

# Effect of alginate on the aggregation kinetics of copper oxide nanoparticles (CuO NPs): bridging interaction and hetero-aggregation induced by $\text{Ca}^{2+}$

Lingzhan Miao<sup>1,2</sup> · Chao Wang<sup>1,2</sup> · Jun Hou<sup>1,2</sup> · Peifang Wang<sup>1,2</sup> · Yanhui Ao<sup>1,2</sup> · Yi Li<sup>1,2</sup> · Bowen Lv<sup>1,2</sup> · Yangyang Yang<sup>1,2</sup> · Guoxiang You<sup>1,2</sup> · Yi Xu<sup>1,2</sup>

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**Abstract** The stability of CuO nanoparticles (NPs) is expected to play a key role in the environmental risk assessment of nanotoxicity in aquatic systems. In this study, the effect of alginate (model polysaccharides) on the stability of CuO NPs in various environmentally relevant ionic strength conditions was investigated by using time-resolved dynamic light scattering. Significant aggregation of CuO NPs was observed in the presence of both monovalent and divalent cations. The critical coagulation concentrations (CCC) were 54.5 and 2.9 mM for  $\text{NaNO}_3$  and  $\text{Ca}(\text{NO}_3)_2$ , respectively. The presence of alginate slowed nano-CuO aggregation rates over the entire  $\text{NaNO}_3$  concentration range due to the combined electrostatic and steric effect. High concentrations of  $\text{Ca}^{2+}$  (>6 mM) resulted in stronger adsorption of alginate onto CuO NPs; however, enhanced aggregation of CuO NPs occurred simultaneously under the same conditions. Spectroscopic analysis revealed that the bridging interaction of alginate with  $\text{Ca}^{2+}$  might be an important mechanism for the enhanced aggregation.

Furthermore, significant coagulation of the alginate molecules was observed in solutions of high  $\text{Ca}^{2+}$  concentrations, indicating a hetero-aggregation mechanism between the alginate-covered CuO NPs and the unabsorbed alginate. These results suggested a different aggregation mechanism of NPs might co-exist in aqueous systems enriched with natural organic matter, which should be taken into consideration in future studies.

**Keywords** CuO nanoparticles · Aggregation kinetics · Alginate · Divalent electrolytes · Bridging interaction · Hetero-aggregation mechanism

## Introduction

Copper oxide nanoparticles (CuO NPs) have been widely used in a variety of industrial products such as anti-bactericide coatings, biomedicines, and toothpastes (Applerot et al. 2012; Zhou et al. 2006). Due to the large quantities produced and their widespread application, engineered nanoparticles will be inevitably released into aqueous environment (Nowack and Bucheli 2007). It has been demonstrated that CuO NPs have an acute toxic effect on bacteria (Karlsson et al. 2008; Zhao et al. 2013), decrease cell viability of activated sludge (Hou et al. 2015; Zhou et al. 2015), and cause oxidative stress in microalgae (Aruoja et al. 2009). According to recent studies, once released into aquatic environments, nanoparticles may interact with naturally occurring water components, which consequently may affect the potential toxicity and fate of nanoparticles in the environment (Burkart et al. 2015; Grillo et al. 2015; Nowack et al. 2012).

Stability of these highly reactive materials in aquatic environment is an essential parameter to evaluate their fate,

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✉ Jun Hou  
hhuhjyhj@126.com

✉ Peifang Wang  
pfwang2005@hhu.edu.cn

<sup>1</sup> Key Laboratory of Integrated Regulation and Resources Development on Shallow Lakes, Ministry of Education, Hohai University, Nanjing 210098, People's Republic of China

<sup>2</sup> College of Environment, Hohai University, 1 Xikang Road, Nanjing 210098, People's Republic of China

bioavailability, and potential toxic effects towards living organisms (Aiken et al. 2011; Miao et al. 2014; Philippe and Schaumann 2014). Existing scientific evidence suggested that a wide range of environmental factors, such as the particle properties (size, surface charge, morphology) and water chemistry (pH, ionic strength, redox conditions, natural organic matter (NOM)) will inevitably interact with CuO NPs and influence their physicochemical properties (aggregation, sedimentation, and dissolution) (Aiken et al. 2011; Chang et al. 2012; Nowack et al. 2012; Yang et al. 2013). It has been demonstrated that the solution pH can influence the surface charge through a pH-dependent protonation and deprotonation of surface hydroxyl groups (Philippe and Schaumann 2014), which affects the aggregation and transport of NPs. Effects of various electrolytes were also investigated, and divalent ions can promote agglomeration of NPs due to compression of the electrical double layer (Son et al. 2015). Typically, higher NOM concentrations result in enhanced NP stability (Adeleye et al. 2014; Grillo et al. 2015): more adsorbed mass of NOM on the NPs induces electrostatic repulsion (attributable to negatively charged carboxylate groups) and electrosteric repulsion for high molecular weight components (Philippe and Schaumann 2014; Zhang et al. 2009).

