RESEARCH ARTICLE



Responses of nitrogen transformation and dissolved oxygen in constructed wetland to biochar and earthworm amendment

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Abstract

Many constructed wetland systems are facing the problem of low dissolved oxygen (DO) and reduced nitrogen removal efficiency. In this study, an experimental constructed wetland system is designed and used to investigate the effect of biochar (rice husk biochar (RHB), coconut shell biochar (CSB), and wood biochar (WB) and earthworm on DO concentration, nitrogen transformation, and ammonia nitrogen removal. Specifically, effects of different biochar and earthworm on NH4+-N in wastewater, N content of *Phragmites australis*, NH_4^+ -N and NO_3^- -N content in substrates, microbial nitrification and denitrification potentials, and the DO concentration were investigated. Results show that the addition of biochar and earthworm increased the removal efficiency of NH4⁺-N from wastewater. The addition of RHB and WB significantly increased the concentration of DO by 21.4% and 25.7% (P < 0.05) respectively in the constructed wetland. The addition of earthworm significantly increased the DO concentration in the constructed wetland system by an average of 30.35% (P < 0.05). The N content of P. australis increased when biochar and earthworm were introduced into the constructed wetland system, with higher relative N content observed in the above-ground biomass. NO_3^- -N content increased, but NH_4^+ -N decreased in the substrate. Addition of both biochar and earthworm increased nitrification and denitrification potentials. However, no significant increase in denitrification potential was observed when only biochar was added. The removal efficiency of NH_4^+ -N from wastewater is significantly positively correlated with the DO, nitrification, and denitrification potential, and nitrogen content of above-ground part of *P. australis* (*P* < 0.05). Results suggest that the DO concentration in constructed wetland systems could be improved by the addition of biochar and earthworm. These findings imply that both biochar and earthworm could be added into constructed wetlands to solve the low DO concentration and improve the removal efficiency of nitrogen.

Keywords Constructed wetland · Biochar · Eisenia fetida · Ammonia nitrogen

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Introduction

Ammonia nitrogen, including free ammonia (NH₃) and ammonium ions (NH₄⁺), is mainly derived from nitrogenous organic matter in domestic sewage, such as fertilizer, excrement of aquatic animals, and decomposition of animal carcasses. Ammonia can be present in water as ionized ammonia nitrogen (NH₄⁺-N) and unionized ammonia nitrogen (NH₃ -N), with the ionized form being most common. The NH₃ can react with water to form a weak base and the reaction is reversible (Goldshleger et al. 2018). The ratio of NH₃-N to NH₄⁺-N is normally in equilibrium depending on water pH and temperature. Aquatic organisms may be at risk of poisoning from ammonia (Egnew et al. 2019; Lu et al. 2019). Zhou and Claude (2015) showed that biological membranes are more permeable to NH₃ than to NH₄⁺, and Cheng et al. (2019) demonstrated that NH₃-N was more toxic to aquatic organisms, especially for benthic crustaceans and mollusks due to its diffusion ability across the gill membranes. However, Liew et al. (2013) reported that NH_4^+ has some degree of toxicity because the high NH_4^+ concentration in the water interferes with the outward movement of ammonia through the gills. Wang et al. (2020) showed that annual emissions of ammonia in China are approximately 2.3 million tons. Therefore, removal of NH_4^+ -N from wastewater is potentially very important to protect human health.

The main methods for removing NH₄⁺-N from wastewater include adsorption, chemical precipitation, and biological action. The adsorption method has been widely used in the treatment of wastewater containing NH4+-N because it has many advantages, such as simple operation, no secondary pollution, and the potential to regenerate the adsorbent (Xu et al. 2019). However, the high cost of adsorbent regeneration and usually low adsorption capacity are disadvantages. NH₄⁺-N can also be removed from wastewater by chemical precipitation. For example, the MgNH₄PO₄·6H₂O (MAP) precipitate occurs when adding Mg^{2+} and PO_4^{3-} into the wastewater with NH4⁺-N, which is a compound fertilizer and has good economic benefits (Ueno and Fujii 2001). However, chemical precipitation requires large amounts of chemical agents, resulting in increased treatment cost. NH4⁺-N can also be removed from wastewater by nitrification-denitrification by microorganisms. This method has many advantages, such as low investment cost and no significant secondary pollution. However, the purifying effectiveness is significantly influenced by temperature and certain substances (pH and toxic metals, etc.) in wastewater.

