ORIGINAL PAPER

# Improvement of growth, under field conditions, of wheat inoculated with Pseudomonas chlororaphis subsp. aurantiaca SR1

Evelin Carlier · Marisa Rovera · Alberto Rossi Jaume · Susana B. Rosas

Received: 11 February 2008 / Accepted: 3 June 2008 / Published online: 27 June 2008 Springer Science+Business Media B.V. 2008

Abstract Effects of inoculation of wheat (Triticum aestivum L.) with the rhizobacterium Pseudomonas chlororaphis subsp. aurantiaca strain SR1 (termed SR1) were studied at an experimental field site in Río Cuarto, Argentina. Treatments involved SR1 inoculation with or without nitrogen/phosphorus fertilization. Inoculation produced a significant increase in plant height and root length in early growth stages. Inoculation plus fertilization with 40 kg ha<sup>-1</sup> urea/30 kg ha<sup>-1</sup> diamonic phosphate ("50% dose") gave a yield increase of 636 kg ha<sup>-1</sup> relative to control, and an increase of 472 kg  $ha^{-1}$  relative to fertilization with 80 kg ha<sup>-1</sup> urea/60 kg ha<sup>-1</sup> phosphate without inoculation. SR1 inoculation without fertilization, compared to control, produced increases of 6% in weight of 1,000 grains, 13% in number of spikes per plant, and 30% in number of grains per spike. Inoculation plus 50% dose fertilization also improved these parameters. Results of the study indicate that inoculation of wheat with SR1 improves various growth and yield parameters, and allows reduced dosage of nitrogen/phosphorus fertilizers in the field.

Keywords Pseudomonas chlororaphis subsp.  $\textit{aurantiaca}$  SR1  $\cdot$  Wheat  $\cdot$  Inoculation  $\cdot$  Plant growth promotion

## Introduction

Bacteria displaying high efficiency in improving plant development and increasing tolerance to pathogenic microorganisms have been designated as ''plant growthpromoting rhizobacteria'' (PGPR) (Kloepper and Schroth [1978](#page-4-0)). Recent controversy has arisen regarding the types of rhizobacteria that should be considered PGPR. Suggested characteristics to define the group include high population density in the rhizosphere after plant inoculation (since a rapidly declining population has low competitive ability against native soil microflora); effective colonization potential on the root surface; promotion of plant development; suppressing effect on other soil microorganisms that are plant pathogens; absence of harmful effects on humans. Experimental and field application of rhizobacteria has resulted in significant promotion of plant development, as observed in terms of emergence, vigor, biomass, development of root systems, and increased yield (up to 30%) of crop species such as corn (Fulchieri and Frioni [1994](#page-4-0)), soybean (Dashti et al. [1998\)](#page-4-0), chickpea (Del Gallo and Fabbri [1990](#page-4-0)), wheat (Luz [2001](#page-4-0)), and others (Okon and Labandera-Gonzalez [1994;](#page-4-0) Bashan [1998;](#page-4-0) Lucy et al. [2004](#page-4-0)).

The positive effects of PGPR have been correlated with increased mobilization of insoluble nutrients and consequent improvement in plant nutrient uptake (Lifshitz et al. [1987](#page-4-0)); production of plant growth regulators (Dubeikovsky et al. [1993\)](#page-4-0); suppression of deleterious bacteria and phytopathogenic fungi (Weller [1998;](#page-5-0) Weller and Thomashow [1993](#page-5-0)), mediated by production of antibiotics (Hebbar et al. [1992](#page-4-0); Thomashow et al. [1990](#page-5-0)), iron-sequestering compounds (Loper and Buyer [1991\)](#page-4-0), extracellular lytic enzymes (Fridlender et al. [1993](#page-4-0)), cyanhydric acid (Voisard et al. [1989](#page-5-0)), induced systemic resistance (ISR) (Andrade et al. [1998\)](#page-4-0), or competition for infection sites and nutrients (Bull et al. [1991\)](#page-4-0). These mechanisms all require direct contact between the bacteria and the surface or interior of root tissues, and active state of the inoculated bacteria (Weger et al. [1995](#page-4-0); Höflich et al. 1995).

