MINING AND THE ENVIRONMENT - UNDERSTANDING PROCESSES, ASSESSING IMPACTS AND DEVELOPING REMEDIATION

# Biogeochemical behaviour and bioremediation of uranium in waters of abandoned mines

Martin Mkandawire

Received: 13 October 2012 /Accepted: 9 January 2013 / Published online: 26 January 2013  $\oslash$  Springer-Verlag Berlin Heidelberg 2013

Abstract The discharges of uranium and associated radionuclides as well as heavy metals and metalloids from waste and tailing dumps in abandoned uranium mining and processing sites pose contamination risks to surface and groundwater. Although many more are being planned for nuclear energy purposes, most of the abandoned uranium mines are a legacy of uranium production that fuelled arms race during the cold war of the last century. Since the end of cold war, there have been efforts to rehabilitate the mining sites, initially, using classical remediation techniques based on high chemical and civil engineering. Recently, bioremediation technology has been sought as alternatives to the classical approach due to reasons, which include: (a) high demand of sites requiring remediation; (b) the economic implication of running and maintaining the facilities due to high energy and work force demand; and (c) the pattern and characteristics of contaminant discharges in most of the former uranium mining and processing sites prevents the use of classical methods. This review discusses risks of uranium contamination from abandoned uranium mines from the biogeochemical point of view and the potential and limitation of uranium bioremediation technique as alternative to classical approach in abandoned uranium mining and processing sites.

Responsible editor: Stuart Simpson

M. Mkandawire  $(\boxtimes)$ 

Verschuren Centre for Sustainability in Energy and the Environment, Cape Breton University, P.O. Box 5300, 1250 Grand Lake Road, Sydney, Nova Scotia, Canada B1P 6L2 e-mail: martin\_mkandawire@cbu.ca

Keywords Ecotoxicology . Tailings . Groundwater . Leaching .Bioreduction .Radionuclides . Uranium minerals

# Introduction

Uranium (U) is not only important because of being the heaviest element occurring in nature in weighable amounts and nuclear fuel but also due to its chemical and radioactive implications to human health and environment. Exposures to environmental U are associated with health effects, such as kidney or liver damage, cancer or birth defects (Schuttmann [1993;](#page-25-0) Sram et al. [1993;](#page-26-0) WHO [2001\)](#page-27-0). Furthermore, U disrupts the endocrine system such that populations exposed to environmental U have increased risk of fertility problems and reproductive cancers. U is one of the most mobile radioactive metals that it are easily washed down through soil together with percolating water to both surface and underlying groundwater. Thus, water pathway is the major exposure route to U, although dust in the air may act as a pathway too. Its geochemical properties especially the chemical form strongly influences the environmental transport. U exists predominantly as hexavalent oxidation state (U(VI)) which forms readily soluble species over a range of pH and redox potential  $(E_H)$ conditions in the aquatic environments. Furthermore, U(VI) ions are exclusively available as species of uranyl ion  $(UO_2^{2+})$ , which sometimes behave like divalent metal ions (Mkandawire et al. [2006a](#page-24-0)).

Small amounts of U are found almost everywhere in soil, rock and water. As a result, it is normal to find dissolved U at very low concentrations in most natural waters, but concern rise only when concentrations increase to levels above  $1 \mu g L^{-1}$  (Kalin et al. [2005a](#page-23-0)). Thus, trace contamination occur naturally, but most of alarming U contaminations are anthropogenic associated with activities of the nuclear fuel cycle, phosphate fertilizer production process and the

Prof. Dr. Martin Mkandawire was Privatdozent (associate professor) in Institute of Materials Science in Technische Universität Dresden, Germany until September 2012

improper disposal of U mine tailings (Jerden and Sinha [2006;](#page-23-0) Chen et al. [2005\)](#page-21-0). The cycle for nuclear fuel, include the mining, milling and processing of U ores or U products. In the production of phosphate fertilizers, the phosphorus is usually extracted from phosphate rocks containing U (Shumilin et al. [2012;](#page-26-0) Rodgher et al. [2012;](#page-25-0) Giri et al. [2010;](#page-22-0) Arogunjo et al. [2009;](#page-20-0) Vanhoudt et al. [2008\)](#page-26-0). U may also be released from fly ash or stack emissions during coal combustion and from the military use of 235U-depleted in warheads (WHO [2001](#page-27-0); Giannardi and Dominici [2003](#page-22-0)). However, major contamination concerns currently stems from U waste and tailing dumps in abandoned sites of ore mining and processing (Carvalho et al[.2005](#page-21-0); Mkandawire [2005\)](#page-24-0). In most abandoned U mining sites, the discharge of U contamination is also accompanied by other toxic heavy metals and metalloids like Cd, Pb, Ra and As (Al Hashimi et al. [1996;](#page-20-0) Trontelj and Ponikvar-Zorko [1998](#page-26-0)).

There are considerable efforts to reclaim most of abandoned and decommissioned U mining sites. Frequently, classical techniques based on civil and chemical engineering strategies are employed in the restoration efforts. The strategies include: covering of tailing and waste damps with either chemical neutralising or physical stabilising material, or even both, and, treatment of contaminated waters using chemical and electrochemical procedures (Kalin et al. [2005a](#page-23-0)). These classical remediation techniques are said to be too expensive to meet the remediation demand due to the number of sites requiring restorations, and the technologies are beyond capacity of most poor nations (Benders et al. [2000;](#page-21-0) Salt et al. [1995;](#page-25-0) Schnoor et al. [1995](#page-25-0); Adler [1996](#page-20-0)). Additionally, it is claimed that the contamination problems are merely suspended or transferred to elsewhere (Mkandawire et al. [2004b](#page-24-0)). Furthermore, the discharges of the contaminants from the tailing and waste heaps into the water pathway are mostly non-point source. Moreover, the patterns of discharge of contaminant loads from mine wastes and tailings are usually unpredictable where they are either (a) high volume and concentrated but a short-term, (b) very low concentrated but long-term or (c) low concentrated but short-term discharge. These criticisms have pointed to the need for alternative remediation technology that would be cheap, easy to run, responsive with long-term effect preferably where natural processes are used. Bioremediation fulfils most of these conditions, and it is currently advocated as a promising alternative technology with potential application in abandoned U mining sites. Therefore, the aim of this review is to relate knowledge of U biogeochemistry with opportunities and challenged in application of bioremediation in abandoned U mines. The scope of this review is limited to discussion on toxicology as well as potential and limitation of bioremediation of natural U in abandoned U mines with special consideration to its biogeochemical behaviours in water.

#### Abandoned U mining site

Not every decommissioned U mine is an abandoned mine. However, only sites where advanced exploration, mining or mines production ceased without comprehensive rehabilitation or restoration are classified as abandoned mines, also called orphaned mines (Hoffmann [2000;](#page-23-0) Mkandawire [2005;](#page-24-0) Mkandawire et al. [2004a\)](#page-24-0). All U abandoned mining sites share identical environmental problems ranging from disturbed physical landscapes to large piles of rocks, mining wastes and tailing to environmental contamination with radionuclides and heavy metals (Diehl [2003;](#page-21-0) Diehl [1995\)](#page-21-0). Consequently, the overlying characteristics of abandoned U mines are public health and safety, environmental safety and aesthetic concerns. However, the mining and processing procedure determines the characteristics of an abandoned mining site.

# Ore mining and processing

#### Ore extraction

There are several techniques used to extract U from its ore deposits but frequently used are open-pit, underground, in situ leaching  $(ISL)^1$  and borehole mining techniques. The open-pit mining of U was predominant until the 1960s (Pettersson and Koperski [1991](#page-25-0); Abdelouas [2006](#page-20-0)). It was used to extract U from ore deposits located near the surface while deposits located deeper are extracted in underground mining. In both open pits and underground mines, traditional mining techniques of drilling and blasting are used to excavating mineral bearing rock. The ISL mining involves injecting either acid or alkaline solutions into the ground to dissolve the mineral ores, and then, pumping the pregnant solution to the treatment plant to recover the U. Alkaline leaching, using a combination of sodium bicarbonate and carbon dioxide, is suitable for ore bodies containing significant calcium (Ca) like limestone or gypsum (Erdem et al. [1995;](#page-22-0) Beddow et al. [2006](#page-21-0); Misaelides et al. [1995](#page-24-0)). Otherwise, acid leaching using sulphuric acid enriched with oxygen is generally ideal. The ISL technique is ideal in permeable ore bodies especially in secondary mineral deposits (e.g. uraninite or coffinite silicate) that occur in permeable sand or sandstones below the water table (Ulrich et al. [2006](#page-26-0)). Lastly, the borehole mining of U is the least frequently used technology although it is potentially applicable in both surface and underground mining. The procedure involves injection of water at high pressure

<sup>&</sup>lt;sup>1</sup> ISL technology was developed independently in both the USSR and USA in the mid-1970s. The Soviets adopted the acid leaching system, while alkaline, primarily carbonate-based leaching system was widely used in the USA. In the USA, the technology is also known as in situ recovery.

through boreholes into the ground. The pressure breaks up the ore-bearing rocks, which results into formation of slurry composed of dirt, rock and minerals. The slurry is pumped to the surface where it is dewatered to recover the U (Salbu [2013](#page-25-0)); Thompson [2011;](#page-26-0) Boice et al. [2010;](#page-21-0) SENES and Alberta Environment [2008](#page-25-0)).

# U processing

The process of extracting U from ore is summarised in Fig. 1. For open-pit and underground mining, the first step is to crush the ore-bearing rocks into a fine powder. Then, huge amounts of water, leaching agents and thickener are added to the pulverised ore to oxidise and dissolve the U minerals. The U minerals usually available as  $UO<sub>2</sub>$  are oxidised to  $UO<sub>3</sub>$  followed by formation of either a uranyl sulphate  $(UO<sub>2</sub>(SO<sub>4</sub>)<sub>3</sub><sup>4</sup>^-)$ or a uranyl carbonate (predominantly  $UO_2(CO_3)_{3}^{4-}$ ) in acid leaching or carbonate leaching, respectively. Obviously, there is no crushing in the ISL and borehole mining procedure. After the leaching step, the U is concentrated and purified using either ion exchange or solvent extraction processes. Solvent extraction is the most applied procedure, which uses tertiary amines  $(R_3N)$  in kerosene diluents (El-Nadi et al. [2003\)](#page-22-0). The amines react with sulphuric acid, which removes



Fig. 1 The U extraction chemistry from ore mining to production of yellow cake

impurities from the loaded solvents. Cations are removed using sulphuric acid while anions are removed with gaseous ammonia (Juznic et al. [1989;](#page-23-0) Levy and Kearney [1999](#page-23-0)). The solvents are stripped with ammonium sulphate solution. Then, exposure to gaseous ammonia to neutralise the solution leads to the precipitation of ammonium diuranate. The diuranate is subsequently dewatered and dried to produce hydrated U peroxide ( $UO<sub>4</sub>·2H<sub>2</sub>O$ ). Lastly, centrifugation and precipitation of  $UO_4$ :  $2H_2O$  yields the yellowcake  $(U_3O_8)$  containing at least 75 % U oxides (Abdelouas [2006](#page-20-0); Moyes et al. [2000\)](#page-24-0).

Characteristics of abandoned U mines

## Mining wastes and tailing

Huge piles of mine waste and tailing are the first characteristics of abandoned open-pit and underground mines and U content of the ores is often between 0.1 and 0.7 % (Diehl [1995;](#page-21-0) Diehl [2003;](#page-21-0) Meinrath et al. [2003](#page-24-0)). Thus, large amounts of rocks are excavated and large amounts of tailings are generated when extracting the U. A mine can generate up to 40 tonnes of waste rock for every tonne of U ore produced, while underground mines produce about 1 tonne of waste rock/tonne of ore (Morgenstein [2003;](#page-24-0) Mkandawire [2005;](#page-24-0) Tomasek et al. [1993;](#page-26-0) Enderle and Friedrich [1995\)](#page-22-0). Mountains of wastes and debris as well as lakes resulting from flooding of open pits disturb the landscape. Apart from creating undesired physical features, wastes, debris and tailings create massive stockpiles of radioactive and toxic waste rock and sand-like tailings, which become a source of radioactive, toxic metals and radioactive air pollutants. With most of the U removed, the mine wastes and tailings remain very radioactive because almost 85 % of the radioactive elements and U decay progenies contained in the original U ore end up in the tailings and wastes (Mkandawire [2005;](#page-24-0) Enderle and Friedrich [1995\)](#page-22-0).

# Mine water quality

Water contamination is a common characteristic of abandoned U mining sites. The tailings and wastes from the U milling process consist of ground rock particles, non-extracted radionuclides, toxic metals, metalloids, remnants of U as well as leach and mill chemicals. As a result, chemically hazardous substances in the wastes and tailings like lead, cadmium, arsenic and mercury are discharged more easily into the environment through leaching and blown dust into the water and air pathway (Al Hashimi et al. [1996;](#page-20-0) Pressyanov et al. [1995\)](#page-25-0). The excavated and ground rock particles have high surface area that increased their contact with air and water, which naturally favours geochemical process especial redox and acidification. Hence, precipitation leaches out radionuclide (i.e. progeny of U and U debris), heavy metals and metalloids, which may contaminate surface and groundwater. Depending on the type of rock, these wastes may be acid generating. For instance, surface mine water discharged from a few abandoned mining sites in eastern Germany (which the author has studied extensively) have pH values between 2.7 to 3.4. The concentrations of dissolved arsenic ranges from 0.8 to 700 mgL<sup>-1</sup>, iron (Fe) from 750 to 3,000 mgL<sup>-1</sup> and sulphate from 2,000 to 8,000 mgL<sup>-1</sup>. However, 20 to 60 % of the dissolved arsenic is immobilised by co-precipitation with Fe (III) (Clara and Magalhães [2002;](#page-21-0) Fritzsche et al. [2006;](#page-22-0) Munoz et al. [2002\)](#page-25-0). Nevertheless, this results in high groundwater concentrations of arsenic and enrichment into the sediments, which can reach as high as 500 % more than the background (Carvalho et al. [2005](#page-21-0); Belzile and Tessier [1990\)](#page-21-0).

## Radon gas emanation and air quality

Another characteristic of abandoned U mining is air pollution, which is manifested either through odours from the waters, dust or both. However, the most dangerous air pollution associated with abandoned U mining is odourless—radon  $(^{222}Rn)$ emanation. The original source of  $222$ Rn gas is from the natural radioactive decay of <sup>238</sup>U to its daughter element radium  $(^{226}$ Ra). U-rich mineral rocks like granite, phosphate, shale and pitchblende store natural high deposits of  $^{222}$ Rn. Therefore, the piles of rock and tailings in abandoned U mines continuously release Rn to the environment. Apart from being significant sources of Ra gas, winds may also blow dust containing radionuclides, heavy metals and particulate matter from the rock debris and tailing piles. Furthermore, waste rock and tailings damps of abandoned U mines release nitrogen oxides, volatile organic compounds and carbon dioxide  $(CO<sub>2</sub>)$ . Acid plants producing acid for milling operations release large amounts of sulphur dioxide—a major contributor to acid rain (Hole et al. [2008](#page-23-0); Fowler et al. [2005;](#page-22-0) Jeffries et al. [2003\)](#page-23-0).

# Acid mine drainage

The generation of acid mine drainage (AMD) is a common in most abandoned U mining and processing sites because most U ore are associated with pyrite minerals  $(FeS<sub>2</sub>)$ . Thus, pyrite minerals are abundant in the mine waste rock (Carvalho et al. [2005\)](#page-21-0). Exposure to water and air breaks down the pyrite-rich waste rocks and tailings leading to the formation of sulphuric acid  $(H_2SO_4)$  and  $FeSO<sub>4</sub>$ . Then, the  $FeSO<sub>4</sub>$  undergoes further reactions, which eventually end up in formation of ferrous hydroxide (Fe(OH)<sub>3</sub>). The Fe(OH)<sub>3</sub> is responsible for reddish or orange colour in the seepage water through mine waste and tailings, which is a symbol that acid drainage is taking place. The whole breakdown process can be summaries in following chemical reactions:

$$
2FeS_2 + 7O_2 + 2H_2O \rightarrow 2FeSO_4 + 2H_2SO_4 \tag{1}
$$

$$
2Fe^{2+} + \frac{1}{2}O_2 + 2H + \rightarrow 2Fe^{3+} + H_2O
$$
 (2)

$$
Fe^{3+} + 3H_2O \rightarrow Fe(OH)_3 + 3H^+ \tag{3}
$$

$$
FeS2(s) + \frac{15}{4}O2 + \frac{7}{2}H2O \rightarrow 4H+ + 2SO42- + Fe(OH)3(s)
$$
\n(4)

Logically, the AMD affects the acidity as well as pH of the receiving waters due to the high Fe and H ion they contain. However, the pH measurements may not detect heavy AMD in some streams with high dissolved carbonates because pH only indicates the concentration of H ions. AMD depletes the buffering ability of water by neutralizing carbonate and bicarbonate ions to form carbonic acid  $(H<sub>2</sub>CO<sub>3</sub>)$ , which readily breaks down into water and carbon dioxide (see reactions 5 and 6).

$$
\mathbf{H}^+ + \mathbf{CO_3}^{2-} \Leftrightarrow \mathbf{HCO_3}^- + \mathbf{H}^+ \Leftrightarrow \mathbf{H_2CO_3} \tag{5}
$$

$$
H_2CO_3 \rightarrow H_2O + CO_2 \tag{6}
$$

## Aquifer contamination from in situ leaching

Much as there is also little surface disturbance and no generation of waste rocks and tailing, the abandoned ISL mines have serious groundwater contaminations that are difficult to remediate. During ISL mining, bacterial contamination are common that sometimes facilitate in the leaching process. When the mines are abandoned, the bacterial activities continue to affect the leaching of minerals into the aquifers. Accompanied by prevailing unsaturated conditions, the pyrite oxidise generating significant quantities of sulphuric acid, which further mobilises heavy metals, U and radionuclides trapped in the pore spaces of the rocks thus adding to the contaminant load in the groundwater. The trapped liquids are characterised by elevated amount of heavy metals and metalloids which can reach between 100 and 400 % higher in Cd, As, Ni and U than before the ISL begins (IAEA [2001](#page-23-0); Carvalho et al. [2005\)](#page-21-0). In acid ISL mining, an important characteristic of using sulphuric acid in leaching is the deterioration of the ore zone permeability due to chemical and gaseous plugging and rising of salinity levels. In the alkaline ISL, the large quantities of solutions escape beyond the mining zone due to gaseous oxygen plugs. However, the alkaline agents tend to increase the salinity of the groundwater but only slightly. Nevertheless, the contamination potential of surrounding waters with U, Ra, SO4, Fe and other heavy metals remains high.

# Geochemistry

# General properties

# Physicochemical

U is the heaviest naturally occurring element, with an atomic number of 92. Despite its high atomic number, it is an ubiquitous element with an average concentration of 3 mgkg<sup>-1</sup> in the Earth's crust (IAEA [1998\)](#page-23-0). The concentrations of U in geological materials are highest in continental-type rocks. Pure U is a silvery white, ductile and malleable heavy metal, which melts at 1,132.3 °C and boils at about 3,818 °C. Its density is about 19.05  $gcm^{-1}$  (Kleykamp [1991\)](#page-23-0). However, elemental U hardly exists in nature, because U metal is readily subjected to surface oxidation (Beddow et al. [2006](#page-21-0); Whitehead et al. [1971](#page-27-0)). Consequently, it ignites spontaneously upon contact with air at ambient temperature forming U oxides with six potential oxidation states of U. However, U(IV) and U(VI) are the most common oxidation states of U in nature because U(III) easily oxidises to U(IV) under most redox conditions while U(V) readily disproportionates to U(IV) and U(VI) in nature (Lovley et al. [1993;](#page-23-0) Benders et al. [2000](#page-21-0); Markich [2002\)](#page-24-0). U forms more than 234 mineral species and accounts for only 5 % of all known minerals (Barthelmy [2008](#page-20-0)). The most common forms of U oxide pitchblende  $(U_3O_8)$  and uraninite  $(U_2O_8)$ , have low solubility in water and are relatively stable over a wide range of environmental conditions. The  $U_3O_8$  exist mostly as  $U_2O_5 UO_3$  (Wersin et al. [1994](#page-26-0)). Generally, U forms many compounds that tend to have yellowish or greenish colours. In respect to this chemical property, U has been historically used as a colorant for glass and ceramic (i.e. producing orange-red to lemon yellow hues) and for tinting and shading in early photography.

