



Biological-based methods for the removal of volatile organic compounds (VOCs) and heavy metals

Mukesh Meena¹ · Priyankaraj Sonigra¹ · Garima Yadav¹

Received: 3 June 2020 / Accepted: 4 October 2020 / Published online: 23 October 2020
© Springer-Verlag GmbH Germany, part of Springer Nature 2020

Abstract

The current scenario of increased population and industrial advancement leads to the spoliation of freshwater and tapper of the quality of water. These results decrease in freshwater bodies near all of the areas. Besides, organic and inorganic compounds discharged from different sources into the available natural water bodies are the cause of pollution. The occurrence of heavy metals in water and volatile organic compounds (VOCs) in the air is responsible for a vast range of negative impacts on the atmosphere and human health. Nonetheless, high uses of heavy metals for human purposes may alter the biochemical and geochemical equilibrium. The major air contaminants which are released into the surroundings known as VOCs are produced through different kinds of sources, such as petrochemical and pharmaceutical industries. VOCs are known to cause various health hazards. VOCs are a pivotal group of chemicals that evaporate readily at room temperature. To get over this problem, biofiltration technology has been evolved for the treatment of heavy metals using biological entities such as plants, algae, fungi, and bacteria. Biofiltration technology is a beneficial and sustainable method for the elimination of toxic pollutants from the aquatic environment. Various types of biological technologies ranging from biotrickling filters to biofilters have been developed and they are cost-effective, simple to fabricate, and easy to perform. A significant advantage of this process is the pollutant that is transformed into biodegradable trashes which can decompose within an average time period, thus yielding no secondary pollutants. The aim of this article is to scrutinize the role of biofiltration in the removal of heavy metals in wastewater and VOCs and also to analyze the recent bioremediation technologies and methods.

Keywords Volatile organic compounds · Biological methods · Bioremediation · Toxic pollutants · Biofiltration · Phytoremediation

Abbreviations

WHO World Health Organization
Hg Mercury
Pb Lead

As Arsenic
Zn Zinc
Cu Copper
Ni Nickel
Co Cobalt
Cd Cadmium
Cr Chromium
Fe Iron
Sn Stannum
Se Selenium
H₂S Hydrogen sulfide
VOCs Volatile organic compounds
BTF Biotrickling filters
mm Millimeter
MBBR Moving bed biofilm reactor
PBBR Packed bed bioreactor
PCBs Polychlorinated biphenyls
PAHs Polycyclic aromatic hydrocarbons

Responsible editor: Elena Maestri

✉ Mukesh Meena
mukeshmeenamsu@gmail.com; mukeshmeenabhu@gmail.com

Priyankaraj Sonigra
priyankarajsonigra124@gmail.com

Garima Yadav
garima.y1995@gmail.com

¹ Laboratory of Phytopathology and Microbial Biotechnology, Department of Botany, Mohanlal Sukhadia University, Udaipur, Rajasthan 313001, India

Introduction

Most of the earth's surface is covered with water while less than 0.002% is consumable for humans. According to the reports of the World Health Organization (WHO), one-sixth part of the total world population does not have access to freshwater. The reason behind this problem is water pollution. Water pollution comes from human activities such as industry, agriculture, or household waste that have a large negative impact on the water bodies (rivers and ocean), environment, and human health also (Owa 2014; Alrumman et al. 2016). Since industrialization, many freshwater bodies' rivers have been receiving a higher amount of pollutants (industrial and many other wastes) in which metals are also important topics of environmental concern. These heavy metals are deposited in the environment owing to anthropogenic activities. For many years, heavy metals have been growing concern over the harmful effects of humans and aquatic ecosystems (Tiwari et al. 2016). Heavy metals refer to trace metals that are toxic in nature at low concentrations with quite high density. Generally, the atomic mass range of heavy metals is found from 63 to 200 (Srivastava and Majumder 2008; Barupal et al. 2020a, 2020b). Metals that are toxic to a living being and refer as heavy metals are listed as follows: mercury (Hg), lead (Pb), arsenic (As), zinc (Zn), copper (Cu), nickel (Ni), cobalt (Co), cadmium (Cd), chromium (Cr), stannum (Sn), etc. (Wang and Chen 2009; Liang et al. 2019; Qu et al. 2020). The foremost sources of heavy metals are electroplating industries waste and mining and many more that are discharged into water bodies and soil (Ahmady-Asbchin et al. 2008; Zhou et al. 2020). In contrast to organic waste, heavy metal removal is very difficult due to its persistent and incorruptible nature (Gupta et al. 2001; Shukla and Pai 2005; Acheampong et al. 2010). Heavy metal's indiscriminate disposal into water resources causes accumulation of toxic metal in living organism's tissues, leading to diseases and disorders. The exposure of heavy metals to humans can occur through different routes, for instance, inhalation, vaporization, and food and drink ingestion. Apart from this, these heavy metals have several environmental impacts such as the destruction of aquatic life and habitat and algal blooms (Nghah and Hanafiah 2008; Akpor et al. 2014). Besides heavy metals, volatile organic compounds (VOCs) are well-known air pollutants and a topic of concern too.

VOCs are compounds with a wide variety of chemicals that have less water solubility and volatility at ambient room temperature because of the high vapor pressure (Cicolella 2008). VOCs include industrial solvents like toluene, benzene, esters, methane, perfluorocarbons, chlorohydrocarbons (Tiwari et al. 2019), methyl tert-butyl ether (Liang et al. 2020), trichloroethylene (Yang et al. 2019), chloroform, paraffin, and ketones (Lomonaco et al. 2020). VOCs also include hydraulic fluids, petroleum fuels, paints, ink, and dry cleaning agents, aromatics that are released in the air from the chemical and refinery industries. Some of these compounds are carcinogenic and some

are mutagenic and toxic for living beings and hazardous to the environment. When VOCs oxidized in the presence of lights, they produce aromatic, chlorinated, and other compounds that are more toxic than their parent compounds (Zhao et al. 2014; Barupal et al. 2020c). VOCs like oxides of nitrogen generate ozone that forms smog which is harmful to living beings and the environment as well (Malakar et al. 2017).

VOCs pollute not only air but also water and soil. That is why it is a growing environmental concern. It is necessary to confiscate or alleviate VOCs and heavy metal from the environment. For this purpose, industries need some innovative technologies for the removal of heavy metal and VOCs from the wastewaters before discharging them into water bodies or environment. A number of physicochemical methods are used for heavy metal such as chemical precipitation, solvent extraction, adsorption, membrane separation, and ion exchange (Eccles 1995; Kurniawan et al. 2006) and for VOCs, ozonation, condensation, absorption, membrane separations, adsorption, catalytic oxidation, and incineration are employed (Kumar et al. 2011). Nonetheless, all these methods have some drawbacks like incomplete removal, required expensive equipment, monitoring system, reagent, and high energy requirements, and also generate undesirable by-products that require disposal as well. Furthermore, these methods might not be much effective and cost-effective. The biological process is an exquisite method for the management of heavy metal and VOCs and can also help to overcome some of the drawbacks of physiochemical treatment methods and provide economical, eco-friendly, and safe removal of pollutants (Mehta and Gaur 2005; Kumar et al. 2011). In this article, we discuss biological methods for pollutant removal (heavy metals, VOCs) like biofiltration, biofilters, microorganisms, biotrickling filters, bioscrubber, and biosorption.

Biological methods

Biological methods involve microorganisms for the elimination of wastewater. This method can be used for the exclusion of VOCs and heavy metals using microorganisms. Table 1 shows various microbes for the removal of different heavy metals. There are different kinds of biological methods that are widely used for pollutant removal such as biofilters, activated sludge, trickling filters, bioscrubber, and bioremediation; stabilization ponds are widely used for treating wastewater (Gunatilake 2015; Fig. 1). Here, Table 2 shows the various biological methods for the elimination of various contaminants, kinds of processes, and related plants/microbes. Biological methods gradually got popularity due to the following reasons: fewer chemicals and equipment and operating system required and its eco-friendly nature (Srivastava and Majumder 2008).

Table 1 Microbes mediated removal of heavy metal

Microorganisms	Methods	Heavy metals and VOCs	References
<i>Eichhornia crassipes</i>	Biofiltration	Pb, Cr, Zn, Mn, Cu	Tiwari et al. (2007)
<i>Azolla</i>	Biofiltration	Sr(II)	Cohen-Shoel et al. (2002)
<i>Pseudomonas taiwanensis</i> , <i>Acinetobacter guillouiae</i> , <i>Klebsiella pneumoniae</i>	Biofiltration	Cr(VI), Cu(II), Zn(II)	Majumder (2015)
<i>Gracilaria</i> sp.	Biofilter	Al, Cr, Zn	Kang and Sui (2010).
<i>Rhizopus nigricans</i>	Biosorption	Pb	Hassan and El-Kassas (2012)
<i>Pleurotus ostreatus</i>	Biosorption	Pb	Barros Júnior et al. (2003)
<i>Aspergillus cristatus</i>	Biosorption	Cd	Martinez-Juarez et al. (2012)
<i>Aspergillus niger</i>	Biosorption	Cd	Bunluesin et al. (2007)
<i>Hydrilla verticillata</i>	Biosorption	Cd	Acosta Rodríguez et al. (2013)
<i>Aspergillus flavus</i> I–V, <i>Aspergillus fumigatus</i> I–II	Biosorption	Hg	Murugesan et al. (2006)
<i>Penicillium chrysogenum</i>	Biosorption	As	Mamisahebei et al. (2007)
Waste tea fungal	Biosorption	As(III), As(V)	Shoaib et al. (2012)
Waste tea fungal	Biosorption	As	Velkova et al. (2012)
<i>Aspergillus niger</i>	Biosorption	Ni	Tay et al. (2012)
<i>Pleurotus ostreatus</i>	Biosorption	Cu	Shipra et al. (2012)
<i>Aspergillus lentulus</i>	Biosorption	Cu	Sutherland and Venkobachar (2010)
<i>Fomes fasciatus</i>	Biosorption	Cu	Parungao et al. (2007)
<i>Penicillium canescens</i>	Biosorption	Cr(VI)	Chhikara et al. (2010)
Fungal (living) mycelium of <i>Phanerochaete chrysosporium</i>	Biosorption	Cr(VI), Cu	Sethi et al. (2010)
<i>Mucor</i>	Biosorption	Cr(VI)	El-Kassas and El-Taher (2009)
<i>Trichoderma viride</i>	Biosorption	Cr(VI)	Sala Cossich et al. (2002)
<i>Aspergillus niger</i>	Biosorption	Cr(VI)	Javaid and Bajwa (2007)
<i>Pleurotus ostreatus</i>	Biosorption	Cr(III), Cr(VI)	Park et al. (2005)
<i>Aspergillus niger</i>	Biosorption	Cr(VI)	Kujan et al. (2006)
<i>Saccharomyces cerevisiae</i>	Biosorption	Cu	Anaemene (2012)
<i>Candida</i> spp.	Biosorption	Cu	Mapolelo and Torto (2004)
<i>Thiobacillus thiooxidans</i>	Biosorption	Cu	Nagashetti et al. (2013)
<i>Saccharomyces cerevisiae</i>	Biosorption	Cr(VI)	Abd-Elsalam (2011)
<i>Saccharomyces cerevisiae</i>	Biosorption	Cr(VI), Cu	Davis et al. (2003)
<i>Candida utilis</i>	Biosorption	Cr(VI)	Arakaki et al. (2011)
<i>Schizosaccharomyces pombe</i>	Biosorption	Cr(VI)	Prakash and Kumar (2013)
Spent yeast	Biosorption	Cr(III)	Pandey et al. (2009)
Yeast biomass	Biosorption	Cr(VI)	Hamuda and Tóth (2012)
<i>Saccharomyces cerevisiae</i>	Biosorption	Cu, Cd, Pb, Ni	Van Wyk (2011)
<i>Saccharomyces cerevisiae</i>	Biosorption	Cr, Sn	Machado et al. (2009).
<i>Saccharomyces cerevisiae</i>	Biosorption	Cu, Ni, Zn	Das et al. (2007)
Mycelial biomass of <i>Pleurotus florida</i>	Biosorption	Cd	Yan and Viraraghavan (2000)
<i>Saccharomyces cerevisiae</i>	Biosorption	Cd	Hamuda and Tóth (2012)
Strain of <i>Saccharomyces cerevisiae</i>	Biosorption	Cr, Cu, Ni, Zn	Machado et al. (2008)
<i>Mucor rouxii</i>	Biosorption	Pb, Cd, Ni, Zn	El-Sayed (2013); Meena et al. (2017f, b)
<i>Saccharomyces cerevisiae</i>	Biosorption	Cd	El-Sayed (2013)
<i>Rhodospirillum</i> sp.	Biofiltration	Cd, Hg, Pb, Ni	Chatterjee (2002)
<i>Gallionella ferruginea</i>	Biofiltration	As, Mn	Fe Katsoyiannis and Zouboulis (2004)
<i>Leptothrix</i> sp.	Biofiltration	As, Mn	Fe Katsoyiannis and Zouboulis (2004)

Table 1 (continued)

Microorganisms	Methods	Heavy metals and VOCs	References
<i>Pseudomonas</i> sp.	Bioadsorption	Cr, As	Valls et al. (2000); Meena et al. (2017a, d)
<i>Desulfovibrio</i> sp.	Biofiltration	Cu, Zn, Ni, Fe, As	Jong and Parry (2003)
<i>Thiomonas</i> sp.	Biofiltration	As, Fe	Casiot et al. (2003)
<i>Escherichia coli</i>	Bioaccumulation	Hg, Ni	Deng et al. (2003)
<i>Thauera selenatis</i>	Biofiltration	Zn, Cd, Co, Cu, Ni, Pb, Cr, Hg	Mergeay et al. (2003)
<i>Penicillium chrysogenum</i>	Biological method	Zn, Cu, Ni, As	Loukidou et al. (2003)
<i>Aspergillus niger</i>	Bioaccumulation	Ni, Cu, Pb, Cr	Dursun et al. (2003); Meena et al. (2017c, f)
<i>Coriolus hirsutus</i>	Activated sludge	Cd	Miyata et al. (2000)
<i>Trametes versicolor</i>	Biotransformation	Cr, Co	Blázquez et al. (2004)
<i>Mucor rouxii</i>	Bioadsorption	Pb, Cd, Zn, Ni	Yan and Viraraghavan (2000)
Brown algae	Biosorption	Cd, Cu, Zn, Pb, Cr, Hg	Davis et al. (2003); Meena et al. (2017e)
Green algae	Bioaccumulation	Cu, Hg, Fe, Zn, Pb, Cd	Haritonidis and Malea (1999)
<i>Scenedesmus</i> genus	Biological method	Cu, Ni, Cd, Cr, Cu	Pena-Castro et al. (2004)
Sulfate-reducing bacteria (SRB)	Biofiltration	Zn	Suriya et al. (2013).
<i>Enterobacter cloacae</i>	Biofiltration	Pb, Cu, Cr(VI), Hg, Cd	Rani et al. (2010)
<i>Bacillus</i> sp., <i>Pseudomonas</i> sp., <i>Micrococcus</i> sp.	Biofiltration	Cu, Cd, Pb	Hussein et al. (2004).
<i>Pseudomonas</i> sp.	Biofiltration	Cr(VI), Cu, Cd, Ni	Liu et al. (2004)
<i>Thiobacillus thiooxidans</i>	Biofiltration	Zn, Cu	Nagashetti et al. (2013)
<i>Sargassum filipendula</i>	Biofiltration	Cu, Ni	Rinku et al. (2012)
<i>Sargassum</i> sp.	Biofiltration	Cr(III)	Yavuz et al. (2006)
Green algae (<i>Spirogyra</i> spp.)	Biofiltration	Cr(VI)	Subhashini et al. (2011)
Microalgae	Biofiltration	Cu, Zn	Saravanan et al. (2011)
Brown algae (<i>Fucus vesiculosus</i>)	Biofiltration	Cu, Pb, Ni, Cd	Brinza et al. (2007)
<i>Pseudomonas</i> sp.	Biofiltration	Benzene	Sene et al. (2002)
<i>Alcaligenes xylosoxidans</i>	Biofiltration	Benzene	Yeom and Daugulis (2001); Meena et al. (2017g)
<i>Cladosporium sphaerospermum</i>	Biofiltration	Benzene	Qi et al. (2002)
<i>Cladosporium resinae</i> , <i>C. sphaerospermum</i> , <i>Exophiala lecanii-corni</i> , <i>Mucor rouxii</i> , <i>Phanerochaete chrysosporium</i>	Biofiltration	Butylacetate	Qi et al. (2002); Meena et al. (2017i)
<i>Pseudomonas</i> sp.	Biofiltration	Chlorobenzene	Seigneur et al. (2001)
<i>Candida utilis</i>	Biofiltration	Ethanol	Christen et al. (2002)
<i>Rhodococcus fascians</i>	Biofiltration	Ethylacetate	Hwang et al. (2002)
<i>Cladosporium resinae</i> , <i>C. sphaerospermum</i> , <i>Exophiala lecanii-corni</i> , <i>Phanerochaete chrysosporium</i>	Biofiltration	Ethylbenzene	Qi et al. (2002); Meena and Zehra (2019)
<i>Cladosporium resinae</i> , <i>C. sphaerospermum</i> , <i>Exophiala lecanii-corni</i>	Biofiltration	Methylethylketone	Qi et al. (2002)
<i>Rhodococcus</i> sp.	Biofiltration	Methylethylketone	Amanullah et al. (2000)
<i>Cladosporium resinae</i> , <i>C. sphaerospermum</i> , <i>Exophiala lecanii-corni</i> , <i>Phanerochaete chrysosporium</i>	Biofiltration	Methylethylketone	Qi et al. (2002)
<i>Pseudomonas aeruginosa</i>	Biofiltration	Methyl-tertbutyl-ether, pentane	Dupasquier et al. (2002)
<i>Aspergillus</i> sp.	Biofiltration	α -Pinene	

Table 1 (continued)