In addition to humic substances, a non-humic aquagenic (i.e., produced in the water column) fraction also accounts for an important fraction of NOM in natural environments. A representative non-humic fraction NOM is polysaccharides, which are widespread in aquatic systems and make up 10–30 % of the NOM in lakes (Chen et al. 2006; Wilkinson et al. 1997). Given the presence of both nanoparticles and polysaccharides in the environment, it is important to understand how they interact with each other and how those interactions may affect the transport and fate of NPs. Although some studies have demonstrated that polysaccharides can destabilize nanoparticle aggregates (Chen et al. 2006; Loosli et al. 2015), their role in influencing the aggregation kinetics of nanoparticles in complex water conditions is still not well understood. Furthermore, polysaccharide molecules can interact with themselves, leading to the formation of molecular aggregates dependent on the environmental conditions. In the presence of multivalent cations,  $\text{Ca}^{2+}$  for example, aggregation of polysaccharides is induced by charge neutralization and cation bridge formation between different polysaccharide molecules (Liu et al. 2010; Li and Yu. 2014; Wang et al. 2013). Consequently, the aggregates of these biomacromolecules may influence the aggregation process of CuO NPs in divalent electrolytes. Therefore, it is of significant interest to systematically study how the polysaccharides affect fundamental processes such as the aggregation of CuO NPs, both in the presence and absence of divalent electrolytes, which is imperative to elucidate the toxicity and risks of CuO NPs in natural environments.

The overall objective of this study is to systematically investigate the interaction between alginate with CuO NPs in monovalent and divalent electrolyte solutions. To this end, alginate (model polysaccharides) was comprehensively characterized, and the impacts on CuO NP aggregation were investigated in monovalent ( $\text{Na}^+$ ) or divalent ( $\text{Ca}^{2+}$ ) electrolyte solutions at varying concentrations. The aggregation kinetics of CuO NPs in the absence or presence of alginate was examined through dynamic light scattering (DLS) and zeta potential. Batch sorption experiments were conducted to measure both the equilibrium and kinetics of alginate sorption onto CuO NPs, and FTIR analysis was performed to elucidate the interactions between alginate and nano-CuO. Transmission electron microscopy (TEM) and scanning electron microscope (SEM) were used to delineate the mechanisms for CuO NP stabilization and destabilization under various conditions.

## Materials and methods

### Chemical reagents

The chemical reagents used in this study, including HCl, NaOH,  $\text{NaNO}_3$ , and  $\text{Ca}(\text{NO}_3)_2$ , were purchased from Aladdin Reagent (Shanghai, China) and at least p.a. grade. The chemical stock solutions were prepared separately by dissolving chemicals into Milli-Q water ( $18.2 \text{ M}\Omega \text{ cm}^{-1}$ , Millipore) and filtered through  $0.22 \mu\text{m}$  filters (Millipore). All samples, unless otherwise specified, were prepared at pH 6.8 (nearly neutral solution). The pH of all solutions was adjusted to 6.8 with  $1 \text{ mmol L}^{-1}$  NaOH or HCl, measured by a pH meter (Hitachi, Japan). Sodium alginate (lyophilized powder, >98 %) was obtained from Sigma-Aldrich (St. Louis, MO) and used as received (model polysaccharides). The molecular weight of alginate ranged from 12 to 80 kDa, with 61 % mannuronic acid and 39 % guluronic acid (Liu et al. 2010). The powder was analyzed by FTIR spectroscopy (Tensor 27, Bruker). To obtain the stock solutions, 200 mg powder sodium alginate was weighed into 500 mL Milli-Q water ( $18.2 \text{ M}\Omega \text{ cm}^{-1}$ , Millipore) and stirred overnight to ensure total dissolution. Then, the alginate solution was filtered through a  $0.22 \mu\text{m}$  cellulose membrane and kept in the dark at  $4 \text{ }^\circ\text{C}$ . The total organic carbon content in the alginate stock solution was measured at  $106.8 \text{ mg/L}$  TOC (Liqui TOC II, Elementar, Germany).

### Preparation of CuO NP dispersions

In this study, CuO nanopowder (vendor-reported size  $<100 \text{ nm}$ ) was purchased from Sigma-Aldrich (St. Louis, MO) (Table 1 for detailed properties). Stock suspension of CuO NPs was prepared by adding  $0.5 \text{ g}$  of CuO NPs to  $1 \text{ L}$

of Milli-Q water. Then, the stock suspension was probe-sonicated in an ice-water bath to avoid uncontrolled heating at 25 °C. Firstly, the effects of ultrasonic time (5, 10, 15, 20, 25, and 30 min) and ultrasonic power (100, 200, 400, and 600 W) (ultrasonic cell crusher, Biosafe 1200-98) on the particle size distribution of CuO NPs were investigated in order to obtain the best dispersion procedure. Then, according to the optimal treatment, the stock suspension was prepared and the physicochemical properties of CuO NPs were characterized using dynamic light scattering (DLS) using Malvern Zetasizer (Nano ZSP, Malvern Instruments, UK) and UV-vis spectroscopy. The zeta potential of CuO NPs was measured in Milli-Q water over a wide range of pH values (5–10.5) at 25 °C. The concentration of the suspended nanoparticles used was adequate to create sufficient scattered intensity. The morphology and particle size of the CuO NPs in the stock suspension were characterized using a JEM-2100 transmission electron microscopy (TEM, JEOL Co., Japan).

### Aggregation kinetics of CuO NPs

Time-resolved DLS (TR-DLS) measurements using a Malvern Zetasizer (Nano ZSP, Malvern Instruments, UK) were performed to study the increase of nano-CuO intensity-weighted averaged hydrodynamic diameter (HDD) with time (Shen et al. 2015). The detector employed a He-Ne laser at a wavelength of 633 nm and detection angle of 173° with each autocorrelation function being accumulated for 10 s. High polydispersity limits the reliability of nanoparticle HDD values (Kroll et al. 2014), and thus, data sets with a polydispersity index (PDI) above 0.7 were not taken into account. The measurements were conducted at 25 °C, and HCl and NaOH were used to adjust pH to 6.8.