Constructed wetland systems (CWs) consisting of substrate, aquatic plants, and microorganisms, have an efficient pollutant purification capacity by synergy of physical, chemical, and biological interactions (Ilyas and Masih 2017). Biochar refers to a fine-grained, porous carbon material formed by high-temperature pyrolysis of carbon-rich biomass under anaerobic conditions. Biochar can possess strong adsorption and anti-biodegradation properties (Manyà et al. 2018). At present, biochar has been used in constructed wetlands to increase purification capacity. Kuno et al. (2018) found that addition of wood biochar into CWs increased the removal of total nitrogen (TN) by 20.0%. Zhou et al. (2017) demonstrated that addition of bamboo biochar into intermittently aerated vertical flow CWs improved removal of NH₄⁺-N and TN by 99% and 53%, respectively. de Rozari et al. (2018) found an 81% reduction in ammonia following the addition of biochar to a CWs. This improvement was attributed to the following main factors: (1) Improved nitrification by biochar. Nitrification is the first step in the nitrogen transformation process in NH₄⁺-N for nitrogen removal of wastewater, and the high DO concentration can promote nitrifying bacteria for nitrification (Lu et al. 2016). Compared to the traditional media, biochar has an extensive surface area for the adsorption of pollutants, microorganism growth, and development to improve the pollutant degradation (Feng et al. 2020). (2) Increasing adsorption NH_4^+ -N by biochar due to its composition of coarse and heterogeneous surfaces, shallow channels (Kasak et al. 2018), and its porosity (Li et al. 2018). (3) Enhanced plant uptake NH_4^+ -N by biochar. Uptake of NH_4^+ -N by *Lythrum Salicaria* improved growth and the removal of NH_4^+ -N provided improved conditions (in terms of oxygen secretion and root surface) for nitrification (Kasak et al. 2018). Therefore, the much larger specific surface area, total pore volume, and micropore volume of biochar were favorable for NH_4^+ -N adsorption for subsequent nitrification and uptake by plant.

Earthworm is a common terrestrial annelid in soil and plays a key role in the ecosystem as decomposers. Previous research shows that adding earthworm into CWs effectively improves the clogging problem of wetland systems and increases the purification efficiency (Davison et al. 2005). Li et al. (2011) showed that the quantity of solid particles accumulated in the substrate was reduced by 50%, when earthworm was added into a subsurface flow CWs. The addition of earthworm can also affect the growth of wetland plants, which means more N could be absorbed and continuously removed by harvesting of mature plants. Xu et al. (2013a, b) found that the absorption of N in the above-ground part of *P. australis, Typha*, and *Canna* in CW increased by 185%, 216%, and 108%, respectively. Therefore, the removal of N improved with the addition of earthworms into CWs.

In general, NH₄⁺-N removal efficiency is mainly related to nitrification in aerobic systems, and the improvement of dissolved oxygen (DO) content, e.g., through mechanical aeration, is conducive to the enhancement of removal of NH₄⁺-N by nitrification. DO content is therefore a significant factor for effective NH4⁺-N removal in CWs. In CWs, substrate layers also play a vital role in the elimination of pollutants from wastewater. Substrates such as biochar, earthworms, gravel, activated carbon, and zeolite have been used in CWs (Abedi and Mojiri 2019; Xu et al. 2013a, 2013b). Previous studies have, however, tended not to look at the combined impact of biochar and earthworms in CWs, and there is a little information on the effect of DO content on removal of NH4⁺-N in CWs where both biochar and earthworm have been added. Therefore, the objectives of this study were to (1) analyze changes in DO content in CWs with biochar and earthworms; (2) determine the response of removal efficiency of NH₄⁺-N to CWs with biochar and earthworms; (3) assess NH₄⁺-N, NO₃⁻-N content, and nitrification and denitrification potentials in CWs with biochar and earthworms.

Materials and methods

Materials

Biochar, including rice husk biochar (RHB), coconut shell biochar (CSB), and wood biochar (WB), were obtained from

a market. Sand, with diameter less than 3 mm, was obtained from a local building supply company. *Phragmites australis* was chosen as the test plant and the seedlings were selected from the pond of Nanjing University of Information Science and Technology. Earthworms (*Eisenia fetida*) were purchased from a local farming market and *E. fetida* was chosen as the test earthworm because it has proven to be effective in vermifiltration (Taylor et al. 2003). *E. fetida* with an average weight of 0.2–0.3 g were selected.

Design of horizontal subsurface flow constructed wetland

An experimental CWs consisting of two plastic basins was designed (Fig. 1). All CWs were placed outdoors under a transparent shelter at Agriculture and Meteorology Experimental Station, Nanjing University of Information Sciences and Technology, Jiangsu Province, China (32.160 N, 118.860 E). Four holes in the bottom of the inner basin (each 1 cm in diameter) allowed free movement of water between the basins. Within the inner basin, a plastic tube with holes (dissolved oxygen tube) was placed inside a rhizobag made from nylon mesh (< 30 μ m) to allow exchange of water and air. The dissolved oxygen tube could effectively detect the DO content in substrate of CWs. Eight versions of the system were constructed using different combinations of substrate, biochar, and earthworm, as follows (Table 1). In each version of the system, 1 L and 7 L of the tested substrate were added to the inside and outside of the rhizobag, respectively. Two seedlings of P. australis were planted in the rhizobag after 1-2 weeks of laboratory cultivation. E. foetida with an average weight of 0.2-0.3 g were selected and added to the inner basin



Fig. 1 Schematic diagram of horizontal subsurface flow constructed wetland

of the relevant systems (0.75 g/L) after 3-5 days of training in laboratory. The inner basin was then placed in the outer basin containing synthetic wastewater. The quality index of the synthetic wastewater is shown in Table 2 and is similar to the pollutant concentration of domestic wastewater in southeast China.