Microorganisms	Methods	Heavy metals and VOCs	References
			Diehl and Saileela (2000); Meena and Swapnil (2019)
<i>C. sphaerospermum</i>	Biofiltration	Styrene	Qi et al. (2002)
<i>Tsukamurella, Pseudomonas, Sphingomonas, Xanthomonas Pseudomonas putida</i>	Biofiltration	Styrene	Arnold et al. (1997)
<i>Pseudomonas pseudoalcaligenes</i>	Biofiltration	Toluene	Park et al. (2002); Meena and Samal (2019)
<i>Exophiala lecanii-corni</i>	Biofiltration	Toluene	Oh and Park (2000)
<i>Scedosporium apiospermum</i>	Biofiltration	Toluene	Woertz et al. (2001)
<i>Paecilomyces variotii Exophiala oligosperma</i>	Biofiltration	Toluene	García-Peña et al. (2001)
<i>Fusarium solani</i>	Biofiltration	Hexane	Estévez et al. (2005); Meena et al. (2016a, 2016b, 2016c)
Bacterial/fungal consortium <i>Fusarium solani</i>	Biofiltration	Toluene n-pentane	Vergara-Fernández et al. (2006)
Fungal consortium	Biofiltration	n-Hexane	Dorado et al. (2008)
<i>Corynebacterium jeikeium, Corynebacterium nitrilophilus, Micrococcus luteus, Pseudomonas mendocina, Sphingobacterium thalophilum, Turicella otitidis</i>	Biofiltration	Toluene	Iranmanesh et al. (2015)
<i>Pseudomonas putida</i>	Biofiltration	Benzene, Toluene and Phenol	Strauss et al. (2000)
<i>Paecilomyces variotii</i>	Biofiltration	Toluene	Abuhamed et al. (2004)
<i>Pseudomonas putida</i>	Biofiltration	Toluene	Aizpuru et al. (2005); Meena et al. (2015)
<i>Exophiala spp. Mytilus edulis</i>	Biofiltration	Benzene, Toluene, Ethylbenzene, and Xylene (BTEX)	Park and Jung (2006); Maestre et al. (2007)
<i>Pseudomonas aeruginosa, Aspergillus oryzae, Penicillium sp., Pseudomonas putida, Aspergillus oryzae, Pseudomonas putida, Penicillium sp., Chryseobacterium indologenes, Halosporangium sp.</i>	Biofiltration	Styrene	Mohammad et al. (2007); Torretta et al. (2015); Raboni et al. (2017)
<i>Cladophialophora sp.</i>	Biofiltration	Toluene	Paca et al. (2009); Meena et al. (2013)
<i>Bacillus cereus</i>	Biotrickling	Toluene	Prenafeta-Boldú et al. (2008)
<i>Stenotrophomonas maltophilia</i>	Biofiltration	Toluene	Li et al. (2008); Kumari et al. (2018a, 2018b); Raboni et al. (2017)
			Ryu et al. (2008)

Biofiltration technologies

Harm to environmental systems and human health by different types of pollutants release has led to the development of various methodologies for the treatment of pollutants and their source. In this regard, biofiltration has been known as a viable option for the removal of pollutants (Vikrant et al. 2018). It has the basic design like a fixed-bed bioreactor in which a microorganism is immobilized to a bed of inorganic or organic

porous-supporting medium (Berenjian et al. 2012). The pollutants passed through the medium (Gómez-Borraz et al. 2017), degraded by microorganisms (Carpenter and Helbling 2017; Jeong et al. 2017). Microorganisms behave as a biocatalyst and developed biofilm. Supporting medium provides large surface areas and additional nutrients (Srivastva et al. 2017; Rene et al. 2018), homogenous distribution of the pollutants (Cassini et al. 2017), and water retention required to keep the biofilm alive (Mačaitis et al. 2014). These conditions

Table 2 Different biological methods for the removal of pollutants

Method(s)	Process	Pollutant(s)	Plant(s)\microbe(s)	Reference(s)
Phytotransformation	Plant uptake and degradation of organic Compounds	Xenobiotic substances	<i>Cannas</i>	Subramanian et al. (2006)
Phytoextraction	Remove metals pollutants and organics from soil by accumulate in plant parts	Cd, Pb, Zn, As, petroleum, hydrocarbons and radionuclides	<i>Viola baoshanensis</i> , <i>Sedum alfredii</i> , <i>Rumex crispus</i>	Macek et al. (2000); Zhuang et al. (2007)
Phytostabilization	Use of plants to reduce the bioavailability of pollutants in the environment	Cu, Cd, Cr, Ni, Pb, Zn	<i>Anthyllis vulneraria</i> , <i>Festuca arvernensis</i>	Vázquez et al. (2006)
Rhizofiltration	Roots absorb Groundwater with pollutants, mainly metals such as Zn, Pb, Cd from water and aqueous waste streams	Zn, Pb, Cd, As	<i>Brassica juncea</i>	Verma et al. (2006); Meena et al. (2018)
Phytodegradation	Plants and associated microorganisms degrade organic pollutants	DDT, explosives, waste and nitrates	<i>Elodea Canadensis</i> , <i>Pueraria</i>	Garrison et al. (2000); Newman and Reynolds (2004)
Bioaugmentation	Addition of exogenous microorganisms with the ability of degrading the contaminants that are recalcitrant to the indigenous microbiota	PAHs, Cr, Pb	<i>Bacillus cereus</i> , <i>Fusarium oxysporum</i> , <i>Actinobacteria</i> , <i>Agaricus bisporus</i>	Jacques et al. (2008); Polti et al. (2014); García-Delgado et al. (2015); Meena et al. (2020)
Bioleaching	Specific microorganisms like <i>Thiobacillus ferrooxidans</i> and <i>T. thiooxidans</i> promote metal solubilization	Cr, Cu, Pb, Zn	<i>Acidithiobacillus</i> spp., <i>Aspergillus niger</i>	Akinci and Guven (2011); Ren et al. (2009); Zehra et al. (2020)
Biofilters	Application of bacteria in filters for the decontamination of polluted water and wastes	Cu, Cd	Micro-algal/bacterial biomass	Loutseti et al. (2009)
Bioventing	Combination of venting of soil to remove the volatile compounds with bioremediation that uses oxygen to degrade the organic contaminants	Phenanthren toluene	Multispecies system	Frutos et al. (2010); Hong and Xingang (2011); Zehra et al. (2015)
Composting	Nutrients are added to soil that is mixed to increase aeration and activation of indigenous microorganisms	Phenolic compounds heavy metals	<i>Thermoascus aurantiacus</i> , <i>Eudrilus eugeniae</i>	Ghaly et al. (2011); Soobhany et al. (2015)
Biosparging	Involving the injection of air below the water table to increase groundwater oxygen concentrations and enhance the rate of biological degradation of contaminants by naturally occurring bacteria	Trichloroethene and <i>cis</i> -dichloroethene	<i>Pseudomonas cepacia</i> G4, <i>Pseudomonas cepacia</i> G4PR1, <i>Pseudomonas mendocina</i> KR1, <i>Pseudomonas putida</i> F1, <i>Methylosinus trichosporium</i> OB3b	Tovanabootr et al. (2001); Meena et al. (2019a, 2019b)
Bioreactors	Slurry reactors (aqueous reactors)	Pyrene PAHs	<i>Pseudomonas</i> spp., <i>Pseudomonas aeruginosa</i>	Nasseri et al. (2010); Mohan et al. (2008)

altogether give suitable internal environments to pollutant-degrading microorganisms (Ibanga et al. 2018). In this process, separation is present between the microorganisms and the wastewater, while the wastewater flows through a support material on that microorganisms are immobilized (Cohen 2001; Chen and Hoff 2012). In biofiltration, the removal efficiency of VOC is also related to its bioavailability (Yang et al. 2019; Pettit et al. 2019). The removal efficiency of VOCs is directly correlated with Henry's constant. According to this, the lower solubility ratio of pollutants in water shows less

biodegradability in the biofiltration process (Miller et al. 2019). The order of degradation is as follows: alcohols > esters > ketones > aromatic hydrocarbons > aliphatic hydrocarbons (Jaber et al. 2017; Vikrant et al. 2018).

Biofilters

The biofilters are easy to manufacture and operate and are economical as well. Using biofilter, the pollutants are transformed into a biodegradable form which can be decomposed

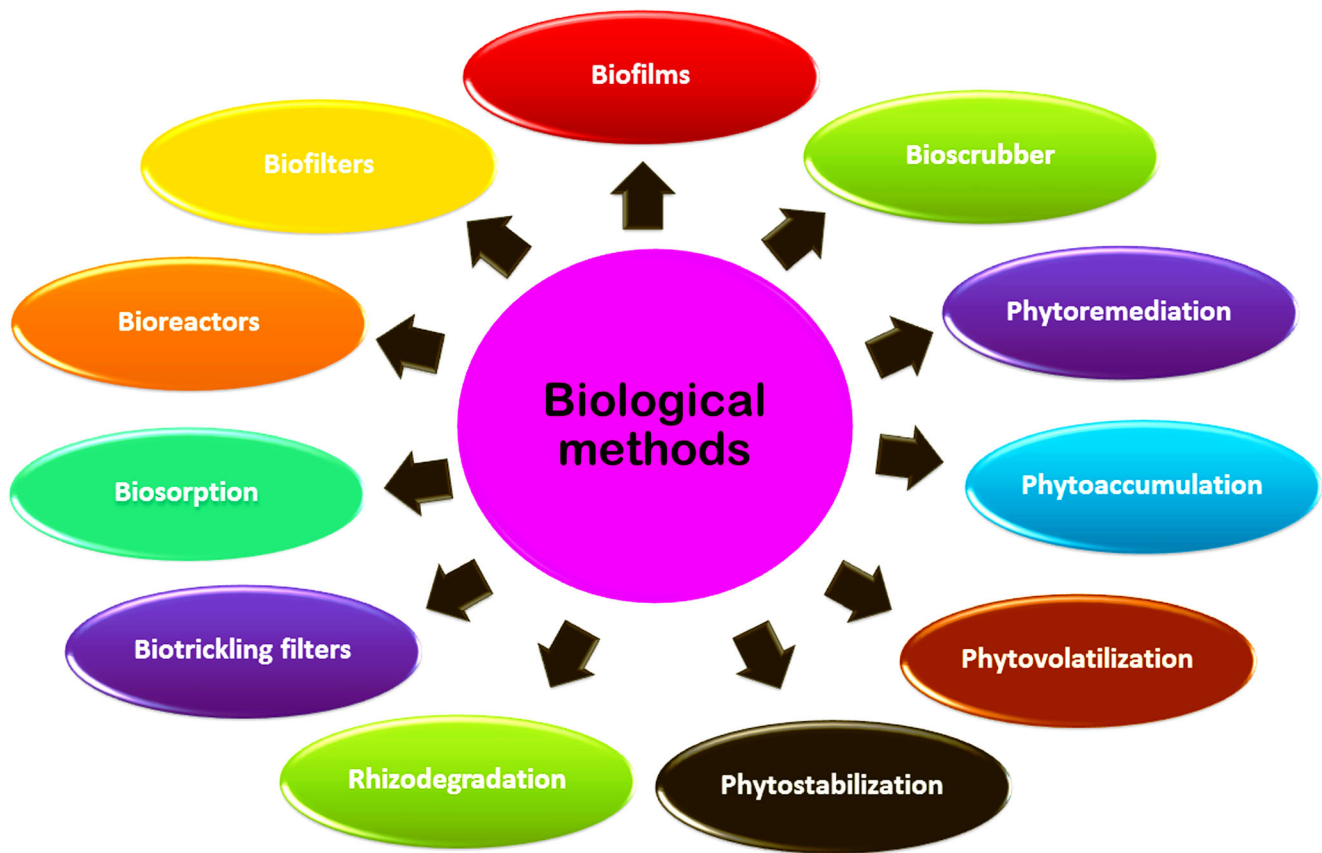


Fig. 1 Various kinds of biological methods for the removal of heavy metals and VOCs

within a moderate timeframe without any secondary pollutant production (Gopinath et al. 2018). A basic design of biofilter is constructed with a supporting medium which provides the required nutrients to the growing microorganisms (Fig. 2). Microorganisms have important roles in the process of pollutant breakdown present in the environment (Srivastava and Majumder 2008). A biofilter consists of biologically active material which is the 1 m in height (Kumar et al. 2013). Materials that are usually used in filters comprise compost, soil, bamboo, or peat activated carbon-based packing material (Lim et al. 2015; Kumar et al. 2019). The maintenance of a biologically active layer, known as the biofilm support medium, has to be kept wet (Zou et al. 2012). Biofilm is gradually developed by microorganisms (algae, protozoa, aerobic, anaerobic, and facultative bacteria, fungi, etc.) and surrounds the biofilter materials. It gives favorable conditions for the microbe’s growth. Compost obtained from the leaves, bark, or other trees’ parts is usually used as a basic filter material. Other materials which can be used as a support medium may include porous clay or polystyrene spheres. These mediums are sometimes used to increase active surface area and reliability, reduce backpressure, and increase the useful life of filter material (Leson and Winer 1991; Barupal et al. 2019). Biofilter’s pollutant removal efficiency mostly depends on the

attribute and nature of its filter media. Porosity and degree of compaction should be minimum to reduce the need for filter material replacement, ability to provide an optimum environment (medium pH, temperature, and moisture) and attachment surface for microbial populations, capacity to maintain high degradation rates, etc. (Leson and Winer 1991; Srivastava and Majumder 2008, 2015). These properties maintain healthy biomass of microorganisms on the surface of the filter media for the better completion of the biofilters.

For the attachment and colonization on the surface of the filter media, these microorganisms use transportation, preliminary adhesion, firm attachment, and colonization. Microorganisms use transportation, preliminary adhesion, firm attachment, and establishment to attach and colonize on the surface of the filter media (Chaudhary et al. 2003; Kumar et al. 2013). Biological methods that take place in a biofilter are as follows: attachment and growth of microorganisms, decay, and detachment (Srivastava and Majumder 2008; Berenjian et al. 2012). Microorganisms present in the biofilter are mostly aerobic; hence, they required O₂ which is supplied by incoming natural air. Oxygen must be dissolved into the water phase so that it can be easily available to the microorganism growing in the biofilter (Chandran et al. 2020). It has been found that most of the bacteria which form club-shaped

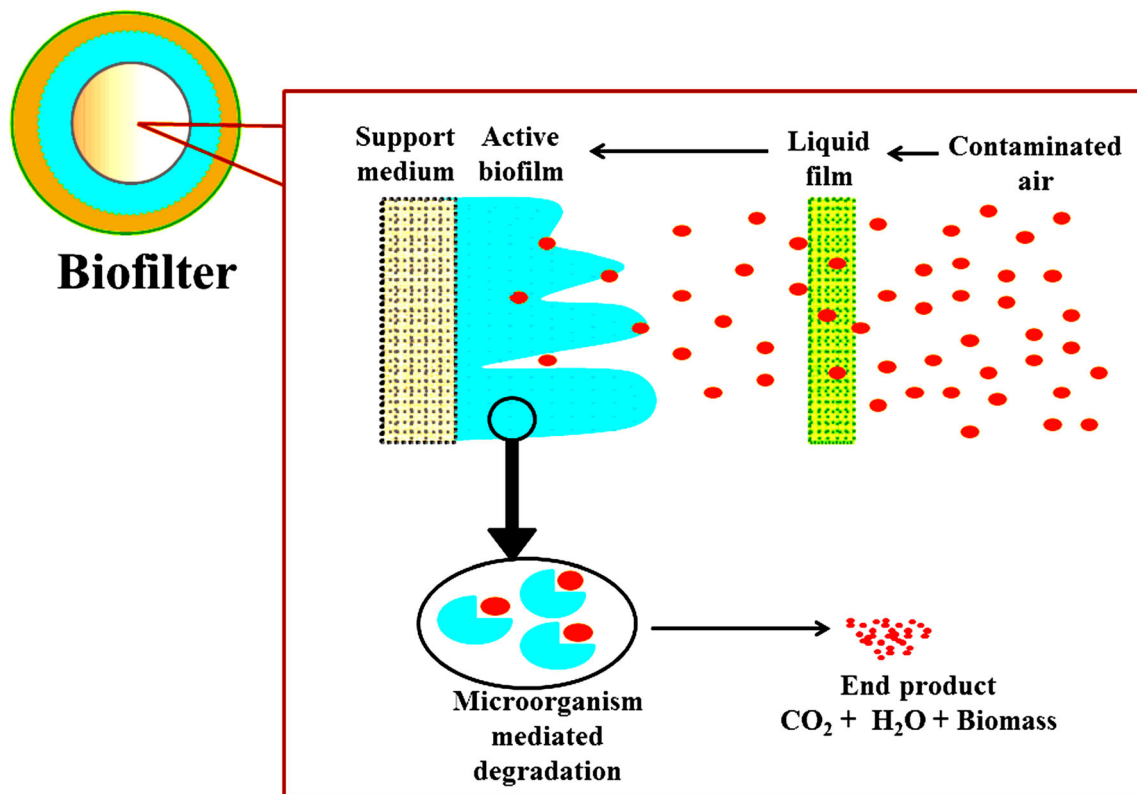


Fig. 2 Phenomena involved in the biofiltration process

(coryneform) endospore and occasionally pseudomonads are present in biofilters. *Streptomyces* spp. some time show their presence in a filter that is actinomycetes. Generally, some fungi such as yeast are not in biofilters. Some filamentous fungi which belong to the Deuteromycetes (*Aspergillus*, *Penicillium*, *Cladosporium*, *Alternaria*, *Fusarium*, *Trichoderma*, and *Botrytis*) and Mucorales (*Mortierella* and *Rhizopus*) commonly show their presence and contribution to the biofiltration process (Kumar et al. 2013).

