RESEARCH ARTICLE



Bioavailability of pollutants sets risk of exposure to biota and human population in reservoirs from Iguaçu River (Southern Brazil)

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Abstract The Iguaçu River, located at the Southern part of Brazil, has a great socioeconomic and environmental importance due to its high endemic fish fauna and its potential to generate hydroelectric power. However, Iguaçu River suffers intense discharge of pollutants in the origin of the river. In a previous report, the local environmental agency described water quality to improve along the river course. However, no study with integrated evaluation of chemical analysis and biological responses has been reported so far for the Iguaçu River. In the current study, three different Brazilian fish species (*Astyanax bifasciatus, Chrenicicla iguassuensis*, and *Geophagus brasiliensis*) were captured in the five cascading reservoirs of Iguaçu River for a multi-biomarker study.

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Chemical analysis in water, sediment, and muscle indicated high levels of bioavailable metals in all reservoirs. Polycyclic aromatic hydrocarbons (PAHs) were detected in the bile of the three fish species. Integration of the data through a FA/PCA analysis demonstrated the poorest environmental quality of the reservoir farthest from river's source, which is the opposite of what has been reported by the environmental agency. The presence of hazardous chemicals in the five reservoirs of Iguaçu River, their bioaccumulation in the muscle of fish, and the biological responses showed the impacts of human activities to this area and did not confirm a gradient of pollution between the five reservoirs, from the source toward Iguaçu River's mouth. Therefore, diffuse source of pollutants present along the river course are increasing the risk of exposure to biota and human populations.

Keywords Iguaçu River · Fish · Multi-biomarkers · Metals · PAHs · Risk of exposure

Introduction

Aquatic ecosystems receive a diverse set of compounds from many anthropogenic sources. Chemical interactions in complex mixtures represent a challenge for assessing the health risks of organisms (Contreras López 2003; Tian et al. 2015), but represent the real situation of exposure.

Human activities have deteriorated water quality in the last century through discharge of poorly treated urban and industrial waste. A critical consequence is the scarcity of potable water even in regions where freshwater is abundant. With the continuous growth of human population, the demand for potable water will increase. The Iguaçu River is an important river of Southern Brazil (Fig. 1), due to its potential for hydroelectric power generation, human water supply, and the occurrence of many endemic fish species. Nevertheless, Iguaçu River is described as the second most polluted river in Brazil (IBGE, I. B. de G. e E 2010). It rises in the Metropolitan Region of Curitiba (MRC, 1.8 million inhabitants - IBGE 2010) and receives large amounts of urban and industrial wastes without adequate treatment, resulting in frequent cyanobacteria blooms in reservoirs closest to the city (Carneiro et al., 2014) and high concentrations of polycyclic aromatic hydrocarbons (PAHs) in the sediment (Leite et al. 2011).

Approximately 200 km from MRC, there is a sequence of five major cascading reservoirs used for hydropower plants and intense agricultural activity (e.g., soybeans and wheat farms) developed along the riverbank. After crossing Parana State, approximately 1000 km from the MRC, the Iguaçu river reaches the Iguaçu Falls, which works as a natural barrier and is responsible for generating a high endemic fish fauna in the Iguaçu River (about 70 % of fish species) (Baumgartner et al. 2012). The lower part of the Iguaçu river will then flow into the Parana River. This significant ichthyologic endemism represents an important ecologic issue for the Iguaçu River, since susceptibility of extinction increases with the construction of dams, introduction of non-native species, aquaculture, and pollution (Daga and Gubiani, 2012).

The State Environmental Agency described in its most recent online report that an improvement of the water quality index of the studied sites upgraded from moderately to less degraded (IAP 2008). However, these regulatory monitoring programs usually consider only analysis of physicochemical parameters, which are inadequate to predict potential toxic effects on the ecosystem (Dos Santos et al. 2016). Correlations among environmental concentrations of contaminants, bioaccumulation, and early biological responses should be integrated to better comprehend the environmental risks (Van der Oost et al. 2003).

In this context, a multi-biomarker approach is a more suitable analysis to detect earlier biological responses triggered by pollutants (de Andrade Brito et al. 2012; Linde-Arias et al. 2008; Osório et al. 2014). This approach will provide a better ecotoxicological scenario necessary for environmental management (Mills and Chichester, 2005).

Biochemical biomarkers have considerable potential for measuring effects of chemicals under field conditions since they are the earliest sensitive response (Carvalho et al. 2012). Genetic biomarkers inform levels of DNA damage which can be parameter used to assess the susceptibility of an organism to mutations and cancer (Araldi et al. 2015; Collins et al. 2014). Histopathological analysis is another important tool for evaluating lesions in target organs that can detect both acute and chronic exposure (Mela et al. 2007; Oliveira Ribeiro et al. 2006; Rabitto et al. 2005). Evaluation of these biomarkers along with the chemical profile of the water provides a reliable risk assessment for the exposed biota.

In the present study, the water quality of five reservoirs along the Iguaçu River was investigated for the first time through a multi-biomarkers approach in three Brazilian freshwater fish species. Three different fish species were used to compare their specific responses. The fish used are from different trophic levels: the *Astyanax bifasciatus* (Characidae) or



Fig. 1 Map of Iguaçu River and the major cascading reservoirs, according to the proximity from the metropolitan region of Curitiba city (MRC), the main source of pollutants. *FA* Foz do Areia, *SE* Segredo, *SS* Salto Santiago, *SO* Salto Osório, *SC* Salto Caxias

lambari an endemic herbivore and the most abundant fish species, the *Chrenicicla iguassuensis* (Cichlidae) or *joaninha* an endemic and carnivorous group, and the *Geophagus brasiliensis* (Cichlidae) or *cará* an omnivorous and widely distributed fish (Baumgartner et al. 2012).

Materials and methods

The Iguaçu River is located in Parana, a southern state in Brazil. It extends 1.060 km and is the most important river in a basin of approximately 72.000 km² (Baumgartner et al. 2012). Along the river, there are five reservoirs that were analyzed in this study (Fig. 1). The Foz do Areia Reservoir (FA, 26° 00' 60.82″ S 51° 65' 68.22″ W) has the largest hydropower plant and is the closest to the metropolitan region of Curitiba (MRC). The other reservoirs, toward the river mouth, are Segredo (SE, 25° 79' 13.36″ S, 52° 11' 89.36″ W), Salto Santiago (SS, 25° 63' 73.08″ S, 52° 60' 20.19″ W), Salto Osório (SO, 25° 53' 71.06″ S, 53° 00' 33.31″ W), and Salto Caxias (SC, 25° 53' 64.43″ S, 53° 49' 90.70″ W).

Fish sampling

A total of 346 adult fish (*A. bifasciatus*, *C. iguassuensis*, and *G. brasiliensis*) were captured in the reservoirs during summer (January–February) and winter (July–August) of 2013

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(Table 1). Fish were captured through fishing nets (4–6 cm mesh) and trawl net with sampling every 4 h during a period of 24 to 48 h. Fish were then anesthetized with 25–30 mg I^{-1} benzocaine and killed in the field to avoid stress due to fish transportation. The same procedure and capture effort was exerted in all studied reservoirs.

Physicochemical parameters, microcystin-LR, and metals detection

Temperature, pH, and oxygen dissolved data were provided by the State of Parana Electric Company - COPEL and TRACTEBEL Energy companies (Table 2).

