RESEARCH ARTICLE



Determination and environmental risk assessment of synthetic musks in the water and sediments of the Jiaozhou Bay wetland, China

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Abstract

Human activity in estuarine areas has resulted in pollution of the aquatic environment, but little is known about the levels of synthetic musks (SMs) in river water and sediments in estuarine areas. This study investigated the concentrations and distribution of SMs in the Jiaozhou Bay wetland, including celestolide, phantolide, traseolide, galaxolide (HHCB), tonalide (AHTN), musk xylene and musk ketone (MK). The SMs HHCB, AHTN and MK were detected at concentrations of 10.7–208, not detected (ND)–59.2 and ND–13.6 ng/L, respectively, in surface water samples and 13.1–27.3, 3.06–14.5 and 1.33–18.8 ng/g (dry weight; dw), respectively, in sediment samples. Based on the calculated total organic carbon (TOC) concentrations, there was no significant correlation between SMs and TOC in sediment samples (p > 0.05). The hazard quotients were 0.204, 0.386 and 0.059 for AHTN, HHCB and MK, respectively, which indicated no serious environmental impact, because these values are all less than 1. The concentrations of SMs decreased as the distance to the Xiaojianxi refuse landfill increased in both surface water and sediments. Compared with previous studies, the concentration of SMs in the Jiaozhou Bay wetland was relatively high. Therefore, more attention should be paid to SMs because of their persistent impact on human health and the environment.

 $\label{eq:Keywords} Keywords \ Galaxolide (HHCB) \cdot Tonalide (AHTN) \cdot Musk \ ketone (MK) \cdot Total \ organic \ carbon (TOC) \cdot Hazard \ quotient (HQ) \ (HQ) \ (HCB) \ ($

Introduction

Synthetic musks (SMs), including nitro musks (NMs), polycyclic musks (PCMs), alicyclic musks and macrocyclic musks, are organic chemicals that are widely used as fragrance additives in many household products and consumer goods (Heberer 2003). Among the PCMs, galaxolide (1,3,4,6,7,8hexahydro-4,6,6,7,8,8,-hexamethylcyclopenta[g]-2benzopyran; HHCB) and tonalide (6-acetyl-1,1,2,4,4,7hexamethyltetraline; AHTN) are the most heavily produced and used worldwide, almost completely replacing NMs. The use of musk xylene and musk ketone (MK) has decreased because of their potential toxicity and bioaccumulation

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Ling Wang qddxhjkx520@163.com (Nakata et al. 2007), but they can still be detected in the effluent from some wastewater treatment plants (WWTPs) in China (Liu et al. 2014).

HHCB and AHTN have potential oestrogen effects and low-dose effects (Yamauchi et al. 2008). The predicted no effect concentrations (PNECs) of HHCB and AHTN are 6.80×103 and 3.50×103 ng/L in surface water and 8.40×103 and 5.20×103 ng/g (dry weight; dw) in sediments, respectively (HERA 2004). The concentrations of HHCB and AHTN in WWTP sewage sludge reflect the production and consumption patterns of SMs in an area.

According to a toxicology report, in most parts of China, SMs were detected at concentrations ranging from below the limit of quantification (LOQ) to 4.14×104 ng/g for HHCB and below the LOQ to 2.20×104 ng/g (dw) for AHTN (Liu et al. 2014). Removal by WWTPs is one of the main elimination processes for HHCB and AHTN, with removal efficiencies of 80.0-90.0% (Luo et al. 2014). Many SMs exist in domestic sewage, and as a result of their incomplete removal by WWTPs or direct dumping, SMs are discharged into lakes and rivers (Clara et al. 2010). Due to their extensive production and use and their persistence, lipophilicity and bioaccumulation (Kannan et al. 2005), SMs have been widely

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detected in the environment, including surface waters (Hu et al. 2011; Reiner and Kannan 2011), snow (Hu et al. 2012), sediments and sewage sludges (Guo et al. 2010; Villa et al. 2012), aquatic biota (Moon et al. 2011) and even breast milk and blood (Moon et al. 2012).