Biofilter technologies for heavy metal removal

Heavy metals are removed through the biological remediation process occurring in a biofilter. Lethal heavy metals are passed through a moist biologically active film present in the filter medium (Majumder 2015; Gallardo-Rodríguez et al. 2019). Some biological waste material such as water treatment sludge, coconut, rice husk, swine hairs, and tea leaves were also used in biofiltration systems and they showed potential heavy metal removal capacity via ion exchange, surface adsorption, and complexation process (Lim et al. 2015; De Paris et al. 2019). These lethal heavy metals are either oxidized or reduced by microorganisms present in the active layer of the biofilter and produce less soluble and less toxic species. These less-soluble forms of metals formed by the process of precipitation/co-precipitation adsorption, absorption, and

bioremediation (Kumari and Tripathi 2015) on the extracellular protein of the microorganisms and the surface of the adsorbent (Valls and De Lorenzo 2002; Li et al. 2016). Some heavy metals are aerobically transformed into less-toxic species, water, and biomass. An alternative vital route for heavy metals removal from water is the methylation of metals (White et al. 1997; Srivastava and Majumder 2008).

Biofilter technologies for VOC removal

It is an economical and excellently efficient treatment method for VOC removal. It is an effective technique over the conventional methods commonly recovered for air pollution management (Jantschak et al. 2004; Alinezhad et al. 2019). For the treatment of polluted air with VOCs, contaminated off-gas is discharged and sustained from the bottom. Contaminated air passes through the biologically active layer of the filter. Due to the turbulent nature of VOCs, the mass transport takes place through convection and diffusion. The substrate passes to the biofilm by passive diffusion or actively via enzymes (Miller and Allen 2004; Miller et al. 2019). Once pollutants are present in the air, they are diffused into the liquid-phase or absorbed directly on the biofilm. The target pollutants go through aerobic degradation in the biofilm. The cell membrane of the microorganism is conveyed VOCs into the cell, where the reaction occurs and VOCs metabolized. The final

products of the comprehensive biodegradation of pollutants are water and carbon dioxide along with microbial biomass (Adler 2001; Showqi et al. 2016). The oxidations of chlorinated organic and reduced sulfur compounds also generate inorganic acids (Leson and Winer 1991). Different types of VOCs present in the air can react with one another, alter one another's chemical properties, and also affect one another's bioavailability and the rate of biodegradation. Especially in the biofiltration process, these interactions might impact the efficiency of the purification process (Balasubramanian et al. 2012; Miller et al. 2019; Ghasemi et al. 2020). The oxygen availability is also an important factor as well as the retrodiffusion of metabolic by-products such as carbon dioxide, water, and biomass (Vergara-Fernández et al. 2018a, b).

Biotrickling filters methods

Biotrickling filters (BTFs) are structurally quite identical to biofilters. However, they have continuous re-circulation (co-currently or counter-currently) of the liquid medium through the filter instead of pre-humidification in biofilters and also there is synthetic packing material used (Omil 2014). The procedure of gas absorption, gas diffusion into the biofilm, and liquid-phase, regeneration and successive biodegradation take place concomitantly in one procedure equipment. The waste gas being treated is carried through a packed bed in a biotrickling filter either co-currently or counter-currently to the liquid flow. An uninterrupted stream of the recirculating aqueous solution carrying the requisite nutrients for microbial growth is diffused evenly into the packed filter bed (Wambugu et al. 2017). The filtering material should have to resist crushing and constriction of biotrickling filter packing, and also it has to enable the gas and liquid flows through the bed to promote the growth of the microbes. Generally, the packing media are made up of granular activated carbon, ceramics, rocks, plastics, and resins (Mudliar et al. 2010).

During the development of biofilm, microorganisms are inoculated on the surface of the synthetic bed or organic packing material where microbes occur in the liquid phase and also grow primarily on the synthetic bed. An aqueous film surrounded the biofilm where contaminants are primarily absorbed by aqueous film then degraded by the biofilm as biotrickling filter comprises a liquid flow phase; it allows us to dispose of the by-products of decomposition, control pH, and nutrient concentrations. Nevertheless, biotrickling filters are great for the treatment of easily water-soluble pollutants, yet it is inappropriate for less-soluble pollutants (Omil 2014). The interaction between the microbes and the contaminants takes place after the dispersion of the contaminants in the liquid phase. Therefore, the liquid phase flow rate and the recycling rate are known to be acute parameters for biotrickling filter procedure.

The biotrickling filter has the benefit of having the capacity to react with acidic degradation entities of volatile organic compounds, acidic or alkaline compounds, and acidic odorous gases; small operating and capital costs; and lower pressure drop throughout continuous operation (Lebrero et al. 2012). On account of this, when combining this advantage with its cost-efficacy, biotrickling filter technique could be an excellent choice to manage VOCs and odor emissions from various industrial operations (Zehraoui et al. 2012). In spite of all, there are also some constraints with BTF methods such as complex in construction and operation, plenty of biomass accumulation, and low mass-transfer rate that may impact the contaminant elimination efficiency of BTFs. The design, trickling recycling rate to guarantee improved mass transfer, nutrient enrichment, packing materials, operating conditions, and biodegradation of pollutants by microorganisms play a crucial role during biotrickling filter operation (Prachuabmorn and Noppaporn 2010). The degree of degradation of pollutants by microorganisms may vary according to a particular pollutant to be treated. As distinct microbes have their own pros and cons, it becomes crucial to select an appropriate microbial consortium (Wu et al. 2018). The biofilms in biotrickling filters are generally composed of a high amount of bacteria than fungi; thus, most of the researchers have been the focus on the analysis of bacterial communities for their potential (Zhao et al. 2014). Bacterial members of the genera *Pseudomonas*, *Bacillus*, *Staphylococcus*, and *Rhodococcus* are frequently used in the BTF system. *Pseudomonas* has been recognized as the superior species of the bacterial community in various bioreactors employed to eliminate H₂S and the number of VOCs (Giri et al. 2014; Li et al. 2014; Zheng et al. 2016). *Staphylococcus* has the capacity to reduce nitrate to nitrite whereas *Bacillus* can be found simultaneously under aerobic nitrification and denitrification processes; *Rhodococcus* has the capability to degrade lethal environmental contaminants comprising naphthalene, toluene, herbicides, and other compounds (Baltrėnas et al. 2015). BTF method plays a crucial role in the exclusive treatment of VOCs and odorants that present in large volumes and lesser concentrations (Wu et al. 2018). To the author's knowledge, nearly all of these studies have been focused on the elimination of VOCs (Chen et al. 2010; Zhang et al. 2010; Lebrero et al. 2012; Yang et al. 2013), hydrogen sulfide (H₂S) (Montebello et al. 2013; Chen et al. 2014), and trimethylamine (Schiavon et al. 2016).

In another study, four biotrickling filters have been packed with polyurethane foam to determine the reaction amid four aromatic compounds (xylene, toluene, benzene, and styrene). The 90% removal efficiency is reported for distinct toluene, styrene, and xylene (Treesubsumtom and Thiravetyan 2018). The complete elimination capabilities for binary, ternary, and quaternary gases considerably lowered largely in all biotrickling filters. Almost all samples were predominated

either by the genus *Achromobacter* or *Burkholderia*. Samples of biotrickling filters treating single and binary gases were dominated by the genus *Achromobacter* with little *Burkholderia* inside. The rest of the samples drawn from biotrickling filters treating ternary and quaternary gases were dominated by the genus *Burkholderia* with little *Achromobacter* detected inside. These genera were involved in the breakdown of the benzene series in biotrickling filters (Liao et al. 2018). Elimination performance for trichloroethylene and H₂S was estimated to be in the range of 50–90% and 95–98%, respectively (Vikrant et al. 2017).

Biofilm

Biofilm is a group of microbes (protozoa, fungi, algae, bacteria) or biological film or a thin film of a viscous, gelatinous complex in which microbes adhere themselves on the surface of the packing with the help of several forces like electrostatic properties, covalent bond formation, and hydrophobic forces. The strength and the combination of forces are relying on diverse environmental factors that are oxygen supply, gas flow rate, type of microorganisms' species and their surface characteristics, nutrient availability, and pollutant concentration (Kumar et al. 2011; Gafri et al. 2019). The formation of biofilm might take certain days to months depending upon the microbe's concentration. There are three major biological methods that take place in the biofiltration systems; those are (1) adherence of microbes on the surface, (2) growth of microbes, and (3) decomposition and detachment of microbes. In a biofilm, organics or food to the microbes is provided by the bulk and substrate transport methods as the microbes have adhered to the surface. The substrate is metabolized after diffusion onto the outer facet of the biofilm. There are mainly three attributes that affect the rate of substrate utilization within a biofilm which are (1) transport of substrate mass to the biofilm, (2) substrate distribution over the biofilm, and (3) utilization kinetics of the biofilm (Durgananda et al. 2003). Microbes attached to the biofilm decompose the organic compounds in liquid biofilm, where water-soluble contaminants dispersed into the biofilm. Microbes form slim layers upon un-rough surfaces; each treatment process has a standard thickness of biofilm. Biofilm thickness ranges from 10 to 10,000 mm; usually, it averages around 1000 mm (Fulazzaky et al. 2014). The thickness of the biofilm increases during the biofiltration operation and above particular thickness, it is called active thickness (Malakar et al. 2017). Nevertheless, the activity of the biofilm enhances with the thickness up to a level termed the "active thicknesses" but the entire biofilm is not active. Above this level, the distribution of nutrients becomes a limiting factor to differentiate an "active" biofilm from an "inactive" biofilm (Kumar et al. 2011).

Aggregation of microorganisms in sessile or non-submersed (activated sludge) atmosphere has a great benefit of enhanced resistance towards exposure to lethal chemicals contaminants in higher concentration, atmospheric stress environments like change in temperature, pH, and salt concentration and change in environmental conditions viz. nutrients and predation. Hence, these characteristics and stiffness of biofilm can be exploited to come up with approaches for bioremediation of organic contaminants (Edwards and Kjellerup 2013). A multi-species association of *Pseudomonas* strains amplifies the biofilm development in comparison with pure cultures. During the biodegradation process, it also performs 100% and 78% elimination of phenanthrene and pyrene, respectively, after 7 days of progression (Isaac et al. 2017). Recently, in the occurrence of secondary carbon sources, for instance, glucose, starch, sucrose, and L-arginine, biodegradation of naphthalene was made. It was deduced that the *Pseudomonas putida* KD9 strain improves the biofilm development in the presence of sucrose (0.5% wt) during the naphthalene degradation. It has been observed that sucrose serves as a biostimulating agent for the breakdown of naphthalene (Dutta et al. 2018). In another study, employing a moving bed biofilm reactor biodegradation of polychlorinated biphenyls (PCBs) carrying wastewater was accomplished with the elimination efficiency of PCB77 in aerobic and anaerobic portions were 73% and 84.4%, respectively (Dong et al. 2015). The integrated approach of activated sludge and biofilm method along with a moving bed biofilm reactor decomposes the pharmaceuticals adequately (Gaur et al. 2018).

Consistent with these observations, it has been observed that Zn was uniformly distributed across thin biofilms (approx. 12 µm) but passes through less than 20 µm into thick biofilms (approx. 350 µm) (Hu et al. 2005). According to previous studies, the interactions amid biofilm and heavy metals have chiefly concentrated on the absorption of heavy metals. Many investigators have investigated the use of biofilms to eliminate heavy metals from contaminated water owing to the capability of biofilms in the elimination of metals from bulk liquid (Labrenz et al. 2000). Electron microscopic study of *Pseudomonas aeruginosa* biofilm has been reported that it has the capability to separate heavy metals while Hg-declining *Pseudomonas putida* biofilms were observed to store elemental Hg on the extrinsic of the biofilms. *Burkholderia cepacia* biofilms were also found to accumulate Pb (Meliani and Bensoltane 2016).

The bacteria of the phylum Chloroflexi (*Dehalococcoides*, *Dehalobium chloro-coercia*) can be exploited for the biodegradation of halogenated hydrocarbons due to the presence of organohalide respiration (Löffler et al. 2013). Moreover, bacteria of the genus *Dehalococcoides* have been reported to show the reductive dechlorination of trichloroethene in a biofilm reactor (Chung and Rittmann 2008). Selenium (Se) has

been accumulated in biofilms on nutrients incorporating tubes (Williams et al. 2013). As and Fe (iron) at gold-quartz mining sites have been passively oxidized by biofilms (Edwards and Kjellerup 2013). The packed bed bioreactor (PBBR) was chiefly devised for the successful removal of organic and nutrient employing a biofilm complex. The maximum adsorption potential of Cu, Ni, and Cd ions onto activated sludge is in the following order $Cu > Ni > Cd$, which was documented by Ong et al. (2013); that is, Cu and Zn are more easily removed than Cd and Ni ($Cu > Zn > Ni > Cd$). It is therefore deduced that PBBR is capable of removing heavy metal contamination from industrial wastewater outflows (Azizi et al. 2016).

The mixed-species biofilm has more tolerance capacity against disinfectants, antibiotics, heavy metals, etc., than the single-species biofilm (Golby et al. 2014; Jahid and Ha 2014). To the writers' knowledge (Golby et al. 2014), one of the first observations on the impact of heavy metals on mixed bacterial biofilm was published. Besides bacteria, the yeast biofilms have also been studied for the efficient elimination of heavy metal (Basak et al. 2014). The mixed-species biofilms of *Rhodotorula mucilaginosa* and *Escherichia coli* show efficient elimination of heavy metals like Cd, Zn, Ni, Cu, Pb, and Hg from polluted environments. Basak et al. (2014) have observed a decrease in the concentrations of the heavy metal ions in the substrate during the removal of Zn using the *Candida rugosa* and *Cryptococcus laurentii* biofilms (Grujić et al. 2017).

Bioscrubber

Bioscrubber's basic design comprises two subunits; one is a bioreactor unit and another is an absorption unit (Rene et al. 2012). In the absorption unit, contaminated gases are converted to the gaseous phase to the dispersed aqueous phase (aerosol). In a column which contains a packing material, gas and liquid phases flow cross currently. In the bioreactor present in the aqueous phase, contaminants are aerobically degraded by microorganisms (Schlegelmilch et al. 2005). The addition of inert material (such as ceramic) provides a developed transfer surface area amid the gas phase (VOCs) and the liquid phase (Van Groenestijn and Hesselink 1993). The separated contaminated liquid phase is pumped to an aerated bioreactor for agitation and the washed gas is liberated from the column. This reactor unit contains nutrient solution (media) and the appropriate microorganism suspended in the liquid phase and gets nutrients from the media which is essential for their growth and maintenance. Presently, most of the bioscrubbers are based on the stimulated sludge principle (Delhoménie and Heitz 2005). In some bioscrubber methods, a specific type of degrading microbial strain is introduced into the bioreactor. The residence time for management differs from 20 to 40 days depending on the nature and absorption of VOCs. After complete biodegradation of the pollutants, the medium of

bioreactor is filtered and a portion of the waste liquid solution can be recycled through this process again while a part of the sediment biomass re-introduced into the system.

Some research studies illustrate that the accumulation of silicon oil, phthalate to the liquid solution, can expressively improve the removal of less soluble pollutants because they act as emulsifying agents who facilitate the VOC bulk transmission from gas to liquid phases (Mortgat 2001; Artiga et al. 2005; Delhoménie and Heitz 2005). There are some advantages with this process as follows: they have enough control of the biological parameters such as pH, nutrient level, and also good operational stability; relatively lower pressure drops (Rho 2000); and no need for large spaces for their installation. There are also some major limitations with bioscrubbers as follows: adapted to treat VOC which have low Henry coefficients (< 0.01) such as alcohols and ketones (Le Cloirec and Humeau 2013) and at low concentrations ($< 5 \text{ g/m}^3$) (Frederickson et al. 2013). It means a narrow band of VOCs is treatable; mass transfer areas available for gas/liquid are quiet less ($< 300 \text{ m}^{-1}$); excess sludge generation; and required two units for the treatment process (Mortgat 2001; Berenjian et al. 2012). There are several forms of bioscrubbers designs available to boost the performance of the VOC treatment are as follows: sorptive-slurry bioscrubbers, anoxic bioscrubbers, two-liquid phase bioscrubbers, airlift bioscrubbers, and spray column bioscrubbers (Mudliar et al. 2010).

Biosorption processes

The biosorption process is based on the metal-binding capabilities of biological materials. The sorption process converts metal ions from soluble form to the solid phase which includes a group of adsorption and precipitation reactions. Generally, this process uses microorganisms to recover or remove heavy metals from the aqueous phase. It is a cost-effective, eco-friendly, efficient new emerging technology. This process uses the physicochemical interactions between metal ions and cellular compounds of biological species. This interaction results in the uptake of heavy metals (Ahalya et al. 2003). It is a metabolic independent passive process. Metals are primarily bound to the functional groups present on the cell wall of the biological species. These functional groups may comprise carboxyl, phosphate, hydroxyl, and amine groups (Sardrood et al. 2013). Some mechanisms which contribute to this process are as follows: ion exchange, adsorption, electrostatic interaction, complexation, and precipitation (Loukidou et al. 2004; Acheampong et al. 2010; Abbas et al. 2014; Li and Yu 2014). Particularly, the cell wall organization of some specific fungi, bacteria, and algae was found responsible for this method. The biosorption method can remove materials (metal ions) by using inactive, non-living or living biomass (materials of biological origin) owing to attractive forces existing amid absorbent and metal (Volesky and Holan 1995; Gadd 2010).

Microbial cells have the ability to bind up dissolved metals but the sorption affinities and capacities may differ from microbial to non-microbial biomass and between the microorganisms themselves as well (Acheampong et al. 2010). The biosorption process comprises two phases: One is a solid phase which may include sorbent/biosorbent/biomass/biological material and another one is an aqueous phase which may include solvent (water), which contain dissolved metal ions to be sorbed. Microorganisms are great sources of biosorbent; for example, algae, bacteria, fungi, and yeast (Kumar et al. 2014; Mustapha and Halimoon 2015; Ayangbenro and Babalola 2017) possess the metal-sequestering property and by using this method, heavy metal ion concentration can be reduced from ppm to ppb level in aqueous solution. It can efficiently eliminate dissolved metal ions from the contaminant solutions; thus, it is an ideal method for the wastewater treatment as well (Chigondo et al. 2013).