Microcystin-LR was extracted according to MATTHIENSEN et al. (1999) with modifications. The intracellular toxin was determined in 1 L of water collected per reservoir and filtered in a glass fiber filter (GF/C 45 mm, Millipore), which was frozen and lyophilized for 24 h. Twenty-five milliliters of pure methanol (HPLC grade, J.T. Baker) was added to extract the toxin at room temperature for 1 h. In order to optimize the extraction, this step was repeated. After the second extraction, the toxin was concentrated with a vacuum rotary evaporator at 80 rpm at 40 °C. The extract was solubilized in 1 ml of methanol and stored at -20 °C. On the other hand, the toxins dissolved in the column of water of the reservoirs were determined in 1 L of filtered water (GF/C 45 mm, Millipore) with addition of 10 %

| | Fish species | Number | | Weight (g) | |
|----|-----------------|--------|--------|-------------------------------|------------------|
| | | Summer | Winter | Summer | Winter |
| FA | A. bifasciatus | 20 | 20 | 14.31 (±1.85) a | 22.78 (±3.53) ab |
| | G. brasiliensis | 20 | 4 | 13.7 (±4.71) a | 24.19 (±2.52) |
| | C. iguassuensis | 6 | 1 | 15.3 (±18.85) a | 88.8 |
| SE | A. bifasciatus | 20 | 20 | 31.4 (±7.07) b | 26.48 (±2.93) ac |
| | G. brasiliensis | 20 | 4 | 29.82 (±4.75) b | 23.71 (±2.98) |
| | C. iguassuensis | 3 | 0 | 51.16 (±87.52) | |
| SS | A. bifasciatus | 20 | 20 | 14.45 (±5.58) a | 13.67 (±2.12) b |
| | G. brasiliensis | 20 | 4 | 12.39 (±12.39) ^a a | 39.58 (±10.11) |
| | C. iguassuensis | 2 | 4 | 79,33 (±37.14) | 26.12 (±3.94) |
| SO | A. bifasciatus | 9 | 20 | 5.32 (±1.36) ^a a | 25.04 (±4.48) c |
| | G. brasiliensis | 9 | 0 | 17.98 ± 18.08 | |
| | C. iguassuensis | 17 | 0 | 17.77 (±8.94) a | |
| SC | A. bifasciatus | 20 | 20 | 34.38 (±5.73) b | 33.81 (±5.92) c |
| | G. brasiliensis | 3 | 0 | 260 (±32.4) b | |
| | C. iguassuensis | 20 | 20 | 96.68 (±31.04) b | 96.68 (±31.04) |

Table 1Fish number and weightaverage of each species capturedin the studied reservoirs of IguaçuRiver during the two samplingseasons

Lowercase letters indicate significant differences between sites in the same season

FA Foz do Areia Reservoir, SE Segredo Reservoir, SS Salto Santiago Reservoir, SO Salto Osório Reservoir, SC Salto Caxias Reservoir

^a Indicates seasonal differences for the same site

Table 2Physicochemicalparameters of temperature, pHand dissolved oxygen andmicrocystin-LR intracellular inthe water of five reservoirs ofIguaçu River

| Reservoir | Season | Temperature (°C) | рН | Dissolved oxygen (mg L^{-1}) | Mc-LR Intrac. (μg L ⁻¹) |
|-----------|--------|------------------|------|---------------------------------|---|
| Conama | | | 6–9 | >5 | |
| FA | Summer | 26.25 | 7.71 | 7.36 | 65.5 |
| | Winter | 7.13 | 6.73 | 7.63 | <lq< td=""></lq<> |
| SE | Summer | 23.70 | 5.90 | 7.17 | 0.27 |
| | Winter | 19.23 | 4.93 | 7.47 | <lq< td=""></lq<> |
| SS | Summer | 22.22 | 7.47 | 4.59 | 18.42 |
| | Winter | 17.35 | 7.13 | 9.21 | <lq< td=""></lq<> |
| SO | Summer | 24.57 | 7.30 | 5.61 | - |
| | Winter | 18.06 | 6.86 | 9.57 | - |
| SC | Summer | 27.50 | 6.53 | 7.60 | <lq< td=""></lq<> |
| | Winter | 19.86 | 6.63 | 9.13 | <lq< td=""></lq<> |

Italicized values indicate maximum or minimum values for each parameter analyzed

FA Foz do Areia Reservoir, SE Segredo Reservoir, SS Salto Santiago Reservoir, SO Salto Osório Reservoir, SC Salto Caxias Reservoir, Mc-LR Intrac Microcystin-LR intracellular, <LQ Lower than limit of quantification

trifluoroacetic acid (TFA) and filtered again. A solid phase column (C18) was used to extract the toxin from the filtered water, which was eluted with 3 ml of 0.1 % methanol/TFA (ν/ν) at a flux of 10 ml min⁻¹. The collected extracts were dried with nitrogen gas at 45 °C and suspended in methanol. Finally, the extracts were analyzed with a LC-MS ion trap (HPLC Shimadzu Prominence) through electrospray source.

Metals (Ag, Co, Cd, Cr, Cu, Mn, Ni, Pb and Zn) were measured in water, sediment, and fish muscle from the five reservoirs. The water was sampled at 30 cm below the surface and preserved with 0.5 % of HNO₃, while the sediment was collected 10 m away from the shore. Samples were protected from light and kept at 4 °C until analysis. For analysis of toxic metals bioaccumulation, muscle samples of all fish were pooled per group and stored at -75 °C until analysis according to Cotta et al. (2006). After acid digestion (Method US EPA 3005A for water; Method US EPA 3050B for sediment and tissue samples), a flame atomic absorption spectrometer was used to detect metals (FAAS, Varian, AA 240FS).

Polycyclic aromatic hydrocarbons detection in bile

The concentration of polycyclic aromatic hydrocarbons (PAHs) in bile was determined following a protocol from Hanson, et al. (2009), with minor modifications. Bile of three individuals was pooled and stored in amber glass vials at -75 °C. Bile samples were diluted (1:1000) in 48 % methanol and added onto 96-well black microplate for PAHs through different excitation/emission wavelengths (288/330, 334/376, 364/406, and 380/422) in spectrophotometer. For quantification, a PAH mix (Cod. 47930-U, SUPELCO) was utilized to establish a PAHs standard curve.

Biological sampling

After anesthesia, some biometric parameters such as total length (TL) and total weight (TW) were obtained for a condition factor of Fulton (K) determination. Blood samples were obtained through caudal vein (for *C. iguassuensis* and *G. brasiliensis*) or cardiac puncture (for *A. bifasciatus*) with heparinized syringes and stored in fetal bovine serum for Comet assay and smeared in a slide for micronucleus counting. After euthanasia, the spinal cord was severed and liver and gill samples were dissected for light and scanning electron microscopy respectively. For biochemical analyses, the liver, muscle, and brain samples were stored at -75 °C. The weight of liver was obtained to determine the hepatosomatic index (HSI) and using the formula HSI=liver mass /(100 × total fish mass).

Biochemical biomarkers

Liver, brain, and muscle samples were thawed and homogenized at 1:10 (w/v) ratio with the following buffers: A Tris–HCl/ EDTA buffer (20 mM Tris–HCl, 1 mM ethylenediamine tetraacetic acid, 1 mM phenylmethylsulfonyl fluoride, pH 7.4) was used for the liver samples and a 0.1-M potassium phosphate buffer was used for muscle (pH 7.5) and brain (pH 8.0) samples. Homogenized samples were then centrifuged at 12000g, 4 °C for 30 min; the supernatants were collected and stored at -75 °C. Total protein concentration was determined (Bradford et al., 1976 with minor modifications of volumes for microplates) and utilized to normalize protein concentrations in supernatants. For *A. bifasciatus*, liver samples of two fish were pooled due to the small size and only catalase, glutathione *S*-transferase, and lipid peroxidation assays were performed. Acetylcholinesterase (AChE) activity assay (Ellman et al. 1961) was performed with the muscle and brain supernatants. *Catalase* (Aebi 1984) and *glutathione S-transferase* (GST, Keen et al. 1976) activity, *non-protein thiols* (NPT) / GSH concentration (Sedlak and Lindsay 1968), *lipid peroxidation* (LPO, Jiang et al. 1991, 1992) and *protein carbonylation* (PCO, Levine et al. 1994) assays were performed with liver supernatants. *Glutathione peroxidase* (GPx) activity was analyzed only for *G. brasiliensis* liver supernatants. Previously published protocols (Costa et al. 2010a, 2010b; de Andrade Brito et al. 2012; Osório et al., 2014) were utilized for analyzing all biochemical biomarkers.

Genotoxicity biomarkers

Comet assay (Speit and Hartmann, 1999) and *piscine micronucleus* test (Heddle, 1973) were performed with erythrocytes, according to previously published protocols (Cestari et al., 2004; Ferraro et al., 2004; Ramsdorf et al. 2009).

