**RESEARCH ARTICLE** 



# Oxidative stress in liver of turtle *Mauremys reevesii* caused by cadmium

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#### Abstract

The research was designed to examine oxidative stress of the liver of turtle *Mauremys reevesii* caused by cadmium (Cd). Turtles were injected intraperitoneally with cadmium at the concentration of 7.5, 15, and 30 mg/kg, and 5 turtles were taken from each group after exposure for 1 week (1 w), 2 weeks (2 w), and 3 weeks (3 w). The activities of SOD and CAT as well as the contents of GSH and MDA in liver tissues were detected by using a kit. The results showed that the difference between the control group and the Cd-treated group was statistically significant with the increase of Cd concentration and the prolongation of exposure time, which suggested that Cd caused oxidative stress on the liver of turtles.

Keywords Cadmium · Turtles · Liver · Oxidative stress

## Introduction

Cadmium (Cd) is a worldwide freshwater aquatic pollutant (Mehinto et al. 2014; Novelli et al. 2000). Cd excretion by the body is very slow, so its biological half-life is relatively long, up to 15–20 years (Joseph. 2009). Therefore, Cd is able to be plentiful through the food web (Rose et al. 2015). While the turtle is in the upper layer of the food chain, Cd is likely to be enriched higher in the turtle. However, there are only a few studies on Cd poisoning of turtle (Dayna et al. 2016; Huo et al. 2017a, b, 2018, 2020a, b; Yu et al. 2013).

The mechanism of Cd poisoning is not very clear, and several interpretations are presented (Varoni et al. 2017). The studies of toxicological Cd indicate that it can cause an increase in reactive oxygen species (ROS), which is able to induce multiple

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<sup>2</sup> Bureau of agriculture and rural affairs of Qianan, Tangshan, Hebei Province, China structural and functional damages, such as cellular lipid peroxidation, protein destruction, and DNA mutation (Cuypers et al. 2010; Huo et al. 2017a, b, 2018, 2020a, b; Wu et al. 2015). One of the products of lipid is malondialdehyde (MDA), which is able to bind to the amino acids of protein and trigger the internal or mutual connection of protein, leading to cell damage (Li et al. 2012). Therefore, the content of MDA can reflect the attack degree of ROS on cells, and it can be referenced as one marker of membrane damage (Amin et al. 2018). There is an antioxidant system in the body, which includes superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GSH-Px) (Jiao et al. 2017; Koim-Puchowska et al. 2020; Laurent et al. 2017; Serdar et al. 2018; Yuan et al. 2016). ROS can be cleared by the antioxidant system. SOD can convert ROS into hydrogen peroxides (H<sub>2</sub>O<sub>2</sub>), and GSH-Px or CAT is able to degrade H<sub>2</sub>O<sub>2</sub> (Afonso et al. 2007). Cd regulates the activities of SOD, CAT, and GSH-Px of the antioxidant system (Yuan et al. 2016).

The purpose of this research was to investigate the effects of Cd on oxidative stress in liver of turtle *Mauremys reevesii*.

### **Materials and methods**

#### Animals and treatments

After acclimation, healthy, similar-weight  $(130 \pm 10 \text{ g})$ Mauremys reevesii were selected for experiment. Mauremys

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*reevesii* were randomly divided into one control group (5 individuals) and three experiment groups (45 individuals, 15 turtles in each group). The control group turtles were injected with 0.85% sodium chloride solution, and the experiment group turtles were injected with 7.5, 15, and 30-mg/kg cadmium chloride respectively for only once.

### Sample collection

Five turtles were randomly sacrificed from each group after 1 week (1 w), 2 weeks (2 w), and 3 weeks (3 w) of Cd exposure. Samples of liver tissue were excised and weighed immediately. Add 9 times medium (0.85% saline solution) to liver tissue by weight (g):volume (ml) equal to the proportion of 1:9, homogenize, and centrifuge at 2500 r/min for 10 min at 4 °C. Then, the supernate was stored at - 80 °C for detecting the oxidation index.

#### **Biochemical assays**

Under the manufacturer's protocols, the content of GSH and MDA and the activities of SOD and CAT in supernatant were detected with the detection kits. All the detections were performed by using a microplate reader.

#### Statistical analysis

Statistical analyses were performed with the SPSS 20.0 software package. The data was representative of mean values of five animals of each group, and the results were shown as means  $\pm$  standard deviations (SD). The probability value less than 0.05 was statistically different. If the probability value was less than 0.01, the difference was statistically significant.