# Radioactivity

U has sixteen radioactive isotopes, but only three isotopes  $-$ <sup>238</sup>U, <sup>235</sup>U and <sup>234</sup>U—occur naturally (Jerden and Sinha [2006\)](#page-23-0). Figure [2a, b](#page-5-0) summarises the decay series of  $^{238}$ U and <sup>235</sup>U detailing the type of radiation given off at each step and the half-life of each step gone through to ultra-stable and non-radioactive  $^{206}Pb$  or  $^{207}Pb$  isotopes. The U isotopes emit mostly alpha and less beta particles accompanied by little emission of gamma radiation especially during the decay of 235U (Cherdyntsev et al. [1982;](#page-21-0) Ivanovich and Harmon [1992\)](#page-23-0). The emissions are all ionising radiation, strong enough to break chemical bonds, thereby possessing the ability to damage or destroy living cells (ATSDR [1999](#page-20-0); WHO [2001,](#page-27-0) [2004](#page-27-0)). In terms of the amount of radioactivity in nature, 235U contributes about 2.2 %, 238U about 48.6 % and 234U about 49.2 %. The total activity level of U depends

on the isotopic composition ratio. The  $^{235}$ U and  $^{234}$ U isotopes pose a greater radiological toxicity risk than  $^{238}$ U because they have much shorter half-lives.

# U classifications

In nature, the U isotopes are distributed as 99.284 % of <sup>238</sup>U, 0.72 % of <sup>235</sup>U and a very small amount (~0.0054 %) of 234U. Thus, U with this isotope distribution is called natural U. The  $^{235}$ U has high fission capacity; consequently,  $235$ U is used as fuel of nuclear reactors. However, a nuclear reactor requires higher than 0.72  $\%$  content of <sup>235</sup>U to function. Thus, the natural U is centrifuged to enrich the content of  $^{235}$ U from 0.72 % to about 1.5–3.0 % (IAEA [1999](#page-23-0), [2007](#page-23-0)). Then, the <sup>234</sup>U and <sup>235</sup>U isotopes are extracted to produce high radioactive enriched U, which is primarily used as fuel in nuclear reactors. Consequently, the remaining U after removal of the enriched fraction is called depleted U and it contains at least three times less  $^{234}$ U and  $^{235}$ U by mass than before commencement of enrichment (Table [1\)](#page-5-0). The depleted U is weakly radioactive and a radiation dose from it is about 60 % of natural U with the same mass (WHO [2001](#page-27-0); Meinrath et al. [2003\)](#page-24-0). Natural and depleted U differ only in their radioactivity but their chemical properties are the same.

## Aquatic chemistry of U

#### Behaviour in surface mine waters

The aquatic chemistry of U is governed by a linear dioxocation  $UO_2^{2+}$  ([O=U=O]<sup>2+</sup>) (Meinrath [1996;](#page-24-0) Meinrath [1999](#page-24-0)), which is mainly controlled by the pH, the  $E_{\rm H}$  and the type of the available complexing ligand such as carbonates, phosphates, vanadates, fluorides, sulphates and silicates (Bernhard et al. [1998](#page-21-0); Duff et al. [2002;](#page-22-0) Martinez Aguirre et al. [1995\)](#page-24-0). The U(III) is very easily oxidised whilst the U(V) is prone to disproportionation (Steele and Taylor [2007;](#page-26-0) Shilov et al. [2007](#page-26-0)). In oxidised subsurface waters with  $E_H$ >200 mV, dissolved  $UO_2^{2+}$  form carbonate complexes of varying stoichiometry as a function of the pH and the partial pressure of  $CO<sub>2</sub>$  (g) (Meinrath [1996](#page-24-0)). The uranyl carbonate species prevail in the neutral to alkaline range, while the  $UO_2^{2+}$  hydrolyses in low pH values. Most of the hydrolysis species are oligomeric nature. Thus, oligomeric hydrolysis  $(UO_2)_2(OH)_2^{2+}$  species are usually the most dominants. These are followed by other oligomeric and monomeric hydrolysis species, which include  $[(UO<sub>2</sub>)<sub>3</sub>(OH)<sub>5</sub>]<sup>+</sup>, [(UO<sub>2</sub>)<sub>3</sub>(OH)<sub>4</sub>]<sup>2+</sup>, [(UO<sub>2</sub>)<sub>2</sub>OH<sub>3</sub>]<sup>+</sup>,$  $[(UO<sub>2</sub>)<sub>3</sub>(OH)<sub>7</sub>]<sup>-</sup>, (UO<sub>2</sub>)<sub>4</sub>(OH)<sub>7</sub><sup>+</sup>, UO<sub>2</sub>OH<sup>+</sup>, UO<sub>2</sub>(OH)<sub>2</sub>,$  $UO_2(OH)_3$ <sup>-</sup> and  $UO_2(OH)_4^{2-}$  (Bernhard et al. [1998;](#page-21-0) Brendler et al. [1998](#page-21-0); Markich [2002](#page-24-0); Meinrath [1997](#page-24-0); Ulrich et al. [2006](#page-26-0); Zanonato et al. [2004](#page-27-0)). The monomeric carbonate

<span id="page-5-0"></span>

Fig. 2 The decay series of a <sup>235</sup>U and b<sup>238</sup>U. The  $\alpha$  and  $\beta$  symbols indicate the alphas and beta decay, while the time indicated the half-life. The asterisk next to the isotope indicates the element that emit  $\gamma$  particles too. Modified from Peterson et al. ([2007\)](#page-25-0)

species include  $UO_2CO_3^0$ ,  $UO_2(CO_3)_2^{2-}$  and  $UO_2(CO_3)_3^{4-}$ while oligomeric carbonate compound include  $(UO<sub>2</sub>)<sub>3</sub>(CO<sub>3</sub>)<sub>6</sub><sup>6</sup>$ (Bernhard et al. [1998](#page-21-0); Meinrath [1998](#page-24-0); Meinrath [1996\)](#page-24-0). The distribution and occurrence of the uranyl species in water is summarised in Fig. [3](#page-6-0).

Under the reduced conditions with  $E_H$  below zero volts, the dissolved U(VI) species are reduced to the U(IV) and precipitate as  $UO<sub>2</sub> 2H<sub>2</sub>O$  (Meinrath [1999](#page-24-0); Luo et al. [2007](#page-24-0); Liu and Xu [2007\)](#page-23-0). This occurs mostly in sediments and very deep reservoirs. The U(IV) has a strong tendency to bind to organic material (Meinrath [1999](#page-24-0)). U in the mine water is derived

Table 1 Distribution of isotopes in natural and depleted uranium

| Isotope            | Fractions of the isotopes |                          |
|--------------------|---------------------------|--------------------------|
|                    | Natural uranium $(\%)$    | Depleted uranium $(\% )$ |
| $^{234}$ U         | 0.006                     | 0.001                    |
| $^{235}$ U         | 0.72                      | 0.20                     |
| $238$ <sup>T</sup> | 99.28                     | 99.80                    |

mainly from oxidation of uraniferous bitumen or dissolution of carbonates (Meinrath [1996](#page-24-0); Gavshin et al. [2005;](#page-22-0) Milodowski et al. [1990](#page-24-0)). Consequently, the dissolved U is associated with the concentrations of  $NO<sub>3</sub><sup>-</sup>, CO<sub>3</sub><sup>2</sup>$ ,  $SO<sub>4</sub><sup>2</sup>$ , Cl<sup>−</sup> and not silica (Petrescu and Bilal [2006\)](#page-25-0).

# U behaviour in oxidising aquifers

The chemistry of U in oxic aquifers is very much similar to the chemistry in the surface waters. Under oxidising conditions, U(VI) is the most stable form of U and it mostly exists as uranyl ion  $(UO_2^{2+})$ . It forms complexes readily with carbonate, phosphate or sulphate ions. In low carbonate-free systems, the important species include UO<sub>2</sub><sup>2+</sup>, UO<sub>2</sub>(OH)<sub>2</sub><sup>0</sup>(aq), UO<sub>2</sub>(OH)<sub>3</sub><sup>-</sup>, (UO<sub>2</sub>)<sub>3</sub>(OH)<sub>5</sub><sup>+</sup> and  $(UO<sub>2</sub>)<sub>2</sub>(OH)<sub>2</sub><sup>2+</sup>$  (Meinrath [1999;](#page-24-0) Meinrath [1997](#page-24-0)). In predominately carbonate-containing water, major uranyl species are carbonate anions with  $UO_2(CO_3)3^{4-}$  accounting for almost 60 %,  $UO_2(CO_3)_2^2$  for 30 % and  $(UO_2)_3(OH)_5^+$ (Meinrath [1999\)](#page-24-0). However, the real distributions of the species in relation to other U(VI) species are a function of

<span id="page-6-0"></span>Fig. 3 Distribution of U(VI) aqueous species at 25 °C and ionic strength  $\mu$ =0.01 M NaCl as a function of pH at atmospheric CO<sub>2</sub> partial pressure. Dashed lines give 90 % confidence limits. The speciation diagram has been modified and used with courtesy from Meinrath ([1999\)](#page-24-0)



pH and the total concentration of dissolved U(VI). Concentrations of dissolved U are controlled by the aqueous speciation of U(VI) and adsorption of U(VI) species onto mineral and organic material (Murakami et al. [1997](#page-25-0); Davis et al. [2004](#page-21-0); Ohnuki et al. [2004](#page-25-0)). The adsorption of U(VI) onto specific mineral phases primarily controls U mobility in oxic aquifers. The minerals prevalent in oxic conditions include carnotite  $[(K_2(UO_2)_2(VO_4)_2)]$ , schoepite  $(UO_3 \cdot$  $2H<sub>2</sub>O$ ), rutherfordine (UO<sub>2</sub>CO<sub>3</sub>), tyuyamunite [Ca  $(UO<sub>2</sub>)<sub>2</sub>(VO<sub>4</sub>)<sub>2</sub>$ ], autunite [Ca(UO<sub>2</sub>)<sub>2</sub>(PO4)<sub>2</sub>], potassium autunite  $[K_2 (UO_2)_2 (PO_4)_2]$  and uranophane [Ca  $(UO<sub>2</sub>)<sub>2</sub>(SiO<sub>3</sub>OH)<sub>2</sub>]$  (Allard et al. [1999;](#page-20-0) Jerden and Sinha [2006;](#page-23-0) Hiess et al. [2012](#page-23-0); Luo and Gu [2011;](#page-23-0) Stefaniak et al. [2009\)](#page-26-0). However, adsorption onto colloidal organic matter facilitates U transport and dispersal in the oxic aquifers. Furthermore, the solubility of U(VI)-containing minerals controls the maximum concentrations of dissolved U. In most abandoned U mines, the dissolution of U is dependent on the equilibrium between partially oxidised uraninite and the precipitation of coffinite (Cai et al. [2007\)](#page-21-0). U coprecipitates with Fe oxyhydroxides, which leads to the Fe oxyhydroxides taking up nearly all the U dissolved in the mine waters before they reach the surface (Gomez et al. [2006\)](#page-22-0).

# U behaviour in reducing aquifer

Under the reducing conditions of negative  $E_{\rm H}$  values, the dissolved U(VI) species are reduced to the less soluble U (IV) (Duff et al. [1999;](#page-22-0) Bots and Behrends [2008\)](#page-21-0). Consequently, there are sparingly soluble U(IV) species or mixed U(IV)–U(VI) solids (e.g.  $U_4O_9$ ) in reducing and anoxic conditions like in sediments and groundwater. The speciation of these sparingly dissolved U(IV) is dominated by hydrolytic species  $U(OH)_3^+$  and  $U(OH)_4^0(aq)$  in extremely low pH, especially of values below pH. The solid phases that control the U concentration in the groundwater are partially oxidised pitchblende. Other important U(IV) minerals associated with reducing conditions include coffinite (USiO<sub>4</sub>) and ningyoite (CaU(PO<sub>4</sub>)<sub>2</sub>⋅2H<sub>2</sub>O) (Jerden and Sinha [2006](#page-23-0)). There are usually accumulations of U(IV) precipitates in the reducing aquifers because the dissolved U(VI) species are in most cases, transported through the permeable where they get into reduced state. This is how the U roll-front deposits in sandstone rocks come about. These ore bodies occur when U(VI) carried by oxic groundwater contacts a zone containing a naturally occurring reductant such as natural organic matter and  $H_2S$  (g) (Duff et al. [1999;](#page-22-0) Gadelle et al. [2001](#page-22-0)). The resulting U(IV)-containing minerals include uraninite and coffinite.

## U in waste-rocks and tailings heaps

Generally, primary U ore in the waste rocks and tailing weathers into saprolites, which retain U as uranyl phosphates of the meta-autunite group mainly as metauranocircite in saturated zones (Jerden and Sinha [2006;](#page-23-0) Vinay Kumar Reddy et al. [2012](#page-26-0); Rodgher et al. [2012](#page-25-0)). The U is retained in the abandoned mine wastes and tailings containing primary ore by two processes:

- 1 Incorporation into barium–strontium–Ca aluminium phosphate minerals of the crandallite group (mainly gorceixite); and
- 2 Sorption of U with phosphorous onto Fe oxides that coat the surfaces of other soil minerals.

Thermodynamically, the meta-autunite group minerals are present in the saprolites in pH values below 4.5. Once exposed to the unsaturated environment, the metaautunite react to form U(VI)-bearing aluminium phosphates and U(VI) surface complexes or nano-precipitates associated with ferric oxides (Jerden and Sinha [2006;](#page-23-0) Shvareva and Albrecht-Schmitt [2006;](#page-26-0) Raicevic et al. [2006](#page-25-0)).

#### Hazard transfer from abandoned U mines

The prime environmental concerns in abandoned U mining and processing sites are the dissolution of minerals containing radioactive elements and heavy metals from waste-rock piles and tailings (Landa [2004](#page-23-0)). All chemical elements exist in conditions close to secular equilibrium in ores deposits, but the state of secular equilibrium is altered during the mining and processing of the ore material. The ore mining and processing lead to both physical and chemical equilibrium changes, which enhance the entry of radionuclides and the heavy metals hazards into human and environmental pollution pathways. The physical changes are responsible for the following major hazards transfers: (1) atmospheric dispersion of radioactive aerosol particles from mining waste and tailings heaps to adjacent sites by blowing winds; (2) increased erosion and seepage of water through loose waste-rock and tailing heaps, which may result in transport of contaminants by surface water; and (3) emanation of radon gas into the atmosphere.

The physical stability of the tailings and waste-rock heaps decreases over time due to internal natural processes, including physical and chemical digenesis and effects of root penetration from a vegetative cover (Riley [1995](#page-25-0); Roussel et al. [2000](#page-25-0)). Thus, the abandoned U mining and processing sites become prone to erosion, leaching as well as emanation of radioactive gases. Erosion occurrences can expose the underlying waste materials to either air or water dispersion. Seepage of water through the tailings pile may transport the radionuclides and other substances into groundwater. Seepage at near-surface groundwater can lead the contamination to re-enter the surface water bodies or air pathways (Biswas and Wu [1998;](#page-21-0) Sims et al. [1996\)](#page-26-0). The seepage to deeper groundwater may act as a long-term source of contamination. Extreme events such as earthquakes, floods and dam failures, though rare, may lead to a short-term but large discharge. Therefore, the physical changes facilitate alteration of chemical stability that is responsible for the transfer of both radionuclide and heavy metal hazards to environmental and human exposure pathways.

# Human health concerns

## Exposure routes

There are several pathways through which humans and the entire environment can be exposed to U toxicity hazard. These include inhalation of aerosol particles, dermal exposures and ingestion through contaminated water and foodstuffs. Inhalation, ingestion and external radiation are the most considered in human exposure risk. Although

absorption of some soluble compounds through the skin is possible, such dermal exposures generally are not significant. The inhalation of insoluble compounds such as U dioxide (UO<sub>2</sub>) and triuranium octaoxide (U<sub>3</sub>O<sub>8</sub>) lead to deposition in the lungs that can remain for long periods. In such conditions, inhaling aerosols with short-lived U progeny may lead to radiation toxicity as well as chemical toxicity of progeny heavy metals like Pb. For ingested U, insoluble compounds are poorly absorbed from the gastrointestinal tract and they are only retained in the body for a short time, thus generally having a low toxicity. However, ingested soluble U compounds enter the bloodstream and eventually reach the kidneys and other internal organs where chemical toxicity is of primary importance. About 98 % of U entering the body via ingestion is not absorbed (Hoffmann et al. [1993;](#page-23-0) Kusiak et al. [1993;](#page-23-0) Simon and Garnier-Laplace [2005](#page-26-0)). Typical gut absorption rates for U in food and water are about 2 % for soluble and about 0.2 % for insoluble U compounds (Veiga et al. [1998](#page-26-0)). The fraction of U absorbed into the blood is generally greater following inhalation than following ingestion of the same chemical form. The fraction will also depend on the particle size distribution. For some soluble forms, more than 20 % of the inhaled material could be absorbed into blood. Of the U absorbed into the blood, approximately 70 % is filtered by the kidney and excreted in the urine within 24 h. This amount may increase to 90 % within 48 to 96 h. On average, approximately 90 μg of U exists in the human body from normal intakes of water, food and air (Piao and Hei [1993;](#page-25-0) Morris and Meinhold [1995](#page-24-0)). The distribution of U in the human body is estimated to 66 % in the skeleton, 16 % in the liver, 8 % in the kidneys and 10 % in other tissues (Henge Napoli et al. [1995;](#page-23-0) Russell et al. [1996](#page-25-0)).

#### Chemical toxicity

U has been reported to cause kidney malfunction, respiratory disorders, DNA damage, mutagenicity, cancer and neurological defects (Stearns et al. [2005](#page-26-0); Mitrovic et al. [2005;](#page-24-0) Yazzie et al. [2003](#page-27-0)). Insoluble forms of U exhibit the least chemical toxicity risks because they are readily excreted via the digestive tract, while soluble forms of U interact with biological ligands and lead to toxicological effects. Once in the bloodstream, the kidneys filter the U compounds out of the body system. U ions, specifically uranyl cations are verified classic nephrotoxins (Dang et al. [1995](#page-21-0); Russell et al. [1996;](#page-25-0) Periyakaruppan et al. [2007](#page-25-0)). They bind to at least ten different proteins in the serum; its tissue distribution is affected by bicarbonate. Among the identified proteins are ceruloplasmin and hemopexin as binding uranyl with a 1:1 stoichiometry (Rybicka [1996;](#page-25-0) Miller et al. [1998](#page-24-0); Van Horn and Huang [2006](#page-26-0)). Generally, uranyl cation is partitioned in the serum into the uranyl bis- and tris-carbonate complexes

and  $UO<sub>2</sub>$ –protein–carbonate complexes with human serum albumin, transferrin and other protein. The uranyl–carbonate complexes are more diffusible into tissue especially the liver, kidneys and bones, while the portion bound to protein is removed from the serum in the kidneys (Dang et al. [1995](#page-21-0); Russell et al. [1996\)](#page-25-0). The uranyl compounds damage the proximal tubules, which are the main filtering component of the kidney. The renal injury is characterised by cellular injury and tubular necrosis (Cooley et al. [2000;](#page-21-0) Bellés et al. [2007](#page-21-0)). One mechanism of U toxicity may relate to the disruption of the first step in glycolysis by uranyl displacing magnesium (Mg) in the enzyme, hexokinase (Van Horn and Huang [2006\)](#page-26-0). Uranyl may also disrupt the paraaminohippurate transporter system and damage to peritubular cell membrane (Miller et al. [1995](#page-24-0)). Thus, U intakes of above 50 mg is fatal in humans (Van Horn and Huang [2006](#page-26-0); Tsuruta [2006\)](#page-26-0).