Histopathological biomarkers

Light microscopy Samples of liver were fixed (70 % ethanol, 4 % formaldehyde, 5 % glacial acetic acid) for 16 h, dehydrated in a graded series of ethanol and xylene, and embedded in Paraplast-Plus (Sigma[®] St Louis, USA). Sections of 5 μ m were obtained using a microtome (Leica), stained with Hematoxylin/Eosin, and mounted with Entellan (Merck[®]). Analysis was performed according to Bernet et al. (1999) with a few modifications.

Scanning electron microscopy The second left gill arch was washed with phosphate buffer, fixed (2.5 % glutaraldehyde and 2 % paraformaldehyde in 0.1 M sodium cacodylate buffer, pH 7.4) for at least 24 h, dehydrated in ethanol series (Merck[®]) and liquid CO₂, and metalized with gold for further analysis in scanning electron microscope JEOL KAL-6360LV.

Statistical analysis

All data were checked for outliers (Grubb's test), normality (Shapiro's test), and homoscedasticity (Bartlett's test) before analysis. Either *two-way* ANOVA test followed by the *Tukey-Kramer* post-test (parametric data) or *Kruskal-Wallis* test followed by the *Dunn's* test (nonparametric data) were utilized, with p < 0.05. Correlation analysis (Spearman test) was performed between the variables and age parameters (weight and total length), in order to evaluate the influence of age on biological responses.

Biological data was integrated by two different methods. First, PERMANOVA was applied on biological responses to assess the differences between the two studied seasons (summer and winter). The second integration method was aimed to highlight associations among the variables measured in this study (both biomarker responses and contaminant levels in water, sediment, and body burden) during each of the two seasons (summer and winter) and for the three studied fish species (A. bifasciatus, G. brasiliensis, and C. iguassuensis). Factor analysis with principal component analysis as the extraction method (FA/PCA) was utilized. Associations between the different biomarkers (Comet assay, micronucleus assay, GST, CAT, GSH, GPx, PCO, AChE, LPO, Histopathological index, K, and HSI), metal loads in water (Ni, Cd, Zn, Cr, and Mn), sediment (Ni, Cd, Zn, Cr, Mn, Ag, Pb, and Cu), muscle body burden (Cd, Zn, Cr, Mn, Pb, Cu, and Co), and PAH metabolites in bile were assessed. Three matrices with original data (one for each fish species) were integrated. Since the matrices presented some missing data, the FA/PCA integration for each of the data matrix was performed in two steps: First, the variables showing missing data were omitted, and the FA/PCA was done with all cases; second, the cases showing missing data were omitted, and the analysis was done with all variables.

The variables were auto-scaled (standardized) and treated with equal importance. The selected variables to be interpreted were those associated with the factors with a loading cutoff of at least 0.50, a value which is more conservative than the loading cutoff recommended by Tabachnic and Fidell (1996). The relevance of the observed associations to each of the six sampling stations (cases) was estimated by calculating the factor score from each case for the centroid of all cases for the original data. All the statistical and multivariate analyses were performed using STATISTICA 12 software (StatSoft Inc. USA).

Results

Condition factor (K) and hepatosomatic index (HSI)

Differences of condition factors between groups were observed only for individuals from the species *A. bifasciatus*. The individuals from SS reservoir presented lower condition factor (1.18) than those of FA (1.31, winter), SE (1.34, summer), and SC (1.33, summer and winter) reservoirs. In general, fish from the five reservoirs had different ages, as estimated by their weight and total length (Table 1). For the three fish species, the weights of the collected fish from SE and SC were higher than those from FA, SS, and SO for the summer samples. Correlation analysis showed a weak negative correlation (r=0.3-0.6) for parameters of age (weight and total length) only with the response of AChE in the muscle of *G. brasiliensis* and *A. bifasciatus*.

The HSIs of *G. brasiliensis* and *A. bifasciatus* from the FA reservoir were significantly higher in the fish captured in the winter (3.33 and 0.97, respectively) than in the summer (1.36

and 0.60). *G. brasiliensis* from the FA reservoir had higher HSI than those from the other reservoirs (SE: 0.58; SS: 1.97).

Physicochemical parameters

Water temperature, pH, dissolved oxygen (DO), and intracellular Microcystin-LR concentrations are summarized in Table 2. The highest registered temperature was 27.5 °C (average of 24.9 °C) in SC reservoir (summer), and the lowest was 7.13 °C (average of 16.3 °C) in FA reservoir (winter). SE reservoir had the lowest pH value (5.90—summer and 4.93 winter). No variation in DO was observed during the seasons for FA and SE reservoirs (around 7 to 8 mg L⁻¹), but remarkable variations were recorded at SS, SO, and SC, with the lowest concentration of DO for the SS reservoir (4.59 mg L⁻¹ summer). Intracellular Microcystin-LR concentrations were higher in FA and SS reservoirs during the summer. In the column of water, Microcystin-LR was only detected in FA (summer) at low concentrations (1.08 ng L⁻¹, data not shown).

Chemical analysis of metals

Water Chromium was above the levels permitted by Brazilian legislation (CONAMA 2012) in the water from FA, SS and SO (summer), and SS (winter) reservoirs. Cd, Zn and Mn were detected in the water of all reservoirs at both seasons, but at concentrations below the limit established by legislation. The highest concentrations of Ni and Cd were detected in SC reservoir, whereas that of Mn was highest in FA at both studied seasons (Table 3). Other metals such as Ag, Cu, Co

and Pb were below the limit of quantification in the water of all reservoirs.

Sediments Concentrations of Pb and Cu were higher in SO than in the other reservoirs (Table 3).

Fish From all studied species and reservoirs had high concentrations of Zn in the muscle. Elevated levels of Cr, Cu, Mn, Cd and Pb were also observed in the samples (Table 4). *A. bifasciatus* and *C. iguassuensis* from SO reservoir (summer) had the highest Pb concentration. Relatively high concentrations of Cr were also detected in fish from almost all reservoirs, whereas Cd was mainly bioaccumulated in the muscle of fish from SC reservoir (summer).

Chemical speciation of metals was not distinguished by the method of detection performed.

PAHs in bile

The 2-ring PAHs were the ones predominantly detected in the bile of the three species from the five reservoirs. The fish from FA reservoir had the highest amounts of the 2-ring PAHs (Table 5). *A. bifasciatus* from SS and *G. brasiliensis* from SO reservoir showed the highest concentrations of larger PAHs.

Biochemical biomarkers

A. bifasciatus showed the more conclusive results when analyzing the livers of individuals from both seasons, but no clear effect of contamination gradient was verified. GST activity

Table 3 Metals concentrations detected in the water and in the sediment of the five studied reservoirs of Iguaçu River

