



DArTseq-derived SNPs for the genus *Psidium* reveal the high diversity of native species

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Abstract

In this study, we characterized DArTseq-derived SNPs for species of *Psidium*, an important Neotropical genus within the family Myrtaceae. Nine *Psidium* species were used for the identification and characterization of an interspecific set of SNPs. The samples chosen represented a wide range of variation regarding agronomic traits and geographical origin. The samples were genotyped using the DArTseq methodology. This approach, associated with annotation analysis, generated a set of conserved, polymorphic, and possibly functional SNPs. Most of the SNPs in coding regions of *Psidium* were annotated for the gene ontology classes. We observed high interspecific diversity among species and considerable intraspecific genetic diversity in native species (*P. cattleianum*, *P. myrtoides*, *P. acidum*, and *P. oblongatum*). *Psidium guajava*, a cultivated species, shows low diversity and shares genetic similarities with the tetraploid *P. guineense*. The group of markers selected in this study will contribute to the development of a panel of interspecific SNPs for *Psidium*. Our investigation will support further studies regarding genetic variability, diversity, phylogeny, cytogenetics, and other relationships among *Psidium* species, constituting essential information for the creation, conservation, management, and use of genetic resources in breeding programs.

Keywords Myrtaceae · Polyploidy · Molecular markers · Genome · Gene ontology

Introduction

Psidium is one of the most important and explored genera within the family Myrtaceae, including about one hundred species distributed from Mexico and the Caribbean to Uruguay and northern Argentina (Thornhill et al., 2015).

The genus belongs to the tribe Myrteae (subtribe Pimentinae), which represents almost all Myrtaceae diversity in Neotropical environments (Wilson et al., 2005; Lucas et al., 2007). Approximately 60 taxa occur in Brazil, and the Atlantic Forest is particularly rich in *Psidium* species, with a total of 38, of which 18 are endemic to this biome (Flora do Brasil, 2020). Guava (*Psidium guajava* L.) is prominent among the species of this genus owing to its commercial and medicinal value. However, *Psidium* comprises other promising species (e.g., the *araças*), with some significant attributes such as early maturation, resistance to certain pests and diseases, and exotic fruit flavor. These attributes may be used in guava breeding programs (Dias et al., 2015; Mendes et al., 2017; Vitti et al., 2020). In addition, natural interspecific hybrids were found to occur between phylogenetically close species, *P. guajava* and *P. guineense* (Landrum et al., 1995). Moreover, an artificial interspecific hybrid of these species has recently become available for use as nematode-resistant rootstock in the guava crop (Costa et al., 2016).

Guava and some other *araça* species (*P. guineense*, *P. cattleianum*, and *P. myrtoides*) have been characterized

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for agronomic traits in Brazil (Nimisha et al., 2013). Chemical analyses have also demonstrated the value of substances from these plants in co-treatments of different illnesses, such as diabetes mellitus, cardiovascular diseases, cancer, and parasitic infections (Cerio et al., 2017). *Psidium* species also contain functional compounds with antioxidant activities (Mallmann et al., 2020), and they are sources of essential oils rich in terpenes, which are of agronomic, industrial, and medicinal interest (Mendes et al., 2017).

The genus *Psidium* is karyologically variable, with a suggested basic number of chromosomes of $x = 11$ (Tuler et al., 2019a). *Psidium guajava*, *P. oblongatum*, and *P. cauliflorum* have a diploid nature ($2n = 2x = 22$) (Souza et al., 2015; Marques et al., 2016; Tuler et al., 2019a), whereas *P. guineense* and *P. friedrichsthalianum* are described as tetraploids ($2n = 4x = 44$), and *P. myrtooides* and *P. longipetiolatum* as hexaploids ($2n = 6x = 66$), confirming a trend towards polyploidy in the genus. In addition, intraspecific polyploidy (i.e., within the same species) also occurs, as found in *P. cattleyanum*, with chromosome numbers varying between 44 and 88 (Machado et al., 2020; Tuler et al., 2019a; Souza et al., 2015).

Genetic diversity and phylogenetic relationships of *Psidium* species have been elucidated through the use of different molecular techniques, such as random amplified polymorphic DNA (RAPD), transferability of simple sequence repeats (SSR) and inter-simple sequence repeats (ISSR), and phylogenetic markers (Abdelmigid and Morsi, 2018; Bernardes et al., 2018; Tuler et al., 2019b; Han et al., 2020; Machado et al., 2020). However, the use of genomic approaches, including SNP markers, is still incipient for these species. Advances in the genetic and genomic tools for distinct *Psidium* species may provide more reliable taxonomic information and support for the development of cultivars with improved agronomic traits (Costa and Santos, 2017), as well as for studies on diversity, conservation, phylogenetics, and evolution of the species.

SNP markers have been increasingly used owing to their high performance, precision, resolution, and reproducibility (Elshire et al., 2011; Kilian et al., 2012; Sansaloni et al., 2020). They are abundant, widely distributed, and in high density in the genome, favoring the detection of multiple haplotypes by the combination of tightly linked SNP alleles. In addition, these markers have been widely used because of their automated genotyping in next-generation sequencing (NGS) platforms (Deschamps et al., 2012; Ren et al., 2015; Al-Beyroutiová et al., 2016; Garavito et al., 2016; Nimmakayala et al., 2016; Egea et al., 2017; Ndjioudjop et al., 2017; Valdisser et al., 2017; Sukumaran et al., 2018).

DArTseq, a high-throughput whole-genome genotyping technology, enables the sequencing and genotyping of hundreds or thousands of SNPs by using methods that reduce the genome complexity that enriches genic regions (Kilian et al.,

2012; Fellers, 2008). The so-called STAGs, fragments sequenced in the population that contain the SNPs, also allow the alignment of these sequences with the genome of the target (or close) species, and the annotation (functional and structural) of the marker regions in the genome. This *in silico* analysis has been used to identify conserved genomic regions among species, to enable the construction of genotyping panels for multiple species, and to detect and characterize genomic regions with signals of selection (Shams et al., 2019; Merot-L'Anthoene et al., 2019; Valdisser et al., 2017).