The synthetic wastewater was added during the operation to ensure the normal growth of the *P. australis*, and 8 L of wastewater was added into outer basin in May 2017. Water was sampled from the CWs after a 7-day hydraulic retention time (HRT) to analyze NH_4^+ -N content as well as dissolved oxygen (DO) content on October 20, 2017. The DO content of CWs was measured by dissolved oxygen analyzer (JPB-607A, INESA, China).

The rhizobag was taken from CWs on November 5, 2017. The substrate and *P. australis* were taken from root bags, respectively. Some substrate samples were used to measure the exchanged NO_3^- and NH_4^+ . Other substrate samples were prepared to determine nitrification and denitrification potentials. *P. australis* were washed thoroughly with tap water and cleaned with deionized water. Fresh *P. australis* was dried at 60 °w in an oven for 48 h to a constant weight. The roots (under-ground), stems and leaves (above-ground) samples were sub-sampled and ground to powder for the determination of the N concentrations.

Analytical methods

Measure removal efficiency of NH4+-N in wastewater

The NH_4^+ -N concentration in wastewater was analyzed according to standard methods (APHA 1992), and its removal efficiency was then calculated by the following equation:

$$R = \frac{100(I_C I_V - E_C E_V)}{I_C I_V}$$
(1)

where *R* is removal efficiency (%), Iv is initial volume (L), Ic is initial concentration (mg/L), E_v is remaining volume (L), E_c is remaining concentration (M).

Measure N content of plants

Subsamples of 0.2 g were digested with H_2SO_4 - H_2O_2 at 260 °C. The N concentration in the digest was then determined by the Kjeldahl method (Lu 1999).

Measure NH₄⁺-N and NO₃⁻-N content in substrates

The concentration of NH_4^+ -N and NO_3^- -N in substrates was determined after mixing 10 g freshly sampled substrate with 50 mL 2 M KCl solution and allowing it to equilibrate overnight (Robertson et al. 1999). The supernatant was filtered and

 Table 1
 Type of constructed

 wetland

System of constructed wetland	Type of biochar	Earthworm	
System with only sand as substrate (S)	-	-	
System with sand and <i>E. fetida</i> $(S + E)$	-	E. fetida	
System with a 9:1 (v/v) mixture of sand and RHB as substrate (S + RHB)	RHB	-	
System with sand, RHB, and E. fetida (S + RHB + E)	RHB	E. fetida	
System with a 9:1 (v/v) mixture of sand and CSB as substrate (S + CSB)	CSB	-	
System with sand, CSB, and <i>E. fetida</i> (S + CSB + E)	CSB	E. fetida	
System with a 9:1 (v/v) mixture of sand and WB as substrate (S + WB)	WB	-	
System with sand, WB, and <i>E. fetida</i> $(S + WB + E)$	WB	E. fetida	

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then analyzed for NH_4^+ -N and NO_3^- -N contents. NH_4^+ -N was determined at a wavelength of 697 nm using spectrophotometer according to the salicylic acid method (Ministry of Environmental Protection 2006). NO_3^- -N contents were determined at a wavelength of 220 and 275 nm in ultraviolet spectrophotometer (UV-752N, YOKE, China).

Determine nitrification and denitrification potentials in substrates

Nitrification potentials were carried out as described by Hart et al. (1994) with slight modifications. Ten grams of substrate were put in 500-mL Erlenmeyer flasks containing 100 mL of pH 7.2 of incubation solution (in M) (volume ratio was 3:7:30 for 0.2 KH₂PO₄, 0.2 K₂HPO₄, and 0.05 (NH₄)₂SO₄). Erlenmeyer flasks were plugged by absorbent cotton and were rotary shaken at 25 °. (140 rpm). The concentration of NO₃⁻-N was measured after 48 h incubation. Nitrification potentials were analyzed using the following equation:

$$N = \frac{1000(C_1 - C_2)(V_1 + V_2)}{TM}$$
(2)

where *N* is nitrification potential (mg/(kg h)), C_2 and C_1 were NO₃⁻-N concentration after 48-h incubation solution and initial concentration of substrate (mg/L). V_1 and V_2 represent the volume of incubation solution and the volume of water in substrate (L). *T* is incubation time. *M* is dry weight of substrate (g).

Denitrification potentials were carried out as described by Mathew et al. (1982). Briefly, 10 g of substrate was put in 500-mL Erlenmeyer flasks containing 100 mL of pH 7.2 of incubation solution (in M) (volume ratio was 3:7:30:10 for 0.2 KH₂PO₄, 0.2 K₂HPO₄, 0.03 KNO₃, and 0.02 glucose). The C/

Indicator	COD _{cr}	NH4 ⁺ -N	TN
Concentration (mg/L)	400-481.3	30.4–32.2	59.9–69.6

N ratio of incubation solution was 3:1. Erlenmeyer flasks were plugged by rubber and incubated at 20 °r. The denitrification potential was analyzed according to the change in nitrate concentration. Denitrification potentials use a similar approach as Eq. (2).