Some studies of SMs in lakes and laboratory studies have been conducted, but little is known about SMs in wetlands, which is essential for addressing the connection between SM pollution and human health. Therefore, we analysed SM pollution in the Jiaozhou Bay wetlands. When wastewater flows through a wetland, complex migration and transformation processes may occur, including evaporation, photochemical degradation, adsorption, accumulation, plant uptake and microbial degradation (Reyes-Contreras et al. 2012). Photochemical degradation has almost no effect on HHCB, and in spring and winter, HHCB is removed mainly by the evaporation of surface water (Quednow and Puttmann 2008). HHCB has a longer photolysis half-life (~135 h) than that of AHTN (~4 h) (Buerge et al. 2003). In addition, MK undergoes strong photochemical degradation in surface waters. After exposure to light for 1 h, the MK concentration decreased to 1% of the original level (Butte et al. 1999). In sediments, AHTN is more readily absorbed than is HHCB (Wang et al. 2010). The major mechanism of HHCB and AHTN removal is absorption by organic matter in the substrate due to the high log Kow values. The reported removal rates of HHCB and AHTN exceeded 80.0% in subsurface flow constructed wetlands and ranged from 70.0% to 90.0% in a vertical flow constructed wetland (Hijosa-Valsero et al. 2011; Zhang et al. 2014).

The objectives of this study were to (1) investigate and characterise the levels of SMs in the Jiaozhou Bay wetland and compare them with global levels, (2) conduct a preliminary environmental risk assessment, (3) examine the relationships between SMs and total organic carbon (TOC) in this area, and (4) analyse the removal of SMs and the effect of wetlands on SM pollution.

Materials and methods

Sampling site

Sediment samples were collected from the Jiaozhou Bay wetland in Qingdao, Shandong Province, eastern China. This is a typical estuary wetland covered with reeds and strongly influenced by the tides. The Dagu River is the largest water system in the Jiaozhou Bay wetland, and Taoyuan River is one of its branches. As the largest estuary bay wetland in the Shandong Peninsula, the Jiaozhou Bay wetland is a heavily industrialised and urbanised region involving a large number of industries. The Xiaojianxi landfill is located in the upper reach of the Taoyuan River, and it processes approximately 1710 tons of rubbish collected from Qingdao City every day. Consequently, rubbish leaching liquor mixed with large amounts of pollutants flows into the Jiaozhou Bay wetland. The Xiaojianxi refuse landfill may be the main source of pollution in this area. It provides an ideal model to investigate the contamination and spatial distribution of SMs in water and sediments from wetlands.

Sampling

A total of 18 sediment samples and 14 surface water samples were collected in March 2014. When we collected the samples, the riverbed at sites 15–18 was bone dry, and therefore water samples could not be collected from these sites. Figure 1 is a map of the sampling sites, which are located mainly in the Taoyuan (sites 1–7) and Dagu (sites 8–18) Rivers. These rivers join downstream of site 7. Both surface water samples and sediment samples were collected at sites 15–18, while only sediment samples were collected at sites 15–18.

The sediment samples were collected using a spade (preconditioned with hexane and deionised water) at a depth of approximately 10 cm. Three parallel samples were collected at each site. After collection, the samples were immediately wrapped in several layers of aluminium foil and placed in sealed plastic bags to avoid irradiation by light. Then, the samples were placed in an icebox before transporting to the laboratory and storing at -20 °C until analysis. When collecting the surface water samples, we divided the river section into three parts, and three samples of the same volume were collected from each part and then mixed as a composite sample for each water sampling site.

Methanol was added to all surface water samples to prevent decomposition of organic material by microorganisms in the water. Samples were extracted within 5 days of collection. The samples were filtered through glass-fibre film (0.45 μ m) and stored at 4 °C before treatment. The sediment samples were frozen for 12 h and dried in a vacuum freeze drier for at least 48 h. Then, the samples were crushed and sieved through a 60-mesh screen and stored at – 20 °C until analysis.

Sample preparation and analysis

Chemicals

Five PCMs standards, including celestolide (ADBI), phantolide (AHMI), traseolide (ATII), HHCB, AHTN, and two NMs standards, including MX, MK, and the internal standard, C13 isotope labelled hexachlorobenzene (HCB-C13), were all purchased from Dr. Ehrenstorfer, (Augsburg, Germany). Surrogate standard, Deuterated fluoranthene (d10-fluoranthene) and solid phase extraction (SPE) column (C-18) were purchased from Supeclo (Bellefonte, PA, USA). All solvents were of the high-performance liquid chromatography (HPLC) grade, including methanol, n-hexane (HEX),



Fig. 1 The sampling stations in the Jiaozhou Bay wetland

dichloromethane (DCM) and methanol. Anhydrous sodium sulphate was baked at 450 °C for 4 h. Silica gel (100–200 mesh) and neutral Al_2O_3 were extracted with HEX/DCM (1:1, *V/V*) for 24 h, baked at 180 and 250 °C for 12 h, respectively, and wetted with distilled water to reduce their activation prior to use. Copper powder was activated by 35%–37% hydrochloric acid.