E. Carlier ( $\boxtimes$ ) · M. Rovera · A. Rossi Jaume · S. B. Rosas Universidad Nacional de Rı´o Cuarto, Rı´o Cuarto, Argentina e-mail: evelincarlier@gmail.com

Studies to date suggest that positive growth responses of wheat (Triticum aestivum L.) to inoculation with PGPR are due in part to increased root absorption capacity. Bacterial genera studied in this regard include Azospirillum (Bashan and Levanony [1990](#page-4-0); Caballero-Mellado et al. [1992\)](#page-4-0), Azotobacter (Rai and Gaur [1988](#page-5-0)), Bacillus (de Freitas [2000](#page-4-0)), Pseudomonas (Zaidi and Khan [2005](#page-5-0)), Clostridium (Gasoni et al. [2001\)](#page-4-0), and Herbaspirillum (Baldani et al. [2000\)](#page-4-0).

Pseudomonas spp. have been shown to contribute to increased plant yield and reduced levels of deleterious microorganisms, in field and greenhouse studies (Rovera et al. [2008](#page-5-0)), and have therefore been extensively studied (Duijff et al. [1997;](#page-4-0) Raunskov et al. [1999](#page-5-0); Sastry et al. [2000](#page-5-0); Vázquez et al. [2000](#page-5-0); Kohler et al. [2006\)](#page-4-0).

The species *Pseudomonas aurantiaca* was recently reclassified as P. chlororaphis subsp. aurantiaca (Peix et al. [2007\)](#page-4-0). P. chlororaphis subsp. aurantiaca strain SR1 (hereafter termed ''SR1'') was isolated from soybean rhizosphere by our group, in the area of Río Cuarto, Córdoba, Argentina. SR1 has the abilities to inhibit various fungal species in vitro (Rosas et al. [2001\)](#page-5-0); to produce siderophores, HCN, and phytohormone-like substances; to solubilize phosphates; and to colonize root systems of various crops (Rosas et al. [2005](#page-5-0); Rovera et al. [2008\)](#page-5-0). We showed previously that SR1 maintains stable populations in the rhizosphere and the internal structure of plants (Rosas et al. [2005\)](#page-5-0).

In the present study, we evaluated the effects of inoculating wheat plants with SR1 under field conditions, in terms of the above properties.

## Materials and methods

Study site and field inoculation technique

Studies were performed at the experimental field site of the Universidad Nacional de Río Cuarto, Río Cuarto, Argentina, (35°07'S, 64°14'W, 421 m.s.n.m.). The soil was Haplustol fine franc-sandy type, with the following physicochemical characteristics: pH (in water), 6.30; electrical conductivity (dS/m), 0.28; organic matter, 2.56%; nitrogen from nitrates, 11.60 ppm; nitrates, 51 ppm; available P, 19.7 ppm. The site has a temperate climate, with average annual rainfall 800 mm and mean annual temperature  $16-17^{\circ}$ C.

We used a randomized complete block design with seven blocks. Each block consisted of six plots (one per treatment; each 7.20  $\text{m}^2$ ); plots were separated by a distance of 1 m. Six rows (separated by 0.20 m) per block were sowed using a plot seed drill. Seed sowing density was 120 kg  $ha^{-1}$ . Seeds were inoculated with SR1, according to a formulation prepared by *Laboratorios Biagro S.A.* (10<sup>9</sup> CFU  $g^{-1}$  of peat), by the following procedure: 40 g inoculant, 20 g S2 adherent (Laboratorios Biagro S.A), and 5 g cell protector S1

(Laboratorios Biagro S.A.) were mixed in 80 ml water. Twelve grams of this mixture was added to 1 kg wheat seeds, to obtain a colony count of  $10^5$  CFU g<sup>-1</sup> seeds.