Eucalyptus grandis, an important representative of Australian Myrtaceae species, is a reference in the family for omics approaches (Grattapaglia et al., 2012; Myburg et al., 2014). Thus, the identification of STAGs in a suite of evolutionarily conserved genes of *E. grandis*, shared with non-traditional model species of Neotropical *Psidium* lineages, is important for genome evolutionary studies in Myrtaceae. In addition, a set of SNPs for multiple species of *Psidium* is also important for the polyploid species of the genus, as the complexity of the genome and the difficulties in developing high-performance genotyping platforms create challenges in the efforts of carrying out breeding and conservation programs, as well as evolutionary studies, for these species.

The identification of SNPs for multiple *Psidium* species can promote advances in the characterization of the structure and function of common genomic regions for these species. In addition, a set of SNPs with high discriminatory potential between species, as well as between genotypes, is useful for routine characterization of accessions in germplasm banks and in breeding programs. To support the construction of a multi-species panel of SNPs, the chosen markers must be conserved among species, preferably selected within the gene regions, and should be polymorphic and useful in the study of intra- and interspecific variation. Therefore, the aim of the present study was to identify and characterize a set of SNP markers that may be useful for breeding, phylogenetics, and further approaches regarding the genome of *Psidium* species.

Materials and methods

Plant material A total of 94 samples from nine species of *Psidium* were evaluated in the present study. *P. guajava* (Voucher: Tuler.A.C 563) samples were the most numerous, totaling 63. Of these, 19 were cloned cultivated genotypes, including 11 cultivars registered in the Brazilian Ministry of Agriculture (Ministério da Agricultura, Pecuária e Abastecimento - MAPA) (<http://sistemas.agricultura.gov.br/>) (Paluma, Pedro Sato, Século XXI, Cortibel LG, Cortibel LM, Cortibel Branca LG, Cortibel RM, Cortibel Branca RM, Cortibel RM2, Cortibel RG and Cortibel SLG) and 8 cultivated materials (Chinesa Branca, Indiana, Kuse, Maçã, Roxa, Tailandesa, Tailandesa Branca, and Petri). Thirty-two

(32) were Cortibel® (C) genotypes obtained by selection from the orchard of seminal origin in the state of Espírito Santo (Frucafé) (VII, XI, XIII, XVIII, XIX, XX, XXII, XXIII, XXIV, XXV, XXVI, XXVII, XXVIII, XXIX, XXX, XXXI, XXXII, XXXIII, XXXIV, XXXV, XXXVI, XXXVII, XXXVIII, XXXIX, XL, XLI, XLII, XLIII, XLV, XLVI, XLVII, and XLVIII). All these *P. guajava* cultivars and selected genotypes were collected from adult plants in the Frucafé Nursery, certified by the Brazilian Ministry of Agriculture. In addition, 12 samples of naturally occurring guava trees were also used from different states of Brazil: Alagoas (1), Bahia (2), Ceará (1), Espírito Santo (2), Pernambuco (1), Paraíba (1), Sergipe (1), Paraná (2), and São Paulo (1). We also included an individual with morphological traits of a natural hybrid (*P. guajava* × *P. guineense*) sampled in the state of Sergipe.

Eight (8) more *Psidium* species with different numbers of samples were added (number of samples/voucher): *P. gaudichaudianum* (01/Tuler.A.C 637), *P. acidum* (01/Tuler.A.C 524), *P. cattleyanum* (10/Tuler.A.C 553), *P. friedrichsthalianum* (01/UFES collection), *P. guineense* (05/Tuler.A.C 548), *P. myrtoides* (07/Tuler.A.C 510), *P. oblongatum* (04/Tuler.A.C 567), and one unidentified sample, *P. sp1*. The samples were collected from adult plants in the municipality of Alegre, ES, most of which are found in the germplasm collection of the Universidade Federal do Espírito Santo (UFES), which maintains naturally occurring plants collected from different states of Brazil. The samples chosen represent a wide range of variation based on agronomic traits and different geographical origins (Table 1).

Genotyping by sequencing on the DArTseq platform The DNA of each sample was extracted from leaves macerated with liquid nitrogen according to the CTAB method (Doyle and Doyle, 1990). After DNA extraction, all quality parameters were checked according to recommendations of the Diversity Arrays Technology Pty. Ltd. company (Canberra, Australia) (<https://www.diversityarrays.com/faq/>). The quantity and quality of the DNA samples were verified using NanoDrop and visualized in 0.8% agarose gel. Only intact DNA samples were used. A volume of 60 µL of each sample, containing 60 ng/µL of DNA, was pipetted into PCR plates and sent to the Diversity Arrays Technology Pty. Ltd. company, according to recommendations (<https://www.diversityarrays.com/faq/>), for production of the library, genotyping by sequencing, and marker identification (Kilian et al., 2012). The genome representation was obtained from a genomic library attained from digestion with the restriction enzymes *Pst*I and *Mse*I. The ends of the cleaved fragments were connected to a barcode adapter and to a common adapter. The mixed fragments were amplified with two primers with sequences

complementary to the attached adapters and to the oligonucleotides of the Illumina sequencing platform. All the successful amplifications were clustered and applied to a flow cell for amplification (Kilian et al., 2012). The clusters were sequenced on the Illumina HiSeq2500 sequencing platform, and the sequences were processed using proprietary DArT analytical pipelines (Sansaloni et al., 2020). The sequences per barcode/sample were identified and used in marker calling. Poor-quality sequences were filtered out, and identical ones were collapsed into fastqcall files. These files were used in a pipeline for DArT PL's proprietary SNP-calling algorithms (DArTsoft-seq14), as described by Sansaloni et al. (2020). The DArTseq quality markers were determined by the reproducibility parameter (the proportion of technical replicate assay pairs for which the marker score is consistent) of at least 0.95 and call rate parameter (the proportion of samples with genotypic score, i.e., not recorded as missing data) with a threshold of 0.7, which constitute the main parameters for marker selection. In addition, the statistics of the filtered DArTseq markers included the expected heterozygosity (H_e), the minor allele frequency (MAF), and the polymorphic information content (PIC).