Furthermore, a few studies have shown that exposure to chemical U lead to genomic instability in lymphocytes (Müller et al. [2004;](#page-24-0) Kryscio et al. [2001\)](#page-23-0). This is attributed to breaks and cross-linkages of DNA strands because hydrolysis in the backbone of the DNA molecule caused by uranyl ions, acting as a hard Lewis acid (Periyakaruppan et al. [2007;](#page-25-0) Geissler et al. [2009;](#page-22-0) Stearns et al. [2005](#page-26-0); Yazzie et al. [2003](#page-27-0)). U also alters the proteome in lung cells (Periyakaruppan et al. [2007](#page-25-0)). U induces significant oxidative stress in lung epithelial cells followed by concomitant decrease in the antioxidant potential of the cells due to loss of total glutathione and superoxide dismutase induced by the presence of U (Periyakaruppan et al. [2007](#page-25-0); Pressyanov et al. [1995;](#page-25-0) Piao and Hei [1993](#page-25-0)).

## Radiological toxicity

Radiological toxicity of U is primarily manifest in bone sarcomas, leukaemia, lung cancer and general increase of cancer risk, whereas combined chemical and radiological effects are responsible for nephrotoxicity (Bellés et al. [2007](#page-21-0); Nagarkatti et al. [1996;](#page-25-0) Veiga et al. [1998\)](#page-26-0). The main radiation hazard from U occurs through injection of insoluble U compounds and inhalation of U containing aerosols because all U isotopes mainly emit alpha particles that have little penetrating ability (Lloyd et al. [1996\)](#page-23-0). However, communities near abandoned U mines are exposed radiation from U decay products, which are sometimes stronger and more dangerous than the radiation from U. For instance, an increased risk of lung cancer is attributed to exposure from radon decay products.

The cancer cases induced by radiation are generally indistinguishable from other naturally occurring cancers. The development of cancer occurs years after the exposure takes place. The probability of developing a radiation-induced cancer rises with increasing U intakes. A high radiation dose increases lung tissue damage, which may lead to lung cancer. Risks for other radiation-induced cancers, including leukaemia, are lower than for lung cancer. Generally, the WHO and ILO recommends that exposure to U radiation should not exceed a dose of more than 1 mSv in a year (WHO [2001](#page-27-0); Simpson et al. [2004](#page-26-0)). In special circumstances, an effective dose of up to 5 mSv in a single year is permitted if the average dose over five consecutive years does not exceed 1 mSv/year. An equivalent dose to the skin should not exceed 50 mSv in a year.

# Ecotoxicity

The chemical ecotoxicity risks are greater than radiological because U has high solubility in various ecosystems. For instance, the Predicted No Effect Concentration (PNEC) is listed in Table 2 showing the toxicity in different compartments and organisms. Generally, the detrimental effects of U on soil bacteria are less than the effects upon plant cells (Chmielowski et al. [1994](#page-21-0)). Based on information from studies in plant communities that colonise natural U ore bodies, toxicity to U may occur at concentrations of 50 ppm in the rhizosphere, and acute toxicity symptoms occurs in higher plants at about ten times this concentration. A study in a U anomaly site in Kayerekera, Malawi showed that visible poisoning symptoms appear at concentrations above 50 ppm U, while acute toxicity symptoms appear at ten times the concentrations (Zeman et al. [2008;](#page-27-0) Vanhoudt et al. [2008](#page-26-0)). High U content may lead to special vegetation formations in some sites. For instance, nitrogen fixing (leguminous) plant species dominated the hotspots where

Table 2 A selection of predicted no effect concentration (PNEC) for the types of non-human biota showing interesting difference in ecosystem compartments

| Compartment                           | PNEC   |
|---------------------------------------|--|
| Terrestrial plants                    | $250 \text{ mg Ukg}^{-1}$ dry soil   |
| Soil biota (from microbes to animals) | 100 mgkg $^{-1}$ dry soil  |
| Freshwater plants                     | $0.005 \text{ mgL}^{-1}$   |
| Freshwater invertebrates              | $0.005 \text{ mgL}^{-1}$   |
| Freshwater benthos                    | 100 mg $kg^{-1}$ dry sediment  |
| Freshwater fish at water              | 0.4 mgL <sup><math>^{-1}</math></sup> in very soft water<br>$(<10 \text{ mg } CaCO3 L^{-1})$ |
|                                       | 2.8 mgL $^{-1}$ in soft water<br>$(10-100 \text{ mg } \text{CaCO}_3\text{L}^{-1})$           |
|                                       | 23 mg $L^{-1}$ in hard water<br>$(>100 \text{ mg } \text{CaCO}_{3}L^{-1})$                   |
| Mammals                               | 0.1 mgkg <sup>-1</sup> body weightday <sup>-1</sup>  |

Sheppard et al. ([2005\)](#page-26-0), data source, are the only research group in literatures (up to now, October 2012) that have approached uranium ecotoxicity from this perspective; therefore, there is no any other information and data to compare with

surface mineralisation of U occurred within 0.5 m at Kayerekera U anomaly site in Malawi. This is probably a long-term response of plant communities to an environmental gradient, the most important aspect being the chemical properties of U.

However, the ecotoxicity of U cannot be generalised. The potential for development of critical ecotoxicological conditions is different from one ecosystem compartment to another as well as the type of organism. In all, the most important is the endpoint used to access the U ecotoxicity because effective assessment depends on the endpoint used at a specific ecological complexity level (Sheppard et al. [2005;](#page-26-0) Franklin et al. [2000;](#page-22-0) Charles et al. [2002b](#page-21-0); Hogan et al. [2005\)](#page-23-0). Consequently, information of U ecotoxicity is scarce and controversial. Assessment of U ecotoxicity is controversial because most organisms exhibit hermetic responses to low concentration and chronic exposure to U while as toxicity is observed only at acute exposure (Mkandawire et al. [2006b\)](#page-24-0). Some organisms relocate their resources (like energy or multiply rapidly) to withstand the U toxicity in low concentration (Mkandawire and Dudel [2005](#page-24-0)). Some invertebrates as well as some macrophytes secrete or exude organic compounds that protect them from U uptake as well as toxicity (Schaller et al. [2008](#page-25-0)). As a result actual effect of U is debatable—is it the lethal or inhibition effect or just presence of stress? The hermetic response can itself be an endpoint depending on the level of complexity at which U ecotoxicity is assessed. For instance, the effect of U at the organism metabolic level can be assessed effectively using molecular biomarker, inhibition of specific and important metabolic process, absence of some proteins and energy chemical molecules like ATPs, etc. Other endpoints are oxidative stress at cell molecular level and growth rate inhibition at population level. At community level, U ecotoxicity seems to be invisible. Thus, several areas of information need further definition to describe adequately the ecological consequences of U. Long-term observations are fundamental to accurate evaluation of the environmental impact of U because of the gradual modified response of ecosystems to U toxicity that follows acute effects (Yang and Volesky [1999a;](#page-27-0) Mkandawire et al. [2006a](#page-24-0); Mkandawire et al. [2007;](#page-24-0) Charles et al. [2006](#page-21-0)).

Furthermore, biomagnifications of U via the trophic contamination route is also scarce. The clay and organic matter content, acidity, and the presence of cations of Ca and Mg as well as Fe in the soil (Saric et al. [1995](#page-25-0); Fresquez et al. [1998\)](#page-22-0) influence the U uptake by terrestrial plants. Similarly, carbonate content and presence of cation (e.g.  $H^+$ ,  $Ca^{2+}$  and  $Mg^{2+}$ ) mostly determines U uptake in aquatic ecosystem because of the formation of soluble U carbonate complexes and the competitive interaction between cations with the uranyl ion  $(UO_2^{2+})$ , respectively (Charles et al. [2002a;](#page-21-0) Fox et al. [2006;](#page-22-0) Zanonato et al. [2012](#page-27-0); Kim et al. [2012](#page-23-0); Doudou et al. [2012;](#page-22-0) Kerisit and Liu [2012;](#page-23-0) Doudou et al. [2011\)](#page-22-0). Most of the U penetrate the roots surface through the epidermis, and then precipitates within the root, which explains the localisation of U in plant roots (Dushenkov et al. [1997;](#page-22-0) Dushenkov [2003;](#page-22-0) Rodriguez et al. [2006\)](#page-25-0). The highest concentrations are usually found in seeds and roots of most plant species, rather than leaves (Saric et al. [1995;](#page-25-0) Dushenkov et al. [1997](#page-22-0); Yong and Macaskie [1998](#page-27-0)).

# U bioremediation $2$

Unlike many organic compounds, U and all other metals do not disintegrate into small components that are non-toxic. Logically, biological activity can (1) only transform the metal from toxic to non-toxic species or (2) immobilise the metals from the pathway. Figure [4](#page-10-0) demonstrates the interactions of U with biotic component and summarise the mechanisms involved metal bioremediation. Uptake, sorption and precipitation by purified biopolymers derived from microbial cells provide alternative methods and additive processes for conventional physicochemical bioremediation (Malekzadeh et al. [2002](#page-24-0)). Some plants and algae species can sequestrate U and other metals into their biomass, while other species exude bioligands that trap U into biometalic colloids. A few bacterial species are capable of reducing mobile U(VI) to immobile U(IV) species (Smeaton et al. [2008](#page-26-0)). Furthermore, the algae-microbial bio-films have the ability to sequestrate U and other metals (Allan et al. [2002;](#page-20-0) Kalin et al. [2005b;](#page-23-0) Hsieh et al. [1994](#page-23-0)). The bioreduction, sequestration and trapping of U in biocolloids can nucleate biomineralisation processes of U. However, practical applications of bioremediation techniques under natural conditions are still far from reality.

For instance, U can be immobilised through adsorption onto cell surfaces, extracellular polysaccharides and low molecular weight organic acids including biogenic colloids (Lloyd and Renshaw [2005;](#page-23-0) Malekzadeh et al. [2002](#page-24-0)). Furthermore, U can be internalised into cells through metabolic active procedures. In the cells, U may be sequestrated as inert compounds in cell organelles such as vacuoles, cell membrane and cell wall. Some species excrete the U compounds back into the water as precipitates or organometalic colloids. Under conducive conditions, the interaction may result into U biomineralisation. For instance, some marine and freshwater algae are able to incorporate U into aragonite  $(CaCO<sub>3</sub>)$  mineral structures.

<sup>2</sup> Bioremediation is defined in this chapter as any process that uses microorganisms, fungi, green plants or their enzymes to remove contaminants from the contamination pathway. Under this definition, phytoremediation belongs to bioremediation.

<span id="page-10-0"></span>Fig. 4 The interaction of microorganism with metals emphasising the important processes that lead to U immobilisation in aquatic environments. The structure is modified from Bruins et al. ([2000\)](#page-21-0)



## Processes of U bioremediation

## Biosorption

U biosorption involves passive deposition of the metal ions into the biosorbent through various sorption processes, such as ion exchange, complexation, chelation and microprecipitation (Akhtar et al. [2007;](#page-20-0) Bhainsa and D'Souza [1999;](#page-21-0) Bustard and McHale [1997](#page-21-0)). The cell walls of most plant and algal or microbial materials are composed mainly of polysaccharides and carbohydrates, e.g. cellulose, xylan and mannans (Chmielowski et al. [1994](#page-21-0)). The backbone of polysaccharide include ligands, such as amino, carboxyl or hydroxyl, sulphide groups, which eventually has an overall negative charge influence on the cell wall (Gonzalez-Munoz et al. [1997](#page-22-0); Malekzadeh et al. [2002\)](#page-24-0). The negatively charged groups attract and bind U cations (O'Flaherty [1995;](#page-25-0) Sar and D'Souza [2001](#page-25-0)). The chemistry involved on the cell surface can be simplified as follows:

$$
\mathbf{R} - \mathbf{COOH} \Leftrightarrow \mathbf{R} - \mathbf{COO}^- + \mathbf{H}^+ \tag{7}
$$

$$
\mathbf{R} - \mathbf{O} - \mathbf{H} \Leftrightarrow \mathbf{R} - \mathbf{O}^- + \mathbf{H}^+ \tag{8}
$$

$$
\mathbf{R} - \mathbf{PO} - \mathbf{H} \Leftrightarrow \mathbf{R} - \mathbf{COO}^- + \mathbf{H}^+ \tag{9}
$$

Deprotonation leads to negatively charged sites, which are convenient for uranyl adsorption:

$$
\mathbf{R} - \mathbf{COOH} \Leftrightarrow \mathbf{R} - \mathbf{COO}^- + \mathbf{UO_2}^{2+} \tag{10}
$$

The cell wall structure and the characteristics of the U ion determine the capacity and efficiency of bioremediation of U water using sorption techniques. The capacity of U adsorption onto cell surface depends on the number of available ligand binding sites, U speciation and the physicochemical properties of the media. The chemical composition and cell size are reasonably constant for a given species, implying that the number of ligand groups on a surface is also fixed. Furthermore, the complexation is limited to a single layer (Ariff et al. [1999](#page-20-0); Prikryl et al. [2001](#page-25-0); Kalin et al. [2005a](#page-23-0)). Therefore, the surface of the cells carries a constant concentration of U with respect to the cell composition, even if the U ions are in excess of the concentration of affinity sites (Kalin et al. [2005a\)](#page-23-0). The number of available surface binding sites can only increase with cell growth, thus the rate at which U ions are removed from water by biosorption procedure is a function of growth rate. A few studies have estimated the plausible number of high affinity surface binding sites on algae and bacteria cells. For instance, phytoplankter have about 108 U binding sites per cell (Kalin et al. [2005a;](#page-23-0) Charbonneau [2009\)](#page-21-0), while on the bacterium Shewanella putrefaciens has approximately 2.0×  $10^{19}$  carboxyl sites,  $5.5 \times 10^{18}$  phosphoryl sites and  $2.3 \times$  $10^{19}$  amine sites per gram of bacteria, which translates to a biosorption capacity of up to 100 mg of U (Kalin et al. [2005a](#page-23-0); Daković et al. [2008](#page-21-0); Yang and Volesky [1999a](#page-27-0)).

There are many ligand types on a cell surface, which result in selective binding of U cation species. Some ligand groups may carry a positive charge and are thus able to absorb and complex anionic metal species (Bonfada et al. [2005](#page-21-0); Haas et al. [1998](#page-22-0)). The mixture of positively and negatively charged groups on a cell surface determines the number of cations and anions complexation. The U cation adsorbed onto most cell surfaces, while binding anionic, complexed species of U are more difficult, as there are fewer positively charged ligands on cell surface (Kalin et al. [2005b](#page-23-0); Markich [2002;](#page-24-0) Worms et al. [2006\)](#page-27-0). In acidic aqueous solutions of low pH value, U is present mainly as free  $UO_2^{2+}$  and adsorbs poorly to cell surface due to competition with protons for binding sites on the biomass (Yang and Volesky [1999a;](#page-27-0) Yang and Volesky [1999b](#page-27-0)). At pH values higher than 6, sorption is higher because of ion exchange between hydrolysed uranyl ions,  $UO_2OH^+$ ,  $(UO_2)^3(OH)_5^+$ ,  $(UO<sub>2</sub>)<sub>2</sub>(OH)<sub>2</sub><sup>2+</sup>$  and protons (Kalin et al. [2005a](#page-23-0); Meinrath [1998\)](#page-24-0).

Living and dead cells behave in a similar fashion with respect to adsorption of U, provided the cell wall structure remains intact (Donnellan et al. [1995;](#page-22-0) Mkandawire et al. [2003](#page-24-0); Volesky and May Phillips [1995\)](#page-26-0). Thus, the dead algae, microbes and plants biomass still contain negatively charged ligands, capable to adsorb positively charged U. With the help of invertebrate shredders, the biomass is reduced to particulate organic matter to which more U is adsorbed because of the increase in the surface to volume ratio (Kalin et al. [2005a\)](#page-23-0). Furthermore, the dead biomass decays into humic and fulvic substances through microbial activities. The humic and fulvic substances are rich in dissolved ligands probably because of the carboxyl and hydroxyl groups associated with the breakdown of lignin. The carboxyl group has a dissociation constant  $(pK_s)$  of about five, which facilitate the complexation with U at wide range of pH values (Dodge and Franscis [1997](#page-22-0); Pollmann et al. [2006;](#page-25-0) Shanbhag and Chopping [1981](#page-25-0)).

# Bioprecipitation

Bioprecipitation occurs when U species interact with biogenic compounds. The processes can take place either internal or external of the cell. Bioprecipitation of U is involved in biosorption as well as in active or metabolic internalisation. Most plant and microbe populations release extra-cellular polysaccharides as a by-product of metabolism (Francis and Dodge [1998;](#page-22-0) Macaskie and Basnakova [1998;](#page-24-0) Macaskie et al. [2000](#page-24-0)). Additionally, when imbalance in resource stoichiometry occurs, homeostatic regulation causes the affected organisms to exude chelates, low molecular weight organic substance and polyphenolic substances (Charbonneau [2009](#page-21-0); Kalin et al. [2005a;](#page-23-0) Schmeide et al. [2003\)](#page-25-0). These excreted by-products or exudates interact with U resulting in precipitate formation but not necessarily on the cell surface. Furthermore, their introduction into the water alters the chemical speciation, which also results in immobilisation of U. For instance, microbes such as Pseudomonas (Gamma Proteobacteria) and Citrobacter (Omnibacteria) also produce large quantities of extracellular polysaccharides (Allan et al. [2002;](#page-20-0) Malekzadeh et al. [2002\)](#page-24-0), while some macrophytes like Lemna sp., and Myriophyllum aquaticum exude oxalates and polyphenolic substance that complex U ions (Mkandawire [2005](#page-24-0); Schmeide et al. [2003](#page-25-0); Mazen et al. [2003](#page-24-0); Ma [2000](#page-24-0)). There is also evidence that interaction of phosphate with U leads to formation of very stable crystalline structures (Duff et al. [2002;](#page-22-0) Murakami et al. [1997](#page-25-0); Mkandawire et al. [2005](#page-24-0)). For instance, a homeostatic regulation of sudden uptake of U by Peltigera sp. lichen) leads to cellular formation of uranyl phosphate crystals, which are stored in the cellular mycobiont (Haas et al. [1998;](#page-22-0) Branquinho et al. [1997\)](#page-21-0). This formation of U(VI) phosphate crystals is widespread and occurs mostly on the surface of cells like in bacteria Citrobacter sp. (Thomas and Macaskie [1998](#page-26-0)). The mechanism involves production of alkaline phosphatase, which causes the cells to excrete phosphate to the cell surface. The phosphorus acts as the nucleus for the precipitation of U(VI) as uranyl phosphate species (Macaskie et al. [2000;](#page-24-0) Jerden et al. [2003](#page-23-0); Geissler et al. [2009\)](#page-22-0). Another homeostatic regulation of stoichiometry imbalance induced by U uptake is apparent in Lemna gibba. In L. gibba, uranyl oxalate crystals develop to counteract presence of U in the cells (Mkandawire et al. [2005\)](#page-24-0).