Surface water samples

The SPE columns were preconditioned with 5 mL DCM/ methanol (1:1, *V*/*V*), 5 mL methanol and then 5 mL distilled water. Surface water samples (400 mL) were spiked with 5 μ L of 1 μ g/mL d10-fluoranthene, and loaded onto the SPE column at a flow rate of 5–10 mL/min. Thereafter, the SPE columns were dried suing nitrogen, before target analytes were eluted with 6 mL HEX and 4 mL HEX/DCM (1:1, *V*/*V*). The eluates were combined and concentrated to 0.5 mL. Finally, 5 μ L of 1 μ g/mL HCB-C13 was added to the extract before analysis.

Sediment samples

The pretreatment procedure of sediment samples is similar to the methods that had been reported (Hu et al., 2011). Sediment samples (2.50 g, dry weight) were spiked with 5 μ L of 1 μ g/ mL d10-fluoranthene and mixed with 15 g anhydrous sodium sulphate. The samples were then extracted with 105 mL HEX/ DCM (1:1 ν/ν) in a Soxhlet extractor at 60 °C for 24 h. Thereafter, copper was added to the extract to remove sulphur. Then all the extract liquor was concentrated to 1 mL, and silica gel/neutral Al₂O₃ was applied to clean up columns. The columns were eluted with 5 mL HEX, 20 mL HEX/DCM (2:1, *V*/*V*), 30 mL HEX/DCM (1:2, *V*/*V*) and 30 mL HEX/DCM (1:3, *V*/*V*). The combined eluents were concentrated to 0.5 mL and added 5 μ L of 1 μ g/mL HCB-C13 before analysis. During the experimentation, three parallel samples were analysed from each sampling site.

Analysis

All samples were analysed using GC-MS (Agilent 7890A-5975C) in the selective ion monitoring mode and using electron-impact (EI) ionisation source. The mass spectrometer quadrupole, source temperature and transfer line temperature were 150, 230 and 280 °C, respectively. The target compounds were separated by HP-5MS capillary column (30 m × 0.25 mm i.d. × 0.25 μ m) and injected splitless (1 μ L) with the temperature of injector port was 250 °C. The temperature programme was as follows: the start temperature was 90 °C, holding 90 °C for 2 min, reached to 170 °C with the speed of 10 °C/min, then the temperature increased to 180 °C with the speed of 1 °C/min, holding 180 °C for 2 min and temperature increased to 270 °C with the speed of 30 °C/min, holding 270 °C for 5 min.

Determination of TOC

The TOC contents were determined by the potassium bichromate volumetry-external heating method (Bao 1999). During the experimentation, three parallel samples were analysed in each site, and two procedural blanks were processed with each batch of samples at the same time.

Calculation of the hazard quotients (HQ)

We used hazard quotients (HQ) to make a preliminary environmental risk assessment. According to the European Guidelines (European Commission 2003), we can calculate the HQs.

$$HQ = MEC/PNEC$$
(1)

PNEC = (L(E)C50 or NOEC) / Assessment factor (2)

MEC is the measured environmental concentration, PNEC is the predicted no effect concentration. L(E)C50 is the median lethal dose or the half maximal effective concentration, NOEC is the no observed effect concentration. If HQ > 1, pollutants may be harmful to the environment, then additional tests need to be performed to quantify the real risk (Hernando et al. 2006).

Quality control and assurance

In order to avoid the contamination during experimentation, skin care products were not used and nitrile gloves were worn to avoid direct contact with samples and all the instruments. All the glassware were soaked in $K_2Cr_2O_7$ - H_2SO_4 solution then baked at 300 °C for 12 h, all glassware were rinsed with HEX before use. The blank samples were processed in the experimentation, all the SMs contaminations were below detection limits in every blank sample. HCB-C13 was used as internal standard to quantify the concentrations of SMs and d10-fluoranthene was used as surrogate standard.

The recoveries of d10-fluoranthene in surface water samples were 81.6-111% and were 93.9-106% in sediment samples. All the experimental results were corrected with the recoveries of surrogate standards. The different concentrations of SMs standards were analysed, all of the working curves had good linear range, and the correlation coefficients were larger than 0.999 ($R^2 > 0.999$). The limits of detection (LOD), as well as the LOQ of SMs were determined by a signal-tonoise ratio of 3 and 10. In surface water and sediment samples, the LODs were in the range of 0.04-0.34 ng/L, 0.03-0.27 ng/ g, respectively; and the LOQs were in the range of 0.13-1.13 ng/L, 0.10–0.92 ng/g, respectively. The recoveries of SMs in surface water and sediment samples were 80.9-97.5% and 92.4-109%, respectively. At the spiking level of 5 ng, repeatability, the relative standard deviation (% RSD) were 3.40-8.20% in surface water samples and 5.30-9.21% in sediment samples. And the linear equations, correlation coefficients, LOD and LOQ of SMs in samples are in the Table 1.