Data analysis SNP markers from common and specific species were identified for all of the species. For *P. guajava*, *P. cattleyanum*, *P. guineense*, *P. myrtoides*, and *P. oblongatum*, the DArTseq quality markers were determined by call rate = 1 and reproducibility > 0.95 (Wenzl et al., 2004), number of polymorphic SNP markers, monomorphic SNPs, and total SNPs per species. The non-amplified markers (from the total amplified in all species) were also calculated.

DArTseq-derived sequences of 40–69 base pairs (bp) containing SNPs (STAGs) (Shams et al., 2019) obtained from 94 individuals of *Psidium* were aligned against the reference genome of *Eucalyptus grandis*, a pivotal species for genomic studies in Myrtaceae (Grattapaglia et al., 2012; Myburg et al., 2014). This analysis was performed and provided by the DArT Pty. Ltd. company.

Identification and annotation of STAGs present in coding regions We performed an in silico functional annotation of each of the STAGs, which were classified regarding their presence in coding and non-coding regions. *E. grandis* annotation and genomic sequence data were used to obtain the functional annotation. To identify the most conserved regions among Myrtaceae species, the unique STAGs were classified as being present in the *E. grandis* genome (anchored) or not found in the genome (non-anchored). The localization of each anchored STAG was obtained by a search with the Basic Local Alignment Search Tool (BLAST) on *E. grandis* gene sequences, available at the

Table 1 Description of species used in the study. Species occurrence in physiognomies of the Atlantic Domain in Brazilian states. States are indicated by acronyms; those followed by an asterisk (*) are part of the Atlantic Domain. Acronyms for physiognomies: *OF*, ombrophilous

forest; *MF*, mixed ombrophilous forest; *SF*, semi-deciduous forest; *DF*, deciduous forest; *NB*, northeastern *Brejos*; *RV*, rock outcrop vegetation; *RE*, *Restinga*; and *AA*, anthropic area. Morphological traits of fruit and seeds of *Psidium* species. *N*, number of vouchers analyzed per species

Species	Brazil occurrence	Physiognomies	Habit	Fruit size	<i>N</i>	Number of seeds	Length (mm)	Width (mm)
<i>P. guajava</i>	Occurs in almost all states	RE, SF, AA	Tree or shrub 1.8–8 m	23–60 × 8–40 mm	29	277.0	3.5	2.25
<i>P. guineense</i>	Occurs in almost all states	RE, SF, DF, AA	Tree or shrub 0.3–6 m	13–24 × 8–20 mm	55	227.0	2.5	2.75
<i>P. cattleyanum</i> *	AL*, BA*, CE*, PE*, SE*, RJ* ES*, MG*, SP*, PR*, SC*, RS*	RE, SF, OF	Tree or shrub 0.5–15 m	15–40 × 9–30 mm	50	41.4	5.5	4.5
<i>P. myrtoides</i>	TO, BA*, CE, MA, DF, GO, ES*, MG*, RJ*, SP*, PR*, RS*, SC*.	RE, SF, OF, MF	Tree 2–20 m	9–16 × 6.5–14 mm	38	7.0	4.5	3.9
<i>P. oblongatum</i>	ES*, MG*, RJ*,	SF, OF	Tree 4–12 m	38–53 × 25–56 mm	9	17.0	8.0	5.5
<i>P. gaudichaudianum</i>	ES*, RJ*, SP*	OF	Tree 20 m	20–34 × 15–20 mm				
<i>P. acidum</i>	AM	-	Tree 10 m	25–50 × 15–20 mm	8	60	5	3
<i>P. friedrichsthalianum</i>	AC	-	Tree 10 m	45–56 × 40–60 mm	8	73	5	4

Phytozome database (<https://phytozome.jgi.doe.gov/pz/portal.html>). For the STAGs non-anchored in *Eucalyptus*, a BLAST was performed against *P. guajava* gene sequences provided by the research group (<http://guava.ufes.br/>). The BLAST was carried out using the BLASTn software (NCBI n.d.), with default parameters and output format changed to *csv* (comma-separated values) and *e*-value $1e^{-7}$.

The functional annotations of the anchored STAGs present in coding regions were obtained from the annotation of *E. grandis*, available in the Phytozome Database. For the non-anchored STAGs, it was necessary to obtain the amino acid sequence translated by each available gene in the data from the structural annotation of *P. guajava* (from the research group database), then perform BLASTp in the NR database and obtain a predicted gene association in *P. guajava* X gene identifier (*gi*). The functional annotation of each *gi* was obtained with the Uniprot database support (www.uniprot.org/uploadlists/).

Files from the processing were analyzed to generate abundance graphs for GOs (Gene Ontology) in each sequence group. With this data, the STAGs were classified as being present in coding regions, when derived from polymorphic sites inside coding regions, or as present in non-coding regions, when found in polymorphic and random sites of the genome (Andersen and Lubberstedt, 2003).

Marker selection A descriptive analysis was performed for the SNPs of each species using the DARTR package in the R software (R Core Team, 2020). To select SNP markers for multiple species of *Psidium*, we filtered the SNPs with call rate = 1.00 and MAF > 0.01 and PIC parameters. The excess of heterozygosity was verified for each SNP to identify possible highly similar sequences that occupy different chromosomal locations (multicopy loci), as described by Willis et al. (2017).

The analysis within and between species was performed using Euclidean distance for principal coordinate analysis (PCoA) with the R software (R Core Team, 2020). In addition, this set of markers was applied for analysis of UPGMA clustering of the genotypes used (Brazilian cultivars, selected genotypes, and guava trees of natural occurrence) for application in Brazilian guava breeding.