# Chelation

Chelates are organic molecules that trap or encapsulate highly reactive trace metal cations, and consequently prevent them from involvement in unwanted chemical reactions. Sometimes, the binding between metals and chelates may lead to precipitation (i.e. form insoluble compounds). Thus, most plants and microorganism use chelates to aid the uptake and transportation of essential metal nutrients. Due to unique binding properties, chelates are often used to stabilise metals, which makes them critical compounds in bioremediation (Kalin et al. [2005a](#page-23-0)). Chelating agents produced by microorganism are mostly siderophores, which enhance the dissolution of Fe and increase its mobility and bioavailability. Siderophores are low molecular weight, and function primarily as Fe transport compounds. Since there are some similarities in the biological behaviour of ferric and uranyl ions, chelates can bind U in a manner similar to those produced for Fe (Premuzic et al. [1985\)](#page-25-0). Plants first release special chelates known as phytosiderophores that bind to metals in the rhizosphere and allow them to be absorbed by the plant (Chiang et al. [2006;](#page-21-0) Jin et al. [2006;](#page-23-0) Wasaki et al. [2005](#page-26-0)). Functioning of chelates is stereoselective, depending on recognition of the metal ion coordination geometry. For example, some chelates like ethylendiaminetetraacetic acid function hardly on U because of the linear structure of uranyl cation (Meinrath et al. [1999](#page-24-0)). Nonetheless, benefits of chelating agent on U remediation are enormous. Low molecular weight organic acids like citric acids, oxalic acids exuded by most aquatic plants, facilitate the dissolution of U and uptake by macrophytes and microorganism (Ebbs et al. [1998b\)](#page-22-0). Other chelates include metallothionens and phytochelatins, which facilitate both uptake and transportation of metal in organisms. Phytochelatins are heavy metal complexing peptides composed of different amino acids (r-glutamic acid cysteine) nglycine  $n=3-7$ , which are involved in detoxication and homeostatic balance of heavy metals in plant cells (Kaplan et al. [1998;](#page-23-0) Maksymiec [1997](#page-24-0); Klapheck et al. [1995](#page-23-0)). Excess heavy metals are bound to cell wall in a process called metathiolate formation through mercaptide complexes (Shah and Dubey [1998\)](#page-25-0). Consequently, presence of chelating agents facilitates bioreduction of U because of the influence on U bioavailability.

## Active intracellular uptake

Probably the most widely recognized biological processes for metal removal in wetlands is plant uptake. The main route of U uptake in aquatic plants is through roots in emergent and surface floating. In plants with either completely submerged leaves or both floating and submerged leaves or free-floating, metals are taken up through both leaves and roots. In the case of foliar uptake of U, a passive movement in aqueous phase through cracks in the cuticle or through the stomata to the cell wall and then the plasmalemma is involved. Submerged rooted plants potentially extract water or metals from sediments, while rootless plants like *Wolfia* sp. or floating macrophytes like *Lemna* sp. extract metals only from water.

Previously, it was thought that a few aquatic macrophytes and microorganisms including algae and plants actively pump metals across their cell membranes (Curie and Briat [2003](#page-21-0); White and Broadley [2003](#page-27-0); Van Assche and Clijsters [1990\)](#page-26-0). The energy source is usually coupled to photosynthesis and temperature. However, emerging reports indicate clearly that a majority of macrophytes actively avoid uptake of U ions into their system (Duquène et al. [2009](#page-22-0); Daković et al. [2008](#page-21-0); Bhat et al. [2008](#page-21-0)). U uptake takes place when the external U concentrations far exceed an arbitrary threshold point of cell resistance. U may be taken up by some cells when the U species mimic an essential metal or interact closely with nutrient resources like  $PO_4^{3-}$ . Once inside the cell, U may be concentrated and sometimes be precipitated into vacuoles like in some plants and algae, or sequestrated in the cell wall in some U resistant microphytes (Martinoia et al. [2007](#page-24-0); Samardakiewicz and Wozny [2000;](#page-25-0) Neumann et al. [1995\)](#page-25-0). In several plant and algae species, U may also interact with internal inorganic phosphate, to create U-phosphate crystals, which are stored in the cellular mycobiont (e.g. in *Lemna* sp.) (Samardakiewicz and Wozny [2000](#page-25-0)). Some organisms react to potential U toxicity, by changing the U redox status, which induces their binding and subsequent excretion as non-mobile organometalic compounds. Furthermore, some plants species through toxicity-avoidance mechanism take up and sequester U in the roots like Phragmites australis, while other species transfer the U into the above ground parts vis-à-vis shoots. The realisation of the different allocations of bioaccumulated U is important in designing the remediation procedures.

The rate of U uptake by aquatic macrophytes and algae varies widely depending on plant species, growth rate, U speciation and concentration of the U in plant tissue. The pH, organic matter context and plant genotype have marked effects on U uptake. The  $UO_2^{2+}$ , uranyl carbonate complexes and  $UO_2PO_4^-$  are preferentially taken up by roots and transferred to the shoots (Vandenhove et al. [2007](#page-26-0)). Furthermore, when in mixtures, U(VI) competes for uptake with other metallic ions in terms of molecular size and shape and the configuration of the ligand. For instance, Li, Na, K and NH4 do not affect U accumulation. However, metals of the transition group as well as Al, Ca, Fe and Zn ions compete with U(VI) for uptake by biotic system especially macrophytes and algae (Kalin et al. [2005a\)](#page-23-0). U uptake is hindered in the presence of phosphate and carbonate ions but not affected by cations (Na, K, Mg, Ca, manganese (Mn), Co, Ni and Zn), nitrates, sulphates and thiosulphates. Nevertheless, the overall contribution of bioaccumulation procedure in the overall immobilisation of U in an aquatic system is quite small.

#### Biological-induced redox reactions

General mechanism Numerous natural-occurring metal dissimilatory and SRB are capable of obtaining energy through coupling the oxidation of organic matter and  $H<sub>2</sub>$  to the reduction of soluble  $U(VI)$  to insoluble  $U(IV)$  (Fig. [5](#page-13-0)). Hydrocarbons are electron donors that provide a source of electrons to reduce metals in a redox reaction. For example, the half reaction for  $U(VI)$  to  $U(IV)$  is:

$$
UO_2^{2+} + 2e^- \leftrightarrow UO_2 \tag{11}
$$

When metabolised by the bacteria, the hydrocarbons provide the necessary electrons to carry out the reaction 11. Remediation benefits from the process because once reduced, the metals are less soluble and easily precipitate out of solution. For instance, the reduced product, U(IV), is a non-toxic precipitate and a mineral, uraninite  $(UO<sub>2</sub>)$  (Jerden and Sinha [2006;](#page-23-0) Min et al. [2005](#page-24-0)). However, many bacteria are not capable of using U(VI) as a sole terminal electron acceptor. Nevertheless, microbial reduction of soluble U(VI) to insoluble U(IV) is one potential strategy of immobilising U from water pathways in abandoned U mines because it promotes U mineralisation under in situ environmental conditions (Mkandawire et al. [2005;](#page-24-0) Virk et al. [1999](#page-26-0); Ginder-Vogel et al. [2006](#page-22-0)).

Enzymatic reduction mechanism The pathway of enzymatic reduction of U(VI) involves c-type cytochromes as a reductase in presence of an electron donor, such as hydrogen gas (Behrends and Van Cappellen [2005;](#page-21-0) Liger et al. [1999](#page-23-0)). In the  $H_2$ -utilising bacteria, the electron in transferred by the tetraheme cytochromes c3, while the triheme periplasmic cytochrome c7 play a role in the transfer of electrons from

<span id="page-13-0"></span>

Inorganic electron acceptor

acetate to U(VI). There are suggestions that periplasmic and outer membrane cytochromes also play an important role (DiSpirito et al. [1983](#page-22-0); Macaskie et al. [2000](#page-24-0)). The reduction of U(VI) to U(IV) requires two electrons. However, the microorganism can transfer only one electron. The one-electron reduction process initially generates U(V), for instance, the U reduction (Luo et al. [2007;](#page-24-0) Wall and Krumholz [2006\)](#page-26-0). Due to the instability of U(V) complexes, it disproportionates to form U(IV). The enzymatic reduction of U can sometimes be hindered by the presence of some natural occurring chemicals. For instance, bicarbonate ion concentrations reduce enzymatic reduction of U(VI) by lowering the half-cell potential of the U (IV)–U(VI) couple (Ortiz-Bernad et al. [2004;](#page-25-0) Rawlings and Silver [1995\)](#page-25-0). Thus, there is formation of ternary Ca–U(VI)– CO complexes that make it a less energetically favourable electron acceptor than U(VI)-carbonato complexes because of direct interaction of Ca with the cells or electron donor. The U (VI) reduction is sometimes hindered by the presence of ferrihydrite (Raicevic et al. [2006\)](#page-25-0).

Bioreduction through Fe-reducing bacteria The majority of Fe(III)-reducing microorganisms can conserve energy by coupling the oxidation of hydrogen or organic compounds to the reduction of Fe(III). Similarly, they also have the ability to reduce soluble U(VI) to insoluble U(IV) in the same manner (Luo et al. [2007](#page-24-0); Madden et al. [2007](#page-24-0); Wall and Krumholz [2006\)](#page-26-0). For example, the Geobacter sp., which normally gains energy by reducing Fe(III) to Fe(II), can reduce U(VI) to U (IV) instead. They can use U(VI) as an electron acceptor, instead of Fe(III) (Min et al. [2005\)](#page-24-0). Further, bacteria from both the delta and gamma subdivision of the Proteobacteria like Geobacter metallireducens, Geobacter sulfurreducens, Desulfuromonas palmitatis and Desulfuromonas acetoxidans couple complete oxidation of acetate to  $CO<sub>2</sub>$  via the citric acid cycle with Fe(III) as the electron acceptor (Fredrickson et al. [2000b;](#page-22-0) Wall and Krumholz [2006;](#page-26-0) Wilkins et al. [2006](#page-27-0)). In

addition to acetate, Geobacter sp. can also oxidise ethanol, propionate, butyrate, valerate, pyruvate, propanol and toluene amongst others to carbon dioxide with Fe(III) as the electron acceptor (Fredrickson et al. [2000a;](#page-22-0) Wall and Krumholz [2006;](#page-26-0) Davis et al. [2006\)](#page-21-0). The bacterial Shewanella oneidensis, Shewanella alga and Pseudomonas sp. do not use organic electron donors like acetate (Luo et al. [2007\)](#page-24-0) but mostly couple the oxidation of hydrogen to the reduction of Fe(III) (Luo et al. [2007](#page-24-0); Choppin [2006](#page-21-0); Wilkins et al. [2006;](#page-27-0) Takashima et al. [2002](#page-26-0)). Given the wide range of Fe(III) reducing bacteria, it is therefore not surprising that they exhibit a high degree of metabolic diversity, with various species able to reduce U(VI). Despite the apparent diversity of Fe(III) reducing bacteria, Geobacter species are the dominant Fe(III) reducing microorganisms (Anderson et al. [2003;](#page-20-0) Neal et al. [2004;](#page-25-0) Lovley [1995;](#page-23-0) Lovley [2003\)](#page-23-0).

Therefore, the capability to couple the oxidation of organic matter to the reduction of Fe(III) and alternatively U (VI) is through the following typical reactions:

$$
[\mathbf{CH}_{3}\mathbf{COCO}_{2}]^{-} + 2\mathbf{Fe}(\mathbf{III}) + 2\mathbf{H2O} \cong [\mathbf{CH}_{3}\mathbf{COO}]^{-} \quad (12)
$$

$$
+\mathbf{HCO}_{3}^{-} + 2\mathbf{Fe}(\mathbf{II}) + 3\mathbf{H}^{+}
$$

In addition, then:

$$
[\mathbf{CH}_3\mathbf{COO}]^- + 8\mathbf{Fe(III)} + 4\mathbf{H}_2\mathbf{O} \Leftrightarrow 2\mathbf{HCO}_3^-
$$

$$
+8\mathrm{Fe}(\mathrm{II})+9\mathrm{H}^{+} \tag{13}
$$

Concurrently or alternatively existence of Fe(III) with U (VI), the reactions include the following:

[CH3COO]<sup>-</sup> + 4U<sup>6+</sup> + 4H<sub>2</sub>O 
$$
\approx
$$
 4U<sup>4+</sup> + 2HCO<sub>3</sub><sup>-</sup> + 9H<sup>+</sup>  
(14)

$$
\mathbf{H}_2 + \mathbf{U}^{6+} \Leftrightarrow \mathbf{U}^{4+} + 2\mathbf{H}^+ \tag{15}
$$

The Fe(III) dissimilatory procedure involves the ferric Fe reductase, probably located on the outer membrane in Gramnegative bacteria (Creanga et al. [2004;](#page-21-0) Lovley [2008;](#page-23-0) Ortiz-Bernad et al. [2004](#page-25-0)). The ferric Fe reductase acts as the terminal reductase of an electron transport chain, which is linked to the cytoplasmic membrane (Lovley [2003](#page-23-0); Anderson and Lovley [2002](#page-20-0)). Electrons are transferred down the transport chain to the ferric Fe reductase, which then transfers them onto insoluble extra-cellular Fe(III) oxides. The transfer of electrons is a mode of conserving energy through the generation of ATP (Vauclare et al. [2002](#page-26-0); Cohen et al. [1998](#page-21-0)). The Fe(III) is reduced by the ferric reductase either before or after uptake into the cell, thus forming a weak Fe(II)-chelate complex from which the Fe can be easily dissociated for use by the cell. Three possible mechanisms employed through which the Fe(III)-reducing bacteria reduce Fe(III) oxides as well as U(VI) have been identified as follows:

- 1. Direct contact, where the microorganism contact the insoluble Fe oxide and transfer an electron from the cell onto the Fe(III) oxide surface with outer membrane bound c-type cytochromes playing some role in this process (Lovley [2008](#page-23-0); Anderson et al. [2003](#page-20-0); Lloyd and Lovley [2001](#page-23-0); Lovley [1995\)](#page-23-0). Some bacteria utilise pili in electron transfer from the cell to the mineral surface as highly conductive 'nanowires' (a pilus is typically 6 to 7 nm in diameter);
- 2. Fe(III) chelators, where the bacteria produce siderophre to dissolve Fe(III) oxides which become more accessible to Fe(III) reductase than insoluble Fe(III) oxides (Lovley [2008;](#page-23-0) Anderson et al. [2003;](#page-20-0) Lloyd and Lovley [2001](#page-23-0); Lovley [1995](#page-23-0)); and,
- 3. Extra cellular electron-shuttling molecules, such as quinone-containing compounds or c-type cytochromes, to transfer electrons from the cellular electron transport chain to the metal terminal electron acceptors (Yang et al. [2001](#page-27-0); Fredrickson et al.[2000a;](#page-22-0) Koike et al. [1996](#page-23-0)). The extracellular electron shuttles may either be already present in the environment due to decomposition of organic material or produced by cellular processes and excreted into the environment (Lovley [2008\)](#page-23-0). Proteins never act as electrons-shuttling compound while humic substances are very good electron-shuttling compounds.

Bioreduction through sulphur-reducing bacteria Sulphurreducing bacteria (SRB) equally reduce U(VI) to U(IV) like it sulphur that it is salvaged for bioremediation in abandoned U mines. Just like the Fe(III)-reducing bacteria, the chemical basis of SRB remediation involves microbialmediated sulphate reduction coupled with organic matter oxidation. The reaction occurs as follows:

$$
2CH2O(aq) + SO42- + H+ \rightarrow H2S + 2HCO3
$$
  

$$
\rightarrow H2S + H2O + CO2
$$
 (16)

where CH<sub>2</sub>O can be replaced by other organic substance. Hydrogen sulphide  $(H_2S)$  then reacts with U, to form a sulphide that precipitates out of the effluent:

$$
H_2S + UO_2^{2+} \to UO_2S + 2H^+ \tag{17}
$$

As indicated in reaction 17, some bicarbonate  $(HCO<sub>3</sub><sup>-</sup>)$ may be accompanied by  $CO<sub>2</sub>$  production in the sulphur redox reaction. This consumes acidity, raising the pH, which facilitates the precipitation of U and related heavy metals. SRB generally rely on simple carbon compounds such as organic acids or alcohols to serve as electron donors for sulphate reduction, though some are capable of using hydrogen (De Schamphelaere et al. [2010;](#page-21-0) Lovley [2008](#page-23-0); Spear et al. [1999](#page-26-0); Basnakova et al. [1998](#page-20-0); Charbonneau [2009](#page-21-0)). When organic matter is used as a carbon source, other heterotrophic bacteria degrade it into simple carbon compounds (Fig. 6). Similar to mechanism in Fe(III)-reducing bacteria, the SRB transfers one electron to U(VI), reducing it to U(V). As mentioned earlier, U(V) is unstable and prone to disproportionate, forming U(VI) and U(IV).

Pre-conditions for bioreduction processes Generally, bacteria couple the oxidation of organic matter to the reduction of oxygen as the terminal electron acceptor in aerobic environments. Under



Fig. 6 Microbial processes of involving oxidation of organic carbon substance coupled to sulphatereduction, which further reduces U(VI) to U(IV). Modified from Doshi [\(2006](#page-22-0))

anaerobic conditions, microbes can utilise  $NO<sub>3</sub><sup>-</sup>$ , Mn(IV), Fe (III),  $SO_4^2$ <sup>-</sup> and  $CO_2$  as terminal electron acceptors in the oxidation of organic matter (Wilkins et al. [2006\)](#page-27-0). Thus, amount of  $NO<sub>3</sub><sup>-</sup>$ , Mn, Ca (because of its relationship to  $CO<sub>2</sub>$  in water) influence the U(VI) reduction because of competition. Fortunately, Fe(III) reduction accounts for the oxidation of up to 65 % of organic matter in some anaerobic sediment (Seidel et al. [2002;](#page-25-0) Finneran et al. [2002;](#page-22-0) Rittle et al. [1995](#page-25-0); Charbonneau [2009\)](#page-21-0). However, the reduction of Fe(III) to Fe(II) lead to other significant chemical changes, such as the release of trace metals and radionuclides into the water pathway and the inhibition of methane production (Mishra et al. [1999](#page-24-0); Zayed and Winter [2000\)](#page-27-0), as well as Fe(II) functioning as a catalyst in reduction on U(VI) to U(IV). Nevertheless, the bioreduction of U occurs much faster than abiotic reduction. For SRB, anaerobic conditions where the  $E_{\rm H}$  is less than −200 mV are required because oxygen, nitrate, Mn and Fe reduction all yield more energy per equivalent than sulphate (Doshi [2006](#page-22-0)). The redox conditions are also suitable for Fe reduction to the ferrous ( $Fe^{2+}$ ) state, which co-precipitate with sulphide. When electron donors are in sufficient to drive the reduction of U, some bioremediation strategies like addition of nutrients or organic compounds, especially alcohol, are done to be electron source. In some situations, there may be a more suitable electron acceptor than the metal itself, such as oxygen. In such case, bioremediation strategies can focus on creating an anaerobic environment such as a constructed wetland.

## Phytoremediation

Phytoremediation is the use of plants to clean-up contaminated sites (Pilon-Smits [2005\)](#page-25-0). Metals can be phytoremediated by several different processes including phytoextraction, rhizofiltration and phytostabilization. Plants also have the ability to remove trace metals from the water through biological uptake and surface adsorption (Slaveykova and Wilkinson [2003;](#page-26-0) Slaveykova and Wilkinson [2002\)](#page-26-0). In phytoextraction, plants take up and concentrate the U and other metals in their tissues. Certain plants species like Lemna sp. and Phragmatis sp. have evolved the capacity to extract particularly high levels of U. Once U and other metals taken up, they are either stored in the root or transported to the shoots through the xylem (Overall and Parry [2004](#page-25-0); Ebbs et al. [1998a\)](#page-22-0). Normally, U has difficulty migrating through the high cation exchange capacity of the xylem. Most metals are chelated to facilitate translocation process, but the transport process for U is not yet clear. However, U is probably immobilised and stored in the cell vacuoles as well as cell wall. Inside the vacuole, U is bound to organic compounds, probably chelates to prevent further damage to the plant. Furthermore, the U may form crystals after uptake in the cell, for instance U oxalates observed in L. gibba (Mkandawire et al. [2005](#page-24-0)). Some plant species use their roots to filter U and other toxic

metals from the water. With this technique, U is deposited in the rhizosphere of the macrophytes either via precipitation, ion exchange or chelation (Kalin et al. [2005a\)](#page-23-0). The processes can be direct when the plant exudes organic substances that facilitate symbiotic, allelopathic associations with microorganisms. For some plant species, the filtering of U is through the rhizospheric microflora, which associates with the plants.