### Results

# Correlations between SOD activities and Cd treatments in liver

From Fig. 1, SOD activities in liver of turtles exposed to 7.5 mg/kg gradually increased with the extension of exposure time, and the activities of SOD for 2 weeks and 3 weeks after Cd treatment were markedly increased (P < 0.01). But SOD activities in the liver of turtles exposed to 15 and 30 mg/kg declined with prolongation of exposure time, even SOD activities in the 30 mg/kg group after 3 weeks of Cd exposure were significantly lower than that of the control turtles (P < 0.01).

The activities of SOD in the liver gradually increased as the dose increases for 1 week. In comparison with the control group, the activities of SOD in the 7.5 mg/kg group had no statistical difference, but SOD activities in the 15 and 30 mg/kg groups were markedly increased (P < 0.01). SOD



**Fig. 1** Effects of Cd on the activities of SOD in the liver of treated turtles. Compared with the control group, \*P < 0.05, \*\*P < 0.01

activities in the 7.5, 15, and 30 mg/kg groups after 2 weeks of Cd treatment were significantly increased (P < 0.01), but the activities of SOD in liver gradually decreased in a dose-dependent mode for 2 weeks. The activities of SOD in the liver increased first and then decreased in a dose-dependent mode after the turtles being exposed to Cd for 3 weeks. Compared with the control group, the activities of SOD in the 7.5 mg/kg group were markedly increased (P < 0.01), while the activities of SOD in the 15 mg/kg group had no statistical difference, but the activities of SOD in the 30 mg/kg group were markedly reduced (P < 0.01).

Overall, the activities of SOD of the liver of turtles *Mauremys reevesii* increased except that those of the 30 mg/kg group decreased after 3 weeks of Cd treatment. In brief, low-dose Cd could induce the increase of SOD activities in the turtle liver; however, high dose of Cd-induced SOD activities in the turtle liver increased first then decreased. The results showed that the changes of SOD activities were dosedependent and time-dependent.

# Correlations between CAT activities and Cd treatments in the liver

As shown in Fig. 2, all of the CAT activities in the liver of turtles *Mauremys reevesii* decreased. The activities of CAT in liver of turtles exposed to 7.5 and 30 mg/kg gradually decreased with prolongation of exposure time. However, the downward trend of the CAT activities in the liver of turtles exposed to 15-mg/kg CAT is not typical. The activities of CAT in the liver decreased for 1 week. The declining degree of CAT activities in the 15 mg/kg group was the most obvious. Compared with the control group, the activities of CAT in the 7.5 mg/kg group had no statistical difference, but the activities of CAT in the 15 and 30 mg/kg groups were markedly decreased (P < 0.01). CAT activities in liver decreased for 2



Fig. 2 Effects of Cd on the activities of CAT in the liver of treated turtles. Compared with the control group, \*P < 0.05, \*\*P < 0.01

weeks. The declining degree of CAT activities in the 15 mg/kg group was the most obvious. The activities of CAT in the 7.5, 15, and 30 mg/kg groups were significantly decreased (P < 0.01). The activities of CAT in the liver decreased dose-dependently after being exposed to Cd for 3 weeks. Compared with the control group, the activities of CAT in the 7.5, 15, and 30 mg/kg group were markedly reduced (P < 0.01).

In short, CAT activities decreased obviously with the increase of dose and time. The outcomes indicated that changes of the activities of CAT were dose-dependent and timedependent.

# Correlations between the content of GSH and Cd treatments in the liver

GSH content in the liver of turtles exposed to 7.5 mg/kg declined slightly (1 W) and then significantly increased (2 W) followed by a little decrease (3 W) with prolongation of exposure time (Fig. 3). GSH content in the liver of turtles exposed to 15 and 30 mg/kg increased first and then gradually decreased with the extension of exposure time. The content of GSH in the liver decreased first (7.5 mg/kg) then increased (15 and 30 mg/kg) as the dose increases for 1 week. In comparison with the control group, GSH content in the 7.5 mg/kg group had no statistical difference, but the contents of GSH in the 15 and 30 mg/kg groups were markedly increased (P < 0.01). GSH content in the liver increased first (7.5 and 15 mg/kg) then decreased (30 mg/kg) for 2 weeks. The content of GSH in the 7.5 mg/kg group was significantly increased (P < 0.01); the 15 and 30 mg/kg groups had no statistical difference with that in the control group. GSH content in the liver increased first (7.5 mg/kg) then decreased (15 and 30 mg/kg) after being exposed to Cd for 3 weeks. Compared with the control group, GSH content in the 7.5 and 15 mg/kg groups had no statistical



**Fig. 3** Effects of Cd on the content of GSH in the liver of treated turtles. Compared with the control group, \*P < 0.05, \*\*P < 0.01

difference, but the content of GSH in the 30 mg/kg group was markedly reduced (P < 0.01).