Statistical analyses

The concentrations of SMs in surface water and sediment samples, as well as the TOC in sediment samples were examined using the non-parametric Kolmogorov-Smirnov test (K-S) (Lou et al. 2016). Pearson correlation analysis was used to assess correlations among the different kinds of SMs and TOC in sediment samples (Lou et al. 2016). The p value below 0.05 was considered to be significant. All statistical analyses were performed using the software of SPSS 17.0 and Origin 7.5.

Results and discussion

Levels of SMs in the Jiaozhou Bay wetland

Concentrations, composition profiles and correlation analysis of SMs in samples

HHCB was detected in all samples, accounting for 53–100% of the total SMs in surface water samples (Table 2) and 48–71% in the sediment samples (Table 3). AHTN and MK were detected in 57% and 71% of the water samples, respectively, while no other SM residues were detected in any water sample. All of the sediment samples also contained AHTN and MK but no other reportable SM residues. The respective concentrations of HHCB, AHTN and MK were in the ranges of 10.7–208 (mean 42.2), not detected (ND)–59.1 (mean 10.4) and ND–13.6 (mean 3.60) ng/L in the surface water samples and 13.1–27.4 (mean 19.2), 3.06–14.5 (mean 5.03) and 1.33–18.8 (mean 9.05) ng/g dw in the sediment samples.

HHCB and AHTN may have common sources and similar environmental fates, because the concentrations of HHCB and AHTN were significantly correlated (p < 0.05) in both the surface water and sediment samples. However, the AHTN-to-HHCB ratios were not significantly correlated (p > 0.05), implying that the production and use of AHTN are lower or that AHTN is degraded more easily compared with HHCB in the wetland. There was also a significant correlation between MK and HHCB concentrations in the sediment samples (p < 0.05). The difference between MK and PCMs may be caused by different modes of transport and enrichment in water–sediment environments. Further investigation is needed.

Global comparison of SM levels

The concentrations of HHCB and AHTN detected in the surface water of Jiaozhou Bay (Table 4) are similar to those in Suzhou Creek (Zhang et al. 2008) and the Haihe River in China (Hu et al. 2011), the Hudson River in the USA (Reiner and Kannan 2011) and Ruhr River and Hessen (four small freshwater river systems in Hessen) in Germany (Bester 2005; Quednow and Puttmann 2008). However, the levels are slightly higher than those in Meiliang Bay, China and the

Compounds	Linear equations	Correlation coefficients (R^2)	Surface	Surface water samples(ng/L)			Sediment samples(ng/g)		
			LOD	LOQ	Linear range	LOD	LOQ	Linear range	
ННСВ	$y = 0.3338 \times -0.0120$	0.999	0.13	0.43	0.5–300	0.10	0.33	0.5–100	
AHTN	$y = 0.1071 \times -0.0097$	0.999	0.23	0.77	1.0-300	0.18	0.62	1.0-100	
MK	$y = 0.1721 \times +0.0451$	0.999	0.09	0.30	0.5–300	0.07	0.24	0.5-100	

Table 1 Method's linear equations, correlation coefficients, LOD, and LOQ

Michigan River, Canada (Ma et al. 2014; Peck and Hornbuckle 2004), but slightly lower than those detected in Korea, the Molgora River in Italy (Lee et al. 2010; Villa et al. 2012) and the Someş River in Romania (Moldovan 2006). The level of MK pollution was higher in this area than in other regions, except for Korea (Table 4).

The contamination of sediments by HHCB and AHTN was similar to that reported for the Haihe River, Liangtan River and Suzhou Creek in China (Hu et al. 2011; Sang et al. 2012; Zhang et al. 2008) and the Lippe River in Germany (Kronimus et al. 2004). However, the concentrations were higher than those in Taihu Lake, Meiliang Bay and the intertidal zone of Jiaozhou Bay in China (Che et al. 2010; Ma et al. 2014; Wang et al. 2015) and in Lake Erie and Lake Ontario in the USA (Peck et al. 2006). However, they were lower than the concentrations in the Hudson River in the USA and Molgora River in Italy (Reiner and Kannan 2011; Villa et al. 2012). The level of MK pollution in the sediments was similar to those in the Liangtan River (Sang et al. 2012) and intertidal zone of Jiaozhou Bay and Haihe River in China (Che