Six treatments were established in order to evaluate the promoting effect of P. chlororaphis subsp. aurantiaca SR1: (1) seed inoculated with SR1 in soil without fertilization; (2) seed without inoculation in soil fertilized with 80 kg ha<sup>-1</sup> urea and 60 kg ha<sup>-1</sup> diamonic phosphate (''100% dose''); (3) seed inoculated with SR1 in soil fertilized with 100% dose; (4) seed without inoculation in soil fertilized with 40 kg ha<sup>-1</sup> urea and 30 kg ha<sup>-1</sup> diamonic phosphate (''50% dose''); (5) seed inoculated with SR1 in soil fertilized with 50% dose; (6) seed without inoculation in soil without fertilization (control).

Weeds were removed manually. Plants were watered by a sprinkler, as needed, in all growth stages.

### Growth and yield parameters

Parameters were recorded at the growth stages termed emergence of seedlings, 1.5 (5 leaves), 3.0 (tillering), and 11.4 (ripe for harvest) (Feekes International Scale—Large [1954](#page-4-0)). At emergence of seedlings stage, the number of seedlings emerging per  $m^2$  was evaluated using a  $\frac{1}{4}$  m<sup>2</sup> iron ring. At Feekes 1.5 and 3.0 stages, the following growth parameters were measured: length from base to tip of leaf (cm), root length (cm) (Newman [1966](#page-4-0)), number of tillers, root volume  $(cm<sup>3</sup>)$  (measured by volume displace-ment of water) (Díaz Vargas et al. [2001\)](#page-4-0), shoot and root fresh and dry weight (72 h at  $60^{\circ}$ C). For each parameter, mean value was calculated from 5 plants per plot (seven plots per treatment).

Yield parameters evaluated were:  $kg \text{ ha}^{-1}$ , weight of 1,000 grains, number of spikes per plant, and number of grains per spike. These parameters were determined after creating clearances of 1 m at the edges of each plot, and 2 sowing lines at each side.

#### Statistical analysis

Data were subjected to analysis of variance (ANOVA). When ANOVA showed treatment effects ( $P < 0.05$ ), the least significant difference test (LSD) was applied to make comparisons among the means ( $P < 0.05$ ). The Statgraphics Plus 4.1 program (Statistical Graphics Corp.) was used for all analyses.

## Results

Effects of inoculation of wheat with SR1 were evaluated over all stages of the crop cycle. Inoculation had no effect on germination or emergence of seedlings. The number of

plants per  $m<sup>2</sup>$  was larger for the inoculation treatment than for fertilization without inoculation (Table 1).

For Feekes 1.5 stage, increase in shoot length (up to 14%) was observed for the inoculated/ unfertilized treatment and for 50% dose fertilization. For Feekes 3.0 stage, statistically significant increases relative to control were observed, particularly for inoculation plus 100% dose fertilization, in which shoot length increased 60% (Table 2).

SR1-inoculated plants showed root length increase up to 78% in Feekes 1.5 and 75% in Feekes 3.0 stages. Root volume increased significantly in Feekes 1.5 stage for inoculation plus 100% dose fertilization (Table 3).

Results for shoot biomass were variable, but all mean values for inoculation and/or fertilization treatments were higher than those of control. Root biomass at Feekes 1.5 stage was greatly increased by inoculation plus 50% dose fertilization, to the same degree as 100% dose fertilization without inoculation. At Feekes 3.0 stage, root biomass was significantly increased by inoculation even without fertilization (Table [4](#page-3-0)).

Number of tillers was increased relative to control at Feekes 3.0 stage by inoculation with or without fertilization (Table [5](#page-3-0)).

Regarding the yield parameters, kg  $ha^{-1}$  value was significantly higher than control (by 636) for SR1 inoculation plus 50% dose fertilization, and by 865 for inoculation plus 100% dose fertilization. Inoculation with or without fertilization increased yield up to 217 kg ha<sup>-1</sup> relative to control. Relative to fertilization-only treatments, inoculation increased yield by 5.5 kg ha<sup>-1</sup> (Table [6](#page-3-0)). Yield-promoting effects of inoculation were also reflected in weight of 1,000 grains and number of spikes per plant. Values of these parameters for inoculation plus 50% dose fertilization were consistently higher than for control, although the differences were not statistically significant. Regarding number of grains per spike, values for inoculation treatments were always higher than for control; the highest value was observed for inoculation plus 50% dose fertilization (Table [6\)](#page-3-0).