Statistical analysis

Data was processed with Microsoft Office Excel 2016, OriginLab 2018, and SPSS 22.0 software. The effect of different treatments was tested by analysis of one-way ANOVA, and comparisons were made using mean \pm standard deviation (LSD's multiple range). Significance values are for $P \le 0.05$ unless stated otherwise. The correlation between removal efficiency of NH₄⁺-N, NH₄⁺-N, NO₃⁻-N, microbial nitrification and denitrification potential in substrate, nitrogen content of above-ground and under-ground parts of plants, and DO content were also analyzed in this study.

Results and discussion

Effect of biochar and earthworm on DO content in CWs

Content of DO in different CWs ranged from 3.5 to 5.9 mg/L (Fig. 2). Compared to the system with only sand as substrate, the addition of RHB and WB significantly increased the concentration of DO by 21.4% and 25.7% (P < 0.05), respectively. The content of DO significantly increased by 31.4%, 30.6%, 25.3%, and 34.1% (P < 0.05), respectively with the addition of earthworm into the four systems containing only sand, sand and RHB, sand and CSB, and sand and WB. Therefore, the addition of both biochar and earthworm may significantly improve the DO content of CWs. Some DO in CWs come from atmosphere by atmospheric diffusion. Kizito et al. (2017) demonstrated that there is higher total pore volume and micropore volume for biochar than ceramsite. Therefore, the increasing DO content in CWs could be related



Fig. 2 The concentration of dissolved oxygen (DO) in different constructed wetlands. S, S + RHB, S + CSB, and S + WB represent constructed wetlands with different substrates of sand, sand with rice husk biochar, sand with coconut shell biochar, sand with wood biochar, respectively. S + E, S + E + RHB, S + E + CSB, and S + E + WB represent constructed wetlands with earthworms added and substrates of sand, sand with rice husk biochar, sand with coconut shell biochar, sand with wood biochar, respectively. Different lowercase letters indicate the difference with significance value for P < 0.05. Data are means \pm SD (n = 3)

to higher pore volume of biochar than those of sand. In addition, porosity of CWs increased by earthworm excavation resulted in increasing DO content.

Effect of biochar and earthworm on nitrogen content of plant

The effect of different biochars on N content in above-ground part of P. australis varied (Fig. 3a). The addition of RHB and WB significantly increased N content by 54.3% and 69.4% (P < 0.05), respectively. Compared with the CWs with only sand (S), the addition of different biochars (RHB, CSB, and WB) increased the N content in the under-ground part of P. australis by 9.0%, 22.1%, and 22.9% (Fig. 3b), respectively. Addition of earthworms into CWs with only sand as substrate increased the N content of above-ground part of P. australis by 28.6%. This result was similar to those of Xu et al. (2013a, b), who demonstrated that addition of earthworms into CWs increased the N content of plant. However, the N content in above-ground part of P. australis was increased on average by 7.56% when earthworms were added to CWs with biochar. Mahtab et al. (2012) showed that content of nitrogen of biochar is 1.14–1.91%, but N is not usually found in sand. Therefore, the response of increased concentration of N in above-ground part of P. australis to addition of earthworm was higher in CWs with only sand as substrate than with sand and biochar. Similarly, the increased N concentration in the under-ground part of *P. australis* was lower when earthworms were added into CWs with only sand as substrate or with sand and biochar. Results showed that the effect of biochar and earthworm on N content of *P. australis* differs.

Addition of biochar into CWs obviously increased the N content of *P. australis*. Chan et al. (2007) showed that biochar could improve soil properties, such as porosity. Blackwell et al. (2010) observed that biochar could increase the parasitism of mycorrhizal fungi, thus enhancing the absorption of nutrients. In addition, Kolton et al. (2011) reported that the addition of biochar could effectively alter the population structure of soil microbes, which contributes to plant growth. Therefore, the increasing N content of *P. australis* could be related to improving substrate properties, or microorganism growth after addition of biochar into CWs. The N content of *P. australis* was not obviously increased by the addition of earthworm into CWs with biochar.

It can be seen from Fig. 3, that the addition of biochar into the constructed wetland systems increased the N content of *P. australis*, especially in the above-ground part. Zhu et al. (2019) demonstrated that addition of biochar into soil will increase migration of NO_3^- -N from roots to shoots. Therefore, the higher level of N in the above-ground part compared to the under-ground part of *P. australis* can be attributed to greater migration of NO_3^- -N from roots to shoots due to adding biochar to the constructed wetland system. These results therefore suggest that the harvesting of plants could be an effective and sustainable way of removing N from wastewater in constructed wetlands due to the higher N content in above-ground than under-ground part of *P. australis*.

Effect of biochar and earthworm on nitrification potentials in substrates

Compared to the CWs with only sand as substrate, the nitrification potential significantly increased by 45.5% and 226.2% (P < 0.05), respectively for CWs with addition of RHB and WB (Fig. 4). Zhou et al. (2018) found that adding wheat straw biochar into metal-contaminated farmland significantly increased the nitrification potential. Ball et al. (2010) demonstrated that the content of biochar was significantly positively correlated with the number of ammoniated bacteria in soil. Deluca et al. (2006) reported that biochar could adsorb some nitrification inhibitors in soil, thereby alleviating the inhibition of nitrification and promoting the nitrification process of microorganisms. In this study, it can be concluded that biochar can enhance the nitrification potential of substrate in CWs.