 Table 2
 The concentrations of SMs in surface water samples (ng/L)

Site	ADBI	AHMI	ATII	ННСВ	AHTN	MX	MK
1	ND	ND	ND	80.3	25.1	ND	ND
2	ND	ND	ND	208	35.1	ND	4.92
3	ND	ND	ND	115	59.1	ND	3.21
4	ND	ND	ND	25.5	< LOD	ND	6.13
5	ND	ND	ND	23.4	6.85	ND	13.6
6	ND	ND	ND	14.9	1.99	ND	ND
7	ND	ND	ND	12.8	ND	ND	4.52
8	ND	ND	ND	13.1	< LOD	ND	ND
9	ND	ND	ND	18.0	6.42	ND	ND
10	ND	ND	ND	16.9	ND	ND	6.53
11	ND	ND	ND	18.7	6.51	ND	4.40
12	ND	ND	ND	19.7	4.93	ND	2.75
13	ND	ND	ND	13.7	ND	ND	1.30
14	ND	ND	ND	10.7	ND	ND	3.02

ND not detected (the concentration of the pollutant is lower than the detection limit of the instrument), *LOD* limit of detection, *ADBI* celestolide, *AHMI* phantolide, *ATII* traseolide, *HHCB* galaxolide, *AHTN* tonalide, *MX* musk xylene, *MK* musk ketone

et al. 2010; Hu et al. 2011), but higher than those in other areas evaluated. The difference in concentrations was largely due to the different consumption patterns of SMs in different areas: MK is used in cosmetics and soap, HHCB to treat myocardial infarction, and AHTN in topgrade cosmetics, detergents and fabric softeners (Zhou 2016). MK was detected in almost all samples, which suggested that MK is used widely in many household products in this area (Sang et al. 2012; Ma et al. 2014).

Preliminary environmental risk assessment and source identification

Studies of the toxicity of musk to organisms (Vera et al. 2017) indicated that PNECAHTN = PNECHHCB = 71 ng/ g dw, and PNECMK = 320 ng/g dw. Considering the

 Table 3
 The concentrations of SMs and TOC in the sediment samples (ng/g, dw)

Site	ADBI	AHMI	ATII	ННСВ	AHTN	MX	MK	TOC (%)
1	ND	< LOD	ND	20.3	4.68	<lod< td=""><td>15.2</td><td>1.06</td></lod<>	15.2	1.06
2	ND	< LOD	<lod< td=""><td>27.4</td><td>14.5</td><td>ND</td><td>11.7</td><td>0.924</td></lod<>	27.4	14.5	ND	11.7	0.924
3	ND	< LOD	ND	24.8	4.62	<loq< td=""><td>15.5</td><td>1.21</td></loq<>	15.5	1.21
4	ND	<loq< td=""><td><lod< td=""><td>24.1</td><td>8.03</td><td><lod< td=""><td>10</td><td>1.22</td></lod<></td></lod<></td></loq<>	<lod< td=""><td>24.1</td><td>8.03</td><td><lod< td=""><td>10</td><td>1.22</td></lod<></td></lod<>	24.1	8.03	<lod< td=""><td>10</td><td>1.22</td></lod<>	10	1.22
5	ND	ND	ND	20.6	3.77	ND	18.8	1.38
6	ND	< LOD	ND	20.4	3.06	ND	11.7	0.77
7	ND	<lod< td=""><td>ND</td><td>16.6</td><td>4.44</td><td>ND</td><td>12.4</td><td>1.21</td></lod<>	ND	16.6	4.44	ND	12.4	1.21
8	ND	< LOD	ND	16.7	3.42	ND	11.5	0.88
9	ND	< LOD	ND	21.2	4.03	ND	17.3	0.792
10	ND	< LOD	ND	19.1	3.85	ND	13.1	0.539
11	ND	<LOQ	ND	27.1	5.19	ND	7.19	1.36
12	ND	<lod< td=""><td>ND</td><td>17.7</td><td>4.29</td><td>ND</td><td>5.17</td><td>1.28</td></lod<>	ND	17.7	4.29	ND	5.17	1.28
13	ND	<lod< td=""><td>ND</td><td>14</td><td>4.36</td><td>ND</td><td>1.33</td><td>0.385</td></lod<>	ND	14	4.36	ND	1.33	0.385
14	ND	< LOD	<lod< td=""><td>19</td><td>4.8</td><td>ND</td><td>3.74</td><td>0.33</td></lod<>	19	4.8	ND	3.74	0.33
15	ND	< LOD	ND	15	4.45	<lod< td=""><td>2.28</td><td>0.264</td></lod<>	2.28	0.264
16	ND	< LOD	<lod< td=""><td>13.1</td><td>3.78</td><td><lod< td=""><td>1.96</td><td>0.429</td></lod<></td></lod<>	13.1	3.78	<lod< td=""><td>1.96</td><td>0.429</td></lod<>	1.96	0.429
17	ND	< LOD	ND	13.1	4.16	ND	1.93	1.14
18	ND	< LOD	ND	14.7	5.06	ND	2.02	0.924