# Biogenic facilitated abiogenic reduction of U

Based on strict thermodynamics, Fe(II) has high potential of reducing U from U(VI) to U(IV). Many subsurface environments in abandoned U mines, especially sediments, are saturated with biogenic Fe(II) due to long-term microbial reduction of Fe(III) (Behrends and Van Cappellen [2005;](#page-21-0) Lack et al. [2002\)](#page-23-0). Logically, the Fe(II)-rich condition should facilitate the immobilisation of U and put abiotic reduction above direct microbial reduction process in U immobilisation in wetland ponds of abandoned U mines (Antunes et al. [2007](#page-20-0); Zänker et al. [2007;](#page-27-0) Mkandawire et al. [2004b](#page-24-0)). Additionally, Fe(II) species adsorbed onto ferrihydrite, magnetite and hematite surfaces are able to reduce U(VI) to U (IV). The reduced  $U(VI)$  vis-à-vis  $U(IV)$  is then present as a  $UO<sub>2</sub>(s)/Fe(OH)<sub>3</sub>(S)$  precipitate in the water. Therefore, abiotic reductions of U(VI) by biogenic Fe(II) are likely the dominant reductive pathway. However, the Fe(II) does not significantly contribute to U(VI) reduction in the sediments because , the reduction of U(VI) by Fe(II) is only a minor process due to the complexation of U(VI) with carbonate.

## Applicable bioremediation strategies in abandoned U mines

There are varieties of bioremediation strategies that can be explored to reduce U and other contamination associated with abandoned U mining and processing sites. Here, three commonly applied strategies in abandoned U mining sites that offer some of the most promising bioremediation treatments are discussed. These strategies provide cheap and efficient alternatives to traditional methods of metal decontamination such as 'pump and treat' soil washing systems, excavation and reburial of contaminated areas.

# Constructed wetlands

One strategy for bioremediation metals in abandoned U mines is the creation of artificial wetlands (Vymazal [2005](#page-26-0); Batty and Younger [2004;](#page-21-0) Lüderitz and Gerlach [2002](#page-23-0); Zayed et al. [1998\)](#page-27-0), which is applicable for treatment of contaminated surface and groundwater as well as acid drainage. Groundwater is pumped into constructed wetland for passive treatment. The wetland strategy relies on using several basic physical, chemical and biological processes to remove or immobilise U from the aquatic system. Thus, an effective strategy can involve all

the bioremediation processes once well designed. The U remediation occurs in the three main compartments of a wetland vis-à-vis the substrate (sediment), hydrology and vegetation.

The substrate harbours consortia of detritus microbial, which are responsible for uranyl reduction, and transformation, which may nucleate U biomineralisation. Wetlands with appropriate substrate promote the growth of sulphatereducing bacteria in anaerobic conditions. Thus in acid mine water which is rich in sulphates, these bacteria will generate hydrogen sulphide, which reacts with metals that leads to formation of insoluble metals sulphides (Hallberg and Johnson [2003;](#page-22-0) Seidel et al. [2002](#page-25-0)). This is very important in controlling generation of AMD from abandoned U mines.

The hydrology compartment contains heterogeneous polyligands like fulvic, humic and tannic acids, amorphous metal oxyhydroxides, clay, bacterial surfaces and associated exocopolymers, suspended particles and macromolecules (e.g. polysaccharides and proteins) (Gomez et al. [2006](#page-22-0); Müller et al. [2002;](#page-24-0) Martinez Aguirre et al. [1995\)](#page-24-0). Anaerobic conditions also develop easily in the upper portion of the sediment because of hydric condition that supports hydrophytic vegetation. Uranyl ion can coordinate with most biological material, living or dead. It is also transported through the water column to the sediments. This is slow in well-oxygenated water and fast in anoxic water.

While sediments of wetlands form primary sinks for U, macrophytes absorb U through roots and shoots. The wetland plants are also autotrophic organisms, creating a biomass of reduced carbon compounds that provide nutrients to a variety of both micro- and macroscopic (Lüderitz and Gerlach [2002\)](#page-23-0). The expansive rhizosphere of wetland plant species provides an enriched culture zone for microbes involved in U species biotransformation (Sheoran and Sheoran [2006\)](#page-26-0). Physical processes in constructed wetlands through which U is removal from water pathway include settling and sedimentation following adsorption on particulate matter (Schaller et al. [2008](#page-25-0); Selig et al. [2002](#page-25-0); Markich [2002](#page-24-0)). To some extent, mats of floating plants serve as sediment traps while rooted and floating plants regulate the flow rate of the water by providing resistance. For particles, which are light or less dense than water, sedimentation occurs after flocculation. Flocculates generally settle more rapidly in a wetland than do individual particles. The flocculation is enhanced by high pH, concentration of suspended matters, ionic strength and high algal concentration (Kalin et al. [2005a\)](#page-23-0). Before sedimentation, other chemical processes like precipitation, adsorption and coprecipitation should occur first. More than 50 % of the U is easily adsorbed onto particulate matter in the wetland (Dudel et al. [2004](#page-22-0); Fritzsche et al. [2006;](#page-22-0) Dienemann et al.[2003](#page-21-0)). In sediments, U adsorbed to the particles by either cation exchange or chemisorptions. The U speciation changes with time as the sediment conditions change.

#### Injection wells

Well injection is another promising bioremediation strategy applicable in abandoned U mine (Anderson et al. [2003\)](#page-20-0). This well injection strategy is illustrated in Fig. [7](#page-17-0), and it is one of the major procedures for U bioremediation in groundwater especial in abandoned ISL mining procedure as well as leaching contaminated water and AMD from the tailing and waste heaps. Mostly, bioremediation in the groundwater occurs through the bioreduction of U by Fe and sulphur-reducing microorganism. In most cases, the underground environments are rich with microorganisms, but the environmental conditions are usually not conducive for their growth and reduction of U. The environments usually lack carbon sources and contain high nitrates. Thus, biostimulation of microorganism activities is necessary for the U bioremediation to occur. A common method to deliver carbon sources are injection wells. For such constructions, a series of wells are installed to control groundwater flow and allow the injection of solutes, such as ethanol, required to create a geochemical environment conducive for microbial growth and subsequent U(VI) reduction. For instance, ethanol is added to stimulate in situ denitrification of residual nitrate and reduction of U(VI) (Wu et al. [2006\)](#page-27-0). The addition of acetate to groundwater promotes the growth and activity of dissimilatory Fe(III)-reducing microorganisms and results in the removal of soluble U(VI) from contaminated groundwater (Vrionis et al. [2005\)](#page-26-0).

# Bioreactive barrier

This is a bioremediation strategy targeting leaching and ground water. The strategies capitalises on the process of bioaugumentantion, where microorganisms are introduced in a form of a barrier to clean the leaching water (Ellen [2006;](#page-22-0) Fuller et al. [2002\)](#page-22-0). Figure [8](#page-17-0) illustrates how the barriers are constructed so that the contaminated waters passes through the barrier and is treated through the same process of U reduction using dissimilating Febacteria or SRB. To stimulate bioremediation, a SRB source such as cow manure or organic matter is added to passive treatment systems.

Challenges in application of bioremediation in AUM

The challenge to apply bioremediation effectively in abandoned U mines lies in the ability to overcome several limitations, uncertainties and failures related to bioremediation systems.

# Stoichiometry and homeostasis limits

There are several limitations to application of bioremediation in contaminated waters of abandoned U mining sites. <span id="page-17-0"></span>Fig. 7 Schematic illustration of well injection strategy of bioremediation where biostimulants like ethanol are injected into underground to stimulate U reduction by bacteria. The well can also be used for bioaugmentation where U-reducing bacteria are injected into the wells (Vrionis et al. [2005](#page-26-0))



One major limitation is with the nature of the organisms, whose immobilisation of U is not a benevolent gesture but a strategy for survival. Most bioremediation agents only work under environmental conditions that suit the organisms' needs. Thus, unless conducive bioremediation conditions are set, organisms use alternative substrates as source of energy than U. Consequently, some type of environmental modification is needed to encourage the organisms to take up or bio-reduce the U at an acceptable rate. Furthermore, the organism must be presented with low levels of the pollutant over a period to induce the organism to develop metabolic pathways needed to reduce the U. As a result, bioremediation can also become very costly due to modifications, labour intensive and can take several months for the remediation to achieve acceptable levels.

Fig. 8 Schematic illustration of how passive treatment using a permeable reactive barrier functions in treatment of tailing heap leachates especially AMD. For AMD, the reactive barrier can be filled with immobilised microorganisms

# Environmental condition limits

Some environmental factors may also limit the applicability and effectiveness of the U bioremediation process. There are occasions where sediment or soil matrices prohibit contaminant microorganisms contact. The matrices form a barrier that the contaminated water may not reach. A worse situation occurs when there is preferential colonisation by microbes, which may cause clogging of nutrient, and water flow in the pores of the sediments. Furthermore, preferential flow paths may severely decrease contact between the biostimulants and U as well as the bioremediation agents. Similarly, bioremediation in clay and highly layered or heterogeneous subsurface environments limit the application because such environments interfere with oxygen or other electron acceptor transfer processes (Luo et al.



[2007;](#page-24-0) Ginder-Vogel et al. [2006\)](#page-22-0). Therefore, the rate of U immobilisation from the aqueous phase depends on the rates of mass transfer of contaminants from the low permeability matrix blocks to higher permeability regions within fractures.

## Discharge pattern limits

Application of bioremediation is limited to the type of contaminant discharge in abandoned U mining sites because the procedure is not responsive to sudden changes. High concentrations of heavy metals are likely to be toxic to microorganisms. Low contaminant concentrations may be below the survival threshold for the organism. Consequently, bioremediation attempts fail and become costly because the microorganisms are sometimes site specific. Each new site represents a different set of conditions, therefore microorganisms used for bioaugmentation need to be adapted to each site. Bioremediation of air in particular is very inefficient, and therefore radon emanated from abandoned U mines is hardly tackled.

## Ecological implication fears

Another problem is that the applicable bioremediation technologies can cause environmental disruption beyond the damage done by the pollution. In most cases, biostimulation and bioaugmentation is unavoidable. These induce environmental conditions can be disruptive to other organisms thereby disrupting the ecological balance. For instance during biostimulation procedures, continued addition of acetate may result in a shift in the dominant terminal electron accepting process from Fe(III) reduction to sulphate reduction, as well as complete degradation of the acetate under sulphate-reducing conditions and an apparent decrease in the rate of removal of soluble U(VI) from groundwater (Vrionis et al. [2005](#page-26-0)). Therefore, the maintenance of Fe(III)-reducing conditions is critical for sustaining reductive precipitation of U(IV), and highlights the need to correlate U (VI) removal with alterations in subsurface microbial community. Furthermore, the modification of the environment can equally be expensive as classical treatments due to the cost, for example of acetate or ethanol in biostimulation. In bioaugmentation, the long-term effects of introducing naturally occurring non-native bioremediation organisms into an area can alter the ecological balance when they become invasive. Worse, most of high and fast acting are genetically altered bioremediation organisms, long term environmental effects are not yet clear. As a result, there are always uncertainty of horizontal transfer of the modified gene, creation of new pathogens and possibility of mutations that allow the organism to become invasive.

## Physicochemical characteristics limits

In many abandoned U mining and processing sites, ambient groundwater has a very low pH and high levels of

aluminium, Ca and nitrate. The pH values lower than 3.4 are not favourable for the growth of most organisms, which requires manipulation of subsurface pH to a value better suited for microbial activity. However, increasing the pH in situ is not desirable because aluminium hydroxide precipitates form at approximate pH values above 4.5 and Ca solids precipitate at near-neutral pH. Ca is present in the groundwater at levels that inhibit U(VI) reduction especially at neutral pH conditions. The formations of U-carbonate-Ca complexes also retard bioreduction (Wilkins et al. [2006\)](#page-27-0). The aqueous species,  $CaUO_2(CO_3)_3^{2-}$  and  $Ca_2UO_2(CO_3)_3$ also have an important role on inhibition of microbial reduction and aqueous speciation of U(VI) in contaminated groundwater.  $Ca^{2+}$  have a significant impact on the aqueous speciation of U(VI), and consequently, on the sorption and mobility of U(VI) in aquifers (Fox et al. [2006\)](#page-22-0).

Nitrate also inhibits U(VI) reduction and is present as a co-contaminant with U in groundwater (Franklin et al. [2000](#page-22-0)). Prior to the reduction of U, persistent nitrate must first and continuously be removed or transformed prior to U because nitrate is a preferred electron acceptor to U, and the stimulation of metal-reducing bacteria are hindered by lowpH environments (Madden et al. [2007\)](#page-24-0). In situ denitrification of bulk nitrate has high probability of incomplete denitrification because of nitrate levels are usually very high in abandoned U mining sites. Any nitrite formed can oxidise U (IV) back to the more soluble U(VI). In addition, complete denitrification of nitrate generates  $N_2$  in excess of its solubility limit. The presence of a gas phase can alter flow direction and decrease solid surface area accessible for sorption as well as desorption and redox reactions.

#### Processes and counteraction limits

The performance of wetland treatment system depends upon the input water quality and the specific mechanism of metal removal processes. The wetland plants translocation oxygen from the shoots to the root rhizomes through their internal gas space parenchyma. The roots and rhizomes in turn leak the oxygen to the reduced environment. Much as this is advantageous, it also limits U bioremediation. If the U immobilisation is aerobic, these oxidised conditions promote precipitation of oxyhydroxides of  $Fe<sup>3+</sup>$  and Mn<sup>2+</sup> that may adsorb or co-precipitate with the U ions (Davis et al. [2006](#page-21-0)). However, the volume available to collect and store chemical precipitates or the number of available removal sites governs elimination rate in aerobic systems. The primary removal processes, which include sedimentation, adsorption, ion exchange and complexation, are finite. Thus, U removal or immobilisation ceases unless new removal sites are generated. Hence, periodic removal of precipitated U from wetlands is necessary. Only systems that are well vegetated can generate sufficient new removal sites to offer

the possibility of long term, low maintenance treatment. Therefore, only site conditions may determine whether it is more economical to simply let the wetland system develop into a new ecosystem or to excavate away the accumulated solids for proper disposal. Since the generation of new removal sites is a slow process, long-term treatment require a large wetland with a relatively small input load. Further, the wetland concentrates the U in one place, which increases the radiation dangers and radon emanation.

If the primary U immobilisation mechanism uses anaerobic systems, oxygenating macrophytes affect the performance of U bioremediation agents in the sediments and subsurfaces. The system also works efficiently as long as sulphate-reducing bacteria reduce sulphate to sulphide thus precipitating U sulphides complexes and generating alkalinity. The mechanism is limited by the amount of available carbon for sulphate-reducing bacteria, but the oxygen availability provides a more preferable electron acceptor than U. Therefore, the bioreduction of U ceases. Furthermore, in the absence of organic-reducing sediments, U-bearing particles are released back into the water column when the macrophytes decompose. In reality, both aerobic and anaerobic processes take place simultaneously in the wetlands. This is a big challenge to designing of a constructed wetlands where these processes do not counteract once another.

## New perspectives—a paradigm of the eco-remediation

In most cases, the laboratory success stories have failed to be applied in the real situation (Bennett [1995](#page-21-0)). The main reason is that laboratory procedures do not completely apply the multiple interactions that take place in nature, especially the roles of different ecological interactions and hierarchy. A natural system is self-organised in an ecological hierarch where the interaction determines the role of different organism in the system. Hence, use of bioremediation procedure should be used at higher ecological interaction, where the multiple interactions are part of the processes. Thus, U remediation in abandoned U mines can be effective when of ecological engineering is employed to enhance natural ecological processes (Kalin et al. [2005a\)](#page-23-0) vis:

- 1. Association of the U with organic particles in the water column;
- 2. Provision of conditions whereby organic particles sink onto and into organic sediments; and,
- 3. Provision of low  $E_H$  conditions in the sediments for metal-reducing microbial populations.

Consequently, oxidised U associates with particulates, settles, is reduced and is bio-mineralised in the sediments effectively creating biogenic ores (Ketterer et al. [2000](#page-23-0); Martinez Aguirre et al. [1995](#page-24-0); Kalin et al. [2005a\)](#page-23-0). The advantage of using living material in the initial step is that it continually grows, providing new organic material for sequestration. This is especially useful in flow-through systems, where the supply of U to be removed is continuous. The removal of U using ecological approach follows a three-step process: (1) first is the sequestration of U on, in and around aquatic plants and periphyton biofilms. Periphytons are mixed communities, which are composed of algae, fungi and bacteria that grow on macrophytic-vascular plants and on sediment surfaces (Kalin et al. [2005a](#page-23-0)). Biogenic ligands produced by plants and microorganism facilitate the removal U(VI) from the mine water. The biofilms continuously renew the cellular surface area; (2) the next is the removal of the biological sequestrated U units from the water column. The macrophytes, phytoplankton and bacteria become aggregated into larger particulates, which sink rapidly to the bottom. Invertebrate shredders (e.g. zooplankton, diatoms etc.) and small aquatic vertebrates (e.g. herbaceous fish) play an important role in formulation of the U rich organic particulates. The faecal pellets are part of biogenic organic particles that bind U and are together transferred into the sediments (Kalin et al. [2005a\)](#page-23-0). However, the production of biogenic organic particles is attributed to high primary productivity. This organic particulate matter adsorbs the U and other metals from the water pathway. Thus, U(VI) is reduced to U(IV) which forma stable precipitates in the sediments. Once the biomass of U-biofilms reaches the sediment interface, it continues to decompose, releasing nutrients and colloidal-bound U. The dead organic material provides organic carbon and other nutrients to heterotrophic microbial consortia to maintain the low  $E_{\rm H}$ , within which the U is transformed and consequently bio-mineralised (Kalin et al. [2005a\)](#page-23-0). The nutrients that sustain microbial populations in the anaerobic sediments come from natural organic matter or algal particles. The biomass of the plant and biofilms encounters sediment primarily as decomposing material. The decomposition of the biomass in the sediments provides nutrients to sustain heterotrophic microbial communities which further degrade organic matter, and provide dissolved organic material for anaerobic metal-reducing microbial consortia (Kalin et al. [2005a](#page-23-0); Wielinga et al. [2000\)](#page-27-0).

Dissolved oxygen is reduced first, followed by denitrification, followed by  $UO_2CO_3$  reduction to uraninite and sulphate reduction to sulphide. At each step in the redox ladder, different microbial enzymes may catalyze reactions. There is a succession of microbial activity ranging from denitrifies, to metal reducers, to sulphate reducers. Denitrification process is complete before sulphate reducers are activated. Thus, the U (VI) reduction does not occur until the nitrate has been utilised by nitrate-reducing microorganisms (Finneran et al. [2002\)](#page-22-0). When nitrate enter a reduced zone where  $U(V)$  is present, the subsequent reduction of nitrate may remobilise the U though facilitation of oxidation of U(IV) to U(VI). Therefore, the anaerobic pond should be planted with high nitrogen demanding macrophytes like Lemna sp. and Eichonia sp. and

<span id="page-20-0"></span>denitrifying bacteria. Further, the intermediates of dissimilatory nitrate reduction (nitrite, nitrous oxide and nitric oxide) can also oxidise and mobilise U(IV). Therefore, the accumulation of carbonate arising from microbial respiration may promote the formation of highly stable carbonato-U(VI) complexes, thereby increasing the thermodynamic favourability of U(IV) oxidation (Gavshin et al. [2005](#page-22-0)).