| | | Water (µg | L^{-1}) | | | | Sediment (| mg kg ⁻¹) | | |
|--------|--------|--|-------------------|--------------------|---|--|--|-----------------------|----------------|--------------------|
| | | Ni | Cd | Zn | Cr | Mn | Pb | Cu | Mn | Fe |
| | Conama | 25 | 10 | 5000 | 50 | 500 | 91.3 | 197 | | |
| Summer | FA | <lq< td=""><td>1.60 ± 1.15</td><td>72.56 ± 20.85</td><td>75.70 ± 18.66</td><td>111.46 ± 5.20</td><td><lq< td=""><td>$5.3\pm\!0.2$</td><td>39,.±2.0</td><td>104.0 ± 3.3</td></lq<></td></lq<> | 1.60 ± 1.15 | 72.56 ± 20.85 | 75.70 ± 18.66 | 111.46 ± 5.20 | <lq< td=""><td>$5.3\pm\!0.2$</td><td>39,.±2.0</td><td>104.0 ± 3.3</td></lq<> | $5.3\pm\!0.2$ | 39,.±2.0 | 104.0 ± 3.3 |
| | SE | <lq< td=""><td>2.23 ± 0.96</td><td>11.66 ± 5.20</td><td><lq< td=""><td><lq< td=""><td><lq< td=""><td>8.9 ± 0.3</td><td>330.7 ± 24.4</td><td>1136.0 ± 158.0</td></lq<></td></lq<></td></lq<></td></lq<> | 2.23 ± 0.96 | 11.66 ± 5.20 | <lq< td=""><td><lq< td=""><td><lq< td=""><td>8.9 ± 0.3</td><td>330.7 ± 24.4</td><td>1136.0 ± 158.0</td></lq<></td></lq<></td></lq<> | <lq< td=""><td><lq< td=""><td>8.9 ± 0.3</td><td>330.7 ± 24.4</td><td>1136.0 ± 158.0</td></lq<></td></lq<> | <lq< td=""><td>8.9 ± 0.3</td><td>330.7 ± 24.4</td><td>1136.0 ± 158.0</td></lq<> | 8.9 ± 0.3 | 330.7 ± 24.4 | 1136.0 ± 158.0 |
| | SS | <lq< td=""><td>0.43 ± 0.25</td><td>23.25 ± 3.74</td><td>69.63 ± 15.16</td><td>55.30 ± 4.57</td><td><lq< td=""><td>6.6 ± 0.6</td><td>67.4 ± 6.5</td><td>312.5 ± 11.0</td></lq<></td></lq<> | 0.43 ± 0.25 | 23.25 ± 3.74 | 69.63 ± 15.16 | 55.30 ± 4.57 | <lq< td=""><td>6.6 ± 0.6</td><td>67.4 ± 6.5</td><td>312.5 ± 11.0</td></lq<> | 6.6 ± 0.6 | 67.4 ± 6.5 | 312.5 ± 11.0 |
| | SO | <lq< td=""><td>0.27 ± 0.05</td><td>42.75 ± 8.98</td><td>72.53 ± 26.16</td><td>16.83 ± 1.55</td><td>30.6 ± 1.8</td><td>76.3 ± 1.7</td><td>18.3 ± 0.8</td><td>1670 ± 80</td></lq<> | 0.27 ± 0.05 | 42.75 ± 8.98 | 72.53 ± 26.16 | 16.83 ± 1.55 | 30.6 ± 1.8 | 76.3 ± 1.7 | 18.3 ± 0.8 | 1670 ± 80 |
| | SC | 5.2 ± 0.56 | 3.06 ± 0.61 | 60.50 ± 41.43 | <lq< td=""><td>5.36 ± 1.87</td><td><lq< td=""><td>$4.1\pm\!0.5$</td><td>40.8 ± 4.0</td><td>92.9 ± 9.0</td></lq<></td></lq<> | 5.36 ± 1.87 | <lq< td=""><td>$4.1\pm\!0.5$</td><td>40.8 ± 4.0</td><td>92.9 ± 9.0</td></lq<> | $4.1\pm\!0.5$ | 40.8 ± 4.0 | 92.9 ± 9.0 |
| Winter | FA | <lq< td=""><td>0.56 ± 0.40</td><td>43.30 ± 23.72</td><td>46.06 ± 14.93</td><td>72.23 ± 5.39</td><td><lq< td=""><td>$3.9\pm\!0.2$</td><td>22.2 ± 2.6</td><td>$23.4\!\pm\!2.1$</td></lq<></td></lq<> | 0.56 ± 0.40 | 43.30 ± 23.72 | 46.06 ± 14.93 | 72.23 ± 5.39 | <lq< td=""><td>$3.9\pm\!0.2$</td><td>22.2 ± 2.6</td><td>$23.4\!\pm\!2.1$</td></lq<> | $3.9\pm\!0.2$ | 22.2 ± 2.6 | $23.4\!\pm\!2.1$ |
| | SE | 4.8 ± 3.82 | $2.96\!\pm\!0.37$ | 106.66 ± 19.87 | <lq< td=""><td>14.00 ± 2.00</td><td><lq< td=""><td>3.9 ± 0.2</td><td>22.2 ± 2.6</td><td>1527. 3±95.5</td></lq<></td></lq<> | 14.00 ± 2.00 | <lq< td=""><td>3.9 ± 0.2</td><td>22.2 ± 2.6</td><td>1527. 3±95.5</td></lq<> | 3.9 ± 0.2 | 22.2 ± 2.6 | 1527. 3±95.5 |
| | SS | <lq< td=""><td>1.00 ± 0.72</td><td>37.90 ± 12.58</td><td>62.53 ± 18.71</td><td>21.66 ± 3.30</td><td><lq< td=""><td>$3.2\pm\!0.4$</td><td>195.5 ± 23.2</td><td>138.9 ± 27.2</td></lq<></td></lq<> | 1.00 ± 0.72 | 37.90 ± 12.58 | 62.53 ± 18.71 | 21.66 ± 3.30 | <lq< td=""><td>$3.2\pm\!0.4$</td><td>195.5 ± 23.2</td><td>138.9 ± 27.2</td></lq<> | $3.2\pm\!0.4$ | 195.5 ± 23.2 | 138.9 ± 27.2 |
| | SO | <lq< td=""><td>1.50 ± 0.4</td><td>35.00 ± 12.0</td><td><lq< td=""><td>7.56 ± 1.20</td><td>21.1±2.1</td><td>51.0 ± 1.6</td><td>17.6 ± 0.8</td><td>1214.6 ± 71.6</td></lq<></td></lq<> | 1.50 ± 0.4 | 35.00 ± 12.0 | <lq< td=""><td>7.56 ± 1.20</td><td>21.1±2.1</td><td>51.0 ± 1.6</td><td>17.6 ± 0.8</td><td>1214.6 ± 71.6</td></lq<> | 7.56 ± 1.20 | 21.1±2.1 | 51.0 ± 1.6 | 17.6 ± 0.8 | 1214.6 ± 71.6 |
| | SC | 6.5 ± 2.55 | 4.03 ± 0.80 | 2.55 ± 5.72 | <lq< td=""><td>16.9 ± 2.94</td><td><lq< td=""><td>11.9 ± 0.7</td><td>250.8 ± 9.1</td><td>2586.1±515</td></lq<></td></lq<> | 16.9 ± 2.94 | <lq< td=""><td>11.9 ± 0.7</td><td>250.8 ± 9.1</td><td>2586.1±515</td></lq<> | 11.9 ± 0.7 | 250.8 ± 9.1 | 2586.1±515 |

Italicized values indicate higher concentrations for each metal analyzed. Data expressed as average \pm standard deviation

FA Foz do Areia Reservoir, SE Segredo Reservoir, SS Salto Santiago Reservoir, SO Salto Osório Reservoir, SC Salto Caxias Reservoir, <LQ Lower than limit of quantification