LOD limit of detection, *LOQ* limit of quantification, *ND* not detected, *ADBI* celestolide, *AHMI* phantolide, *ATII* traseolide, *HHCB* galaxolide, *AHTN* tonalide, *MX* musk xylene, MK musk ketone

Aleas	ннсв	AHTN	MX	MK	Source
Ruhr River, Germany Haihe River, China Korea Meiliang Bay, China Somes River, Romania Michigan River, Canada Hessen,(four small freshwater river systems in Hessen, the concentration is mean concentration) Germany	< 3.00-600 3.50-32.0 $100-2.72 \times 10^{3}$ 0.340-1.39 300 4.70 5.00-678	<1.00-120 2.30-26.7 30.0-520 0.0400-0.230 100 1.00 3.00-299	ND ND 0.0800–0.540 ND 0.0500 ND	ND ND-34.6 < LOQ-420 0.340-1.38 ND 0.0800 ND	Bester (2005) Hu et al. (2011) Lee et al. (2010) Ma et al. (2014) Moldovan (2006) Peck and Hornbuckle (2004) Quednow and Puettmann (2008)
Hudson River, USA Molgora River, Italy Suzhou Creek, China This study Taibu Laka China	3.95–25.8 463 20.0–93.0 10.7–208 0.160_2_10	5.09–22.8 97.0 8.00–20.0 ND-35.1	ND ND ND ND	ND ND ND-13.6	Reiner and Kannan (2011) Villa et al. (2012) Zhang et al. (2008)
Iaihu Lake, China Haihe River, China Lippe River, Germany Meiliang Bay, China Lake Erie and Ontario, USA Hudson River, USA Liangtan River, China Molgora River, Italy Intertidal zone of Jiaozhou Bay Suzhou Creek, China This study	$\begin{array}{c} 0.160 - 3.10 \\ 1.50 - 32.3 \\ 0.500 - 20.0 \\ 0.270 - 0.790 \\ 3.20 - 16.0 \\ 72.8 - 388 \\ < 10.0 - 269 \\ 1.35 \times 10^3 \\ 1.84 - 4.35 \\ 7.00 - 78.0 \\ 13.1 - 27.4 \end{array}$	0.180-1.21 2.00-21.9 23.0-90.0 < LOD-0.0600 0.960 133 - 544 < 5.00-99.7 927 ND-10.9 2.00-21.0 3.06-14.5	0.0800-0.350 ND-47.5 ND < LOD-0.0400 ND ND ND ND ND ND ND ND	0.0300-0.0800 ND-22.6 ND < LOD ND < 5.00-22.0 ND 6.90-10.9 ND 1 33-18.8	Che et al. (2010) Hu et al. (2011) Kronimus et al. (2004) Ma et al. (2014) Peck et al. (2006) Reiner and Kannan (2011) Sang et al. (2012) Villa et al. (2012) Wang et al. (2015) Zhang et al. (2008)
	Ruhr River, Germany Haihe River, China Korea Meiliang Bay, China Somes River, Romania Michigan River, Canada Hessen,(four small freshwater river systems in Hessen, the concentration is mean concentration) Germany Hudson River, USA Molgora River, Italy Suzhou Creek, China This study Taihu Lake, China Lippe River, Germany Meiliang Bay, China Lake Erie and Ontario, USA Hudson River, USA Liangtan River, China Molgora River, Italy Intertidal zone of Jiaozhou Bay Suzhou Creek, China This study	RuhrRiver, Germany Haihe River, China $< 3.00-600$ $3.50-32.0$ $100-2.72 \times 10^3$ Korea $100-2.72 \times 10^3$ Meiliang Bay, China $0.340-1.39$ Somes River, Romania 300 Michigan River, Canada 4.70 Hessen,(four small freshwater river systems in Hessen, the concentration) Germany $5.00-678$ Hudson River, USA $3.95-25.8$ Molgora River, Italy 463 Suzhou Creek, China $20.0-93.0$ This study $10.7-208$ Taihu Lake, China $1.50-32.3$ Lippe River, Germany $0.500-20.0$ Meiliang Bay, China $0.270-0.790$ Lake Erie and Ontario, USA $3.20-16.0$ Hudson River, USA $72.8-388$ Liangtan River, China 1.35×10^3 Intertidal zone of Jiaozhou Bay $1.84-4.35$ Suzhou Creek, China $7.00-78.0$ This study $13.1-27.4$	RuhrRinnerRuhrRinnerRuhrRiver, Germany Haihe River, China $3.50-32.0$ $3.50-32.0$ $2.30-26.7$ $2.30-26.7$ Korea $100-2.72 \times 10^3$ $30.0-520$ $30.0-520$ Meiliang Bay, China $0.340-1.39$ $0.0400-0.230$ $0.0400-0.230$ Somes River, Romania 300 Michigan River, Canada 4.70 1.00 1.00 Hessen, (four small