The nitrification potentials increased by 62.4%, 151.9%, 61.8%, and 26.4%, respectively for addition of earthworm into the CWs with only sand as substrate, sand and RHB, sand and CSB, and sand and WB. These results are similar to those of Xu et al. (2013a, b), who demonstrated that nitrification



Fig. 3 Nitrogen content in above-ground (**a**) and under-ground (**b**) part of *P. australis.* S, S + RHB, S + CSB, and S + WB represent constructed wetlands with only sand as substrate, sand and rice husk biochar, sand and coconut shell biochar, and sand and wood biochar, respectively. S + E, S + E + RHB, S + E + CSB, and S + E + WB represent constructed

increased with the addition of earthworm into CWs. Increased aeration and consequently DO concentration from the addition of both biochar and earthworm into CWs resulted in higher rates of nitrification (Fig. 2).



Fig. 4 Nitrification potentials in substrates. S, S + RHB, S + CSB, and S + WB represent constructed wetlands with different substrates of sand, sand with rice husk biochar, sand with coconut shell biochar, and sand with wood biochar, respectively. S + E, S + E + RHB, S + E + CSB, and S + E + WB represent constructed wetlands with earthworms added and substrates of sand, sand with rice husk biochar, sand with coconut shell biochar, and sand with wood biochar, respectively. Different lowercase letters indicate the difference with significance value for P < 0.05. Data are means \pm SD (n = 3)



wetlands with sand and earthworms, with sand, earthworm and rice husk biochar, with sand, earthworm and coconut shell biochar, and with sand, earthworm, and wood biochar, respectively. Different letters indicate the difference with significance value for P < 0.05. Data are means \pm SD (n = 3)

Effect of biochar and earthworm on denitrification potentials in substrates

Denitrification potentials in substrate of different CWs range from 4.38 to 19.41 mg/(kg h) (Fig. 5). The denitrification potential significantly increased by 184.7%, 127.2%, and 270.6%, respectively with the addition of RHB, CSB, and WB into CWs (P < 0.05). The denitrification process mainly refers to the biochemical process in which bacteria in soil convert nitrate (NO_3) into nitrogen (N_2) through a series of intermediate products (NO2, NO, N2O). The porous structure of biochar can provide a suitable habitat for the growth and reproduction of denitrifying bacteria (Shamim and Whalen 2016), thereby enhancing the denitrification process. Results suggest that biochar can significantly affect denitrification potential in substrates in CWs. The denitrification potentials increased by 23.2%, 28.6%, 28.1%, and 19.7%, respectively for addition of earthworms into CWs with only sand as substrate, sand and RHB, sand and CSB, and sand and WB. However, in this study, addition of earthworm did not significantly increase denitrification potential.

Effect of biochar and earthworm on ammonia and nitrate content in substrate

Figure 6 a shows that the NH₄⁺-N content in substrate of different CWs ranges from 0.36 to 0.48 mg/kg. Addition of RHB and WB into the CWs significantly decreased NH₄⁺-N content in substrate (P < 0.05). The content of NH₄⁺-N in



Fig. 5 Denitrification potentials in substrate. S, S + RHB, S + CSB, and S + WB represent constructed wetlands with different substrates of sand, sand with rice husk biochar, sand with coconut shell biochar, and sand with wood biochar, respectively. S + E, S + E + RHB, S + E + CSB, and S + E + WB represent constructed wetlands with earthworms added and substrates of sand, sand with rice husk biochar, sand with coconut shell biochar, and sand with wood biochar, respectively. Different lowercase letters indicate the difference with significance value for P < 0.05. Data are means \pm SD (n = 3)

substrate was decreased on average by 8.9% with the addition of earthworm into CWs. Wu et al. (2015) reported that NH_4^+ -N in CWs is mainly influenced by DO content, and NH_4^+ -N can be easily converted to other forms of nitrogen through nitrification and oxidation reactions where sufficient DO is present. Therefore, the addition of biochar and earthworm into



Fig. 6 The content of NH_4^+ -N (a) and NO_3^- -N (b) in substrate. S, S + RHB, S + CSB, and S + WB represent constructed wetlands with different substrates of sand, sand with rice husk biochar, sand with coconut shell biochar, and sand with wood biochar, respectively. S + E, S + E + RHB, S + E + CSB, and S + E + WB represent constructed

the CWs decreased the NH_4^+ -N content, which could be related to increasing DO content (Fig. 2).

According to Fig. 6b, the addition of WB into the CWs significantly increased NO₃⁻-N content (P < 0.05). Knowles et al. (2011) demonstrated that biochar can reduce the leaching of NO₃⁻-N in soil. Mizuta et al. (2004) also found that biochar can reduce NO₃⁻-N leaching by direct adsorption. Therefore, the increasing NO₃⁻-N content after the addition of biochar into the CWs could be related to NO₃⁻-N adsorption by biochar.