| Mus | cle (mg kg ⁻¹) | Cr | | Cu | | Mn | | Cd | | Zn | | Pb | |
|---------------|----------------------------|----------------|---|----------------|--|---|---|---|---|------------------|--------------------|-----------------|-------------------|
| | | Summer | Winter | Summer | Winter | Summer | Winter | Summer | Winter | Summer | Winter | Summer | Winter |
| FA | A. bifasciatus | 4.10 ± 3.0 | <lq< td=""><td>1.23 ± 0.8</td><td>0.94 ± 0.7</td><td>5.52</td><td>0.48 ± 0.6</td><td>0.27 ± 0.1</td><td>1.39 ± 0.2</td><td>52.93 ± 0.3</td><td>58.92 ± 7.4</td><td>15.99 ± 0.9</td><td>8.75 ± 3.1</td></lq<> | 1.23 ± 0.8 | 0.94 ± 0.7 | 5.52 | 0.48 ± 0.6 | 0.27 ± 0.1 | 1.39 ± 0.2 | 52.93 ± 0.3 | 58.92 ± 7.4 | 15.99 ± 0.9 | 8.75 ± 3.1 |
| | G. brasiliensis | 5.11 ± 1.4 | 2.76 | NA | <lq< td=""><td>12.50</td><td><lq< td=""><td>0.79 ± 0.1</td><td>2.14 ± 3.0</td><td>25.79 ± 3.0</td><td>30.10 ± 11.0</td><td>2.75</td><td><lq< td=""></lq<></td></lq<></td></lq<> | 12.50 | <lq< td=""><td>0.79 ± 0.1</td><td>2.14 ± 3.0</td><td>25.79 ± 3.0</td><td>30.10 ± 11.0</td><td>2.75</td><td><lq< td=""></lq<></td></lq<> | 0.79 ± 0.1 | 2.14 ± 3.0 | 25.79 ± 3.0 | 30.10 ± 11.0 | 2.75 | <lq< td=""></lq<> |
| | C. iguassuensis | NA | 12.05 | NA | 3.48 | 20.35 ± 10.7 | <lq< td=""><td>NA</td><td><pre>CLQ</pre></td><td>14.77 ± 1.9</td><td>15.08</td><td></td><td>51.70</td></lq<> | NA | <pre>CLQ</pre> | 14.77 ± 1.9 | 15.08 | | 51.70 |
| \mathbf{SE} | A. bifasciatus | <pre>CLQ</pre> | 13.17 ± 11.9 | 1.58 ± 0.2 | 0.21 ± 0.3 | 4.82 | <lq< td=""><td><lq< td=""><td>2.79 ± 0.8</td><td>84.77</td><td>72.87 ± 22.8</td><td>8.75 ± 1.8</td><td><lq< td=""></lq<></td></lq<></td></lq<> | <lq< td=""><td>2.79 ± 0.8</td><td>84.77</td><td>72.87 ± 22.8</td><td>8.75 ± 1.8</td><td><lq< td=""></lq<></td></lq<> | 2.79 ± 0.8 | 84.77 | 72.87 ± 22.8 | 8.75 ± 1.8 | <lq< td=""></lq<> |
| | G. brasiliensis | 7.23 ± 0.7 | 10.53 ± 0 | 3.71 ± 0.4 | <lq< td=""><td>0.90 ± 0.3</td><td>≤LQ</td><td>0.90 ± 0.3</td><td>0.41 ± 0.6</td><td>30.49 ± 3.4</td><td>12.05 ± 2.2</td><td>NA</td><td><lq< td=""></lq<></td></lq<> | 0.90 ± 0.3 | ≤LQ | 0.90 ± 0.3 | 0.41 ± 0.6 | 30.49 ± 3.4 | 12.05 ± 2.2 | NA | <lq< td=""></lq<> |
| SS | A. bifasciatus | 3.76 ± 4.1 | 1.65 ± 1.9 | 1.76 ± 0.4 | 1.07 ± 1.3 | 0.96 | 2.90 ± 0.9 | 0.79 ± 0.2 | <pre>CLQ</pre> | 59.68 ± 13.3 | 46.11 ± 4.0 | 13.16 ± 0.4 | 12.79 ± 1.2 |
| | G. brasiliensis | 7.82 ± 0.2 | <lq< td=""><td>3.20 ± 0.1</td><td>NA</td><td>11.84 ± 2.3</td><td>7.87 ± 0.3</td><td>0.93 ± 0.2</td><td>NA</td><td>33.21 ± 4.4</td><td>31.03 ± 0</td><td>NA</td><td><lq< td=""></lq<></td></lq<> | 3.20 ± 0.1 | NA | 11.84 ± 2.3 | 7.87 ± 0.3 | 0.93 ± 0.2 | NA | 33.21 ± 4.4 | 31.03 ± 0 | NA | <lq< td=""></lq<> |
| SO | A. bifasciatus | 2.94 | 3.55 ± 0.3 | 2.30 | 1.18 ± 0.1 | <lq< td=""><td>21.59 ± 15.9</td><td><lq< td=""><td><lq< td=""><td>60.25</td><td>37.31 ± 0.6</td><td>56.06</td><td>11.30 ± 2.0</td></lq<></td></lq<></td></lq<> | 21.59 ± 15.9 | <lq< td=""><td><lq< td=""><td>60.25</td><td>37.31 ± 0.6</td><td>56.06</td><td>11.30 ± 2.0</td></lq<></td></lq<> | <lq< td=""><td>60.25</td><td>37.31 ± 0.6</td><td>56.06</td><td>11.30 ± 2.0</td></lq<> | 60.25 | 37.31 ± 0.6 | 56.06 | 11.30 ± 2.0 |
| | G. brasiliensis | 7.10 ± 0.6 | 3.76 ± 0.4 | 3.65 ± 0.2 | 0.45 ± 0.2 | 16.77 ± 2.2 | <lq< td=""><td>1.12 ± 0.3</td><td><pq <-=""></pq></td><td>34.83 ± 5.4</td><td>51.12 ± 15.3</td><td>NA</td><td>$9,.5\pm0.4$</td></lq<> | 1.12 ± 0.3 | <pq <-=""></pq> | 34.83 ± 5.4 | 51.12 ± 15.3 | NA | $9,.5\pm0.4$ |
| | C. iguassuensis | 1.61 | 5.67 ± 1.5 | 7.74 | 0.77 ± 0.2 | NA | 8.38 ± 5.4 | NA | 0.86 | 41.71 | 47.83 ± 13.9 | 87.94 | 7.54 ± 0.9 |
| $^{\rm SC}$ | A. bifasciatus | 7.96 ± 0.3 | <lq< td=""><td>2.06 ± 0.1</td><td>0.94 ± 0.7</td><td><lq< td=""><td>0.48 ± 0.6</td><td>4.18 ± 1.9</td><td></td><td>28.01 ± 3.4</td><td></td><td>1.27 ± 1.4</td><td></td></lq<></td></lq<> | 2.06 ± 0.1 | 0.94 ± 0.7 | <lq< td=""><td>0.48 ± 0.6</td><td>4.18 ± 1.9</td><td></td><td>28.01 ± 3.4</td><td></td><td>1.27 ± 1.4</td><td></td></lq<> | 0.48 ± 0.6 | 4.18 ± 1.9 | | 28.01 ± 3.4 | | 1.27 ± 1.4 | |
| | G. brasiliensis | 0.63 | 2.76 | <pre>cLQ</pre> | <lq <<="" td=""><td><pq <</pq </td><td><lq< td=""><td>12.68 ± 1.6</td><td></td><td>30.44 ± 4.1</td><td></td><td>>LQ</td><td></td></lq<></td></lq> | <pq <</pq | <lq< td=""><td>12.68 ± 1.6</td><td></td><td>30.44 ± 4.1</td><td></td><td>>LQ</td><td></td></lq<> | 12.68 ± 1.6 | | 30.44 ± 4.1 | | >LQ | |
| | | | | | | | | | | | | | |
| Italic | sized values indicate | e higher conce | entrations for eac | h metal analy | 'zed. Data exl | pressed as avera | ige ± standard de | viation | | | | | |
| FA F | oz do Areia Reserv | oir, SE Segree | do Reservoir, SS | Salto Santiag | o Reservoir, | SO Salto Osóric | Reservoir, SC S | alto Caxias Re | servoir, <lq< td=""><td>Lower than limi</td><td>it of quantificati</td><td>on, NA Not and</td><td>lyzed</td></lq<> | Lower than limi | it of quantificati | on, NA Not and | lyzed |
| | | | | | | | | | | | | | |

 Table 4
 Metals concentrations detected in the muscle of fish from the five reservoirs of Iguaçu River