freshwater river systems in Hessen, the concentration) Germany $5.00-678$ $3.00-299$ $3.00-299$ Hudson River, USA Molgora River, Italy $3.95-25.8$ 463 97.0 $5.09-22.8$ $8.00-20.0$ This study Taihu Lake, China $0.160-3.10$ $0.180-1.21$ $0.180-1.21$ Haihe River, ChinaHaihe River, China $1.50-32.3$ $2.00-21.0$ $2.00-21.9$ Lippe River, Germany Meiliang Bay, China $0.270-0.790$ $2.20-0.790$ $CDD-0.0600$ $2.30-90.0$ Lake Erie and Ontario, USA Hudson River, USA $3.20-16.0$ 1.35×10^3 927 927 Intertidal zone of Jiaozhou Bay $1.84-4.35$ $1.84-4.35$ ND-10.9Suzhou Creek, China Tines study $7.00-78.0$ $2.00-21.0$ $2.00-21.0$	RuhrRuhrRuhrRuhrRuhrRuhrRince $3.00-600$ $<1.00-120$ NDHaihe River, China $3.50-32.0$ $2.30-26.7$ NDKorea $100-2.72 \times 10^3$ $30.0-520$ NDMeiliang Bay, China $0.340-1.39$ $0.0400-0.230$ $0.0800-0.540$ Somes River, Romania 300 100 NDMichigan River, Canada 4.70 1.00 0.0500 Hessen,(four small freshwater $5.00-678$ $3.00-299$ NDriver systems in Hessen, the concentration) Germany $concentration Germany$ NDHudson River, USA $3.95-25.8$ $5.09-22.8$ NDMolgora River, Italy 463 97.0 NDSuzhou Creek, China $20.0-93.0$ $8.00-20.0$ NDTaihu Lake, China $0.160-3.10$ $0.180-1.21$ $0.0800-0.350$ Haihe River, China $1.50-32.3$ $2.00-21.9$ ND-47.5Lippe River, Germany $0.500-20.0$ $23.0-90.0$ NDMeiliang Bay, China $0.270-0.790$ $LOD-0.0600$ $Lake Erie and Ontario, USA3.20-16.00.960NDHudson River, USA72.8-388133-544NDLiangtan River, China<1.00-269<5.00-99.7NDMolgora River, Italy1.35 \times 10^3927NDIntertidal zone of Jiaozhou Bay1.84-4.35ND-10.9NDSuzhou Creek, China7.00-78.02.00-21.0NDThis study13.1-27.43.06-14.5RuleRuleRuleRuleRuleRuleRulrRuleRuleRuleRuleRuleRuleRulrRiver, Germany3.00-6002.30-26.7NDNDHaihe River, China0.340-1.390.0400-0.2300.0800-0.5400.340-1.38Somes River, Romania300100NDNDMichigan River, Canada4.701.000.05000.0800Hessen, (four small freshwater5.00-6783.00-299NDNDriver systems in Hessen, theconcentration is meanconcentration Germany5.00-6785.09-22.8NDNDHudson River, USA3.95-25.85.09-22.8NDNDNDMolgora River, Italy46397.0NDNDSuzhou Creek, China20.0-93.08.00-20.0NDNDThis study10.7-208ND-35.1NDND-13.6Taihu Lake, China0.160-3.100.180-1.210.0800-0.3500.0300-0.0800Haihe River, China1.50-32.32.00-21.9ND-47.5ND-22.6Lippe River, Germany0.500-20.023.0-90.0NDNDMeiliang Bay, China0.270-0.790Lake Erie and Ontario, USA72.8-388133-544NDNDHudson River, USA72.8-388133-544NDNDLiangtan River, China<1.00-269<5.00-99.7ND<5.00-22.0Molgora$	RuleRuleRuleRuleRuleRuleRulrRuleRuleRuleRuleRuleRuleRulrRiver, Germany $3.00-600$ $2.30-26.7$ NDNDHaihe River, China $0.340-1.39$ $0.0400-0.230$ $0.0800-0.540$ $0.340-1.38$ Somes River, Romania 300 100 NDNDMichigan River, Canada 4.70 1.00 0.0500 0.0800 Hessen, (four small freshwater $5.00-678$ $3.00-299$ NDNDriver systems in Hessen, the concentration is mean concentration Germany $5.00-678$ $5.09-22.8$ NDNDHudson River, USA $3.95-25.8$ $5.09-22.8$ NDNDNDMolgora River, Italy 463 97.0 NDNDSuzhou Creek, China $20.0-93.0$ $8.00-20.0$ NDNDThis study $10.7-208$ ND-35.1NDND-13.6Taihu Lake, China $0.160-3.10$ $0.180-1.21$ $0.0800-0.350$ $0.0300-0.0800$ Haihe River, China $1.50-32.3$ $2.00-21.9$ ND-47.5ND-22.6Lippe River, Germany $0.500-20.0$ $23.0-90.0$ NDNDMeiliang Bay, China $0.270-0.790$ $Lake Erie and Ontario, USA72.8-388133-544NDNDHudson River, USA72.8-388133-544NDNDLiangtan River, China<1.00-269<5.00-99.7ND<5.00-22.0Molgora$