# Concluding remarks

Bioremediation of U shows high potential as alternative to classic remediation under laboratory conditions. However, the crossing from pure academic research to actual marketable technology applicable in abandoned U mines is slow to come. Like many technologies with good scientific foundations, its merits are marred by over-optimistic speculations and fraudulent claims. Furthermore, U bioremediation is usually considered in the absence of other influential ecological interactions. This undermines its potential application in situ because natural aquatic environment including abandoned U mining sites are heterogeneous; and complex interactions involving water column, bed sediments, air and biotic compartment continuously take place. It is characterised with presence of multiple stressors, ligand heterogeneity and polydispersity and non-equilibrium conditions. Consequently, metals and metalloids undergo an array of dynamic physical, chemical and biological processes. For instance, the sulphides minerals, acid-buffering capacity, presence of organic matter, colloids, Fe and Mn oxide minerals affect the solubility, mobility and bioavailability of U and other metal ions. The toxicity of U is a function of its ability to cross biological barriers (e.g. plasma membrane) which is partially regulated by the concentration or flux of internalised metals and the physicochemical conditions of the medium. When the homeostasis is disrupted due to the interactions chemical interactions, the resulting stoichiometry imbalance is reversed by homeostatic regulation of the biotic compartment. Thus, the homeostatic regulation of resource stoichiometry in an aquatic system is one of the prime regulatory processes that can be salvaged for enhancing bioremediation technology as well as toxicity alleviation. The idea of stimulating these biological processes for the purposes of stabilising U in the subsurface is therefore promising as a basis for U remediation technologies. Therefore, a realistic functioning is only feasible if the bioremediation procedures are encapsulated in ecological engineering processes where multiple interactions are manipulated but left to emulate the rule of nature. For instance, cascade wetland procedure would be more effective than just monoculture constructed wetlands in treating U. The long-term stability of biologically reduced U will be determined by the complex interplay of sediment mineralogy, aqueous geochemistry, microbial and invertebrate activity, stabilisation by aquatic plants

and potential U(IV) oxidants. Many of these factors have been studied under laboratory conditions; however, the impact of these factors on U cycling in natural, subsurface environments is yet to be understood.

Acknowledgements The author is very much grateful to Prof. Dr. Wolfgang Pompe and Prof. Dr. E. Gert Dudel for the everlasting support, advice and encouragement. This review is based on scientific discussions and projects throughout a long-time collaboration between Prof. Gert Dudel and the author. Prof. Dr. Günther Meinrath kindly provided information and allowed the use of his material and intellectual property on uranium geochemistry. Dr. Klaus P. Kuhn and Dr. Msau Mkandawire read and constructively criticised the drafts. Finally yet importantly, the author is very grateful to the two anonymous reviewers who provided very constructive criticisms that helped improve the original draft.

## References

- Abdelouas A (2006) Uranium mill tailings: geochemistry, mineralogy, and environmental impact. Elem 2(6):335–341
- Adler T (1996) Botanical cleanup crews—using plants to tackle polluted water and soil. Sci News 150:42–43
- Akhtar K, Waheed Akhtar M, Khalid AM (2007) Removal and recovery of uranium from aqueous solutions by Trichoderma harzianum. Water Res 41(6):1366–1378
- Al Hashimi A, Evans GJ, Cox B (1996) Aspects of the permanent storage of uranium tailings. Water Air Soil Pollut 88(1–2):83–92
- Allan VJM, Callow ME, Macaskie LE, Paterson-Beedle M (2002) Effect of nutrient limitation on biofilm formation and phosphatase activity of a Citrobacter sp. Microbiol-Sgm 148:277–288
- Allard T, Ildefonse P, Beaucaire C, Calas G (1999) Structural chemistry of uranium associated with Si, Al, Fe gels in a granitic uranium mine. Chem Geol 158(1–2):81–103
- Anderson RT, Lovley DR (2002) Microbial redox interactions with uranium: an environmental perspective. In: Keith-Roach MJ, Livens FR (eds) Interacfions of microorganisms with radionuclides. Elsevier, Amsterdam, pp 205–223
- Anderson RT, Vrionis HA, Ortiz-Bernad I, Resch CT, Long PE, Dayvault R, Karp K, Marutzky S, Metzler DR, Peacock A, White DC, Lowe M, Lovley DR (2003) Stimulating the in situ activity of geobacter species to remove uranium from the groundwater of a uranium-contaminated aquifer. Appl Environ Microbiol 69(10):5884–5891
- Antunes SC, de Figueiredo DR, Marques SM, Castro BB, Pereira R, Goncalves F (2007) Evaluation of water column and sediment toxicity from an abandoned uranium mine using a battery of bioassays. Sci Total Environ 374(2–3):252–259
- Ariff AB, Mel M, Hasan MA, Karim MIA (1999) The kinetics and mechanism of lead(II) biosorption by powderized Rhizopus oligosporus. World J Microbiol Biotechnol 15:291–298
- Arogunjo AM, Hollriegl V, Giussani A, Leopold K, Gerstmann U, Veronese I, Oeh U (2009) Uranium and thorium in soils, mineral sands, water and food samples in a tin mining area in Nigeria with elevated activity. J Environ Radioact 100(3):232–240
- ATSDR (1999).Toxicological profile for uranium. 462 (Ed A. f.T. S. a. D. Registry). U.S. Department of Health and Human Services
- Barthelmy D. (2008). Mineralogy Database. <http://webmineral.com>
- Basnakova G, Spencer AJ, Palsgard E, Grime GW, Macaskie LE (1998) Identification of the nickel uranyl phosphate deposits on Citrobacter sp. cells by electron microscopy with electron probe

<span id="page-21-0"></span>X-ray microanalysis and by proton induced X-ray emission analysis. Environ Sci Technol 32(6):760–765

- Batty LC, Younger PL (2004) Growth of Phragmites australis (Cav.) Trin ex. Steudel in mine water treatment wetlands: effects of metal and nutrient uptake. Environ Pollut 132(1):85–93
- Beddow H, Black S, Read D (2006) Naturally occurring radioactive material (NORM) from a former phosphoric acid processing plant. J Environ Radioact 86(3):289–312
- Behrends T, Van Cappellen P (2005) Competition between enzymatic and abiotic reduction of uranium(VI) under iron reducing conditions. Chem Geol 220(3–4):315–327
- Bellés M, Linares V, Albina ML, Sirvent J, Sánchez DJ, Domingo JL (2007) Melatonin reduces uranium-induced nephrotoxicity in rats. J Pineal Res 43(1):87–95
- Belzile N, Tessier A (1990) Interactions between arsenic and iron oxyhydroxides in lacustrine sediments. Geochim Cosmochim Acta 54(1):103–109
- Benders J, Duff MC, Phillips P, Hill M (2000) Bioremediation and bioreduction of dissolved U(VI) by microbial mat consortium supported on silica gel particles. Environ Sci Technol 34:3235–3241
- Bennett JP (1995) Abnormal chemical element concentrations in lichens of Isle Royale National Park. Environ Exp Bot 35(3):259–277
- Bernhard G, Geipel G, Brendler V, Nitsche H (1998) Uranium speciation in waters of different uranium mining areas. J Alloys Compd 271–273:201–205
- Bhainsa KC, D'Souza SF (1999) Biosorption of uranium(VI) by Aspergillus fumigatus. Biotechnol Tech 13:695–699
- Bhat SV, Melo JS, Chaugule BB, D'Souza SF (2008) Biosorption characteristics of uranium(VI) from aqueous medium onto Catenella repens, a red alga. J Hazard Mater 158(2–3):628–635
- Biswas P, Wu CY (1998) Control of toxic metal emissions from combustors using sorbents: a review. J Air Waste Manag Assoc 48(2):113–127
- Boice JD Jr, Mumma MT, Blot WJ (2010) Cancer incidence and mortality in populations living near uranium milling and mining operations in grants, New Mexico, 1950–2004. Radiat Res 174(5):624–636
- Bonfada E, Oliveira GMD, Back DF, Lang ES (2005) Metallation of ligands with biological activity: synthesis and X-ray characterization of  $[UO_2(PN)_2(H_2O)]Cl_2$  {PN=vitamin B6 pyridoxine[2methyl-3-hydroxy-4,5-bis(hydroxymethyl) pyridine]}. Z Anorg Allg Chem 631(5):878–881
- Bots P, Behrends T (2008) Uranium mobility in subsurface aqueous systems: the influence of redox conditions. Mineral Mag 72(1):381–384
- Branquinho C, Brown DH, Catarino F (1997) The cellular location of Cu in lichens and its effects on membrane integrity and chlorophyll fluorescence. Environ Exp Bot 38(2):165–179
- Brendler V, Geipel G, Nitsche H (1998) Possible impact of phosphate influx on the uranium speciation and migration in seepage waters. Forschungszentrum Rossendorf e.V, Institute of Radiochemistry, Dresden
- Bruins MR, Kapil S, Oehme FW (2000) Microbial resistance to metals in the environment. Ecotoxicol Environ Saf 45(3):198–207
- Bustard M, McHale AP (1997) Biosorption of uranium by cross-linked and alginate immobilized residual biomass from distillery spent wash. Bioprocess Eng 17:127–130
- Cai C, Dong H, Li H, Xiao X, Ou G, Zhang C (2007) Mineralogical and geochemical evidence for coupled bacterial uranium mineralization and hydrocarbon oxidation in the Shashagetai deposit, NW China. Chem Geol 236(1–2):167–179
- Carvalho IG, Cidu R, Fanfani L, Pitsch H, Beaucaire C, Zuddas P (2005) Environmental impact of uranium mining and ore processing in the Lagoa Real District, Bahia, Brazil. Environ Sci Technol 39(22):8646–8652
- Charbonneau M (2009) Current problems and recent advances of in situ biore-mediation of uranium-contaminated sites. MMG 445 Basic. Biotechnology 5:13–18
- Charles AL, Markich SJ, Stauber JL, De Filippis LF (2002a) The effect of water hardness on the toxicity of uranium to a tropical freshwater alga (Chlorella sp.). Aquat Toxicol 60(1–2):61–73
- Charles AL, Markich SJ, Stauber JL, Filippis LFD (2002b) The effect of wawter hardness on the toxicity of uranium to tropical freshwater alga (Chlorella sp.). Aquat Toxicol 60(1–2):61–73
- Charles AL, Markich SJ, Ralph P (2006) Toxicity of uranium and copper individually, and in combination, to a tropical freshwater macrophyte (Lemna aequinoctialis). Chemosphere 62(8):1224– 1233
- Chen B, Roos P, Borggaard OK, Zhu Y-G, Jakobsen I (2005) Mycorrhiza and root hairs in barley enhance acquisition of phosphorus and uranium from phosphate rock but mycorrhiza decreases root to shoot uranium transfer. New Phytol 165 (2):591–598
- Cherdyntsev VV, Ivanovich M, Harmon RS (1982) Uranium series disequilibrium: applications to environmental problems. Clarendon Press, Oxford
- Chiang P-N, Wang MK, Chiu CY, Chou S-Y (2006) Effects of cadmium amendments on low-molecular-weight organic acid exudates in rhizosphere soils of tobacco and sunflower. Environ Toxicol 21 (5):479–488
- Chmielowski J, Woznica A, Klapcinska B, Pytlak M, Piech W (1994) Binding of uranium by yeast cell wall polysaccharides. Bull Pol Acad Sci Biol Sci 42(2):147–149
- Choppin GR (2006) Actinide speciation in aquatic systems. Mar Chem 99(1–4):83–92
- Clara M, Magalhães F (2002) Arsenic. An environmental problem limited by solubility. Pure Appl Chem 74(10):1843–1850
- Cohen CK, Fox TC, Garvin DF, Kochian LV (1998) The role of irondeficiency stress responses in stimulating heavy-metal transport in plants. Plant Physiol 116(3):1063–1072
- Cooley HM, Evans RE, Klaverkamp JF (2000) Toxicology of dietary uranium in lake whitefish (Coregonus clupeaformis). Aquat Toxicol 48(4):495–515
- Creanga DE, Poiata A, Morariu VV, Tupu P (2004) Zero-magnetic field effect in pathogen bacteria. J Magn Magn Mater 272–276 (Part 3):2442–2444
- Curie C, Briat J-F (2003) Iron transport and signaling in plants. Annu Rev Plant Biol 54(1):183–206
- Daković M, Kovačević M, Andjus P, Bačić G (2008) On the mechanism of uranium binding to cell wall of Chara fragilis. Eur Biophys J 37(7):1111–1117
- Dang HS, Pullat VR, Sharma RC (1995) Distribution of uranium in human organs of an urban Indian population and its relationship with clearance half-lives. Heal Phys 68(3):328–331
- Davis JA, Meece DE, Kohler M, Curtis GP (2004) Approaches to surface complexation modeling of uranium(VI) adsorption on aquifer sediments. Geochim Cosmochim Acta 68(18):3621–3641
- Davis JA, Curtis GP, Wilkins MJ, Kohler M, Fox P, Naftz DL, Lloyd JR (2006) Processes affecting transport of uranium in a suboxic aquifer. Phys Chem Earth Parts A/B/C 31(10–14):548–555
- De Schamphelaere KAC, Stubblefield W, Rodriguez P, Vleminckx K, Janssen CR (2010) The chronic toxicity of molybdate to freshwater organisms. I. Generating reliable effects data. Sci Total Environ 408(22):5362–5371
- Diehl P (1995) Uranabbau in Europa—Die Folgen für Mensch und Umwelt. In: BBU Argumente. BBU, Bonn. p. 58
- Diehl P (2003) Uranium Mining in Eastern Germany: the WISMUT Legacy. Vol. 2003, Arnsdorf, Germany: WISE Uranium Project. Available at: <http://www.antenna.nl/wise/uranium/uwis.html>
- Dienemann C, Dienemann H, Dudel GE, Brackhage C, Stolz L (2003) Selektive Akkumulation von Radionuklide und Arsen in naturlichen Feuchtgebieten und künstlichen Fließstrecken. In: Coring E, Bäthe J (eds) der Deutschen Gesellschaft für Limnologie (DGL) und der deuschen und österreichen Section der Societes

<span id="page-22-0"></span>Internationalis Limnologie (SIL), vol. II. Eingenverlag der DGL, Braunschweig, pp 779–784

- DiSpirito AA, Talnagi JW, Tuovinen OH (1983) Accumulation and cellular distribution of uranium in Thiobacillus ferrooxidans. Arch Microbiol 135(4):250–253
- Dodge CJ, Franscis AJ (1997) Biotransformation of binary and ternary citric acid complexes of iron and uranium. Environ Sci Technol 31:3062–3067
- Donnellan N, Rollan A, Mchale L, Mchale AP (1995) The effect of electric field stimulation on the biosorption of uranium by nonliving biomass derived from Kluyveromyces marxianus IMB3. Biotechnol Lett 17(4):439–442
- Doshi SM (2006) Bioremediation of acid mine drainage using sulfatereducing bacteria. Environmental Protection Agency, Washington
- Doudou S, Arumugam K, Vaughan DJ, Livens FR, Burton NA (2011) Investigation of ligand exchange reactions in aqueous uranyl carbonate complexes using computational approaches. Phys Chem Chem Phys 13(23):11402–11411
- Doudou S, Vaughan DJ, Livens FR, Burton NA (2012) Atomistic simulations of calcium uranyl(VI) carbonate adsorption on calcite and stepped-calcite surfaces. Environ Sci Technol 46(14):7587– 7594
- Dudel GE, Brackhage C, Dienemann H, Mkandawire M, Weiske A (2004) Optimising natural attenuation of trace contaminants from uranium mine tailing waters by nature like constructed wetlands. In: Jarvis AP, Dudgeon BA, Younger PL (eds) Mine water process, policy and progress, vol. 2. University of Newcastle upon Tyne Press, Newcastle upon Tyne
- Duff MC, Hunter DB, Bertsch PM, Amrhein C (1999) Factors influencing uranium reduction and solubility in evaporation pond sediments. Biogeochemistry 45(1):95–114
- Duff MC, Coughlin JU, Hunter DB (2002) Uranium co-precipitation with iron oxide minerals. Geochim Cosmochim Acta 66 (20):3533–3547
- Duquène L, Vandenhove H, Tack F, Meers E, Baeten J, Wannijn J (2009) Enhanced phytoextraction of uranium and selected heavy metals by Indian mustard and ryegrass using biodegradable soil amendments. Sci Total Environ 407(5):1496–1505
- Dushenkov S (2003) Trends in phytoremediation of radionuclides. Plant Soil 249(1):167–175
- Dushenkov S, Vasudev D, Kapulnik Y, Doloressa G, Fleisher D, Ting KC, Ensley B (1997) Removal of uranium from Waters using terrestrial plants. Environ Sci Technol 31:3468–3474
- Ebbs SD, Brady DJ, Kochian LV (1998a) Role of uranium speciation in the uptake and translocation of uranium by plants. J Exp Bot 49 (324):1183–1190
- Ebbs, S. D., Brady, D. J., Norvell, W. A. and Kochian, L. V. (1998b). Role of uranium speciation in plant uranium uptake and the phytoremediation of uranium-contaminated soil. In: Membrane transport. American Society of Plant Biologists, Rockville
- Ellen CE (2006) Treatment of uranium-contaminated waters using organic-based permeable reactive barriers. Fed Facil Environ J 17(1):19–35
- El-Nadi YA, El-Hefny NE, Daoud JA (2003) Mechanism of extraction of hexavalent uranium from alkaline medium by aliquat-336/ kerosene solution. J Nucl Radiochem Sci 4(2):19–22
- Enderle GJ, Friedrich K (1995) East German uranium miners (Wismut): exposure conditions and health consequences. Stem Cells 13(suppl 1):78–89
- Erdem E, Tinkilic N, Yilmaz VT, Uyanik A, Olmez H (1995) Distribution of uranium in the production of triple superphosphate (TSP) fertilizer and phosphoric acid. Fertil Res 44(2):129–131
- Finneran KT, Housewright ME, Lovley DR (2002) Multiple influences of nitrate on uranium solubility during bioremediation of uranium-contaminated subsurface sediments. Environ Microbiol 4(9):510–516