The addition of earthworm significantly increased NO₃⁻-N content by 54.9% and 38.4%, respectively for CWs with RHB and WB (P < 0.05). Earthworm activity enhanced DO content in the substrate (Fig. 2), which helps to stimulate the process of nitrification, and thereby NH₄⁺-N can be rapidly oxidized and converted to NO₃⁻-N under aerobic conditions. As shown in Fig. 6, the addition both earthworm and biochar into CWs decreased NH₄⁺-N content, but increased NO₃⁻-N content. This is similar to the findings of Fu et al. (2019), who found that with increasing biochar application rate into soil, the content of NH₄⁺-N decreased, but the content of NO₃⁻-N increased.

Effect of biochar and earthworm on removal efficiency of ammonia nitrogen

The addition of RHB, CSB, and WB into CWs significantly increased the removal efficiency of NH_4^+ -N by 8.4%, 6.2%, and 9.6% (P < 0.05) (Fig. 7), respectively. These results show that the addition of biochar can significantly improve the removal efficiency of NH_4^+ -N. The observed increase in



wetlands with earthworms added and substrates of sand, sand with rice husk biochar, sand with coconut shell biochar, and sand with wood biochar, respectively. Different lowercase letters indicate the difference with significance value for P < 0.05. Data are means \pm SD (n = 3)



Fig. 7 The removal efficiency of NH_4^+ -N in wastewater. S, S + RHB, S + CSB, and S + WB represent constructed wetlands with different substrates of sand, sand with rice husk biochar, sand with coconut shell biochar, and sand with wood biochar, respectively. S + E, S + E + RHB, S + E + CSB, and S + E + WB represent constructed wetlands with earthworms added and substrates of sand, sand with rice husk biochar, sand with coconut shell biochar, and sand with wood biochar, respectively. Different lowercase letters indicate the difference with significance value for P < 0.05. Data are means \pm SD (n = 3)

removal of NH_4^+ -N associated with the addition of biochar could be attributed to (1) biochar improving the physical and chemical properties of substrates, thus promoting the growth of plants, which is beneficial to improving N uptake by *P. australis* (Fig. 3), (2) biochar producing a carbon resource and habitat for microorganisms, thus enhancing nitrification and denitrification potentials (Figs. 4 and 5). As shown in Fig. 7, the addition of earthworm into CWs increased removal efficiency of NH_4^+ -N by 5.3% on average. The improved removal efficiency of NH_4^+ -N was related to increasing DO content caused by earthworm action in the CWs.

Table 3 Correlation coefficient between removal efficiency of NH_4^+-N , NH_4^+-N , and NO_3^--N in substrate, nitrification and denitrification in substrate, nitrogen content in both above and below ground part of *P. australis*, and DO in constructed wetland

Response of removal efficiency of NH₄⁺-N to DO, nitrification, and denitrification potential

The correlation coefficients between the removal efficiency of NH₄⁺-N and DO in wastewater, NH₄⁺-N, NO₃⁻-N, nitrification and denitrification potential in substrate, and nitrogen content of uptake by *P. australis* are shown in Table 3. The removal efficiency of NH₄⁺-N in wastewater is significantly negatively correlated with the content of NH4+-N in substrate (P < 0.01). However, it is significantly positively correlated with the DO content, nitrification and denitrification potential, and nitrogen content of above-ground part of P. australis (P <0.05). Maltais-Landry et al. (2009) reported that artificial aeration supplied oxygen to the wetland substrate, stimulating nitrification and resulting in a higher removal of NH₄⁺-N. Kasak et al. (2018) showed that the growth of Lythrum Salicaria was improved by biochar, and NH₄⁺ removal benefited from plant uptake and the provision of improved conditions (in terms of oxygen secretion and root surface) for nitrification. Therefore, the addition of biochar and earthworm into constructed wetland systems increased the removal efficiency of NH₄⁺-N, which can be attributed to higher DO, nitrification and denitrification potential, and uptake N by P. australis.

Conclusions

The addition of biochar and earthworms into CWs increased DO content, nitrification and denitrification potentials in substrates, and nitrogen accumulation in above-ground of *P. australis*. The addition of biochar and earthworm into CWs increased the removal efficiency of NH_4^+ -N, and this was related to enhanced DO, nitrification and denitrification potentials, and N uptake by plant. The addition of biochar and

Index	REN	NH4 ⁺ -N (S)	NO ₃ ⁻ -N (S)	Nitri	Denitri	N (A-G)	N (U-G)	DO
REN	1							
NH4 ⁺ -N (S)	- 0.949**	1						
NO3 ⁻ -N (S)	0.669	- 0.545	1					
Nitri	0.842*	- 0.815*	0.860*	1				
Denitri	0.840*	-0.856*	0.504	0.820*	1			
N (A-G)	0.819*	- 0.736	0.716	0.744	0.473	1		
N (U-G)	0.623	- 0.634	0.410	0.463	0.512	0.256	1	
DO	0.845*	- 0.737	0.843*	0.749	0.527	0.790*	0.717	1

REN represents removal efficiency of NH_4^+ -N in wastewater. NH_4^+ -N (S) and NO_3^- -N (S) represent NH_4^+ -N and NO_3^- -N in substrate, respectively. Nitri and Denitri represent nitrification potential and denitrification potential, respectively. N (A-G) and N (U-G) represent nitrogen content of above and under-ground part of *P. australis*, respectively. DO represent dissolved oxygen. ** and * represent significant values of $P \le 0.01$ and $P \le 0.05$, respectively earthworm could therefore be an effective option to improve removal of NH_4^+ -N in CWs.