In brief, low-dose Cd had complicated effect on the content of GSH in the turtle liver; however, high dose of Cd-induced GSH content in the turtle liver increased first then decreased with prolongation of exposure time and increase of dose.

# Correlations between the content of MDA and Cd treatments in the liver

In comparison with the control group, the content of MDA in each exposure group and each exposure time increased (Fig. 4). The content of MDA in the liver of turtles exposed to 7.5 and 30 mg/kg increased first and



**Fig. 4** Effects of Cd on the content of MDA in the liver of treated turtles. Compared with the control group, \*P < 0.05, \*\*P < 0.01

then decreased with prolongation of exposure time. MDA content in the liver of turtles exposed to 15 mg/kg had no clear trend. In comparison with the control group, MDA content in the 7.5 mg/kg group had no statistical difference, but the contents of MDA in the 15 and 30 mg/kg group were markedly increased (P < 0.01) for 1 week. The content of MDA in the 7.5, 15, and 30 mg/kg groups increased significantly (P < 0.01) for 2 weeks. In comparison with the control group, MDA content in the 7.5 mg/kg group increased significantly (P < 0.01) for 2 weeks. In comparison with the control group, MDA content in the 7.5 mg/kg group increased significantly (P < 0.01), but the 15 and 30 mg/kg groups had no statistical difference.

In conclusion, Cd could induce the increase of MDA content in the turtle liver.

### Discussion

Incomplete reduction of oxygen at electron donor sites on several mitochondrial and extra-mitochondrial enzymes produces ROS in cells (Brand, 2016; Mailloux, 2018; Wong et al., 2019). ROS is involved in many cellular functions and regulatory networks (Sies 2017; Mailloux 2018).

Excessive production of ROS causes cells to enter the oxidative stress state, followed by cell necrosis or apoptosis (Wu et al. 2014; Gilgun-Sherki et al. 2002). MDA is one of the widely recognized biomarkers of oxidative stress and cellular lipid peroxidation (Lawal et al. 2011; Zhou et al. 2017).

ROS produced by heavy metal poisoning can be cleared by the cellular defense enzyme system, such as SOD, CAT, and GSH (Sevcikova et al. 2011). The proximal ROS produced by the mitochondria is superoxide anion  $(\cdot O_2^{-})$  and  $H_2O_2$  (Mailloux, 2018).  $O_2^{-}$  results from single electron reduction of molecular oxygen, then  $O_2^{-1}$  is dismutated to H<sub>2</sub>O<sub>2</sub> and O<sub>2</sub> spontaneously or catalyzed into  $H_2O_2$  and  $O_2$  by manganese superoxide dismutase (Mn-SOD) (mitochondrial matrix) and copper-zinc superoxide dismutase (Cu/Zn-SOD) (cytosol and mitochondrial intermembrane space) (Okoye et al., 2019).  $H_2O_2$  is also produced by paired electron reduction of oxygen directly (Andreyev et al., 2015; Brand, 2016). H<sub>2</sub>O<sub>2</sub> is a secondary messenger to coordinate oxidative metabolism with changes in cell physiology. The level of H<sub>2</sub>O<sub>2</sub> is regulated through its production and degradation (Mailloux, 2018). CAT is able to catalyze H<sub>2</sub>O<sub>2</sub> into H<sub>2</sub>O and O<sub>2</sub> (Fomazier et al., 2002). GSH plays an important role in neutralizing ROS and directly countering Cd toxicity (Varoni et al. 2017).

Cd cannot stimulate the body to produce ROS directly, but it can indirectly induce the body to produce excessive ROS and destroy the antioxidant defense system (Cuypers et al. 2010). Cd increases the levels of lipid peroxidation and decreases total antioxidant capacity in hepatic tissue (Moradkhani et al. 2020). Cd can induce the body to produce  $O_2^-$ ,  $H_2O_2$ , and hydroxyl radicals (·OH) by altering the expression of ROS-related genes (Patra et al. 2011). When there are no enough GSH and CAT enzymes, excessive ·OH is produced by Fenton's reaction (Abdeen et al. 2019). ·OH has the highest rate of reactivity compared with other ROS, so it is the most harmful radical (Sies 2017). It results from the breakdown of the unsaturated fatty acid content (Avery 2011). ·OH can cause lipid peroxidation and production of MDA by diffusing or reacting with distant molecules such as cell membranes (Avery 2011).