 $\textcircled{2}$  Springer

- Fowler D, Smith RI, Muller JB, Hayman G, Vincent KJ (2005) Changes in the atmospheric deposition of acidifying compounds in the UK between 1986 and 2001. Environ Pollut 137(1):15–25
- Fox PM, Davis JA, Zachara JM (2006) The effect of calcium on aqueous uranium(VI) speciation and adsorption to ferrihydrite and quartz. Geochim Cosmochim Acta 70(6):1379–1387
- Francis AJ, Dodge CJ (1998) Remediation of soils and wastes contaminated with uranium and toxic metals. Environ Sci Technol 32 (24):3993–3998
- Franklin NM, Stauber JL, Markich SJ, Lim RP (2000) pH-dependent toxicity of copper and uranium to a tropical freshwater alga (Chlorella sp.). Aquat Toxicol 48(2–3):275–289
- Fredrickson JK, Kostandarithes HM, Li SW, Plymale AE, Daly MJ (2000a) Reduction of Fe(III), Cr(VI), U(VI), and Tc(VII) by Deinococcus radiodurans R1. Appl Environ Microbiol 66 (5):2006–2011
- Fredrickson JK, Zachara JM, Kennedy DW, Duff MC, Gorby YA, Li S-M, Li S-MW, Krupka KM (2000b) Reduction of U(VI) in goethite ([alpha]-FeOOH) suspensions by a dissimilatory metal-reducing bacterium. Geochim Cosmochim Acta 64(18):3085–3098
- Fresquez PR, Armstrong DR, Mullen MA, Naranjo L (1998) The uptake of radionuclides by beans, squash, and corn growing in contaminated alluvial soils at Los Alamos National Laboratory. J Environ Sci Health B 33(1):99–121
- Fritzsche A, Dienemann H, Dudel E (2006) Arsenic fixation on ironhydroxide-rich and plant litter-containing sediments in natural environments. Environ Geol 51(1):133–142
- Fuller CC, Barger JR, Davis JA, Piana MJ (2002) Mechanisms of uranium interactions with hydroxyapatite: implications for groundwater remediation. Environ Sci Technol 36(2002):158–165
- Gadelle F, Wan J, Tokunaga TK (2001) Removal of uranium(VI) from contaminated sediments by surfactants. J Environ Qual 30 (2):470–478
- Gavshin VM, Melgunov MS, Sukhorukov FV, Bobrov VA, Kalugin IA, Klerkx J (2005) Disequilibrium between uranium and its progeny in the Lake Issyk-Kul system (Kyrgyzstan) under a combined effect of natural and manmade processes. J Environ Radioact 83(1):61–74
- Geissler A, Merroun M, Geipel G, Reuther H, Selenska-Pobell S (2009) Biogeochemical changes induced in uranium mining waste pile samples by uranyl nitrate treatments under anaerobic conditions. Geobiology 7(3):282–294
- Giannardi C, Dominici D (2003) Depleted uranium, the longterm effects of the US bombing of Iraq. Ponte 59(3–4):227–238
- Ginder-Vogel M, Wu W-M, Carley J, Jardine P, Fendorf S, Criddle C (2006) In situ biological uranium remediation within a highly contaminated aquifer. SSRL Science Highlights. Available at: [http://www.ssrl.slac.stanford.edu/research/highlights\\_archive/u\\_](http://www.ssrl.slac.stanford.edu/research/highlights_archive/u_remed.html) [remed.html](http://www.ssrl.slac.stanford.edu/research/highlights_archive/u_remed.html)
- Giri S, Singh G, Jha VN, Tripathi RM (2010) Natural radionuclides in fish species from surface water of Bagjata and Banduhurang uranium mining areas, East Singhbhum, Jharkhand, India. Int J Radiat Biol 86(11):946–956
- Gomez P, Garralon A, Buil B, Turrero MJ, Sanchez L, de la Cruz B (2006) Modeling of geochemical processes related to uranium mobilization in the groundwater of a uranium mine. Sci Total Environ 366(1):295–309
- Gonzalez-Munoz MT, Merroun ML, BenOmar N, Arias JM (1997) Biosorption of uranium by Myxococcus xanthus. Int Biodeterior Biodegrad 40(2–4):107–114
- Haas JR, Bailey EH, Purvis OW (1998) Bioaccumulation of metals by lichens: uptake of aqueous uranium by Peltigera membranacea as a function of time and pH. Am Mineral 83:1494–1502
- Hallberg KB, Johnson DB (2003) Passive mine water treatment at the former Wheal Jane tin mine, Cornwall: important biogeochemical and microbiological lessons. Land Contam Reclam 11(2):213–220
- <span id="page-23-0"></span>Henge Napoli MH, Archimbaud M, Ansoborlo E, Metivier H, Gourmelon P (1995) Efficacy of 3,4,3-LIHOPO for reducing the retention of uranium in rat after acute administration. Int J Radiat Biol 68(4):389–393
- Hiess J, Condon DJ, McLean N, Noble SR (2012) <sup>238</sup>U/<sup>235</sup>U Systematics in terrestrial uranium-bearing minerals. Science 335 (6076):1610–1614
- Hoffmann D (2000) In the laboratory of the planned economy—from labor allocation to labor recruitment in the Russian Zone of Occupation/German Democratic Republic (1945–1961). Vierteljahrsh Zeitgesch 48(4):531–666
- Hoffmann W, Kranefeld A, Schmitz-Feuerhake I (1993) Radium-226 contaminated drinking water: hypothesis on an exposure pathway in a population with elevated childhood leukemia. Environ Heal Perspect 101(Suppl 3):113–115
- Hogan AC, van Dam RA, Markich SJ, Camilleri C (2005) Chronic toxicity of uranium to a tropical green alga (Chlorella sp.) in natural waters and the influence of dissolved organic carbon. Aquat Toxicol 75(4):343–353
- Hole LR, Brunner SH, Hanssen JE, Zhang L (2008) Low cost measurements of nitrogen and sulphur dry deposition velocities at a semialpine site: gradient measurements and a comparison with deposition model estimates. Environ Pollut 154(3):473–481
- Hsieh KM, Murgel GA, Lion LW, Shuler ML (1994) Interactions of microbial biofilms with toxic trace metals: 1. Observation and modeling of cell growth, attachment, and production of extracellular polymer. Biotechnol Bioeng 44(2):219–231
- IAEA (1998) Guidebook on good practice in the management of uranium mining and mill operations and the preparation for their closure. International Atomic Energy Agency, Vienna, p 81
- IAEA (1999). Technologies for the treatment of effluents from uranium mines, mills and tailings—part 1. In: Technical Committee. International Atomic Energy Agency, Vienna. p. 14
- IAEA (2001) Manual of acid in situ leach uranium mining technology. International Atomic Energy Agency, Vienna, p 294
- IAEA (2007) Management of reprocessed uranium current status and future prospects. Nuclear fuel cycle and materials section, international atomic energy agency, Vienna, p 108
- Ivanovich M, Harmon RS (1992) Uranium-series disequilibrium: applications to earth, marine, and environmental science. Clarendon Press, Oxford
- Jeffries DS, Brydges TG, Dillon PJ, Keller W (2003) Monitoring the results of Canada/U.S.A. acid rain control programs: some lake responses. Environ Monit Assess 88(1–3):3–19
- Jerden JJL, Sinha AK (2006) Geochemical coupling of uranium and phosphorous in soils overlying an unmined uranium deposit: Coles Hill, Virginia. J Geochem Explor 91(1–3):56–70
- Jerden JL, Sinha AK, Zelazny L (2003) Natural immobilization of uranium by phosphate mineralization in an oxidizing saprolitesoil profile: chemical weathering of the Coles Hill uranium deposit, Virginia. Chem Geol 199(1–2):129–157
- Jin CW, He YF, Tang CX, Wu P, Zheng SJ (2006) Mechanisms of microbially enhanced Fe acquisition in red clover (Trifolium pratense L.). Plant Cell Environ 29(5):888–897
- Juznic K, Koponen MH, Juznic A (1989) Transfer of <sup>210</sup>Pb and <sup>210</sup>Po to the environment during mining and processing of uranium ores. J Radioanal Nucl Chem Lett 135(4):293–298
- Kalin M, Wheeler WN, Meinrath G (2005a) The removal of uranium from mining waste water using algal/microbial biomass. J Environ Radioact 78:151–177
- Kalin M, Wheeler WN, Meinrath G (2005b) The removal of uranium from mining waste water using algal/microbial biomass. J Environ Radioact 78(2):151–177
- Kaplan D, Prakash NT, Abeliovich A (1998) Glutathione-induced recovery in Chlorella cells from metal toxicity. Fresenius Environ Bull 7(3–4):153–159
- Kerisit S, Liu C (2012) Diffusion and adsorption of uranyl carbonate species in nanosized mineral fractures. Environ Sci Technol 46 (3):1632–1640
- Ketterer ME, Wetzel WC, Layman RR, Matisoff G, Bonniwell EC (2000) Isotopic studies of source of uranium in sediments of Ashtabula River, Ohio, USA. Environ Sci Technol 34:966–972
- Kim KW, Lee KY, Chung DY, Lee EH, Moon JK, Shin DW (2012) Evaluation of the stability of uranyl peroxo-carbonato complex ions in carbonate media at different temperatures. J Hazard Mater 233–234:213–218
- Klapheck S, Schlunz S, Bergmann L (1995) Synthesis of phytochelatins and homo-phytochelatins in Pisum sativum L. Plant Physiol 107(2):515–521
- Kleykamp H (1991) Thermodynamics of the uranium–platinum metals systems. Pure App Chem 63(10):1401–1408
- Koike H, Yoneyama K, Kashino Y, Satoh K (1996) Mechanism of electron flow through the QB site in photosystem II. 4. Reaction mechanism of plastoquinone derivatives at the QB Site in spinach photosystem II membrane fragments. Plant Cell Physiol 37 (7):983–988
- Kryscio A, Müller WUU, Wojcik A, Kotschy N, Grobelny S, Streffer C (2001) A cytogenetic analysis of the long-term effect of uranium mining on peripheral lymphocytes using the micronucleuscentromere assay. Int J Radiat Biol 77(11):1087–1193
- Kusiak RA, Ritchie AC, Muller J, Springer J (1993) Mortality from lung cancer in Ontario uranium miners. B J Ind Med 50(10):920– 928
- Lack JG, Chaudhuri SK, Kelly SD, Kemner KM, O'Connor SM, Coates JD (2002) Immobilization of radionuclides and heavy metals through anaerobic bio-oxidation of Fe(II). Appl Environ Microbiol 68(6):2704–2710
- Landa ER (2004) Uranium mill tailings: nuclear waste and natural laboratory for geochemical and radioecological investigations. J Environ Radioact 77(1):1–27
- Levy DB, Kearney WF (1999) Irrigation of native rangeland using treated wastewater from in situ uranium processing. J Environ Qual 28(1):208–217
- Liger E, Charlet L, Van Cappellen P (1999) Surface catalysis of uranium(VI) reduction by iron(II). Geochim Cosmochim Acta 63(19–20):2939–2955
- Liu Y, Xu H (2007) Equilibrium, thermodynamics and mechanisms of  $Ni<sup>2+</sup>$ biosorption by aerobic granules. Biochem Eng J 35(2):174–182
- Lloyd JR, Lovley DR (2001) Microbial detoxification of metals and radionuclides. Curr Opin Biotechnol 12:248–253
- Lloyd JR, Renshaw JC (2005) Bioremediation of radioactive waste: radionuclide–microbe interactions in laboratory and field-scale studies. Curr Opin Biotechnol 16(3):254–260
- Lloyd RD, Polig E, Taylor GN, Bruenger FW, Miller SC (1996) Uranium skeletal dosimetry and distribution in young adult beagles: a guide for calculating uranium skeletal doses in humans. Heal Phys 70(3):396–401
- Lovley DR (1995) Bioremediation of organic and metal contaminants with dissimilatory metal reduction. J Ind Microbiol 14(2):85–93
- Lovley DR (2003) Cleaning up with genomics: appling molecular biology to bioremediation. Nat Rev 1:35–44
- Lovley DR (2008) Extracellular electron transfer extracellular electron transfer: wires, capacitors, iron lungs, and more. Geobiology 5:225–231
- Lovley DR, Widman PK, Woodward JC, Phillips EJP (1993) Reduction of uranium by cytochrome C3 of Desulfovibrio vulgaris. Appl Environ Microbiol 59(11):3572–3576
- Lüderitz V, Gerlach F (2002) Phosphorus removal in different constructed wetlands. Acta Biotechnol 22(1–2):91–99
- Luo W, Gu B (2011) Dissolution of uranium-bearing minerals and mobilization of uranium by organic ligands in a biologically reduced sediment. Environ Sci Technol 45(7):2994–2999
- <span id="page-24-0"></span>Luo J, Weber F-A, Cirpka OA, Wu W-M, Nyman JL, Carley J, Jardine PM, Criddle CS, Kitanidis PK (2007) Modeling in-situ uranium (VI) bioreduction by sulfate-reducing bacteria. J Contam Hydrol 92(1–2):129–148
- Ma JF (2000) Mini review role of organic acids in detoxification of aluminum in higher plants. Plant Cell Physiol 41(4):383–390
- Macaskie LE, Basnakova G (1998) Microbially enhanced chemisorption of heavy metals: a method for the bioremediation of solutions containing long lived isotopes of neptunium and plutonium. Environ Sci Technol 32(1):184–187
- Macaskie LE, Bonthrone KM, Yong P, Goddard DT (2000) Enzymically mediated bioprecipitation of uranium by a Citrobacter sp.: a concerted role for exocellular lipopolysaccharide and associated phosphatase in biomineral formation. Microbiology 146(8):1855–1867
- Madden AS, Smith AC, Balkwill DL, Fagan LA, Phelps TJ (2007) Microbial uranium immobilization independent of nitrate reduction. Environ Microbiol 9(9):2321–2330
- Maksymiec W (1997) Effect of copper on cellular processes in higher plants. Photosynthetica 34(3):321–342
- Malekzadeh F, Latifi AM, Shahamat M, Levin M, Colwell RR (2002) Effects of selected physical and chemical parameters on uranium uptake by the bacterium Chryseomonas MGF-48. World J Microbiol Biotechnol 18(7):599–602
- Markich SJ (2002) Uranium speciation and bioavailability in aquatic systems: an overview. Scientific World Journal 2:707–729
- Martinez Aguirre A, Garcia Leon M, Ivanovich M (1995) U and Th speciation in river sediments. Sci Total Environ 173–174(1– 6):203–209
- Martinoia E, Maeshima M, Neuhaus HE (2007) Vacuolar transporters and their essential role in plant metabolism. J Exp Bot 58(1):83– 102
- Mazen AMA, Zhang D, Franceschi VR (2003) Calcium oxalate formation in Lemna minor: physiological and ultrastructural aspects of high capacity calcium sequestration. New Phytol 161:435–448
- Meinrath G (1996) Coordination of uranyl (VI) carbonate complexation in aqueous solution. J Radioanal Nucl Chem 211:349–362
- Meinrath G (1997) Uranium speciation by spectroscopy. J Radioanal Nucl Chem 224:119
- Meinrath G (1998) Aquatic chemistry of uranium: a review focusing on aspects of environmental chemistry. In: Freiberg On-line Geosciences Freiberg
- Meinrath G (1999) Aquatic Chemistry of Uranium: A review focusing on aspects of environmental chemistry. Freib On-line Geosci 1:1– 112
- Meinrath G, Volke P, Helling C, Dudel E-G, Merkel BJ (1999) Determination and interpretation of environmental water samples contaminated by uranium mining activities. Fresenius J Anal Chem 364:191–202
- Meinrath A, Schneider P, Meinrath G (2003) Uranium ore and depleted uranium in the environment, with reference to uranium in the biosphere from Erzgebirge/Sachsen, Germany. J Environ Radioact 64:175–193
- Miller RC, Marino SA, Brenner DJ, Martin SG, Richards M, Randers Pehrson G, Hall EJ (1995) The biological effectiveness of radonprogeny alpha particles: II. Oncogenic transformation as a function of linear energy transfer. Radiat Res 142(1):54–60
- Miller AC, Blakely WF, Livengood D, Whittaker T, Xu JQ, Ejnik JW, Hamilton MM, Parlette E, St John T, Gerstenberg HM, Hsu H (1998) Transformation of human osteoblast cells to the tumorigenic phenotype by depleted uranium uranyl chloride. Environ Heal Perspect 106(8):465–471
- Milodowski AE, West JM, Pearce JM, Hyslop EK, Basham IR, Hooker PJ (1990) Uranium-mineralized micro-organisms associated with uraniferous hydrocarbons in southwest Scotland. Nature 347:465– 467
- $\textcircled{2}$  Springer
- Min M, Xu H, Chen J, Fayek M (2005) Evidence of uranium biomineralization in sandstone-hosted roll-front uranium deposits, northwestern China. Ore Geol Rev 26(3–4):198–206
- Misaelides P, Godelitsas A, Filippidis A, Charistos D, Anousis I (1995) Thorium and uranium uptake by natural zeolitic materials. Sci Total Environ 173–174(1–6):237–246
- Mishra SR, Bharati K, Sethunathan N, Adhya TK (1999) Effects of heavy metals on methane production in tropical rice soils. Ecotoxicol Environ Saf 44(1):129–136
- Mitrovic SM, Allis O, Furey A, James KJ (2005) Bioaccumulation and harmful effects of microcystin-LR in the aquatic plants *Lemna* minor and Wolffia arrhiza and the filamentous alga Chladophora fracta. Ecotoxicol Environ Saf 61(3):345–352
- Mkandawire M (2005) Fate and effect of uranium and arsenic in surface waters of abandoned uranium mines: studies with Lemna gibba L. Shaker, Aachen
- Mkandawire M, Dudel EG (2005) Accumulation of arsenic in Lemna gibba L. (duckweed) in tailing waters of two abandoned uranium mining sites in Saxony, Germany. Sci Total Environ 336(1–3):81–89
- Mkandawire M, Taubert B, Dudel EG (2003) Biosorption capacity and limits in uranium immobilisation by dead and living Lemna gibba L. biomass. In: Kalogerakis N (ed) Second European Bioremediation Conference, vol 2. Technical University of Crete and European Community (High-Level Scientific Conference Programme), Crete, pp 357–360
- Mkandawire M, Dudel EG, Taubert B (2004a) Accumulation of uranium in Lemna gibba L. in relation to milieu conditions of tailing waters in abandoned uranium mines in Germany. In: Jarvis AP, Dudgeon BA, Younger PL (eds) Mine water—process, policy and progress, vol. 2. University of Newcastle upon Tyne Press, Newcastle upon Tyne, pp 9–18
- Mkandawire M, Taubert B, Dudel EG (2004b) Capacity of Lemna gibba L. (Duckweed) for uranium and arsenic phytoremediation in mine tailing waters. Int J Phytoremediat 6(4):347–362
- Mkandawire M, Dudel EG, Müller C (2005) Possible biomineralisation of uranium in Lemna gibba G3. In: Merkel BJ, Hasche-Berger A (eds) Uranium in the environment. Springer, Berlin, pp 495–505
- Mkandawire M, Pompe W, Dudel EG (2006a) Prediction of uranium (VI) toxicity to Lemna gibba L. G3 using ion and ligand characteristics. In: Coring E, Bäthe J (eds) der Deutschen Gesellschaft für Limnologie (DGL) und Societas Internationalis Limnologie (SIL). der Deutschen Gesellschaft für Limnologie (DGL), Dresden, pp 124–125
- Mkandawire M, Taubert B, Dudel EG (2006b) Limitation of growth parameters in Lemna gibba bioassay for arsenic and uranium under variable phosphate availability. Ecotoxicol Environ Saf 65:118–128
- Mkandawire M, Vogel K, Taubert B, Dudel EG (2007) Phosphate regulates uranium (VI) toxicity to Lemna gibba L. G3. Environ Toxicol 22(1):9–16
- Morgenstein ME (2003) Uranium-series and luminescence dating of volcanic lithic artefacts. Archaeometry 45:503–518
- Morris S, Meinhold AF (1995) Probabilistic risk assessment of nephrotoxic effect of uranium in drinking water. Heal Phys 69(6):897–908
- Moyes LN, Parkman RH, Charnock JM, Vaughan DJ, Livens FR, Hughes CR, Braithwaite A (2000) Uranium uptake from aquous solution by intrasction with geothite, lepidocrocite, muscoviote and macknawite: an X-ray absorption spectroscopy study. Environ Sci Technol 34:1062–1068
- Müller B, Müller B, Granina L, Schaller T, Ulrich A, Wehrli B (2002) P, As, Sb, Mo, and other elements in sedimentary Fe/Mn layers of Lake Baikal. Environ Sci Technol 36:411–420
- Müller W-U, Kryscio A, Streffer C (2004) Micronuclei in lymphocytes of uranium miners of the former Wismut SDAG. Cytogenet Genome Res 104:295–298
- <span id="page-25-0"></span>Munoz JA, Gonzalo A, Valiente M (2002) Arsenic adsorption by Fe (III)-loaded open-celled cellulose sponge. Thermodynamic and selectivity aspects. Environ Sci Technol 36:3405–3411
- Murakami T, Ohnuki T, Isobe H, Sato T (1997) Mobility of uranium during weathering. Am Mineral 82:888-899
- Nagarkatti M, Nagarkatti PS, Brooks A (1996) Effect of radon on the immune system: alterations in the cellularity and functions of Tcells in lymphoid organs of mouse. J Toxicol Environ Heal 47  $(6)$ :535–552
- Neal AL, Amonette JE, Peyton BM, Geesey GG (2004) Uranium complexes formed at hematite surfaces colonized by sulfatereducing bacteria. Environ Sci Technol 38:3019–3027
- Neumann D, Zurnieden U, Lichtenberger O, Leopold I (1995) How does Armeria maritima tolerate high heavy metal concentrations? J Plant Physiol 146(5–6):704–717
- O'Flaherty EJ (1995) PBK modeling for metals: examples with lead, uranium, and chromium. Toxicol Lett 82–83:367–372, Special issue
- Ohnuki T, Kozai N, Samadfam M, Yasuda R, Yamamoto S, Narumi K, Naramoto H, Murakami T (2004) The formation of autunite (Ca  $(UO<sub>2</sub>)2(PO<sub>4</sub>)2[square root]nH<sub>2</sub>O)$  within the leached layer of dissolving apatite: incorporation mechanism of uranium by apatite. Chem Geol 211(1–2):1–14
- Ortiz-Bernad I, Anderson RT, Vrionis HA, Lovley DR (2004) Resistance of solid-phase U(VI) to microbial reduction during in situ bioremediation of uranium-contaminated groundwater. Appl Environ Microbiol 70(12):7558–7560
- Overall RA, Parry DL (2004) The uptake of uranium by Eleocharis dulcis (Chinese water chestnut) in the ranger uranium mine constructed wetland filter. Environ Pollut 132(2):307–320
- Periyakaruppan A, Kumar F, Sarkar S, Sharma C, Ramesh G (2007) Uranium induces oxidative stress in lung epithelial cells. Arch Toxicol 81(6):389–395
- Peterson J, MacDonell M, Haroun L, Monette F, Hildebrand R D, Taboas A (2007) Radiological and Chemical fact sheets to support health risk analyses for contaminated areas. Vol. Human health fact sheet, 4 (Ed A. N. Laboratory). Argonne National Laboratory, EVS in collaboration with U.S. Department of Energy
- Petrescu L, Bilal E (2006) Speciation of natural uranium and thorium in surface waters around a uranium mine (Bistrita Mts., Romania). Geochim Cosmochim Acta Geochimica et Cosmochimica Acta 70(18, Supplement 1):A486
- Pettersson HBL, Koperski J (1991) Investigation of aerial dispersion of radioactive dust from an open-pit uranium mine by passive vinyl collectors. Heal Phys 60(5):681–690
- Piao CQ, Hei TK (1993) The biological effectiveness of radon daughter alpha particles. I. Radon, cigarette smoke and oncogenic transformation. Carcinogenesis 14(3):497–501
- Pilon-Smits E (2005) Phytoremediation. Annu Rev Plant Biol 56:15– 39
- Pollmann K, Raff J, Merroun M, Fahmy K, Selenska-Pobell S (2006) Metal binding by bacteria from uranium mining waste piles and its technological applications. Biotechnol Adv 24  $(1):58-68$
- Premuzic ET, Francis AJ, Lin M, Schubert J (1985) Induced formation of chelating agents by Pseudomonas aeruginosa grown in presence of thorium and uranium. Arch Environ Contam Toxicol 14 (6):759–768
- Pressyanov DS, Guelev MG, Sharkov BG (1995) Radon and radon progeny outdoors in a valley of enhanced natural radioactivity. Atmos Environ 29(23):3433–3439
- Prikryl JD, Jain A, Turner DR, Pabalan RT (2001) UraniumVI sorption behavior on silicate mineral mixtures. J Contam Hydrol 47(2– 4):241–253
- Raicevic S, Wright JV, Veljkovic V, Conca JL (2006) Theoretical stability assessment of uranyl phosphates and apatites: selection