 Table 5
 Polycyclic aromatic

 hydrocarbons detected in the bile
 of fishes from the five studied

 reservoirs of Iguaçu River in
 summer

| | | PAHs ($\mu g.g^{-1}$) | 2 rings | 4 rings | 5 rings | 6 rings |
|--------|----|-------------------------|--------------------|-------------------|------------------|------------------|
| Summer | FA | A. bifasciatus | 115.95 | 5.91±19.77 | 4.57 | 0.37 ± 39.57 |
| | | G. brasiliensis | 133.52±29.62 | $5.76\!\pm\!2.08$ | 1.23 ± 0.3 | 0.21 ± 0.07 |
| | | C. iguassuensis | 81.54 ± 14.27 | 7.96 ± 4.00 | 3.06 ± 2.19 | 0.55 ± 0.23 |
| | SE | A. bifasciatus | 65.38 ± 24.20 | 9.04 ± 3.95 | 10.49 ± 5.69 | 0.96 ± 0.54 |
| | | G. brasiliensis | 65.76 ± 10.81 | 9.38 ± 1.14 | 3.99 ± 1.40 | 0.60 ± 0.085 |
| | SS | A. bifasciatus | 86.35 ± 24.54 | 11.59 ± 0.94 | 18.71 ± 4.51 | 2.17 ± 0.94 |
| | | G. brasiliensis | 94.35 ± 15.55 | 7.20 ± 4.23 | 2.26 ± 0.20 | 0.38 ± 0.18 |
| | SO | G. brasiliensis | 54.16 ± 25.50 | 15.16±9.61 | 5.8 ± 0.83 | 1.18 ± 0.23 |
| | | C. iguassuensis | 39.59 | 8.56 | 2.42 | 0.45 |
| | SC | A. bifasciatus | 42.78 ± 32.24 | 16.95 ± 2.24 | 17.83 ± 5.07 | 0.92 ± 0.32 |
| | | G. brasiliensis | 22.70 | 4.50 | 1.83 ± 14.2 | 0.30 |
| | | C. iguassuensis | 112.33 ± 69.93 | $5.24\!\pm\!2.15$ | 1.45 ± 0.33 | 0.20 ± 0.03 |

Italicized values indicate higher concentrations for each PAH group analyzed. Data expressed as average \pm standard deviation

FA Foz do Areia Reservoir, SE Segredo Reservoir, SS Salto Santiago Reservoir, SO Salto Osório Reservoir, SC Salto Caxias Reservoir

was higher in samples from the SE reservoir when compared to SS, SO (winter), and SC (summer) (Fig. 2). Elevated lipid peroxidation (LPO) was observed in A. bifasciatus captured during the winter from the FA and SO reservoirs compared to SS and SC reservoirs. The liver of G. brasiliensis had higher activity of CAT in individuals from the SO reservoir when compared to FA and SS (summer), while for winter it was higher in the individuals from SE (Fig. 2). The GPx activity was enhanced in G. brasiliensis individuals from the SS and SO reservoir compared to individuals from the FA site (summer; data not shown). Corroborating with this result, concentrations of non-protein thiols in individuals from FA reservoir were higher than SS individuals. Protein carbonyls were higher in SE than in FA (summer) and SS (winter). No statistical differences were observed between the groups for GST activity and lipid peroxidation in G. brasiliensis (data not shown). The CAT activity in C. iguassuensis was also higher in individuals from SO than SE and SC (summer), but no significant differences were observed for GST activity, PCO (Fig. 2) and LPO analysis (data not shown).

In general, the AChE activity was lower in individuals from SC reservoir than from the other reservoirs, particularly in the muscle of *G. brasiliensis* and *C. iguassuensis* (Fig. 3). For *A. bifasciatus*, brain AChE activity was the highest in the fish from SO (summer) and SC (winter). AChE enzymatic activity was typically higher in summer than winter (Fig. 3).

Genotoxicity biomarkers

A. bifasciatus and *C. iguassuensis* from SC reservoir (summer) had the highest scores of DNA damage in red blood

cells (Fig. 4). Nuclear morphological alterations (NA), such as notched and micronuclei in erythrocytes, were more frequent in *G. brasiliensis* fish from SC reservoir (summer). For *A. bifasciatus*, NA was more frequent in the fish from SE (summer) reservoir.

Histopathological analysis

According to Bernet's Lesion Index for the liver samples, no differences were observed among individuals captured in the studied sites. However, *C. iguassuensis* fish from SC reservoir had higher index than those from SO reservoir (summer; data not shown). In general, hepatic lesions were more frequent in *G. brasiliensis* than in *A. bifasciatus* and *C. iguassuensis*. However, the alterations were more frequent in summer than winter (Table 6). Despite of this, some important histopathological findings were observed in individuals from all reservoirs, such as necrosis, inflammatory responses (leukocytes infiltration, perivascular and perinuclear granulomatosis) and presence of parasites (Fig. 5). Other alterations, such as intracellular deposits and vacuolization that were also observed, indicated physiological and circulatory disturbances in the liver.

Hyperplasia, lamellar fusion, structural alterations of the lamellae and epithelium, and parasites were frequent in the gills of fish from the studied species (Table 6, Fig. 6). The fish captured in FA reservoir had a high incidence of fused lamellae, indicating an alteration in gills normal function. In general, the occurrence of lesions was homogeneously distributed between the species and the reservoirs in the summer, while in winter the variation was restricted between the groups of the same species.

Multivariate analysis

The PERMANOVA test revealed significant differences between sites and seasons for the interaction of all measured variables for the studied species. The principal component analysis (PCA), based on individual biomarker values, was performed in order to plot a unified view of the responses of all studied species (Figure S1, ESM 1).

Integrative analyses

Three different datasets were used to perform FA/PCA, metals levels in the water column and sediment, biomarker responses, and contaminant levels (metals and PAHs in the bile) in the three different fish species for an integrated description of environmental quality in the different sampling sites. Tables showing the factor loadings and the estimated scores of the factors for each case are presented in detail in the ESM 1 (Tables S1 and S2).

The total explained variance ranged from 69.03 to 100 % in all analyses. In the summer, the FA, SE, and SC sites showed relevant scores which were associated with the effects seen in the blood tissue, antioxidant responses (induction or inhibition), biomarkers of damage (including increased AChE levels), and low condition factor. In the results obtained for summer, the sites SS and SO showed no relevant effects in the blood for the three studied species. In these sites, antioxidant responses were not found in *A. bifasciatus*, although G. *brasiliensis* and C. *iguassuensis* showed catalase activity relatively higher in SO.

The results obtained from FA/PCA and the estimation of the relevance of factors for each studied site showed that the FA reservoir, followed by SC, presented the highest metal concentrations in water (summer sampling). On the other hand, the SE and SO site showed the highest number of metals



Fig. 2 Biochemical biomarkers in the liver. *Letters* indicate significant differences between sites in the same season, *a*, *b*. *Asterisk* indicates seasonal differences for the same site



Fig. 3 Acetylcholinesterase activity (AChE) in the brain and muscle. *Letters* indicate significant differences between sites in the same season, *a b. Asterisk* indicates seasonal differences for the same site

in the sediment associated with factors of higher relevance. Different patterns of metals body burden were verified between fish species, with higher bioaccumulation of metals in *A. bifasciatus* from FA and SC, and G. *brasiliensis* collected in the SO reservoir. Regarding the environmental quality in the winter, the results of the FA/PCA also showed a consistency in the damage responses (disturbed AChE activity and higher levels of LPO) and effects in the blood tissue (relatively higher frequencies of blebbed, notched, and mononuclear erythrocytes) in



Fig. 4 Comet assay and nuclear morphological alterations in the blood cells. *Letters* indicate significant differences between sites in the same season, *a b*. Asterisk indicates seasonal differences for the same site