The aggregation kinetics of CuO NPs was first tested in two electrolyte solutions (0–500 mM NaNO<sub>3</sub> and 0–50 mM Ca(NO<sub>3</sub>)<sub>2</sub>) without alginate addition. The effect of alginate on the aggregation of CuO NPs was investigated with aqueous alginate concentrations of 5 mg/L C (corresponding to typical freshwater natural organic matter concentration) (Loosli et al. 2015). In these experiments, a certain amount of electrolyte stock solution was added into the DLS cuvette which contained a predetermined volume of diluted nano-CuO stock solution and alginate so that the total volume of the final suspension was 1 mL. The concentration of CuO NPs in the DLS vial was maintained at 100 mg/L. Then, the tested suspensions were vortexed (WH-2, Huxi Analysis Instrument Factory, Shanghai) for 1 s immediately after mixing and then placed in the Zetasizer (Huangfu et al. 2013). At the initial stage of aggregation, the aggregation rate constant is directly proportional to the initial aggregation rate for Rh with time. Then, the particle attachment efficiency ( $\alpha$ , i.e., inverse

stability ratio,  $1/W$ ) was evaluated, which was defined as the  $k$  normalized by the aggregation rate constant measured under a diffusion-limited (fast) regime (Chen et al. 2006; Shen et al. 2015). Details of the TR-DLS measurements, calculation methods for attachment efficiency ( $\alpha$ ), and critical coagulation concentration (CCC) are given in the [supplementary material](#).

### Zeta potential measurement

The electrophoretic mobility of CuO nanoparticles was evaluated by a Zetasizer (Nano ZSP, Malvern Instruments, UK) and converted to zeta potential using the Smoluchowski equation (Baalousha 2009). EPMs of CuO NPs in alginate-free electrolyte solutions were measured at a range of NaNO<sub>3</sub> and Ca(NO<sub>3</sub>)<sub>2</sub> concentrations. Sample preparation was performed in the same way as the samples for the aggregation experiments. At least ten measurements were conducted for each sample employed. For measurements in the presence of polysaccharides, a concentration of 5 mg/L C alginate was employed. The measurements were taken at room temperature (25 °C), and all the experiments were run in duplicate or triplicate.

### Spectroscopic analysis

TEM (JEM-2100), SEM (Hitachi S-4800), and energy-dispersive spectroscopy (EDS) analyzer were used to analyze the aggregates of the CuO NPs in the Ca(NO<sub>3</sub>)<sub>2</sub> solution in the presence and absence of alginate. For the sample preparation, several drops of the CuO NP suspension were deposited onto a 300-mesh carbon-coated copper grid (Nanjing Zhongjing Technology Co., China) in a dust-free condition (Huangfu et al. 2013). The excess water was then removed with filter papers and the grid was air-dried for 30 min. Then, at least 20 representative images were taken for each sample. Simultaneously, EDS spectra were recorded at selected areas on the aggregate surface to obtain information on the surface atomic distribution and chemical composition.

Alginate-adsorbed nano-CuO powder was recovered by centrifuging the suspension and drying in a vacuum oven at 60 °C overnight. FTIR spectra of sample pellets with KBr were recorded on a Fourier transform infrared (FTIR) spectrometer (Tensor 27, Bruker). Infrared spectra of all nanoscale materials were collected from 4000 to 400 cm<sup>-1</sup> range at room temperature (25 °C).

## Results and discussion

### Characterization of CuO NPs

Major physicochemical properties of CuO nanoparticles are summarized in Table 1. A TEM image of CuO NPs is provided

**Table 1** Summary of CuO nanoparticle properties used in this study

Parameter	Value
Density (g/cm <sup>3</sup> )	6.372
Vendor-reported size (nm)	60–100
TEM particle size measured (nm) ( <i>n</i> = 20)	92 ± 12
DLS HDD measured in Milli-Q water (nm) ( <i>n</i> = 12)	281 ± 27
BET specific surface area measured (m <sup>2</sup> /g) ( <i>n</i> = 3)	29 ± 3
Point of zero charge	8.8
Zeta potential (mV) in Milli-Q water (pH = 7)	+21.3 ± 1.6
Purity by ICP-MS (wt%)	98.81

The measured nano-CuO sizes showed significantly different between TEM and DLS measurements ( $p < 0.05$ ). The purity of CuO NPs used in this study was determined through acid (HNO<sub>3</sub>) digestion, and the concentrations of Cu<sup>2+</sup> were analyzed by inductively coupled plasma mass spectrometry (ICP-MS, Agilent 7700×)

HDD hydrodynamic diameter

in Fig. S1, and the presence of CuO was evidenced by the FTIR spectra in Fig. S2 in the SI. The effects of ultrasonic time (5, 10, 15, 20, 25, and 30 min) and ultrasonic power (100, 200, 400, and 600 W) on the particle size distribution of CuO NP stock (500 mg/L) in Milli-Q water were investigated by DLS measurements. As shown in Figure S3(A), the dispersion stability of CuO NP suspension was jointly affected by both the sonication time and the ultrasonic power. The best dispersion procedure was obtained by 25-min sonication with ultrasonic power of 400 W, and no significant difference in aggregate size distributions was observed increasing the ultrasonic time over 25 min with ultrasonic power over 400 W.