Biosorption processes for metal removal using living cells

This process comprises two important steps. In the first step, the metal ions are adsorbed by biological systems (biosorbent) on their surface. The bioadsorption process takes place owing to the presence of interaction amid heavy metal and functional groups of the biological systems. Second, metal ions are absorbed intracellularly because of active biosorption through the cell membrane and metal enters into the cells. This is a metabolism-dependent process which is responsible for metal transportation and accumulation or deposition (Abbas et al. 2014).

Biosorption processes for metal removal using dead cells

Numerous removal studies were performed on dead cells as they are unaffected by metal toxicity, unlike live cells (Chen et al. 2020). In this process, metal is removed mainly by passive mode with non-living cells which means it is independent of the energy process (Mohapatra et al. 2019). It is a simple physicochemical process that shows resemblance with some conventional methods such as adsorption or ion exchange. This process is accomplished by the different chemical groups present on the surface of the dead cells. The biosorption process initiates with metal uptake on the cell surface (extracellular binding) and occurs by the processes that include physical adsorption, ion exchange, van der Waals forces, complexation, or inorganic microprecipitation (Gin et al. 2002; Kotrba 2011; Abbas et al. 2014). Some reported metal-binding chemical groups belong to carboxylates, amines, and imidazoles family with cellulose (algal cell wall), chitin, and chitosan (fungal cell wall) and teichoic acids or lipopolysaccharides (gram-positive and gram-negative bacteria, respectively) (Chen et al. 2019; Hansda et al. 2016). The biosorption capability of

dead cells may be less, equivalent, or greater than that of living cells (Gallardo-Rodríguez et al. 2019). Therefore, it is a metabolism-independent process. It rapidly removes metal by anyone or a group of metal-binding mechanisms, for instance, ion exchange, adsorption, physical, and complexation (Gin et al. 2002; Kotrba 2011; Abbas et al. 2014).

Phytoremediation technologies

Plants have been used to eliminate pollutants from the surrounding dates for 300 years. Baumann (1885) first ever reported bioaccumulation of heavy metals by the plant species *Thlaspi caerulescens* and *Viola calaminaria* at the end of the nineteenth century. Later, the term “phytoremediation” was introduced in the earlier 1990s (Shackira and Puthur 2019). Phytoremediation employed plants and cooperating soil microbes. Moreover, in situ or ex situ methods are used to eliminate or reduce pollutants in distinct environmental forms like air, soil, and water. In which in situ phytoremediation is most frequently used as it can manage the interference of soil and atmosphere and it also helps in lowering the expansion of pollution through airborne and waterborne wastes (Kumar and Gunasundari 2018). Phytoremediation can be used to deal with nearly all types of pollutants, radionuclides (Dubchak and Bondar 2019), toxic metals (Xiao et al. 2019), and recalcitrant organic contaminants (Cameselle and Gouveia 2019) such as organophosphate insecticides, sulfonated aromatics, nitroaromatics and explosives, chlorinated pesticides, polynuclear aromatic hydrocarbons, polychlorinated biphenyls petroleum hydrocarbons, phenolics, and chlorinated solvents (Wu et al. 2012). Phytoremediation is a rising branch of biofiltration science that utilizes several plants for the degradation, extraction, accumulation, or immobilization pollutants from soil and water. Later, this technique has been recognized as an innovative, cost-effective unconventional treatment method. Figure 3 provides the schematic representation of the phytoremediation of heavy metals and VOCs. This method is usually depending exclusively on the use of microbes which is complementary to conventional bioremediation methods (Abhilash et al. 2009; Thijs et al. 2017). Plants have been efficiently employed not only for soil remediation but also to treat polluted air, industrial, and municipal wastewaters (Zurita et al. 2009).

Phytoremediation for heavy metals removal

Plant species that have a capacity to absorb and store the pollutants in the plant tissues like roots, aerial parts, or shoots can be employed for the phytoremediation of metals in soils. During the process of phytoremediation, it is important to use hyperaccumulator plant species that has the capability to accumulate an elevated amount of concentrations of heavy metals a hundred times higher than non-hyperaccumulator

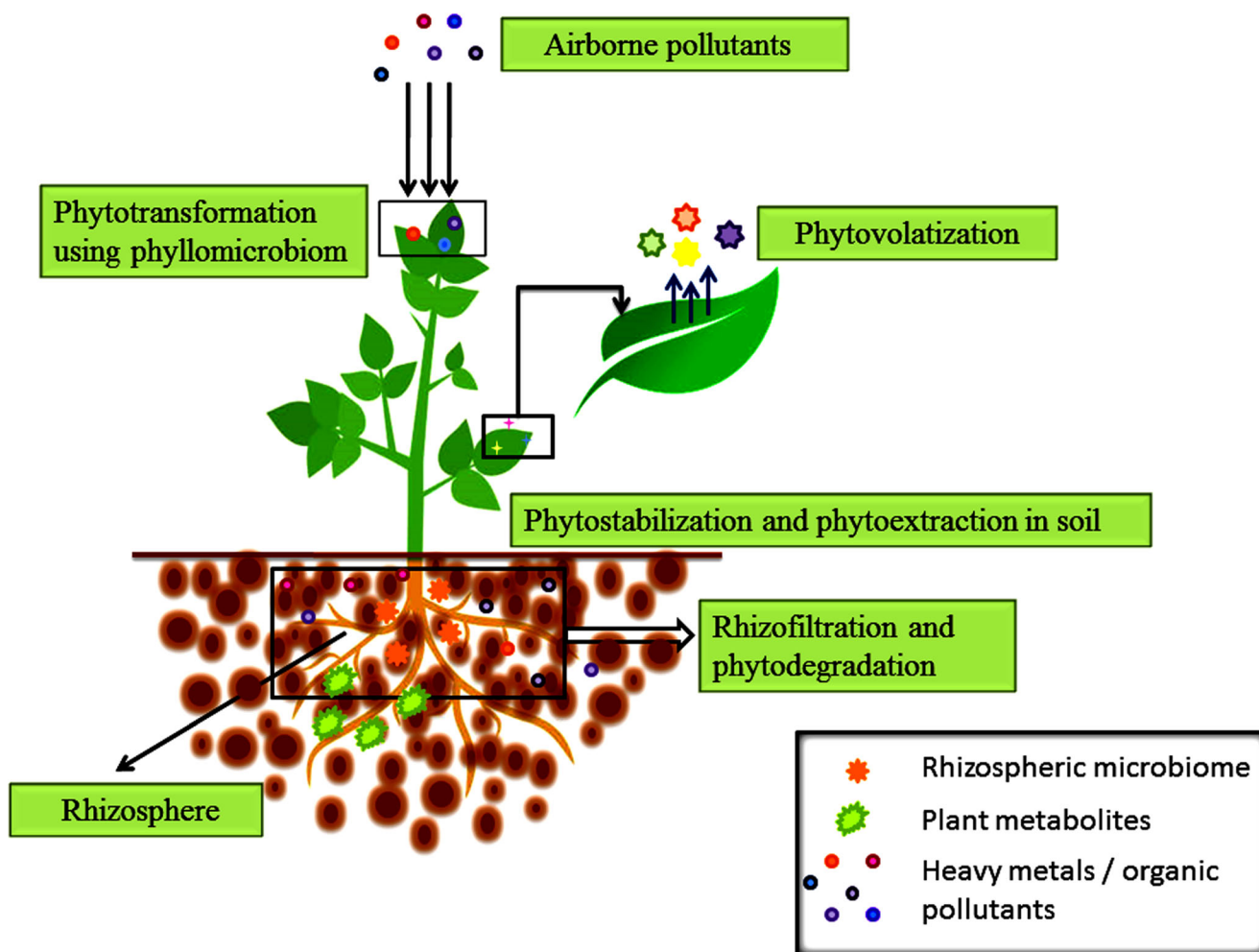


Fig. 3 Schematic representation of phytoremediation of heavy metals and VOCs

plants with no considerable adverse effect on their growth and development. Hyperaccumulators have greatly pronounced metal removal mechanisms and, occasionally, higher internal needs for specific metals, and mobilized and solubilized heavy metals and converts into low-soluble forms than the non-hyperaccumulating plant species (Rascio and Navari-Izzo 2011; Hesami et al. 2018).

Nevertheless, their efficacy also relies upon the kind of heavy metal. Different metals have different types of activity and mobility within plant tissues such as Cd, Ni, and Zn which are more readily translocated to the aboveground plant parts, whereby Pb, Cr, and Cu immobilized and translocated in the root system (Pulford and Watson 2003; Xu et al. 2019). After uptake of metals by the plant, metals commonly accumulated in vacuoles where they are attached to organic acids or tend to bind to cell wall constituents or to sulfur ligands in the cytosol (Callahan et al. 2006). In addition to this, it may also form precipitates with carbonate, phosphate, or sulfate and accumulates within intracellular or extracellular spaces of cells (Chirakkara et al. 2016).

Phytoremediation of VOC removal

Phytoremediation of volatile organic compounds is principal element of the environment, chiefly as the precursor of ground-level ozone and secondary organic aerosols. Hydroxyl radicals serve as the superior VOC sink; at the same time, volatile organic compounds may also nucleate to very fine particulates through the moderate procedure of oxidation (Singh et al. 2017). Various plants liberate VOCs into the environment; hence, only those plants should be selected for phytoremediation, which have low VOC-emitting capacity. Various plant species have been identified to have the capability to eliminate VOCs from the air (Teiri et al. 2018). The elimination of VOCs comprises the incorporation of direct and indirect methods, which may be obtained either by growing medium, leaves, shoot system, root system, or by microbes existing in the rhizosphere. Both stomata and cuticles play a vital role during the removal process of VOC. The stomatal uptake of VOCs is predominantly depended upon types of VOCs. Benzene passes readily through the cuticle whereas formaldehyde passes more readily through stomatal apertures (Cruz et al.

2014). After entering into the plant body, VOCs undergo breakdown, accumulation, or excretion either at the place of penetration or after translocation to vacuoles of the cell. Certain volatile organic compounds, like xylene, toluene, and formaldehyde, are occupied by plants leaves and are eventually transferred to the root system where microbes are capable to digest them. Ornamental plants are reported as an efficient system for decomposition of volatile organic compounds and as well as lower the volatile organic compound concentration for indoor atmosphere (Kim et al. 2010). Volatile organic compounds are one of the most indoor air pollutants and there are so many predominant sources within the indoor environment with high concentrations than outdoor conditions. The most often volatile organic compounds are ethyl-benzene, formaldehyde, naphthalene, benzene, toluene, xylene, and polycyclic aromatic hydrocarbons (PAHs) including trichloroethylene and benzo[α]pyrene. Hanging pot plants also help in lowering the carbonyl concentration in the indoor atmosphere (Pegas et al. 2012). Other researches have been recommended that *Spathiphyllum* and *Dracaena deremensis* successfully eliminate toluene, even though *Dracaena marginata*, *D. deremensis*, *Spathiphyllum floribundum* “Petite”, *S. floribundum* “Sensation,” and *Schefflera actinophylla* “Amate” were identified to be beneficial for the elimination of benzene (Orwell et al. 2006). Moreover, *D. marginata*, *S. floribundum*, and *D. deremensis* have been employed to eliminate xylene and ethylbenzene from indoor air (Wood et al. 2006). In addition to this, the greater removal capacity has been investigated in *Chrysanthemum morifolium* and *Calathea rotundifolia* cv. “Fasciata” out of 94 studied potted plant species for the removal of benzene and toluene (Yang and Liu 2011). Some plants have been identified to lower the formaldehyde concentration in the range of 47–70% than the controlled atmosphere. Some plants such as *Chlorophytum comosum* L. consume formaldehyde as a carbon and energy resource. Industrial, traffic, and commercial activities are the prime sources of outdoor pollutants as they liberate the number of air contaminants. Later these contaminants, either through primary conversion or gas-to-particle conversion, might react with plant bodies and their microbiomes where they have the capabilities to alter the future of pollutants (Agarwal et al. 2019). There are distinct methods used for the phytoremediation of heavy metal and VOCs like phytoextraction, phytodegradation, rhizofiltration, phytotransformation, phytovolatilization, and phytostabilization.

Phytoaccumulation or phytoextraction

Phytoextraction is the process acquired by the plants to store contaminants into their aboveground plant parts like shoots or leaves and the root system. It comprises the absorption, translocation, and accumulation of heavy metals by the

aboveground and belowground plant parts along with other nutrients. In this direction, plants are capable of storage of heavy metals. These plants are grown in contaminated places where heavy metal accumulated aboveground plant parts are collected for the elimination of traces of the heavy metal. Phytoextraction is also defined as phytoabsorption, phytoaccumulation, or phytosequestration. Contradictory to the degradation procedures, this method generates a chunk of plants and pollutants that can be transferred for discarding or reprocessing (Sharma 2012). The benefits of the phytoextraction are high tolerance to heavy metals, excessive biomass, rapid growth rate, and great root system (Suman et al. 2018). There are two methods that have been recognized for the phytoextraction based upon the plant properties. The first method is the utilization of natural hyperaccumulator plants that have a greater capacity to store heavy metals. These hyperaccumulators are successfully accumulating many of the heavy metal that is 10–500 times higher than ordinary plants (Chibuikwe and Obiora 2014). However, in the second method, chelates or soil amendments are used with the high-biomass plant to boost the capability to store heavy metals from the atmosphere. Few plants have the capabilities for the storage of more heavy metals than one in the same respective plant such as the hyperaccumulation of Zn and Cd by *Sedum alfredii*. Frequently available plants for the hyperaccumulation of metals are *Haumaniastrum robertii* (Co), *Aeollanthus subacaulis* (Cu), *Lecythis ollaria* (Se), *Agrostis tenuis* (Pd), *Streptanthus polygaloides* (Ni), *Maytenus bureaviana* (Mn), *Pteris vittata* (As), *Thlaspitatreuse*, *Thlaspi caerulescens* (Zn), etc. Nonetheless, the phytoextraction method also has some constraints such as the utilization of hyperaccumulators which include the possibility of polluting the food chain. In spite of such limitation, the hyperaccumulators of metal of the family *Brassicaceae* have more amounts of thiocyanates that make them unappetizing to animals, and therefore, these plants can reduce the chances of availability of metal ions in the food chain.

Phytostabilization

This process is relying upon the immobilization of metal ions in the soil by the plant rather than their degradation. Leachable components are uptaken by the plants. These components are attached to the plant structures so that they build a solid mass of plants that taper of the bioavailability of heavy metals in the surroundings through erosion and infiltration from which the pollutants will not turn back into the environment. Phytostabilization of heavy metals using plants can be accomplished through precipitation, absorption, and metal valence reduction. This phytostabilization is more advantageous for the rapid immobilization of heavy metal from soil and groundwater, although the entire elimination of heavy metals from the environment is not unachievable (Kumar and Gunasundari

2018). Phytostabilization is an unsophisticated, economical process that employs plants to restrict the movement of toxic metal contaminants within the root tissues and in the rhizosphere. In spite of the physical elimination of the pollutants, this process involves the deactivation and immobilization of the contaminants, consequently ceasing the further transfer of contaminants to the same food chain. The microbial community that has been associated with roots and rhizospheric regions of plants engaged in performing many functions such as recycling of nutrients, detoxification of toxic pollutants, and sustainment of soil. It has been studied that bacteria namely *Sphingomonas macrogoltabidus*, *Microbacterium liquefaciens*, *Microbacterium arabinogalactanolyticum*, and *Alyssum murale* upon incorporation into the soil promote the phytostabilization by reducing the pH of the soil, thereby remarkably enhancing the phytoavailability of heavy metals together with Ni too (Shackira and Puthur 2019). Phytostabilization is a highly appropriate approach for the immobilization of the noxious pollutants in a heavily contaminated site. It abruptly stops the movement of potentially harmful pollutants by acting as a strong barrier for the filtration of water within the soil; thus, contaminants remain in the soil (Ali et al. 2020). It has been proved that it is truly efficient in the rapid immobilization for the preservation of ground and surface waters; also it is a pertinent process for the removal of Cd, Cu, As, Zn, and Cr (Ekta and Modi 2018).

Rhizodegradation: phytoremediation using root system

This method comprises the degradation of pollutants. The specific activity in the rhizospheric region is responsible for the breakdown of contaminants which is present in the plant- and microbe-derived proteins and enzymes. Rhizodegradation is an outcome of the symbiotic association between plants and microbes. Rhizofiltration is a type of phytoremediation, which is used to purify extracted water with a lower concentration of pollutants by using root system. It can also be termed as phytostimulation as degradation of pollutants being stimulated by rhizospheric microflora (Lee et al. 2020). This method can be used for other metals like Cd, Cu, Pb, Zn, Ni, and Cr that are voluminously accumulated into the roots. Sunflowers, rye, tobacco, corn, Indian mustard, spinach, and pulse are broadly used for the elimination of Pb from water or soil (Luciano et al. 2013). Basically, both terrestrial and aquatic plants engaged in the rhizofiltration for in situ or ex situ purposes. The bigger disadvantage of this method is the pH settlement required for a regular time span (Pinto et al. 2016).

Phytovolatilization

In this method, plants absorbed water and carried organic contaminants from the soil. These organic contaminants are transformed into volatile components after being dispersed

into the aboveground plant parts. Hence, the degraded volatile compounds were released into the environment via aerial parts of the plant (Lee et al. 2020). Later, these organic pollutants are transformed into volatile skeletons and are liberated into the air through their leaves. The phytovolatilization can be employed for the elimination of organic contaminants as well as some heavy metals such as Se and Hg. This method could not completely eliminate contaminants. It only transformed the lethal form of contaminants into the least toxic form. The limitation of this method is the productions of additional products that may re-accumulate into the water bodies. For this process, genetically modified plants are predominantly used for the absorption of pollutants; particularly, *Brassica juncea* and *Brassica napus* have been employed for the phytovolatilization of Hg and Se from soil (Chibuike and Obiora 2014). Since, Hg and Se are considered most convenient to be remediated through phytovolatilization, it has been reported that Indian mustard and canola are very helpful in the phytovolatilization of Se (Ali et al. 2020). Mercury is the foremost metal contaminant that has been phytoremediated by using phytovolatilization process together with trichloroethene and volatile inorganic chemicals like Se and As (Bisht et al. 2020). This method extremely relies upon the physical state of the pollutant itself and the availability of the contaminant in rhizosphere of the plant to be absorbed by the roots (Gupta et al. 2020).