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References

- APHA (1992) Standard methods for the examination of water and wastewater, eighteenth edn. American Public Health Association, Washington, DC
- Ball P, MacKenzie MD, Deluca TH, Holben WE (2010) Wildfire and biochar enhance nitrification and ammonium-oxidizing bacterial abundance in dry montane forest soils. Environ Qua 139(4):1243– 1253
- Blackwell P, Krull E, Bulter G, Herbert A, Solaiman Z (2010) Effect of banded biochar on dry land wheat production and fertiliser use in southwestern Australia: an agronomic and economic perspective. Aust J Soil Res 48(7):531–545
- Chan KY, Van ZL, Meszaros I, Downie A, Joseph SD (2007) Agronomic values of green waste biochar as a soil amendment. Aust J Soil Res 45:629–634
- Cheng CH, Ma HL, Su YL, Deng YQ, Feng J, Xie JW, Chen XL, Guo ZX (2019) Ammonia toxicity in the mud crab (*Scylla paramamosain*): the mechanistic insight from physiology to transcriptome analysis. Ecotoxicol Environ Saf 179:9–16
- Davison L, Headley TR, Pratt K (2005) Aspects of design, structure and performance and operation of reed beds-eight years' experience in northeastern New South Wales, Australia. Water Sci Technol 51(10):129–138
- de Rozari P, Greenway M, El Hanandeh A (2018) Nitrogen removal from sewage and septage in constructed wetland mesocosms using sand media amended with biochar. Ecol. Eng. 111:1–10
- Deluca TH, MacKenzie MD, Gundale MJ, Holben WE (2006) Wildfireproduced charcoal directly influences nitrogen cycling in ponderosa pine forests. Soil Sci Soc Am J 70(2):448–453
- Egnew N, Renukdas N, Ramena Y, Yadav AK, Kelly AM, Lochmann RT, Sinha AK (2019) Physiological insights into largemouth bass (*Micropterus salmoides*) survival during long-term exposure to high environmental ammonia. Aquat Toxicol 207:72–82
- Feng L, Wang R, Jia L, Wu H (2020) Can biochar application improve nitrogen removal in constructed wetlands for treating anaerobicallydigested swine wastewater. Chem Eng J DOI 379:122273. https:// doi.org/10.1016/j.cej.2019.122273
- Fu Q, Yan JW, Li H, Li TX, Hou RJ, Liu D, Ji Y (2019) Effects of biochar amendment on nitrogen mineralization in black soil with different moisture contents under freeze-thaw cycles. Geoderma 353:459– 467
- Goldshleger N, Grinberg A, Harpaz S, Shulzinger A, Abramovich A (2018) Real-time advanced spectroscopic monitoring of ammonia concentration in water. Agricultural Engineering 83:103-108
- Hart SC, Stark JM, Daridson EA, Firestone MK (1994) Nitrogen mineralization, immobilization, and nitrification. In: Weaver RW, Angle JS, Bottomley PS (eds) Method of Soil Analysis, Part 2. Microbiological and Biochemical Properties. SSSA Book Series, no. 5. SSSA, Madison, pp 987–1016
- Ilyas H, Masih I (2017) The performance of the intensified constructed wetlands for organic matter and nitrogen removal: a review. J Environ Manag 198:372–383