Phylogenetic reconstruction We analyzed the SNP data with the SNPhylo software (Lee et al., 2014). A *vcf* file was used as data input and analyzed with the script *snphylo.sh -v VCF_file A -b -B 1500*, where *-A* performs multiple alignments by MUSCLE, *-b* performs the bootstrap analysis and generates a tree, and *-B* is the number of bootstrap samples. After that, the Newick file (tree format file) was generated using the iTol software (Letunic and Bork, 2019). Finally, the modified file was edited using the Affinity Designer® program.

Results

A total of 124,069 *loci* were obtained with SNPs, representing 64,672 unique sequences. Of these, 22,050 STAGs were anchored on the eleven chromosomes and scaffolds of the reference genome *E. grandis* (Myburg et al., 2014). Considering only the anchored SNPs, the averages of PIC and the frequency of homozygote and heterozygote *loci* per chromosome were similar, whereas the average of heterozygosity of the SNPs in the chromosomes was low (0.026) (Supplementary File S1). The SNPs anchored to chromosomes 10 and 11 had an average PIC above 0.2, revealing variability. SNPs with high PIC values (0.5) were detected in chromosomes 06, 08, 10, and 11, demonstrating the possibility of conserving selection, but selection of polymorphic *loci* markers. The SNPs exclusive to *Psidium* species (102,019) had an average PIC of 0.2 (Supplementary File S1).

Psidium guajava and *P. guineense* had similar numbers of SNPs and STAGs anchored to the *E. grandis* genome, whereas *P. oblongatum* and *P. myrtilloides* had lower numbers of markers anchored in this manner (Fig. 1A).

Sixty percent of the total SNPs (75,074 STAGs) were recorded in coding regions, indicating the effectiveness of the DArTseq methodology in selecting the fraction of the genome that corresponds to coding regions. For the SNPs anchored to the *E. grandis* genome, 82% were recorded in coding regions,

while for non-anchored markers this percentage was 55% (Fig. 1A).

A similar pattern was found for the parameters evaluated in *P. acidum* and *P. friedrichsthalianum*. *Psidium oblongatum*, exceptionally, had a similar number of *loci* in coding and non-coding regions, while the other species had a higher number of *loci* in coding regions (Fig. 1A). *Loci* monomorphic within species and polymorphic among species were verified. A large number of monomorphic *loci* per species indicate genomic regions shared between species, but do not indicate intraspecific polymorphism (Fig. 1B, C, D). The number of monomorphic *loci* was higher in *P. guajava*, *P. guineense*, and *P. cattleyanum*. These markers can be investigated for inter-specific differences.

Psidium guajava, *P. guineense*, and *P. cattleyanum* had a larger number of polymorphic *loci* than *P. myrtilloides* and *P. oblongatum* did. These species also had the smallest number of *loci* in shared coding regions and the highest marker absence. The large number of *loci* not shared with the other species showed the divergence of *P. oblongatum* and *P. myrtilloides*. The number of non-shared regions for *P. cattleyanum* was greater than the number observed in *P. guajava* and *P. guineense* (Fig. 1B, C, and D). For indication of a multispecies set of SNP markers, those conserved in the species (with intra- and interspecific polymorphisms) were prioritized.

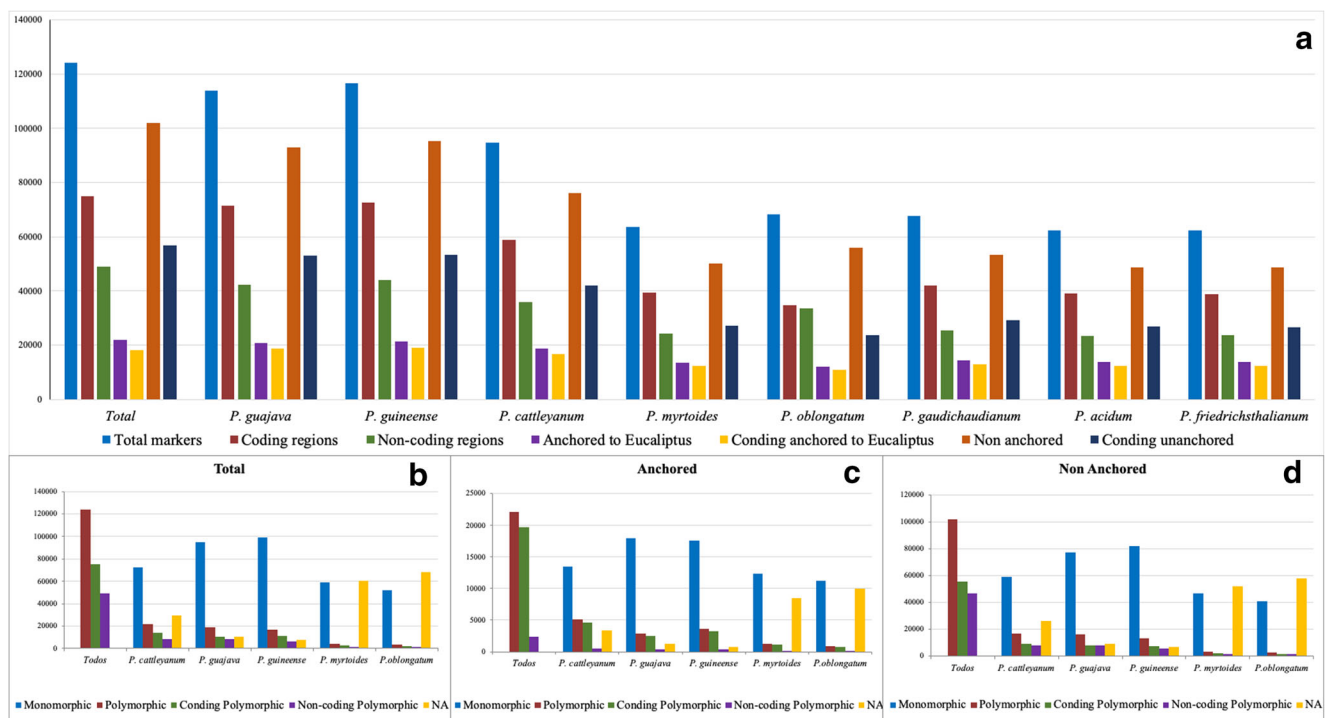


Fig. 1 (A) Distribution of 124,069 SNP markers identified in nine species into the classes: Coding and non-coding regions; anchored in *E. grandis* genome and coding anchors; non-anchored and non-anchored in coding regions. Distribution of SNP markers: (B) total, (C) anchored in