Conclusions

This review gives a thumbnail sketch of the biological-based methods for the elimination of VOCs and heavy metal from water wastes. Throughout the last few decenniums, tremendous heed has been paid to control the environmental pollution that is attributed to the excessive amounts of lethal heavy metals and VOCs. Here, we emphasize that upcoming endeavors are required to explicate methods for the decontamination of heavy metals and volatile organic compounds from the environment which is becoming very much necessary for the maintenance of a healthy and safe environment. Nevertheless, physical-chemical methods are often used for the same purposes; biological processes namely biosorption/bioaccumulation, biofilters, phytoremediation, and bioscrubber seem to be a promising alternative method from the perspective of pollutant removal efficiency, costs, technology requirement, environmental impacts, and energy efficiency. Although, all the above methods can be employed for the management of heavy metal and VOCs, it is pivotal to declare that the adoption of the most suitable treatment techniques relies upon the initial contaminant concentration, capital investment and operational cost, the component of the wastewater, environmental impact, utility of the treated water, etc.

Acknowledgments The author, MM, is thankful to Mohanlal Sukhadia University, Udaipur, for providing the necessary facilities during the course of study.

Author's contributions MM, PS, and GY conceived the idea of the review, provided the general concept and inputs for each specific section, and drafted part of the manuscript. MM, PS, and GY wrote the review after collecting the literature. MM edited, compiled, and finalized the draft. Finally, all the authors read and approved it for publication.

Funding This study was supported by Startup Research Grant (UGC Faculty Research Promotion Scheme; FRPS) and sustained by Mohanlal Sukhadia University, Udaipur, Rajasthan, India.

Data availability The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Compliance with ethical standards

Competing interest The authors declare that there is no conflict of interest.

References

- Abbas SH, Ismail IM, Mostafa TM, Sulaymon AH (2014) Biosorption of heavy metals: a review. *J Chem Sci Technol* 3:74–102
- Abd-El salam IS (2011) Factorial design for some parameters affecting on chromium III uptake by *Saccharomyces cerevisiae*. *Int J Appl Biol Pharm* 2:33–40
- Abhilash PC, Pandey VC, Srivastava P, Rakesh PS, Chandran S, Singh N, Thomas AP (2009) Phytofiltration of cadmium from water by *Limncharis flava* (L.) Buchenau grown in free-floating culture system. *J Hazard Mater* 170:791–797
- Abuhamed T, Bayraktar E, Mehmetoğlu T, Mehmetoğlu Ü (2004) Kinetics model for growth of *Pseudomonas putida* F1 during benzene, toluene and phenol biodegradation. *Process Biochem* 39:983–988
- Acheampong MA, Meulepas RJ, Lens PN (2010) Removal of heavy metals and cyanide from gold mine wastewater. *J Chem Technol Biotechnol* 85:590–613
- Acosta Rodríguez I, Martínez-Juárez VM, Cárdenas-González JF, Moctezuma-Zárate MDG (2013) Biosorption of arsenic (III) from aqueous solutions by modified fungal biomass of *Paecilomyces* sp. *Bioinorg Chem Appl* 2013:376780. <https://doi.org/10.1155/2013/376780>
- Adler SF (2001) Biofiltration—a primer. *Chem Eng Prog* 97:33–41
- Agarwal P, Sarkar M, Chakraborty B, Banerjee T (2019) Phytoremediation of air pollutants: prospects and challenges. In: *Phytomanagement of Polluted Sites*. Elsevier, pp 221–241
- Ahalya N, Ramachandra TV, Kanamadi RD (2003) Biosorption of heavy metals. *Res J Chem Environ* 7:71–79
- Ahmady-Asbchin S, Andres Y, Gérente C, Le Cloirec P (2008) Biosorption of Cu (II) from aqueous solution by *Fucus serratus*: surface characterization and sorption mechanisms. *Bioresour Technol* 99:6150–6155
- Aizpuru A, Dunat B, Christen P, Auria R, García-Peña I, Revah S (2005) Fungal biofiltration of toluene on ceramic rings. *J Environ Eng* 131: 396–402
- Akinci G, Guven DE (2011) Bioleaching of heavy metals contaminated sediment by pure and mixed cultures of *Acidithiobacillus* spp. *Desalination* 268:221–226
- Akpor OB, Ohiobor GO, Olaolu DT (2014) Heavy metal pollutants in wastewater effluents: sources, effects and remediation. *Adv Biosci Bioeng* 2:37–43
- Ali S, Abbas Z, Rizwan M, Zaheer IE, Yavaş İ, Ünay A, Abdel-Daim MM, Bin-Jumah M, Hasanuzzaman M, Kalderis D (2020) Application of floating aquatic plants in phytoremediation of heavy metals polluted water: a review. *Sustainability* 12(5):1927. <https://doi.org/10.3390/su12051927>
- Alinezhad E, Haghighi M, Rahmani F, Keshizadeh H, Abdi M, Naddafi K (2019) Technical and economic investigation of chemical scrubber and bio-filtration in removal of H₂S and NH₃ from wastewater treatment plant. *J Environ Manag* 241:32–43. <https://doi.org/10.1016/j.jenvman.2019.04.003>
- Alrumman S, Keshk S, El-Kott A (2016) Water pollution: source and treatment. *Am J Environ Eng* 6:88–98
- Amanullah MD, Farooq S, Viswanathan S (2000) Effect of adsorption capacity of the solid support on the performance of a biofilter. In: *Adsorption Science and Technology*. World Scientific, pp 209–213
- Anaemene IA (2012) The use of *Candida* sp. in the biosorption of heavy metals from industrial effluent. *Eur J Exp Biol* 2:484–488
- Arakaki AH, Souza Vandenberghe LPD, Soccol VT, Masaki R, Rosa Filho EFD, Gregório A, Soccol CR (2011) Optimization of biomass production with copper bioaccumulation by yeasts in submerged fermentation. *Braz Arch Biol Technol* 54:1027–1034
- Arnold M, Reittu A, von Wright A, Martikainen PJ, Suihko ML (1997) Bacterial degradation of styrene in waste gases using a peat filter. *Appl Microbiol Biotechnol* 48:738–744
- Artiga P, Oyanedel V, Garrido JM, Mendez R (2005) An innovative biofilm-suspended biomass hybrid membrane bioreactor for wastewater treatment. *Desalination* 179:171–179
- Ayangbenro AS, Babalola OO (2017) A new strategy for heavy metal polluted environments: a review of microbial biosorbents. *Int J Environ Res Public Health* 14:94. <https://doi.org/10.3390/ijerph14010094>
- Azizi S, Kamika I, Tekere M (2016) Evaluation of heavy metal removal from wastewater in a modified packed bed biofilm reactor. *PLoS One* 11(5):e0155462. <https://doi.org/10.1371/journal.pone.0155462>
- Balasubramanian P, Philip L, Bhallamudi SM (2012) Biotrickling filtration of VOC emissions from pharmaceutical industries. *Chem Eng J* 209:102–112. <https://doi.org/10.1016/j.cej.2012.04.020>
- Baltrėnas P, Zagorskis A, Misevičius A (2015) Research into acetone removal from air by biofiltration using a biofilter with straight structure plates. *Biotechnol Biotechnol Equip* 29:404–413
- Barros Júnior LM, Macedo GR, Duarte MML, Silva EP, Lobato AKCL (2003) Biosorption of cadmium using the fungus *Aspergillus niger*. *Braz J Chem Eng* 20:229–239
- Barupal T, Meena M, Sharma K (2019) Inhibitory effects of leaf extract of *Lawsonia inermis* on *Curvularia lunata* and characterization of novel inhibitory compounds by GC–MS analysis. *Biotechnol Rep* 23:e00335. <https://doi.org/10.1016/j.btre.2019.e00335>
- Barupal T, Meena M, Sharma K (2020a) *In vitro* assay of antifungal activity of various elicitors and binders against *Curvularia lunata*. *Food Sci Nutr Technol* 5(1):00206. <https://doi.org/10.23880/fsnt-16000206>
- Barupal T, Meena M, Sharma K (2020b) Effect of different physical factors on *Lawsonia inermis* leaf extracts and their herbal formulations efficacy. *Am J Agric Sci* 7(1):01–07
- Barupal T, Meena M, Sharma K (2020c) A study on preventive effects of *Lawsonia inermis* L. bioformulations against leaf spot disease of maize. *Biocatal Agric Biotechnol* 23:101473. <https://doi.org/10.1016/j.bcab.2019.101473>
- Basak G, Lakshmi V, Chandran P, Das N (2014) Removal of Zn (II) from electroplating effluent using yeast biofilm formed on gravels: batch and column studies. *J Environ Health Sci* 12:1–11. <https://doi.org/10.1186/2052-336X-12-8>

- Baumann A (1885) Das Verhalten von Zinksätzen gegen Pflanzen und im Boden. *Landwirtsch. Vers Statn* 31:1–53
- Berenjian A, Chan N, Malmiri HJ (2012) Volatile organic compounds removal methods: a review. *Am J Biochem Biotechnol* 8:220–229
- Bisht R, Chanyal S, Srivastava RK (2020) A Systematic review on phytoremediation technology: removal of pollutants from waste water and soil. *Int J Res Eng Sci Manage* 3(1):54–59
- Blázquez P, Casas N, Font X, Gabarrell X, Sarrà M, Caminal G, Vicent T (2004) Mechanism of textile metal dye biotransformation by *Trametes versicolor*. *Water Res* 38:2166–2172
- Brinza L, Dring MJ, Gavrilescu M (2007) Marine micro and macro algal species as biosorbents for heavy metals. *Environ Eng Manag J* 6: 237–251
- Bunluesin S, Kruatrachue M, Pokethitiyook P, Upatham S, Lanza GR (2007) Batch and continuous packed column studies of cadmium biosorption by *Hydrilla verticillata* biomass. *J Biosci Bioeng* 103: 509–513
- Callahan DL, Baker AJ, Kolev SD, Wedd AG (2006) Metal ion ligands in hyperaccumulating plants. *J Biol Inorg Chem* 11:2–12
- Cameselle C, Gouveia S (2019) Phytoremediation of mixed contaminated soil enhanced with electric current. *J Hazard Mater* 361:95–102. <https://doi.org/10.1016/j.jhazmat.2018.08.062>
- Carpenter CM, Helbling DE (2017) Removal of micropollutants in biofilters: hydrodynamic effects on biofilm assembly and functioning. *Water Res* 120:211–221. <https://doi.org/10.1016/j.watres.2017.04.071>
- Casiot C, Morin G, Juillot F, Bruneel O, Personné JC, Leblanc M, Elbaz-Poulichet F (2003) Bacterial immobilization and oxidation of arsenic in acid mine drainage (Carnoulès creek, France). *Water Res* 37: 2929–2936
- Cassini F, Scheutz C, Skov BH, Mou Z, Kjeldsen P (2017) Mitigation of methane emissions in a pilot-scale biocover system at the AV Miljø Landfill, Denmark: 1. System design and gas distribution. *Waste Manag* 63:213–225. <https://doi.org/10.1016/j.wasman.2017.01.013>
- Chandran H, Meena M, Barupal T, Sharma K (2020) Plant tissue culture as a perpetual source for production of industrially important bioactive compounds. *Biotechnol Rep* 26:e00450. <https://doi.org/10.1016/j.btre.2020.e00450>
- Chatterjee AK (2002) Introduction to environmental biotechnology. Prentice Hall India (PHI) Learning Pvt Ltd, New Delhi, p 105
- Chaudhary DS, Vigneswaran S, Ngo HH, Shim WG, Moon H (2003) Biofilter in water and wastewater treatment. *Korean J Chem Eng* 20(6):1054–1065
- Chen L, Hoff SJ (2012) A two-stage wood chip-based biofilter system to mitigate odors from a deep-pit swine building. *Appl Eng Agric* 28: 893–901
- Chen JM, Zhu RY, Yang WB, Zhang LL (2010) Treatment of a BTo-X-contaminated gas stream with a biotrickling filter inoculated with microbes bound to a wheat bran/red wood powder/diatomaceous earth carrier. *Bioresour Technol* 101:8067–8073
- Chen Y, Fan Z, Ma L, Yin J, Luo M, Cai W (2014) Performance of three pilot-scale immobilized-cell biotrickling filters for removal of hydrogen sulfide from a contaminated air stream. *Saudi J Biol Sci* 21: 450–456
- Chen SH, Chew YL, Ng SL, Ting ASY (2019) Mechanisms for metal removal established via electron microscopy and spectroscopy: a case study on metal tolerant fungi *Penicillium simplicissimum*. *J Hazard Mater* 362:394–402. <https://doi.org/10.1016/j.jhazmat.2018.08.077>
- Chen SH, Chew YL, Ng SL, Ting ASY (2020) Bioaccumulation and biosorption activities of indoor metal-tolerant *Penicillium simplicissimum* for removal of toxic metals. *Int J Environ Sci Technol* 14:235–242. <https://doi.org/10.1007/s41742-020-00253-6>
- Chhikara S, Hooda A, Rana L, Dhankhar R (2010) Chromium (VI) biosorption by immobilized *Aspergillus niger* in continuous flow system with special reference to FTIR analysis. *J Environ Biol* 31: 561–566
- Chibuikwe GU, Obiora SC (2014) Heavy metal polluted soils: effect on plants and bioremediation methods. *Appl Environ Soil Sci* 2014: 752708. <https://doi.org/10.1155/2014/752708>
- Chigondo F, Nyamunda BC, Sithole SC, Gwatidzo L (2013) Removal of lead (II) and copper (II) ions from aqueous solution by baobab (*Adonsonia digitata*) fruit shells biomass. *IOSR J Appl Chem* 5: 43–50
- Chirakkara RA, Cameselle C, Reddy KR (2016) Assessing the applicability of phytoremediation of soils with mixed organic and heavy metal contaminants. *Rev Environ Sci Biotechnol* 15:299–326
- Christen P, Domenech F, Michelena G, Auria R, Revah S (2002) Biofiltration of volatile ethanol using sugar cane bagasse inoculated with *Candida utilis*. *J Hazard Mater* 89:253–265
- Chung J, Rittmann BE (2008) Simultaneous bio-reduction of trichloroethene, trichloroethane, and chloroform using a hydrogen-based membrane biofilm reactor. *Water Sci Technol* 58:495–501
- Cicoellella A (2008) Volatile organic compounds (VOC): definition, classification and properties. *Rev Mal Respir* 25:155–163
- Cohen Y (2001) Biofiltration – the treatment of fluids by microorganisms immobilized into the filter bedding material: a review. *Bioresour Technol* 77:257–274
- Cohen-Shoel N, Barkay Z, Ilzyer D, Gilath I, Tel-Or E (2002) Biofiltration of toxic elements by *Azolla* biomass. *Water Air Soil Pollut* 135:93–104
- Cruz MD, Christensen JH, Thomsen JD, Müller R (2014) Can ornamental potted plants remove volatile organic compounds from indoor air?—a review. *Environ Sci Pollut Res* 21:13909–13928
- Das N, Charumathi D, Vimala R (2007) Effect of pretreatment on Cd²⁺ biosorption by mycelial biomass of *Pleurotus florida*. *Afr J Biotechnol* 6:2555–2558
- Davis TA, Volesky B, Mucci A (2003) A review of the biochemistry of heavy metal biosorption by brown algae. *Water Res* 37:4311–4330
- De Paris JO, Scapini T, Camargo AF, Venturin B, Dalastra C, Kubeneck S, Czapela F, Preczeski KP, Stefanski FS, Korf EP, Valério A, Luccio MD, Mossi AJ, Fongaro G, Treichel H (2019) Removal of chromium from wastewater by swine hair residues applied as a putative biofilter. *Environ Sci Pollut Res* 26:33014–33022. <https://doi.org/10.1007/s11356-019-06313-5>
- Delhoménie MC, Heitz M (2005) Biofiltration of air: a review. *Crit Rev Biotechnol* 25:53–72
- Deng X, Li QB, Lu YH, Sun DH, Huang YL, Chen XR (2003) Bioaccumulation of nickel from aqueous solutions by genetically engineered *Escherichia coli*. *Water Res* 37:2505–2511
- Diehl SV, Saileela B (2000) Biofiltration of selected monoterpenes found in Southern Yellow Pine Wood emissions. *Forest Prod J* 50:43–65
- Dong B, Chen HY, Yang Y, He QB, Dai XH (2015) Biodegradation of polychlorinated biphenyls using a moving-bed biofilm reactor. *CLEAN – Soil Air Water* 43:1078–1083
- Dorado AD, Baquerizo G, Maestre JP, Gamisans X, Gabriel D, Lafuente J (2008) Modeling of a bacterial and fungal biofilter applied to toluene abatement: kinetic parameters estimation and model validation. *Chem Eng J* 140:52–61
- Dubchak S, Bondar O (2019) Bioremediation and phytoremediation: best approach for rehabilitation of soils for future use. In: Gupta D, Voronina A (eds) Remediation measures for radioactively contaminated areas. Springer, Cham, pp 201–221. https://doi.org/10.1007/978-3-319-73398-2_9
- Dupasquier D, Revah S, Auria R (2002) Biofiltration of methyl tert-butyl ether vapors by cometabolism with pentane: modeling and experimental approach. *Environ Sci Technol* 36:247–253
- Durgananda SC, Saravanamuthu V, Huu-Hao N, Wang GS, Hee M (2003) Biofilter in water and wastewater treatment. *Korean J Chem Eng* 20:1054–1065