Cd can combine with SOD sulfhydryl groups to form the Cu-Cd-SOD complex by instead of Zn in the SOD, thus causing the decrease or even disappearance of the SOD activities (Wu et al. 2014). The activities of SOD reduce obviously in the mice liver of the damage group (Gong et al. 2017; Pang et al. 2010). The activities of SOD decline in rats' liver (Wang et al. 2018; Yu et al. 2006). SOD activities in the liver of Bufo gargarizans are declined (Jia et al. 2004). SOD activities in the liver of Orechromis niloticus decrease (Wang et al. 2016). The activities of SOD in the hepatopancreas of crucian carp under the Cd alone-exposed group are lower than that in the control group (Zhuo et al. 2016). SOD activities in the liver of cock decline (Wang et al. 2007B). The activities of SOD in grass carp's liver tissues decrease first then increase and decrease once again when Cd<sup>2+</sup> presents low-concentration pollution, while the activity of SOD is inhibited obviously all along when Cd<sup>2+</sup> presents high-concentration pollution (Wang et al. 2007A). Our results of the 30 mg/kg group in 3 weeks (Fig. 1) were in accordance with these reports, which may be explained that the turtle had a strong tolerance to Cd exposure. There are also reports that SOD activities change in the opposite direction. SOD activities in the liver of the frog Rana nigromaculata are increased (Wang et al. 2006). Our results in the early days (Fig. 1) were in accordance with this report.

The activities of CAT reduce obviously in the mice liver of the Cd group (Gong et al. 2017). CAT activities in the liver of *Bufo gargarizans* are declined (Jia et al. 2004). CAT activities in the liver of *Orechromis niloticus* decrease (Wang et al. 2016). The activities of CAT in the hepatopancreas of crucian carp under the Cd-exposed group are lower than that in the control group (Zhuo et al. 2016). Our outcomes indicated a reduction of the activities of CAT in the turtle liver after Cd exposure (Fig. 2).

The content of GSH in rats' liver increases obviously (Yu et al. 2006). GSH contents are induced significantly in the liver of the frog *Rana nigromaculata* with an increase of exposure time (Wang et al. 2006). Our study also showed an increase of the content of GSH in the turtle liver after Cd treatment (Fig. 3). However, it is not in agreement with some study that animals exposed to Cd decrease the content of GSH. GSH content decreases in the liver of *Bufo gargarizans* (Jia et al. 2004) and *Orechromis niloticus* (Wang et al. 2016).

The difference may come from dosage, exposure duration, the way of administration, species, and age of the experimental animals.

The content of MDA in the liver of mice increases with the prolonged time (Gong et al. 2017; Pang et al. 2010). MDA content in rats' liver increases (Wang et al. 2018). MDA content in the liver of the frog *Rana nigromaculata* increases rapidly with the prolonged time (Wang et al. 2006). The MDA content in the *Bufo gargarizans* liver increases (Jia et al. 2004). MDA content in the hepatopancreas of crucian carp under the Cd-exposed group increases (Zhuo et al. 2016). MDA content in the liver of cock increases (Wang et al. 2007B). Our study indicated that the content of MDA increased in the turtle liver (Fig. 4).

### Conclusions

Low-dose Cd could induce the increase of SOD activities in the turtle liver; however, high dose of Cd-induced SOD activities in the turtle liver increased first then decreased. CAT activities decreased with the increase of dose and time. Lowdose Cd increased the content of GSH in the turtle liver; however, high dose of Cd-induced GSH content in the turtle liver increased first then decreased with prolongation of exposure time. Cd could induce the increase of MDA content in the turtle liver. The results of our experiments indicated that Cd is able to induce oxidative stress and damage in turtles *Mauremys reevesii*. But turtles seemed to have some tolerance to Cd exposure.

Authors' contributions Aiguo Dong and Junfeng Huo designed the study, performed the research, analyzed the data, and wrote the paper. Juanjuan Yan was a major contributor in performing the research. Ailing Dong was a major contributor in writing the manuscript. All authors read and approved the final manuscript.

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Data availability Not applicable.

### **Compliance with ethical standards**

**Competing interests** The authors declare that they have no competing interests.

**Ethics approval and consent to participate** This study was approved by Shanxi University of Chinese Medicine (permit number: SXZYYDXLL022).

**Consent to publish** Not applicable.

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