Eucalyptus genome, and (D) non-anchored. In each class, the SNPs were subdivided by species and recorded as monomorphic, polymorphic, polymorphic in coding regions, polymorphic in non-coding regions, and absence of the marker (NA)

We verified the potential gene ontologies (GO) related to STAGs present in coding regions of anchored (18,211 STAGs) and non-anchored (56,836 STAGs) through functional annotation. A coding region can be present in one or more of three main ontological structure categories: biological process, cellular component, and molecular function. From the 18,211 STAGs anchored in coding regions of the *Eucalyptus* genome, 6325 were aligned in coding regions with functional annotation, represented by 959 GOs. From the remaining non-anchored STAGs in coding regions (56,836), 11,539 annotations were found, which identified 1384 different GOs.

The groups of genes with STAGs of *Psidium* species are represented in Supplementary File S2. The 20 most abundant gene classes within the biological processes ontology (Supplementary File S2-A) showed 4810 GOs associated with STAGs anchored to the *Eucalyptus* genome, and 2499 were specific to the *Psidium* species (non-anchored). For the cellular component graph (Supplementary File S2-B), specific regions of STAGs for *Psidium* (non-anchored) had a larger number of GO terms (6343), demonstrating the diversification of these regions in the *Psidium* genomes. Both anchored and non-anchored STAGs showed the integral components of the membrane class as the most abundant. The STAGs in regions of genes with molecular function are listed in Supplementary File S2-C. In this group, the class of genes related to ATP binding was the most abundant, with 1518 GOs in anchored and 2276 in non-anchored regions. Molecular function was the ontological class most represented by the annotation of STAGs in this study, exhibiting 48% of GOs, followed by cellular components (28.5%) and biological processes (23.5%). When differentiated into anchored STAGs and non-anchored STAGs, only the cellular components showed higher values for the non-anchored STAGs (72%).

The genetic relationship within a species and between species was evidenced by the results of principal coordinate analysis (PCoA), performed with three data groups: all SNPs, anchored SNPs, and non-anchored SNPs (Fig. 2). The results

provided a spatial representation of the relative genetic distances between individuals, with little difference in the proportion of the variation explained in the analysis with all of the SNPs (59.6%) and with non-anchored ones (58.1%). The greater representativeness of the data was observed in the analysis with the anchored SNPs (explained variation = 66.1%), justifying the selection of these markers as a priority for interspecific evaluations.

The PCoA distribution resulted in three distinct groups. *Psidium guajava* and *P. guineense* were grouped on the first axis, showing less intraspecific variation compared to the other wild and polyploid species. *Psidium myrtilloides* and *P. oblongatum* were close on one of the PCoA axes. Less intraspecific variation was observed in individuals of *P. oblongatum* than in individuals of *P. myrtilloides*.

Considerable variability was found for *P. acidum*, *P. cattleyanum*, *P. friedrichsthalianum*, *P. gaudichaudianum*, and *Psidium* sp1, which consist of tree species with large fruit, many of these tree species grown due to their socioeconomic importance. The ten samples of *P. cattleyanum* represented a subdivision. Six samples were close to *P. acutangulum*, *P. friedrichsthalianum*, and *P. gaudichaudianum*, which share many SNPs. The other four samples of *P. cattleyanum* were clustered with a sample harvested in Espírito Santo, identified as *Psidium* sp.1 (not yet identified).

The 5951 SNPs whose call rate was 1 were considered the most conserved of the species. Of these only 14 showed heterozygotes above the expected 50% for bi-allelic SNPs (0.5–0.69) (Willis et al., 2017). For the 5951 SNPs, a new analysis was performed for the anchored and non-anchored SNPs (Supplementary File S1; see Supplemental Data with this article). Similar values between groups of SNPs were observed, and the average PIC increased from 0.20 (Supplementary File S1-A) to 0.24 (Supplementary File S1-B), suggesting *loci* diversity. Of these SNPs, 78% showed functional annotation and 37% were anchored to the *E. grandis* genome (Fig. 3A,

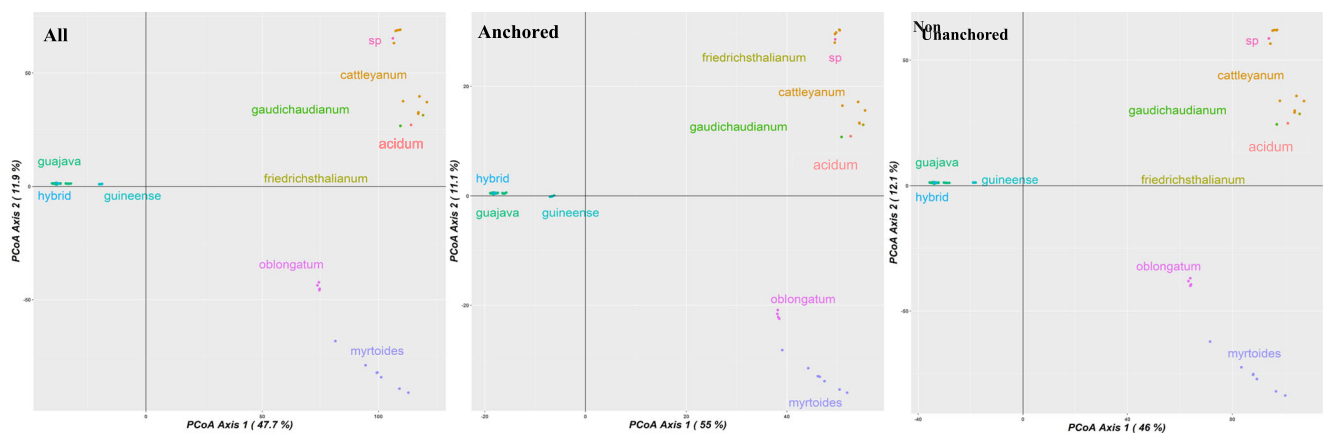


Fig. 2 Principal coordinate analysis (PCoA) based on SNP markers for 94 *Psidium* samples. Each species is represented by a different color for all markers, anchored markers, and non-anchored markers