- Kasak K, Truu J, Ostonen I, Sarjas J, Oopkaup K, Paiste P, KoivVainik M, Mander Ü, Truu M (2018) Biochar enhances plant growth and nutrient removal in horizontal subsurface flow constructed wetlands. Sci Total Environ 639:67–74
- Kizito S, Lv T, Wu S, Ajmal Z, Luo H, Dong R (2017) Treatment of anaerobic digested effluent in biochar-packed vertical flow constructed wetland columns: role of media and tidal operation. Sci Total Environ 592:197–205
- Knowles OA, Robinson BH, Contangelo A, Clucas L (2011) Biochar for the mitigation of nitrate leaching from soil amended with biosolids. Sci Total Environ 409(17):3206–3210
- Kolton M, Harel YM, Pasternak Z, Graber ER, Elad Y, Cytryn E (2011) Impact of biochar application to soil on the root-associated bacterial community structure of fully developed greenhouse pepper plants. Appl Environ Microb 77(14):4924–4930
- Kuno K, Truu J, Ostonen I, Sarjas J (2018) Biochar enhances plant growth and nutrient removal in horizontal subsurface flow constructed wetlands. Sci Total Environ 639:67–74
- Li HZ, Wang S, Ye JF, Xu ZX, Jin W (2011) A practical method for the restoration of clogged rural vertical subsurface flow constructed wetlands for domestic wastewater treatment using earthworm. Water Sci Technol 63(2):283–290
- Li J, Fan J, Zhang J, Hu Z, Liang S (2018) Preparation and evaluation of wetland plant-based biochar for nitrogen removal enhancement in surface flow constructed wetlands. Environ Sci Pollut Res 25: 13929–13937
- Liew HJ, Sinha AK, Nawata CM, Blust R, Wood CM, Boeck G (2013) Differential responses in ammonia excretion, sodium luxes and gill permeability explain different sensitivities to acute high environmental ammonia in three fresh water teleosts. Aquat Toxicol 126: 63–76
- Lu RK (1999) Agrochemical analyzed method of soil. Agricultural Science and Technology Press of China, Beijing
- Lu S, Zhang X, Wang J, Pei L (2016) Impacts of different media on constructed wetlands for rural household sewage treatment. J Clean Prod 127:325–330
- Lu X, Luan S, Dai P, Luo K, Chen B, Cao B, Sun L, Yan Y, Kong J (2019) Insights into the molecular basis of immunosuppression and increasing pathogen infection severity of ammonia toxicity by transcriptome analysis in pacific white shrimp *Litopenaeus vannamei*. Fish Shellfish Immunol 88:528–539
- Mahtab A, Sang SL, Dou XM, Dinesh M, Sung JK, Yang JE, Yong SO (2012) Effects of pyrolysis temperature on soybean stover and peanut shell derived biochar properties and TCE adsorption in water. Bioresour Technol 118:536–544
- Maltais-Landry G, Maranger R, Brisson J, Chazarenc F (2009) Nitrogen transformations and retention in planted and artificially aerated constructed wetlands. Water Res 43:535–545
- Manyà JJ, Azuara M, Manso JA (2018) Biochar production through slow pyrolysis of different biomass materials: seeking the best operating conditions. Biomass Bioenerg 117:115–123
- Mathew K, Newman PWG, Ho GE (1982) Groundwater recharge with secondary sewage effluent. Australian Water Resource Council. Technical Paper, No.71, Canberra
- Mizuta K, Matsumoto T, Hatate Y, Nishihara K, Nakanishi T (2004) Removal of nitrate–nitrogen from drinking water using bamboo powder biochar. Bioresour Technol 195(3):255–257
- Robertson GP, Wedin D, Groffman PM, Blair JM, Holland EA, Nadelhoffer KJ, Harris D (1999) Soil carbon and nitrogen availability: nitrogen mineralization, nitrification, and soil respiration potentials. In: Robertson GP, Coleman DC, Bledsoe CS, Sollins P (eds) Standard soil methods for long-term ecological research. Oxford University Press, New York, pp 258–271
- Shamim G, Whalen JK (2016) Biochemical cycling of nitrogen and phosphorus in biochar-amended soils. Soil Biol Biochem 103:1–15

- Abedi T, Mojiri A (2019) Constructed wetland modified by biochar/ zeolite addition for enhanced wastewater treatment. Environ Technol Inno 16:100472. https://doi.org/10.1016/j.eti.2019.100472
- Taylor M, Clarke WP, Greenfield PF (2003) The treatment of domestic wastewater using small-scale vermicompost filter beds. Ecol Eng 21:197–203
- Ueno Y, Fujii M (2001) Three years operating experience selling recovered struvite from full scale plants. Environ Technol 22(11):1373– 1381
- Wang XN, Li J, Chen J, Cui L, Li WW, Gao XY, Liu ZT (2020) Water quality criteria of total ammonia nitrogen (TAN) and un-ionized ammonia (NH₃-N) and their ecological risk in the Liao River, China. Chemosphere 243:125328
- Wu H, Fan J, Zhang J, Ngo HH, Guo W, Hu Z, Liang S (2015) Decentralized domestic wastewater treatment using intermittently aerated vertical flow constructed wetlands: impact of influent strengths. Bioresour Technol 176:163–168
- Xu DF, Cao JM, Li YX, Howard A, Yu KW (2019) Effect of pyrolysis temperature on characteristics of biochars derived from different feedstocks: a case study on ammonium adsorption capacity. Waste Manage 87:652–660
- Xu DF, Li YX, Howard A, Guan YD (2013b) Effect of earthworm Eiseniafetida and wetland plants on nitrification and denitrification

potentials in vertical flow constructed wetland. Chemosphere 92: 201-206

- Xu DF, Li YX, Howard A (2013a) Influence of earthworm *Eiseniafetida* on removal efficiency of N and P in vertical flow constructed wetland. Environm Sci Pollut R 20:5922–5929
- Zhou HM, Wang P, Chen D, Shi GL, Cheng K, Bian RJ, Liu XY, Zhang XH, Zheng JF (2018) Short-term biochar manipulation of microbial nitrogen transformation in wheat rhizosphere of a metal contaminated Inceptisol from North China plain. Sci Total Environ 640-641: 1287–1296
- Zhou L, Claude EB (2015) An assessment of total ammonia nitrogen concentration in Alabama USA ictalurid catfish ponds and the possible risk of ammonia toxicity. Aquaculture 437:263–269
- Zhou X, Wang XZ, Zhang H, Wu HM (2017) Enhanced nitrogen removal of low C/N domestic wastewater using a biochar amended aerated vertical flow constructed wetland. Bioresour Technol 241:269–275
- Zhu Y, Li H, Wu Y, Yin XA, Zhang GX (2019) Effects of surfacemodified biochars and activated carbon on the transformation of soil inorganic nitrogen and growth of maize under chromium stress. Chemosphere 227:124–132

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