- Dursun AY, Uslu G, Cuci Y, Aksu Z (2003) Bioaccumulation of copper (II), lead (II) and chromium (VI) by growing *Aspergillus niger*. *Process Biochem* 38:1647–1651
- Dutta K, Shityakov S, Khalifa I, Mal A, Moulik SP, Panda AK, Ghosh C (2018) Effects of secondary carbon supplement on biofilm-mediated biodegradation of naphthalene by mutated naphthalene 1, 2-dioxygenase encoded by *Pseudomonas putida* strain KD9. *J Hazard Mater* 357:187–197
- Eccles H (1995) Removal of heavy metals from effluent streams—why select a biological process? *Int Biodeterior Biodegradation* 35:5–16
- Edwards SJ, Kjellerup BV (2013) Applications of biofilms in bioremediation and biotransformation of persistent organic pollutants, pharmaceuticals/personal care products, and heavy metals. *Appl Microbiol Biotechnol* 97:9909–9921
- Ekta P, Modi NR (2018) A review of phytoremediation. *J Pharmacogn Phytochem* 7:1485–1489
- El-Kassas HY, El-Taher EM (2009) Optimization of batch process parameters by response surface methodology for mycoremediation of chrome-VI by a chromium resistant strain of marine *Trichoderma viride*. *Am-Eurasian J Agric Environ Sci* 5:676–681
- El-Sayed MT (2013) Removal of lead (II) by *Saccharomyces cerevisiae* AUMC 3875. *Ann Microbiol* 63:1459–1470
- Estévez E, Veiga MC, Kennes C (2005) Biodegradation of toluene by the new fungal isolates *Paecilomyces variotii* and *Exophiala oligosperma*. *J Ind Microbiol Biotechnol* 32:33–37. <https://doi.org/10.1007/s10295-004-0203-0>
- Frederickson J, Boardman CP, Gladding TL, Simpson AE, Howell G, Sgouridis F (2013) Evidence: biofilter performance and operation as related to commercial composting. Environment Agency, Bristol ISBN: 978-1-84911-299-4
- Frutos FJG, Escolano O, García S, Babín M, Fernández MD (2010) Bioventing remediation and ecotoxicity evaluation of phenanthrene-contaminated soil. *J Hazard Mater* 183:806–813
- Fulazzaky MA, Talaiekhosani A, Ponraj M, Abd Majid MZ, Hadibarata T, Goli A (2014) Biofiltration process as an ideal approach to remove pollutants from polluted air. *Desalination Water Treat* 52:3600–3615
- Gadd GM (2010) Metals, minerals, and microbes: geomicrobiology and bioremediation. *Microbiology* 156:609–643
- Gafri HFS, Zuki FM, Aroua MK, Hashim NA (2019) Mechanism of bacterial adhesion on ultrafiltration membrane modified by natural antimicrobial polymers (chitosan) and combination with activated carbon (PAC). *Rev Chem Eng* 35:421–443. <https://doi.org/10.1515/revce-2017-0006>
- Gallardo-Rodríguez JJ, Rios-Rivera AC, Von Bennevit MR (2019) Living biomass supported on a natural-fiber biofilter for lead removal. *J Environ Manag* 231:825–832. <https://doi.org/10.1016/j.jenvman.2018.11.004>
- García-Delgado C, Yunta F, Eymar E (2015) Bioremediation of multi-polluted soil by spent mushroom (*Agaricus bisporus*) substrate: polycyclic aromatic hydrocarbons degradation and Pb availability. *J Hazard Mater* 300:281–288
- García-Peña EI, Hernández S, Favela-Torres E, Auria R, Revah S (2001) Toluene biofiltration by the fungus *Scedosporium apiospermum* TB1. *Biotechnol Bioeng* 76:61–69
- Garrison AW, Nzungu VA, Avants JK, Ellington JJ, Jones WJ, Rennels D, Wolfe NL (2000) Phytodegradation of p,p'-DDT and the enantiomers of o,p'-DDT. *Environ Sci Technol* 34:1663–1670
- Gaur N, Narasimhulu K, PydiSetty Y (2018) Recent advances in the bioremediation of persistent organic pollutants and its effect on environment. *J Clean Prod* 198:1602–1631
- Ghaly AE, Zhang B, Dave D (2011) Biodegradation of phenolic compounds in creosote treated wood waste by a composting microbial culture augmented with the fungus *Thermoascus aurantiacus*. *Am J Biochem Biotechnol* 7:90–103
- Ghasemi R, Golbabaie F, Rezaei S, Pourmand MR, Nabizadeh R, Jafari MJ, Masoorian E (2020) A comparison of biofiltration performance based on bacteria and fungi for treating toluene vapors from airflow. *AMB Express* 10:1–9. <https://doi.org/10.1186/s13568-019-0941-z>
- Gin KYH, Tang YZ, Aziz MA (2002) Derivation and application of a new model for heavy metal biosorption by algae. *Water Res* 36:1313–1323
- Giri BS, Kim KH, Pandey RA, Cho J, Song H, Kim YS (2014) Review of biotreatment techniques for volatile sulfur compounds with an emphasis on dimethyl sulfide. *Process Biochem* 49:1543–1554
- Golby S, Ceri H, Marques LL, Turner RJ (2014) Mixed-species biofilms cultured from an oil sand tailings pond can biomineralize metals. *Microb Ecol* 68:70–80
- Gómez-Borraz TL, González-Sánchez A, Bonilla-Blancas W, Revah S, Noyola A (2017) Characterization of the biofiltration of methane emissions from municipal anaerobic effluents. *Process Biochem* 63:204–213. <https://doi.org/10.1016/j.procbio.2017.08.011>
- Gopinath M, Pulla RH, Rajmohan KS, Vijay P, Muthukumar C, Gurunathan B (2018) Bioremediation of volatile organic compounds in biofilters. In: *Bioremediation: applications for Environmental Protection and Management*. Springer, Singapore, pp 301–330
- Grujić S, Vasić S, Čomić L, Ostojić A, Radojević I (2017) Heavy metal tolerance and removal potential in mixed-species biofilm. *Water Sci Technol* 76:806–812
- Gunatilake SK (2015) Methods of removing heavy metals from industrial wastewater. *Methods* 1:12–18
- Gupta VK, Gupta M, Sharma S (2001) Process development for the removal of lead and chromium from aqueous solutions using red mud—an aluminium industry waste. *Water Res* 35:1125–1134
- Gupta GS, Yadav G, Tiwari S (2020) Bioremediation of heavy metals: A new approach to sustainable agriculture. In: Upadhyay AK, Singh R, Singh DP (eds) *Restoration of wetland ecosystem: a trajectory towards a sustainable environment*. Springer Nature, Switzerland, pp 195–226
- Hamuda HEB, Tóth N (2012) Functioning of divalent alkaline metal on yeast multiplication in heavy metal contaminated soil. *Tájökológiai Lapok* 10:195–208
- Hansa A, Kumar V, Anshumali (2016) A comparative review towards potential of microbial cells for heavy metal removal with emphasis on biosorption and bioaccumulation. *World J Microbiol Biotechnol* 32(10):170. <https://doi.org/10.1007/s11274-016-2117-1>
- Haritonidis S, Malea P (1999) Bioaccumulation of metals by the green alga *Ulva rigida* from Thermaikos Gulf, Greece. *Environ Pollut* 104:365–372
- Hassan SW, El-Kassas HY (2012) Biosorption of cadmium from aqueous solutions using a local fungus *Aspergillus cristatus* Glaucus group. *Afr J Biotechnol* 11:2276–2286
- Hesami R, Salimi A, Ghaderian SM (2018) Lead, zinc, and cadmium uptake, accumulation, and phytoremediation by plants growing around Tang-e Douzan lead–zinc mine. *Iran Environ Sci Pollut Res* 25:8701–8714. <https://doi.org/10.1007/s11356-017-1156-y>
- Hong SUI, Xingang LI (2011) Modeling for volatilization and bioremediation of toluene-contaminated soil by bioventing. *Chin J Chem Eng* 19:340–348
- Hu Z, Hidalgo G, Houston PL, Hay AG, Shuler ML, Abruna HD, Ghirose WC, Lion LW (2005) Determination of spatial distributions of zinc and active biomass in microbial biofilms by two-photon laser scanning microscopy. *Appl Environ Microbiol* 71:4014–4021
- Hussein H, Ibrahim SF, Kandeel K, Moawad H (2004) Biosorption of heavy metals from waste water using *Pseudomonas* sp. *Electron J Biotechnol* 7:30–37
- Hwang SCJ, Wu SJ, Lee CM (2002) Water transformation in the media of biofilters controlled by *Rhodococcus fascians* in treating an ethyl acetate-contaminated airstream. *J Air Waste Manage Assoc* 52:511–520

- Ibanga IE, Fletcher LA, Noakes CJ, King MF, Steinberg D (2018) Pilot-scale biofiltration at a materials recovery facility: the impact on bioaerosol control. *Waste Manag* 80:154–167. <https://doi.org/10.1016/j.wasman.2018.09.010>
- Iranmanesh E, Halladj R, Zamir SM (2015) Microkinetic analysis of n-hexane biodegradation by an isolated fungal consortium from a biofilter: Influence of temperature and toluene presence. *CLEAN – Soil Air Water* 43:104–111
- Isaac P, Alessandrello MJ, Macedo AJ, Estévez MC, Ferrero MA (2017) Pre-exposition to polycyclic aromatic hydrocarbons (PAHs) enhance biofilm formation and hydrocarbon removal by native multi-species consortium. *J Environ Chem Eng* 5:1372–1378
- Jaber MB, Couvert A, Amrane A, Le Cloirec P, Dumont E (2017) Hydrogen sulfide removal from a biogas mimic by biofiltration under anoxic conditions. *J Environ Chem Eng* 5:5617–5623. <https://doi.org/10.1016/j.jece.2017.10.029>
- Jacques RJ, Okeke BC, Bento FM, Teixeira AS, Peralba MC, Camargo FA (2008) Microbial consortium bioaugmentation of a polycyclic aromatic hydrocarbons contaminated soil. *Bioresour Technol* 99:2637–2643
- Jahid IK, Ha SD (2014) The paradox of mixed-species biofilms in the context of food safety. *Compr Rev Food Sci F* 13:990–1011
- Jantschak A, Daniels M, Paschold R (2004) Biofilter technology: an innovative and cost-effective system to remove VOC. *IEEE T Semiconduct M* 17:255–260
- Javaid A, Bajwa R (2007) Biosorption of Cr (III) ions from tannery wastewater by *Pleurotus ostreatus*. *Mycopathologia* 5:71–79
- Jeong S, Cho K, Jeong D, Lee S, Leiknes T, Vigneswaran S, Bae H (2017) Effect of engineered environment on microbial community structure in biofilter and biofilm on reverse osmosis membrane. *Water Res* 124:227–237. <https://doi.org/10.1016/j.watres.2017.07.064>
- Jong T, Parry DL (2003) Removal of sulfate and heavy metals by sulfate reducing bacteria in short-term bench scale upflow anaerobic packed bed reactor runs. *Water Res* 37:3379–3389
- Kang KH, Sui Z (2010) Removal of eutrophication factors and heavy metal from a closed cultivation system using the macroalgae, *Gracilaria* sp. (Rhodophyta). *Chin J Oceanol Limnol* 28:1127–1130
- Katsoyiannis IA, Zouboulis AI (2004) Application of biological processes for the removal of arsenic from groundwaters. *Water Res* 38:17–26
- Kim KJ, Jeong MI, Lee DW, Song JS, Kim HD, Yoo EH, Jeong SJ, Han SW, Kays SJ, Lim YW, Kim HH (2010) Variation in formaldehyde removal efficiency among indoor plant species. *HortScience* 45:1489–1495
- Kotrba P (2011) Microbial biosorption of metals—general introduction. In: *Microbial biosorption of metals*. Springer, Dordrecht, pp 1–6
- Kujan P, Prell A, Šafář H, Sobotka M, Řezanka T, Holler P (2006) Use of the industrial yeast *Candida utilis* for cadmium sorption. *Folia Microbiol* 51:257–260
- Kumar PS, Gunasundari E (2018) Bioremediation of heavy metals. In: *Bioremediation: Applications for Environmental Protection and Management*. Springer, Singapore, pp 165–195
- Kumar TP, Rahul MA, Chandrajit B (2011) Biofiltration of volatile organic compounds (VOCs): An overview. *Res J Chem Sci* 1:83–92
- Kumar KV, Sridevi V, Harsha N, Lakshmi MC, Rani K (2013) Biofiltration and its application in treatment of air, and water pollutants—a review. *Int J Innov Eng Res Manag* 2:226–231
- Kumar R, Sharma AK, Singh P, Dhir B, Mehta D (2014) Potential of some fungal, and bacterial species in bioremediation of heavy metals. *J Nucl Phy Mat Sci Rad A* 1:213–223
- Kumar M, Giri BS, Kim KH, Singh RP, Rene ER, López ME, Rai BN, Singh H, Prasad D, Singh RS (2019) Performance of a biofilter with compost and activated carbon based packing material for gas-phase toluene removal under extremely high loading rates. *Bioresour Technol* 285:121317. <https://doi.org/10.1016/j.biortech.2019.121317>
- Kumari M, Tripathi BD (2015) Effect of *Phragmites australis* and *Typha latifolia* on biofiltration of heavy metals from secondary treated effluent. *Int J Environ Sci Technol* 12:1029–1038. <https://doi.org/10.1007/s13762-013-0475-x>
- Kumari P, Meena M, Upadhyay RS (2018a) Characterization of plant growth promoting rhizobacteria (PGPR) isolated from the rhizosphere of *Vigna radiata* (mung bean). *Biocatal Agric Biotechnol* 16:155–162. <https://doi.org/10.1016/j.bcab.2018.07.029>
- Kumari P, Meena M, Gupta P, Dubey MK, Nath G, Upadhyay RS (2018b) Plant growth promoting rhizobacteria and their biopriming for growth promotion in mung bean (*Vigna radiata* (L.) R. Wilczek). *Biocatal Agric Biotechnol* 16:163–171. <https://doi.org/10.1016/j.bcab.2018.07.030>
- Kurniawan TA, Chan GY, Lo WH, Babel S (2006) Physico-chemical treatment techniques for wastewater laden with heavy metals. *Chem Eng J* 118:83–98
- Labrenz M, Druschel GK, Thomsen-Ebert T, Gilbert B, Welch SA, Kemner KM, Logan GA, Summons RE, De Stasio G, Bond PL, Lai B (2000) Formation of sphalerite (ZnS) deposits in natural biofilms of sulfate-reducing bacteria. *Science* 290:1744–1747
- Le Cloirec P, Humeau P (2013) Bioscrubbers. In: *Air Pollution Prevention, and Control: Bioreactors, and Bioenergy*. John Wiley & Sons, Ltd, pp 139–153
- Lebrero R, Estrada JM, Muñoz R, Quijano G (2012) Toluene mass transfer characterization in a biotrickling filter. *Biochem Eng J* 60:44–49
- Lee BXY, Hadibarata T, Yuniarto A (2020) Phytoremediation mechanisms in air pollution control: a review. *Water Air Soil Pollut* 231:437. <https://doi.org/10.1007/s11270-020-04813-6>
- Leson G, Winer AM (1991) Biofiltration: an innovative air pollution control technology for VOC emissions. *J Air Waste Manage Assoc* 41:1045–1054
- Li WW, Yu HQ (2014) Insight into the roles of microbial extracellular polymer substances in metal biosorption. *Bioresour Technol* 160:15–23
- Li G, He Z, An T, Zeng X, Sheng G, Fu J (2008) Comparative study of the elimination of toluene vapours in twin biotrickling filters using two microorganisms *Bacillus cereus* S1 and S2. *J Chem Technol Biotechnol* 83:1019–1026
- Li Y, Zhang W, Xu J (2014) Siloxanes removal from biogas by a lab-scale biotrickling filter inoculated with *Pseudomonas aeruginosa* S240. *J Hazard Mater* 275:175–184
- Li H, Huang S, Zhou S, Chen P, Zhang Y (2016) Study of extracellular polymeric substances in the biofilms of a suspended biofilter for nitric oxide removal. *Appl Microbiol Biotechnol* 100:9733–9743. <https://doi.org/10.1007/s00253-016-7824-x>
- Liang CM, Wu XY, Huang K, Yan SQ, Li ZJ, Xia X, Pan WJ, Sheng J, Tao YR, Xiang HY, Hao JH, Wang QN, Tao FB, Tong SL (2019) Trace element profiles in pregnant women's sera and umbilical cord sera and influencing factors: repeated measurements. *Chemosphere* 218:869–878. <https://doi.org/10.1016/j.chemosphere.2018.11.115>
- Liang Y, Liu X, Wu F, Guo Y, Fan X, Xiao H (2020) The year-round variations of VOC mixing ratios and their sources in Kuytun City (Northwestern China), near oilfields. *Atmos Pollut Res* 11(9):1513–1523. <https://doi.org/10.1016/j.apr.2020.05.022>
- Liao D, Li E, Li J, Zeng P, Feng R, Xu M, Sun G (2018) Removal of benzene, toluene, xylene and styrene by biotrickling filters and identification of their interactions. *PLoS One* 13:e0189927. <https://doi.org/10.1371/journal.pone.0189927>
- Lim HS, Lim W, Hu JY, Ziegler A, Ong SL (2015) Comparison of filter media materials for heavy metal removal from urban stormwater runoff using biofiltration systems. *J Environ Manage* 147:24–33. <https://doi.org/10.1016/j.jenvman.2014.04.042>