**Table 1** Wheat emergence (plants per  $m<sup>2</sup>$ )

Treatments	Emergence
1. Inoculated seeds, unfertilized soil	534.86 <sup>a</sup>
2. 100% dose fertilized (80 kg ha <sup>-1</sup> ) urea -- 60 kg ha <sup>-1</sup> diamonic phosphate)	$357.7^{b}$
3. Inoculated plus 100% dose fertilized	$571.43^{\rm a}$
4. 50% dose fertilized (40 kg ha <sup>-1</sup> ) urea - 30 kg ha <sup>-1</sup> diamonic phosphate)	384.57 <sup>b</sup>
5. Inoculated plus 50% dose fertilized	$540.00^{\rm a}$
6. Uninoculated seeds, unfertilized soil (control)	$542.00^{\rm a}$

<sup>a,b</sup> Significant differences by LSD test ( $P < 0.05$ )

Table 2 Shoot length at Feekes 1.5 and 3.0 stages



a,b,c Significant differences by LSD test ( $P < 0.05$ )

Table 3 Root length and volume at Feekes 1.5 and 3.0 stages

Treatments	Root length (cm)		Root (cm <sup>3</sup> )	volume
	<b>Feekes</b> 1.5	Feekes 3.0	Feekes 1.5	Feekes 3.0
1. Inoculated seeds, unfertilized soil		$160.40^a$ 339.64 <sup>a</sup> 1.77 <sup>a</sup>		4.47 <sup>a</sup>
$2.100\%$ dose fertilized $(80 \text{ kg ha}^{-1} \text{ urea})$ $-60 \text{ kg ha}^{-1}$ diamonic phosphate)		$87.15^{\rm b}$ 357.50 <sup>a</sup> 1.29 <sup>b</sup>		$4.23^{\rm a}$
3. Inoculated plus 100% dose fertilized		$181.49^a$ 346.28 <sup>a</sup> 2.09 <sup>a</sup>		$3.97^{\rm a}$
4. 50% dose fertilized $(40 \text{ kg ha}^{-1} \text{ urea})$ $-30 \text{ kg ha}^{-1}$ diamonic phosphate)		$159.59^a$ $235.52^b$ $1.20^b$		$2.46^{b}$
5. Inoculated plus 50% dose fertilized		$150.16^a$ 359.30 <sup>a</sup> 1.66 <sup>a</sup>		$3.69^{\rm a}$
6. Uninoculated seeds, unfertilized soil (control)		$101.89^b$ 204.92 <sup>b</sup> 0.91 <sup>c</sup>		2.71 <sup>b</sup>

a,b,c Significant differences by LSD test ( $P < 0.05$ )

## Discussion

This is the first field study in Argentina of P. chlororaphis subsp. aurantiaca strain SR1 inoculation effects. The results are promising. Effects of SR1 inoculation were variable, depending on the growth or yield parameter, and the plant growth stage recorded.

The decrease in seedling emergence we observed for 50% and 100% dose fertilization without inoculation (treatments 2 and 4, Table 1) may be related to application methodology, e.g., a phytotoxic effect by direct contact of seeds with phosphorus and nitrogen. Perhaps components of some formulations used (peat, S1, S2, SR1) reduced

<span id="page-3-0"></span>



a,b,c,d Significant differences by LSD test ( $P \lt 0.05$ ), ns: no significant differences

Table 5 Number of tillers

<b>Treatments</b>	Number of tillers
1. Inoculated seeds, unfertilized soil	3.00 <sup>a</sup>
2. 100% dose fertilized (80 kg ha <sup>-1</sup> ) urea -- 60 kg ha <sup>-1</sup> diamonic phosphate)	3.17 <sup>a</sup>
3. Inoculated plus 100% dose fertilized	$2.63^{ab}$
4. 50% dose fertilized (40 kg ha <sup>-1</sup> ) urea - 30 kg ha <sup>-1</sup> diamonic phosphate)	$2.46^{ab}$
5. Inoculated plus 50% dose fertilized	$2.69^{ab}$
6. Uninoculated seeds, unfertilized soil (control)	2.00 <sup>b</sup>

a,b Significant differences by LSD test ( $P < 0.05$ )