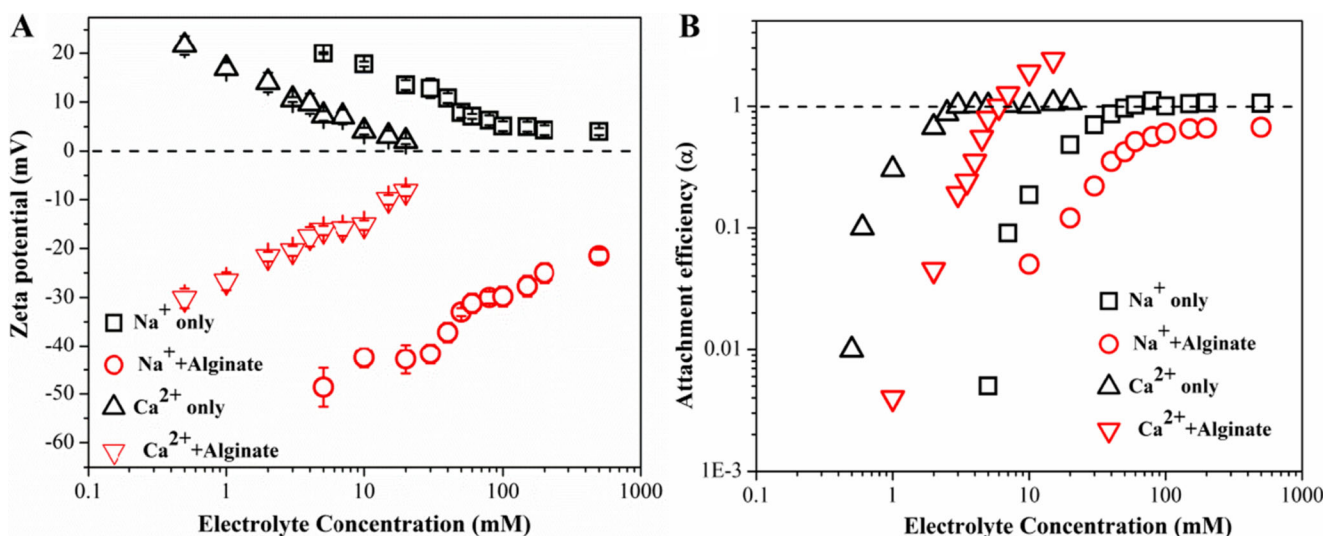
According to the optimal treatment, the stock suspension was prepared after 30-min sonication with ultrasonic power of 600 W. CuO NPs in the stock suspension showed a positive charge with a zeta potential of +21.3 ± 1.6 mV. The intensity-weighted mean hydrodynamic diameter of CuO NPs in the stock suspension measured by DLS was 273 ± 37 nm (polydispersity index (PDI), 0.279) (Fig. S3(B)), indicating the majority were in well dispersion in Milli-Q water. Much larger size of CuO NPs measured by DLS compared to the primary size (60–100 nm) and the TEM measurements ( $p < 0.05$ ) is likely due to the formation of nano-CuO aggregates, and the diameter measured by DLS is a value that refers to how a particle moves within a liquid (called the hydrodynamic diameter) (Majedi et al. 2014; Zhang et al. 2012). Zeta potential was measured at various pH values as shown in Fig. S4. The zeta potential is positive between pH 5 and 6 and is zero at pH 8.8, above which it becomes negative. These results are in good agreement with those reported by Sousa and Teixeira (2013), who have determined the point of zero charge for CuO NPs at pH ~9.

### CuO NP aggregation kinetics with monovalent and divalent cations

The effects of monovalent and divalent cations on CuO NP aggregation were studied with 0–500 mM NaNO<sub>3</sub> and 0–20 mM Ca(NO<sub>3</sub>)<sub>2</sub> (at pH 6.8). Representative aggregation profiles of CuO NPs obtained from TR-DLS measurements are presented in Figure S5. For the two electrolytes, the increase in electrolyte concentrations leads to a corresponding increase in the aggregation rate as a function of time up to a certain concentration above which the aggregation rate does not increase any further. These aggregation behaviors of CuO NPs in NaNO<sub>3</sub> and Ca(NO<sub>3</sub>)<sub>2</sub> solution are consistent with the DLVO theory. The DLVO-type interactions were also discussed for NP aggregation kinetics (e.g., MnO<sub>2</sub> NPs and fullerenes (C<sub>60</sub>)) (Huangfu et al. 2013; Shen et al. 2015).