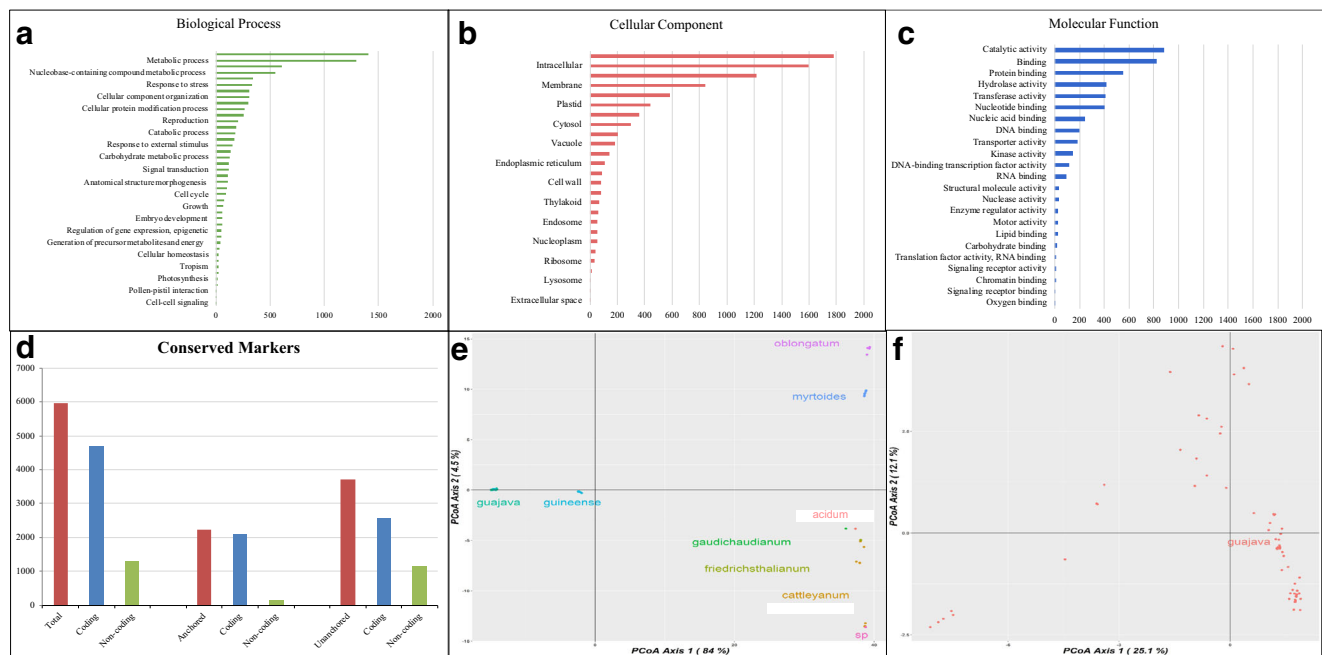


Fig. 3 List of the 5951 SNP markers selected with their gene ontology (GO) for genes related to (A) biological processes (blp), (B) cellular components (cc), and (C) molecular function (mf). (D) Distribution of selected markers in coding and non-coding regions, considering all

B, and C). The percentage of anchored SNPs in coding regions was 93% (Fig. 3D). Of the 5951 sequences, 4846 are present in the assembly of the *P. guajava* genome. A total of 6459 alignments were obtained; of 4258 STAGs aligned against 5719 genes, 2734 exhibit GO annotation.

The biological process and cellular component ontological classes were represented with 39.28% and 39.32% of the GOs found, respectively (Fig. 3A and B). Among the biological processes with most abundant GO terms, responses to stress, responses to external stimuli, and reproduction were most prominent. In turn, the class of molecular function, widely represented previously, corresponded to 21.4% of the results in this analysis (Fig. 3C).

The PCoA analysis performed with the SNPs from the STAGs of all samples reveals the potential of this tool for genomic approaches of interspecific (Fig. 3E) and also intraspecific discrimination, as shown by the guava genotypes used (Fig. 3F). These SNPs increased the power of discrimination between species (only one component explained 84%). As in the first cluster, here we also observed the formation of three distinct groups: (1) *P. guajava* and *P. guineense*, very close species; (2) *P. myrtoides* and *P. oblongatum*; and (3) *P. acidum*, *P. cattleyanum*, *P. friedrichsthalianum*, *P. gaudichaudianum*, and *Psidium* sp1. These SNPs also showed ability to discriminate genotypes within species (with genotypes distributed on four axes) (Fig. 3F). The low variability of *P. guajava* in comparison with the other native species was also evident, indicating high diversity, as well as inter- and intraspecific variability (Fig. 3E).

markers, markers anchored in *E. grandis* genome, and markers exclusive to *Psidium* species (non-anchored). Principal coordinate analysis (PCoA) based on the SNP markers selected (E) for 94 samples of *Psidium* and (F) for *P. guajava* samples only.

A phylogeny was also provided (Fig. 4). The reconstructed tree was formed by five well-supported clades (ML bootstrap = 100), showing that the markers used had sufficient resolution to separate five species of *Psidium* sp.: *P. guajava*, *P. guineense*, *P. oblongatum*, *P. myrtoides*, and *P. cattleyanum*. Three genotypes, previously classified as *P. gaudichaudianum* (x92), *P. acidum* (x93), and *P. friedrichsthalianum* (x94), were allocated to the larger clade of *P. cattleyanum*. The distance of *P. guajava* genotypes to those of other species is shown by the size of the central branch, which demonstrates an evolutionary process distinct from the others.