| | Liver | | | | | | | | | | | | Gills | | | | | | | |
|--------------------|------------|---------|-----------|------------|------------|----------|------------------------|------------|----------------------|-------------|-----------|----------|----------|--------|-------------------------------------|----------------|-------------------------------------|------------------|-------------------------|--------|
| | Necrosi | ~ | Parasite | S | Inflamm | lation | Circulatc Disturbaı | ary nce | Melano-mi centers | acrophage | Vacuolat | ion | Lamellar | fusion | Structura alteratior 1st lame | l Is lae | Structura alteratior 2nd lame | ul 15 clae | Epitheliu alteratior | u s |
| | Summer | Winter | Summe | r Winter | Summer | Winter | Summer | Winter | Summer | Winter | Summer | Winter | Summer | Winter | Summer | Winter | Summer | Winter | Summer | Winter |
| FA A. bifasciatus | 86.6 | 80 | 73.3 | 66.7 | 33.3 | 20 | 62.5 | 100 | 62.5 | I | I | I | 12.5 | 40 | 62.5 | 100 | 62.5 | 100 | 75 | 60 |
| G. brasiliensis | 72.2 | 66.6 | 22.2 | I | 22.2 | 33.3 | 20 | | 100 | 66.6 | 33.3 | 100 | | 25 | 20 | | 100 | 50 | | 50 |
| C. iguassuensis | 100 | | | | | | | | | | 09 | | | | | | | | | |
| SE A. bifasciatus | 85.7 | 80 | 78.6 | 80 | 14.3 | 40 | 30 | 85.7 | 10 | Ι | I | Ι | 10 | 14.2 | 30 | 85.7 | 10 | 28.5 | 40 | 71.4 |
| G. brasiliensis | 73.6 | | 21 | | 15.7 | 25 | | 25 | 40 | 50 | 26.3 | 25 | | | | 25 | 40 | 50 | 20 | 50 |
| C. iguassuensis | 100 | | | | 50 | | | | | I | | I | | | | | | | | |
| SS A. bifasciatus | 100 | 93.3 | 50 | 53.3 | 16.6 | 46.6 | 85.7 | 55 | 28.5 | | I | 100 | | | 85.7 | 55 | 28.5 | | 75 | 55.5 |
| G. brasiliensis | 75 | 100 | 25 | | 25 | | 30 | | 60 | I | 43.7 | I | 20 | | 30 | | 60 | 50 | 30 | 25 |
| SO A. bifasciatus | 77.7 | 53.3 | 100 | 80 | I | 66.6 | 55.5 | 83.3 | 44.4 | I | I | I | 22.2 | 16.6 | 55.5 | 83.3 | 44.4 | 16.6 | 33.3 | 66.6 |
| G. brasiliensis | 75 | | 37.5 | | 25 | | | | 55 | | I | | 22 | | | | 55 | | | |
| C. iguassuensis | 100 | | | | 20 | | | | | | 20 | | | | | | | | | |
| SC A. bifasciatus | 87.5 | 71.4 | 62.5 | 71.4 | 25 | 85.7 | 33.3 | | 33.3 | | Ι | | | | 33.3 | | 33.3 | | 100 | |
| G. brasiliensis | 100 | | Ι | | 33.3 | | | | | | Ι | | 66 | | | | | | 66 | |
| C. iguassuensis | ; 100 | 100 | | | 22 | 33.3 | 5.5 | | | | 83 | 33.3 | | | | | | | | |
| | | | | | | | | | | | | | | | | | | | | |
| FA Foz do Areia Re | servoir, S | E Segre | do Reserv | voir, SS S | Salto Sant | iago Res | ervoir, SC |) Salto O | sório Rese | rvoir, SC S | alto Caxi | as Reser | voir | | | | | | | |
| | | | | | | | | | | | | | | | | | | | | |



Fig. 5 Liver sections of Astyanax bifasciatus (light microscopy). a Liver without alterations. b Necrosis (arrows head). c Tissue differentiation (arrows head). d Perivascular granulomatosis (arrows head). e Granulomatosis peritubular (arrows head) (scale $bar = 50 \mu m$). f Inflammatory response (arrows head) and prenecrotic area (arrow). g Hemorrhagic area (arrows head). h and i Invasive and encapsulated neoplasic areas, respectively (arrows head). Stain: Hematoxylin and Eosin. Bars: 100 μm (c, i), 50 μm (other images)



A. bifasciatus and *C. iguassuensis*. These damage responses were associated with levels of metals in water and sediment from the SC site. In turn, few biological responses of relevance were verified for both *A. bifasciatus* and *G. brasiliensis* from SE reservoir, while SS showed a consistency on antioxidant responses during winter regardless the species, suggesting an intermediate environmental quality.

Discussion

The results of the current study show that all of the reservoirs along the Iguaçu River, especially FA, SE, and SC reservoirs, are jeopardized by human activities. The Metropolitan region of Curitiba is a primary source of pollutants while other diffuse sources are also found along the river. The levels of metals detected in water, sediments, and fish muscle from the five reservoirs were either similar or higher than concentrations reported for other impacted aquatic environments (Authman et al. (2012), Maceda-Veiga et al. (2013), Noël et al. (2013) and Voigt et al. (2014), This shows that majority of toxic metals in impacted environments are bioavailable. Furthermore, relevant biological responses observed in fish through the multi-biomarker approach used in this study demonstrate an impact on the health of these organisms.

No clear gradient of pollution was verified along the Iguaçu River course

According to the results obtained from chemical and biological analyses, two aspects are relevant to consider the degree of pollution in the reservoirs: (i) the proximity to the MRC and (ii) the presence of diffuse sources of pollutants along the Iguaçu River. The integrated analysis was able to conclude that there was no clear gradient of pollution between the five reservoirs. The conditions of FA and SE reservoirs confirmed that MRC and others small cities were important sources of pollutants from domestic and industrial wastes to the Iguaçu River. Additionally, the SC reservoir had the poorest environmental quality found possibly due to the intense agricultural activity in that region. The FA/PCA analysis result reinforces



Fig. 6 Gills of Astyanax bifasciatus (SEM). a FA winter: fusion of secondary lamellae (arrowhead) and the damages on lamellae epithelia (respectively a1 and a2). b FA summer: secondary lamellae fusion and aneurisms (arrowhead). c SE winter: fusion (arrowhead) and disarrange on secondary lamellae. d SE summer: damages on secondary lamellae (arrowhead), epithelial cell death (d1) and secondary lamellae fusion (d2). e SO winter: secondary lamellae fusion (arrowhead) and epithelial cell death. f SO summer: intense epithelial damages (arrow) as detailed in (f). g SS winter: damages in primary (arrowhead) and secondary lamellae epithelia. h SS summer: few damages are observed, except for the occurrence of epithelial effects on secondary lamellae

that SS and SO reservoirs had the better environmental condition compared to the other reservoirs. However, when considering seasonal effects, the best environmental condition was observed for SE and SS reservoirs in the winter. On the other hand, the worst condition in winter showing greater environmental impact was different between the fish species: FA reservoir for *G. brasiliensis*, SO reservoir for *A. bifasciatus*, and SC reservoir *C. iguassuensis*. These findings indicate a difference in the sensitivity of the different species analyzed for the pollutants present in each reservoir.

Seasonal effects and others factors influencing biological responses of the three fish species

Overall, the integrated set of biomarkers and PERMANOVA analysis confirmed that the biological responses suggest that individuals were more exposed to more pollutants during summer than in winter. The concentration of chemicals could be higher due to the lower water level in the reservoirs during the summer, especially in the FA reservoir. Furthermore, biological activity may also be correlated with the higher temperature during this season. Based on the physicochemical analysis, the five reservoirs presented distinct abiotic factors, such as temperature, that may interfere with chemical bioavailability (Fujita et al. 2001; Malczyk and Branfireun 2015) and affect biological responses (Elahee and Bhagwant 2007; de la Haye et al. 2012; Wäge et al. 2015).

Organic matter content in water, associated with domestic sewage, may also influence these responses, lead to eutrophication, and consequently to cyanobacteria proliferation (Yan et al. 2016; Thevenon et al. 2011). According to a recent report (Ide et al., 2016), organic matter content (dissolved organic carbon, nitrite, nitrate, orthophosphate, total nitrogen, and phosphate) and the density of phytoplankton decreased significantly along the Iguaçu River. This study, therefore, suggests a pollution gradient of domestic waste from the origin toward the river's mouth. However, significant variation in these parameters was observed only for the first 100 km range of the Iguaçu River from the MRC; between the five reservoirs, there was no longer evident variation. Therefore, the gradient of pollution was verified only before the reservoirs.

Physicochemical parameters are not enough to predict the chemicals effects on fish health

Although physicochemical analysis is utilized to evaluate water quality by environmental agencies in Brazil, they are not enough to predict the real risk of aquatic organisms that are exposed to many pollutants. According to the previous report published by the State Agency for Environment Control - IAP (IAP 2008), the reservoir with the best water quality in Iguaçu River was SC. However, SC reservoir had the worst water quality in the present study. FA reservoir was considered by the agency (IAP) as the worst in terms of water quality, particularly due to the high deficit of oxygen, chlorophyll a, and cyanobacteria blooms (IAP 2008). Water quality gradually improved in the subsequent reservoirs, as these parameters decreased (IAP 2008). However, the current study presents a different view of the water quality in the reservoirs, providing strong evidences that there are other sources of contamination along the river, which refute the hypothesis of a gradient of pollution between the reservoirs.