Table 6 Wheat yield components

phytotoxicity by avoiding direct contact between the chemical fertilizer and seeds. However, such difference in number of plants per  $m<sup>2</sup>$  did not alter final yield because of the compensation that took place during the Feekes 3.0 stage. Moreover, other authors reported emergence promotion effects in wheat inoculated with fluorescent Pseudomonas (Luz 2001); our results didn't show significant differences on this parameter in the inoculated treatments, relative to control.



The root growth parameters (length, volume, dry biomass) increased  $>60\%$  during Feekes 3.0 stage in SR1inoculated plants, compared to controls. The effect of SR1 plus 50% dose fertilizer was not significantly different from that of 100% dose fertilizer. Previous studies have shown that rhizobacteria increase root absorption capacity of gramineous plants when the dosage of nitrogen fertilizer applied to soil is reduced (Trân Van et al. [2000;](#page-5-0) Whitmore [2000](#page-5-0)). Increased root weight, mediated by rhizobacteria, is generally associated with inoculation, and enhances access of the plant to soil nutrients (Kohler et al. [2006](#page-4-0)).

Values of the yield component kg  $ha^{-1}$  were, on average, 5.5 kg ha<sup>-1</sup> higher for SR1 inoculation than for



a,b,c Significant differences by LSD test ( $P < 0.05$ ), ns: no significant differences

<span id="page-4-0"></span>fertilization treatments. The value for inoculation plus 50% dose fertilization was significantly higher than either control or 100% dose fertilization (Table [6\)](#page-3-0).

Important conclusions from this study are: (i) inoculation with SR1 promoted both growth and yield of wheat; (ii) the dosages of chemical fertilizers currently applied in commercial wheat fields in Argentina could be reduced through proper combination of SR1 inoculation plus fertilization.

Acknowledgements This work was supported by grants from the Secretaría de Ciencia y Técnica of Universidad Nacional de Río Cuarto (UNRC) and the Agencia Nacional de Promoción Científica, Tecnológica y de Innovación Productiva (CONICET) of the República Argentina. We gratefully acknowledge Biagro Laboratories S.A for providing the inoculants and Dr. S. Anderson for editing the manuscript.