The zeta potentials (in mV) and representative  $\alpha$  measured over a range of monovalent (NaNO<sub>3</sub>) and divalent (Ca(NO<sub>3</sub>)<sub>2</sub>) electrolyte concentration (at pH 6.8) are presented in Fig. 1. CuO nanoparticles (with positive charge) exhibited a decrease in surface charge with increasing electrolyte concentration (Fig. 1a). It is proposed that NaNO<sub>3</sub> and Ca(NO<sub>3</sub>)<sub>2</sub> ions could cover the nanoparticle surface and form an ion shield, which can promote nano-CuO aggregation (Khan et al. 2011; Shih et al. 2012). In addition, Ca(NO<sub>3</sub>)<sub>2</sub> showed a higher efficiency in increasing zeta potential than the NaNO<sub>3</sub>, and the zeta potential rapidly reduced from +21 to +2 mV when the concentration of Ca(NO<sub>3</sub>)<sub>2</sub> was increased from 0 to 20 mM (Fig. 1a). Because NPs tend to aggregate when their surface charge is close to zero, the more significant influence of divalent counter-ions on CuO NP aggregation than the monovalent counter-ions was suggested through the changes of zeta potential (Son et al. 2015; Sousa and Teixeira 2013). The aggregation attachment efficiency ( $\alpha$ ) analysis of CuO NPs in the NaNO<sub>3</sub> and Ca(NO<sub>3</sub>)<sub>2</sub> solutions clearly depicted a reaction-limited (slow) and diffusion-limited (fast) aggregation regime (Fig. 1b). The minimum electrolyte concentration required for fast aggregation was defined as critical coagulation concentration (CCC). In this study, extrapolation between the two regimes yielded CCC values of 54.5 and 2.9 mM for NaNO<sub>3</sub> and Ca(NO<sub>3</sub>)<sub>2</sub>, respectively, which were comparable with the reported results for CuO NPs in previous studies (Adeleye et al. 2014; Sousa and Teixeira 2013). Increasing the IS and divalent cation concentrations results in the compression of the electrostatic double layer because inter-particle repulsions diminish (Zhang et al. 2009). The same phenomena were also observed in the case of MnO<sub>2</sub> colloids (Huangfu et al. 2013) and Ag NPs (Khan et al. 2011).





**Fig. 1** **a** Zeta potential of CuO NPs as a function of NaNO<sub>3</sub> and Ca(NO<sub>3</sub>)<sub>2</sub> concentration in the absence or presence of 5 mg/L C alginate; **b** aggregation attachment efficiencies of CuO NPs as a

function of NaNO<sub>3</sub> and Ca(NO<sub>3</sub>)<sub>2</sub> concentration in the absence or presence of 5 mg/L C alginate. Aggregation experiments and zeta potential measurements were conducted at pH 6.8 and 25 °C

**Increased stability of CuO NPs with monovalent cations in the presence of alginate**

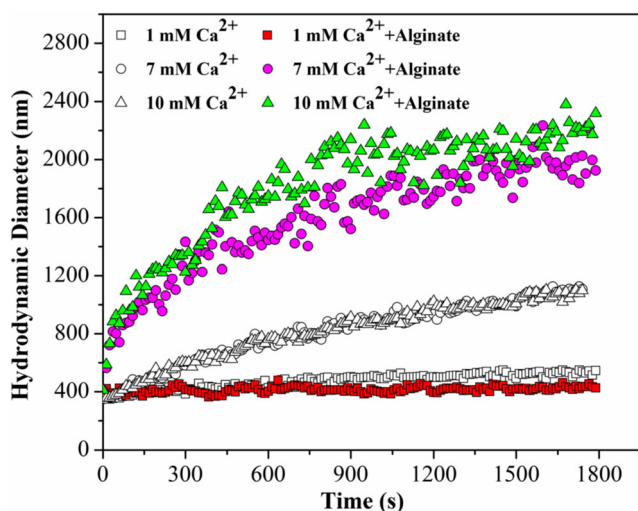
The effects of alginate on the aggregation and stability of CuO NPs were investigated with a wide range of Na<sup>+</sup> and Ca<sup>2+</sup> concentrations. As shown in Fig. 1, nano-CuO aggregation was significantly suppressed in the presence of alginate in Na<sup>+</sup> solution and similar results were also obtained in the case of TiO<sub>2</sub> NPs (Gallego-Urrea et al. 2014; Loosli et al. 2015) and hematite nanoparticles (Chen et al. 2006). Over the entire NaNO<sub>3</sub> concentration range, the values of nano-CuO attachment efficiency in the presence of alginate were lower than those in the absence of alginate (Fig. 1b), indicating that alginate enhanced the stability of CuO NPs. According to the results of zeta potential, the surface charge of CuO NPs changed from positive to negative on the addition of alginate for the NaNO<sub>3</sub> systems (Fig. 1a), which indicated that the negatively charged functional groups of alginate such as carboxylates were attached to the surface of the CuO NPs. These results suggested the relative importance of electrostatic repulsive interactions. Similar effect of alginate on particle charge was observed in the case of hematite as well (Chen et al. 2006). With the electrolyte concentration increasing, however, negative zeta potentials of alginate-coated nanoparticles decreased, since the charges would have been effectively screened by the high NaNO<sub>3</sub> concentration. The right shift of CCC and attenuation of α values strongly indicated that the steric hindrance combined with electrostatic effect was the predominant mechanism affecting the stability of CuO NPs in solutions of alginate/NaNO<sub>3</sub>. Previously, the

stabilizing effect of HA on magnetite nanoparticles was attributed to the combined electrostatic and steric effect (Illés and Tombácz 2006).

