18.9	1.39 ± 0.42	0.12 ± 0.02	5.85 ± 1.75	3.17
<i>T. orientalis</i>	0.62 ± 0.28	0.53 ± 0.26	57.03 ± 15.18	1.03 ± 0.23	0.07 ± 0.02	2.46 ± 0.32	2.26
<i>W. fruticosa</i>	0.91 ± 0.3	1.35 ± 0.16	121.96 ± 28.66	1.6 ± 0.26	0.43 ± 0.01	3.37 ± 1.16	2.57

plant species are shown in Table 5. At the mining area, MAI ranged from 4.5 (*W. fruticosa*) to 17.29 (*C. tomentosa*). The trend followed was *C. tomentosa* (17.29) > *F. hispida* (10.31) > *M. peltata* (10.16) > *H. pubescens* (9.74) > *A. excelsa* (7.46) > *T. arjuna* (7.05) > *T. orientalis* (6.89) > *B. orellana* (5.17) > *W. fruticosa* (4.5). At the control area, the trend followed was *F. hispida* (6.38) > *C. roxburghii* (3.4) > *T. arjuna* (3.17) > *M. peltata* (3.15) > *C. tomentosa* (2.88) > *A. excelsa* (2.82) > *W. fruticosa* (2.57) > *H. pubescens* (2.34) > *T. orientalis* (2.26) > *B. orellana* (1.94). In both areas, *F. hispida* had a higher MAI value (mining area = 10.31, control area = 6.38), indicating that it has higher metal accumulation capacity. However, the MAI of *A. excelsa* (7.46) in the Sukinda chromite mine area is higher than that previously reported by Roy et al. (2020) in an industrial area (2.13) and a commercial area (2.03). In another study in China, a similar kind of result was found by Hu et al. (2014). The MAI value of *Sophora japonica* in Beijing (9.0) was higher than that reported in Yan'an city (2.56). Nadgórska-Socha et al. (2017) and Karmakar and Padhy (2019) also found a positive correlation between heavy metal uptakes in plant leaves and their concentration in soil. So, this fluctuation of MAI value could be due to a higher metal concentration in mining areas compared previously with the industrial and commercial areas. Plant species with higher MAI values have a better metal accumulation

capacity and considered metal-tolerant species; those species can use an impediment between contaminated and uncontaminated areas. Those plant species that have a lower MAI value are considered sensitive to metal and can be used as bioindicators of metal pollution.

Conclusion

Based on dust capturing capacities and metal accumulation index as well as APTI and API, *F. hispida*, *M. peltata* and *H. pubescens* can be used as tolerant species and could be recommended for greenbelt development to mitigate pollution. Those plant species (*A. excelsa*, *B. orellana*, *C. roxburghii*, *W. fruticosa*, *T. arjuna*, *T. orientalis*) have lower APTI and API values which could be used as sensitive species for biomonitoring of environmental pollution. Our study also finds that *C. tomentosa* is the highest metal accumulation capacities, but according to the anticipated performance index it is a poor performer towards pollution. So, these species could be used as hyper-accumulator to mitigate heavy metal contamination from air and soil. The physiological and biochemical responses of plants vary from species to species and also from area to area. But more biochemical and physiological research is needed to confirm the effect

and response of plant species in a particular pollution stress condition.

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Contribution: conceptualization of the work, supervision, preparing the manuscript.

2. Kalicharan Mandal: (first author)

Contribution: methodology, sample collection, data analysis and interpretation, preparation of the manuscript.

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