Table 4 Comparison of SM pollution levels in different regions

HHCB galaxolide, AHTN tonalide, MX musk xylene, MK musk ketone, ND not detected, LOD limit of detection

worst-case scenario, we took the maximum concentration that we detected as the maximum estimated concentration (MEC); thus, the results represent the worst possible contamination in the area. We then derived hazard quotients (HQs) of 0.204, 0.386 and 0.059 for AHTN, HHCB and MK, respectively.

According to the method used by the European Commission (2003), it does not seem likely that our target chemicals will cause much environmental damage theoretically, because their HQs are all less than 1. Nevertheless, more relevant toxicology data for estuarine organisms are needed to confirm this conclusion.

For hydrophobic organic compounds in sediments, TOC is also required for ecological risk assessment (Hu et al. 2011). In this study, the TOC in the sediment samples was in the range of 0.264 - 1.38% (mean 0.894%) (Table 3). There were no significant correlations between SMs and TOC (p > 0.05); however, this relationship requires further research.

Distributions patterns of SMs in the Jiaozhou Bay wetland

The highest concentration of total SMs was detected at site 2 (Fig. 2: 248 ng/L in surface water and Fig. 3: 53.6 ng/g in sediment samples). The lowest concentration of total SMs was

detected at site 8 for the surface water samples and at site 16 for the sediment samples (13.1 ng/L and 18.8 ng/g, respectively). In general, the concentration of SMs in the Jiaozhou Bay wetland decreased in the direction of flow, except at the sites near landfill sites or older villages. This phenomenon occurred in both surface water and sediment samples, which indicated the presence of point source pollution of SMs in the Jiaozhou Bay wetland.

In the Taoyuan River, the concentrations of SMs tended to be higher at sites 1 and 2 because of the continuous input of landfill leachate. Perhaps the accumulation effect was greater than the degradation effect in this area. At sites 3 and 4, the pollutant concentrations dropped rapidly; the flow may be greater there, and the effects of erosion on pollutants were obvious. Because the concentration of pollutants in sediments did not change much, there was no accumulation in sediments at sites 3 and 4 (Fig. 3). Due to the purification effect and current scouring, the concentrations of SMs decreased gradually with the flow, except at sites 4 and 5, which are near villages.

Conclusions

SMs were detected in surface water and sediment samples from the Jiaozhou Bay wetland, China. HHCB, AHTN and

Fig. 2 The distribution of SMs in

the surface water at the different

sites

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MK were the main pollutants detected in both the surface water and sediment samples. The sources of HHCB and AHTN were similar. The distribution of SMs conformed to the characteristics of the population density and sewage drainage. Based on the HQ and TOC content, there may be no significant harm to the environment. More relevant toxicology data for estuarine organisms are still needed for accurate assessment of the environmental risk. The relationships between TOC and SMs require further research. The concentrations of SMs discharged from the landfill were lower in the Dagu River estuary. Therefore, the wetland is a natural barrier in Jiaozhou Bay, as it prevented contaminants from entering the sea.



Fig. 3 The distribution of SMs in sediments at the different sites

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