## References

- Andrade G, De Leij FAAM, Lynch JM (1998) Plant mediated interactions between Pseudomonas fluorescens, Rhizobium leguminosarum and arbuscular mycorrhizae on pea. Lett Appl Microbiol 26:311–316. doi:[10.1046/j.1472-765X.1998.00337.x](http://dx.doi.org/10.1046/j.1472-765X.1998.00337.x)
- Baldani VLD, Baldani JJ, Döbereiner J (2000) Inoculation of rice plants with the endophytic diazotrophs Herbaspirillum seropediacae spp. Biol Fertil Soils 30:485–491. doi[:10.1007/s0037400](http://dx.doi.org/10.1007/s003740050027) [50027](http://dx.doi.org/10.1007/s003740050027)
- Bashan Y (1998) Inoculants of plant growth-promoting bacteria for use in agriculture. Biotechnol Adv 16:729–770. doi[:10.1016/](http://dx.doi.org/10.1016/S0734-9750(98)00003-2) [S0734-9750\(98\)00003-2](http://dx.doi.org/10.1016/S0734-9750(98)00003-2)
- Bashan Y, Levanony H (1990) Current status of Azospirillum inoculation technology: Azospirillum as a challenge for agriculture. Can J Microbiol 36:591–599
- Bull CT, Weller DM, Thomashow LS (1991) Relationships between root colonization and suppression of Gaeumannomyces graminis var tritici by Pseudomonas fluorescens strain 2-79. Phytopathology 81:954–959. doi[:10.1094/Phyto-81-954](http://dx.doi.org/10.1094/Phyto-81-954)
- Caballero-Mellado J, Carcaño-Montiel MG, Mascarúa-Esparza MA (1992) Field inoculation of wheat (Triticum aestivum) with Azospirillum brasilense under temperate climate. Symbiosis 13: 243–253
- da Luz WC (2001) Evaluation of plant growth-promoting and bioprotecting rhizobacteria on wheat crop. Fitopatol Bras 26: 597–600
- Dashti N, Zhang F, Hynes R, Smith DL (1998) Plant growth-promoting rhizobacteria accelerate nodulation and increase nitrogen fixation activity by field grown soybean [Glycine max(L.) Merr.] under short season conditions. Plant Soil 200:205–213. doi[:10.1023/](http://dx.doi.org/10.1023/A:1004358100856) [A:1004358100856](http://dx.doi.org/10.1023/A:1004358100856)
- de Freitas JR (2000) Yield and N assimilation of winter wheat (Triticum aestivum L., var Norstar) inoculated with rhizobacteria. Pedobiologia (Jena) 44:97–104. doi[:10.1078/S0031-4056\(04\)70031-1](http://dx.doi.org/10.1078/S0031-4056(04)70031-1)
- de Weger LA, van der Bij AJ, Dekkers LC, Simons M, Wijffelman CA, Lugtenberg BJJ (1995) Colonization of the rhizosphere of crop plants by plant-beneficial pseudomonads. FEMS Microbiol Ecol 17(4):221–227. doi[:10.1111/j.1574-6941.1995.tb00146.x](http://dx.doi.org/10.1111/j.1574-6941.1995.tb00146.x)
- Del Gallo M, Fabbri P (1990) Inoculation of Azospirillum brasilense Cd on chickpea (Cicer arietinum. Symbiosis 9:283–287
- Díaz Vargas P, Ferrera-Cerrato R, Almaraz-Suárez JJ, Alcántar Gonzalez G (2001) Inoculación de bacterias promotoras de crecimiento en lechuga. Terra Latinoam 19:329–335
- Dubeikovsky AN, Mordukhova EA, Kochetkov VV, Polikarpoca FY, Boronin AM (1993) Growth promotion of blackcurrant softwood cuttings by recombinant strain Pseudomonas fluorescens BSP53a synthesizing and increased amount of indol-3-acetic acid. Soil Biol Biochem 25:1277–1281. doi[:10.1016/0038-0717\(93\)90225-Z](http://dx.doi.org/10.1016/0038-0717(93)90225-Z)
- Duijff BJ, Gianinazzi-Pearson V, Lemanceau P (1997) Involvement of the outer membrane lipopolysaccharides in the endophytic colonization of tomato roots by biocontrol Pseudomonas fluorescens strain WCS417r. New Phytol 135:325–334. doi:[10.1046/j.](http://dx.doi.org/10.1046/j.1469-8137.1997.00646.x) [1469-8137.1997.00646.x](http://dx.doi.org/10.1046/j.1469-8137.1997.00646.x)