In addition, the application of this set of markers for breeding of *P. guajava* was demonstrated by analysis of the UPGMA clustering of the genotypes used, and seven clusters were shown (Fig. 5).

The STAGs shared with *E. grandis* represent a genomic tool for the family. The distribution of these markers in chromosomes revealed enrichment of *Psidium*-conserved coding regions in chromosomes 2, 6, and 11 of *E. grandis*. The STAGs exclusive to the *Psidium* species represent specific evolution in the genus. All information about these SNP markers is available (Supplementary File S3).

Discussion

The DArTseq technology provided a large number of SNPs, which can be anchored to reference genomes (Sansaloni et al.,

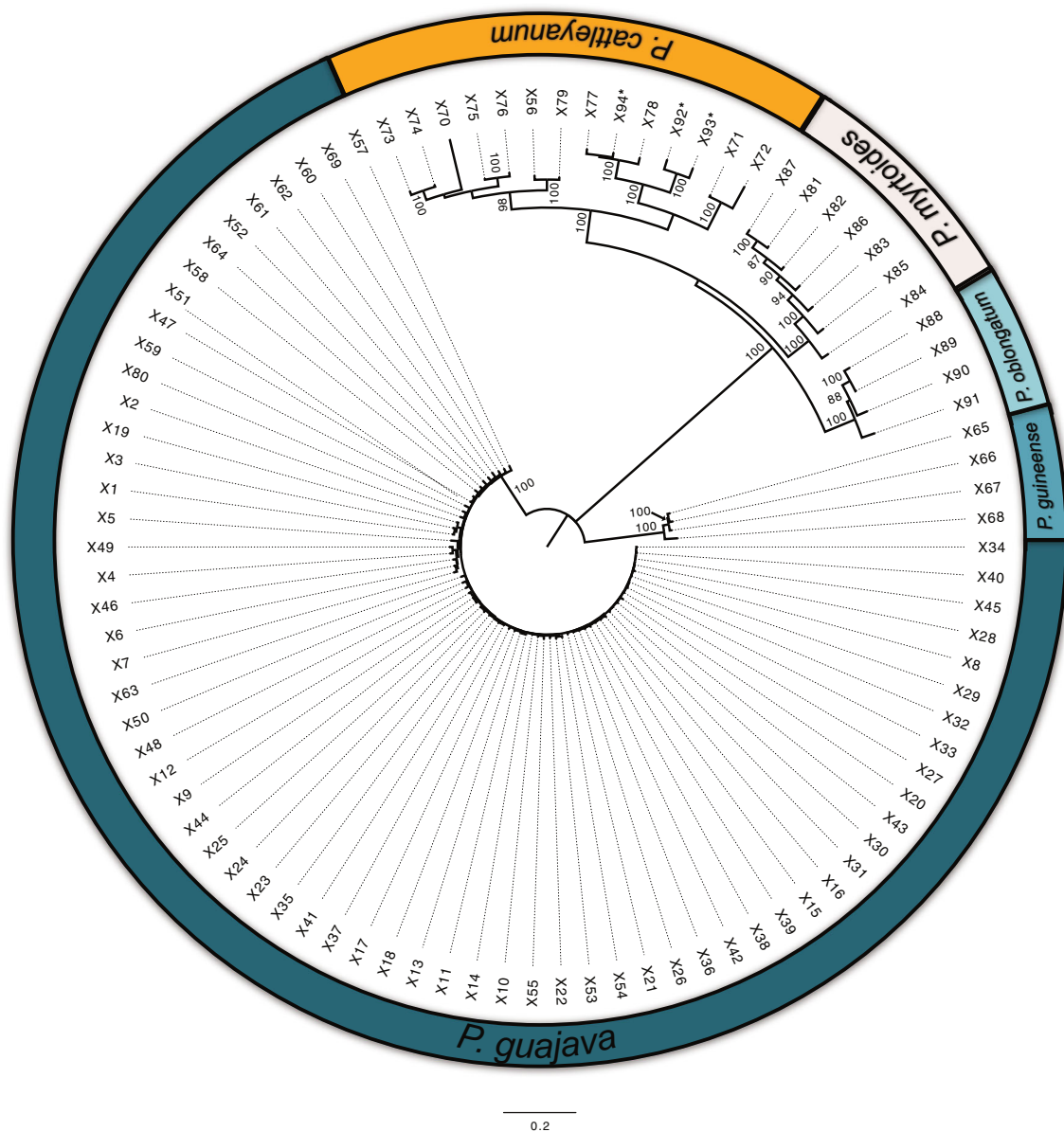


Fig. 4 Genetic relationships among *Psidium* species. Phylogram based on the maximum likelihood inference of 5951 SNPs of *Psidium* sp. The bootstrap probability is indicated near the branch nodes, and only values

greater than 75 are shown. An asterisk denotes genotypes previously classified as *P. gaudichaudianum* (x92), *P. acidum* (x93), and *P. friedrichsthalianum* (x94)

2011). However, in most studies, the STAGs that contain the SNPs remain unused. These sequences provide important information for identification of regions evolutionarily conserved across related species (Shams et al., 2019). Here, a set of STAGs, conserved for *Psidium* species and associated with protein-coding regions and random sequences, revealed an important genomic tool for *Psidium* species, which can help increase knowledge regarding the genetics and evolution of these species, as well as assist in their breeding.

The SNPs provided robust information about the genetic relationships among species, corroborating morphological, cytogenetic, and genetic patterns previously described for these species (Tuler et al., 2015, 2019a, 2019b; Marques

et al., 2016). In addition, they exposed the wide divergence among individuals of native species, particularly *P. cattleianum*, for which intraspecific polyploidization has been reported (Machado et al., 2020). Moreover, the set of SNPs allowed intraspecific discrimination of the species evaluated, which supplements their morphological evaluation and allows the development of tools related to selection and genetic breeding, which is especially relevant for breeding of guava.