The five reservoirs are differently impacted by sources of pollution

The FA reservoir is the closest to MRC, the main source of pollutants. Data from this study confirm that the FA site is a highly impacted, showing high levels of metals (Cr, Zn, and Mn) in the water and fish's muscle, high concentrations of microcystin-LR in the water, and 2-ring PAHs in the fish's bile. Biological responses, such as high LPO, HSI, and gill lesions in the three fish species, corroborate this condition. Zn and Mn are both ubiquitous elements, essential for normal physiological function and are indicators of human activity. However, deficiency or excess of these metals can be harmful (Giardina et al. 2009). They are known to inhibit antioxidant responses (Qu et al. 2014) and affect the nervous system, including changes of behavior and altered cognitive function (ATSDR - Public Health Statement: Cadmium. 2999). The concentrations of Cr in water were above the established by Brazilian legislation. Cr can bind to biomolecules, resulting in lipid peroxidation, protein carbonylation, and DNA damage (Mattagajasingh et al. 2008; Nickens et al. 2010). Finally, the highest levels of 2-ring PAHs were measured in the three species of fish from FA reservoir, indicating the influence of industrial activities in the MRC, such as crude oil refinement (Gallotta and Christensen, 2012; Leite et al., 2011).

The SE reservoir is the second furthest from MRC and, according to our data, also presented poor water quality. The reservoir had low pH, contained Cd in the water, Mn in the sediment, and elevated carbonylated proteins in the fish's liver. Acidic condition increases metal solubility in water and can interfere with normal gill osmoregulation, acid–base balance, and uptake/homeostasis of essential elements like Zn, Fe, and Cu in fish (Levit and Bozeman, 2010; Qu et al. 2014).

Metals are ubiquitous elements, but industrial and agricultural activities can increase metal concentration in the environment. Fertilizers can contain Cd (WHO 2011), as well as Co, Cu, Zn, Pb, and Ni as impurities, and herbicides present high levels of Fe, Mn, Pb, and Ni (Gimeno-García et al. 1996). Culture of soybeans, wheat, and corn increase from SE until SC reservoir, representing a potential source of pesticides for these sites.

SS and SO are, respectively, the third and fourth intermediate reservoirs, located in a region of intense agricultural activity. Considering their distances from MRC, other sources of contaminants may be responsible for the low dissolved oxygen, high levels of Cr in the water, low condition factor of the fish in SS reservoir, as well as toxic concentrations of Pb, Cu, and Zn in the sediment, and Pb in the muscle in SO reservoir. These metals may lead to gills dysfunction, oxidative damage (Wei and Yang, 2015), apoptosis (Luzio et al. 2013; Monteiro et al. 2009), and neurotoxicity (White et al. 2007).

SC reservoir is the furthest from MRC, but it has the high concentrations of Cd and Ni in the water, Cd in the muscle, low AChE activity, and high DNA damage in blood cells in all three fish species analyzed. Non-essential metals like Cd can cause deleterious effects on biota even at low concentrations (Satarug et al., 2009). Although the concentration of Cd was below the established level set by the Brazilian legislation, Cd is a human carcinogen, a potent endocrine disruptor, and exerts toxic effects on kidney, skeletal, respiratory, and reproductive systems (Gerbron et al. 2015; Jiménez-Ortega et al. 2012; Wang et al. 2014; WHO 2011). Mn and Pb can inhibit AchE (Santos et al. 2012), and mixtures of Pb+Cd (detected in SC) can induce neuronal degeneration in the brain even at low concentrations (Cobbina et al. 2015).

In summary, independent of the distance from MRC, the studied reservoirs are impacted, at distinct levels, by chemicals from distinct sources.

The presence of complex mixtures in aquatic ecosystems complicate environmental risk assessment

Assessing chemical interactions (e.g., additive, synergistic, or antagonistic effects) is a great challenge for ecotoxicologists (Norberg et al. 2015). According to Gauthier et al. (2014), metal-PAH mixtures can have additive effects, e.g., alteration of membrane permeability to metals and mutual inhibition of detoxification by CYP biotransformation pathway. Mixture of the carcinogens such as Cd and benzo(a)pyrene (B[a]P) can impair the activity of antioxidant enzymes, though these chemicals can induce them when separate (Costa et al. 2010a, 2010b). This scenario illustrates the complexity in establishing the interaction between natural exposure to pollutants and biological effects. Biogeochemistry can only partially explain the differences of metal concentrations in water, sediment, and muscle of fish, whereas different metabolic roles of essential and non-essential elements, environmental factors (Monroy et al. 2014), and food habit (Kalantzi et al. 2013) are related to the distinct bioaccumulation of metals in the three fish species.

A. bifasciatus reflected the environmental quality better than G. brasiliensis and C. iguassuensis

A. bifasciatus represented a good bioindicator of biomonitoring study to the Iguaçu River, since its biological responses reflected the environmental quality better than the other two fish species in the current study. The same conclusion had been reported by Freire et al. (2015) for this species. *A. bifasciatus* is widely distributed in the Iguaçu River (Daga and Gubiani 2012; Pie et al. 2009), and studies in different regions of Brazil and South America also describe the gender *Astyanax* sp. as a good bioindicator for biomonitoring (Alberto et al. 2005; de Lemos et al. 2008; Paulino et al. 2014; Prado et al. 2011, 2014).

Different biomarkers showed that the health of fish is compromised

Besides the importance of using different species in biomonitoring programs, it is also crucial to evaluate several responses in target organs to better comprehend the health status of the fish. Biochemical and genotoxicity analysis demonstrated alterations of enzymes involved in antioxidant defenses and nervous system, as well as damages in lipids, proteins, and DNA. These responses were correlated to the bioavailability of metals and PAHs, which induce several lesions and alterations in target tissues, as described by Liu et al. (2010, 2011).

Despite of the similar indexes of lesion among the groups, important pathological changes were observed in the liver and gills of the three fish species. Histopathological findings in target organs represent a strong evidence of chemical exposure in the field. The liver is described as a key organ for biotransformation of chemicals in vertebrates and a potential target for chemical-induced injury. Some chemicals as well as metals can lead to necrosis of hepatocyte cells (de Oliveira Ribeiro et al. 2002; Sánchez-Chardi et al. 2009), DNA fragmentation, and redox unbalance (Proskuryakov et al. 2003). In the gills, some pathological alterations such as primary and secondary lamellae damage, fusion of adjacent lamellae, neoplasia, and aneurisms are characteristic of waterborne exposure to chemicals (Liu et al. 2010; Srivastava et al. 2009). Incidence of lesions in target organs, evidences of neurotoxicity, and macromolecules damage associated with bioaccumulation of toxic metals and PAHs indicated the impact on fish health related to low water quality of the five reservoirs.

Integration of multiple biomarkers and chemical analysis is an efficient strategy for evaluation of water quality

The integration of biomarkers in fish and metal detection represents an efficient strategy for evaluation of water quality and risk assessment of human exposure (Carvalho et al. 2012; Omar et al. 2014; Sakuragui et al. 2013; Gusso-Choueri et al. 2015). An integrated analysis was performed through FA/PCA in order to improve the environmental diagnosis, since it can show the cases when relevant responses are significantly associated with contaminants present in the reservoirs to show the environmental quality of a site.

Final considerations

The results of this study provide strong evidence that human activities greatly impact the five reservoirs of the Iguaçu River. Domestic and industrial wastes from MRC and other small cities were considered as the main responsible to impact FA reservoir even with a considerable distance from this source of pollutants. Water quality of the second reservoir, SE, is impacted by what compounds from the MRC and small cities and agricultural activities. SS and SO reservoirs are primarily subject to pesticides inputs, with less impact from MRC. Finally, SC reservoir had the poorest water quality, indicating that there is no gradient of pollution between the five reservoirs from the origin of the river toward the mouth.

The effects of the chemicals described here demand a risk assessment for the biota and human populations in this area since the Iguaçu basin has a high environmental and socioeconomical importance. These sets of results represent the first study performed in the reservoirs of Iguaçu River showing the impacts in fish health and can be used for its management or to establish conservation politics.

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