**Enhanced aggregation of CuO NPs with divalent cations in the presence of alginate**

Accordingly, the aggregation of CuO NPs in the presence of alginate and Ca(NO<sub>3</sub>)<sub>2</sub> is different from that in the presence of alginate and NaNO<sub>3</sub>. As shown in Fig. 1b, at low Ca(NO<sub>3</sub>)<sub>2</sub> concentrations, the CuO NPs are more stable than in the presence of alginate, which can be explained by the steric influence imposed by the adsorbed alginate on the CuO NPs. It is widely reported that the stabilization of many particles was attributed to steric effects in the presence of alginate (Huangfu et al. 2013; Loosli et al. 2015). In addition, it is noteworthy that the adsorption quantity of alginate onto CuO NPs in the presence of Ca(NO<sub>3</sub>)<sub>2</sub> was much higher than that in NaNO<sub>3</sub> solution (Figure S6); however, Ca(NO<sub>3</sub>)<sub>2</sub> showed a higher efficiency in decreasing zeta potential than the NaNO<sub>3</sub> at the same molarity in the presence of alginate in this study (Fig. 1a). Although the adsorbed alginate would impart a negative charge onto the surface of CuO NPs, the interactions between Ca<sup>2+</sup> cations and negative functional groups would impede the increase of zeta potential (Shen et al. 2015).

Nevertheless, as the Ca<sup>2+</sup> concentration was increased, the co-presence of alginate and high Ca<sup>2+</sup> concentration (>6 mM) resulted in significant increase in hydrodynamic diameter of CuO NPs (as shown in Fig. 2). Furthermore, there was a corresponding increase in the attachment efficiency to values above unity (Fig. 1b), indicating that the steric effects were not



**Fig. 2** Aggregation profiles of CuO NPs in various concentrations of  $\text{Ca}(\text{NO}_3)_2$  solutions in the absence and presence of 5 mg/L C alginate at pH 6.8 and 25 °C

the predominant for the CuO NPs at high  $\text{Ca}^{2+}$  concentration (>6 mM) in the presence of alginate in this study. These observations strongly indicated that at high  $\text{Ca}^{2+}$  concentration, different aggregation mechanisms may be involved in the presence of alginate, which results in the enhanced aggregation kinetics, which will be studied next.

#### Alginate bridging leads to enhanced aggregation

TEM images of nano-CuO aggregates in the presence of  $\text{Ca}^{2+}$  and the co-presence of  $\text{Ca}^{2+}$  and alginate were captured to compare the morphology and to delineate the aggregation mechanisms (Shen et al. 2015). As shown in Fig. 3, in the absence of alginate, aggregates of CuO NPs can be observed where the particles are in close proximity to each other at 10 mM  $\text{Ca}^{2+}$ , at approximately 1 h after the initiation of nano-CuO aggregation. The aggregation is probably due to the decreasing surface charge of CuO NPs caused by the addition of  $\text{Ca}(\text{NO}_3)_2$ . However, in the presence of  $\text{Ca}^{2+}$  (10 mM) and alginate (5 mg/L C), it appears that CuO nanoparticles were embedded on the gel-bead structures formed by the clusters of alginate, and condensed shades of  $\text{Ca}^{2+}$ /alginate

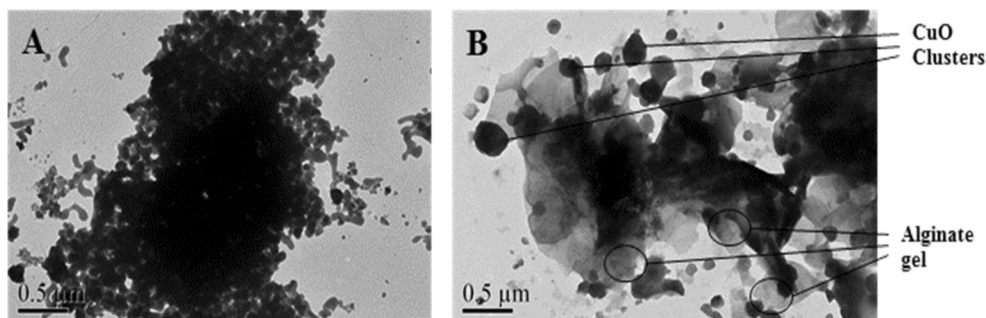
aggregates as bridges among nano-CuO particles in the TEM images were observed.

In addition, the bridging mechanism was further evidenced by elemental mapping obtained from SEM-EDS analysis. In copper mapping, the copper element positions were well consistent with the SEM image, with either absence or presence of alginate (Fig. 4). However, the presence of alginate resulted in an obvious decrease of the correspondence of the calcium elements with the nano-CuO aggregate positions when compared with that of aggregate in the absence of alginate (Fig. 4). This suggested that different mechanisms are predominant in the absence and presence of alginate. In the absence of alginate, CuO nanoparticles formed the aggregates via  $\text{Ca}(\text{NO}_3)_2$  adsorption and the charge screening; thus, the mapping of calcium was in accordance with the area of the nano-CuO aggregate. In contrast, there was a wider distribution of the calcium element than the copper element in the presence of alginate, suggesting the  $\text{Ca}^{2+}$ /alginate complex adsorbed on the surface of CuO NPs, which confirmed the bridging mechanism for CuO nanoparticles. In addition, calcium is known to bridge the molecules in the alginate network ( $\beta$ -D-mannuronate and  $\alpha$ -L-guluronate) to form gel-like aggregates (Gallego-Urrea et al. 2014), which is also possible to see in the EDX mapping (Fig. 4), and will be discussed next.