- Fridlender M, Inbar J, Chet I (1993) Biological control of soil-borne plant pathogens by a  $\beta$ -1,3-glucanase-producing *Pseudomonas* cepacia. Soil Biol Biochem 25:1211–1221. doi:[10.1016/0038-](http://dx.doi.org/10.1016/0038-0717(93)90217-Y) [0717\(93\)90217-Y](http://dx.doi.org/10.1016/0038-0717(93)90217-Y)
- Fulchieri M, Frioni L (1994) Azospirillum inoculation on maize (Zea mays): effect on yield in a field experiment in central Argentina. Soil Biol Biochem 26:921–923. doi:[10.1016/0038-0717\(94\)](http://dx.doi.org/10.1016/0038-0717(94)90308-5) [90308-5](http://dx.doi.org/10.1016/0038-0717(94)90308-5)
- García-González MM, Farías-Rodríguez R, Peña-Cabriales JJ, Sánchez-Yáñez JM (2005) Inoculación del Trigo var. Pavón con Azospirillum spp y Azotobacter beijerinckii. Terra Latinoam 23:65–72
- Gasoni L, Cozzi J, Kobayashi K, Yossen V, Zumelzu G, Babbitt S et al (2001) Yield response of lettuce and potato to bacterial and fungal inoculants under field conditions in Córdoba (Argentina). J Plant Dis Prot 108(5):530–535
- Hebbar KP, Atkinson D, Tucker W, Dart PJ (1992) Suppression of Fusarium moliniforme by maize root-associated Pseudomonas cepacia. Soil Biol Biochem 24(10):1009–1020. doi[:10.1016/](http://dx.doi.org/10.1016/0038-0717(92)90029-W) [0038-0717\(92\)90029-W](http://dx.doi.org/10.1016/0038-0717(92)90029-W)
- Höflich G, Wiehe W, Hecht-Buchholz CH (1995) Rhizosphere colonization of different growth- promoting Pseudomonas and Rhizobium bacteria. Microbiol Res 150:139–147
- Kloepper JW, Schroth MN (1978) Plant growth-promoting rhizobacteria on radishes. In: Station the Pathologie Végétale et Phytobactériologie, INRA, Angers (ed) Proceedings of the fourth international conference on plant pathogenic bacteria, vol 2 . Gilbert Clary, Tours, France, pp 879–882
- Kohler J, Caravaca F, Carrasco L, Roldán A (2006) Contribution of Pseudomonas mendocina and Glomus intraradices to aggregate stabilization and promotion of biological fertility in rhizosphere soil of lettuce plants under field conditions. Soil Use Manage 22(3):298–304. doi:[10.1111/j.1475-2743.2006.00041.x](http://dx.doi.org/10.1111/j.1475-2743.2006.00041.x)
- Large EC (1954) Growth stages in cereals. Illustration of the Feekes scale. Plant Pathol 3(4):128–125. doi:[10.1111/j.1365-3059.1954.](http://dx.doi.org/10.1111/j.1365-3059.1954.tb00716.x) [tb00716.x](http://dx.doi.org/10.1111/j.1365-3059.1954.tb00716.x)
- Lifshitz R, Kloepper JW, Kozlowsky M, Simonson C, Carlson J, Tipping EM et al (1987) Growth promotion of canola (rapeseed) seedlings by strain of Pseudomonas putida under gnotobiotic conditions. Can J Microbiol 33(5):390–395
- Loper JE, Buyer JS (1991) Siderophores in microbial interactions on plant surfaces. Mol Plant Microbe Interact 4:5–13
- Lucy M, Reed E, Glick BR (2004) Applications of free living plant growth-promoting rhizobacteria. Antonie Van Leeuwenhoek 86:1–25. doi[:10.1023/B:ANTO.0000024903.10757.6e](http://dx.doi.org/10.1023/B:ANTO.0000024903.10757.6e)
- Newman E (1966) A method of estimating the total length of root in a sample. J Appl Ecol 2:139–145. doi:[10.2307/2401670](http://dx.doi.org/10.2307/2401670)
- Okon Y, Labandera-González CA (1994) Agronomic applications of Azospirillum: an evaluation of 20 years of worldwide field inoculation. Soil Biol Biochem 26:1591–1601. doi[:10.1016/](http://dx.doi.org/10.1016/0038-0717(94)90311-5) [0038-0717\(94\)90311-5](http://dx.doi.org/10.1016/0038-0717(94)90311-5)
- Peix A, Valverde A, Rivas R, Igual JM, Ramírez-Bahena M, Mateos PF, et al (2007) Reclassification of Pseudomonas aurantiaca as a synonym of Pseudomonas chlororaphis and proposal of three subspecies, P. chlororaphis subsp. chlororaphis subsp. nov., P. chlororaphis subsp. aureofaciens subsp. nov., comb. nov. and