- Liu HL, Chen BY, Lan YW, Cheng YC (2004) Biosorption of Zn (II) and Cu (II) by the indigenous *Thiobacillus thiooxidans*. *Chem Eng J* 97: 195–201
- Löffler FE, Ritalahti KM, Zinder SH (2013) *Dehalococcoides* and reductive dechlorination of chlorinated solvents. In: Bioaugmentation for Groundwater Remediation. Springer, New York, pp 39–88
- Lomonaco T, Manco E, Corti A, La Nasa J, Ghimenti S, Biagini D, Castelvetro V (2020) Release of harmful volatile organic compounds (VOCs) from photo-degraded plastic debris: a neglected source of environmental pollution. *J Hazard Mater* 394:122596. <https://doi.org/10.1016/j.jhazmat.2020.122596>
- Loukidou MX, Matis KA, Zouboulis AI, Liakopoulou-Kyriakidou M (2003) Removal of As(V) from wastewaters by chemically modified fungal biomass. *Water Res* 37:4544–4552
- Loukidou MX, Zouboulis AI, Karapantsios TD, Matis KA (2004) Equilibrium and kinetic modeling of chromium (VI) biosorption by *Aeromonas caviae*. *Colloid Surface A* 242:93–104
- Loutseti S, Danielidis DB, Economou-Amilli A, Katsaros C, Santas R, Santas P (2009) The application of a micro-algal/bacterial biofilter for the detoxification of copper and cadmium metal wastes. *Bioresour Technol* 100:2099–2105
- Luciano A, Viotti P, Torretta V, Mancini G (2013) Numerical approach to modelling pulse-mode soil flushing on a Pb-contaminated soil. *J Soils Sediments* 13:43–55. <https://doi.org/10.1007/s11368-012-0567-0>
- Mačaitis K, Misevičius A, Paškevičius A, Raudonienė V, Repečienė J (2014) Effectiveness research on a wavy lamellar plate-type biofilter with a capillary system for the humidification of the packing material applying introduced microorganisms. *J Environ Eng Landsc* 22:254–263. <https://doi.org/10.3846/16486897.2014.972409>
- Macek T, Mackova M, Káš J (2000) Exploitation of plants for the removal of organics in environmental remediation. *Biotechnol Adv* 18:23–34
- Machado MD, Santos MS, Gouveia C, Soares HM, Soares EV (2008) Removal of heavy metals using a brewer's yeast strain of *Saccharomyces cerevisiae*: the flocculation as a separation process. *Bioresour Technol* 99:2107–2115
- Machado MD, Janssens S, Soares HM, Soares EV (2009) Removal of heavy metals using a brewer's yeast strain of *Saccharomyces cerevisiae*: advantages of using dead biomass. *J Appl Microbiol* 6: 1792–1804
- Maestre JP, Gamisans X, Gabriel D, Lafuente J (2007) Fungal biofilters for toluene biofiltration: evaluation of the performance with four packing materials under different operating conditions. *Chemosphere* 67:684–692
- Majumder S (2015) Studies on removal of heavy metals from wastewater using biofiltration. (thesis, <https://shodhganga.inflibnet.ac.in/bitstream/10603/124741/1/thesis.pdf>)
- Malakar S, Saha PD, Baskaran D, Rajamanickam R (2017) Comparative study of biofiltration process for treatment of VOCs emission from petroleum refinery wastewater—a review. *Environ Technol Innov* 8:441–461
- Mamisahahei S, Khaniki GRJ, Torabian A, Nasseri S, Naddafi K (2007) Removal of arsenic from an aqueous solution by pretreated waste tea fungal biomass. *J Environ Health Sci Eng* 4:85–92
- Mapolelo M, Torto N (2004) Trace enrichment of metal ions in aquatic environments by *Saccharomyces cerevisiae*. *Talanta* 64:39–47
- Martinez-Juarez VM, Cárdenas-González JF, Torre-Bouscoulet ME, Acosta-Rodríguez I (2012) Biosorption of mercury (II) from aqueous solutions onto fungal biomass. *Bioinorg Chem Appl* 2012: 156190. <https://doi.org/10.1155/2012/156190>
- Meena M, Samal S (2019) *Alternaria* host-specific (HSTs) toxins: an overview of chemical characterization, target sites, regulation and their toxic effects. *Toxicol Rep* 6:745–758. <https://doi.org/10.1016/j.toxrep.2019.06.021>
- Meena M, Swapnil P (2019) Regulation of *WRKY* genes in plant defense with beneficial fungus *Trichoderma*: current perspectives and future prospects. *Arch Phytopathol Plant Protect* 52(1-2):1–17. <https://doi.org/10.1080/03235408.2019.1606490>
- Meena M, Zehra A (2019) Tomato: a model plant to study plant-pathogen interactions. *Food Sci Nutr Technol* 4(1):000171. <https://doi.org/10.23880/fsnt-16000171>
- Meena M, Tiwari A, Zehra A, Prasad V, Upadhyay RS (2013) Morphological and molecular identification of *Alternaria alternata* from tomato. Proceeding in International Conference on Global Scenario of Traditional System of Medicine, Ayurveda, Agriculture and Education, RGSC, Barkachha, BHU, 1:506–509
- Meena M, Prasad V, Zehra A, Gupta VK, Upadhyay RS (2015) Mannitol metabolism during pathogenic fungal–host interactions under stressed conditions. *Front Microbiol* 6:1019–1026
- Meena M, Prasad V, Upadhyay RS (2016a) Assessment of the bio-weedicidal effects of *Alternaria alternata* metabolites against *Parthenium* species. *Bull Environ Sci Res* 5(1):1–7
- Meena M, Zehra A, Dubey MK, Aamir M, Gupta VK, Upadhyay RS (2016b) Comparative evaluation of biochemical changes in tomato (*Lycopersicon esculentum* Mill.) infected by *Alternaria alternata* and its toxic metabolites (TeA, AOH, and AME). *Front Plant Sci* 7:1408. <https://doi.org/10.3389/fpls.2016.01408>
- Meena M, Zehra A, Dubey MK, Upadhyay RS (2016c) Mannitol and proline accumulation in *Lycopersicon esculentum* during infection of *Alternaria alternata* and its toxins. *Int J Biomed Sci Bioinformat* 3(2):64–68
- Meena M, Dubey MK, Swapnil P, Zehra A, Singh S, Kumari P, Upadhyay RS (2017a) The rhizosphere microbial community and methods of its analysis. In: Singh HB, Sarma BK, Keswani C. (Eds), Advances in PGPR research. CAB International, 275–295
- Meena M, Gupta SK, Swapnil P, Zehra A, Dubey MK, Upadhyay RS (2017b) *Alternaria* toxins: potential virulence factors and genes related to pathogenesis. *Front Microbiol* 8:1451. <https://doi.org/10.3389/fmicb.2017.01451>
- Meena M, Prasad V, Upadhyay RS (2017c) Evaluation of biochemical changes in leaves of tomato infected with *Alternaria alternata* and its metabolites. *Vegetos* 30:2. <https://doi.org/10.5958/2229-4473.2017.00020.9>
- Meena M, Prasad V, Upadhyay RS (2017d) Evaluation of *Alternaria alternata* isolates for metabolite production isolated from different sites of Varanasi, India. *J Agric Res* 2(1):00012
- Meena M, Swapnil P, Upadhyay RS (2017e) Isolation, characterization and toxicological potential of tenuazonic acid, alternariol and alternariol monomethyl ether produced by *Alternaria* species phytopathogenic on plants. *Sci Rep* 7:8777. <https://doi.org/10.1038/s41598-017-09138-9>
- Meena M, Swapnil P, Zehra A, Dubey MK, Upadhyay RS (2017f) Antagonistic assessment of *Trichoderma* spp. by producing volatile and non-volatile compounds against different fungal pathogens. *Arch Phytopathol Plant Protect* 50(13-14):629–648. <https://doi.org/10.1080/03235408.2017.1357360>
- Meena M, Swapnil P, Zehra A, Aamir M, Dubey MK, Upadhyay RS (2017g) Beneficial microbes for disease suppression and plant growth promotion. In: Singh D, Singh H, Prabha R (eds) Plant-microbe interactions in agro-ecological perspectives. Springer, Singapore, pp 395–432. https://doi.org/10.1007/978-981-10-6593-4_16
- Meena M, Zehra A, Dubey MK, Aamir M, Upadhyay RS (2017h) *Penicillium* enzymes for the food industries. In: Gupta VK, Rodriguez-Couto S. (Eds), New and future developments in microbial biotechnology and bioengineering. Elsevier, 167–186. <https://doi.org/10.1016/B978-0-444-63501-3.00014-4>
- Meena M, Zehra A, Swapnil P, Dubey MK, Patel CB, Upadhyay RS (2017i) Effect on lycopene, β -carotene, ascorbic acid and phenolic content in tomato fruits infected by *Alternaria alternata* and its

- toxins (TeA, AOH and AME). Arch Phytopathol Plant Protect 50(7-8):317–329. <https://doi.org/10.1080/03235408.2017.1312769>
- Meena M, Aamir M, Vikas K, Swapnil P, Upadhyay RS (2018) Evaluation of morpho-physiological growth parameters of tomato in response to Cd induced toxicity and characterization of metal sensitive NRAMP3 transporter protein. Environ Exp Bot 148:144–167. <https://doi.org/10.1016/j.envexpbot.2018.01.007>
- Meena M, Divyanshu K, Kumar S, Swapnil P, Zehra A, Shukla V, Yadav M, Upadhyay RS (2019a) Regulation of L-proline biosynthesis, signal transduction, transport, accumulation and its vital role in plants during variable environmental conditions. Heliyon 5(12):e02951. <https://doi.org/10.1016/j.heliyon.2019.e02952>
- Meena M, Swapnil P, Zehra A, Dubey MK, Aamir M, Patel CB, Upadhyay RS (2019b) Virulence factors and their associated genes in microbes. In: Singh HB, Gupta VK, Jogaiah S. (Eds), New and future developments in microbial biotechnology and bioengineering. Elsevier, 181–208. <https://doi.org/10.1016/B978-0-444-63503-7.00011-5>
- Meena M, Swapnil P, Divyanshu K, Kumar S, Harish TYN, Zehra A, Marwal A, Upadhyay RS (2020) PGPR-mediated induction of systemic resistance and physiochemical alterations in plants against the pathogens: Current perspectives. J Basic Microbiol 60(8):1–34. <https://doi.org/10.1002/jobm.202000370>
- Mehta SK, Gaur JP (2005) Use of algae for removing heavy metal ions from wastewater: progress and prospects. Crit Rev Biotechnol 25: 113–152
- Meliani A, Bensoltane A (2016) Biofilm-mediated heavy metals bioremediation in PGPR *Pseudomonas*. J Bioremediat Biodegrad 7:2. <https://doi.org/10.4172/2155-6199.1000370>
- Mergeay M, Monchy S, Vallaeyts T, Auquier V, Benotmane A, Bertin P, Wattiez R (2003) *Ralstonia metallidurans*, a bacterium specifically adapted to toxic metals: towards a catalogue of metal-responsive genes. FEMS Microbiol Rev 27:385–410
- Miller MJ, Allen DG (2004) Transport of hydrophobic pollutants through biofilms in biofilters. Chem Eng Sci 59:3515–3525. <https://doi.org/10.1016/j.ces.2004.05.011>
- Miller U, Sówka I, Adamiak W (2019) The effect of betaine on the removal of toluene by biofiltration. SN Appl Sci 1:984. <https://doi.org/10.1007/s42452-019-0832-6>
- Miyata N, Mori T, Iwahori K, Fujita M (2000) Microbial decolorization of melanoidin-containing wastewaters: combined use of activated sludge and the fungus *Corioliolus hirsutus*. J Biosci Bioeng 89:145–150
- Mohammad BT, Veiga MC, Kennes C (2007) Mesophilic and thermophilic biotreatment of BTEX-polluted air in reactors. Biotechnol Bioeng 97:1423–1438
- Mohan SV, Prasanna D, Reddy BP, Sarma PN (2008) Ex situ bioremediation of pyrene contaminated soil in bio-slurry phase reactor operated in periodic discontinuous batch mode: Influence of bioaugmentation. Int Biodeterior Biodegradation 62:162–169
- Mohapatra RK, Parhi PK, Pandey S, Bindhani BK, Thatoi H, Panda CR (2019) Active and passive biosorption of Pb (II) using live and dead biomass of marine bacterium *Bacillus xiamenensis* PbRPSD202: Kinetics and isotherm studies. J Environ Manag 247:121–134. <https://doi.org/10.1016/j.jenvman.2019.06.073>
- Montebello AM, Bezerra T, Rovira R, Rago L, Lafuente J, Gamisans X, Campoy S, Baeza M, Gabriel D (2013) Operational aspects, pH transition and microbial shifts of a H₂S desulfurizing biotrickling filter with random packing material. Chemosphere 93:2675–2682
- Mortgat B (2001) Traitement biologique des odeurs et COV. Environ Technol 203:39–42
- Mudliar S, Giri B, Padoley K, Satpute D, Dixit R, Bhatt P, Pandey R, Juwarkar A, Vaidya A (2010) Bioreactors for treatment of VOCs and odours—a review. J Environ Manag 91:1039–1054
- Murugesan GS, Sathishkumar M, Swaminathan K (2006) Arsenic removal from groundwater by pretreated waste tea fungal biomass. Bioresour Technol 97:483–487
- Nagashetti V, Mahadevaraju GK, Muralidhar TS, Javed A, Trivedi D, Bhusal KP (2013) Biosorption of heavy metals from soil by *Pseudomonas aeruginosa*. Int J Inno Technol Explor Eng 2:22–24
- Nasseri S, Kalantary R, Nourieh N, Naddafi K, Mahvi A, Baradaran N (2010) Influence of bioaugmentation in biodegradation of PAHs-contaminated soil in bio-slurry phase reactor. J Environ Health Sci Eng 7:199–208
- Newman LA, Reynolds CM (2004) Phytodegradation of organic compounds. Curr Opin Biotechnol 15:225–230
- Ngah WW, Hanafiah MM (2008) Removal of heavy metal ions from wastewater by chemically modified plant wastes as adsorbents: a review. Bioresour Technol 99:3935–3948
- Oh YS, Park SC (2000) Selection of suitable packing material for biofiltration of toluene, m-and p-xylene vapors. J Microbiol 38: 31–35
- Omil, F., 2014. Biological technologies for the removal of VOCs, odours and greenhouse gases.
- Ong SA, Toorisaka E, Hirata M, Hano T (2013) Comparative study on kinetic adsorption of Cu (II), Cd (II) and Ni (II) ions from aqueous solutions using activated sludge and dried sludge. Appl Water Sci 3: 321–325
- Orwell RL, Wood RA, Burchett MD, Tarran J, Torpy F (2006) The potted-plant microcosm substantially reduces indoor air VOC pollution: II. Laboratory study. Water Air Soil Poll 177:59–80
- Owa FW (2014) Water pollution: sources, effects, control and management. Int Lett Nat Sci 8:1–6
- Paca J, Halecky M, Kozliak E (2009) Styrene biofiltration using two packing materials with different adsorption properties. Environ Eng Sci 26:195–208
- Pandey A, Shukla A, Ray L (2009) Uptake and recovery of lead by agarose gel polymers. Am J Biochem Biotechnol 5:14–20
- Park OH, Jung IG (2006) A model study based on experiments on toluene removal under high load condition in biofilters. Biochem Eng J 28: 269–274
- Park DW, Kim SS, Haam S, Ahn IS, Kim EB, Kim WS (2002) Biodegradation of toluene by a lab-scale biofilter inoculated with *Pseudomonas putida* DK-1. Environ Technol 23:309–318
- Park D, Yun YS, Park JM (2005) Use of dead fungal biomass for the detoxification of hexavalent chromium: screening and kinetics. Process Biochem 40:2559–2565
- Parungao MM, Tacata PS, Tanayan CRG, Trinidad LC (2007) Biosorption of copper, cadmium and lead by copper-resistant bacteria isolated from Mogpog River, Marinduque Philipp. J Sci 136: 155–165
- Pegas PN, Alves CA, Nunes T, Bate-Epey EF, Evtuygina M, Pio CA (2012) Could houseplants improve indoor air quality in schools? J Toxicol Environ Health 75(22-23):1371–1380
- Pena-Castro JM, Martinez-Jerónimo F, Esparza-García F, Canizares-Villanueva RO (2004) Heavy metals removal by the microalga *Scenedesmus incrassatulus* in continuous cultures. Bioresour Technol 94:219–222
- Pettit T, Bettes M, Chapman AR, Hoch LM, James ND, Irga PJ, Torpy FR, Plants, Environmental Quality Research Group (2019) The botanical biofiltration of VOCs with active airflow: is removal efficiency related to chemical properties? Atmos Environ 214:116839. <https://doi.org/10.1016/j.atmosenv.2019.116839>
- Pinto AP, de Varennes A, Lopes ME, Teixeira DM (2016) Biological approaches for remediation of metal-contaminated sites. In: Phytoremediation. Springer, Cham, pp 65–112
- Polti MA, Aparicio JD, Benimeli CS, Amoroso MJ (2014) Simultaneous bioremediation of Cr(VI) and lindane in soil by actinobacteria. Int Biodeterior Biodegradation 88:48–55