A higher number of *loci* shared by *Psidium* species and *E. grandis*, which represent genera belonging to distant tribes of Myrtaceae (Vasconcelos et al., 2017), increases the applications of this study due to the possible transferability of these

in the present study in relation to selected and cultivated genotypes in Brazil. These results confirm, refine, and expand previous reports from our research group for a common set of genotypes and species studied using SSR markers (Coser et al., 2012).

The narrow diversity of *P. guineense* may be related to recent divergence of the species, suggesting an event of self-polyploidy in relation to *P. guajava*, also reported by Marques et al. (2016) and Tuler et al. (2019b). The wide diversity observed for the other species is probably due to their multiple ploidies (Marques et al., 2016; Tuler et al., 2019a). Furthermore, the geographical dispersion of the species may also contribute, as the genetic characteristics of the populations depend on the interaction between genetic drift, gene flow, and natural selection, processes that can be strongly affected by the spatial distribution of the populations (Eckert et al., 2008).

Psidium oblongatum and *P. myrtilloides* were in the same group and were the species that most differed from the others. Moreover, they shared the fewest genomic regions with *Eucalyptus* and the other *Psidium* species, which shows their genetic distance. The genetic relationship between these two species was reported in a previous study of cluster analysis based on microsatellite amplification patterns performed by Tuler et al. (2015), in which they clustered with *P. sartorianum* and *P. brownianum*.

Psidium myrtilloides is a polyploid species known in the Atlantic Forest, Cerrado, and Caatinga in the Southeast, West-Central, and Northeast of Brazil. This wide distribution exposes individuals from this species to different environmental conditions, which may explain the separation of two groups of individuals within this same species, but the two groups were not maintained in the PCoA analysis only with the conserved markers. It is noteworthy that *P. oblongatum* also showed wide genetic diversity, yet it consists of a diploid species endemic to the Atlantic Forest (Tuler et al., 2019a).

Psidium cattleianum and *P. friedrichsthalianum* also formed a cluster. The proximity between these species has already been identified in other studies by different molecular markers (Costa and Santos, 2013). *P. cattleianum* has an intraspecific ploidy variation, which explains the wide genetic variability detected in the samples analyzed (Machado et al., 2020; Tuler et al., 2019a; Souza et al., 2015).

Costa and Santos (2017) also evaluated SNPs in *Psidium* species, using EUChip60Kchip, developed by Silva-Junior et al. (2015) for *Eucalyptus* species, with transfer of 3523 SNPs (around 5%) to *Psidium* samples. The authors concluded that the transfer of SNPs between genera was very reliable, in agreement with previous genetic divergence studies using microsatellites. In the present study, we found around 17.7% of markers of *Psidium* species common to *Eucalyptus*, thus increasing study possibilities.

In this study, we selected SNP markers of greater informativeness for nine *Psidium* species. These markers showed efficiency in detecting intra- and interspecific variations, with 37% shared with *E. grandis* and 78% aligned in sites with annotation in coding regions. These marker regions were delimited in the genome of *P. guajava* and provide data for the development of a large-scale and flexible panel of SNPs for genotyping different species of *Psidium*.

The evolution of *Psidium* gave rise to several endemic species differing in ploidy level, distribution range, and habitat preferences. In the present study, polyploidy contributed to the high diversity observed in the *Psidium* species studied. Furthermore, we verified wide genetic diversity in little-known native species, such as *P. gaudichaudianum*, *P. acidum*, and *P. oblongatum*. Investigating genetic variability and the relationships between *Psidium* species is important for the creation, conservation, management, and use of these genetic resources in breeding programs. The information found here may be used as an aid for these purposes.

The lack of phylogenetic resolution for some species reported above may be due to the small number of individuals, or even related to the markers used. It is believed that very close species need more markers with higher resolving power in order to be grouped in different clades (Parks et al., 2009; Swenson, 2009). Phylogenetic diversity can be directly affected by phylogenetic resolution when working with large datasets, and cryptic species are formed (Swenson, 2009; Santos et al., 2020).

The distance of *P. guajava* genotypes from the other species may be due to the fact that this species has long been domesticated. Moreover, the species may have been separated according to the type of evolution to which each gene was subjected, which can be more or less conserved.

Here, DArTseq-derived SNPs were used for the genetic analysis of diploid and polyploid species of *Psidium*. The present study also made advances in the detection and characterization of intra- and interspecific polymorphic genomic regions. In addition, a set of SNPs with high discriminatory potential among species, as well as among individuals, was proposed for routine characterization of accessions in gene banks and in breeding programs. The STAGs exclusive to the *Psidium* species represent specific evolution in the genus. The associated GO terms reveal classes of genes conserved in evolution of the family. Furthermore, the distribution of a subset of markers in *E. grandis* chromosomes has importance for guava breeding, comparative genomics, and genome evolution studies.

Conclusions

The DArTseq approach, associated with annotation analysis, generated a large set of conserved, polymorphic, and possibly

functional SNPs for *Psidium* species. A large number of markers conserved between *Psidium* and *Eucalyptus* contributes to *Psidium* research, which can benefit from studies on *Eucalyptus*, a pivotal genus in Myrtaceae. Species-specific markers were identified, and most of the functional polymorphisms in *Psidium* were annotated for the GO classes: biological processes, cellular components, and molecular functions. These SNPs generated clusters that showed intra- and interspecific relationships. These clusters revealed that *P. guajava* shows substantial genetic similarity to the tetraploid *P. guineense* and low variability in relation to the other species. A group of SNPs was selected and annotated, allowing the development of a panel of interspecific SNPs for *Psidium*. The association of STAGs with a suite of evolutionarily conserved genes reveals the conserved nature of orthologs along the genome of *E. grandis*, which are shared by the *Psidium* lineages. This result is important for guava breeding and evolutionary genomics studies in the Myrtaceae family, and highlights how STAGs can be used for comparative genomics and genome evolution studies in these non-traditional model species of *Psidium* for which whole-genome sequences are unavailable.

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Data archiving statement The sequence data of the 5951 STAGs are available in Supplementary File S5.

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