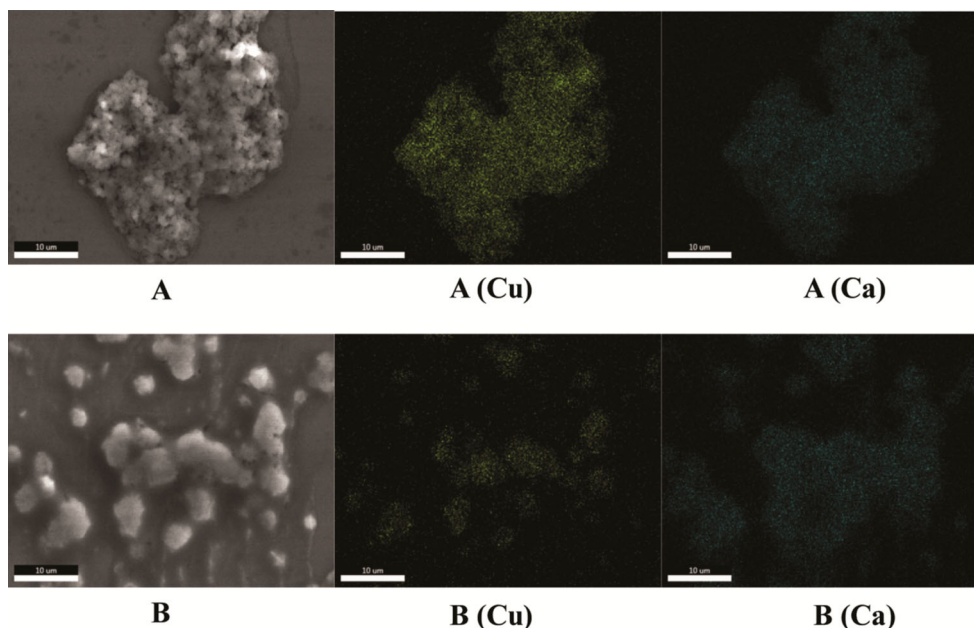
#### Hetero-coagulation mechanism between CuO NPs and the unadsorbed alginate

Besides, as calculated from the adsorption tests, only about 20–35 % of alginate could be adsorbed onto 100 mg/L CuO NPs (Figure S6), leaving the majority of alginate in both the electrolyte solutions in this study. According to the literature,  $\text{Ca}^{2+}$  has the ability of producing aggregation of alginate molecules above a certain cation concentration due to the complexation of the carboxylic groups (Kloster and Avena 2015). Thus, the above quantitative comparison of the amounts of adsorbed alginate and the unadsorbed alginate allows us to hypothesize a hetero-coagulation mechanism between the alginate-covered CuO NPs and the unadsorbed alginate under high concentration of  $\text{Ca}(\text{NO}_3)_2$  in this study.

**Fig. 3** Representative TEM images of CuO NPs in the presence of **a**  $\text{Ca}^{2+}$  and **b**  $\text{Ca}^{2+}$  and alginate ( $\text{Ca}^{2+}$  = 10 mM; alginate = 5 mg/L C). TEM images were captured from samples collected 1 h after the initiation of aggregation



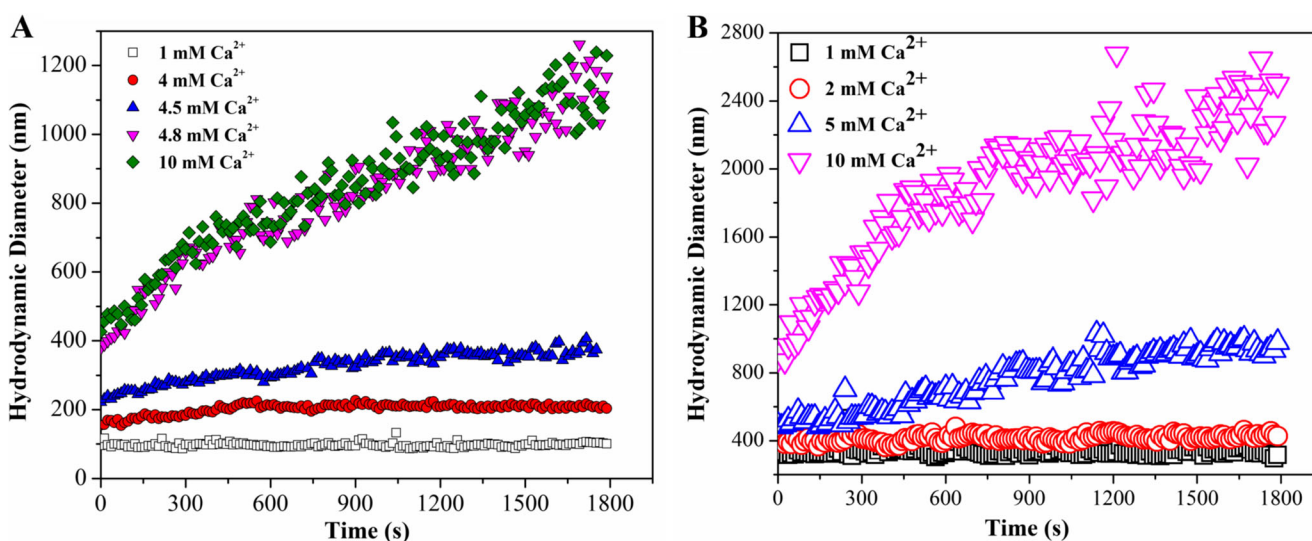
**Fig. 4** Representative SEM-EDS images with element mapping of CuO NPs in the presence of **a**  $\text{Ca}^{2+}$  and **b**  $\text{Ca}^{2+}$  and alginate ( $\text{Ca}^{2+} = 10 \text{ mM}$ ; alginate = 5 mg/L C). SEM images were captured from samples collected 1 h after the initiation of aggregation



To further investigate the role of  $\text{Ca}^{2+}$ -induced alginate aggregation in the  $\text{Ca}(\text{NO}_3)_2$  systems, the scattered light intensities from separately prepared alginate solutions with different electrolytes were measured with respect to time by employing the light scattering setup following the method described by Chen and Elimelech (2007). The alginate solutions were prepared at the same concentration as the DLS experiments of 5 mg/L TOC at pH 6.8, and the aggregation experiments of alginate were conducted following the procedures described in section “Aggregation kinetics of CuO NPs.”