- Prachuabmorn A, Noppaporn P (2010) Isolation and identification of xylene degrading microorganisms from biofilter. *J Appl Sci* 10: 585–589
- Prakash BS, Kumar SV (2013) Batch removal of heavy metals by biosorption onto marine algae—Equilibrium and kinetic studies. *Int J Chem Technol Res* 5:1254–1262
- Prenafeta-Boldú FX, Illa J, van Groenestijn JW, Flotats X (2008) Influence of synthetic packing materials on the gas dispersion and biodegradation kinetics in fungal air biofilters. *Appl Microbiol Biotechnol* 79:319–327
- Pulford ID, Watson C (2003) Phytoremediation of heavy metal-contaminated land by trees—a review. *Environ Int* 29:529–540
- Qi B, Moe W, Kinney K (2002) Biodegradation of volatile organic compounds by five fungal species. *Appl Microbiol Biotechnol* 58:684–689
- Qu KC, Li HQ, Tang KK, Wang ZY, Fan RF (2020) Selenium mitigates cadmium-induced adverse effects on trace elements and amino acids profiles in chicken pectoral muscles. *Biol Trace Elem Res* 193:234–240. <https://doi.org/10.1007/s12011-019-01682-x>
- Raboni M, Torretta V, Viotti P (2017) Treatment of airborne BTEX by a two-stage biotrickling filter and biofilter, exploiting selected bacterial and fungal consortia. *Int J Environ Sci Technol* 14:19–28. <https://doi.org/10.1007/s13762-016-1127-8>
- Rani MJ, Hemambika B, Hemapriya J, Kannan VR (2010) Comparative assessment of heavy metal removal by immobilized and dead bacterial cells: a biosorption approach. *Afr J Environ Sci Technol* 4:77–83
- Rascio N, Navari-Izzo F (2011) Heavy metal hyperaccumulating plants: how and why do they do it? And what makes them so interesting? *Plant Sci* 180:169–181
- Ren WX, Li PJ, Geng Y, Li XJ (2009) Biological leaching of heavy metals from a contaminated soil by *Aspergillus niger*. *J Hazard Mater* 167:164–169
- Rene ER, Veiga MC, Kennes C (2012) Combined biological and physicochemical waste-gas cleaning techniques. *J Environ Sci Heal A* 47:920–939
- Rene ER, Sergienko N, Goswami T, López ME, Kumar G, Saratale GD, Venkatachalam P, Pakshirajan K, Swaminathan T (2018) Effects of concentration and gas flow rate on the removal of gas-phase toluene and xylene mixture in a compost biofilter. *Bioresour Technol* 248: 28–35. <https://doi.org/10.1016/j.biortech.2017.08.029>
- Rho D (2000) La bioépuration de l'air, biofiltres et biolaveurs: un univers à la rencontre de la microbiologie et de l'ingénierie. *Vecteur Environ* 33:22–31
- Rinku NH, Patel V, Jasrai RT (2012) Removal of cadmium, chromium and lead from filamentous alga of *Pithophora* sp. of industrial wastewater. *Int J Environ Sci* 3:408–411
- Ryu HW, Kim SJ, Cho KS, Lee TH (2008) Toluene degradation in a polyurethane biofilter at high loading. *Biotechnol Bioprocess Eng* 13:360–365
- Sala Cossich E, Granhen Tavares CR, Kakuta Ravagnani TM (2002) Biosorption of chromium (III) by *Sargassum* sp. biomass. *Electron J Biotechnol* 5:6–7
- Saravanan A, Brindha V, Krishnan S (2011) Studies on the structural changes of the biomass *Sargassum* sp. on metal adsorption. *J Adv Bioinf* 2:193–196
- Sardood BP, Goltapeh EM, Verma A (2013) An introduction to bioremediation fungi as bioremediation. Springer, Heidelberg, pp 3–27
- Schiavon M, Ragazzi M, Rada EC, Torretta V (2016) Air pollution control through biotrickling filters: a review considering operational aspects and expected performance. *Crit Rev Biotechnol* 36:1143–1155
- Schlegelmilch M, Streese J, Stegmann R (2005) Odour management and treatment technologies: an overview. *Waste Manag* 25:928–939
- Seigneur C, Vuillemin A, Adler N, Peringer P (2001) A procedure for production of adapted bacteria to degrade chlorinated aromatics. *J Hazard Mater* 84:265–277
- Sene L, Converti A, Felipe MGA, Zilli M (2002) Sugarcane bagasse as alternative packing material for biofiltration of benzene polluted gaseous streams: a preliminary study. *Bioresour Technol* 83:153–157
- Sethi BK, Kanungo S, Rout JR, Nanda PK, Sahoo SL (2010) Effect of chromium on *Mucor* species and optimization of growth conditions. *Nat Sci* 8:29–32
- Shackira AM, Puthur JT (2019) Phytostabilization of heavy metals: Understanding of principles and practices. In: Srivastava S, Srivastava A, Suprasanna P (eds) *Plant-metal interactions*. Springer, Cham, pp 263–282. https://doi.org/10.1007/978-3-030-20732-8_13
- Sharma S (2012) Bioremediation: features, strategies and applications. *Asian J Pharm Life Sci* 2:202–213
- Shirpa J, Dikshit SN, Pandey G (2012) Comparative study of growing/immobilized biomass versus resting biomass of *A. lentulus* for the effect of pH on Cu²⁺ metal removal. *Res J Pharm, Biol Chem Sci* 3: 421–427
- Shoab A, Naureen A, Tanveer F, Aslam N (2012) Removal of Ni (II) ions from substrate using filamentous fungi. *Int J Agric Biol* 14(5): 831–833
- Showqi I, Lone FQ, Ashraf M, Mehmood MA, Rashid A (2016) Biofilters in mitigation of odour pollution - A review. *Nat Environ Pollut Technol* 15(4):1177–1185
- Shukla SR, Pai RS (2005) Adsorption of Cu (II), Ni (II) and Zn (II) on modified jute fibres. *Bioresour Technol* 96:1430–1438
- Singh N, Mhawish A, Deboudt K, Singh RS, Banerjee T (2017) Organic aerosols over Indo-Gangetic Plain: sources, distributions and climatic implications. *Atmos Environ* 157:59–74
- Soobhany N, Mohee R, Garg VK (2015) Comparative assessment of heavy metals content during the composting and vermicomposting of municipal solid waste employing *Eudrilus eugeniae*. *Waste Manag* 39:130–145
- Srivastava NK, Majumder CB (2008) Novel biofiltration methods for the treatment of heavy metals from industrial wastewater. *J Hazard Mater* 151:1–8
- Srivastava N, Singh RS, Dubey SK (2017) Efficacy of wood charcoal and its modified form as packing media for biofiltration of isoprene. *J Environ Manag* 196:252–260. <https://doi.org/10.1016/j.jenvman.2017.03.006>
- Strauss JM, Plessis CAD, Riedel KHJ (2000) Empirical model for biofiltration of toluene. *J Environ Eng* 126:644–648
- Subhashini SS, Kaliappan S, Velan M (2011) Removal of heavy metal from aqueous solution using *Schizosaccharomyces pombe* in free and alginate immobilized cells. In: 2nd International Conference on Environmental Science and Technology. 6:108–111
- Subramanian M, Oliver DJ, Shanks JV (2006) TNT phytotransformation pathway characteristics in *Arabidopsis*: role of aromatic hydroxylamines. *Biotechnol Prog* 22:208–221
- Suman J, Uhlík O, Viktorova J, Macek T (2018) Phytoextraction of heavy metals: a promising tool for clean-up of polluted environment? *Front Plant Sci* 9:1476. <https://doi.org/10.3389/fpls.2018.01476>
- Suriya J, Bharathiraja S, Rajasekaran R (2013) Biosorption of heavy metals by biomass of *Enterobacter cloacae* isolated from metal-polluted soils. *Int J Chem Tech Res* 5:1229–1238
- Sutherland C, Venkobachar C (2010) A diffusion-chemisorption kinetic model for simulating biosorption using forest macro-fungus, *fomes fasciatus*. *Int Res J Plant Sci* 1:107–117
- Tay CC, Liew HH, Yong SK, Surif S, Redzwan G, Abdul-Talib S (2012) Cu (II) removal onto fungal derived biosorbents: biosorption performance and the half saturation constant concentration approach. *Int Res J Chem Environ* 2:138–143

- Teiri H, Pourzamani H, Hajizadeh Y (2018) Phytoremediation of VOCs from indoor air by ornamental potted plants: A pilot study using a palm species under the controlled environment. *Chemosphere* 197: 375–381. <https://doi.org/10.1016/j.chemosphere.2018.01.078>
- Thijs S, Sillen W, Weyens N, Vangronsveld J (2017) Phytoremediation: State-of-the-art and a key role for the plant microbiome in future trends and research prospects. *Int J Phytoremediat* 19:23–38 19(1): 23–38. <https://doi.org/10.1080/15226514.2016.1216076>
- Tiwari S, Dixit S, Verma N (2007) An effective means of biofiltration of heavy metal contaminated water bodies using aquatic weed *Eichhornia crassipes*. *Environ Monit Assess* 129:253–256
- Tiwari AK, Singh PK, Singh AK, De Maio M (2016) Estimation of heavy metal contamination in groundwater and development of a heavy metal pollution index by using GIS technique. *Bull Environ Contam Toxicol* 96:508–515. <https://doi.org/10.1007/s00128-016-1750-6>
- Tiwari A, Alam T, Kumar A, Shukla AK (2019) Control of odour, volatile organic compounds (VOCs) & toxic gases through biofiltration—an overview. *Int J Tech Innov Mod Eng Sci* 5:1–6
- Torretta V, Collivignarelli MC, Raboni M, Viotti P (2015) Experimental treatment of a refinery waste air stream, for BTEX removal, by water scrubbing and biotrickling on a bed of *Mitilus edulis* shells. *Environ Technol* 36(18):2300–2307. <https://doi.org/10.1080/09593330.2015.1026289>
- Tovanabootr A, Semprini L, Dolan ME, Azizian M, Magar VS, DeBacker D, Lesson A, Kempisty CD (2001) Cometabolic air sparging field demonstration with propane to remediate trichloroethene and cis-dichloroethene. In: Sixth International *In Situ* and On Site Bioremediation Symposium, pp 145–153
- Treesubstorn C, Thiravetyan P (2018) Botanical biofilter for indoor toluene removal and reduction of carbon dioxide emission under low light intensity by using mixed C3 and CAM plants. *J Clean Prod* 194:94–100. <https://doi.org/10.1016/j.jclepro.2018.05.141>
- Valls M, De Lorenzo V (2002) Exploiting the genetic and biochemical capacities of bacteria for the remediation of heavy metal pollution. *FEMS Microbiol Rev* 26:327–338
- Valls M, de Lorenzo V, González-Duarte R, Atrian S (2000) Engineering outer-membrane proteins in *Pseudomonas putida* for enhanced heavy-metal bioadsorption. *J Inorg Biochem* 79:219–223
- Van Wyk CS (2011) Removal of heavy metals from metal-containing effluent by yeast biomass. *Afr J Biotechnol* 10:11557–11561
- Vázquez S, Agha R, Granado A, Sarro MJ, Esteban E, Penalosa JM, Carpena RO (2006) Use of white lupin plant for phytostabilization of Cd and As polluted acid soil. *Water Air Soil Pollut* 177:349–365
- Velkova Z, Stoytcheva M, Gochev V (2012) Biosorption of Cu (II) onto chemically modified waste mycelium of *Aspergillus awamori*: equilibrium, kinetics and modeling studies. *J Biosci Biotechnol* 1:163–169
- Vergara-Fernández A, Van Haaren B, Revah S (2006) Phase partition of gaseous hexane and surface hydrophobicity of *Fusarium solani* when grown in liquid and solid media with hexanol and hexane. *Biotechnol Lett* 28:2011–2017
- Vergara-Fernández A, Revah S, Moreno-Casas P, Scott F (2018a) Biofiltration of volatile organic compounds using fungi and its conceptual and mathematical modeling. *Biotechnol Adv* 36:1079–1093. <https://doi.org/10.1016/j.biotechadv.2018.03.008>
- Vergara-Fernández A, Yáñez D, Morales P, Scott F, Aroca G, Diaz-Robles L, Moreno-Casas P (2018b) Biofiltration of benzo[α]pyrene, toluene and formaldehyde in air by a consortium of *Rhodococcus erythropolis* and *Fusarium solani*: effect of inlet loads, gas flow and temperature. *Chem Eng J* 332:702–710. <https://doi.org/10.1016/j.cej.2017.09.095>
- Verma P, George KV, Singh HV, Singh SK, Juwarkar A, Singh RN (2006) Modeling rhizofiltration: heavy-metal uptake by plant roots. *Environ Model Assess* 11:387–394
- Vikrant K, Kim KH, Szulejko JE, Pandey SK, Singh RS, Giri BS, Brown RJ, Lee SH (2017) Bio-filters for the treatment of VOCs and odors—a review. *Asian J Atmos Environ* 11:139–152
- Vikrant K, Kailasa SK, Tsang DC, Lee SS, Kumar P, Giri BS, Singh RS, Kim KH (2018) Biofiltration of hydrogen sulfide: trends and challenges. *J Clean Prod* 187:131–147. <https://doi.org/10.1016/j.jclepro.2018.03.188>
- Volesky B, Holan ZR (1995) Biosorption of heavy metals. *Biotechnol Prog* 11:235–250
- Wambugu C, Marvins AD, Das J, Rene ER (2017) Conventional bioprocesses for the removal of gas-phase contaminants. *Res Rev Insights* 1:1–2
- Wang J, Chen C (2009) Biosorbents for heavy metals removal and their future. *Biotechnol Adv* 27:195–226
- White C, Sayer JA, Gadd GM (1997) Microbial solubilization and immobilization of toxic metals: key biogeochemical processes for treatment of contamination. *FEMS Microbiol Rev* 20:503–516
- Williams KH, Wilkins MJ, N'Guessan AL, Arey B, Dodova E, Dohnalkova A, Holmes D, Lovley DR, Long PE (2013) Field evidence of selenium bioreduction in a uranium-contaminated aquifer. *Environ Microbiol Rep* 5:444–452
- Woertz JR, Kinney KA, McIntosh NDP, Szanislo PJ (2001) Removal of toluene in a vapor-phase bioreactor containing a strain of the dimorphic black yeast *Exophiala lecanii-corni*. *Biotechnol Bioeng* 75: 550–558
- Wood RA, Burchett MD, Alquezar R, Orwell RL, Tarran J, Torpy F (2006) The potted-plant microcosm substantially reduces indoor air VOC pollution: I. Office field-study. *Water Air Soil Pollut* 175: 163–180
- Wu L, Li Z, Han C, Liu L, Teng Y, Sun X, Pan C, Huang Y, Luo Y, Christie P (2012) Phytoremediation of soil contaminated with cadmium, copper and polychlorinated biphenyls. *Int J Phytoremediat* 14:570–584
- Wu H, Yan H, Quan Y, Zhao H, Jiang N, Yin C (2018) Recent progress and perspectives in biotrickling filters for VOCs and odorous gases treatment. *J Environ* 222:409–419
- Xiao R, Zhang H, Wang Z, Zhang Z, Du J, Li R, Luo N, Ali A, Sun Z, Zhang Z (2019) Foliar litters: sources of contaminants in phytoremediation sites by returning potentially toxic metals (PTMs) back to soils. *Chemosphere* 222:9–14. <https://doi.org/10.1016/j.chemosphere.2019.01.090>
- Xu L, Xing X, Liang J, Peng J, Zhou J (2019) *In situ* phytoremediation of copper and cadmium in a co-contaminated soil and its biological and physical effects. *RSC Adv* 9:993–1003. <https://doi.org/10.1039/C8RA07645F>
- Yan G, Viraraghavan T (2000) Effect of pretreatment on the bioadsorption of heavy metals on *Mucor rouxii*. *Water Sa-Pretoria* 26:119–124
- Yang H, Liu Y (2011) Phytoremediation on air pollution. In: Khallaf M (ed) The impact of air pollution on health, economy, environment and agricultural sources. *Intech Open*, 1:281–294
- Yang B, Niu X, Ding C, Xu X, Liu D (2013) Performance of biotrickling filter inoculated with activated sludge for chlorobenzene removal. *Procedia Environ Sci* 18:391–396
- Yang K, Wang C, Xue S, Li W, Liu J, Li L (2019) The identification, health risks and olfactory effects assessment of VOCs released from the wastewater storage tank in a pesticide plant. *Ecotoxicol Environ Saf* 184:109665. <https://doi.org/10.1016/j.ecoenv.2019.109665>
- Yavuz H, Denizli A, Güngüneş H, Safarikova M, Safarik I (2006) Biosorption of mercury on magnetically modified yeast cells. *Sep Purif Technol* 52:253–260
- Yeom SH, Daugulis AJ (2001) Development of a novel bioreactor system for treatment of gaseous benzene. *Biotechnol Bioeng* 72:156–165
- Zehra A, Dubey MK, Tiwari A, Meena M, Kumari P, Singh VK, Gupta VK, Upadhyay RS (2015) Fungal biomolecules and their implications. In: Gupta VK, Mach RL, Sreenivasaprasad S (eds) *Fungal*

- biomolecules: source applications and recent developments. Wiley Blackwell, John Wiley & Sons Ltd., USA, pp 363–375
- Zehra A, Meena M, Swapnil P, Raytekar NA, Upadhyay RS (2020) Sustainable approaches to remove heavy metals from water. In: Singh J, Vyas A, Wang S, Prasad R (eds) *Microbial biotechnology: basic research and applications*. Springer, Singapore, pp 127–146. <https://doi.org/10.1007/978-981-15-2817-0>
- Zehraoui A, Hassan AA, Sorial GA (2012) Effect of methanol on the biofiltration of n-hexane. *J Hazard Mater* 219:176–182
- Zhang L, Jia Y, Zhang H, Yang T (2010) Pilot study on removal of benzene by using high efficient biotrickling filter. In: 4th International Conference on Bioinformatics and Biomedical Engineering, IEEE. pp 1–4.
- Zhao L, Huang S, Wei Z (2014) A demonstration of biofiltration for VOC removal in petrochemical industries. *Environ Sci Process Impacts* 16:1001–1007
- Zheng M, Li C, Liu S, Gui M, Ni J (2016) Potential application of aerobic denitrifying bacterium *Pseudomonas aeruginosa* PCN-2 in nitrogen oxides (NO_x) removal from flue gas. *J Hazard Mater* 318:571–578
- Zhou Q, Yang N, Li Y, Ren B, Ding X, Bian H, Yao X (2020) Total concentrations and sources of heavy metal pollution in global river and lake water bodies from 1972 to 2017. *Glob Ecol Conserv* 22:e00925. <https://doi.org/10.1016/j.gecco.2020.e00925>
- Zhuang P, Yang QW, Wang HB, Shu WS (2007) Phytoextraction of heavy metals by eight plant species in the field. *Water Air Soil Pollut* 184:235–242
- Zou JL, Xu GR, Pan K, Zhou W, Dai Y, Wang X, Ma M (2012) Nitrogen removal and biofilm structure affected by COD/NH₄⁺-N in a biofilter with porous sludge-ceramsite. *Sep Purif Technol* 94:9–15
- Zurita F, De Anda J, Belmont MA (2009) Treatment of domestic wastewater and production of commercial flowers in vertical and horizontal subsurface-flow constructed wetlands. *Ecol Eng* 35:861–869

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.