of amendments for in situ remediation of uranium. Sci Total Environ 355(1–3):13–24

- Rawlings DE, Silver S (1995) Mining with microbes. Biotechnology 13(8):773–778
- Riley SJ (1995) Geomorphic estimates of the stability of a uranium mill tailings containment cover: Nabarlek, Northern Territory, Australia. Land Degrad Rehab 6(1):1–16
- Rittle KA, Drever JI, Colberg PJS (1995) Precipitation of arsenic during bacterial sulfate reduction. Geomicrobiol J 13(1):1–11
- Rodgher S, de Azevedo H, Ferrari C R, Roque C V, Ronqui L B, de Campos M B, Nascimento M R (2012) Evaluation of surface water quality in aquatic bodies under the influence of uranium mining (MG, Brazil). Environ Monit Assess. doi[:10.1007/](http://dx.doi.org/10.1007/s10661-012-2719-5) [s10661-012-2719-5](http://dx.doi.org/10.1007/s10661-012-2719-5)
- Rodriguez PB, Tome FV, Fernandez MP, Lozano JC (2006) Linearity assumption in soil-to-plant transfer factors of natural uranium and radium in Helianthus annuus L. Sci Total Environ 361(1–3):1–7
- Roussel C, Bril H, Fernandez A (2000) Arsenic speciation: involvement in evaluation of environmental impact caused by mine wastes. J Environ Qual 29(1):182–188
- Russell JJ, Kathren RL, Dietert SE (1996) A histological kidney study of uranium and non-uranium workers. Heal Phys 70(4):466–472
- Rybicka KK (1996) Glycosomes—the organelles of glycogen metabolism. Tissue Cell 28(3):253–265
- Salbu B (2013) Preface: uranium mining legacy issue in Central Asia. J Environ Radioact. doi[:10.1016/j.jenvrad.2011.12.010](http://dx.doi.org/10.1016/j.jenvrad.2011.12.010)
- Salt DE, Blaylock M, Kumar NPBA, Dushenkov V, Ensley BD, Chet I, Raskin I (1995) Phytoremediation: a novel strategy for the removal of toxic metals from the environment using plants. Biotechnology 13(5):468–474
- Samardakiewicz S, Wozny A (2000) The distribution of lead in duckweed (Lemna minor L.) root tip. Plant Soil 226(1):107–111
- Sar P, D'Souza SF (2001) Biosorptive uranium uptake by a Pseudomonas strain: characterization and equilibrium studies. J Chem Technol Biotechnol 76(12):1286–1294
- Saric MR, Stojanovic M, Babic M (1995) Uranium in plant species grown on natural barren soil. J Plant Nutr 18(7):1509–1518
- Schaller J, Weiske A, Mkandawire M, Dudel EG (2008) Enrichment of uranium in particulate matter during litter decomposition affected by Gammarus pulex L. Environ Sci Technol 42(23):8721–8726
- Schmeide K, Sachs S, Bubner M, Reich T, Heise KH, Bernhard G (2003) Interaction of uranium(VI) with various modified and unmodified natural and synthetic humic substances studied by EXAFS and FTIR spectroscopy. Inorg Chim Acta 351:133–140
- Schnoor JL, Licht LA, McCutcheon SC, Wolfe NL, Carriera LH (1995) Phytoremediation: an emerging technology for contaminated sites. Department of Civil and Environmental Engineering of the University of Iowa and Environmental Research Laboratory of the EPA, Athens
- Schuttmann W (1993) Schneeberg lung disease and uranium mining in the Saxon Ore Mountains (Erzgebirge). Am J Ind Med 23(2):355–368
- Seidel H, Mattusch J, Wennrich R, Morgenstern P, Ondruschka J (2002) Mobilization of arsenic and heavy metals from contaminated sediments by changing the environmental conditions. Acta Biotechnologica 22(1–2):153–160
- Selig U, Hübener T, Michalik M (2002) Dissolved and particulate phosphorus forms in a eutrophic shallow lake. Aquat Sci 64:97–105
- SENES Consultants Limited. and Alberta. Alberta Environment (2008) Environmental impacts of different uranium mining processes. Alberta Environment, Edmonton
- Shah K, Dubey RS (1998) A 18 kDa cadmium inducible protein complex: its isolation and characterisation from rice (Oryza sativa L.) seedlings. J Plant Physiol 152(4–5):448–454
- Shanbhag PM, Chopping GR (1981) Binding of uranyl by humic acid. J Inorg Nucl Chem 43(12):3369–3372
- <span id="page-26-0"></span>Sheoran AS, Sheoran V (2006) Heavy metal removal mechanism of acid mine drainage in wetlands: a critical review. Miner Eng 19 (2):105–116
- Sheppard SC, Sheppard MI, Gallerand M-O, Sanipelli B (2005) Derivation of ecotoxicity thresholds for uranium. J Environ Radioact 79(1):55–83
- Shilov V, Yusov A, Fedoseev A (2007) Disproportionation of nonoxygenated U(V) bound in the complex with heteropoly anions P2W17O 6110−. Radiochemistry 49(4):361–366
- Shumilin E, Rodriguez-Figueroa G, Sapozhnikov D, Sapozhnikov Y, Choumiline K (2012) Anthropogenic and authigenic uranium in marine sediments of the Central Gulf of California adjacent to the Santa Rosalia mining region. Arch Environ Contam Toxicol 63 (3):309–322
- Shvareva TY, Albrecht-Schmitt TE (2006) General route to threedimensional framework uranyl transition metal phosphates with atypical structural motifs: the case examples of  $Cs<sub>2</sub>$  {(UO<sub>2</sub>)<sub>4</sub>[Co  $(H_2O)_2]^2$ (HPO<sub>4</sub>)(PO<sub>4</sub>)<sub>4</sub>) and Cs<sup>3+</sup>x[(UO<sub>2</sub>)<sub>3</sub>CuH<sup>4-</sup>x(PO<sub>4</sub>)<sub>5</sub>]·H<sub>2</sub>O. Inorg Chem 45(5):1900–1902
- Simon O, Garnier-Laplace J (2005) Laboratory and field assessment of uranium trophic transfer efficiency in the crayfish Orconectes limosus fed the bivalve C. fluminea. Aquat Toxicol 74(4):372–383
- Simpson SL, Angel BM, Jolley DF (2004) Metal equilibration in laboratory-contaminated (spiked) sediments used for the development of whole-sediment toxicity tests. Chemosphere 54(5):597– 609
- Sims R, Lawless TA, Alexander JL, Bennett DG, Read D (1996) Uranium migration through intact sandstone: effect of pollutant concentration and the reversibility of uptake. J Contam Hydrol 21 (1–4):215–228
- Slaveykova V, Wilkinson KJ (2002) Physicochemical aspects of lead bioaccumulation by Chlorella vulgaris. Environ Sci Technol 36:969–975
- Slaveykova V, Wilkinson K (2003) Effect of pH on Pb biouptake by the freshwater alga Chlorella kesslerii. Environ Chem Lett 1 (3):185–189
- Smeaton CM, Weisener CG, Burns PC, Fryer BJ, Fowle DA (2008) Bacterially enhanced dissolution of meta-autunite. Am Mineral 93 (11–12):1858–1864
- Spear JR, Figuaroa LA, Honeyman B (1999) Modeling the removal of uranium U(VI) from aqueous solutions in the presence of sulfate reducing bacteria. Environ Sci Technol 33:2667–2675
- Sram RJ, Binkova B, Dobias L, Rossner P, Topinka J, Vesela D, Vesely D, Stejskalova J, Bavorova H, Rericha V (1993) Monitoring genotoxic exposure in uranium miners. Environ Heal Perspect 99:303–305
- Stearns DM, Yazzie M, Bradley AS, Coryell VH, Shelley JT, Ashby A, Asplund CS, Lantz RC (2005) Uranyl acetate induces hprt mutations and uranium-DNA adducts in Chinese hamster ovary EM9 cells. Mutagenesis 20(6):417–423
- Steele H, Taylor RJ (2007) A theoretical study of the inner-sphere disproportionation reaction mechanism of the pentavalent actinyl ions. Inorg Chem 46(16):6311–6318
- Stefaniak EA, Alsecz A, Frost R, Mathe Z, Sajo IE, Torok S, Worobiec A, Van Grieken R (2009) Combined SEM/EDX and micro-Raman spectroscopy analysis of uranium minerals from a former uranium mine. J Hazard Mater 168(1):416–423
- Takashima T, Fukunishi N, Nishiki T, Konishi Y (2002) Anaerobic oxidation of dissolved hydrogen sulfide in continuous culture of the chemoautotrophic bacterium Thiobacillus denitrificans. Kagaku Kogaku Ronbunshu 28(1):25–30
- Thomas RAP, Macaskie LE (1998) The effect of growth conditions on the biodegradation of tributyl phosphate and potential for the remediation of acid mine drainage waters by a naturallyoccurring mixed microbial culture. Appl Microbiol Biotechnol 49(2):202–209

Thompson T (2011) Uranium mining. Greenhaven Press, Detroit

- Tomasek L, Darby SC, Swerdlow AJ, Placek V, Kunz E (1993) Radon exposure and cancers other than lung cancer among uranium miners in West Bohemia [see comments]. Lancet 341 (8850):919–923
- Trontelj A, Ponikvar-Zorko P (1998) Influence of a uranium mine on the macrozoobenthic communities of the streams in the nearest environs, Slovenia. Water Sci Technol 37(8):235–241
- Tsuruta T (2006) Bioaccumulation of uranium and thorium from the solution containing both elements using various microorganisms. J Alloys Compd 408–412:1312–1315
- Ulrich K-U, Rossberg A, Foerstendorf H, Zanker H, Scheinost AC (2006) Molecular characterization of uranium(VI) sorption complexes on iron(III)-rich acid mine water colloids. Geochim Cosmochim Acta 70(22):5469–5487
- Van Assche F, Clijsters H (1990) Effects of metals on enzyme activity in plants. Plant Cell Environ 13:195–206
- Van Horn JD, Huang H (2006) Uranium(VI) bio-coordination chemistry from biochemical, solution and protein structural data. Coord Chem Rev 250(7–8):765–775
- Vandenhove H, Van Hees M, Wannijn J, Wouters K, Wang L (2007) Can we predict uranium bioavailability based on soil parameters? Part 2: soil solution uranium concentration is not a good bioavailability index. Environ Pollut 145(2):577–586
- Vanhoudt N, Vandenhove H, Smeets K, Remans T, Van Hees M, Wannijn J, Vangronsveld J, Cuypers A (2008) Effects of uranium and phosphate concentrations on oxidative stress related responses induced in Arabidopsis thaliana. Plant Physiol Biochem 46(11):987–996
- Vauclare P, Kopriva S, Fell D, Suter M, Sticher L, von Ballmoos P, Krahenbuhl U, den Camp RO, Brunold C (2002) Flux control of sulphate assimilation in Arabidopsis thaliana: adenosine 5'-phosphosulphate reductase is more susceptible than ATP sulphurylase to negative control by thiols. Plant J 31(6):729–740
- Veiga LHS, Amaral ECS, Fernandes HM (1998) Human health risk screening of radioactive and nonradioactive contaminants due to uranium industry operation. J Environ Radioact 39(1):69–85
- Vinay Kumar Reddy K, Gopal Reddy C, Vidya Sagar D, Yadagiri Reddy P, Rama Reddy K (2012) Environmental radioactivity studies in the proposed Lambapur and Peddagattu uranium mining areas of Andhra Pradesh, India. Radiat Prot Dosimetry 151 (2):290–298
- Virk HS, Sharma N, Bajwa BS (1999) Environmental radioactivity: a case study in Himachal Pradesh, India. J Environ Radioact 45 (2):119–127
- Volesky BA, May Phillips HA (1995) Biosorption of heavy metals by Saccharomyces cerevisiae. Appl Microbiol Biotechnol 42 (5):797–806
- Vrionis HA, Anderson RT, Ortiz-Bernad I, O'Neill KR, Resch CT, Peacock AD, Dayvault R, White DC, Long PE, Lovley DR (2005) Microbiological and geochemical heterogeneity in an in situ uranium bioremediation field site. Appl Environ Microbiol 71 (10):6308–6318
- Vymazal J (ed) (2005) Natural and constructed wetlands: nutrient, metal and management. Backhuys Publishers, Prague
- Wall JD, Krumholz LR (2006) Uranium reduction. Annu Rev Microbiol 60:149–166
- Wasaki J, Rothe A, Kania A, Neumann G, Romheld V, Shinano T, Osaki M, Kandeler E (2005) Root exudation, phosphorus acquisition, and microbial diversity in the rhizosphere of white lupine as affected by phosphorus supply and atmospheric carbon dioxide concentration. J Environ Qual 34(6):2157–2166
- Wersin P, Hochella MF, Persson P, Redden G, Leckie JO, Harris DW (1994) Interaction between aqueous uranium(VI) and sulfide minerals: spectroscopic evidence for sorption and reduction. Geochim Cosmochim Acta 58(13):2829–2843
- <span id="page-27-0"></span>White PJ, Broadley MR (2003) Calcium in plants. Ann Bot 92(4):487– 511
- Whitehead NE, Brooks RR, Peterson PJ (1971) The nature of uranium occurrence in the leaves of Coprosma australis (A. Rich) Robinson. Aust J Biol Sci 24:67–73
- WHO (2001) Depleted uranium sources, exposure and health effects. Department of Protection of the Human Environment, World Health Organization, Geneva
- WHO (2004) Uranium in drinking-water: background document for development of WHO guidelines for drinking-water quality. World Health Organization, Geneva, p 23
- Wielinga B, Bostick B, Hansel CM, Rosenzweig RF, Fendorf S (2000) Inhibition of bacterial promoted uranium reduction: ferric (Hydr)oxide as competitive electron accepter. Environ Sci Technol 34:2190–2195
- Wilkins M, Livens F, Vaughan D, Lloyd J (2006) The impact of Fe (III)-reducing bacteria on uranium mobility. Biogeochemistry 78 (2):125–150
- Worms I, Simon DF, Hassler CS, Wilkinson KJ (2006) Bioavailability of trace metals to aquatic microorganisms: importance of chemical, biological and physical processes on biouptake. Biochimie 88 (11):1721–1731
- Wu WM, Carley J, Fienen M, Mehlhorn T, Lowe K, Nyman J, Luo J, Gentile ME, Rajan R, Wagner D, Hickey RF, Gu B, Watson D, Cirpka OA, Kitanidis PK, Jardine PM, Criddle CS (2006) Pilotscale in situ bioremediation of uranium in a highly contaminated aquifer. 1. Conditioning of a treatment zone. Environ Sci Technol 40(12):3978–3985
- Yang J, Volesky B (1999a) Biosorption and evolution of uranuim with seaweed biomass. In: Conference IBS'99-Spain, Spain
- Yang J, Volesky B (1999b) Modelling uranium-proton ion exchange in biosorption. Environ Sci Technol 33:4079–4085
- Yang DH, Andersson B, Aro EM, Ohad I (2001) The redox state of the plastoquinone pool controls the level of the light-harvesting

chlorophyll a/b binding protein complex II (LHC II) during photoacclimation—cytochrome b(6)f deficient Lemna perpusilla plants are locked in a state of high-light acclimation. Photosynth Res 68(2):163–174

- Yazzie M, Gamble SL, Civitello ER, Stearns DM (2003) Uranyl acetate causes DNA single strand breaks in vitro in the presence of ascorbate (vitamin C). Chem Res Toxicol 16(4):524–530
- Yong P, Macaskie LE (1998) Bioaccumulation of lanthanum, uranium and thorium, and use of a model system to develop a method for the biologically-mediated removal of plutonium from solution. J Chem Technol Biotechnol 71(1):15–26
- Zänker H, Ulrich K-U, Opel K, Brendler V (2007) The role of colloids in uranium transport: a comparison of nuclear waste repositories and abandoned uranium mines. In: Cidu R, Frau F (eds) IMWA Symposium 2007: water in mining environments. Cagliari, Italy
- Zanonato P, Bernardo PD, Bismondo A, Liu G, Chen X, Rao L (2004) Hydrolysis of uranium(VI) at variable temperatures (10–85 °C). J Am Chem Soc 126:5515–5522
- Zanonato PL, Di Bernardo P, Szabo Z, Grenthe I (2012) Chemical equilibria in the uranyl(vi)-peroxide-carbonate system; identification of precursors for the formation of poly-peroxometallates. Dalton Trans 41(38):11635–11641
- Zayed G, Winter J (2000) Inhibition of methane production from whey by heavy metals—protective effect of sulfide. Appl Microbiol Biotechnol 53(6):726–731
- Zayed A, Gowthaman S, Terry N (1998) Phytoaccumulation of trace elements by wetland plants: I Duckweed. J Environ Qual 27 (3):715–721
- Zeman FA, Gilbin R, Alonzo F, Lecomte-Pradines C, Garnier-Laplace J, Aliaume C (2008) Effects of waterborne uranium on survival, growth, reproduction and physiological processes of the freshwater cladoceran Daphnia magna. Aquat Toxicol 86(3):370–378