Figure S7 shows the scattered light intensities from the alginate solutions normalized by the incident laser intensity in the presence of the  $\text{NaNO}_3$  and  $\text{Ca}(\text{NO}_3)_2$  electrolytes. At

high  $\text{Na}^+$  concentrations (200 mM), the normalized scattered light intensities remained constant at slightly higher values than that of water, suggesting that the relatively dilute alginate solutions did not form aggregates. In contrast, in the presence of 10 mM  $\text{Ca}^{2+}$ , the normalized scattered light intensity increases with time, indicating significant aggregation of alginate macromolecules. Then, the concentration of alginate was increased to 25 mg TOC, in order to observe the aggregation of alginate in the  $\text{NaNO}_3$  and  $\text{Ca}(\text{NO}_3)_2$  electrolytes. Over the entire  $\text{NaNO}_3$  concentration range,  $\langle D_h \rangle$  remained about 100 nm, indicating the stability of alginate colloids (data not shown). In contrast, as the  $\text{Ca}^{2+}$  concentration increased from 1 to 4.5 mM, the  $\langle D_h \rangle$  exhibited a slightly increase (below 400 nm), while a rapid rise of  $\langle D_h \rangle$  was observed when the



**Fig. 5** **a** Aggregation profiles of alginate (25 mg/L C) in various concentrations of  $\text{Ca}(\text{NO}_3)_2$  solutions at pH 6.8 and 25 °C. **b** Aggregation profiles of CuO NPs in the presence of alginate (25 mg/L C) in various concentrations of  $\text{Ca}(\text{NO}_3)_2$  solutions at pH 6.8 and 25 °C



$\text{Ca}(\text{NO}_3)_2$  concentration exceeded 4.8 mM (Fig. 5a). Bound divalent cations could form intra- and/or intermolecular bridges between adjacent alginate colloids, and as a result increase the attractive intermolecular forces (Wang et al. 2013). Thus, divalent  $\text{Ca}^{2+}$  is considered to rapidly interact with alginate molecules, and the combined effects of charge screening, calcium complexation as well as bridging lead to the rapid formation and growth in the size of the alginate aggregates.

Based on the above observations and discussion, another experimental group was conducted, in which the aggregation kinetics of CuO NPs (100 mg/L) was determined with the presence of alginate (25 mg/L TOC) in various concentrations of  $\text{Ca}(\text{NO}_3)_2$  solutions at pH 6.8 and 25 °C. The measured  $\langle D_h \rangle$  of the aggregates in the solution increased significantly with the increasing of  $\text{Ca}^{2+}$  concentrations (Fig. 5b), consistent with the results obtained from the alginate aggregation. The results may indicate hetero-aggregation of alginate molecules and CuO NPs in solution of high concentration of  $\text{Ca}^{2+}$ . Although it is very difficult to discriminate between  $\text{Ca}^{2+}$ -induced aggregation of alginate molecules and direct aggregation of nano-CuO/alginate complex, it is possible to infer that hetero-aggregation would be the prevailing process when the NPs were released into the aqueous systems enriched with NOM. Thus, further studies should be conducted to investigate the NP stability under more complex and environmentally relevant conditions in the future.

## Conclusion and implications

This study provides an insight into the stability of CuO nanoparticles in various ionic strength conditions with and without the presence of alginate. It appears that, in the absence of alginate, significant aggregation of CuO NPs was observed in the two electrolyte solutions ( $\text{NaNO}_3$  and  $\text{Ca}(\text{NO}_3)_2$ ), especially for  $\text{Ca}(\text{NO}_3)_2$ . The presence of alginate had pronounced effects on the particle size distribution of CuO NPs and greatly enhanced the aggregation of CuO NPs in high concentration of  $\text{Ca}^{2+}$  solutions, confirmed by DLS measurements. The morphology of the aggregates of CuO NPs formed under different solutions was observed by TEM analysis. Besides, significant coagulation of the alginate molecules was observed in solutions of high  $\text{Ca}^{2+}$  concentrations, suggesting a hetero-coagulation mechanism between the alginate-covered CuO NPs and the unabsorbed alginate under high concentration of  $\text{Ca}(\text{NO}_3)_2$ . The results presented in this study suggested that not only the presence of NOM but also the ratio between NP and NOM should be taken into consideration in studying the NP stability in aqueous systems, due to the interaction between the multivalent cations and the ubiquitous organic matter (Kloster et al. 2013; Wang et al. 2013). More studies should be conducted to extend to more complex and environmentally relevant conditions in the future. Thus, specific

conditions in aquatic environments play a very important role in evaluating the fate and transport of engineered nanoparticles, which ultimately have impacts on uptake and therefore bioavailability and toxicity of ENPs to aquatic organisms.

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