<span id="page-5-0"></span>P. chlororaphis subsp. aurantiaca subsp. nov., comb. nov. Int J Syst Evol Microbiol 57:1286–1290. doi:10.1099/ijs.0.64621-0

- Raunskov S, Nybroe O, Jakobsen I (1999) Influence of an arbuscular mycorrhizal fungus on Pseudomonas fluorescens DF57 in rhizosphere and hyphosphere soil. New Phytol 142:113–122. doi[:10.1046/j.1469-8137.1999.00374.x](http://dx.doi.org/10.1046/j.1469-8137.1999.00374.x)
- Rai SN, Gaur AC (1988) Characterization of Azotobacter spp. and effect of Azotobacter and Azospirillum as inoculant on the yield and N-uptake of wheat crop. Plant Soil 109(1):131–134. doi: [10.1007/BF02197592](http://dx.doi.org/10.1007/BF02197592)
- Rosas SB, Altamirano F, Schroder E, Correa N (2001) In vitro biocontrol activity of Pseudomonas aurantiaca. **Dyton-Interna**tional. J Exp Bot 67:203–209
- Rosas S, Rovera M, Andrés JA, Pastor NA, Guiñazú LB, Carlier E, et al (2005) In: Sorvari S, Toldo O (eds) Proceeding prospects and applications for plant associated microbes. 1st International conference on plant–microbe interactions: endophytes and biocontrol agents. Lapland, Finland, pp 91–99
- Rovera M, Carlier E, Pasluosta C, Avanzini G, Andres J, Rosas S (2008) Pseudomonas aurantiaca: plant growth promoting traits, secondary metabolites and inoculation response. In: Ahmad I, Pichtel J, Hayat S (eds) Plant–bacteria interactions. Strategies and techniques to promote plant growth. Wiley–VCH, Germany, pp 155–164
- Sastry MSR, Sharma AK, Johri BN (2000) Effect of an AM fungal consortium and Pseudomonas on the growth and nutrient uptake of Eucalyptus hybrid. Mycorrhiza 10:55–61. doi[:10.1007/s0057](http://dx.doi.org/10.1007/s005720000057) [20000057](http://dx.doi.org/10.1007/s005720000057)
- Thomashow LS, Weller DM, Bonsall RF, Pierson LSIII (1990) Production of the antibiotic Phenazine-1-carboxylic acid by fluorescent Pseudomonas species in the rhizosphere of wheat. Appl Environ Microbiol 56(4):908–912
- Trân Van V, Berge O, Ngô Kê J, Balandrau J, Heulin T (2000) Repeated beneficial effects of rice inoculation with a strain of Burkholderia vietnamiensis on early and late yield components in low fertility sulphate acid soils of Vietnam. Plant Soil 218:273–284
- Vázquez MM, César S, Azcón R, Barea JM (2000) Interactions between arbuscular mycorrhizal fungi and other microbial inoculants (Azospirillum, Pseudomonas, Trichoderma) and their effects on microbial population and enzyme activities in the rhizosphere of maize plants. Appl Soil Ecol 15:261–272. doi: [10.1016/S0929-1393\(00\)00075-5](http://dx.doi.org/10.1016/S0929-1393(00)00075-5)
- Voisard C, Keel C, Hass D, Defago G (1989) Cyanide production by Pseudomonas fluorescens helps suppress black root rot of tobacco under gnotobiotic conditions. EMBO J 8:351–358
- Weller DM (1998) Biological control of soil-borne plant pathogens in the rhizosphere with bacteria. Annu Rev Phytopathol 26:379– 407. doi[:10.1146/annurev.py.26.090188.002115](http://dx.doi.org/10.1146/annurev.py.26.090188.002115)
- Weller DM, Thomashow LS (1993) Advances in rhizobacteria for biocontrol. Curr Opin Biotechnol 4:306–311. doi:[10.1016/0958-](http://dx.doi.org/10.1016/0958-1669(93)90100-B) [1669\(93\)90100-B](http://dx.doi.org/10.1016/0958-1669(93)90100-B)
- Whitmore AP (2000) The biological management of soil fertility project. Neth J Agric Sci 48:115–122
- Zaidi A, Khan S (2005) Interactive effect of rhizotrophic microorganisms on growth, yield, and nutrient uptake of wheat. J Plant Nutr 28(12):2079–2092. doi:[10.1080/01904160500320897](http://dx.doi.org/10.1080/01904160500320897)